

1 **A comparison of epibenthic reef communities settling on**
2 **commonly used experimental substrates: PVC versus**
3 **ceramic tiles.**

5 ^{1*}Mallela, J., ¹Milne, B.C., ¹Martinez-Escobar, D.

7 ¹Research School of Biology, Australian National University, Canberra, ACT 0200,
8 Australia

10 * Corresponding author email: j.a.mallela93@members.leeds.ac.uk

12 **Abstract:**

14 Artificial substrates are routinely used in coral reef research to model the recruitment
15 and growth responses of benthic organisms (e.g. coral recruitment and encrusting
16 organisms) to environmental change. Two commonly used, but structurally different,
17 artificial substrates include cylindrical PVC pipes and flat ceramic tiles. Various
18 ecosystem based models extrapolate data from these substrates interchangeably based
19 on the assumption that results are directly comparable. In order to test this
20 assumption we deployed these commonly used artificial substrate materials, PVC
21 poles and ceramic tiles, in shallow patch reefs for 34 months at One Tree Island,
22 Great Barrier Reef. Tiles were positioned to mimic upwards facing, well-lit substrates
23 (exposed), and downwards facing, shaded (cryptic) substrates. Multivariate analyses

1 demonstrated that the community composition differed significantly between all three
2 treatments. The majority of artificial substrate, coral reef experiments focus on key
3 groups of calcifying organisms, primarily: coralline algae, scleractinian coral and/or
4 total calcareous encruster cover. Interestingly, significant differences in the
5 recruitment, colonisation and community composition of these organisms were
6 detected for our three treatments. When compared to ceramic tiles, PVC poles had
7 greater coverage of crustose coralline algae but reduced levels of coral recruits
8 (<1mm diameter) and turf algae. We suggest that comparisons between studies that
9 utilise data from different substrate types should be used with caution. Additionally,
10 large scale modelling and forecasting exercises utilising these data sets should adjust
11 for the inherent biases of each method.

12
13 **Key words:** crustose coralline algae, encruster, turf, coral, recruitment, artificial
14 substrate

15 **1. Introduction:**

16
17 Real world, *in situ* data on the life histories, growth and development of different
18 coral reef organisms are critical if we are to be able to model reef development and
19 forecast how reefs in the future will respond to changing environmental conditions
20 (Stearn et al. 1977; Mallela and Perry 2007; Kennedy et al. 2013; Hepburn et al. 2015;
21 Jones et al. 2015). The deployment of artificial substrates on the reef, such as PVC
22 pipes and ceramic tiles, provide us with a valuable, potentially non-destructive tool to
23 collect quantitative reef growth data. In reef research, artificial substrates are
24 increasingly being used to assess reef development and assess the impacts of changing

1 environmental conditions (Kennedy et al. 2013; Mallela 2013). In particular, the use
2 of ceramic tiles to study coral settlement and life history traits has been widely
3 adopted (e.g. English et al. 1997; Mundy 2000). In contrast, studies focusing on the
4 growth and development of other epibenthic organisms, in particular calcareous reef
5 building organisms, are characterised by a diverse array of experimental materials
6 which include natural materials such as slices of dead coral (Harriott and Fisk 1987;
7 Klumpp 1992; Pari et al. 1998), and commonly available manmade products
8 including concrete, ceramic tiles, PVC poles, cattle ear tags and glassware (Adey and
9 Vassar 1975; Bak 1976; Martindale 1992; Field et al. 2007; Mallela 2007; Kuffner et
10 al. 2013; Hepburn et al. 2015; Roik et al. 2016). Unsurprisingly, experimental
11 substrates were often selected according to their affordability and local availability
12 (Field et al. 2007).

13
14 The diverse range of methods detailed in the literature highlighted a need for a
15 standardised method to be introduced for the analysis of encruster assemblages and
16 resulting coral reef carbonate budget models. Mallela in 2004 suggested a low impact
17 (non-destructive) carbonate budget method which incorporated the use of ceramic
18 tiles to assess encruster assemblages and their rates of carbonate production (Mallela
19 2004). This built on comprehensive methods conducted in Barbados in the 1970's
20 (Stearn et al. 1977) which also used settlement plate data. This low impact method
21 was successfully trialled and ground truthed in Jamaica (Mallela 2004,2007; Mallela
22 and Perry 2007). Subsequently, a rapid assessment method for assessing encruster
23 assemblages and coral reef carbonate budgets has been proposed using PVC poles.
24 This alternate method builds on pioneering work developed in Curacao (Bak 1976).
25 Known as ReefBudget, this method relies on multiple PVC poles being inserted into

1 the reef and subsequently lifted for analysis (e.g. Morgan and Kench 2014; Perry et al.
2 online resource). Interestingly, data resulting from these two artificial substrate
3 methods have never been directly compared. There has also been some debate about
4 whether or not experimental substrates provide real world information that can be
5 extrapolated to naturally occurring marine assemblages (Glasby and Connell 2001;
6 Perkol-Finkel et al. 2006; Mallela 2007; Burt et al. 2009). If we are to be able to
7 extrapolate from artificial substrate data in a meaningful manner we need to know
8 their caveats. This includes assessing if inter-substrate data sets are comparable, if
9 they simulate real-world (reef-scape) data, and if not, what their inherent biases are.

10

11 The effects of different artificial substrates on coral recruitment have been widely
12 documented (Harriott and Fisk 1987; Petersen et al. 2005; Burt et al. 2009; Miller et
13 al. 2009). Currently, we know less about their impacts on calcareous encrusting
14 organisms (*sensu* Taylor 1990) and the wider epibenthic reef community structure
15 (Field et al. 2007). Substrate orientation (Taylor 1990; Mallela 2007; Hepburn et al.
16 2015) and morphology (Martindale 1992) are known to influence encruster
17 settlement. For instance, nodular colonies show a preference for convex reef surfaces
18 whilst laminar colonies tend to colonise concave or planar surfaces (Martindale
19 1976,1992). Whilst many epibenthic organisms also display rugophilic behaviour
20 preferentially settling in cracks, crevices and shaded habitats on the reef (Taylor
21 1990) some deployment methods omit this important reef parameter (Adey and
22 Vassar 1975; Bak 1976; Nozawa et al. 2011). Caribbean reef comparisons of
23 experimental reef substrates deployed in different orientations over one to two years
24 also note clear differences in recruitment and community composition between
25 vertical and horizontal substrates, and well-lit and shaded substrates (Mallela 2013;

1 Hepburn et al. 2015). Interestingly, Mallela's (2013) fore-reef study in Tobago found
2 total encruster cover was greatest on exposed-horizontal substrates when compared to
3 vertical or horizontal low-light substrates. In contrast, Hepburn's 2015 Mexico study
4 across a range of reef sites, at various depths and orientations, observed both inter-site
5 variability and especially high encruster cover in cryptic/vertical habitats at some
6 locations. Such findings highlight, irrespective of substrate type, a degree of inter-site
7 variability occurring in the early stages (e.g. initial 24 months) of benthic recruitment
8 and encruster development (Burt et al. 2009).

