

## Supplementary Information

### A Multi-proxy approach to exploring *Homo sapiens*' arrival, environments and adaptations in Southeast Asia

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## Material and Methods

**Context and nature of the fossiliferous deposits:** The fossil localities studied are located in the northern parts of Vietnam (Duoi U’Oi and Coc Muoi) and Laos (Tam Hang South, Nam Lot, and Tam Hay Marklot), in the same 23°-20° latitudinal belt (Bacon et al., 2008, 2015, 2018a; Bourgon et al., 2020) (Fig. S1). The landscape of this region contains typical tower karsts that emerge from the alluvial plain covered by cultivated fields and houses. All five karstic sites formed in massive limestone beds, Carboniferous to Triassic in age, and present the same type of sedimentary infillings in the caves and cavities.

The deposits consist of either well-cemented breccias plastered as relics on the walls and roofs of caves and cavities and/or silty to sandy clays located on the cave floor. The analysis of the sites suggests that the breccias were created principally by reworked carbonate clasts from the limestone massif, some speleothems and sandy to silty clays, along with fossil remains. These constituents were carried and deposited by water inside the cave network (mainly endokarstic processes mixed with variable exokarstic material) (Durringer et al., 2012). Fossiliferous deposits are most likely a result of a long transportation process of remains through the subterranean cave networks, over several thousands of years or more. The detailed description of sites can be found elsewhere in original publications.

**Conservation of assemblages and taphonomic biases:** The five assemblages contain mainly isolated teeth of a large array of mammals including Artiodactyla, Perissodactyla, Proboscidea, Carnivora, Primates and Rodentia (> 5 kg). Small-bodied microvertebrates are lacking. The analysis of these assemblages revealed that they share taphonomic pathways, due to the action of biotic (rodents and carnivores) and abiotic (water flows) agents, through comparable processes of deposition (Durringer et al., 2012).

The high percentages of teeth (> 70%) showing gnawing marks in all assemblages indicate the significant role of porcupines (*Hystrix* sp.), as the last accumulator agent of remains of carcasses at the sites, before being buried in the sediments and transported into the cave network. The differential preservation of lower *versus* upper teeth has been associated with the capacity of porcupines to collect all detachable and transportable remains of carcasses of animals of various body-sizes (~5 kg up to 5,000 kg), either mandibles for large-sized ungulates or cranial remains with maxillae for smaller ones (muntjacs), or complete jaws in some cases (wild pigs) (Bacon et al., 2015). Factors involved in the selection of remains by porcupines might be weight, size, and density of elements (Brain, 1981). Therefore, this capacity of porcupines to collect a wide range of animals > 5kg, means that assemblages are representative of the species diversity. Furthermore, as previously noticed by Brain (1981) for African sites “*It is evident that the minimum numbers of individuals animal represented by the porcupine collected remains do indeed mirror the actual abundance of the antelope species*”, it can be reasonably proposed that assemblages are also representative of the abundance of species at a local scale.

**Brief description of sites and dating:** The five mammal assemblages are bracketed by ages obtained from the dating of flowstones (Uranium-series) and/or cave sediments (red thermoluminescence [TL], single-grain optically stimulated luminescence [SG-OSL], and post-infrared infrared-stimulated luminescence [pIR-IRSL]). Only the Marklot fauna has been dated by direct dating on a few numbers of teeth (combined ESR/U-series) due to the lack of suitable

sediments to date. Therefore, these multiple and independent dating techniques provide a reliable age for the fauna assemblages. We present a brief description of sites and dating results from the oldest to the youngest:

**Coc Muoi cave, Northeast Vietnam:** The site is close to the Chinese border 155 km NNE from Hanoi in the Lang Son province. The site is a cave situated in a small isolated hill, 361 m above the sea level (Bacon et al., 2018a). The results of both the SG-OSL and pIR-IRSL dating of the sandy clays containing bones and teeth indicate that the sediments were deposited in the cave between 148 and 117 ka. The results of two U-series dating of the overlying flowstone, 114 and 108 ka, represent the minimum age for breccia and fossil accumulation (Fig. S2a).

**Tam Hang South rock shelter, Northeast Laos:** The site is a group of rock shelters situated along the wall of the Mountain of Pà Hang, in the Huà Pan province, at an elevation of 1120 m. The Mountain also includes other major Pleistocene to Holocene sites, Nam Lot cave, Tam Pà Ling, and Tam Hang Central (Bacon et al., 2015; Patole-Edoumba & Demeter, 2019). The fossiliferous breccias of Tam Hang South were located at the basal part of the Pà Hang cliff, plastered against the wall of the rock shelter. The sedimentary infilling consists of argillaceous-rich breccias separated by flowstones, the dating of which was based on different techniques: red TL, SG-OSL and U-series results provided an age range of 94 - 60 ka (Fig. S2b).

