PROTOZOOLOGY - ORIGINAL PAPER



Seroprevalence of *Toxoplasma gondii* and associated risk factors in domestic pigs raised from Cuba

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Abstract

A cross-sectional study was carried out to determine the seroprevalence of *Toxoplasma gondii* and associated risk factors in pigs in the largest pork-producing region in Cuba. Serum samples from 420 pigs, including 210 sows and 210 post-weaning pigs, were tested for antibodies against *T. gondii* using a commercial indirect enzyme-linked immunosorbent assay. Anti-*T. gondii* antibodies were detected in 56 animals (13.3%, 95% CI: 10.1–16.6). A generalized estimating equations model revealed that the risk factors associated with higher seropositivity in pigs were altitude (higher in farm's location < 250 m above sea level (masl) versus \geq 250 masl) and age (higher in sows compared to post-weaning pigs). The results indicated that this protozoan parasite is widely distributed on pig farms in the study area, which is a public health concern since the consumption of raw or undercooked pork meat products containing tissue cysts is considered one of the main routes of *T. gondii* transmission worldwide. Control measures should be implemented to reduce the risk of exposure to *T. gondii* in pigs in Cuba.

Keywords Toxoplasma gondii · Seroprevalence · Domestic pigs · Public health · Cuba

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Introduction

Toxoplasmosis is a worldwide zoonotic disease caused by the obligate intracellular protozoan parasite, *Toxoplasma gondii*, which infects virtually all warm-blooded species including human beings (Dubey et al. 2020). Approximately one-third of the human population is considered to be infected by this protozoan parasite (Behnke et al. 2020). Although *T. gondii* infection is usually asymptomatic, it can cause abortion, as well as blindness, neuromuscular disease, and even death in immunocompromised people (Dubey 2010). Moreover, an association between toxoplasmosis and neuropsychiatric disorders, including schizophrenia, has been suggested (Flegr and Horáček 2017).

Toxoplasma gondii is an important food-borne pathogen (EFSA 2018). The consumption of raw or undercooked meat products containing tissue cysts is considered one of the main routes of *T. gondii* transmission worldwide (Dubey 2010; Almeria and Dubey 2021). In this respect, pork is one of the major sources of human toxoplasmosis in some countries (Dubey et al. 2020; Almeria and Dubey 2021). Public concerns associated with *T. gondii* clearly indicate the need for epidemiological investigation in animals that can be used as a source of food. Previous immunological studies



on the presence of T. gondii antibodies in domestic pigs carried out in Latin America showed wide variations in the seroprevalence among countries and between regions within the same country (Cañón-Franco et al. 2014; Feitosa et al. 2014; Foroutan et al. 2019; Dubey et al. 2020). In Cuba, anti-T. gondii antibodies have been found in human patients with acquired immunodeficiency syndrome (Alfonso et al. 2009) and retinochoroiditis (Regalado Andújar et al. 2013), in pregnant women (González-Morales et al. 1995), neonates (Goya Batista et al. 2014), and blood donors (Sánchez-Artigas et al. 2009). Seropositivity has also been reported in domestic animals, including cats (Grandía et al. 2012) and dogs (Navarrete et al. 2017) in this country. Nevertheless, information regarding T. gondii in domestic pigs in Cuba is very scarce. The only previous survey in this species was carried out in Ciego de Ávila province (Central Cuba) between 1980 and 2002 (Suárez-Hernández et al. 2005). Hence, the aim of this study was to assess the current seroprevalence and risk factors associated with *T. gondii* in pigs in the largest pork-producing region in Cuba.