9
10 Two of the most commonly deployed artificial substrates used in reef research to
11 study the growth and development of encrusters and epibenthic organisms include
12 PVC poles and ceramic tiles. Few comparisons of these substrates with natural reef
13 habitats exist (see Table 1 for a summary of these studies). Adey (Adey and Vassar
14 1975) found that when PVC poles were positioned on the reef to mimic dead
15 branching coral substrates coralline communities were faster to develop on the PVC
16 when compared to the natural dead substrate. Additionally, PVC poles positioned in
17 the reef pavement zone did not attract some of the less dominant coralline algae
18 species observed naturally in the shallow pavement zone. Studies in St Croix (Adey
19 and Vassar 1975) and Curacao (Bak 1976) also noted that PVC poles, including their
20 shaded undersides if orientated accordingly, did not attract certain sclerobionts (e.g.
21 encrusting forams, bryozoans, and specific coralline algae species) typical of
22 shaded/cryptic environments (e.g. the undersides of corals or cryptic pavement areas).
23 A comparison of settlement plates with naturally occurring reef substrates in
24 Barbados provided anecdotal evidence stating that no evidence was seen for a marked
25 difference in the crusts between natural and artificial substrates (Martindale 1976).

1 Whilst a study in Jamaica that compared assemblages on experimental, cryptically
2 orientated ceramic tiles with the undersides of adjacent platy corals, found no
3 significant difference with regards to community composition and benthic cover
4 (Mallela 2007). The Jamaican study concluded that cryptically orientated ceramic
5 tiles were a good proxy for naturally cryptic (e.g. shaded) reef substrates.

6
7 Carbonate budget models, which predict reef accretion, also utilise encruster growth
8 data from a variety of artificial substrates, typically PVC or ceramic settlement plates
9 are used to collect site specific data (Stearn et al. 1977; Mallela and Perry 2007;
10 Morgan and Kench 2014; Perry et al. online resource). In the absence of site specific,
11 *in situ* measurements additional data are often extrapolated from experiments in other
12 regions of the world using various methods and different experimental substrates (e.g.
13 Hart and Kench 2007; Kennedy et al. 2013). While direct comparisons are then made
14 between these different studies data are rarely cross checked or validated between
15 these different approaches. The effects of artificial substrate type, and the deployment
16 method used, on encruster and non-calcareous community development still remains
17 generally overlooked and poorly understood. In addition, the implications of
18 upscaling data sets gleaned from different methods to input into reef scale models
19 (e.g. carbonate budgets) has largely been ignored.

20
21 Based on a review of the literature, two of the most common methods used for
22 generating epibenthic data that is then extrapolated for use in reef accretion models
23 utilise data gleaned from two physically different, but readily available, artificial
24 substrates types: 1) flat, ceramic (terracotta) tiles, (also called settlement plates) and
25 2) hollow, white PVC poles. In order to determine if these two approaches are

1 comparable, the variation in community composition and the abundance (% cover) of
2 epibenthic communities (both calcareous and non-calcareous) recruiting to these
3 different, commonly used, artificial substrates were assessed. Horizontally orientated
4 PVC poles and ceramic tiles in a horizontal, downwards facing orientation (cryptic)
5 and ceramic tiles in a horizontal, upwards facing (exposed) orientation were compared
6 and apparent biases examined.

8 **2. Methods:**

9
10 **Study sites:** Artificial substrates were placed at three patch reef sites (microatolls) at
11 One Tree Island in the Southern Great Barrier Reef. These shallow microatolls form
12 part of an extensive patch reef system within the lagoon at One Tree Island.
13 Microatolls in this study were characterised by having a fully enclosed perimeter of
14 living reef composed primarily of coral and coralline algae. Inside the perimeter, the
15 microatolls were characterised by sand, rock, coralline algae and small coral colonies
16 (depth at low tide < 2m), the outer walls of the microatolls fall steeply to lagoon floor
17 (depth: 2-5 m adjacent to microatoll). At low tide the inner 'pond' of the microatolls
18 were isolated from the rest of the lagoon by their circular reef walls. At high tide the
19 water rises above the living walls (common name: piecrust) of the microatolls (≤ 1.5
20 m) enabling water exchange and free movement of reef organisms. These microatolls
21 have been described in great detail in earlier work and previously used site names will
22 be used here for continuity: microatoll 1) Kinsey (described in Kinsey and Domm
23 1974), microatoll 2) ENCORE 4, and microatoll 3) ENCORE 11 (ENCORE sites
24 described in Larkum and Steven 1994; Steven and Atkinson 2003).

1

2 **Experiment design:** Two common artificial substrate types were used in this

3 experiment: 1) unglazed, square, ceramic tiles (13 x 13 cm) and standard, white PVC
4 poles (smooth surface, hollow, 50 cm long, 7cm in circumference). In order to remove
5 the confounding effects of orientation (e.g. horizontal versus vertical substrate effects
6 (e.g. Glasby and Connell 2001; Mallela 2013; Hepburn et al. 2015) all substrates were
7 positioned horizontally and randomly, without touching, inside the three microatolls
8 at depths of 1 - 2 m at low tide. All artificial substrates were positioned to reflect the
9 natural gradient of the lagoon floor (angle of slope) on which it had been placed,
10 typically 0° to 10° in slope. All substrates were secured to PVC frames attached to the
11 floor of the microatolls using cable ties threaded through pre-drilled holes in the
12 artificial substrates. Ceramic tiles were attached horizontally in close fitting, overlying
13 pairs leaving only the two outer faces available to recruitment and subsequent
14 colonisation. Each pair of ceramic tiles had one upwards facing, well-lit surface, from
15 here-in referred to as “exposed” (TE) and one downwards facing, shaded, surface
16 from herein referred to as “cryptic” (TC). PVC substrates were also positioned
17 horizontally with the circumference (outer surface) of the PVC tube available for
18 subsequent colonisation.

19

20 Substrates remained *in situ* for a period of almost three years in order to be
21 representative of established (multi-year) encruster assemblages. All substrates were
22 deployed in May 2012 and lifted in March 2015 after a total underwater deployment
23 of 34 months. The number of intact replicates analysed at the end of the experiment
24 were: Tiles-Exposed: 30; Tiles-Cryptic: 31; PVC poles: 30.

25

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

1 On collection, artificial substrates were labelled and air dried. A 100 point grid was
2 superimposed over each substrate (TE, TC and PVC), each data point was ≥ 1 cm
3 apart. For the square tiles we used a square 10 x 10 grid, for the PVC tubes we used a
4 rectangular 50 x 2 grid. Using a dissection microscope all organisms under each point
5 were identified to taxonomic group (e.g. crustose coralline algae, coral recruit,
6 calcareous worm, turf) and counted (Mallela 2013), see supplementary Table 1 which
7 details all identification categories.