**Nam Lot cave, Northeast Laos:** The cave is located along the cliff of the Pà Hang Mountain, 250 m to the east of the Tam Hang site (Düringer et al., 2012; Bacon et al., 2015, 2018b). The fauna assemblage was derived from the breccia and from the silty to sandy clays in the lower part of the cave close to the entrance. Based on red TL, SG OSL and U-series dating, the age range for the assemblage is 86 - 72 ka (Bacon et al., 2015) (Fig. S3a).

**Duoi U'Oi cave, Northwest Vietnam:** The cave is located on the other side of the border from Laos, around 100 km away from the Pà Hang Mountain (Bacon et al., 2008, 2018b). It is a lowland site situated around ~5 m above the alluvial plain. The red TL and SG-OSL dating of the sedimentary breccia unit which contains the faunal assemblage, and the U-series dating of the calcitic floor that crosses the fossiliferous breccia, provide an age range of 70 – 60 ka (Bacon et al., 2015) (Fig. S3b). The age constraints on the fauna are also supported by additional radiocarbon dates on teeth, which are > 32 cal kBP (Wood et al. 2016, 2021).

**Tam Hay Marklot cave, Northeast Laos:** The cave is located in the Huà Pan province, northeastern Laos (Bourgon et al., 2020). Remains of the faunal assemblage have been found in a soft sandy to gravelly clays that covered almost the entire surface of the cave (Fig. S3c). Based on combined ESR and U-Th dating of five mammalian teeth, the age range of the fauna is 38.4 – 13.5 ka.

**Carbon stable isotopes:** The carbon isotopic composition of bioapatite reflects that of diet, whereas the oxygen mainly reflects ingested water (DeNiro & Epstein, 1978; Lee-Thorp et al., 1989; Sponheimer & Lee-Thorp, 1999; Cerling & Harris 1999; Sullivan & Krueger, 1981; Longinelli, 1984; Luz et al., 1984; Passey et al., 2005).

For the geographic area in this study, a  $\delta^{13}\text{C}$  range of -37 ‰ to -23 ‰ is considered to represent  $\text{C}_3$  plants, as higher values (> -23 ‰) are restricted to particular context and species (Smith & Epstein, 1971; O'Leary, 1988; Kohn, 2010).  $\text{C}_4$  plants present higher  $\delta^{13}\text{C}$  values of -17 ‰ to -10 ‰ (Smith and Epstein 1971; O'Leary 1988). The burning of fossil fuel over the past 150 years has decreased the  $\delta^{13}\text{C}$  of atmospheric  $\text{CO}_2$ , consequently this has influenced the  $\delta^{13}\text{C}$  of all living organisms. As the  $\delta^{13}\text{C}$  values of plants are based on modern samples, a correction must be applied (~ +1.7 ‰) (Friedli et al., 1986) when comparing with organisms

that predate the fossil-fuel-induced atmospheric CO<sub>2</sub> shift. Therefore, the upper  $\delta^{13}\text{C}$  limit for C<sub>3</sub> plants used for the present study has been shifted from -23 ‰ to -21.3 ‰, and the lower limit for C<sub>4</sub> plants from -17 ‰ to -15.3 ‰.

The composition of the fauna isotopic diet is then metabolized and incorporated into the tissues of animals (Lee-Thorp et al., 1989). However, trophic fractionation occurs when the food's carbon is incorporated into the tissues of animals (bone collagen, enamel carbonate, hairs, etc.) and also varies according to the animals (DeNiro & Epstein, 1978; Lee-Thorp et al., 1989; Cerling & Harris 1999; Passey et al., 2005). Although arguably little is known of the digestive physiologies of fossil species to accordingly adjust discrimination factors, some comparable modern data are nevertheless available (Cerling & Harris 1999; Balasse, 2002; Passey et al., 2005; Lee-Thorp & Van der Merwe, 1987; Bocherens, 2002; Bocherens et al., 2011; Cerling et al., 2004; Fox-Dobbs et al., 2006; Sponheimer et al., 2013). Furthermore, Tejada-Lara et al. (2018) recently established a relationship where body mass can be used to predict  $^{13}\text{C}$  isotope enrichment in mammals. As mentioned by the authors, a single systematical enrichment factor, rather than multiple ones, can alter interpretations of animal ecologies and thus potentially lead to less precisely reconstructed environments. In this paper, species-specific enrichment factors were calculated for each taxon according to mean body mass following the equations proposed by Tejada-Lara et al. (2018).

