

1 **Combining geometric morphometric analyses of multiple 2D**
2 **observation views improves interpretation of evolutionary allometry**
3 **and shape diversification in monitor lizard (*Varanus*) crania**

4

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7 **Key words:** two-dimensional geometric morphometrics; cranium; *Varanus*; landmark;
8 observation view.

9 **ABSTRACT**

10 Geometric morphometrics is a powerful tool for the quantification, visualization and analysis of
11 morphological variation and change. This approach is being applied more frequently in a
12 phylogenetic comparative context to assess the relative influence of size, ecology, function, and
13 developmental constraints on morphological evolution. Geometric morphometric methods rely on
14 homologous landmarks as the source of shape data, and the level of detail and accuracy increases
15 with the amount of information contained in a landmark configuration. However, it may be possible
16 to capture particular elements of shape variation by concentrating on different observation angles of
17 a complex structure like the vertebrate cranium. Our study examines how observation view (dorsal,
18 ventral and lateral) influences 2D geometric morphometric analysis of interspecific cranial shape
19 variation in monitor lizards. We recover strong phylogenetic signal in all three views and general
20 concordance in patterns of size-corrected shape diversification within the genus. However, we also
21 find subtle but important differences among views in analyses of evolutionary allometry and shape
22 variation, which may reflect both landmark configuration design and adaptive functional trends of
23 the study system. Our study shows that studies restricted to a 2D geometric morphometric analysis
24 of a complex 3D biological structure can combine carefully designed 2D landmark configurations
25 describing alternative planes to maximize shape coverage.

26 INTRODUCTION

27 The field of morphometrics is a central component of biology that quantifies biological shape,
28 shape variation, and its correlation with other variables (Bookstein, 1991; Adams, Rohlf & Slice,
29 2004; Webster & Sheets, 2010; Adams, Rohlf & Slice, 2013). Geometric morphometrics are a
30 popular set of methods that restrict data to locations of discrete anatomical points (landmarks), and
31 rely on a constellation of points (the landmark configuration) to sample morphology (Bookstein,
32 1986; Bookstein, 1993; Corti, 1993; O'Higgins, 2000). Designing an appropriate landmark
33 configuration is a critical early step in geometric morphometric analysis and ideally, three broad
34 goals are achieved through appropriate selection of landmarks - a configuration should cover the
35 variation clearly observable among objects, provide a sufficiently comprehensive and biologically
36 significant sampling of morphology, and deliberately test the hypotheses proposed (Roth, 1993;
37 O'Higgins, 2000; Oxnard & O'Higgins, 2009; Zelditch, Swiderski & Sheets, 2013).

38
39 Geometric morphometric studies frequently use a two-dimensional landmark configuration to
40 quantify form, even though the structure of interest is typically three-dimensional (Roth, 1993;
41 Cardini, 2014; Klingenberg, 2015). **While the 3D approach is becoming more accessible, for many
42 it is still a more expensive and time consuming processes that requires large data storage facilities.**

43 The 2D approach has several advantages over 3D, in particular the ease of capturing 2D coordinates
44 by placing points on a photograph using freely available, user-friendly software such as tpsDig
45 (Rohlf, 2010a), and the intuitively understood visualizations of shape variation produced using the
46 thin-plate spline (Klingenberg, 2013a). It is also much faster and so it is easier to obtain much larger
47 sample sizes to quantify variation. The obvious disadvantage of a 2D approach is the loss of
48 information regarding depth, and both correlational and comparative studies of 2D versus 3D
49 datasets suggest that including the third coordinate may lead to different results for *some* biological
50 structures (Cardini & Thorington, 2006; Álvarez & Perez, 2013; Cardini, 2014). Despite the fact
51 that 2D morphometric analyses of 3D structures mean the loss of tridimensional information, the

52 relative increased costs, time and difficulty associated with 3D data collection, storage and analysis
53 still make 2D geometric morphometrics a realistic, inexpensive, and time-effective choice for many
54 researchers (Cardini, 2014; and references therein).

55

56 The vertebrate head is a complex structure that has received much attention in morphometric
57 studies, perhaps due to the enormous disparity in form that has evolved among groups. At the
58 broadest scale, the skull is comprised of three parts: the cranial vault, the cranial base, and face (de
59 Beer, 1937). Although these different parts originate in embryologically distinct regions, they
60 apparently grow in a morphologically integrated manner through numerous developmental and
61 functional interactions (Lieberman, Ross & Ravosa, 2000; Bookstein *et al.* 2003; Sanger *et al.*
62 2012). This has implications for 2D geometric morphometric analyses of head shape diversification
63 regarding which observation view (or combination of views) is most informative for any vertebrate
64 group. An observation view is the orientation of the specimen in relationship to observer, and the
65 view(s) chosen by a researcher may influence the nature of the morphometric data yielded from a
66 given specimen. This is problematic as the numerous structural subunits of the head observable
67 from different views may be semi-independent in ontogeny or function, and are thus expected to
68 vary in conservative properties and respond differently to ecological and evolutionary factors
69 (Klingenberg, 1996; Lu *et al.* 2014). It is not understood how profoundly 2D shape analyses of the
70 vertebrate head are influenced by observation view(s) and the landmark configuration used to
71 characterize them.