Material and methods

Study design

A cross-sectional study was carried out to determine seroprevalence against *T. gondii* in domestic pigs in Villa Clara (Central Cuba) (Fig. 1). This region accounts for the highest number of domestic pigs and is the largest pork producer in Cuba, with annual production at around 49,332 tons (ONEI 2017). Pork production in Cuba is characterized by an agreement management system. This means that all breeding farms, including sows, reproductive males, and piglets (from post-farrowing to post-weaning), are managed by the State government in intensive production systems. Fattening

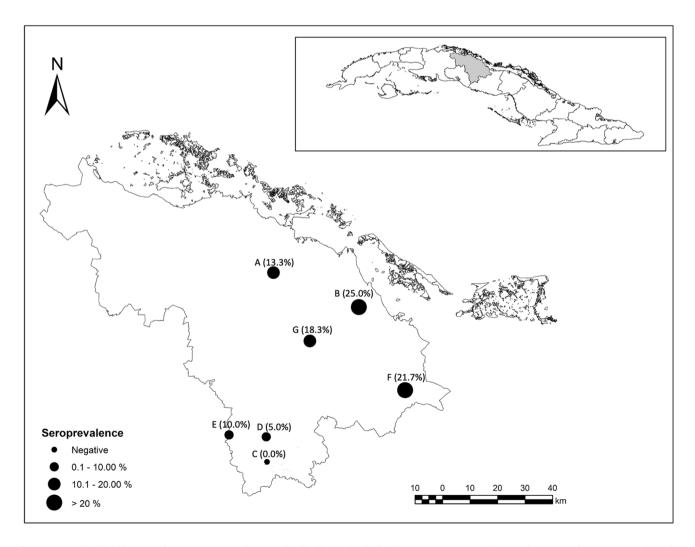


Fig. 1 Map of Villa Clara province (Cuba) showing the distribution and within-farm seroprevalence of *Toxoplasma gondii* on the sampled pig farms



is then carried out by private swine farmers until the pigs are ready for slaughter.

The total population of all breeding farms in the study area was used to calculate sampling size. The size of the sampled farms ranged from 500 to 1,600 sows (mean: 1,050 sows). The breeds of the sows and post-weaning pigs were Yorkshire/Landrace and Yorkshire/Landrace X CC21 (Cuban paternal breed), respectively. Sample size was calculated assuming a prevalence of 50% (which provides the highest sample size in studies with unknown prevalence) with a 95% confidence level (95% CI) and desired precision of $\pm 5\%$, resulting in 384 domestic pigs to be sampled (Thrusfield et al. 2018). Sixty animals, including 30 sows and 30 post-weaning pigs, were randomly selected from each pig farm in order to detect infection with 95% probability and assuming a minimum within-farm prevalence of 6%. A total of 420 pigs were finally sampled in seven (A-G) farms managed under governmental intensive production management. All sampled farms presented very similar biosecurity measures including self-replacement gilts (replacement of breeding sows using gilts from the same herd), all-in-all out management, absence of cats, absence of other animal species, perimetral fence around the farm, rodent and insect control, disinfection and disinfestations protocols, sanitary ford, and water chlorination, among others. Farms A, B, F, and G were located < 250 m above sea level (masl), while farms C, D, and E were located at altitudes \geq 250 masl.

Sample collection and serological analysis

The collection of samples analyzed in the present study was part of the official Animal Health Campaigns under Cuban legislation. No animals were specifically sampled for this study; therefore, no ethical approval was necessary. Blood samples (about 10 ml) were collected using the orbital sinus puncture method. Samples were then centrifuged at 4,800 rpm for 10 min. Serum samples were separated and stored at – 20 °C until analysis. To obtain the presence of the antibodies against *T. gondii*, serum samples were analyzed using a commercial indirect ELISA (PrioCHECK® *Toxoplasma* Ab porcine, Thermo Fisher Scientific Prionics Lelystad BV) in accordance with the manufacturer's recommendations (Castillo-Cuenca et al. 2020). The sensitivity and specificity of this ELISA according to the manufacturer are 98% and 99.6%, respectively.