As explained in Tejada-Lara et al. (2018), the formulae use log transformed (ln) body mass. Additionally, in order to obtain the ‰ isotope enrichment value, the obtained diet-bioapatite ( $\epsilon^*$ ) needs to be inverted ( $e^x$ ). For all intent and purposes and intent, this results in the following formulae:

$$\text{General: } \epsilon^*_{\text{diet-bioapatite}} = e^{2.4 + 0.034 (\ln \text{BM})}$$

$$\text{Foregut fermenter: } \epsilon^*_{\text{diet-bioapatite}} = e^{2.34 + 0.05 (\ln \text{BM})}$$

$$\text{Hindgut fermenter: } \epsilon^*_{\text{diet-bioapatite}} = e^{2.42 + 0.032 (\ln \text{BM})}$$

The enrichment factor of the giant panda (*Ailuropoda melanoleuca*) was not calculated, as was Tejada-Lara et al. (2018) clearly demonstrated that the giant panda's calculated enrichment factor was incorrect, and also not suited for calculating enrichment factor of carnivores. As such, a  $\epsilon^*_{\text{diet-bioapatite}}$   $\delta^{13}\text{C}$  enrichment of +10.51 ‰, found in the literature (Han et al., 2016) was used for the giant panda (*Ailuropoda melanoleuca*). While calculating the enrichment factor is clearly not suited to carnivores, an average carnivore-herbivore enamel spacing of -1.3 ‰ was established (Clementz et al., 2009). This spacing can then be added to their  $\delta^{13}\text{C}_{\text{apatite}}$ , resulting in an averaged apatite value of their prey, which can be transformed using the general formulae from Tejada-Lara et al. (2018). This effectively obtains estimates of the  $\delta^{13}\text{C}$  values of the environment in which carnivores hunted at the time. Finally, no  $\epsilon^*_{\text{diet-bioapatite}}$   $\delta^{13}\text{C}$  enrichment are available for ursids and calculating enrichment factor was not suited due to their carnivore digestive physiology. Since ursids are not strict carnivore, obtaining  $\delta^{13}\text{C}_{\text{source}}$  values, in a similar way to carnivores as described above, was also not possible. As such, we used a  $\epsilon^*_{\text{diet-bioapatite}}$   $\delta^{13}\text{C}$  enrichment of 13.3 ‰ from pigs (Passey et al., 2005), as the closest and most suitable alternative for omnivorous taxa.

Such conversion accounts for differences in faunal assemblages between sites, thus ensuring better comparisons between taxa that can be relating directly to the environment. Finally, we use the notation " $\delta^{13}\text{C}_{\text{carbon source}}$ " instead of the commonly seen " $\delta^{13}\text{C}_{\text{diet}}$ " (Louys & Roberts, 2020; Tejada-Lara et al., 2020) as  $\delta^{13}\text{C}_{\text{apatite}}$  of carnivores were also tentatively converted to  $\delta^{13}\text{C}_{\text{carbon source}}$  values (*i.e.* plant  $\delta^{13}\text{C}$  values). The " $\delta^{13}\text{C}_{\text{carbon source}}$ " thus effectively reflects the initial carbon uptake derived from plants, a terrestrial food web's primary carbon

source, and not always (namely in the case of carnivores) the  $\delta^{13}\text{C}$  value of the consumer's diet. While  $\delta^{13}\text{C}_{\text{apatite}}$  values were converted to  $\delta^{13}\text{C}_{\text{carbon source}}$ , the  $\delta^{18}\text{O}_{\text{apatite}}$  values were not similarly converted to drinking water  $\delta^{18}\text{O}$  values because empirically determined water-enamel  $^{18}\text{O}$  fractionation (or closely related ones) is not determined for too many modern-day analogue species of the investigated fossil taxa (or closely related ones).

During every mass spectrometer run (15 runs in total for the three sites), an internal laboratory standard (Marble LM, accepted  $\delta^{13}\text{C} = +2.13\text{‰}$  and  $\delta^{18}\text{O} = -1.83\text{‰}$ ) was analysed that has been normalised to the International Atomic Energy Agency reference material (NBS 19). It was used for tooth sample correction and for controlling the precision of the mass spectrometer ( $1\sigma$  ( $\delta^{13}\text{C}$ ) = 0.031 ‰ and  $1\sigma$  ( $\delta^{18}\text{O}$ ) = 0.044 ‰ for the 15 runs). Each tooth sample was usually analysed one or two times except for 6 individuals from Coc Muoi and Duoi U'Oi that were analysed three times. The repeated analyses were used to test for intra-individual heterogeneity and analytical reproducibility of enamel analysis (Table S1). Maximum standard deviations were 0.136 ‰ and 0.165 ‰ for  $\delta^{13}\text{C}$  and  $\delta^{18}\text{O}$  enamel analysis, respectively.