72

73 Lizards are ideal for such examination, as they have a remarkable diversity in cranial form that is
74 tightly linked to the functional and constructional demands of the skull, and show a unique
75 evolutionary trend toward increased fenestration and reduced cranial ossification (Evans, 2003;
76 Stayton, 2005; Herrel *et al.* 2007; Daza *et al.* 2008). The ventral morphology of lizard crania,
77 formed by parts of the dermatocranium and neurocranium, is directly involved in the mechanics of

78 feeding, affecting bite force through connections to the jaw musculature (Herrel *et al.* 2007). Jaw
79 musculature is also associated with structures most clearly observed from a lateral view (for
80 example the postorbital-postfrontal, quadrate, coronoid and jugal). Consequently, much shape
81 variation observed in lateral view is strongly linked with diet, for example herbivorous lizards tend
82 to have short snouts and high skulls, which is consistent with a slow but powerful bite relative to
83 carnivorous lizards (Evans, 2003; Stayton, 2005). Palate morphology and the septomaxilla also
84 reflect the evolution of the chemosensory system in general, and the vomeronasal system in
85 particular (Rieppel, Gauthier & Maisano, 2008), and the skull roofing bones are greatly influenced
86 by the development of the brain and other sensory organs (Haas, 1973). The most highly fenestrated
87 **cranial** morphology belongs to the monitor lizards (family: Varanidae), and is known as a ‘space
88 frame’ type of construction in engineering terms (McCurry *et al.* 2015). The monitor cranium has a
89 noteworthy mixture of extremely reduced bones (such as those forming the skull roof, particularly
90 the snout region), derived bones (such as the palpebral and dermal palate elements), and composite
91 bones (such as the fused postorbitofrontal) (Bellairs, 1949; Frazzetta, 1962; Auffenberg, 1988;
92 Rieppel *et al.* 2008; Werneburg *et al.* 2015). Further, 2D geometric morphometric analysis of lateral
93 **cranial** shape among lizard families reveals monitors have **conspicuous** patterns of interspecific
94 cranial shape disparity (Stayton, 2005).

95

96 Our study examines how observation view and landmark configuration design influence 2D
97 geometric morphometrics analysis of interspecific cranial shape variation in monitor lizards.
98 Specifically, we quantify the covariation of species **cranial** shapes with phylogeny and allometry,
99 and characterize major patterns of interspecific cranial shape variation and size-corrected
100 diversification in morphospace. We then identify and evaluate differences among **2D** views to
101 **determine** the utility of combining multiple views, and to assess whether choosing different views
102 for landmark-based morphometric analyses could influence our quantification of **shape** differences
103 among specimens or species, **providing complementary morphological information to each other.**

104 We expect congruence among views for quantification of shape covariation with phylogeny to
105 reflect developmental constraints within the cranium. A pervasive influence of allometry on shape
106 is predicted, and should be exemplified in monitor lizard morphology, as species differ in body
107 mass over four orders of magnitude (Pianka, 1995). However, we predict the anatomical points
108 sampled to reflect functionally different characters among views, and therefore to recover
109 discrepant patterns of interspecific cranial shape variation and size-corrected diversification.

110

111 **MATERIALS AND METHODS**

112 **Morphometric data**

113 Our morphometric dataset comprises images of the cranium for 29 *Varanus* species (n = 152
114 specimens) in lateral, dorsal and ventral views (Supp. Table S1). We only used cranial material of
115 adult specimens (skeletal maturity judged by specimen size) and we did not know the sex of the
116 specimens. Within a species, male and female varanids have the same shaped heads but subtle
117 sexual dimorphism in head size has been detected in some of the larger monitor species (Frýdlová,
118 Velenský, Šimková *et al.* 2011). Not knowing the sex in our study is unlikely to influence our
119 results because we correct for head size and our primary interest is in interspecific shape variation
120 rather than intraspecific size variation. A majority of the dataset is photographs of preserved crania,
121 obtained from collections in the American Museum of Natural History (AMNH), the Florida
122 Museum of Natural History (FLMNH), the Field Museum of Natural History (FMNH), the National
123 Museum of Victoria (NMV), the University of Michigan Museum of Zoology (UMMZ) and the
124 Smithsonian Institution National Museum of Natural History (USNM). Photographs were taken
125 with a mounted and leveled Olympus ® C-765 Ultra Zoom. Skulls were situated against a dark
126 background with a scale bar in the same plane. The lateral view involved positioning the crania so
127 the camera was perpendicular to the mid-sagittal plane. The dorsal view involved placing the
128 cranium ventral side down with the teeth and palate against the surface. The ventral view involved
129 positioning the skull so the palate was parallel to the camera lens. The dataset was supplemented

130 with 2D images obtained from micro X-ray computed tomography (CT) scans of crania from
131 whole, alcohol-preserved specimens from the Western Australian Museum (WAM) (11 species).
132 All scans were made with the Xradia microCT x400 system at the Australian Microscopy and
133 Microanalysis Research Facility (University of Sydney). CT scan data were rendered using Drishti
134 v2.0 (<http://anusf.anu.edu.au/Vizlab/drishti/>). Non-cranial volumes were digitally removed, and
135 images of the lateral, dorsal and ventral views of the cranium were then saved (including a scale
136 bar).