Statistical analysis

Individual seroprevalence against *T. gondii* was calculated as the ratio of seropositive animals to the total number of animals examined, using two-sided exact binomial confidence intervals (95% CI). Analysis of means (ANOM) applied to proportions was used to identify farms with a significantly

different within-farm seroprevalence relative to the overall mean combining all the sampled farms ("grand mean"), enabling detection of groups that deviate significantly from the overall mean (Rao 2005). The analysis was performed using the "ANOM" package (Pallmann and Hothorn 2016) of the statistical software R (R v. 3.5.2). If a statistically significant difference between the farms was found by ANOM, a *dummy* variable was created ("significantly different farm" vs "other farms") and included in the bivariate analysis.

Epidemiological information including age, sex, farm (from A to G), altitude, and farm size was gathered for each sampled animal. For sows, data on offspring per birth, number of parities, weaning piglets, and stillbirths were also recorded. Bivariate chi-square and Fisher's exact tests were performed to obtain the statistical significance of independent variables regarding individual T. gondii status (dependent variable). Variables with P < 0.20 in the bivariate analysis were selected as potential risk factors. Collinearity between pairs of variables was tested by Cramer's V coefficient. Finally, a generalized estimating equation (GEE) was carried out to study the effect of the variables selected on the basis of bivariate analysis (Thrusfield et al. 2018). The number of seropositive animals was assumed to follow a binomial distribution and the "farm" was included as random effect. A logarithmic link function was considered. A forward introduction of variables was used, starting with the variable with the lowest Pvalue in bivariate analysis. At each step, the confounding effect of the included variable was assessed by computing the change in the odds ratios (ORs). The model was re-run until all remaining variables presented statistically significant values (likelihood-ratio Wald's test, P < 0.05) and a potential relationship with the response variable existed. The choice of the best model was based on the quasi-likelihood under independence model criterion (Hanley et al. 2003). Statistical analyses were performed using SPSS v25.0 software (Statistical Package for Social Sciences, Inc., Chicago, IL, USA).

Results

Antibodies against *T. gondii* were detected in 56 of 420 pigs tested (13.3%, 95% CI: 10.1–16.6). Seropositivity was found in six of the seven (85.7%) tested farms, and the withinfarm seroprevalence ranged between 5.0 and 25%: with the highest seroprevalence observed in pigs from farm B and the lowest value in pigs from farm D had the lowest value. Interestingly, antibodies against *T. gondii* were not found in samples from farm C (Table 1) (Fig. 1). ANOM showed a significant lower seroprevalence on farm "C," which was negative to the presence of anti-*T. gondii* antibodies, in relation to the overall mean of the other farms tested (Fig. 2).



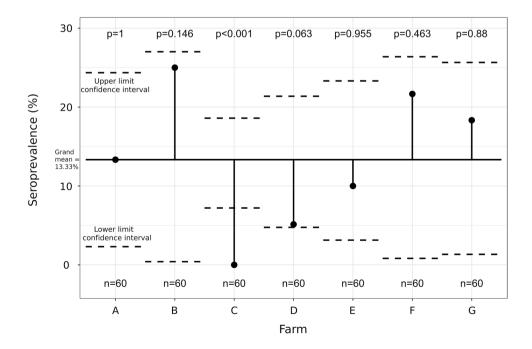
Table 1 Distribution of the prevalence of antibodies against *Toxoplasma gondii*, using ELISA, on pig farms in Villa Clara province (Cuba) by category. Variables with *P*-value < 0.20 in the bivariate

analysis were included in the multivariate analysis (generalized estimating equation) to determine potential risk factors

