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## **Epigenome-wide association study of global cortical volumes in Generation Scotland**

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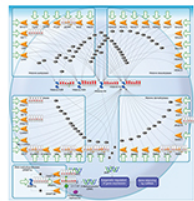
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 Epigenetic variation in the genome and its role in disease  
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**Epigenome-wide association study of global cortical volumes in Generation Scotland:  
Scottish Family Health Study**

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## Abstract

A complex interplay of genetic and environmental risk factors influence global brain structural alterations associated with brain health and disease. Epigenome-wide association studies (EWAS) of global brain imaging phenotypes have the potential to reveal the mechanisms of brain health and disease and can lead to better predictive analytics through the development of risk scores.

We perform an EWAS of global brain volumes in Generation Scotland using peripherally measured whole blood DNA methylation (DNAm) from two assessments, (i) at baseline recruitment, ~6 years prior to MRI assessment (N=672) and (ii) concurrent with MRI assessment (N=565). Four CpGs at baseline were associated with global cerebral white matter, total grey matter, and whole-brain volume (Bonferroni  $p \leq 7.41 \times 10^{-8}$ ,  $\beta_{\text{range}} = -1.46 \times 10^{-6}$  to  $9.59 \times 10^{-7}$ ). These CpGs were annotated to genes implicated in brain-related traits, including psychiatric disorders, development, and ageing. We did not find significant associations in the meta-analysis of the EWAS of the two sets concurrent with imaging at the corrected level.

These findings reveal global brain structural changes associated with DNAm measured ~6 years previously, indicating a potential role of early DNAm modifications in brain structure. Although concurrent DNAm was not associated with global brain structure, the nominally significant findings identified here present a rationale for future investigation of associations between DNA methylation and structural brain phenotypes in larger population-based samples.

**Keywords:** DNA methylation; epigenome-wide association study; cortical volumes; Generation Scotland

## 1 Introduction

2 Global brain structure is influenced by genetic and environmental factors, and has  
3 previously been associated with health and disorder traits across the lifetime (1–3). For  
4 instance, changes in global grey and white matter have been observed in a number of  
5 psychiatric and neurological disorders, including schizophrenia (4), major depressive disorder  
6 (MDD) (3), bipolar disorder (5), Rett syndrome (6), and Alzheimer’s disease (7). Previous  
7 studies have also found age-related reductions in both grey and white matter (8,9).

8 Such global brain structural changes in both health and disease may reflect genetic  
9 and environmental factors and their impact. While previous studies have focused on  
10 revealing the genetic architecture of brain structure, there are now opportunities to explore  
11 genetic and environmental risk factors through epigenetics, which correlate with changes in  
12 gene expression by modulating the genome in different cell types, without altering the  
13 underlying genome sequence (10). One such process, DNA methylation (DNAm), implicates  
14 the covalent addition of a methyl group to a cytosine nucleotide followed by guanine in DNA,  
15 resulting in Cytosine-phosphate-Guanine (CpG) sites (10).

16 DNAm is modulated by both genetic and environmental factors, and may thus aid in  
17 identifying genetic and environmental contributions to health and disease (11). Several brain-  
18 related traits and diseases are associated with variation in DNAm. MDD, a moderately  
19 heritable disorder, has been associated with differential methylation at several CpG sites,  
20 with a methylation risk score explaining 1.75% of the variance in the disorder (12). Further, in  
21 an epigenome-wide association study (EWAS) using blood, CpG sites associated with  
22 depressive symptoms were annotated to genes involved in axonal guidance (13).  
23 Schizophrenia has been associated with epigenetic variation at multiple loci that contribute  
24 to the polygenicity of the disorder (14,15). Finally, growing evidence has shown that DNAm  
25 can act as a proxy for the biological age of multiple tissues across life (16). These studies  
26 indicate that it may be possible, in future, to utilise DNAm modifications as biomarkers for  
27 brain-related healthy traits and diseases and to identify novel mechanisms contributing to  
28 these traits.

29 In recent years, increasing efforts have been made to identify epigenetic correlates of  
30 brain phenotypes, using both blood and brain tissue (17,18). To maximise statistical

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32 previous studies have focused on candidate genes and candidate epigenetic markers in  
33 relation to specific brain regions of interest, such as subcortical volumes in the hippocampus  
34 and amygdala, as well as cortical thickness and volume in Freesurfer-derived brain regions  
35 (18), although consistency between study findings is modest. Recent advances in high-  
36 throughput array technologies that can identify DNAm levels at over 450K and 850K locations  
37 along the genome have enabled researchers to identify DNAm-brain associations using a  
38 hypothesis-free approach using EWAS (19). DNAm modifications in relation to brain  
39 phenotypes have also been identified in patients as opposed to healthy individuals, including  
40 in the frontal cortex in schizophrenia (20,21), hippocampal volume in MDD (22), in the  
41 cerebral cortex in Alzheimer's disease (23), and in the frontal cortex in Parkinson's disease  
42 (24). Structural brain measures may therefore function as endophenotypes that can be used  
43 to assess the association between epigenetic modifications and brain health and disease.

44 The pathogenesis of psychiatric and neurodegenerative disorders has been associated  
45 with a multitude of cortical and subcortical brain regions with inconsistent results across  
46 studies (3,25–27), potentially indicating a role for whole-brain abnormalities in these  
47 disorders. Peripheral DNAm alterations associated with clinically relevant global brain  
48 structure may therefore further our mechanistic understanding of brain anatomy in both  
49 health and disease, may help to identify modifiable risk factors and may form a basis for the  
50 development of more accurate predictive risk scores capturing a wider array of potential  
51 influences.

52 The majority of the studies mentioned above used whole blood as a surrogate tissue  
53 for the brain due to inaccessibility of the brain ante-mortem. Although DNAm is reported to  
54 be tissue- and cell type-specific, similarities between blood and brain DNAm have also been  
55 identified (28). In addition, whole blood has successfully been used in the past to identify  
56 meaningful epigenetic differences in brain-related traits, as shown above (18).

57 Here, we sought to assess DNAm associations with Magnetic Resonance Imaging (MRI)  
58 global brain structural phenotypes, including cerebral white matter, total grey matter, and  
59 whole-brain volume using the Illumina Infinium MethylationEPIC array, capturing DNAm at  
60 approximately 850K CpG sites (29). Using DNAm measured ~6 years prior to MRI data  
61 collection, we examined whether CpG sites were associated with global brain structure at a  
62 later timepoint in N=672 individuals. We then investigated whether concurrently measured



63 DNAm was associated with global brain structure in N=565 individuals.

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## 64 **Methods**

65

### 66 **Study population: Generation Scotland: Scottish Family Health Study (GS:SFHS)**

67 GS:SFHS is a large, family-based epidemiological study aiming to investigate the  
68 genetics of health and disease in approximately 24,000 individuals aged 18-98 years across  
69 Scotland. Data collected between 2006 and 2011 consists of genetic, DNA methylation, and  
70 environmental variables (30,31). GS:SFHS received ethical approval from NHS Tayside  
71 Research Ethics Committee (REC reference number 05/S1401/89) and has Research Tissue  
72 Bank Status (reference: 20/ES/0021). Written informed consent was obtained from all  
73 participants.

74 A total of N=9,618 participants from GS responded when re-contacted at a later  
75 timepoint, and further data on mental health, specifically depression, was obtained. N=1,188  
76 were recruited for brain scanning, and approximately N=700 with both DNAm and  
77 neuroimaging data were available at the time of the current study. Details of recruitment and  
78 study information have been reported previously (32,33). The study was supported by the  
79 Wellcome Trust through a Strategic Award (reference 104036/Z/14/Z). Written consent at  
80 each stage of the study was obtained from all participants.

81 Two timepoints were used for the current study: blood samples were collected at  
82 baseline measurement (2006-2011), and concurrently with neuroimaging data (2015-2019).

83

## 84 **Phenotypes**

85

### 86 *Global brain volumes*

87 T1 images were processed using standard ENIGMA protocols (34) with FreeSurfer 5.3  
88 and all output was visually quality checked. Manual edits were applied as required to  
89 correct for inclusion of skull tissue, exclusion of brain tissue or for errors in parcellation.  
90 Global measures were extracted from the final output following all edits. Manual editing,  
91 although necessary, did introduce a degree of subjective bias, therefore 'editing' was  
92 included as a binary covariate (values: yes/no). Further, as the complete set of T1s was  
93 processed, quality checked and edited in two parts, 'batch' was also included as a covariate.

94 We used 3 global volume measures in the current study. Total cerebral white matter  
95 includes hyperintensities and excludes anything that is not white matter. Total grey matter is  
96 rendered by the sum of the cortex within the left and right hemispheres, as well as subcortical  
97 and cerebellar grey matter. Finally, whole-brain volume includes both grey and white matter,  
98 and corresponds to brain volume without the brain stem, ventricles, cerebrospinal fluid, and  
99 choroid plexus.

100

#### 101 *Baseline lifestyle factors and MDD status*

102 Body mass index (BMI) was calculated using height (m) and weight (kg) as measured  
103 by clinical staff at baseline recruitment. Participants were asked to report the number of units  
104 of alcohol consumed during the past week and their smoking status (never, former, current);  
105 pack years was used to measure heaviness of smoking in current smokers by multiplying the  
106 number of cigarette packs (20 cigarettes/pack) smoked per day by the number of years a  
107 person has smoked (35). MDD status was assessed at baseline using the Structured Clinical  
108 Interview of the Diagnostic and Statistical Manual, version IV (SCID) (36). Participants with no  
109 MDD were defined as those individuals who did not fulfil criteria for a current or previous  
110 MDD diagnosis following the SCID interview.

111

#### 112 *Concurrent lifestyle factors and MDD status*

113 At the follow-up assessment, participants were sent study packages that included  
114 questionnaires. Here, BMI was calculated using height (m) and weight (kg). Participants also  
115 recorded the number of units consumed during the past week, whether they were current,  
116 former, or non-smokers, and (if they smoked) the number of cigarettes smoked in an average  
117 week. Finally, MDD status was ascertained through the Composite International Diagnostic  
118 Interview-Short Form (CIDI-SF) (37), and participants with no MDD were those individuals  
119 who did not fulfil criteria for current or previous MDD diagnoses based on responses.

120

#### 121 **DNA methylation**

122 Baseline DNAm data was pre-processed and quality-checked for all individuals by  
123 Amador et al. in 2019 (38). At the concurrent timepoint, samples were placed on the array

124 at two different time points and were therefore processed separately. The main difference  
125 between processing and analysis pipelines related to how key covariates were adjusted for.  
126 At baseline these were regressed out during pre-processing, whereas for the concurrent  
127 batches they were included as covariates in downstream analyses. However, across all  
128 batches, standard quality check (QC) and pre-processing steps with regards to sample and  
129 probe exclusions were identical (see below). We note however that differences in the  
130 processing resulted in different numbers of final CpG sites included for analysis.

131 Cross-reactive (N=42,558) and polymorphic (N=10,971) CpGs, obtained from  
132 McCartney et al. (2016) were removed from both the baseline and concurrent DNAm datasets  
133 (39).