**Palaeoproteomics, Methods:** Two extracts (13.4 mg and 12.0 mg, respectively) of the lower incisor (NL 433) were prepared and analyzed using an HCl-based demineralization protocol without using a digestive protease, as previously described (Cappellini et al., 2019). Raw data were searched against an enamel-specific proteome database including Hominoidea and *Macaca* entries for the following protein sequences: COL1A1, COL1A2, COL17A1, ALB, TUFT1, ODAM, DSPP, AMELX, AMELY, AMBN, ENAM, KLK4, MMP20, FAM20A, FAM20C, obtained from UniProt and Genbank. Missing isoforms (for AMELX, AMELY, AMBN, ENAM) and predicted protein sequences for ancestral nodes within Hominidae (for ALB, AMBN, AMELX, AMELY, AMTN, COL17A1, ENAM, MMP20) were added to this protein sequence database, as previously described (Welker et al., 2020), to minimize cross-species proteomic effects (Welker, 2018). MaxQuant (v. 1.6.3.4) settings mirrored those described in Welker et al. (2020), with the omission of histidine to aspartic acid (H>D) and histidine to hydroxyglutamate as variable modifications. Deamidation was calculated as described in Mackie et al. (2018), with values grouped at the protein-level.

To assign a biological identity to this specimen, we retained only those proteins with at least 2 unique peptides. Subsequently, we reconstructed the ancient protein sequence of the Nam Lot individual, obtained relevant SAPs, and created a heatmap of the amino acid distances between species pairs, including the Nam Lot individual (packages seqinr v. 3.6.1, (Charif & Lobry, 2007), and pheatmap v. 1.0.12, (Kolde, 2015), on the resulting variant matrix (restricted to ALB, AMELX, AMBN, ENAM and COL17A1). Simultaneously, the variant matrix was interrogated for sites in the Nam Lot individual that are uniquely present in the genus *Homo* or in the genus *Pongo*. The quality of spectra overlapping such positions was then manually validated. A similar procedure was used for the two reported AMELY-specific peptides. These did not represent high-confidence spectra, and as a result we do not assign the specimen to a biological sex.

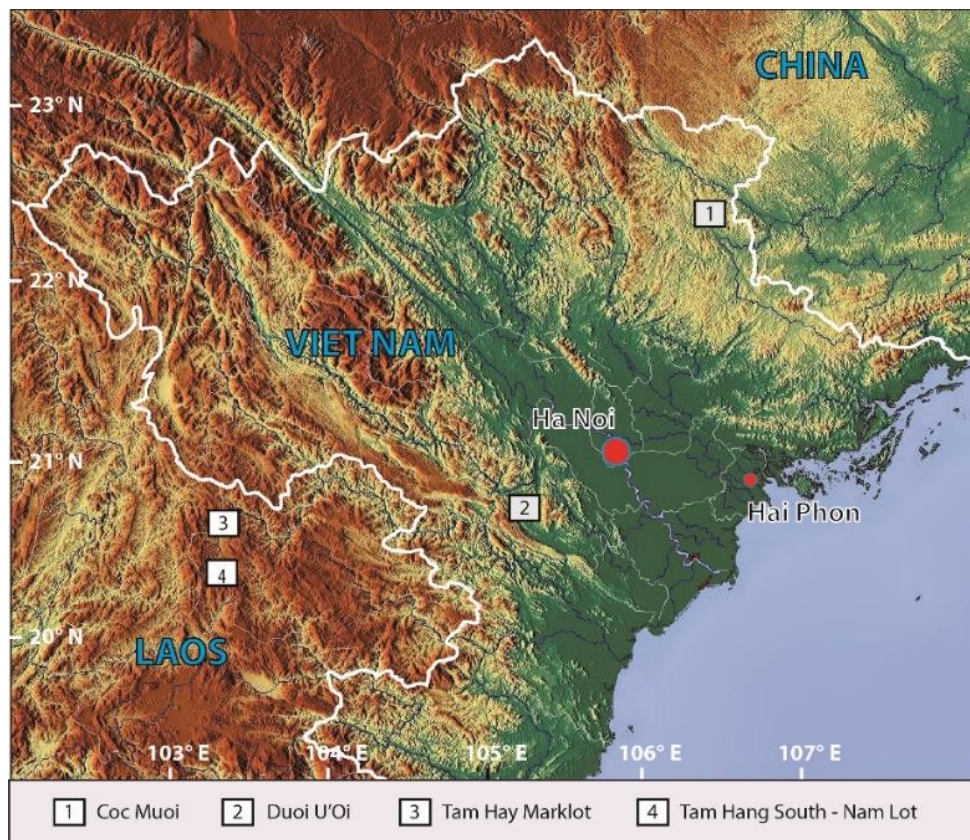
Proteomic data generated for the Nam Lot specimen has been deposited in ProteomeXchange under accession number PXD027426.