Variable	Categories	Number/overall (% positive)	OR	95% CI	Chi-square	P-value
Farm	A	8/60 (13.3)	NA		25.385	< 0.001
	В	15/60 (25.0)				
	C	0/60 (0.0)				
	D	3/60 (5.0)				
	E	6/60 (10.0)				
	F	13/60 (21.7)				
	G	11/60 (18.3)				
Altitude (m above sea level)	< 250	47/240 (19.6)	4.627	2.20-9.72	18.830	< 0.001
	≥250	9/180 (5.0)	*			
Farm size	>501	23/120 (19.2)	1.918	1.07-3.43	4.497	0.022
	< 500	33/300 (11.0)	*			
Age	Sows	46/210 (21.9)	5.610	2.75-11.5	26.703	< 0.001
	Post-weaning pigs	10/210 (4.8)	*			
Sex	Male	4/106 (3.8)	0.627	0.17-2.29	0.505	0.350
	Female	6/102 (5.9)	*			
Offspring per birth	<9	10/32 (31.3)	0.558	0.24-1.28	1.927	0.125
	≥10	36/178 (20.2)	*			
Parity number	<3	30/116 (25.9)	0.588	0.30-1.16	1.927	0.125
	≥4	16/94 (17.0)	*			
Weaning piglets	<9	10/47 (21.3)	1.05	0.48-2.31	0.014	0.541
	≥10	36/163 (22.1)	*			
Still birth	Yes	12/67 (17.9)	0.70	0.34-1.46	0.918	0.220
	No	34/143 (23.8)	*			

NA not applicable; *reference category

Fig. 2 Results of the analysis of means comparing seroprevalences on the sampled farms





No association between seropositivity to *T. gondii* and sex, offspring per weaning piglets, and still birth was found in the bivariate analysis. Farm, altitude, farm size, age, offspring per birth, and parity number were selected for the multivariate analysis (Table 1). The final GEE model showed that the main factors associated with *T. gondii* seropositivity in pigs in Cuba were altitude and age. The prevalence of *T. gondii* antibodies was significantly higher on farm located < 250 masl (19.6%; 95% CI: 14.6–24.6) compared to the farms located at altitude \geq 250 masl (5.0%; 95% CI: 1.8–8.2) (OR = 5.28; P = 0.001; 95% CI: 1.91–14.57). Significantly higher seropositivity was also found in sows (21.9%; 95% CI: 16.3–27.5) compared to post-weaning pigs (4.8%; 95% CI: 1.8–7.6) (OR = 6.05; P < 0.001; 95% CI: 2.53–14.60).

Discussion

Consumption of contaminated undercooked or raw meat from farm animals has been known to be a major risk factor for acquisition of *T. gondii* infection in humans, and among food livestock species, pork is considered one of the main sources of *T. gondii* infection (Almeria and Dubey 2021). The first key step to prevent transmission of this zoonotic parasite in the swine production is to determine the presence of the parasite in the farms. In this regard, serological surveillance is the most commonly used method tool for identifying *T. gondii* exposure in pigs.

The individual seroprevalence detected in pigs raised in Cuba in our study (13%) is of the same magnitude as found previously in Cuba (14%) and in other Latin American countries such as Brazil (13%), Colombia (15%), and Mexico (ranging between 13 and 17%) (Suárez-Hernández et al. 2005; Foroutan et al. 2019; Dubey et al. 2020). Slightly higher mean seroprevalence values were observed in Brazil (ranging between 20 and 26%), while higher seropositivity was found in Argentina (48%), Brazil (ranging between 33 and 52%), Costa Rica (44%), Hawaii (49%), Mexico (ranging between 45 and 97%), Panamá (32%), and Peru (30%) (Cañón-Franco et al. 2014; Foroutan et al. 2019; Dubey et al. 2020). In contrast, lower seroprevalence rates were detected in other studies in Brazil (ranging between 0 and 8%), Chile (9%), and Mexico (ranging between 1 and 9%) (Foroutan et al. 2019; Dubey et al. 2020). Even though statistically accurate comparisons cannot be made given the differences in number of animals tested, the population sampled, and/ or the different serological methods used, we would like to state that the seroprevalence in pigs in the study area should be considered moderate.