137

138 **Phylogeny**

139 The phylogenetic hypothesis used in this study is based on the published molecular phylogeny
140 presented in Vidal *et al.* (2012). We assembled the published data for 26 species included in Vidal
141 *et al.* (2012) and added three additional species from the published work of Welton *et al.* (2014)
142 (total: 29 species) (Fig. 1; Supp. Table S1). In order to include the three additional species we
143 generated a maximum-likelihood phylogenetic tree based on mitochondrial protein-coding genes
144 (ND3 and ND4) and nuclear (brain-derived neurotrophic factor, bone morphogenetic protein 2, and
145 neurotrophin 3) sequence data and constrained the major clades so that the resultant phylogeny
146 would be consistent with the phylogeny presented in Vidal *et al.* (2012). The resulting phylogeny
147 was used for visualizing shape diversification across *Varanus*, identifying shape differences among
148 different groups in a phylogenetic context, and to test for phylogenetic signal.

149

150 **2D Geometric morphometrics**

151 We used a 2D geometric morphometric approach for all views, also using a 2D sliding
152 semilandmark procedure for the ventral view (Bookstein, 1997). Landmarks and semilandmarks
153 were collected using tpsDig v. 2.16 (Rohlf, 2010a (Fig. 2; Supp. Table S2). The semilandmarks
154 were collected as a curve outlining the fused pre-maxillae and paired maxillae in ventral view (Fig.
155 2). These data were subsequently reduced to equidistant landmarks (four on the pre-maxillae and

156 ten on each maxilla), and subsequently defined as semilandmarks using the software tpsUtil v. 1.46
157 (Rohlf, 2010c). We then slid the landmarks using the bending energy method (Bookstein, 1997;
158 Gunz & Mitteroecker, 2013) implemented in the R statistics package *geomorph* (Adams & Otárola-
159 Castillo, 2013). The **original** landmark coordinate data were aligned using a generalized Procrustes
160 superimposition analysis (GPA), **providing the centroid size and shape coordinates** (Adams *et al.*
161 2013). The monitor cranium has matching symmetry in lateral view and object symmetry in dorsal
162 and ventral views. Although shape analysis procedures for both types of symmetry can separate the
163 symmetric (left-right averages) component of variation from left-right asymmetries within
164 individuals (Klingenberg, Barluenga & Meyer, 2002), damage to specimens in lateral view meant
165 we could only digitize landmarks on one side of the cranium. Shape data therefore refers to
166 Procrustes coordinates in lateral view, and the symmetric component of shape variation in dorsal
167 and ventral views. We then calculated the mean of the Procrustes coordinates and centroid sizes of
168 each species, in order to generate phylomorphospace plots and allow phylogenetic comparative
169 analyses on the shape data.

170

171 **Cranium size and shape variation**

172 Ignoring the phylogeny in preliminary transformations of species data results in substantially
173 elevated variance and type I error (false positive) in statistical estimators (Revell, 2009). To
174 quantify the **cranial** shape variation evolutionarily associated with allometry in the monitor lizard
175 cranium, we performed a Procrustes regression of shape on size, based on the Procrustes
176 coordinates averaged by species, using the software *geomorph* (Adams & Otárola-Castillo, 2013).
177 We also used the residuals from the averaged Procrustes coordinates and centroid sizes in order to
178 perform a phylogenetic regression of shape on size in a phylogenetic context, in order to get
179 ‘evolutionary allometry’-corrected shape data, also using *geomorph* (Adams & Otárola-Castillo,
180 2013). Analyzing the residuals from a regression of the size-dependent trait against size is a

181 common method of correcting for allometry in evolutionary studies (Gould, 1966; Monteiro, 1999;
182 Revell, 2009).

183

184 **Phylogenetic signal**

185 To quantify phylogenetic signal in *Varanus* crania for each of the views (dorsal, ventral, and
186 lateral), we used the K_{mult} method, which is Blomberg's K statistic generalization appropriate for
187 high-dimensional and/or multivariate data (Adams, 2014), using the R package *geomorph* (Adams
188 & Otárola-Castillo 2013). We determined the statistical significance of K_{mult} using phylogenetic
189 permutation with 10,000 iterations, which is calculated by permuting the shape data of the
190 Procrustes-aligned specimens among all tips of the phylogenetic tree. We also simulated the
191 confidence intervals of K under a Brownian Motion (BM) model of evolution using *phytools*
192 (Revell, 2012), with 10,000 iterations. Under BM, K_{mult} has an approximate expected value of 1.0,
193 and the higher the K_{mult} value is, the stronger the phylogenetic signal, indicating higher
194 morphological variance among clades instead of within clades (Adams, 2014).