#### 134 *Baseline DNA methylation*

135 Genome-wide DNAm data profiled from whole blood samples was available for  
136 9,873 individuals in GS:SFHS using the Illumina Human-MethylationEPIC BeadChip (29).  
137 Samples were obtained and DNA was extracted between 2006-2011. DNAm profiling using  
138 the Illumina Human-MethylationEPIC BeadChip (29) was performed in two sets (in 2016, set  
139 A<sub>N</sub>=5101; in 2019, set B<sub>N</sub>=4,450) and pre-processing and QC was conducted once the second  
140 set was released, as detailed in Amador et al. (38,40,41). Participants were removed due to  
141 a number of reasons, including sex mismatch (N<sub>removed</sub>=24), having more than 1% CpG sites  
142 with a detection p-value>0.05 (N<sub>removed</sub>=52), being an outlier for bisulphite conversion  
143 control probes (N<sub>removed</sub>=1), having a median methylated signal intensity more than 3  
144 standard deviations lower than expected (N<sub>removed</sub>=74), and other technical and dataset-  
145 specific issues (N<sub>removed</sub>=602, see Supplementary Materials). A total of 10,495 CpG sites were  
146 removed due to low beadcount, poor detection p-value, and sub-optimal binding.

147 R package “minfi” was used to read in the IDAT files, compute M and beta values, and  
148 remove probes with large detection p-values, and to compute principal components (PC) of  
149 control probes. Correction was then applied for (1) technical variation, where M values were  
150 included as outcome variables in a mixed linear model adjusting for appointment date and  
151 Sentrix ID (random effects), jointly with Sentrix position, batch, clinic, year, weekday, and 10  
152 PCs (fixed effects); and (2) biological variation by fitting residuals of (1) as outcome variables  
153 in a second mixed linear model adjusting for genetic and common family shared

154 environmental contributions (random effects classed as G: common genetic; K: kinship; F:  
155 nuclear family; C: couple; and S: sibling) and sex, age, and estimated cell type proportions  
156 (CD8T, CD4T, NK, Bcell, Mono, Gran) (fixed effects) (42). The final number of CpG sites that  
157 converged for these analyses was 674,246 across the 22 autosomes.

158

### 159 *Concurrent DNA methylation*

160 Genome-wide DNAm data profiled from whole blood samples was available for a total  
161 of 710 individuals using the Illumina Human-MethylationEPIC BeadChip (29). Pre-processing  
162 was carried out in two separate sets ( $N_{\text{set } 1}=404$ ;  $N_{\text{set } 2}=306$ ) intended as discovery and  
163 replication datasets, by Walker et al. (43,44). Meffil (45) was used to remove samples if: there  
164 was a mismatch between self-reported and methylation-predicted sex and if >0.5% of probes  
165 failed the detection p-value threshold (>0.01); probes were removed if >1% samples failed  
166 the detection p-value >0.01 and if >5% of samples failed the beadcount threshold (N=3). In  
167 addition, samples were removed if they showed evidence of dye bias and they were outliers  
168 for the bisulphite conversion control probes. ShinyMethyl (46) was then used to plot the log  
169 median intensity of methylated and unmethylated signals per array and inspect the output  
170 from the control probes; outlying samples detected by visual inspection were excluded.  
171 Meffil (45) was then used again to remove any additional samples who had a sex mismatch.  
172 PC plots were made using the first two methylation principal components and any additional  
173 outlying samples on the basis of these plots were removed. Finally, data were normalised  
174 using the dasen method in watermelon, and M-values were generated using the beta2m  
175 function in lumi (47). The final number of CpG sites after pre-processing was N=768,068 (set  
176 1) and N=765,695 (set2) across the 22 autosomes.

177

## 178 **Statistical methods**

179

### 180 *Epigenome-wide association*

181 We used the “limma” package (48) in R to run linear regression models for both  
182 baseline and concurrent DNAm data, where each CpG was included as an outcome variable.  
183 Brain cortical volumes, specifically cerebral white matter, total grey matter, and whole brain  
184 volume were included as predictor variables in separate EWAS at each DNAm timepoint.

185 TheR code for these analyses is available in the Supplementary Materials.

186 Covariates for each model using baseline DNAm were MRI site (to account for  
187 different data collection sites; see Supplementary Materials), age, age<sup>2</sup>, sex, intracranial  
188 volume, and set (to account for different DNAm data pre-processing sets). Due to the impact  
189 of lifestyle factors on DNAm (49–52), BMI, alcohol units, smoking status, and pack years  
190 were also included as covariates. Lastly, due to the increased prevalence of MDD in the  
191 dataset, MDD status was included as a covariate in all models. Technical (batch,  
192 appointment date) and biological (relatedness, cell type estimations, methylation principal  
193 components) variables were regressed out during pre-processing and were not included as  
194 covariates in downstream analyses. After QC, there were 674,246 CpGs and epigenome-  
195 wide significance was determined by a Bonferroni correction ( $0.05/674,246$ ,  $p \leq 7.41 \times 10^{-8}$ ).

196 For both sets at the concurrent DNAm timepoint, covariates for each model were  
197 DNAm batch, 5 cell type proportion estimations (granulocytes, natural killer cells, B-  
198 lymphocytes, CD4+ T-lymphocytes and CD8+ T-lymphocytes), MRI site, age, age<sup>2</sup>, sex,  
199 intercranial volume, BMI, smoking status, number of cigarettes smoked/week, alcohol units,  
200 MDD status, and 20 methylation PCs. Bonferroni correction was applied based on the number  
201 of CpGs remaining in each set after QC (set 1:  $0.05/768,068$  CpGs,  $p \leq 6.51 \times 10^{-8}$ ; set 2:  
202  $0.05/765,695$  CpGs,  $p \leq 6.52 \times 10^{-8}$ ).

203 The Blood Brain DNA Methylation Comparison Tool (53)  
204 (<http://epigenetics.essex.ac.uk/bloodbrain/>) investigates the correlation between DNAm  
205 from whole blood and four brain regions (prefrontal cortex, entorhinal cortex, superior  
206 temporal gyrus, and cerebellum) for all probes on the Illumina 450K array (54). We used this  
207 resource to investigate the strength of correlation between the two tissues for CpGs identified  
208 here.

209

#### 210 *Meta-analysis using METAL – concurrent timepoint*

211 At the concurrent timepoint, in set 1, N=331 individuals were available with global  
212 volume and methylation data after QC and N=234 were available in set 2. Meta-analysis of  
213 these two datasets was performed in METAL (55) using p-value based analysis (N=565). The  
214 meta-analysis was based on N=769,263 CpGs across both sets and a Bonferroni correction

215 (0.05/769,263) was used to define epigenome-wide significance ( $p \leq 6.49 \times 10^{-8}$ ).

#### 216 *Pathway analysis*

217 We annotated CpG sites to genes through the Infinium MethylationEPIC BeadChip  
218 database (29). The database provides information about genes, chromosome location, start  
219 and end sites, and other features.

220 We used missMethyl (56), accessed via methylGSA (57), to assess pathway enrichment  
221 for differentially-methylated CpG sites. The package allows correction for biases in the  
222 representation of genes on the Infinium BeadChip. Gene Ontology (GO) terms were accessed  
223 using the msigdb package (58). Pathways included in the analysis were all GO pathways of  
224 size 1-250 genes inclusive. CpG sites included in the analysis were those significant at a  
225 threshold of  $p < 1 \times 10^{-5}$ , as used in previous studies (59). Information on GO pathways can be  
226 accessed via [www.geneontology.org](http://www.geneontology.org) using Gene Ontology identifiers, comprised of "GO"  
227 followed by a string of numbers (e.g. GO:0000000).

228

#### 229 *Power analysis – concurrent timepoint*

230 Since the concurrent data was formed by two smaller samples of pre-processed  
231 data, we additionally conducted power analysis to determine whether our concurrent  
232 samples had sufficient power to detect a significant effect. This was conducted using effect  
233 sizes from the baseline data to inform the power calculations. We used the "pwr.f2.test"  
234 function in package "pwr" in R and the set parameters were as follows:

235 1. Regression coefficients: DNAm batch, 5 cell type estimations (granulocytes,  
236 natural killer cells, B-lymphocytes, CD4+ T-lymphocytes and CD8+ T-lymphocytes), MRI site,  
237 age, age<sup>2</sup>, sex, intercranial volume, BMI, smoking status, number of cigarettes  
238 smoked/week, alcohol units, MDD status, 20 methylation principal components.

239 2. Effect size: we input the largest effect size identified in EWAS at baseline (N=672)  
240 for each global volume.

241 3. Significance level: to adjust for multiple testing correction (FDR), the p-value for a  
242 single potential test was set based on the number of CpG sites in each dataset (set 1:  
243  $0.05/768,068 = 6.51 \times 10^{-8}$ ; set 2:  $0.05/765,695 = 6.53 \times 10^{-8}$ ).

244 4. Power: to observe different power percentages, we input 60%, 80%, 90%, 95%  
245 and 99% power.

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246 **Results**

247

248 *Demographic characteristics*

249 There were N=672 individuals in the baseline EWAS, N=331 in the set 1 concurrent  
 250 EWAS, and N=234 in the set 2 concurrent EWAS. Demographic characteristics for all  
 251 individuals are presented in Table 1. Further descriptive characteristics regarding global  
 252 volumes are presented in Supplementary Table 1.

<b>Demographic characteristics</b>	<b>Baseline (N=672)</b>	<b>Concurrent set 1 (N=331)</b>	<b>Concurrent set 2 (N=234)</b>
<b>Age – Mean (SD), range</b>	52.29 (9.93), 18-75	60.45 (8.42), 28-78	59.61 (10.21), 28-81
<b>Sex</b>			
Female	406	193	132
Male	266	138	102
<b>Set</b>			
1	621	-	-
2	51	-	-
<b>BMI – Mean (SD), range</b>	27.13 (4.96), 15.96-56.60	27.48 (5.18), 16.42-51.75	28.23 (5.31), 19-20-52.81
<b>Alcohol units – Mean (SD), range</b>	10.53 (16.44), 0-326	7.12 (8.91), 0-60	7.39 (9.67), 0-60
<b>Smoking status</b>			
Current smoker	83	16	12
Former smokers (quit < 1 year ago)	10	124	92
Former smokers (quit > 1 year ago)	208		
Never smoked tobacco	371	191	130
<b>Pack years – Mean (SD), range</b>	7.59 (14.56), 0-111	-	-
<b>Cigarettes smoked/week</b>			
1-10 cigarettes	-	10	6
11-20 cigarettes	-	10	9
<b>MDD status</b>			
Cases	121	83	83
Controls	551	248	151

253 **Table 1.** Demographic characteristics for individuals with global volume data, including  
 254 lifestyle variables and MDD. “-” indicates that there was no data of the sort for the  
 255 respective dataset. Former smokers at the baseline measurement were split into those who  
 256 quit less than a year ago and those who quit more than a year ago; at the concurrent  
 257 timepoint, this division is not made.

258

259

260 *Baseline EWAS*

261 Baseline EWAS identified 1, 3, and 2 CpG sites that were associated with cerebral  
 262 white matter, total grey matter, and whole-brain volume, respectively ( $p \leq 7.41 \times 10^{-8}$ ). Both  
 263 CpGs associated with whole brain volume were also associated with total grey matter and  
 264 were significantly hypermethylated. One CpG site associated with cerebral white matter and  
 265 one associated with total grey matter were hypomethylated. As shown in Figure 1A-C, CpG  
 266 associations with grey matter were stronger than with white matter. Information about each  
 267 CpG site is shown in Table 2.