At least one seropositive pig was detected in six of the seven (85.7%) farms tested, with positive within-farm seroprevalence values ranging between 5.0 and 25.0%. Although

farm "C" was negative to the presence of anti-T. gondii antibodies, the number of samples collected in each farm was calculated assuming a minimum within-farm prevalence of 6% and therefore, the possibility of that particular farm having a seroprevalence lower than 5% could not be discounted. The results indicated that T. gondii infection is widespread on pig breeding farms in Cuba. Since the sampled farms were all managed under a very similar production system, the environmental characteristics may explain differences in the seroprevalences in pigs within the study region. In this regard, farms located < 250 masl showed significantly higher seropositivity compared to the raised at higher altitude. Our results are in agreement with those reported by Villari et al. (2009) who consider the higher altitude (> 200 masl) of the farms as a protective factor of T. gondii exposure; this observation is likely associated with a reduced environmental viability of oocysts with decreasing ambient temperature and, perhaps, also humidity. Higher seroprevalence levels were also found in wild boars (Sus scrofa) sampled in hunting states located < 600 masl compared to those sampled at higher altitude (Calero-Bernal et al. 2016). In contrast, other studies observed higher seropositivity to T. gondii in pigs raised in mountainous regions than those raised in lowlands (Alvarado-Esquivel et al. 2012; Papatsiros et al. 2016). The reason for these differences is unclear; however, environmental, and climatic conditions may impact survival of oocysts in soil, food, and water contaminated with feline feces (Gauss et al. 2006), which are the likely sources of infection for pigs. Further studies are needed to address this issue.

Significantly higher seropositivity was found in sows compared to post-weaning pigs. Age is an important factor affecting T. gondii seroprevalence in pigs because most animals acquire T. gondii infection postnatally (Dubey 2009). The higher prevalence of T. gondii antibodies in sows compared to post-weaning pigs is consistent with those previously reported (García-Bocanegra et al. 2010a; Hill et al. 2014; Djokic et al. 2016; Castillo-Cuenca et al. 2020) and probably reflects the cumulative likelihood of exposure to T. gondii and lifelong persistence of IgG antibodies. Maternalderived antibodies decline after the first week of age, but the decay is dependent on the antibody level of the dam at birth. However, because maternally transferred antibodies can persist until 4 months of age (Dubey 2009; García-Bocanegra et al. 2010b), the presence of maternally transferred antibodies detected in some seropositive post-weaning pigs cannot be ruled out.

Toxoplasmosis outbreaks have been reported in humans by ingestion of infected porcine meat containing tissue cysts (Choi et al. 1997; Vitale et al. 2014; Almeria and Dubey 2021). Even though we are not aware of any report of human toxoplasmosis directly linked to eating infected pork in Cuba, ocular toxoplasmosis (Mesa Hernández et al.



2011; Galbán Lueje et al. 2013; Bustillo et al. 2015; Ginorio Gavito et al. 2017), toxoplasmic encephalitis in patients with acquired immunodeficiency syndrome (Alfonso et al. 2009), acute toxoplasmosis in pregnant women (Lombana et al. 2012), and congenital toxoplasmosis (Amador Morán et al. 2016) have been reported in this country. Moreover, seropositivity values found in pregnant women (38%) (González-Morales et al. 1995), neonates (23%) (Goya Batista et al. 2014), and blood donors (13%) (Sánchez-Artigas et al. 2009) indicate that *T. gondii* is widely distributed in the human population in Cuba. Since viable *T. gondii* can be found in seropositive pigs, our results suggest that the consumption of non-properly cooked pork products may contribute to human toxoplasmosis in Cuba, and studies on the zoonotic impact of this disease are urgently needed in this country.

In summary, this is the first report on T. gondii in pigs in Villa Clara province, the largest pork-producing region in Cuba. Although the number of sampled farms was limited, the results obtained provide a first approximation to T. gondii exposure in domestic pigs in a country where there was no recent information in this animal species. The observed seropositivity indicates that this zoonotic parasite is widespread in pig breeding farms in the largest pork-producing region in Cuba. This finding indicates a public health concern because seropositive pigs can harbor tissue cysts in their meat, therefore representing a tentative zoonotic risk for consumers of raw or undercooked porcine meat or its products. In addition, evisceration and management of carcasses of infected pigs could also constitute a risk of infection for humans. Our results may contribute to the development of improved control strategies against T. gondii in this country. Further immunological and molecular studies on genotypes circulating in pig farms must be conducted to increase the knowledge of the epidemiology of T. gondii in Cuba.