195

196 **Patterns of cranial shape variation and diversification**

197 To examine patterns of cranial shape variation among *Varanus*, we subjected the averaged shape
198 coordinates to a Principal Component Analysis (PCA), both before and after size correction, for all
199 three views. To characterize the evolutionary patterns of cranial shape diversity, we projected the
200 phylogeny into the shape tangent space and **projected** it on plots species mean size-corrected **cranial**
201 shapes. This approach maps PC scores of the species cranial shapes to the phylogeny, computes the
202 PC scores at internal nodes, and subsequently projects the branches of the phylogenetic tree onto
203 the morphospace. Evaluating this 'phylomorphospace' provides insights into the history of
204 morphological diversification for complex traits and allows a multivariate visualization of shape
205 differences between species (Sidlauskas, 2008; Sherratt *et al.* 2014). To quantify the amount of
206 shape variation and dispersion between **original** and size-corrected shape data, we measured the

207 dispersion of all the individuals at **three** levels: (a) for the whole data set – by calculating the
208 dispersion around the mean shape for all the specimens, **(b) by species, and** (c) by the phylogenetic
209 groups identified in Vidal *et al.* (2012). **Finally, we tested the integration between the three views,**
210 **in order to assess the strength of dependency among each landmark configuration, with *geomorph***
211 **(Adams & Otárola-Castillo 2013).**

212

213 **RESULTS**

214 **Shape variation among *Varanus* crania**

215 In dorsal view, **original** shape variation captured by the two first Principal Components (PC 1-2)
216 relates to cranium width, relative length of the snout and parietal units, and angularity of the fronto-
217 parietal suture and nasal-frontal boundary (Fig. 3; Fig. 4). PC 1 (accounting for 54.9% of the overall
218 morphological variance) (Table 1) contrasts crania with a shortened snout complex and widened
219 parietal unit in the negative direction, and crania with a lengthened snout complex and narrowed
220 parietal unit in the positive direction. PC 2 (19.0% variance) contrasts narrow crania with a
221 relatively short snout in the negative direction, and broad crania with a relatively long snout in the
222 positive direction.

223

224 In ventral view, over 70% of the cranial shape variation among monitors relates to cranium width,
225 curvature of the snout, robusticity of the palatine and ectopterygoid, and length of the pterygoid
226 (Fig. 3; Fig. 4). For PC 1 (47.7% variance), a negative change from the mean describes crania with
227 a short and broad snout, distally shifted and broadened suborbital bones, and a broad and long
228 pterygoid. A positive change from the mean describes crania with an elongate and narrow snout,
229 mesially shifted and narrowed suborbital region, and a shortened pterygoid. PC 2 (24.7%) captures
230 relative shifts in landmarks altering pointedness of the snout, width of the posterior maxilla region,
231 mesial-distal shifting in the palatine and length of the pterygoid.

232

233 In lateral view, shape variation captured by PC 1-2 relates to overall cranium height, length and
234 angularity of the snout complex, convex-concave bending and length of the basal units, and
235 robustness of the parietal unit (Fig. 3; Fig. 4). For PC 1 (31.1%), a negative change from the mean
236 shape describes relative shifts in landmarks that flatten the cranium, lengthen the snout complex,
237 and both shorten and project the parietal and basal units mesially. A positive change from the mean
238 shape corresponds to a heightened cranium with a short but steeply rising snout complex, and
239 enlarged parietal and basal units. PC 2 (22.5%) describes more subtle shape changes, contrasting
240 crania with a lengthened snout complex, ventrally projected pterygoid, and steeply rising cranial
241 roof in the negative direction, and crania with a shortened snout complex, mesially projected
242 elongate pterygoid, and flattened cranial roof in the positive direction.

243

244 **Phylogenetic signal**

245 The results of the multivariate K-statistic calculated on the shape data are significant for each view
246 of the *Varanus* cranium (dorsal: $K_{\text{mult}} = 0.99$, $P < 0.001$; ventral: $K_{\text{mult}} = 0.85$; $P < 0.001$; lateral:
247 $K_{\text{mult}} = 0.83$, $P < 0.001$). K 95% confidence interval for values expected under a Brownian Motion
248 model of trait evolution = [0.658, 1.612].

249

250 **Cranial size and shape variation**

251 The multivariate Procrustes regressions of shape data on centroid size were statistically significant
252 ($P < 0.0001$) in dorsal, ventral and lateral view, indicating the presence of allometry in the *Varanus*
253 cranium. In the quantification of evolutionary allometry (when phylogenetic relatedness is
254 incorporated in the regression), the r^2 values decrease substantially but remain significant for all
255 three views (Table 2). The dorsal view recovers the largest r^2 values, followed by lateral view then
256 ventral view, suggesting that shape variation in the monitor lizard cranium is most heavily
257 constrained by size in the skull roof. The amount of shape variation accounted for by PC 1-5 is
258 substantially less after evolutionary allometry is removed from average species cranial shapes

259 (Table 1). The amount of dispersion and shape variation was very similar between original and size-
260 corrected shape data for each of the views, for all three levels (all individuals, by species, and by
261 phylogenetic groups; Table 3). The two-block PLS between different views pointed out very high
262 integration between dorsal and lateral ($r\text{-PLS} = 0.943, P < 0.001$) and between ventral and lateral
263 landmark configurations ($r\text{-PLS} = 0.921, P < 0.001$), and moderately high between dorsal and
264 ventral views ($r\text{-PLS} = 0.878, P < 0.001$).