268

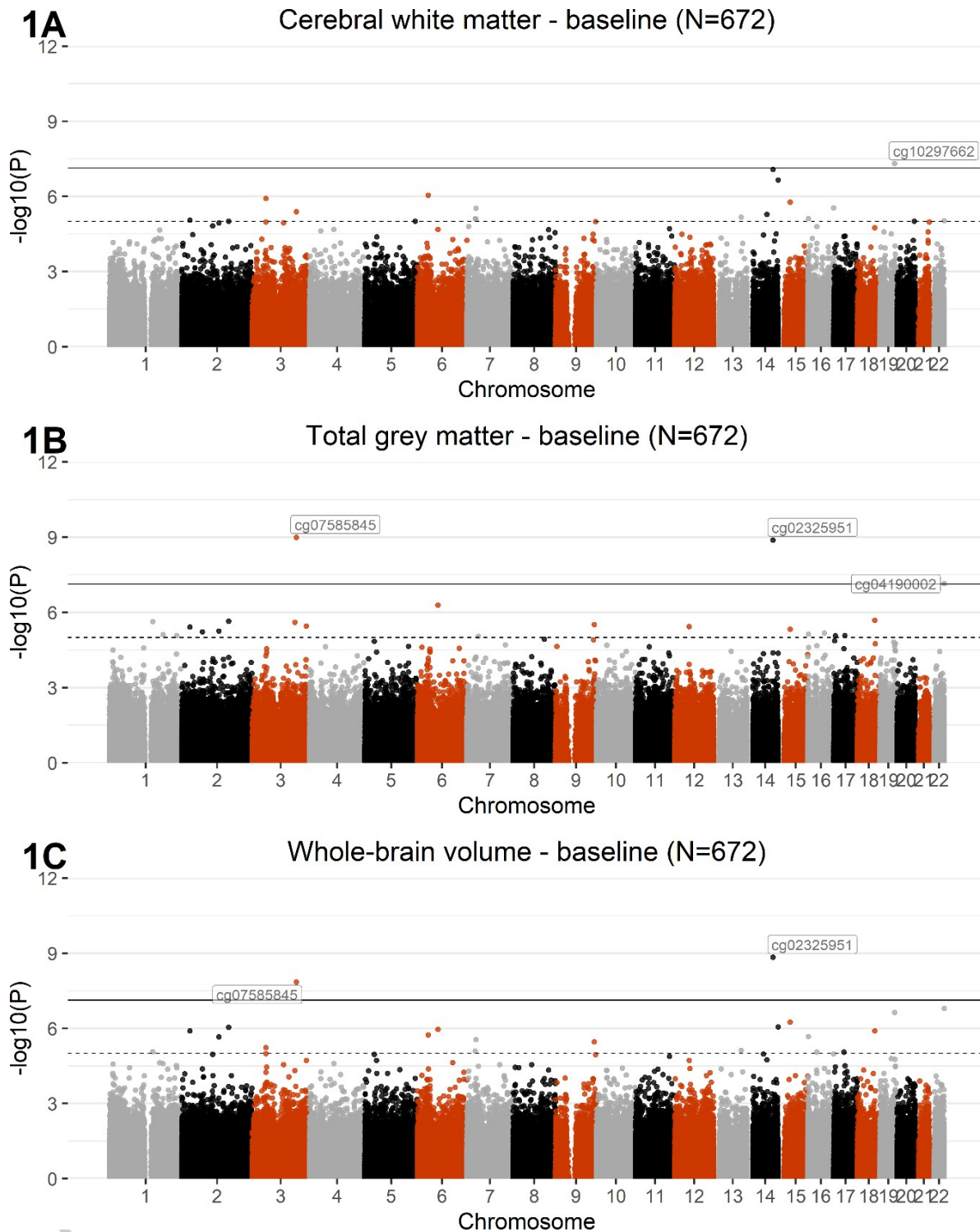
Phenotype	CpG site	Gene	C	$\beta$	P-value	P-corr	CpG – previously associated traits	Gene – previously associated traits
Total grey matter	cg07585845 (EPIC)	-	3	$9.59 \times 10^{-7}$	$1.02 \times 10^{-9}$	0.0007	-	-
Whole-brain volume	cg07585845 (EPIC)	-	3	$4.47 \times 10^{-7}$	$1.38 \times 10^{-8}$	0.009	-	-
Total grey matter	cg02325951 (450K)	<i>FOXN3</i>	14	$6.53 \times 10^{-7}$	$1.31 \times 10^{-9}$	0.0009	-	Acute myeloid leukemia ( $p=8 \times 10^{-21}$ $p=3 \times 10^{-14}$ ; (60)) Heel bone mineral density ( $p=2 \times 10^{-12}$ ; (61)) Intelligence ( $p=1 \times 10^{-11}$ ; (62)) Self-reported educational attainment ( $p=8 \times 10^{-11}$ ; (63)) Cognitive function measurement ( $p=2 \times 10^{-9}$ ; (63)) Mathematical ability ( $p=3 \times 10^{-9}$ ; (63))
Whole-brain volume	cg02325951 (450K)	<i>FOXN3</i>	14	$3.26 \times 10^{-7}$	$1.45 \times 10^{-9}$	0.001	Sex ( $p=2 \times 10^{-54}$ ; $1.8 \times 10^{-42}$ ; (54))	Smoking status measurement ( $p=7 \times 10^{-9}$ ; (64)) Risk-taking behaviour ( $p=8 \times 10^{-9}$ ; (65))
Cerebral white matter	cg10297662 (EPIC)	<i>PNKP</i>	19	$-1.46 \times 10^{-6}$	$4.92 \times 10^{-8}$	0.03	-	Involved in DNA repair; mutations at locus associated with microcephaly, seizures, and developmental delay (66)
Total grey matter	cg04190002 (450K)	<i>SHANK3</i>	22	$-3.75 \times 10^{-7}$	$7.31 \times 10^{-9}$	0.04	Sex ( $p=5.4 \times 10^{-19}$ ; (62))	Self-reported educational attainment ( $p=2 \times 10^{-20}$ ; (63)) Mathematical ability ( $p=1 \times 10^{-17}$ ; (63)) Cognitive function measurement ( $p=3 \times 10^{-12}$ ; (63)) Schizophrenia ( $p=3 \times 10^{-12}$ ; (67))

269

270 **Table 2.** CpG sites significantly associated with cerebral white matter, total grey matter, and  
 271 whole-brain volume (N=672), along with gene annotations (Gene), chromosome (C),

272 standardised effect size ( $\beta$ ), nominal (P-value) and multiple comparison-corrected p-values  
273 (P-corr). Traits previously associated with each CpG site were extracted from EWAS  
274 catalogues (<http://www.ewascatalog.org/>, association between traits and CpGs on Illumina  
275 450K array at  $p \leq 1.0 \times 10^{-4}$ ; and <http://www.bioapp.org/ewasdb/>, (68)), association between  
276 traits and CpGs on Illumina 450K and EPIC arrays at  $p \leq 1.0 \times 10^{-3}$ ). Gene information was  
277 extracted from the GWAS catalogue (<https://www.ebi.ac.uk/gwas/>; associations between  
278 traits and SNPs at  $p < 1.0 \times 10^{-5}$ ). All associations included in the table from these two  
279 catalogues are genome-wide significant.

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282 **Figure 1A, 1B, 1C.** Manhattan plots showing the results from EWASs of cerebral white matter  
 283 (1A), total grey matter (1B), and whole-brain volume (1C), using baseline DNAm data (N=672).  
 284 The black line defines the threshold for epigenome-wide significance ( $p \leq 7.41 \times 10^{-8}$ ) and the  
 285 dotted line defines CpG sites at  $p \leq 1 \times 10^{-5}$ . Epigenome-wide significant hits for each phenotype  
 286 are labelled on the graph.

287 *Correlation between whole blood DNAm and four brain regions*

288 We used the Blood Brain DNA Methylation Comparison Tool (53) to investigate the  
289 correlation between blood and brain methylation measurements for two of the CpGs  
290 identified here, located on the 450K array, and four brain regions. cg04190002 was strongly  
291 correlated with prefrontal cortex ( $r=0.579$ ,  $p=6.55 \times 10^{-8}$ ), entorhinal cortex ( $r=0.564$ ,  
292  $p=2.94 \times 10^{-7}$ ), superior temporal gyrus ( $r=0.598$ ,  $p=1.5 \times 10^{-8}$ ), and cerebellum ( $r=0.663$ ,  
293  $p=3.02 \times 10^{-10}$ ), while cg02325951 was strongly correlated with prefrontal cortex ( $r=0.858$ ,  
294  $p=1.73 \times 10^{-22}$ ), entorhinal cortex ( $r=0.868$ ,  $p=1.19 \times 10^{-22}$ ), and superior temporal gyrus  
295 ( $r=0.871$ ,  $p=3.32 \times 10^{-24}$ ).

296

297 *Baseline Pathway Analysis*

298 Enrichment of differentially methylated regions in biological pathways was analysed  
299 using missMethyl (56), where an over-representation analysis of GO pathways was performed  
300 for sets of genes annotated to CpG sites differentially expressed at  $p < 1 \times 10^{-5}$  ( $N_{\text{cerebral white matter}}$ :  
301 19,  $N_{\text{total grey matter}}$ : 22,  $N_{\text{whole-brain volume}}$ : 21).