**Results:** Extraction of the Nam Lot 433 (NL 433) enamel incisor fragment resulted in the acquisition of 45,237 MS2 spectra combined over two independent LC-MS/MS injections. After quality filtering (see Methods), we identified 7 unique protein groups (AMELX, AMBN,

ENAM, MMP20, ALB, and COL17A1; Table S2), with AMELX represented by two isoforms. The Nam Lot enamel is therefore composed of the same core enamel proteome as other Pleistocene enamel proteomes (Cappellini et al., 2019; Welker et al., 2019, 2020). The deamidation of these proteins is higher than that observed for a small list of contaminants identified (Table S2), providing an indication that these proteins are likely to be of endogenous origin.

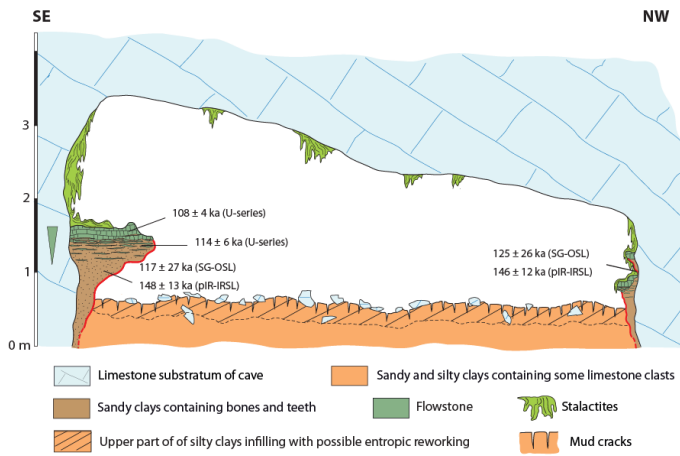
As with previous Pleistocene enamel proteomes, sequence coverage for AMTN and MMP20 is limited to small, uninformative sequence regions. We therefore restricted phylogenetic analysis to AMELX, AMBN, ENAM, ALB, and COL17A1. The reconstructed protein sequence of the NL 433 specimen assigns it unambiguously to the genus *Pongo* (Fig. S4a). All retained peptide matches are 100% matches to the *Pongo* sequences, with no unique matches to the genus *Homo*. In several proteins, the Nam Lot specimen provided protein sequences overlapping uniquely-derived single amino acid polymorphisms of the genus *Pongo* (Fig. S4b-d). For those stretches where we have protein sequence data for the ancient specimens, both species within the genus *Pongo*, *P. abelli* and *P. pygmaeus*, carry the same protein sequence. Therefore, we cannot assign the Nam Lot 433 specimen to a more specific orangutan species.

### Supplementary Figures

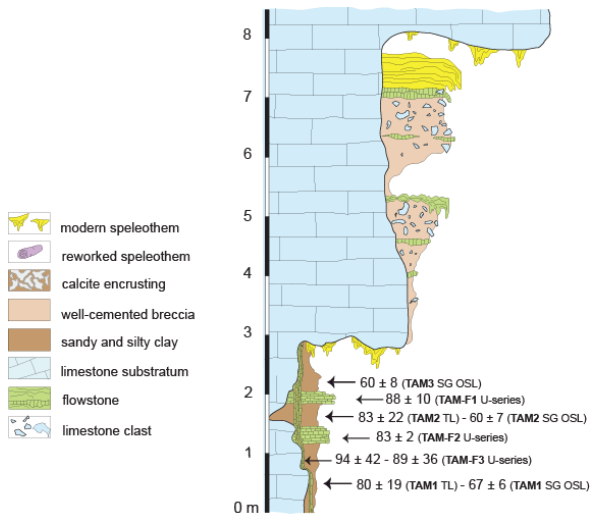


**Figure S1:** Satellite image of the studied area with the location of sites (Figure made by P. Düringer). The satellite image is from the website (<http://www.maps-for-free.com/>), and reworked using the software Illustrator CS5 (version 15.0.0).