8

9 **Statistical analysis:** Community data were analysed using PRIMER 6 statistical
10 software (Clarke and Gorley 2006). Multivariate analyses were used to test for
11 differences in community composition between the treatments. Multivariate data were
12 square root transformed and the Bray-Curtis similarity coefficient employed to
13 construct a similarity matrix for the percentage cover (%) of colonising epibenthic
14 communities. Non-metric multidimensional scaling (MDS) ordinations were used to
15 assess assemblages between microatolls and across different substrate types. No
16 significant microatoll (reef site) impact was found (supplementary Fig. 1) so data sets
17 were pooled in order to further investigate substrate impacts. One-way analyses of
18 similarities (ANOSIM) tests were used to look for differences in epibenthic
19 communities between exposed tiles, cryptic tiles and PVC. The R-statistic indicated
20 the extent of significant differences, R-statistic values <0.1 were considered negligible
21 (Clarke 1993). If ANOSIM indicated a significant difference between substrate types
22 ($R>0.1$), Similarity Percentages Analyses (SIMPER), using 1-way analysis on Bray-
23 Curtis similarities for substrate groups using a 90% dissimilarity threshold, was used
24 to indicate which epibenthic groups were responsible for these observed differences.

25

1 Key epibenthic groups of interest to the wider reef research community using PVC
2 and settlement plate artificial substrates methods were identified during our literature
3 review. Subsequent analysis focused on these key groups: total epibenthic cover,
4 coralline algae, hard (scleractinian) coral cover and total calcareous encruster cover.
5 The abundance, and substrate preferences, of these groups were further explored
6 using IBM-SPSS 22 statistical software. Normality of distribution and homogeneity
7 of variance were tested using Kolmogorov-Smirnov and Levene's test, respectively.
8 As a significant microatoll effect was not found to influence the community
9 composition of these key epibenthic groups (see supplementary Fig. 1 MDS plot of
10 microatoll community composition and supplementary table 2 detailing Scheirer-Ray-
11 Hare test results) data from the three micro-atoll sites were pooled. To test for
12 differences among the three substrate types the Kruskal-Wallis (KW) test was used
13 due to non-normal data distributions. If the KW test revealed a significant difference
14 between the three substrate types, the Mann-Whitney *U* Test was subsequently used
15 for pairwise comparisons to assess which pair was responsible for the difference. This
16 approach has the same logic as an ANOVA *posthoc* LSD test if it is only applied
17 when the KW test reveals a significant result (Dytham 2003). To account for multiple
18 comparisons a Bonferroni correction of alpha was applied where $p \leq 0.01$ was
19 considered significant. Data transformations were not required to meet the
20 assumptions of these tests.

21 **3. Results:**

22 Multivariate analyses were used to compare the community composition of epibenthic
23 assemblages colonising the different artificial substrate types after 34 months of
24

1 deployment. MDS ordinations gave a good representation of community assemblage
2 (2D Stress: 0.08) and depicted differences in epibenthic assemblages between the
3 three test substrates (Fig. 1). The one-way ANOSIM test comparing benthic
4 composition between artificial substrates indicated a significant difference: ANOSIM,
5 Global R of 0.457 ($p = 0.001$). ANOSIM Pairwise Tests also indicated differences
6 between pairs of substrate types: exposed tile v PVC: $R = 0.5$ ($p < 0.001$), cryptic tiles
7 v PVC: $R = 0.6$ ($p < 0.001$), and exposed v cryptic tiles: $R = 0.3$ ($p < 0.001$). One-way
8 Similarity Percentages (SIMPER) indicated that the categories primarily responsible
9 for these differences were: turf, total non-calcareous cover, total calcareous cover,
10 crustose coralline algae and uncolonised (bare) substrate (see supplementary material
11 table 3).

12
13 Total epibenthic cover by all organisms was significantly different between substrates
14 (Kruskall-Wallis: $H = 47.6$, $df = 2$, $p = 0.000$) with PVC having significantly greater
15 cover compared to exposed tiles (Mann-Whitney U test: PVC median = 94%, exposed
16 median = 71%, $U = 38$, $p = 0.000$), Fig. 2a. No difference was observed between PVC
17 and cryptic tiles, whilst cryptic tiles had significantly higher epibenthic cover than
18 exposed tiles (Mann-Whitney U test: cryptic median = 94%, exposed median = 71%,
19 $U = 62.5$, $p = 0.000$), Fig. 2 a. PVC poles displayed 29 and 1 % more total epibenthic
20 cover than exposed and cryptic tiles respectively. Whilst mean total calcareous cover
21 (%) was also 39 and 27 % greater on PVC compared to exposed and cryptic tiles
22 respectively (supplementary material table 1).

23
24 The percentage cover of crustose coralline algae (CCA) was significantly different
25 between the three substrates (Kruskall-Wallis: $H = 51.9$, $df = 2$, $p = 0.000$). CCA

1 cover was significantly different when PVC was compared with cryptic (Mann-
2 Whitney U test: PVC median = 85.5 %, cryptic median = 61 %, $U = 67.5$, $p = 0.000$)
3 and exposed substrates (Mann-Whitney U test: PVC median = 85.5% %, exposed
4 median = 55%, $U = 13.0$, $p = 0.000$), Fig. 2 b. On average, PVC poles displayed 37 &
5 26 % more CCA than exposed and cryptic tiles respectively (supplementary material
6 table 1).

7
8 The percentage cover of coral recruits (scleractinian corals < 1cm in diameter) was
9 also found to be significantly different between substrates types due to coral recruits
10 only being observed on grids superimposed over cryptic settlement plates (Kruskall-
11 Wallis: $H = 8.0$, $df = 2$, $p = 0.018$). Due to the low numbers of coral colonies > 1cm in
12 diameter being recorded, no significant differences were observed for coral colonies
13 (scleractinian coral > 1cm in diameter) or total coral cover (recruits + colonies), Fig. 2
14 c.

15
16 PVC poles had significantly more calcareous cover than both cryptic and exposed
17 tiles, (Kruskall-Wallis: $H = 54.512$, $df = 2$, $p = 0.000$), Fig. 2d. The percentage cover
18 of all combined non-calcareous organisms (e.g. sponges, turf, macro algae, ascidians)
19 was not significantly different between substrates. However, the cover of turf algae
20 (defined as algal assemblages < 1mm in height) was significantly different (Kruskall-
21 Wallis: $H = 49.2$, $df = 2$, $p = 0.000$) as was the proportion of non-colonised substrate
22 (Kruskall-Wallis: $H = 45.9$, $df = 2$, $p = 0.000$). PVC substrates had significantly less
23 turf colonising them when compared to cryptic and exposed settlement plates. Levels
24 of bare substrate were similar between PVC and cryptic plates, whilst exposed plates

1 displayed significantly elevated levels of bare substrate. See supplementary Tables 1
2 for full data set.