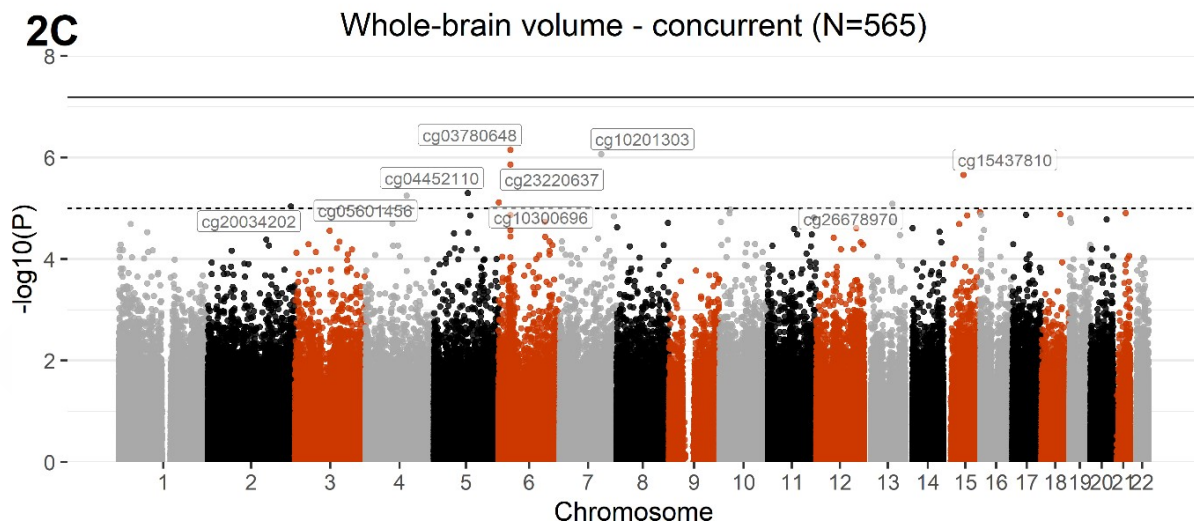
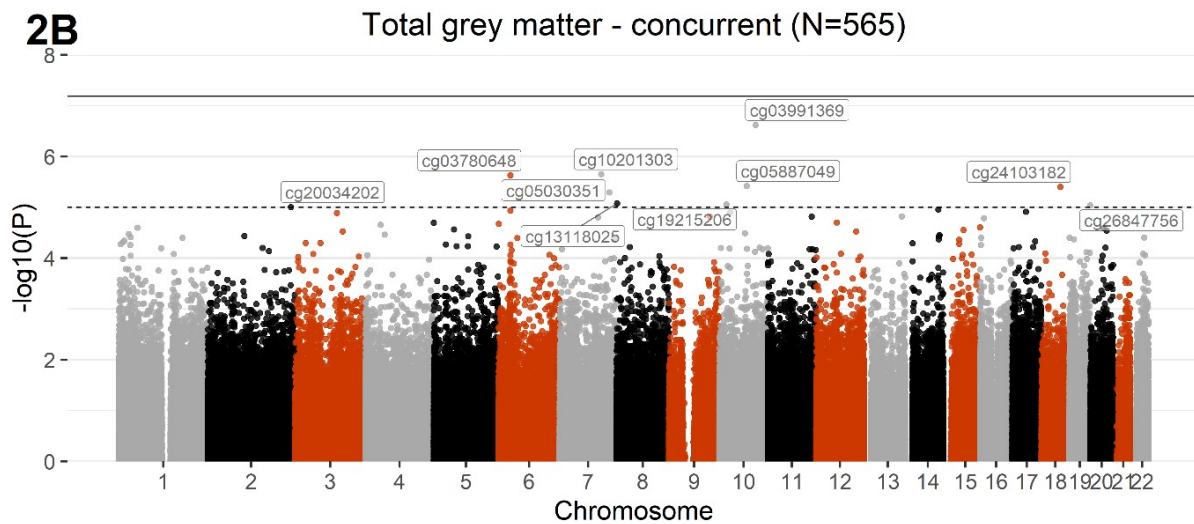
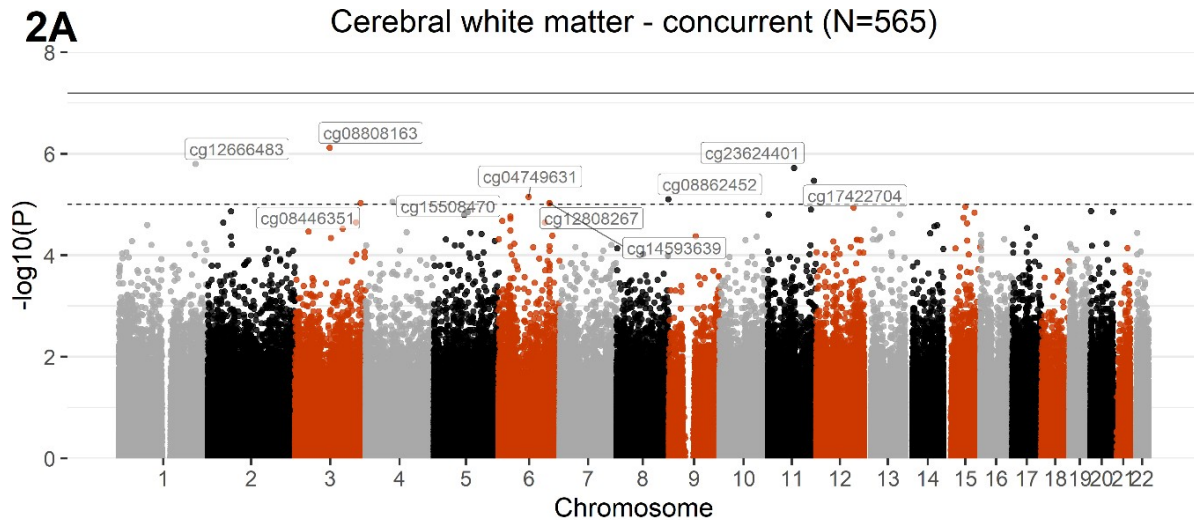
302 There were no over-represented pathways after multiple correction. A number of  
303 brain-related biological processes, molecular functions, and cellular components were  
304 included in the top 10 significant pathways (Supplementary Table 2). For instance, guanylate  
305 kinase-associated protein clustering, which facilitates assembly of post-synaptic density of  
306 neurons (GO:0097117), was found to be over-represented for all three imaging phenotypes  
307 (cerebral white matter nominal  $p$ -value=0.0007; total grey matter nominal  $p$ -value=0.001;  
308 whole-brain volume nominal  $p$ -value=0.0009). Positive regulation of synapse structural  
309 plasticity (GO:0051835) was over-represented in both cerebral white matter (nominal  $p$ -  
310 value=0.002) and total grey matter (nominal  $p$ -value=0.002). Finally, forebrain generation of  
311 neurons (GO:0021872; nominal  $p$ -value=0.001) was over-represented for cerebral white  
312 matter.

313

314 *Concurrent EWAS*

315 Meta-analysis of EWAS across the two concurrent sets did not reveal any Bonferroni-  
316 corrected CpG sites associated with any of the global volumes (Figure 2A-C). A list of the top

317 10 CpGs associated with cerebral white matter (EWAS<sub>set 1</sub>  $\beta_{\text{range}}=4.71 \times 10^{-6}$ -  $6.53 \times 10^{-6}$ ;  
318 EWAS<sub>set 2</sub>  $\beta_{\text{range}}=1.02 \times 10^{-5}$ - $8.75 \times 10^{-6}$ ) total grey matter (EWAS<sub>set 1</sub>  $\beta_{\text{range}}=6.71 \times 10^{-6}$ - $8.03 \times 10^{-6}$ ;  
319 EWAS<sub>set 2</sub>  $\beta_{\text{range}}=1.03 \times 10^{-5}$ - $8.84 \times 10^{-6}$ ), and whole-brain volume (EWAS<sub>set 1</sub>  $\beta_{\text{range}}=2.69 \times 10^{-6}$ -  
320  $4.05 \times 10^{-6}$ ; EWAS<sub>set 2</sub>  $\beta_{\text{range}}=6.23 \times 10^{-6}$ - $6.69 \times 10^{-6}$ ), is presented in Supplementary Tables 3-5.  
321 Genes annotated to these top 10 CpGs have previously been implicated in brain-related  
322 phenotypes, including psychiatric disorders (MDD (69–72), schizophrenia (73)),  
323 neurodegenerative disorders (neurofibrillary tangles and PHF-tau measurement in  
324 Alzheimer’s Disease (74)), and cognitive traits (mathematical ability, self-reported  
325 educational attainment (75)). Results reported here are nominal and should be supported  
326 by further large-scale cohorts.



327

328 **Figure 2A, 2B, 2C.** Manhattan plots showing meta-analysis of EWAS of cerebral white matter  
 329 (2A), total grey matter (2B), and whole-brain volume (2C), across the 2 concurrent sets ( $N_{\text{set } 1}=331$ ;  
 330  $N_{\text{set } 2}=234$ ;  $N_{\text{total}}=565$ ). The black line defines the threshold for epigenome-wide  
 331 significance ( $p \leq 6.5 \times 10^{-8}$ ) and the dotted line defines  $p \leq 1 \times 10^{-5}$ . CpGs that met a significance of  
 332  $p \leq 1 \times 10^{-5}$  are labelled on the graph.

333 *Concurrent Pathway Analysis*

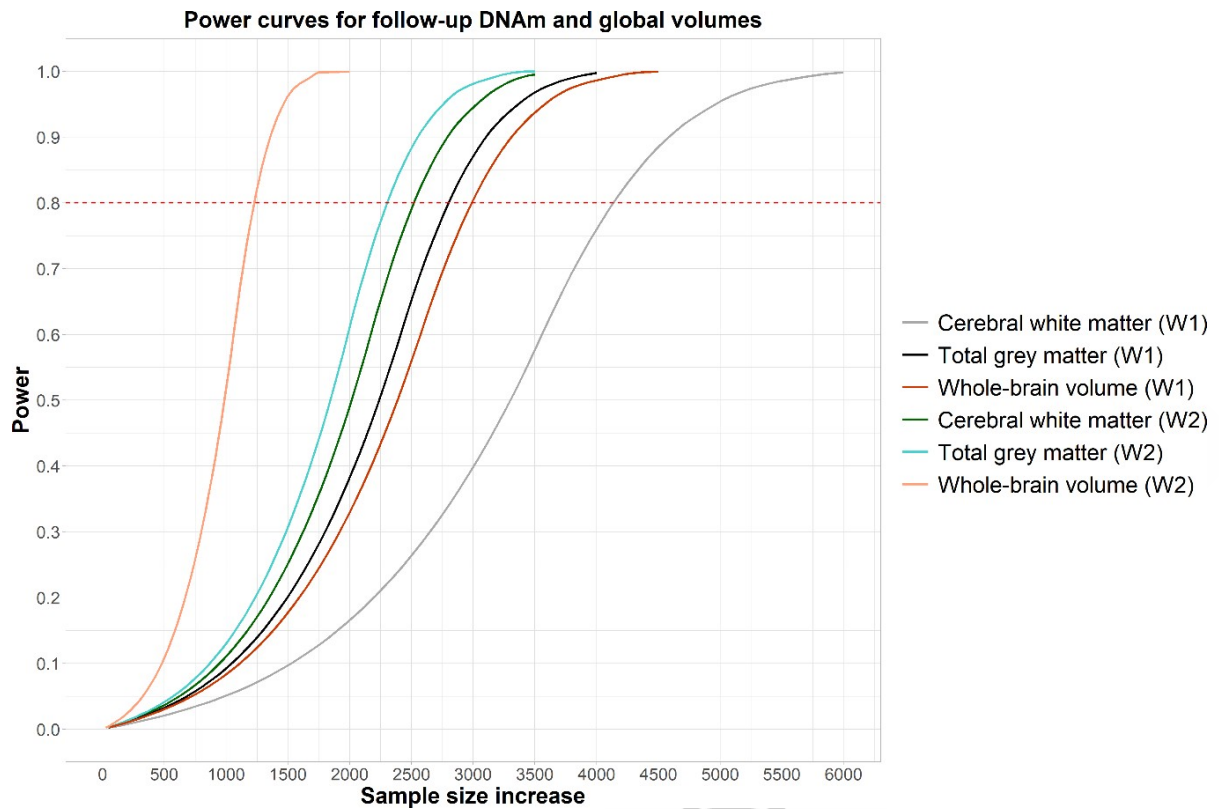
334 As above, enrichment of differentially methylated regions in specific pathways was  
335 assessed using missMethyl (50) for sets of genes annotated to CpG sites differentially  
336 expressed at  $p < 1 \times 10^{-5}$  ( $N_{\text{cerebral white matter}}: 10$ ,  $N_{\text{total grey matter}}: 10$ ,  $N_{\text{whole-brain volume}}: 9$ ). There were  
337 no over-represented pathways following FDR adjustment for multiple comparisons. The top  
338 10 most significant pathways for each phenotype indicated a pattern of phenotype-specific  
339 biological processes, molecular functions, and cellular components (Supplementary Table 6).  
340 For instance, over-represented pathways in cerebral white matter included myelination  
341 (GO:0042552; nominal p-value=0.002), ensheathment of neurons (GO:0007272; nominal p-  
342 value=0.002), axon ensheathment (GO:0008366; nominal p-value=0.001), glial cell  
343 development (GO:0021782; nominal p-value=0.001) and glial cell differentiation  
344 (GO:0010001; nominal p-value=0.004). Total grey matter over-represented pathways  
345 included glutamate catabolic process to aspartate (GO:0019550; nominal p-value=0.0009)  
346 and to 2-oxoglutarate (GO:0019551; nominal p-value=0.0009). Finally, over-represented  
347 pathways in whole-brain volume included several MHC-related biological processes, including  
348 regulation (GO:0002586; nominal p-value=0.001) and negative regulation (GO:0002587;  
349 nominal p-value=0.0009) of antigen processing and presentation of peptide antigen via MHC  
350 class II, negative regulation of antigen processing and presentation of peptide or  
351 polysaccharide antigen via MHC class II (GO:0002581; nominal p-value=0.001), as well as N-  
352 acetyllactosaminide beta-1,3-N-acetylglucosaminyltransferase (GO:0008532, molecular  
353 function, nominal p-value=0.001), an enzyme encoded by the gene *B3GNT2*, which is highly  
354 expressed in whole-brain, hippocampus, amygdala, cerebellum, and caudate nucleus  
355 (<https://www.uniprot.org/uniprot/Q9Z222>).