**Figure S2: Sediments and datings.**

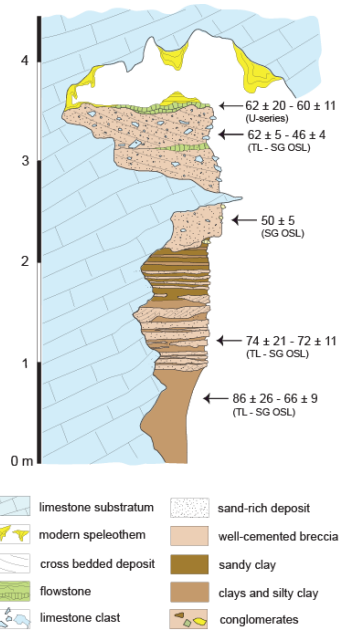


- a. Vertical section of the Coc Muoi cave, with results of U-series dating of the flowstones and luminescence dating (SG-OSL and pIR-IRSL) of the breccia unit (drawing of Philippe Durringer and Jean-Luc Ponche published in Bacon et al., 2018; Fig. 5).

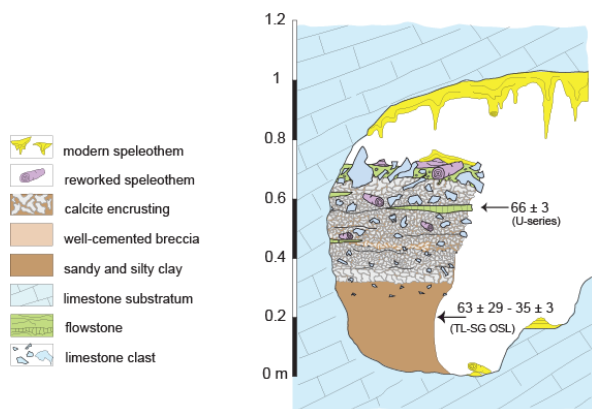


- b. Vertical section of the wall of the Tam Hang rock shelter, with results of U-series dating of the flowstones and luminescence dating (TL and SG-OSL) of the breccia unit (drawing of Philippe Durringer and Jean-Luc Ponche published in Bacon et al., 2015; Fig. 2).

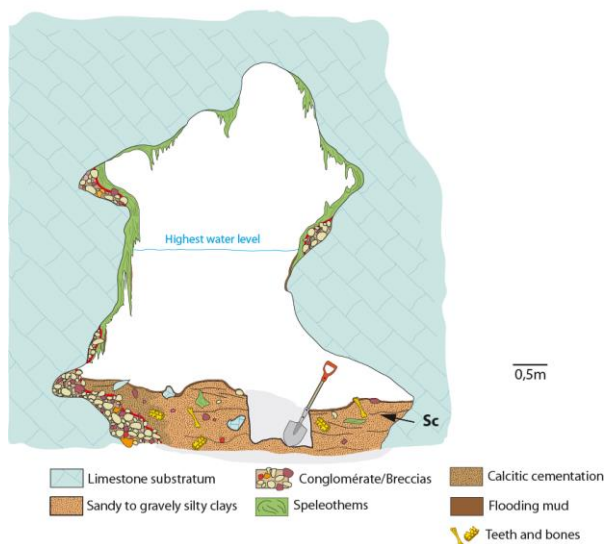
**Figure S3: Sediments and datings.**



a. Vertical section of the breccia unit of Nam Lot cave, with results of U-series dating of the flowstones and luminescence dating (TL and SG-OSL) of the breccia (drawing of Philippe Duringer and Jean-Luc Ponche published in Bacon et al., 2015; Fig. 3).