3

4 **4. Discussion:**

5 Our multi-year data set demonstrates that two of the most commonly used
6 experimental substrates, unglazed ceramic tiles (both cryptic and exposed) and PVC
7 poles, are colonised by different epibenthic assemblages. Consequently, results from
8 studies using different substrates are unlikely to be directly comparable. In particular,
9 we found that recruitment and growth by sclerobionts, which include key calcareous
10 reef building organisms (e.g. scleractinian coral recruits and coralline algae), were
11 significantly different between substrate types. PVC substrates were characterised by
12 greater coverage of photophilic, encrusting, coralline algae, but unlike cryptic tiles,
13 coral recruits were not observed on their 100 point grids. In contrast, settlement plates
14 were characterised by higher turf algae cover. We therefore suggest the choice of
15 artificial substrate and method (e.g. orientation) of deployment be driven by the
16 research aim and suggest caution when selecting and extrapolating data sets for real
17 world models (e.g. predicting ecosystem growth and development).

18

19 The use of ceramic (e.g. terracotta) settlement plates for scleractinian coral
20 recruitment studies have been widely endorsed (e.g. English et al. 1997; Burt et al.
21 2009; Mallela and Crabbe 2009; Humanes and Bastidas 2015). Our results found that
22 in the shallow patch reefs of One Tree Island coral recruits were primarily attracted to
23 cryptically orientated ceramic tiles in contrast to PVC poles. Other studies assessing
24 the recruitment patterns of scleractinian corals to different substrate types (e.g. petri

1 dishes, sliced coral skeletons and ceramic tiles) also note substrate specific results and
2 suggest ceramic tiles as the best substrate for coral recruitment studies based on the
3 premise that they attract the most coral recruits (Harriott and Fisk 1987). Possibly
4 due to their initial small size (e.g. <1mm) and preference for cryptic habitats, which
5 makes *in situ*, underwater observations difficult, we were unable to find any studies
6 that validate this widely accepted approach with naturally occurring, reef substrate
7 data.

8
9 Research focusing on other key reef building organisms such crustose coralline algae
10 and other calcareous encrusters also uses a wide range of readily available substrates
11 including PVC poles, ceramic tiles, cattle ear tags and glass to assess reef growth and
12 calcification questions (Bak 1976; Field et al. 2007; Mallela 2007; Kuffner et al.
13 2013; Mallela 2013; Hepburn et al. 2015; Roik et al. 2015). Whilst many of these
14 studies claim to measure the ‘natural range and variability’ of growth and calcification
15 there is very little literature available to validate or ground-truth these findings. The
16 few validation studies and observations we found were based on Caribbean reefs.

17 Early studies using experimental PVC substrates on reefs to investigate the growth of
18 encrusters noted that PVC, if deployed vertically on the reef surface, or at angles
19 mimicking dead branching coral (*Acropora palmata*), favoured recruitment by
20 crustose coralline algae (Adey and Vassar 1975; Bak 1976). Adey and Vassar (1975)
21 observed how coralline overgrowth occurred more slowly on naturally occurring dead
22 coral branches, when compared to PVC. This was attributed to the more uniform PVC
23 surface being more conducive to coralline settling while not initially providing a good
24 holding surface for mobile reef organisms (e.g. crab and worms), organisms that
25 presumably could hinder early settlement and growth. Both studies (Adey and Vassar

1 1975; Bak 1976) noted how PVC substrates were devoid of shaded (cryptic) reef
2 elements, in particular crustose coralline algae species (e.g. *Neogoniolithon*
3 *accretum*), encrusting foraminifera and bryozoa. Organisms commonly occurring on
4 the cryptic, dead bases of reef building coral colonies (*Montastrea annularis*),
5 accounted for 12 and 8 % of encruster cover respectively but were not observed on
6 vertical PVC (Bak 1976). Another 12 month study at 10m on Jamaican fore-reefs
7 ground truthed findings and found that cryptically orientated, unglazed, ceramic tiles
8 had similar encruster assemblages when compared to the underside of adjacent platy
9 corals (Mallela 2007). With cryptic (shaded) habitats on the reef estimated to account
10 for up to two-thirds of the reef volume and 75% of total available reef space (Jackson
11 et al. 1971; de Goeij and Van Duyl 2007) our results suggest that studies that rely on
12 growth and calcification data sourced only from the outside surface of PVC poles
13 overlook epibenthic communities that are characteristic of cryptic reef habitats (e.g.
14 shaded overhangs and crevices) and indeed can make up the greater portion of the reef
15 (Buss and Jackson 1979; Gischler and Ginsburg 1996). Such findings could result in
16 misleading or biased reef growth interpretations.

17
18 Many experiments that utilise artificial substrates only deploy the substrates for a
19 short period of time (e.g. < 1 year). As a result whilst providing data on settlement
20 and initial growth rates they may not provide data on established or mature
21 communities which are indicative of a large portion of *in situ* reef growth.
22 Observations in St Croix, Caribbean (Adey and Vassar 1975) using PVC substrates at
23 shallow depths (< 3m) noted that a one year deployment period is probably suitable to
24 reach a climax state on substrates positioned on exposed algal ridges. In contrast, in
25 shallow, cryptic habitats (e.g. 1-2 m reef pavement) several years growth on PVC may

1 be required to reach a climax community. We suggest that studies should also
2 consider what stage of growth and development they are measuring and note this in
3 their comparisons, data extrapolations and interpretations.

4
5 Our findings add to a growing number of studies that demonstrate how encruster
6 recruitment and benthic cover varies according to substrate type. However, another
7 potential source of error in reef scape accretion models is when models upscale
8 percentage cover data into reef accretion data (also known as calcification or calcium
9 carbonate production). In order to determine how much calcium carbonate (CaCO_3) is
10 being added to the reef the percentage cover of calcifying organisms is combined with
11 organism specific calcification rates (linear extension and skeletal density) in order to
12 calculate rates of CaCO_3 production $\text{g m}^{-2} \text{y}^{-1}$ (Stearn et al. 1977; Hubbard et al. 1990;
13 Mallela and Perry 2007). Due to the paucity of site specific growth rate data a
14 number of encruster carbonate production studies and reef accretion models
15 extrapolate growth rate and skeletal density data from other studies in order to
16 estimate carbonate production (e.g. Pari et al. 1998; Hart and Kench 2007). By
17 combining percentage cover data gleaned from natural or artificial substrates with
18 encruster growth rate data extrapolated from other locations another level of error is
19 potentially introduced into reef scape accretion models (Mallela 2013).