356

357 *Power curves for concurrent data*

358 Power curves for the three imaging phenotypes are presented in Figure 3. Further  
359 details, including effect size for each phenotype, are included in Supplementary Tables 7 and  
360 8. These indicate that approximately 1,000-6,000 individuals (depending on phenotype)  
361 would be needed to detect an effect after multiple correction.





362

363

364

365 **Figure 3.** Power curves for cerebral white matter, total grey matter, and whole-brain volume  
 366 calculated separately for set 1 and set 2. The x-axis indicates how many participants would  
 367 be needed to detect an effect with 60%, 80%, 90%, 95% or 99% power at  $p < 6.51 \times 10^{-8}$  (set 1  
 368 (W1)) and  $p < 6.53 \times 10^{-8}$  (set 2 (W2)) with 36 regression coefficients included in the linear  
 369 model. Effect sizes were calculated based on the largest effect size obtained in EWAS for each  
 370 phenotype at baseline.

371 **Discussion**

372 We report a number of significant associations between DNAm measured ~6 years  
373 prior to MRI data collection and cerebral white matter ( $N_{\text{significant CpGs}}=1$ ), total grey matter  
374 ( $N_{\text{significant CpGs}}=3$ ), and whole-brain volume ( $N_{\text{significant CpGs}}=2$ ) ( $N=672$ ), annotated to genes  
375 involved in brain-related traits. There were no significant associations between DNAm  
376 collected concurrently with MRI data ( $N=565$ ). In addition, pathway analysis did not uncover  
377 any significant findings for either the baseline or concurrent analyses. Power analysis of the  
378 concurrent data using baseline data for effect size confirmed that approximately 1,000-  
379 6,000 individuals (depending on phenotype) would be needed to detect a statistically  
380 significant effect.

381 For the analysis of associations between DNAm measured at baseline and cortical  
382 volumes ~6 years later, one CpG associated with cerebral white matter, cg10297662, was  
383 annotated to *PNKP*. This CpG site has not previously been associated with any other traits, to  
384 the best of our knowledge. *PNKP* is involved in DNA repair following ionizing radiation or  
385 oxidative damage (76) and is expressed in a number of tissues, including the brain.  
386 Mutations in this gene have been associated with a number of neural conditions, including  
387 microcephaly, developmental delay, seizures, and cerebellar ataxia (66,77). These mutations  
388 have been shown to lead to white matter defects, which is the phenotype investigated here  
389 (78). Previous evidence also indicates that loss of *PNKP* strongly impacts oligodendrocytes,  
390 leading to white matter abnormalities (79). Efforts should be made to identify whether the  
391 relationship between *PNKP* mutations and defects in white matter is mediated by  
392 differential DNAm at specific sites.

393 Two CpGs, cg07585845 and cg02325951, were associated with both total grey matter  
394 and whole-brain volume. cg07585845 has not been previously associated with any traits nor  
395 annotated to any genes. cg02325951 was previously associated with sex in a study  
396 investigating methylation trajectories across human foetal brain development ( $p=2 \times 10^{-54}$ ;  
397 (80)). The gene to which cg02325951 is annotated, *FOXN3*, is involved in several physiological  
398 processes, such as development, ageing, obesity, and cancer and is expressed in multiple  
399 tissues, including the forebrain and midbrain. Further, animal studies show that mutations  
400 within the gene have been associated with craniofacial defects (81). In addition, *FOXN3* has  
401 previously been associated with several brain-related phenotypes in previous GWAS,

402 including intelligence ( $p=1 \times 10^{-11}$ ; (62)), self-reported educational attainment ( $p=8 \times 10^{-11}$ ),  
403 cognitive function measurement ( $p=2 \times 10^{-9}$ ), and mathematical ability ( $p=3 \times 10^{-9}$ ) (63). These  
404 cognition-related phenotypes have previously been associated with whole brain volume,  
405 where higher cognition was associated with a larger brain size (76). Future studies should  
406 investigate whether DNAm localized to *FOXN3* plays a role in cognition development  
407 through modifications in whole-brain volume.

408 Finally, in addition to the two CpGs above, total grey matter was also associated with  
409 cg04190002, a CpG previously associated with sex in newborns ( $p=5.4 \times 10^{-19}$ ; (82)). The CpG is  
410 annotated to *SHANK3*, which encodes multidomain scaffold proteins of the postsynaptic  
411 density connecting neurotransmitter receptors, among other membrane proteins and is  
412 expressed in the cerebral cortex and the cerebellum. The gene has previously been  
413 associated with a host of brain disorders and traits, including self-reported educational  
414 attainment ( $p=2 \times 10^{-20}$ ), mathematical ability ( $p=1 \times 10^{-17}$ ), cognitive function measurement  
415 ( $p=3 \times 10^{-12}$ ) (63) and schizophrenia ( $p=3 \times 10^{-9}$ ; (67)), and mutations have previously been  
416 associated with autism spectrum disorder (83). These disorders in turn have been  
417 associated with changes in grey matter (84), and future studies should investigate whether  
418 these psychiatric disorders are also associated with differential DNAm at cg04190002, and  
419 other probes localized to *SHANK3*, as well as explore whether associations are mediated by  
420 global brain phenotypes.

421 Blood and brain methylation measures for both cg02325951 and cg04190002 (both  
422 CpGs on the 450K array) were strongly correlated, indicating that whole blood is a suitable  
423 proxy tissue for investigating associations with brain phenotypes, at least for these probes.  
424 Future studies exploring DNAm in relation to global brain phenotypes and associated traits  
425 may therefore benefit from whole blood DNAm measurements.

426 DNAm profiled at a different timepoint to phenotype measurement has previously  
427 yielded interesting results. Barbu et al. (2020) found that a methylation risk score calculated  
428 from DNAm profiled 4-11 years prior to MDD diagnosis was significantly associated with  
429 incident cases who were well at DNAm measurement but went on to develop MDD (12). Clark  
430 et al. (2020) similarly associated DNAm profiled in MDD patients at baseline with MDD status  
431 6 years later (85). These previous findings indicate that DNAm measured prior to phenotype  
432 measurement may provide meaningful insight into phenotype development and change

433 across time. The findings above relating DNAm measured previously to MRI scans may  
434 therefore aid in the investigation of epigenetic differences in brain-related disease and health  
435 at a later timepoint, although further longitudinal replication is needed to verify these  
436 associations.

437 Associations between DNAm measured concurrently to MRI scans did not yield any  
438 significant findings. Power calculations using the baseline data to derive effect size showed  
439 that approximately 1,000-6,000 participants (depending on phenotype) would be needed to  
440 uncover a significant effect at epigenome-wide level. This number is supported by previous  
441 studies, such as Jia et al. (2019), who analysed 3,337 individuals across 11 cohorts as part of  
442 ENIGMA to find 2 CpGs significantly associated with hippocampal volume (19). This may  
443 indicate that null findings were due to lack of power at the concurrent timepoint. Null  
444 findings here should serve as a stimulus for larger collaborations and meta-analyses in  
445 future.

446 Further, effect sizes for both timepoints were much smaller than those identified in  
447 previous studies that analysed larger sample sizes in specific brain regions (19) (largest  
448 baseline effect size:  $1.46 \times 10^{-6}$ ; largest concurrent effect size:  $1.06 \times 10^{-6}$ ), which suggests that  
449 findings here should be interpreted with caution. The results here indicate that global  
450 associations with DNAm may be weaker than those at a regional level. Future studies may  
451 therefore benefit from focussing on lobe- and region-specific correlates of DNAm.

452 At the concurrent timepoint, DNAm data was pre-processed and quality-checked in 2  
453 sets, resulting in a different number of final CpGs ( $N_{\text{CpG set 1}}=768,068$ ;  $N_{\text{CpG set 2}}=765,695$ ).  
454 Pearson's correlations between the EWAS betas from set 1 and set 2 across all CpGs were  
455  $r=0.02$  (95% C.I.=0-0.102),  $r=0.04$  (95% C.I.=0-0.122), and  $r=0.03$  (95% C.I.=0-0.112) for  
456 cerebral white matter, total grey matter, and whole brain volume, respectively. When  
457 restricting CpGs to those with a nominal p-value ( $\leq 0.05$ ), the beta correlations were slightly  
458 higher, although not strong:  $r=0.17$  (95% C.I.=0.089-0.249),  $r=0.18$  (95% C.I.=0.099-0.259),  
459 and  $r=0.22$  (95% C.I.=0.14-0.297) for cerebral white matter, total grey matter, and whole-  
460 brain volume, respectively. The low effect size correlations may be a further reflection of the  
461 small sample investigated here.

462 There are limitations to the current study. Firstly, we report DNAm changes in whole

463 blood, which may not be representative of brain phenotypes. However, two of the CpGs  
464 identified here, located on the 450K array, were strongly correlated with DNAm in four brain  
465 regions (53). Although previous studies have shown that there is considerable agreement  
466 between blood and brain (28), future studies should explore DNAm changes in the brain in  
467 post-mortem samples where possible to uncover biological mechanisms underpinning brain  
468 structure within the same tissue. Further, findings at baseline may indicate that some DNAm  
469 changes lie upstream of brain structural changes, although effect sizes for each CpG were  
470 small compared to previous concurrent EWAS of brain regions (18,19). In addition, we  
471 cannot test the direction of association between brain structural changes and DNAm. In  
472 future, studies may apply Mendelian Randomization to investigate whether DNAm may be  
473 on the causal path to brain structure alterations in brain health and disease. Finally, in the  
474 current study we focussed on global brain phenotypes to explore whether global brain-  
475 related changes, previously associated with psychiatric and neurological disorders, are  
476 associated with DNAm alterations. Previous evidence includes DNAm associations at both  
477 global and regional level (18), and it may be that DNAm may provide more insight into  
478 region-specific alterations in relation to brain health and disease.