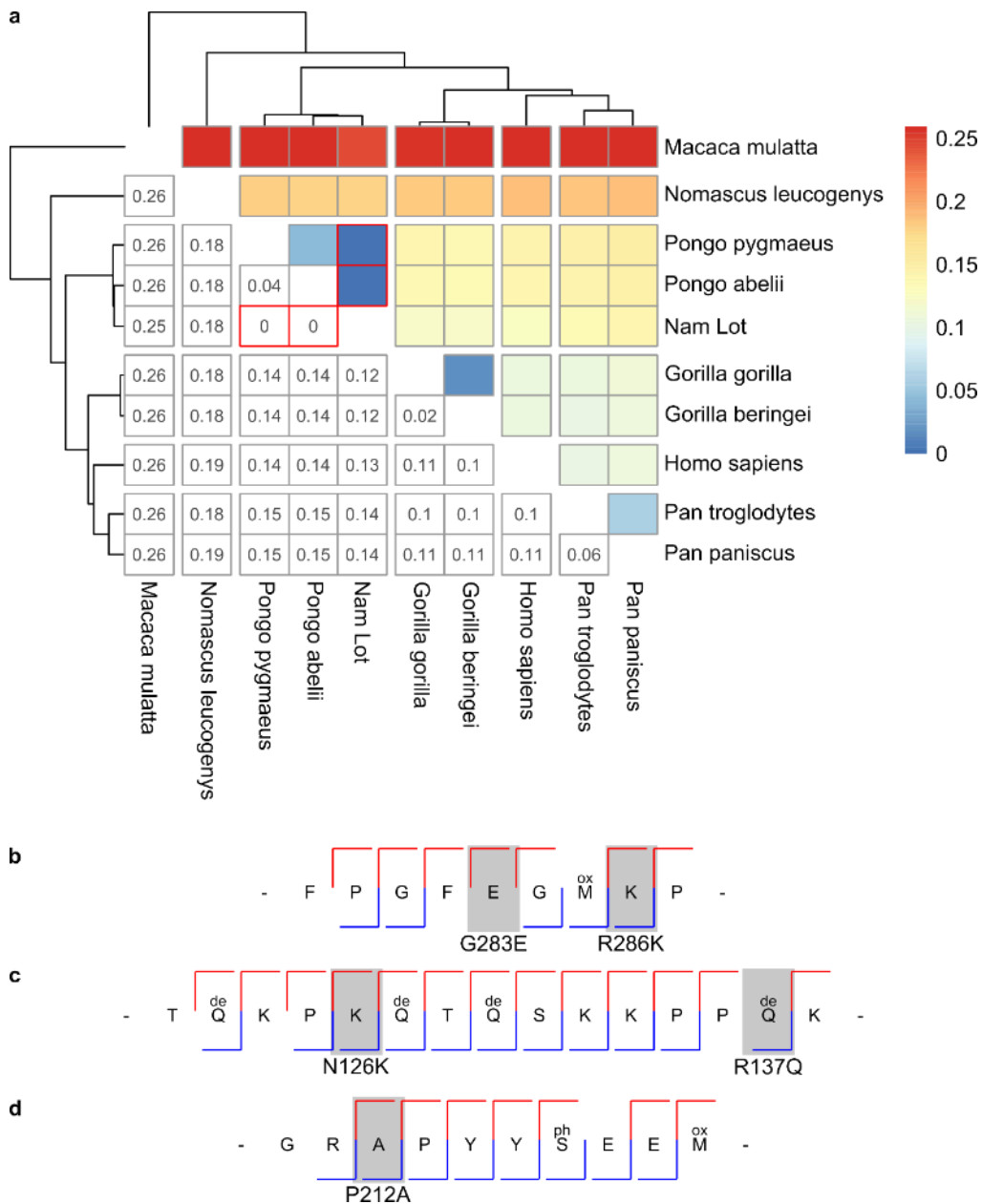


b. Vertical section of the breccia unit of Duoi U'Oi cave, with results of U-series dating of the flowstones and luminescence dating (TL and SG-OSL) of the breccia (drawing of Philippe Duringer and Jean-Luc Ponche published in Bacon et al., 2015; Fig. 4).