265

266 **Size-corrected shape diversification in *Varanus* crania**

267 Superimposing the phylogeny onto a plot defined by PC 1-2 of evolutionary allometry-corrected
268 shape variation shows cranial shape diversification across species, depicting their great
269 morphological differences (Fig. S1). There is a moderate clustering of size-corrected species cranial
270 shapes according to phylogenetic group in ‘phylomorphospace’ for all three-observation views.
271 These clusters are most clearly separated from each other in dorsal view. The major difference in
272 diversification patterns among views is attributed to the African species, which occupy a positive
273 position on PC 2 in dorsal view, but a negative PC 2 position in ventral and lateral views. Cranial
274 shapes of species in the Indo-Australian C phylogenetic group show the greatest divergence from
275 all other species cranial shapes, occupying a distinct region of each phylomorphospace. Cranial
276 shapes of individual species from all other phylogenetic groups have diversified varying amounts in
277 each view, as shown by branches that traverse a large proportion of the region occupied by the
278 whole sample. These include the African species *Varanus exanthematicus* and *Varanus albigularis*,
279 the Indo-Asian species *Varanus olivaceus* and *Varanus rudicollis*, and the Indo-Australian species
280 *Varanus komodoensis* and *Varanus giganteus*.

281

282 **DISCUSSION**

283 The highly fenestrated space frame construction of monitor lizard crania creates clear differences in
284 abundance and spread of candidate landmarks among alternative views of the structure. It follows

285 that the design of each 2D landmark configuration inevitably describes shape in a slightly different
286 way. Phylogeny underpins patterns of gross morphological variation among monitor lizard crania,
287 even when diluted by size-related constraints and presumably adaptive trends, which might be also
288 phylogenetically constrained. The strength of association between cranial size and shape differs
289 among observation angles; however, it is unclear whether these results reflect genuine evolutionary
290 trends or slight differences on the morphological information captured by each landmark
291 configuration design. The patterns of cranial shape variation are broadly concordant among views,
292 suggesting dietary variation has contributed to shape diversification across the entire cranium at a
293 higher phylogenetic level. Below, we evaluate 2D characterization of the monitor cranium, present
294 an overview of cranial shape evolution in monitors, and discuss more broadly how 2D geometric
295 morphometric analyses of vertebrate head shape can be improved through analysis of multiple
296 observation views and careful landmark configuration design.

297

298 **The evolution of cranial morphology in monitor lizards**

299 A careful consideration of landmark configuration design is required when interpreting geometric
300 morphometric results. Our landmark configurations capture a dorsal-ventral ‘silhouette’ in lateral
301 view, the medial paired and fused bones in dorsal view, and palatopterygoid morphology in ventral
302 view (Fig. 2). A 2D characterization of monitor lizard cranial morphology is most comprehensive in
303 ventral view, as abundant type I and type II landmarks (Bookstein, 1991) are reliably digitized, and
304 four curves are traced to outline the fused pre-maxillae and paired maxillae. We find the lateral
305 view to be the least reliable, because of the difficulty in accurately orienting specimens when taking
306 photographs (Klingenberg, 2015). The requirement of 2D landmarks to be coplanar (Zelditch *et al.*
307 2013) is most limiting for landmark configuration design in lateral and dorsal view, and is the key
308 reason why semilandmarks outlining the pre-maxillae and maxillae were only added in ventral
309 view. There are also morphological differences clearly observable among species for numerous

310 bones (such as the palpebral); however, such variation should not be characterized using geometric
311 morphometric methods (Zelditch *et al.* 2004).

312

313 The major monitor lizard clades match biogeographic distribution well, dividing the African, Indo-
314 Asian, and Indo-Australian species (Fig. 1). An Asian origin for crown *Varanus* is likely, with
315 major dispersal events occurring to Africa approximately 41 million years ago (mya), and to
316 Australia approximately 32 mya (Vidal *et al.* 2012). Significant phylogenetic signal is found across
317 observation angles, and interspecific cranial shape variation still retains substantial phylogenetic
318 structure after correcting for evolutionary allometry; Fig. 4 shows that **cranial** shapes of closely
319 related species form moderate clusters in phylomorphospace. The importance of phylogenetic
320 affinity for explaining interspecific variation is recovered in geometric morphometric analyses of
321 lateral, dorsal and/or ventral **cranial** shape in other lizard studies (Stayton, 2005; Daza *et al.* 2009;
322 Openshaw & Keogh, 2014). Such clear phylogenetic structure in the morphometric data across
323 observation angles may indicate a role for developmental constraints in diversification of monitor
324 lizard **cranial** shape.