20
21 Results from this shallow microatoll study demonstrate that PVC poles and ceramic
22 tiles (exposed and cryptic orientations) are characterised by different suites of
23 epibenthic assemblages. Unfortunately, we were unable to validate our findings with
24 data from natural reef settings. Our analysis of epibenthic reef communities on
25 artificial substrates was fine-scale and required the use of a dissection microscope for

1 the identification of small organisms (<1mm in diameter). We were unable to get this
2 level of resolution from underwater, *in situ* observation during this study. Clearly if
3 we are to extrapolate and upscale data from artificial substrates to answer ecosystem-
4 development questions we need to know if our data is comparable to natural reef
5 substrates. Ground-truthing of data in order to calibrate data sets obtained using
6 different approaches and enable the extrapolation of inter-substrate data in a
7 meaningful manner will be our next challenge.

8

9 In conclusion, the results presented here add to small, but growing body of settlement
10 plate literature, indicating that settlement and growth of marine epibenthic
11 communities on different artificial substrates may not be directly comparable (Harriott
12 and Fisk 1987; Field et al. 2007). Key to these findings are that a specific substrate
13 type, positioned in a certain orientation, may bias, promote or deter recruitment of
14 specific organisms (see summary table 1). We know for example, that the outside of
15 PVC poles and upwards facing ceramic tiles attached to the surface of the reef are
16 unlikely to have epibenthic assemblages characteristic of cryptic reef habitats (e.g.
17 foraminifera and bryozoan that overgrow the dead bases and undersides of coral
18 colonies (Bak 1976; Mallela 2007). In spite of this, and due to a paucity of site
19 specific information, such data sets are being extrapolated for use in reef development
20 models without always noting their limitations. We suggests that data sets should be
21 interpreted and extrapolated with care and any caveats made clear. The choice of
22 substrate clearly depends on the research question. If the aim of the study is to use a
23 substrate that promotes recruitment, growth and subsequent survival of your target
24 organism, the ‘more is better’ approach, then this and other studies suggest PVC poles
25 are ideal for initiating and propagating crustose coralline algae communities whilst

1 ceramic tiles facilitate coral recruitment. However, without validation and ground
 2 truthing, such findings may be misleading if they are subsequently used as proxy for
 3 naturally occurring reef habitats and used to assess reef growth and/or carbonate
 4 accretion. Clearly artificial substrates are a useful, non-destructive and affordable tool
 5 in coral reef research. However, if they are to be used to model ecosystem
 6 development (e.g. sclerobiont growth and calcification) the choice of substrate and its
 7 orientation needs to be justified, methods should also be validated, and limitations
 8 noted.

9 **Acknowledgements:**

10 JM was funded by an Australian Research Council (ARC) Discovery Early Career
 11 Researcher Award (DECRA). Fieldwork permission: All work was undertaken with
 12 the permission of the Great Barrier Reef Marine Park Authority (GBRMPA permit
 13 number G12.35021.1) and One Tree Island Research Station. A special thanks for
 14 help in the field at One Tree Island: Adam Leavesley, Chris Bloomfield, Christine
 15 Schoenberg and Rebecca Fox.

17 **References:**

- 18
 19 Adey W, Vassar JM (1975) Colonization, succession and growth rates of tropical crustose coralline
 20 algae (Rhodophyta, Cryptonemiales)*. *Phycologia* 14:55-69
 21 Bak RPM (1976) The growth of coral colonies and the importance of crustose coralline algae and
 22 burrowing sponges in relation with carbonate accumulation. *Netherlands Journal of Sea
 23 Research* 10:285-337
 24 Burt J, Bartholomew A, Bauman A, Saif A, Sale PF (2009) Coral recruitment and early benthic
 25 community development on several materials used in the construction of artificial reefs and
 26 breakwaters. *Journal of Experimental Marine Biology and Ecology* 373:72-78
 27 Buss LW, Jackson JBC (1979) Competitive networks: nontransitive competitive relationships in
 28 cryptic coral reef environments. *American Naturalist* 113:223-234
 29 Clarke KR (1993) Non-parametric multivariate analyses of changes in community structure. *Australian
 30 Journal of Ecology* 18:117-143
 31 Clarke KR, Gorley RN (2006) PRIMER v6: User Manual/Tutorial. PRIMER-E: Plymouth
 32 de Goeij JM, Van Duyl FC (2007) Coral cavities are sinks of dissolved organic carbon (DOC).
 33 *Limnology and Oceanography* 52:2608-2617
 34 Dytham C (2003) *Choosing and using statistics - a biologist's guide*. Blackwell science, Oxford
 35 English S, Wilkinson C, Baker V (1997) *Survey manual for tropical marine resources*, 2nd edition.
 36 Australian Institute of Marine Sciences, Townsville, Australia