479 In conclusion, we report an EWAS of global cortical brain volumes using DNAm data  
480 collected ~6 years prior to MRI data collection in 672 individuals and an EWAS meta-analysis  
481 of cortical brain volumes using DNAm measured concurrently to MRI data in 565 individuals,  
482 both part of a large, population-based cohort. Using baseline DNAm data, we find four CpGs  
483 significantly associated with cortical brain volumes ~6 years later, all of which are annotated  
484 to genes implicated in brain-related phenotypes. We did not find significant associations at  
485 the concurrent timepoint. Findings here should be interpreted with caution, and future  
486 studies should aim to determine further links between DNAm changes and brain structure  
487 and function, to highlight our understanding of this relationship in health and disease.

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507

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513 **References**

- 514 1. DeYoung CG, Hirsh JB, Shane MS, Papademetris X, Rajeevan N, Gray JR. Testing  
515 predictions from personality neuroscience. *Brain structure and the big five*. *Psychol*  
516 *Sci a J Am Psychol Soc / APS* [Internet]. 2010 [cited 2020 Nov 9];21(6):820–8.  
517 Available from: /pmc/articles/PMC3049165/?report=abstract
- 518 2. Zatorre RJ, Fields RD, Johansen-Berg H. Plasticity in gray and white: Neuroimaging  
519 changes in brain structure during learning [Internet]. Vol. 15, *Nature Neuroscience*.  
520 Europe PMC Funders; 2012 [cited 2020 Nov 9]. p. 528–36. Available from:  
521 /pmc/articles/PMC3660656/?report=abstract
- 522 3. Shen X, Reus LM, Cox SR, Adams MJ, Liewald DC, Bastin ME, et al. Subcortical volume  
523 and white matter integrity abnormalities in major depressive disorder: Findings from  
524 UK Biobank imaging data. *Sci Rep* [Internet]. 2017 Dec 1 [cited 2020 Nov 9];7(1).  
525 Available from: /pmc/articles/PMC5514104/?report=abstract
- 526 4. Höistad M, Segal D, Takahashi N, Sakurai T, Buxbaum JD, Hof PR. Linking white and  
527 grey matter in schizophrenia: Oligodendrocyte and neuron pathology in the  
528 prefrontal cortex [Internet]. Vol. 3, *Frontiers in Neuroanatomy*. Frontiers Media SA;  
529 2009 [cited 2020 Nov 9]. Available from:  
530 /pmc/articles/PMC2713751/?report=abstract
- 531 5. Favre P, Pauling M, Stout J, Hozer F, Sarrazin S, Abé C, et al. Widespread white matter  
532 microstructural abnormalities in bipolar disorder: evidence from mega- and meta-  
533 analyses across 3033 individuals. *Neuropsychopharmacology* [Internet]. 2019 Dec 21  
534 [cited 2020 Nov 9];44(13):2285–93. Available from:  
535 <http://www.nature.com/articles/s41386-019-0485-6>
- 536 6. Mahmood A, Bibat G, Zhan AL, Izbudak I, Farage L, Horska A, et al. White matter  
537 impairment in rett syndrome: Diffusion tensor imaging study with clinical  
538 correlations. *Am J Neuroradiol* [Internet]. 2010 Feb [cited 2020 Nov 9];31(2):295–9.  
539 Available from: /pmc/articles/PMC2909097/?report=abstract
- 540 7. Serra L, Cercignani M, Lenzi D, Perri R, Fadda L, Caltagirone C, et al. Grey and white  
541 matter changes at different stages of Alzheimer’s disease. *J Alzheimer’s Dis*. 2010 Jan  
542 1;19(1):147–59.
- 543 8. Salat DH, Buckner RL, Snyder AZ, Greve DN, Desikan RSR, Busa E, et al. Thinning of the  
544 cerebral cortex in aging. *Cereb Cortex* [Internet]. 2004 Jul [cited 2020 Nov  
545 9];14(7):721–30. Available from: <https://pubmed.ncbi.nlm.nih.gov/15054051/>
- 546 9. Cox SR, Ritchie SJ, Tucker-Drob EM, Liewald DC, Hagenaars SP, Davies G, et al. Ageing  
547 and brain white matter structure in 3,513 UK Biobank participants. *Nat Commun*  
548 [Internet]. 2016 Dec 15 [cited 2020 Nov 9];7. Available from:  
549 /pmc/articles/PMC5172385/?report=abstract
- 550 10. Bird A. Perceptions of epigenetics. Vol. 447, *Nature*. Nature Publishing Group; 2007.  
551 p. 396–8.
- 552 11. C Greenberg M V, Bourchis D. The diverse roles of DNA methylation in mammalian  
553 development and disease. *Nat Rev Mol Cell Biol* [Internet]. [cited 2020 Nov 9];



- 554 Available from: [www.nature.com/nrm](http://www.nature.com/nrm)
- 555 12. Barbu MC, Shen X, Walker RM, Howard DM, Evans KL, Whalley HC, et al. Epigenetic  
556 prediction of major depressive disorder. *Mol Psychiatry* [Internet]. 2020 Jun 10 [cited  
557 2020 Jun 11];1–12. Available from: [http://www.nature.com/articles/s41380-020-](http://www.nature.com/articles/s41380-020-0808-3)  
558 0808-3
- 559 13. Jovanova OS, Nedeljkovic I, Spieler D, Walker RM, Liu C, Luciano M, et al. DNA  
560 methylation signatures of depressive symptoms in middle-aged and elderly persons:  
561 Meta-analysis of multiethnic epigenome-wide studies. *JAMA Psychiatry*.  
562 2018;75(9):949–59.
- 563 14. Hannon E, Dempster E, Viana J, Burrage J, Smith AR, Macdonald R, et al. An integrated  
564 genetic-epigenetic analysis of schizophrenia: Evidence for co-localization of genetic  
565 associations and differential DNA methylation. *Genome Biol* [Internet]. 2016 Aug 30  
566 [cited 2020 Nov 9];17(1):176. Available from:  
567 <https://genomebiology.biomedcentral.com/articles/10.1186/s13059-016-1041-x>
- 568 15. Montano C, Taub MA, Jaffe A, Briem E, Feinberg JI, Trygvadottir R, et al. Association  
569 of DNA methylation differences with schizophrenia in an epigenome-wide association  
570 study. *JAMA Psychiatry* [Internet]. 2016 May 1 [cited 2020 Nov 9];73(5):506–14.  
571 Available from: <https://jamanetwork.com/>
- 572 16. Horvath S, Raj K. DNA methylation-based biomarkers and the epigenetic clock theory  
573 of ageing. *Nat Rev Genet* [Internet]. 2018;19(6):371–84. Available from:  
574 <http://dx.doi.org/10.1038/s41576-018-0004-3>
- 575 17. Ladd-Acosta C, Pevsner J, Sabunciyan S, Yolken RH, Webster MJ, Dinkins T, et al. DNA  
576 methylation signatures within the human brain. *Am J Hum Genet* [Internet]. 2007 Dec  
577 1 [cited 2020 Nov 9];81(6):1304–15. Available from: [www.ajhg.org](http://www.ajhg.org)
- 578 18. Wheeler ENW, Stoye DQ, Cox SR, Wardlaw JM, Drake AJ, Bastin ME, et al. DNA  
579 methylation and brain structure and function across the life course: A systematic  
580 review. Vol. 113, *Neuroscience and Biobehavioral Reviews*. Elsevier Ltd; 2020. p. 133–  
581 56.
- 582 19. Jia T, Chu C, Liu Y, van Dongen J, Papastergios E, Armstrong NJ, et al. Epigenome-wide  
583 meta-analysis of blood DNA methylation and its association with subcortical volumes:  
584 findings from the ENIGMA Epigenetics Working Group. *Mol Psychiatry*. 2019;
- 585 20. Wockner LF, Noble EP, Lawford BR, Young RMD, Morris CP, Whitehall VLJ, et al.  
586 Genome-wide DNA methylation analysis of human brain tissue from schizophrenia  
587 patients. *Transl Psychiatry* [Internet]. 2014 Jan 7 [cited 2020 Nov 9];4(1):339.  
588 Available from: [www.nature.com/tp](http://www.nature.com/tp)
- 589 21. Liu J, Siyahhan Julnes P, Chen J, Ehrlich S, Walton E, Calhoun VD. The association of  
590 DNA methylation and brain volume in healthy individuals and schizophrenia patients.  
591 *Schizophr Res* [Internet]. 2015 Dec 1 [cited 2020 Nov 9];169(1–3):447–52. Available  
592 from: [/pmc/articles/PMC4681600/?report=abstract](http://pmc/articles/PMC4681600/?report=abstract)
- 593 22. Davis EG, Humphreys KL, McEwen LM, Sacchet MD, Camacho MC, Maclsaac JL, et al.  
594 Accelerated DNA methylation age in adolescent girls: associations with elevated

- 595 diurnal cortisol and reduced hippocampal volume. *Transl Psychiatry* [Internet]. 2017  
596 Aug 29 [cited 2020 Nov 9];7(8):e1223. Available from: [www.nature.com/tp](http://www.nature.com/tp)
- 597 23. De Jager PL, Srivastava G, Lunnon K, Burgess J, Schalkwyk LC, Yu L, et al. Alzheimer's  
598 disease: Early alterations in brain DNA methylation at ANK1, BIN1, RHBDF2 and other  
599 loci. *Nat Neurosci* [Internet]. 2014 Sep 1 [cited 2020 Nov 9];17(9):1156–63. Available  
600 from: [/pmc/articles/PMC4292795/?report=abstract](http://pmc/articles/PMC4292795/?report=abstract)
- 601 24. Masliah E, Dumaop W, Galasko D, Desplats P. Distinctive patterns of DNA methylation  
602 associated with Parkinson disease: Identification of concordant epigenetic changes in  
603 brain and peripheral blood leukocytes. *Epigenetics* [Internet]. 2013 [cited 2020 Nov  
604 9];8(10):1030–8. Available from: [/pmc/articles/PMC3891683/?report=abstract](http://pmc/articles/PMC3891683/?report=abstract)
- 605 25. van Erp TGM, Walton E, Hibar DP, Schmaal L, Jiang W, Glahn DC, et al. Cortical Brain  
606 Abnormalities in 4474 Individuals With Schizophrenia and 5098 Control Subjects via  
607 the Enhancing Neuro Imaging Genetics Through Meta Analysis (ENIGMA) Consortium.  
608 *Biol Psychiatry* [Internet]. 2018 Nov 1 [cited 2020 Nov 26];84(9):644–54. Available  
609 from: [/pmc/articles/PMC6177304/?report=abstract](http://pmc/articles/PMC6177304/?report=abstract)
- 610 26. Schmaal L, Pozzi E, C. Ho T, van Velzen LS, Veer IM, Opel N, et al. ENIGMA MDD:  
611 seven years of global neuroimaging studies of major depression through worldwide  
612 data sharing. Vol. 10, *Translational Psychiatry*. Springer Nature; 2020.
- 613 27. Wright IC, Rabe-Hesketh S, Woodruff PWR, David AS, Murray RM, Bullmore ET. Meta-  
614 analysis of regional brain volumes in schizophrenia. *Am J Psychiatry* [Internet]. 2000  
615 Jan 1 [cited 2020 Nov 26];157(1):16–25. Available from:  
616 <https://ajp.psychiatryonline.org/doi/abs/10.1176/ajp.157.1.16>
- 617 28. Walton E, Hass J, Liu J, Roffman JL, Bernardoni F, Roessner V, et al. Correspondence of  
618 DNA methylation between blood and brain tissue and its application to schizophrenia  
619 research. *Schizophr Bull* [Internet]. 2016 Mar 1 [cited 2020 Nov 9];42(2):406–14.  
620 Available from:  
621 <https://academic.oup.com/schizophreniabulletin/article/42/2/406/2518914>
- 622 29. Hansen K. *IlluminaHumanMethylationEPICanno.ilm10b2.hg19*: Annotation for  
623 Illumina's EPIC methylation arrays. R Packag version 060 [Internet]. 2016; Available  
624 from: [https://bitbucket.com/kasperdanielhansen/Illumina\\_EPIC](https://bitbucket.com/kasperdanielhansen/Illumina_EPIC)
- 625 30. Smith BH, Campbell H, Blackwood D, Connell J, Connor M, Deary IJ, et al. Generation  
626 Scotland: The Scottish Family Health Study; a new resource for researching genes and  
627 heritability. *BMC Med Genet* [Internet]. 2006 Oct 2 [cited 2020 Nov 9];7(1):74.  
628 Available from: [http://bmcmmedgenet.biomedcentral.com/articles/10.1186/1471-](http://bmcmmedgenet.biomedcentral.com/articles/10.1186/1471-2350-7-74)  
629 [2350-7-74](http://bmcmmedgenet.biomedcentral.com/articles/10.1186/1471-2350-7-74)
- 630 31. Smith BH, Campbell A, Linksted P, Fitzpatrick B, Jackson C, Kerr SM, et al. Cohort  
631 Profile : Generation Scotland : Scottish Family Health Study ( GS : SFHS ). The study ,  
632 its participants and their potential for genetic research on health and illness. *Int J*  
633 *Epidemiol*. 2013;(July 2012):689–700.
- 634 32. Navrady LB, Wolters MK, MacIntyre DJ, Clarke TK, Campbell AI, Murray AD, et al.  
635 Cohort profile: Stratifying Resilience and Depression Longitudinally (STRADL): A  
636 questionnaire follow-up of Generation Scotland: Scottish Family Health Study (GS:

- 637 SFHS). *Int J Epidemiol* [Internet]. 2018 Feb 1 [cited 2020 Nov 9];47(1):13-14g.  
638 Available from: <https://academic.oup.com/ije/article/47/1/13/3978732>
- 639 33. Habota T, Sandu A-L, Waiter GD, McNeil CJ, Steele JD, Macfarlane JA, et al. Cohort  
640 profile for the STRatifying Resilience and Depression Longitudinally (STRADL) study: A  
641 depression-focused investigation of Generation Scotland, using detailed clinical,  
642 cognitive, and neuroimaging assessments. *Wellcome Open Res* [Internet]. 2019 Nov  
643 25 [cited 2020 Nov 9];4:185. Available from:  
644 <https://doi.org/10.12688/wellcomeopenres.15538.1>  
645 <https://doi.org/10.12688/wellcomeopenres.15538.1>
- 646 34. Zugman A, Harrewijn A, Cardinale EM, Zwiebel H, Freitag GF, Werwath KE, et al.  
647 Mega-analysis methods in ENIGMA: The experience of the generalized anxiety  
648 disorder working group [Internet]. *Human Brain Mapping*. John Wiley and Sons Inc.;  
649 2020 [cited 2020 Nov 26]. Available from:  
650 <https://onlinelibrary.wiley.com/doi/full/10.1002/hbm.25096>
- 651 35. Leffondré K, Abrahamowicz M, Siemiatycki J, Rachet B. Modeling smoking history: A  
652 comparison of different approaches. *Am J Epidemiol* [Internet]. 2002 Nov 1 [cited  
653 2020 Nov 9];156(9):813–23. Available from:  
654 <https://pubmed.ncbi.nlm.nih.gov/12396999/>
- 655 36. First MB, Spitzer RL, Gibbon M, Williams JBW, Davies M, Borus J, et al. The structured  
656 clinical interview for DSM-III-R personality disorders (SCID-II). Part II: Multi-site test-  
657 retest reliability study. *J Pers Disord*. 1995 Feb 8;9(2):92–104.
- 658 37. Gigantesco A, Morosini P. Development, reliability and factor analysis of a self-  
659 administered questionnaire which originates from the World Health Organization's  
660 Composite International Diagnostic Interview - Short Form (CIDI-SF) for assessing  
661 mental disorders. *Clin Pract Epidemiol Ment Heal* [Internet]. 2008 Apr 10 [cited 2020  
662 Nov 9];4:8. Available from: [/pmc/articles/PMC2329624/?report=abstract](https://pubmed.ncbi.nlm.nih.gov/12396999/)
- 663 38. Amador C, Zeng Y, Barber M, Walker R, Campbell A, McIntosh AM, et al. Genome-  
664 wide methylation data improves dissection of the effect of smoking on body mass  
665 index. *bioRxiv* [Internet]. 2021 Apr 21 [cited 2021 Oct 12];2020.10.08.329672.  
666 Available from: <https://www.biorxiv.org/content/10.1101/2020.10.08.329672v2>
- 667 39. McCartney DL, Walker RM, Morris SW, McIntosh AM, Porteous DJ, Evans KL.  
668 Identification of polymorphic and off-target probe binding sites on the Illumina  
669 Infinium MethylationEPIC BeadChip. *Genomics Data* [Internet]. 2016 Sep 1 [cited  
670 2020 Nov 9];9:22–4. Available from: [/pmc/articles/PMC4909830/?report=abstract](https://pubmed.ncbi.nlm.nih.gov/12396999/)
- 671 40. Zeng Y, Amador C, Gao C, Walker RM, Morris SW, Campbell A, et al. Lifestyle and  
672 Genetic Factors Modify Parent-of-Origin Effects on the Human Methylome. *bioRxiv*  
673 [Internet]. 2021 Jun 30 [cited 2021 Oct 12];2021.06.28.450122. Available from:  
674 <https://www.biorxiv.org/content/10.1101/2021.06.28.450122v1>
- 675 41. Barbu M, Huider F, Campbell A, Amador C, Adams M, Lynall M, et al. Methylome-wide  
676 association study of antidepressant use in Generation Scotland and the Netherlands  
677 Twin Register implicates the innate immune system. *medRxiv* [Internet]. 2021 Oct 15  
678 [cited 2021 Oct 18];2020.10.06.20207621. Available from:

- 679 <https://www.medrxiv.org/content/10.1101/2020.10.06.20207621v2>
- 680 42. Xia C, Amador C, Huffman J, Trochet H, Campbell A, Porteous D, et al. Pedigree- and  
681 SNP-Associated Genetics and Recent Environment are the Major Contributors to  
682 Anthropometric and Cardiometabolic Trait Variation. *PLoS Genet* [Internet]. 2016 Feb  
683 1 [cited 2020 Nov 9];12(2):e1005804. Available from: [www.ed.ac.uk/](http://www.ed.ac.uk/)
- 684 43. Gadd DA, Hillary RF, McCartney DL, Zaghlool SB, Stevenson AJ, Nangle C, et al.  
685 Epigenetic scores for the circulating proteome replicate protein-disease predictions  
686 as tools for biomarker discovery. *bioRxiv* [Internet]. 2021 Apr 28 [cited 2021 Oct  
687 12];8:2020.12.01.404681. Available from:  
688 <https://www.biorxiv.org/content/10.1101/2020.12.01.404681v2>
- 689 44. Gadd DA, Hillary RF, McCartney DL, Shi L, McGeachan RI, Stolicyn A, et al. Epigenome  
690 and phenome study reveals circulating markers pertinent to brain health. *medRxiv*  
691 [Internet]. 2021 Sep 6 [cited 2021 Oct 12];2021.09.03.21263066. Available from:  
692 <https://www.medrxiv.org/content/10.1101/2021.09.03.21263066v1>
- 693 45. Min JL, Hemani G, Smith GD, Relton C, Suderman M. Meffil: Efficient normalization  
694 and analysis of very large DNA methylation datasets. *Bioinformatics* [Internet]. 2018  
695 Dec 1 [cited 2021 May 31];34(23):3983–9. Available from:  
696 <https://academic.oup.com/bioinformatics/article/34/23/3983/5042224>
- 697 46. Fortin J-P, Fertig E, Hansen K. shinyMethyl: interactive quality control of Illumina 450k  
698 DNA methylation arrays in R. *F1000Research* [Internet]. 2014 Jul 30 [cited 2020 Nov  
699 9];3:175. Available from: <https://doi.org/10.12688/f1000research.4680.1>
- 700 47. Du P, Kibbe WA, Lin SM. lumi: A pipeline for processing Illumina microarray.  
701 *Bioinformatics* [Internet]. 2008 Jul [cited 2020 Nov 9];24(13):1547–8. Available from:  
702 <https://pubmed.ncbi.nlm.nih.gov/18467348/>
- 703 48. Ritchie ME, Phipson B, Wu D, Hu Y, Law CW, Shi W, et al. Limma powers differential  
704 expression analyses for RNA-sequencing and microarray studies. *Nucleic Acids Res*  
705 [Internet]. 2015 Jan 6 [cited 2020 Nov 9];43(7):e47. Available from:  
706 [/pmc/articles/PMC4402510/?report=abstract](http://pmc/articles/PMC4402510/?report=abstract)
- 707 49. McCartney DL, Hillary RF, Stevenson AJ, Ritchie SJ, Walker RM, Zhang Q, et al.  
708 Epigenetic prediction of complex traits and death. *Genome Biol*. 2018;19(1):136.
- 709 50. Mendelson MM, Marioni RE, Joehanes R, Liu C, Hedman ÅK, Aslibekyan S, et al.  
710 Association of Body Mass Index with DNA Methylation and Gene Expression in Blood  
711 Cells and Relations to Cardiometabolic Disease : A Mendelian Randomization  
712 Approach. *PLOS Med*. 2017;1–30.
- 713 51. Liu C, Marioni RE, Hedman AK, Pfeiffer L, Tsai PC, Reynolds LM, et al. A DNA  
714 methylation biomarker of alcohol consumption. *Mol Psychiatry* [Internet]. 2018 Feb 1  
715 [cited 2021 Mar 29];23(2):422–33. Available from: [www.nature.com/mp](http://www.nature.com/mp)
- 716 52. Joehanes R, Just AC, Marioni RE, Pilling LC, Reynolds LM, Mandaviya PR, et al.  
717 Epigenetic Signatures of Cigarette Smoking. *Circ Cardiovasc Genet* [Internet]. 2016  
718 Oct 1 [cited 2021 Mar 29];9(5):436–47. Available from:  
719 <http://circgenetics.ahajournals.org/lookup/suppl/doi:10.1161/CIRCGENETICS.116.001>

720 506/-/DC1.