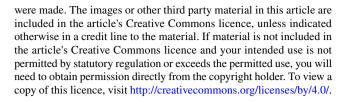
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Declarations

Conflict of interest The authors declare no competing interests.

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References

- Alfonso Y, Fraga J, Fonseca C et al (2009) Molecular diagnosis of *Toxoplasma gondii* infection in cerebrospinal fluid from AIDS patients. Cerebrospinal Fluid Res 6:2. https://doi.org/10.1186/1743-8454-6-2
- Almeria S, Dubey JP (2021) Foodborne transmission of *Toxoplasma* gondii infection in the last decade. An overview. Res Vet Sci 135:371–385. https://doi.org/10.1016/j.rvsc.2020.10.019
- Alvarado-Esquivel C, Estrada-Malacón MA, Reyes-Hernández SO et al (2012) High prevalence of *Toxoplasma gondii* antibodies in domestic pigs in Oaxaca State, Mexico. J Parasitol 98:1248–1250. https://doi.org/10.1645/GE-3184.1
- Amador Morán R, Coutos Ramos MJ, Peña Cedeño A et al (2016)
 Presentación de un caso con toxoplasmosis congénita. Rev Cuba
 Obstet y Ginecol 42. http://revginecobstetricia.sld.cu/index.php/gin/article/view/47/31. Accessed Feb 2021
- Behnke MS, Saeij JPJ, Boyle JP (2020) Development and application of classical genetics in *Toxoplasma gondii*. In: *Toxoplasma gondii*, Thirth. Wiess, L.M. and Kim, K. Academic Press, Elsevier, p 859–896. https://doi.org/10.1016/B978-0-12-815041-2. 00019-0
- Bustillo JL, Diaz JD, Pacheco IC, Gritz DC (2015) Cuban ocular toxoplasmosis epidemiology study (COTES): incidence and prevalence of ocular toxoplasmosis in Central Cuba. Br J Ophthalmol 99:382–386. https://doi.org/10.1136/bjophthalmol-2014-305843
- Calero-Bernal R, Pérez-Martín JE, Reina D et al (2016) Detection of zoonotic protozoa *Toxoplasma gondii* and *Sarcocystis suihominis* in wild boars from Spain. Zoonoses Public Health 63:346–350. https://doi.org/10.1111/zph.12243
- Cañón-Franco WA, López-Orozco N, Gómez-Marín JE, Dubey JP (2014) An overview of seventy years of research (1944–2014) on toxoplasmosis in Colombia. South America Parasit Vectors 7:427. https://doi.org/10.1186/1756-3305-7-427
- Castillo-Cuenca JC, Díaz-Cao JM, Martínez-Moreno Á et al (2020) Seroepidemiology of *Toxoplasma gondii* in extensively raised Iberian pigs in Spain. Prev Vet Med 175:104–854. https://doi.org/10.1016/j.parint.2010.06.001
- Choi W-Y, Nam H-W, Kwak N-H et al (1997) Foodborne outbreaks of human toxoplasmosis. J Infect Dis 175:1280–1282. https://doi.org/10.1086/593702
- Djokic V, Fablet C, Blaga R et al (2016) Factors associated with *Toxoplasma gondii* infection in confined farrow-to-finish pig herds in western France: an exploratory study in 60 herds. Parasit Vectors 9:466. https://doi.org/10.1186/s13071-016-1753-5
- Dubey JP (2010) Toxoplasmosis of animals and humans. Second. CRC Press Taylor & Francis Group, New York
- Dubey JP (2009) Toxoplasmosis in pigs—the last 20 years. Vet Parasitol 164:89–103. https://doi.org/10.1016/j.vetpar.2009.05.018
- Dubey JP, Cerqueira-Cézar CK, Murata FHA et al (2020) All about *Toxoplasma gondii* infections in pigs: 2009–2020. Vet Parasitol 288:109–185. https://doi.org/10.1016/j.vetpar.2020.109185
- EFSA (2018) Public health risks associated with food-borne parasites. EFSA J 16:e05495. https://doi.org/10.2903/j.efsa.2018.5495