325

326 Monitors occupy a wide range of environments across their distribution, and the selective demands
327 imposed by habitat have driven the evolution of extreme body size disparity in the genus (with
328 extant species ranging in total body length as adults from 23 cm to 3 m) (Collar, Schulte & Losos,
329 2011). Strong size-shape **relationships** are known for monitor lizard body and head morphology
330 (Christian & Garland, 1996; Thompson & Withers, 1997; Collar *et al.* 2011; Openshaw & Keogh,
331 2014). Initially, we found strong and significant allometry in each view of the cranium; however,
332 controlling for non-independence of shape data due to shared ancestry reduced the estimation of
333 evolutionary allometry in all views. This also reflects the strong phylogenetic signal in body size
334 evolution for the group (Collar *et al.* 2011; Openshaw & Keogh, 2014). The inconsistency in
335 strength of evolutionary allometry among views could reflect discrepancy in the way each

336 observation view quantifies size: dorsal view recovers the strongest influence of evolutionary
337 allometry ($r^2 = 0.2072$), and is described by a landmark configuration that effectively captures
338 cranium length, but only covers cranium width at the posterior-most edges of the cranial table (Fig.
339 2).

340

341 The three observation views share notable similarities in patterns of interspecific cranial shape
342 variation (Fig. 3) readily linked with diversity in diet through particular influences on cranial
343 performance and strength. First, the shape changes captured by the primary axis of shape variation
344 (PC 1) reveals two cranial phenotypes: (1) relatively tall and broad crania with a short and steeply
345 rising snout; and (2) relatively flat and gracile crania with an elongate, slender snout. Monitor lizard
346 crania with greater widths and heights perform better at biting, pulling and shaking (as exemplified
347 by *V. komodoensis*) (Moreno *et al.* 2008; D'Amore *et al.* 2011). The most extreme examples of this
348 morphotype belong to durophagous species like *V. exanthematicus*, which are characteristically
349 short and wide with a tall snout (McCurry *et al.* 2015). These traits are often coupled with deep
350 mandibles and robust dentition (Rieppel and Lambhardt 1979; D'Amore 2015; McCurry *et al.*
351 2015). The gracile morphotype is most clearly observed in 2D in *V. giganteus*. The elongate cranial
352 shape likely reflects a diet consisting of large prey and carrion (McCurry *et al.* 2015). Second, there
353 are finer scale shape differences that are expected to be important in determining the strength of a
354 space frame construction. For example, a greater ventral projection of the palatopterygoid
355 articulation to form an obtuse angle, and a more vertical positioning of the epipterygoid increases
356 structural support, and is observed in semi-frugivorous species feeding like *V. olivaceous*
357 (Auffenberg, 1988). We also recover variation in angularity of the frontoparietal suture or
358 mesokinetic joint. Cranial sutures are forms of articulation in which the bones are rigidly joined by
359 fibrous tissue (synarthroses) (Di Ieva *et al.* 2013) and are expected to have important roles in cranial
360 kinesis and reducing stresses during biting feeding (Moazen *et al.* 2009). The concordance among
361 observation angles and links with diet suggests feeding performance imposes selective pressure

362 across each view, resulting in **high integration between the different views, and to certain degree**
363 among the cranial subunits described by each landmark configuration (Harmon *et al.* 2005; Perez,
364 Bernal & Gonzalez, 2006; Klingenberg, 2008; Meloro *et al.* 2011).

365

366 Our analysis of dorsal, ventral and lateral cranium shape in monitors leads to subtle differences in
367 interpretation of morphological diversification of the cranium as a whole. Given the dorsal
368 landmark configuration describes fused and paired bones, we expect to recover a more conserved
369 pattern of morphological diversification. Increased conservatism is evident through the strong
370 phylogenetic signal, the greatest size-shape constraint among views, and a more clear separation of
371 phylogenetic groups in phylomorphospace. The interplay of lateral and ventral views is apparent,
372 further supporting a role for some developmental constraints in morphological diversification of the
373 monitor cranium, but also highlighting the role of diet.

374

375 **Implications for 2D landmark configuration design**

376 Geometric morphometric methods have been widely applied in studies of head shape diversification
377 for numerous vertebrate groups (Slice, 2007). External head morphology is primarily landmarked
378 using the facial features in primates, and the scales of lizards, snakes and fish (Kaliontzopoulou
379 2011; Kerschbaumer & Sturmbauer, 2011; Baab, McNulty & Rohlf, 2012). The skull is frequently
380 broken down into its constituent parts (including dentition), depending on the question of interest,
381 and described by landmarks at the contacts between bones, tips of processes, origins and
382 attachments of muscles, locations of joints and tips of lever systems (Slice, 2007). Although
383 landmarks are frequently collected as surface marks on one tissue (i.e. scales or bone), they may
384 relate to different tissues whose relationships vary with evolution, development, and growth (Slice,
385 2007; Oxnard & O'Higgins, 2009). For example, 2D landmark configurations on the surface of
386 bone have been utilized in marmot skulls for all three cranial observation views, and the mandible.
387 The ventral cranium **seems to be** the best morphological feature for recovering phylogenetic