721 53. Hannon E, Lunnon K, Schalkwyk L, Mill J. Interindividual methylomic variation across  
722 blood, cortex, and cerebellum: Implications for epigenetic studies of neurological and  
723 neuropsychiatric phenotypes. *Epigenetics* [Internet]. 2015 [cited 2021 Jan  
724 22];10(11):1024–32. Available from: /pmc/articles/PMC4844197/?report=abstract

725 54. Hansen K. IlluminaHumanMethylation450kanno. ilmn12. hg19: annotation for  
726 Illumina’s 450k methylation arrays. 2016;

727 55. Willer CJ, Li Y, Abecasis GR. METAL: Fast and efficient meta-analysis of genomewide  
728 association scans. *Bioinformatics* [Internet]. 2010 Jul 8 [cited 2020 Nov  
729 9];26(17):2190–1. Available from: <https://pubmed.ncbi.nlm.nih.gov/20616382/>

730 56. Phipson B, Maksimovic J, Oshlack A. MissMethyl: An R package for analyzing data  
731 from Illumina’s HumanMethylation450 platform. *Bioinformatics* [Internet]. 2016 Jan  
732 15 [cited 2020 Nov 9];32(2):286–8. Available from:  
733 <https://pubmed.ncbi.nlm.nih.gov/26424855/>

734 57. Ren X, Kuan PF. methylGSA: a Bioconductor package and Shiny app for DNA  
735 methylation data length bias adjustment in gene set testing. *Bioinformatics*.  
736 2019;35(11):1958–9.

737 58. Dolgalev I. msigdb: MSigDB Gene Sets for Multiple Organisms in a Tidy Data Format.  
738 R package version 7.1.1. [Internet]. 2020 [cited 2020 Jul 29]. Available from:  
739 <https://cran.r-project.org/web/packages/msigdb/index.html>

740 59. Chu AY, Tin A, Schlosser P, Ko YA, Qiu C, Yao C, et al. Epigenome-wide association  
741 studies identify DNA methylation associated with kidney function. *Nat Commun*  
742 [Internet]. 2017 Dec 1 [cited 2020 Jul 29];8(1):1–12. Available from:  
743 [www.nature.com/naturecommunications](http://www.nature.com/naturecommunications)

744 60. H L, M Z, Z S, J L, S Z, D L, et al. Genome-wide haplotype association study identify the  
745 FGFR2 gene as a risk gene for acute myeloid leukemia. *Oncotarget* [Internet]. 2017  
746 [cited 2021 Sep 16];8(5):7891–9. Available from:  
747 <https://pubmed.ncbi.nlm.nih.gov/27903959/>

748 61. G K, G B, PR L, S G, K B, MK F, et al. Leveraging Polygenic Functional Enrichment to  
749 Improve GWAS Power. *Am J Hum Genet* [Internet]. 2019 Jan 3 [cited 2021 Sep  
750 16];104(1):65–75. Available from: <https://pubmed.ncbi.nlm.nih.gov/30595370/>

751 62. Hill WD, Marioni RE, Maghzian O, Ritchie SJ, Hagenaars SP, McIntosh AM, et al. A  
752 combined analysis of genetically correlated traits identifies 187 loci and a role for  
753 neurogenesis and myelination in intelligence. *Mol Psychiatry* [Internet].  
754 2019;24(2):169–81. Available from: <http://dx.doi.org/10.1038/s41380-017-0001-5>

755 63. JJ L, R W, A O, E K, O M, M Z, et al. Gene discovery and polygenic prediction from a  
756 genome-wide association study of educational attainment in 1.1 million individuals.  
757 *Nat Genet* [Internet]. 2018 Aug 1 [cited 2021 Sep 16];50(8):1112–21. Available from:  
758 <https://pubmed.ncbi.nlm.nih.gov/30038396/>

759 64. Liu M, Jiang Y, Wedow R, Li Y, Brazel DM, Chen F, et al. Association studies of up to  
760 1.2 million individuals yield new insights into the genetic etiology of tobacco and

- 761 alcohol use [Internet]. Vol. 51, Nature Genetics. Nature Publishing Group; 2019 [cited  
762 2020 Nov 9]. p. 237–44. Available from: /pmc/articles/PMC6358542/?report=abstract
- 763 65. Karlsson Linnér R, Biroli P, Kong E, Meddens SFW, Wedow R, Fontana MA, et al.  
764 Genome-wide association analyses of risk tolerance and risky behaviors in over 1  
765 million individuals identify hundreds of loci and shared genetic influences. Nat Genet  
766 [Internet]. 2019 Feb 1 [cited 2020 Nov 9];51(2):245–57. Available from:  
767 /pmc/articles/PMC6713272/?report=abstract
- 768 66. Shen J, Gilmore EC, Marshall CA, Haddadin M, Reynolds JJ, Eyaid W, et al. Mutations  
769 in PNKP cause microcephaly, seizures and defects in DNA repair. Nat Genet [Internet].  
770 2010 Mar [cited 2020 Nov 9];42(3):245–9. Available from:  
771 <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2835984/>
- 772 67. Lam M, Hill WD, Trampush JW, Yu J, Knowles E, Davies G, et al. Pleiotropic Meta-  
773 Analysis of Cognition, Education, and Schizophrenia Differentiates Roles of Early  
774 Neurodevelopmental and Adult Synaptic Pathways. Am J Hum Genet [Internet]. 2019  
775 Aug 1 [cited 2020 Nov 9];105(2):334–50. Available from:  
776 /pmc/articles/PMC6699140/?report=abstract
- 777 68. Liu D, Zhao L, Wang Z, Zhou X, Fan X, Li Y, Xu J, Hu S, Niu M, Song X LY. EWASdb:  
778 epigenome-wide association study database. Nucleic Acids Res. 2019;47:989–93.
- 779 69. Howard DM, Adams MJ, Clarke T-K, Hafferty JD, Gibson J, Shirali M, et al. Genome-  
780 wide meta-analysis of depression identifies 102 independent variants and highlights  
781 the importance of the prefrontal brain regions. bioRxiv. 2018.
- 782 70. Day FR, Ong KK, Perry JRB. Elucidating the genetic basis of social interaction and  
783 isolation. Nat Commun [Internet]. 2018 Dec 1 [cited 2020 Nov 10];9(1). Available  
784 from: <https://pubmed.ncbi.nlm.nih.gov/29970889/>
- 785 71. Nagel M, Jansen PR, Stringer S, Watanabe K, De Leeuw CA, Bryois J, et al. Meta-  
786 analysis of genome-wide association studies for neuroticism in 449,484 individuals  
787 identifies novel genetic loci and pathways. Nat Genet [Internet]. 2018 Jul 1 [cited  
788 2020 Nov 10];50(7):920–7. Available from:  
789 <https://pubmed.ncbi.nlm.nih.gov/29942085/>
- 790 72. BML B, R J, HF I, J van D, A A, MP van de W, et al. Multivariate genome-wide analyses  
791 of the well-being spectrum. Nat Genet [Internet]. 2019 Mar 1 [cited 2021 Sep  
792 16];51(3):445–51. Available from: <https://pubmed.ncbi.nlm.nih.gov/30643256/>
- 793 73. Meta-analysis of GWAS of over 16,000 individuals with autism spectrum disorder  
794 highlights a novel locus at 10q24.32 and a significant overlap with schizophrenia. Mol  
795 Autism [Internet]. 2017 [cited 2020 Nov 10];8:21. Available from:  
796 <https://pubmed.ncbi.nlm.nih.gov/28540026/>
- 797 74. Wang H, Yang J, Schneider JA, De Jager PL, Bennett DA, Zhang HY. Genome-wide  
798 interaction analysis of pathological hallmarks in Alzheimer’s disease. Neurobiol Aging  
799 [Internet]. 2020 Sep 1 [cited 2020 Nov 19];93:61–8. Available from:  
800 <https://pubmed.ncbi.nlm.nih.gov/32450446/>
- 801 75. Lee JJ, Wedow R, Okbay A, Kong E, Maghziyan O, Zacher M, et al. Gene discovery and

- 802 polygenic prediction from a genome-wide association study of educational  
803 attainment in 1.1 million individuals. *Nat Genet.* 2018;50(8):1112–21.
- 804 76. Jilani A, Ramotar D, Slack C, Ong C, Yang XM, Scherer SW, et al. Molecular cloning of  
805 the human gene, PNKP, encoding a polynucleotide kinase 3'-phosphatase and  
806 evidence for its role in repair of DNA strand breaks caused by oxidative damage. *J Biol*  
807 *Chem* [Internet]. 1999 Aug 20 [cited 2020 Nov 9];274(34):24176–86. Available from:  
808 <http://www.jbc.org/>
- 809 77. Gatti M, Magri S, Nanetti L, Sarto E, Di Bella D, Salsano E, et al. From congenital  
810 microcephaly to adult onset cerebellar ataxia: Distinct and overlapping phenotypes in  
811 patients with *PNKP* gene mutations. *Am J Med Genet Part A* [Internet]. 2019 Nov 22  
812 [cited 2020 Nov 9];179(11):2277–83. Available from:  
813 <https://onlinelibrary.wiley.com/doi/abs/10.1002/ajmg.a.61339>
- 814 78. Dumitrache LC, McKinnon PJ. Polynucleotide kinase-phosphatase (PNKP) mutations  
815 and neurologic disease. *Mech Ageing Dev* [Internet]. 2017 Jan 1 [cited 2021 Jul  
816 2];161(Pt A):121–9. Available from: </pmc/articles/PMC5161711/>
- 817 79. Shimada M, Dumitrache LC, Russell HR, McKinnon PJ. Polynucleotide kinase–  
818 phosphatase enables neurogenesis via multiple DNA repair pathways to maintain  
819 genome stability . *EMBO J* [Internet]. 2015 Oct [cited 2021 Jul 2];34(19):2465–80.  
820 Available from: </pmc/articles/PMC4601665/>
- 821 80. Spiers H, Hannon E, Schalkwyk LC, Smith R, Wong CCY, O'Donovan MC, et al.  
822 Methyloomic trajectories across human fetal brain development. *Genome Res*  
823 [Internet]. 2015 Mar 1 [cited 2020 Nov 9];25(3):338–52. Available from:  
824 <http://www.genome.org/cgi/doi/10.1101/gr.180273.114>.
- 825 81. Samaan G, Yugo D, Rajagopalan S, Wall J, Donnell R, Goldowitz D, et al. Foxn3 is  
826 essential for craniofacial development in mice and a putative candidate involved in  
827 human congenital craniofacial defects. *Biochem Biophys Res Commun.* 2010;400:60–  
828 5.
- 829 82. Yousefi P, Huen K, Davé V, Barcellos L, Eskenazi B, Holland N. Sex differences in DNA  
830 methylation assessed by 450K BeadChip in newborns. *BMC Genomics* [Internet]. 2015  
831 Nov 9 [cited 2020 Nov 9];16(1):911. Available from:  
832 <https://bmcbgenomics.biomedcentral.com/articles/10.1186/s12864-015-2034-y>
- 833 83. Moessner R, Marshall CR, Sutcliffe JS, Skaug J, Pinto D, Vincent J, et al. Contribution of  
834 SHANK3 mutations to autism spectrum disorder. *Am J Hum Genet* [Internet]. 2007  
835 Dec 1 [cited 2020 Nov 9];81(6):1289–97. Available from: [www.ajhg.org](http://www.ajhg.org)
- 836 84. Mancuso L, Fornito A, Costa T, Ficco L, Liloia D, Manuello J, et al. A meta-analytic  
837 approach to mapping co-occurrent grey matter volume increases and decreases in  
838 psychiatric disorders. *Neuroimage.* 2020 Nov 15;222:117220.
- 839 85. Clark SL, Hattab MW, Chan RF, Shabalin AA, Han LKM, Zhao M, et al. A methylation  
840 study of long-term depression risk. *Mol Psychiatry* [Internet]. 2020 Jun 1 [cited 2020  
841 Nov 9];25(6):1334–43. Available from: </pmc/articles/PMC7061076/?report=abstract>
- 842