- 1 Field SN, Glassom D, Bythell J (2007) Effects of artificial settlement plate materials and methods of
2 deployment on the sessile epibenthic community development in a tropical environment.
3 Coral Reefs 26:279-289
- 4 Gischler E, Ginsburg N (1996) Cavity dwellers (coelobites) under coral rubble in Southern Belize
5 barrier and atoll reefs. Bulletin of Marine Science 58:570-589
- 6 Glasby TM, Connell JH (2001) Orientation and position of substrata have large effects on epibiotic
7 assemblages. Marine Ecology Progress Series 214:127-135
- 8 Harriott VJ, Fisk DA (1987) A comparison of settlement plate types for experiments on the recruitment
9 of scleractinian corals. Mar Ecol Prog Ser 37:201-208
- 10 Hart DE, Kench PS (2007) Carbonate production of an emergent reef platform, Warraber Island, Torres
11 Strait, Australia. Coral Reefs 26:53-68
- 12 Hepburn LJ, Blanchon P, Murphy G, Cousins L, Perry CT (2015) Community structure and
13 palaeoecological implications of calcareous encrusters on artificial substrates across a
14 Mexican Caribbean reef. Coral Reefs 34:189-200
- 15 Hubbard DK, Miller AI, Scaturo D (1990) Production and cycling of calcium carbonate in a shelf-edge
16 reef system (St. Croix, U.S. Virgin Islands): applications to the nature of reef systems in the
17 fossil record. Journal of Sedimentary Petrology 60:335-360
- 18 Humanes A, Bastidas C (2015) In situ settlement rates and early survivorship of hard corals: a good
19 year for a Caribbean reef. Marine Ecology Progress Series 539:139
- 20 Jackson JBC, Goreau TF, Hartman WD (1971) Recent Brachiopod-Coralline Sponge Communities and
21 Their Paleocological Significance. Science 173:623-625
- 22 Jones NS, Ridgwell A, Hendy EJ (2015) Evaluation of coral reef carbonate production models at a
23 global scale. Biogeosciences 12:1339-1356
- 24 Kennedy Emma V, Perry Chris T, Halloran Paul R, Iglesias-Prieto R, Schönberg Christine HL,
25 Wisshak M, Form Armin U, Carricart-Ganivet Juan P, Fine M, Eakin CM, Mumby Peter J
26 (2013) Avoiding Coral Reef Functional Collapse Requires Local and Global Action. Current
27 Biology 23:912-918
- 28 Kinsey DW, Domm A (1974) Effects of fertilization on a coral reef environment- primary production
29 studies. Proceedings of the Second International Symposium on Coral Reefs 1
- 30 Klumpp DW (1992) Community structure, biomass and productivity of epilithic algal communities on
31 the Great barrier Reef: dynamics at different spatial scales. Marine Ecology Progress Series
32 86:77-89
- 33 Kuffner IB, Hickey TD, Morrison JM (2013) Calcification rates of the massive coral *Siderastrea*
34 *siderea* and crustose coralline algae along the Florida Keys (USA) outer-reef tract. Coral Reefs
35 32:987-997
- 36 Larkum AWD, Steven ADL (1994) ENCORE: the effect of nutrient enrichment on coral reefs. 1.
37 Experimental design and research programme. Marine Pollution Bulletin 29:112-120
- 38 Mallela J (2004) Coral reef communities and carbonate production in a fluvially-influenced
39 embayment, Rio Bueno, Jamaica. PhD Ph.D. thesis, Manchester Metropolitan University, UK,
40 p224
- 41 Mallela J (2007) Coral reef encruster communities and carbonate production in cryptic and exposed
42 coral reef habitats along a gradient of terrestrial disturbance. Coral Reefs 26:775-785
- 43 Mallela J (2013) Calcification by Reef-Building Sclerobionts. PLOS ONE 8:e60010
- 44 Mallela J, Perry CT (2007) Calcium carbonate budgets for two coral reefs affected by different
45 terrestrial runoff regimes, Rio Bueno, Jamaica. Coral Reefs 26:53-68
- 46 Mallela J, Crabbe MJC (2009) Hurricanes and coral bleaching linked to changes in coral recruitment in
47 Tobago. Marine Environmental Research 68:158-162
- 48 Martindale W (1976) Calcareous encrusting organisms of the recent and pleistocene reefs of Barbados,
49 West Indies. Ph.D. Ph.D. thesis, The University of Edinburgh, p156
- 50 Martindale W (1992) Calcified epibionts as palaeoecological tools: examples from the recent and
51 Pleistocene reefs of Barbados. Coral Reefs 11:167-177
- 52 Miller MW, Valdivia A, Kramer KL, Mason B, Williams DE, Johnston L (2009) Alternate benthic
53 assemblages on reef restoration structures and cascading effects on coral settlement. Marine
54 Ecology Progress Series 387:147-156
- 55 Morgan KM, Kench PS (2014) Carbonate production rates of encruster communities on a lagoonal
56 patch reef: Vabbinfaru reef platform, Maldives. Marine and Freshwater Research 65:720-726
- 57 Mundy C, N. (2000) An appraisal of methods used in coral recruitment studies. Coral Reefs 19:124-
58 131

- 1 Nozawa Y, Tanaka K, Reimer JD (2011) Reconsideration of the surface structure of settlement plates
 2 used in coral recruitment studies. *Zoological Studies* 50:53-60
- 3 Pari N, Peyrot-Clausade M, Le Campion-Alsumard T, Fontaine M, Hutchings P, A., Chazottes V,
 4 Golubic S, Le Campion J, Fontaine M (1998) Bioerosion of experimental substrates on high
 5 islands and on atoll lagoons (French Polynesia) after two years of exposure. *Marine Ecology*
 6 *Progress Series* 166:119-130
- 7 Perkol-Finkel S, Shashar N, Benayahu Y (2006) Can artificial reefs mimic natural reef communities?
 8 The roles of structural features and age. *Marine Environmental Research* 61:121-135
- 9 Perry CT, Murphy G, Edinger E, Kench P, Mumby PJ, Smithers SG, Steneck RS (online resource)
 10 ReefBudget:
 11 [http://geography.exeter.ac.uk/media/universityofexeter/schoolofgeography/reefbudget/docume](http://geography.exeter.ac.uk/media/universityofexeter/schoolofgeography/reefbudget/documents/ReefBudget_Methodology.pdf)
 12 [nts/ReefBudget_Methodology.pdf](http://geography.exeter.ac.uk/media/universityofexeter/schoolofgeography/reefbudget/documents/ReefBudget_Methodology.pdf)
- 13 Petersen D, Laterveer M, Schuhmacher H (2005) Innovative substrate tiles to spatially control larval
 14 settlement in coral culture. *Marine Biology* 146:937-942
- 15 Roik A, Roder C, Röthig T, Voolstra CR (2015) Spatial and seasonal reef calcification in corals and
 16 calcareous crusts in the central Red Sea. *Coral Reefs*:1-13
- 17 Roik A, Roder C, Röthig T, Voolstra CR (2016) Spatial and seasonal reef calcification in corals and
 18 calcareous crusts in the central Red Sea. *Coral Reefs* 35:681-693
- 19 Stearn CW, Scoffin TP, Martindale W (1977) Calcium carbonate budget of a fringing reef on the west
 20 coast of Barbados. Part 1: Zonation and productivity. *Bulletin of Marine Science* 27:479-510
- 21 Steven ADL, Atkinson MJ (2003) Nutrient uptake by coral-reef microatolls. *Coral Reefs* 22:197-204
- 22 Taylor PD (1990) Encrusters. In: Briggs DEG, & Crowther, P. R. (ed) *Palaeobiology*. Blackwell
 23 Scientific Publications, Boston, pp346-351
- 24
- 25

26 **Figure Legends:**

27

28 **Fig. 1** Multidimensional scaling (MDS) ordination of Bray-Curtis similarities between
 29 epibenthic organisms recruiting to three artificial substrate types: cryptic tiles (C),
 30 exposed (E) tiles and PVC. The MDS is based on square root transformed benthic
 31 cover (%) data. The 2D stress value indicates that the plot is a good representation of
 32 multidimensional community similarity.

33 **Fig. 2** Box-whisker plots detailing total percentage cover on cryptic ceramic tiles
 34 (TC), exposed ceramic tiles (TE) and PVC poles. The thick bar represents the median
 35 value, the interquartile range is represented by the box, and the full range as the
 36 whiskers, raw values given in Supplementary table 1. *Significant differences noted
 37 on graphs, to account for multiple comparisons a Bonferroni correction of alpha was
 38 applied where $p \leq 0.01$ was considered significant.