- Feitosa TF, Vilela VLR, Bezerra de Melo LR et al (2014) *Toxoplasma gondii* and *Neospora caninum* in slaughtered pigs from Northeast, Brazil. Vet Parasitol 202:305–309. https://doi.org/10.1016/j.vet-par.2014.03.015
- Flegr J, Horáček J (2017) *Toxoplasma*-infected subjects report an Obsessive-compulsive disorder diagnosis more often and score higher in Obsessive-Compulsive Inventory. Eur Psychiatry 40:82–87. https://doi.org/10.1016/j.eurpsy.2016.09.001
- Foroutan M, Fakhri Y, Riahi SM et al (2019) The global seroprevalence of *Toxoplasma gondii* in pigs: a systematic review and metaanalysis. Vet Parasitol 269:42–52. https://doi.org/10.1016/j.vetpar. 2019.04.012
- Galbán Lueje T de las M, Lima León CE, Fariñas Falcón Z et al (2013) Caracterización de la toxoplasmosis ocular en pacientes de consulta externa. Acta Med 7:12–22. https://www.medigraphic.com/pdfs/medicadelcentro/mec-2013/mec134c.pdf. Accessed Oct 2020
- García-Bocanegra I, Simon-Grifé M, Dubey JP et al (2010a) Seroprevalence and risk factors associated with *Toxoplasma gondii* in domestic pigs from Spain. Parasitol Int 59:421–426. https://doi. org/10.1016/j.parint.2010.06.001
- García-Bocanegra I, Simon-Grifé M, Sibila M et al (2010b) Duration of maternally derived antibodies in *Toxoplasma gondii* naturally infected piglets. Vet Parasitol 170:134–136. https://doi.org/10.1016/j.vetpar.2010.01.042
- Gauss CBL, Dubey JP, Vidal D et al (2006) Prevalence of *Toxoplasma* gondii antibodies in red deer (*Cervus elaphus*) and other wild ruminants from Spain. Vet Parasitol 136:193–200. https://doi.org/10.1016/j.vetpar.2005.11.013
- Ginorio Gavito DE, Vilches Lescaille D, Gracial X et al (2017) Toxoplasmosis ocular: Algunos hallazagos clínicos y seroepidemiológicos. Rev AACC 7:4–13. http://revistaccuba.sld.cu/index.php/revacc/article/view/352/352. Accessed Nov 2020
- González-Morales T, Bacallo-Gallestey J, García-Santana CA, Molina-García JR (1995) Prevalencia de anticuerpos anti-*Toxoplasma gondii* en una población de mujeres embarazadas en Cuba. Gac Méd Méx 131:400. https://www.anmm.org.mx/bgmm/1864_2007/1995-131-5-6-499-503.pdf. Accessed Mar 2021
- Goya Batista Y, Sánchez Artigas R, Cobos Valdes D et al (2014) Determinación de anticuerpos IgG contra Toxoplasma gondii en neonatos de la Sala de Neonatología del Hospital General Universitario "Vladimir Ilich Lenin", Holguín. Rev Cuba Investig Biomed 33:12–18. http://scielo.sld.cu/pdf/ibi/v33n1/ibi02114.pdf. Accessed Oct 2020
- Grandía R, Entrena Á, Cruz J et al (2012) Seroprevalencia de *Toxo-*plasma gondii y factores de riesgo asociados en *Felis catus* en La
 Habana. Rev Salud Anim 34:201. http://scielo.sld.cu/scielo.php?
 script=sci_arttext&pid=S0253-570X2012000300013. Accessed
 Nov 2020
- Hanley JA, Negassa A, Edwardes MD, Forrester JE (2003) Statistical analysis of correlated data using generalized estimating equations: an orientation. Am J Epidemiol 157:364–375. https://doi.org/10.1093/aje/kwf215
- Hill DE, Dubey JP, Baroch JA et al (2014) Surveillance of feral swine for *Trichinella spp.* and *Toxoplasma gondii* in the USA and host-related factors associated with infection. Vet Parasitol 205:653–665. https://doi.org/10.1016/j.vetpar.2014.07.026