388 relationships in varanids, and it seems that the largest shape modifications occur in regions directly
389 involved in the mechanics of mastication in several other groups (Cardini & O'Higgins, 2004;
390 Caumul & Polly, 2004; Cardini & Thorington, 2006). The lateral cranium and mandible are least
391 strongly correlated with phylogeny, and dorsal and lateral cranium morphology together reflects
392 brain size and dermal bones (Caumul & Polly, 2004; Cardini, Hoffman & Thorington, 2005).

393

394 The efficiency of geometric morphometrics methods to provide a detailed and accurate quantitative
395 description of form ultimately increases with the amount of information contained within a
396 landmark configuration (Cardini & Thorington, 2006). Outline-based geometric morphometric
397 techniques are therefore a powerful addition to a 2D landmark configuration, as they can capture
398 structural information on large areas of morphology only represented by surfaces, curves or outlines
399 (Oxnard, 1978; Roth, 1993; Adams *et al.* 2004; Perez *et al.* 2006; Webster & Sheets, 2010). The
400 outline-based approach is less widely applied than the landmark-based approach, having mainly
401 been utilized in anthropological studies (Slice, 2007; Baab *et al.* 2012). However, three limitations
402 of outline-based methods should be considered: (1) protrusions on one individual that do not appear
403 on another can be poorly captured by the resulting semilandmark alignment; (2) it is often not clear
404 how many semilandmarks should be used for any given outline; and (3) outline methods only define
405 relative features (MacLeod, 1999; McCane, 2013; Finlay & Cooper, 2015). Regardless, their utility
406 is highlighted in studies like ours, where the clarity of results is considerably improved in the
407 landmark configuration(s) that utilize both landmarks and outlines (Baab *et al.* 2012; Finlay &
408 Cooper, 2015). By increasing the number of coordinates along a homologous contour or surface, it
409 is ultimately possible to obtain more information on the shape of a structure. The optimal 2D
410 observation view for analyzing cranial shape variation may thus be the view in which both
411 landmarks and semilandmarks may be utilized.

412

413 In conclusion, our study adds to a growing body of evidence suggesting multiple carefully designed
414 2D landmark configurations can provide fairly accurate descriptions of interspecific variation in
415 complex structures like the vertebrate head (Cardini & Thorington, 2006; Perez *et al.* 2009; Álvarez
416 & Perez, 2013; Cardini, 2014; Finlay & Cooper, 2015; Klingenberg, 2015). The key benefits of 3D
417 techniques may instead be in the extensions to **assessing morphological patterns in internal**
418 **structures (e.g. brain case)**, functional morphology (finite element modeling and mechanical strain)
419 and reconstruction of ancestral shapes (evolutionary warping or morphing) that they offer (Wiley *et*
420 *al.* 2007; Parr *et al.* 2012).

421

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618 **FIGURE CAPTIONS**

619 **Fig. 1.** Phylogenetic relationships of the 26 *Varanus* species included in this study. The tree is
620 primarily based on the phylogeny of Vidal *et al.* (2012), with additional information from Welton *et*
621 *al.* (2014) (see text for details).

622

623 **Fig. 2.** The three 2D landmark configurations used in this study to characterize cranial shape for (a)
624 dorsal, (b) ventral, and (c) lateral views. A scale bar equal to 10 mm is shown, and numbers refer to
625 the anatomical definitions of landmarks in Supp. Table S2. The example skull is *Varanus glauerti*
626 specimen [WAM_R77266].

627

628 **Fig. 3.** The two major principal axes of cranial shape variation in *Varanus*, visualized as
629 deformation grids, in dorsal, ventral and lateral views. PC axes are from a PCA of species means,
630 and shape changes associated with the PCs are shown as extreme cranial shapes representative of
631 the positive and negative ends of each axis. In each case the magnitude of the shape change from
632 the mean is indicated by the extent of deformation.

633

634 **Fig. 4.** Cranial shape diversification among *Varanus*, in (a) dorsal, (b) ventral, and (c) lateral views,
635 for original shape data. The phylogenetic tree is superimposed onto a plot of the first two PCs of the
636 covariance matrix based on residuals computed using the regression vector of independent
637 contrasts. The tips of terminal branches are at the locations of species means, and are coloured
638 according to phylogenetic group.

TABLES

Table 1. Cranial shape variation in the three observation views captured in PC 1-5, both for the original and size-corrected shape data. The tabled values are the percentages of total variance (%) for which each principal component accounts.