1 **Table 1.** A summary of studies that ground truth data on artificial substrates (PVC
2 and ceramic tiles) with natural, in situ reef substrates
3
4
5

Artificial Substrate	Compared to natural reefs substrate	Findings	Location	Reference
PVC poles mimicking dead coral branches	Dead branching coral substrates	Coralline communities develop faster on PVC	St Croix	Adey & Vassar 1975
PVC poles in reef pavement	Reef pavement	PVC did not attract less dominant coralline algae species	St Croix	Adey & Vassar 1975
PVC poles attached to reef	Cryptic reef areas (e.g. undersides of corals/shaded locations)	PVC did not attract certain sclerobionts typical of cryptic reef substrates	Curacao	Adey & Vassar 1975, Bak 1976
Settlement plates	Natural reef substrate	Anecdotal evidence: no difference in crusts observed	Barbados	Martindale 1976
Cryptic settlement plates on reef	The underside of adjacent platy corals	No significant difference in community composition or benthic cover	Jamaica	Mallela 2007

31
32
33

34 **Supplementary Material:**

35
36
37 **Supplementary Fig. 1** Multidimensional scaling (MDS) ordinations of matrices
38
39 constructed from percentage cover of all benthic organisms recruiting to artificial
40
41 substrates: cryptic tiles (C), exposed tiles (e) and PVC at each microatoll site (E11, E4
42
43 and K). Multivariate analyses does not find a site affect.
44
45
46
47
48

49 **Supplementary Table 1.** Descriptive statistics detailing epibenthic percentage cover
50
51 (%) of key groups of benthic cover (raw data available on request).
52
53
54

55
56 **Supplementary Table 2.** Scheirer-Ray-Hare (SRH) test results
57
58
59
60
61
62
63
64
65

1 **Supplementary Table 3.** SIMPER analysis for comparison between epibenthic
2 communities recruiting to three artificial substrate types: exposed tiles (e), cryptic
3 tiles (c) and PVC poles. Analyses based on Bray-Curtis similarities, calculated from
4 square-root transformed benthic cover data and the benthic categories listed in
5 supplementary table 1. The low contributions cut-off was 90%. Abbreviations:
6 Av.Diss: average Bray–Curtis dissimilarity between substrates; Diss/SD: average
7 dissimilarity divided by its standard deviation; Contrib%: percentage contribution to
8 average dissimilarity; Cum.%: cumulative percentage contribution to dissimilarity.