- Lombana RS, Cabrera MPC, Gavito DG et al (2012) Toxoplasmosis y embarazo. Rev Cuba Obstet Ginecol 38:99–106. http://scielo.sld.cu/pdf/gin/v38n1/gin12112.pdf. Accessed Nov 2020
- Mesa Hernández E, González Peña O, Padilla González C et al (2011) Comportamiento de la toxoplasmosis ocular activa en el Instituto Cubano de Oftalmología "Ramón Pando Ferrer". Rev Cuba Oftalmol 24:124–135. http://scielo.sld.cu/pdf/oft/v24n1/oft12111.pdf. Accessed Oct 2020
- Navarrete MG, Cordeiro MD, Batista Y et al (2017) Serological detection of *Toxoplasma gondii* in domestic dogs in the western region of Cuba. Vet Parasitol Reg Stud Reports 9:9–12. https://doi.org/10.1016/j.vprsr.2017.03.005
- ONEI (2017) Anuario Estadístico de la Provincia de Villa Clara. La Habana. http://www.onei.gob.cu/mapa/provincia/villa-clara. Accessed March 2021
- Pallmann P, Hothorn LA (2016) Analysis of means: a generalized approach using R. J Appl Stat 43:1541–1560. https://doi.org/10.1080/02664763.2015.1117584
- Papatsiros VG, Athanasiou LV, Stougiou D et al (2016) Cross-sectional serosurvey and risk factors associated with the presence of *Toxoplasma gondii* antibodies in pigs in Greece. Vector Borne Zoonotic Dis 16:48–53. https://doi.org/10.1089/vbz.2015.1845
- Rao CV (2005) Analysis of means—a review. J Qual Technol 37:308–315. https://doi.org/10.1080/00224065.2005.11980334
- Regalado Andújar B, Rodríguez Peña MS, Fraga Nodarse J, et al (2013) Aplicación de herramientas serológicas y moleculares para el diagnóstico de coriorretinitis por *Toxoplasma gondii*. Rev Cuba Med Trop 65:13–25. http://scielo.sld.cu/scielo.php?script=sci_ arttext&pid=S0375-07602013000100003. Accessed Feb 2021
- Sánchez-Artigas R, Góngora-Amores W, Torres-Ponce Z, Castañeda-Comerón B (2009) Seroprevalencia de anticuerpos IgG anti-Tox-oplasma gondii en donantes especiales de sangre en el municipio de Holguín. Ciencias Holguín 15:1–8. http://www.ciencias.holguin.cu/index.php/cienciasholguin/article/view/529. Accessed Feb 2021
- Suárez-Hernández M, González-Fernández A, Gardón-Quirola BY, Martínez-Sánchez R (2005) Infección y enfermedad por *Toxo*plasma gondii en animales y humanos en 23 años de observación en la provincia de Ciego de Ávila, Cuba. Rev Biomed 16:21– 27. https://www.medigraphic.com/cgi-bin/new/resumen.cgi? IDARTICULO=7396. Accessed Mar 2021
- Thrusfield M, Christley R, Brown H et al (2018) Veterinary epidemiology. Fourth. John Wiley & Sons, Oxford
- Villari S, Vesco G, Petersen E et al (2009) Risk factors for toxoplasmosis in pigs bred in Sicily, Southern Italy. Vet Parasitol 161:1–8. https://doi.org/10.1016/j.vetpar.2009.01.019
- Vitale M, Tumino G, Partanna S et al (2014) Impact of traditional practices on food safety: a case of acute toxoplasmosis related to the consumption of contaminated raw pork sausage in Italy. J Food Prot 77:643–646. https://doi.org/10.4315/0362-028X.JFP-13-285
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