Axis	Original Shape Data			Size-corrected Shape Data		
	Dorsal	Ventral	Lateral	Dorsal	Ventral	Lateral
PC 1	54.939	47.742	31.077	15.242	21.246	14.443
PC 2	18.995	24.666	22.514	12.078	11.425	10.838
PC 3	6.361	7.426	13.142	9.283	8.190	8.719
PC 4	5.094	5.803	6.781	7.330	6.924	6.810
PC 5	3.745	3.174	5.734	6.319	6.367	6.510
Cumulative (PC1-5)	89.133	88.811	79.248	50.252	54.152	47.320

Table 2. Summary table of multivariate Procrustes regressions and phylogenetic regressions of shape data on centroid size, for each of the three views, using *geomorph* (Adams & Otárola-Castillo 2013).

	Dorsal		Ventral		Lateral	
	R ²	<i>p</i>	R ²	<i>p</i>	R ²	<i>p</i>
Regression	0.3052	<0.0001	0.1666	<0.0001	0.2294	<0.0001
Phylogenetic regression	0.2072	<0.0001	0.1121	<0.0001	0.1367	<0.0001

Table 3. Summary of Procrustes Variances, accounting for the morphological disparity of all the samples for the whole data set, grouped by species (dispersion calculated within species), and grouped by regions (dispersion within each geographical region). Procrustes Variances were calculated for both the original morphological GPA (Generalised Procrustes Analysis)-aligned morphological dataset, and the size-corrected one (accounting for allometry on the GPA-aligned dataset). For each analysis, we performed 1000 permutations, using the R-package *geomorph* (Adams & Otárola- Castillo, 2013).

Groups	Original shape data			Size-corrected shape data		
	Dorsal	Ventral	Lateral	Dorsal	Ventral	Lateral
All specimens	0.0054	0.0057	0.0068	0.0038	0.0048	0.0052
Species:						
<i>Varanus acanthurus</i>	0.0088	0.0062	0.0085	0.0036	0.0038	0.0041
<i>Varanus albigularis</i>	0.0080	0.0066	0.0120	0.0078	0.0066	0.0120
<i>Varanus bengalensis</i>	0.0053	0.0041	0.0069	0.0057	0.0039	0.0068
<i>Varanus brevicauda</i>	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001
<i>Varanus bushi</i>	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001
<i>Varanus caudolineatus</i>	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001
<i>Varanus dumerilii</i>	0.0066	0.0032	0.0056	0.0076	0.0031	0.0061
<i>Varanus eremius</i>	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001
<i>Varanus exanthematicus</i>	0.0085	0.0125	0.0097	0.0072	0.0122	0.0092
<i>Varanus giganteus</i>	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001
<i>Varanus gilleni</i>	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001
<i>Varanus glauerti</i>	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001
<i>Varanus gouldii</i>	0.0028	0.0039	0.0051	0.0023	0.0033	0.0052
<i>Varanus griseus</i>	0.0028	0.0034	0.0032	0.0027	0.0028	0.0026
<i>Varanus indicus</i>	0.0021	0.0020	0.0040	0.0017	0.0022	0.0050
<i>Varanus komodoensis</i>	0.0094	0.0098	0.0087	0.0016	0.0043	0.0036
<i>Varanus mertensi</i>	0.0014	<0.0001	<0.0001	0.0013	<0.0001	<0.0001
<i>Varanus mitchelli</i>	0.0024	0.0021	0.0058	0.0013	0.0011	0.0037
<i>Varanus niloticus</i>	0.0022	0.0035	0.0038	0.0025	0.0032	0.0033
<i>Varanus olivaceus</i>	0.0042	0.0030	0.0037	0.0053	0.0028	0.0038
<i>Varanus prasinus</i>	0.0041	0.0035	0.0047	0.0023	0.0036	0.0043
<i>Varanus rosenbergi</i>	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001
<i>Varanus rudicollis</i>	0.0183	0.0146	0.0150	0.0158	0.0145	0.0148
<i>Varanus salvadorii</i>	0.0055	0.0068	0.0096	0.0048	0.0075	0.0090
<i>Varanus salvator</i>	0.0048	0.0042	0.0054	0.0030	0.0049	0.0049
<i>Varanus scalaris</i>	0.0056	0.0036	0.0087	0.0018	0.0024	0.0040
<i>Varanus storri</i>	0.0127	<0.0001	<0.0001	0.0059	<0.0001	<0.0001
<i>Varanus tristis</i>	0.0041	0.0050	<0.0001	0.0031	0.0025	<0.0001
<i>Varanus varius</i>	0.0034	0.0027	0.0044	0.0045	0.0029	0.0045
Region:						
African	0.0043	0.0056	0.0065	0.0042	0.0053	0.0060
Indo-Asian A	0.0062	0.0055	0.0069	0.0055	0.0056	0.0066
Indo-Asian B	0.0038	0.0030	0.0041	0.0037	0.0029	0.0041
Indo-Australian A	0.0070	0.0076	0.0077	0.0029	0.0044	0.0046
Indo-Australian B	0.0029	0.0045	0.0053	0.0024	0.0040	0.0051
Indo-Australian C	0.0071	0.0078	0.0094	0.0032	0.0060	0.0049