9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

Figure 1

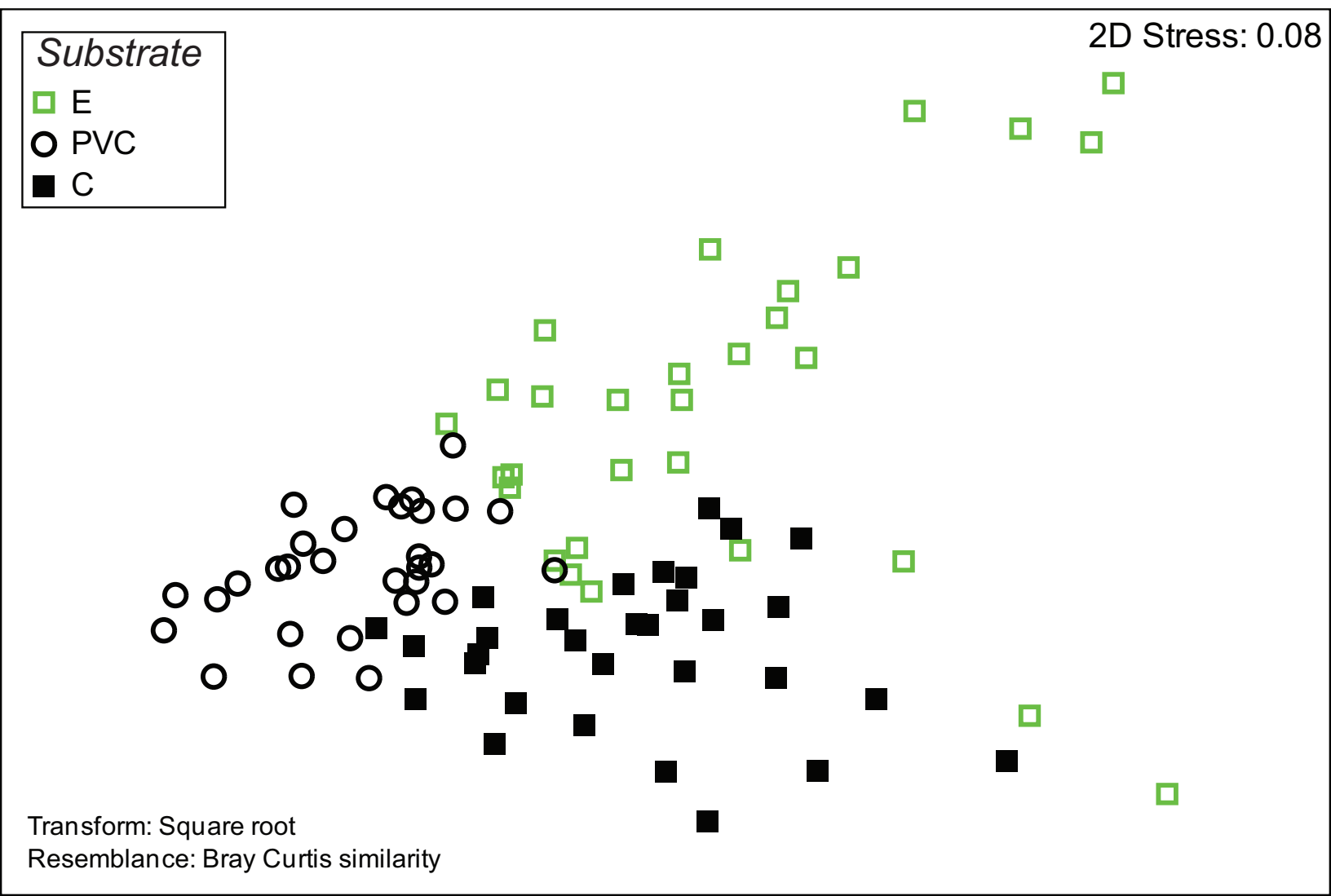
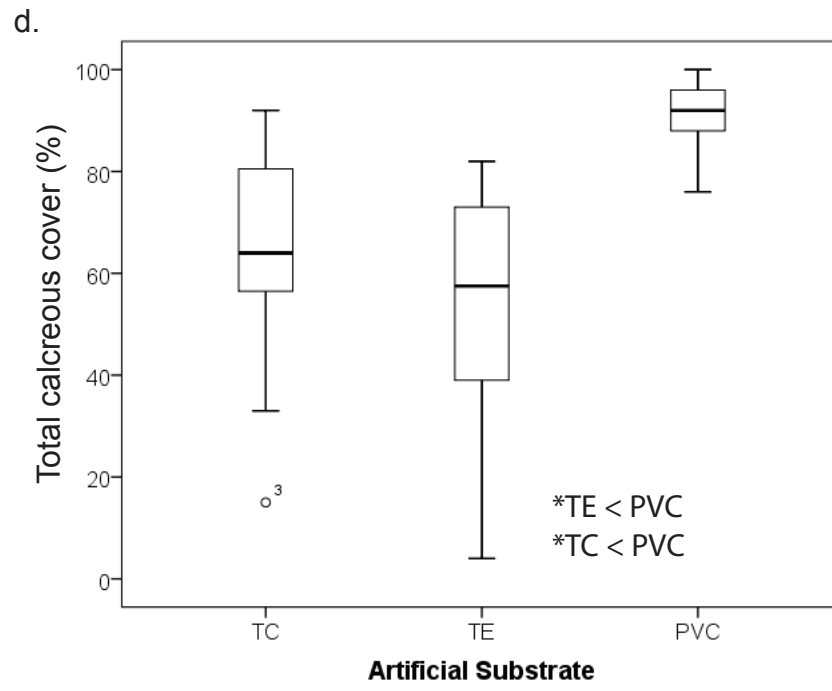
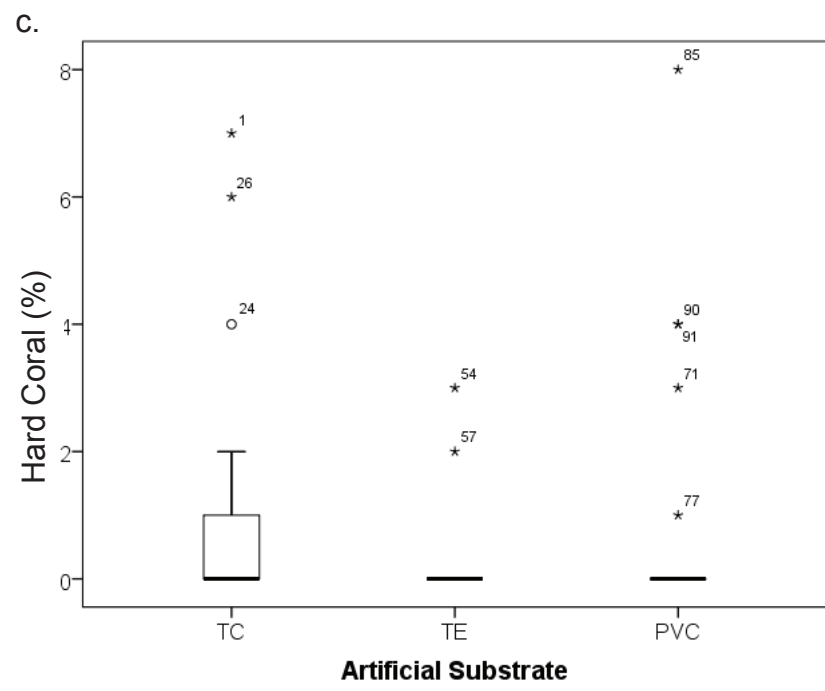
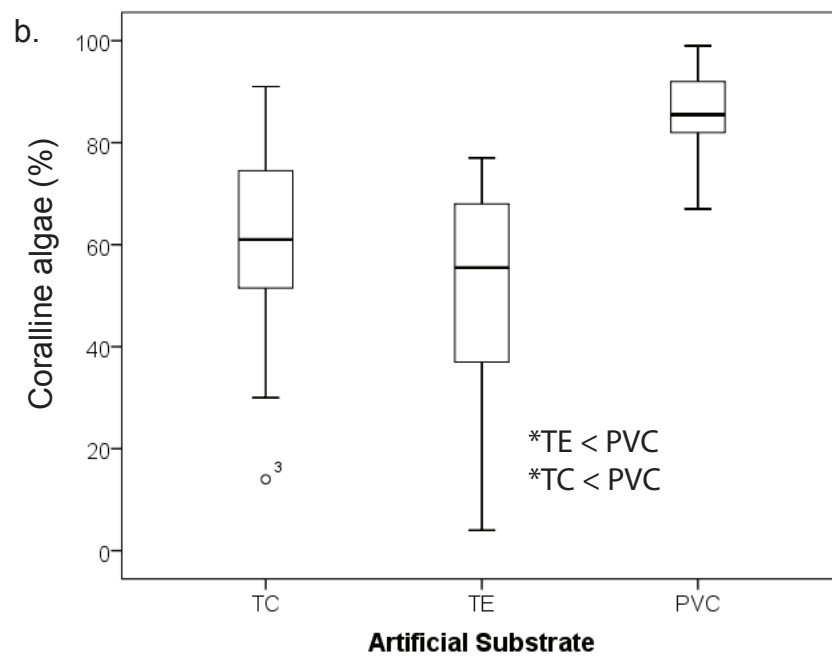
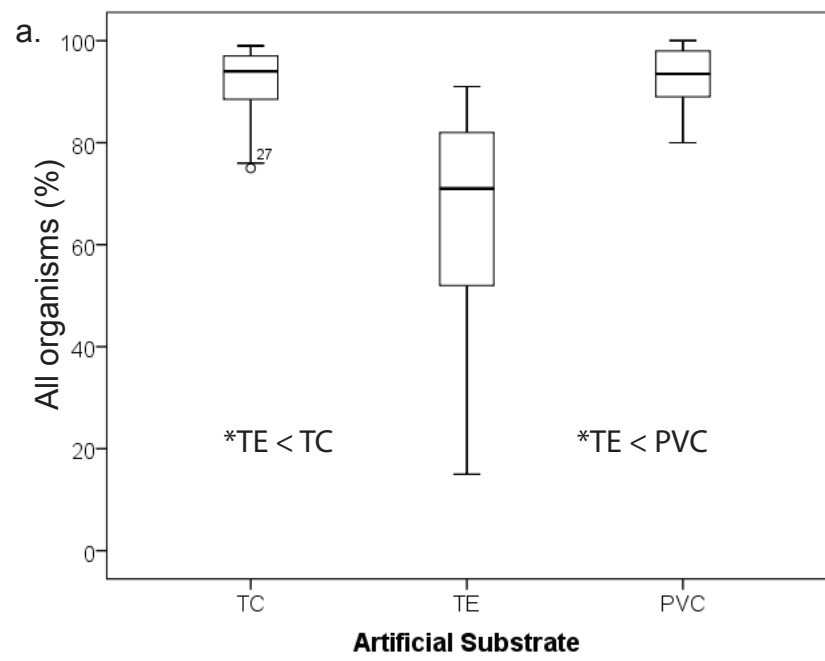


Figure 2



Supplementary Material Tables

[Click here to download Supplementary Material: Suppl. T1 T2 T3.docx](#)

Supplementary Material Fig 1

[Click here to download Supplementary Material: Suppl. Fig 1 MDS by site.eps](#)