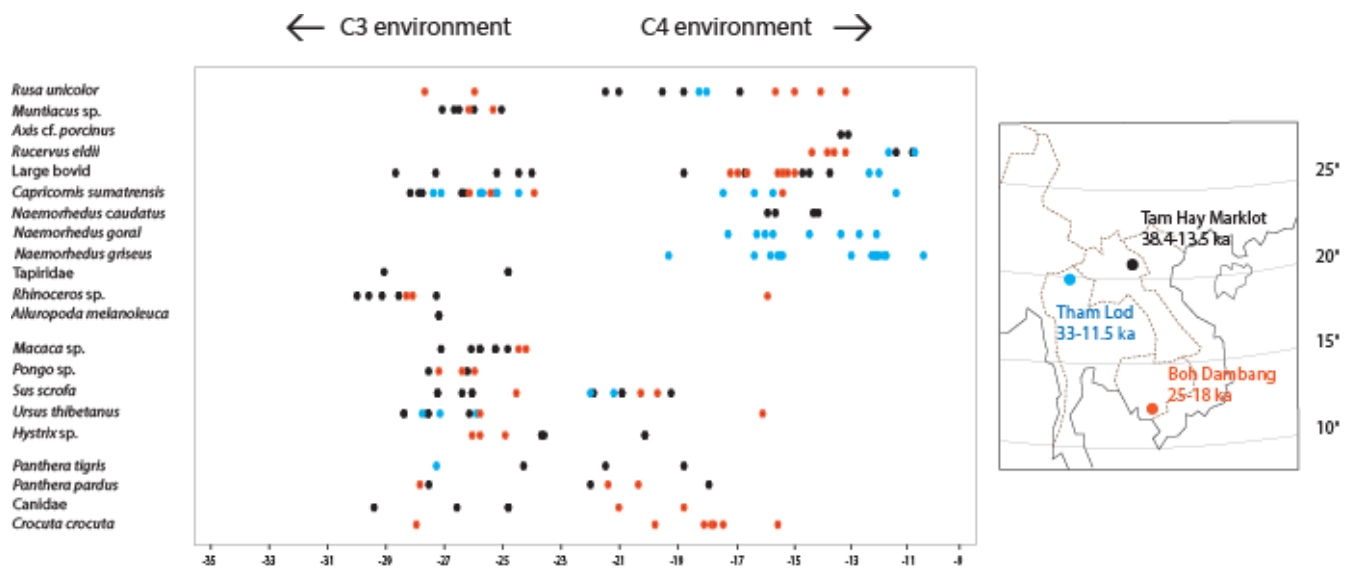


c. Section of the corridor in the Tam Hay Marklot cave. Specimens of the faunal assemblage have been found in the soft sandy to gravelly clays that covered almost the entire soil of the cave (drawing of Philippe Duringer and Jean-Luc Ponche published in Bourgon et al., 2020; Fig. S17). The age range of the fauna is 38.4 – 13.5 ka.

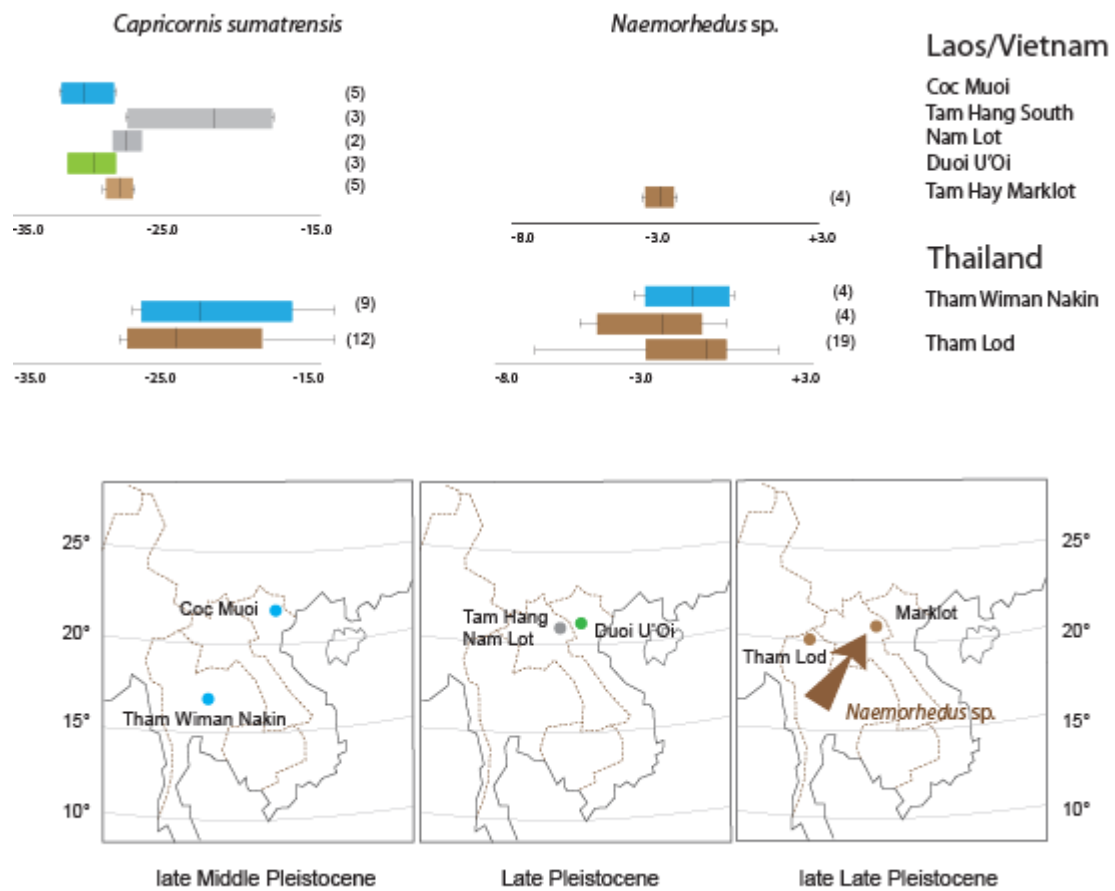




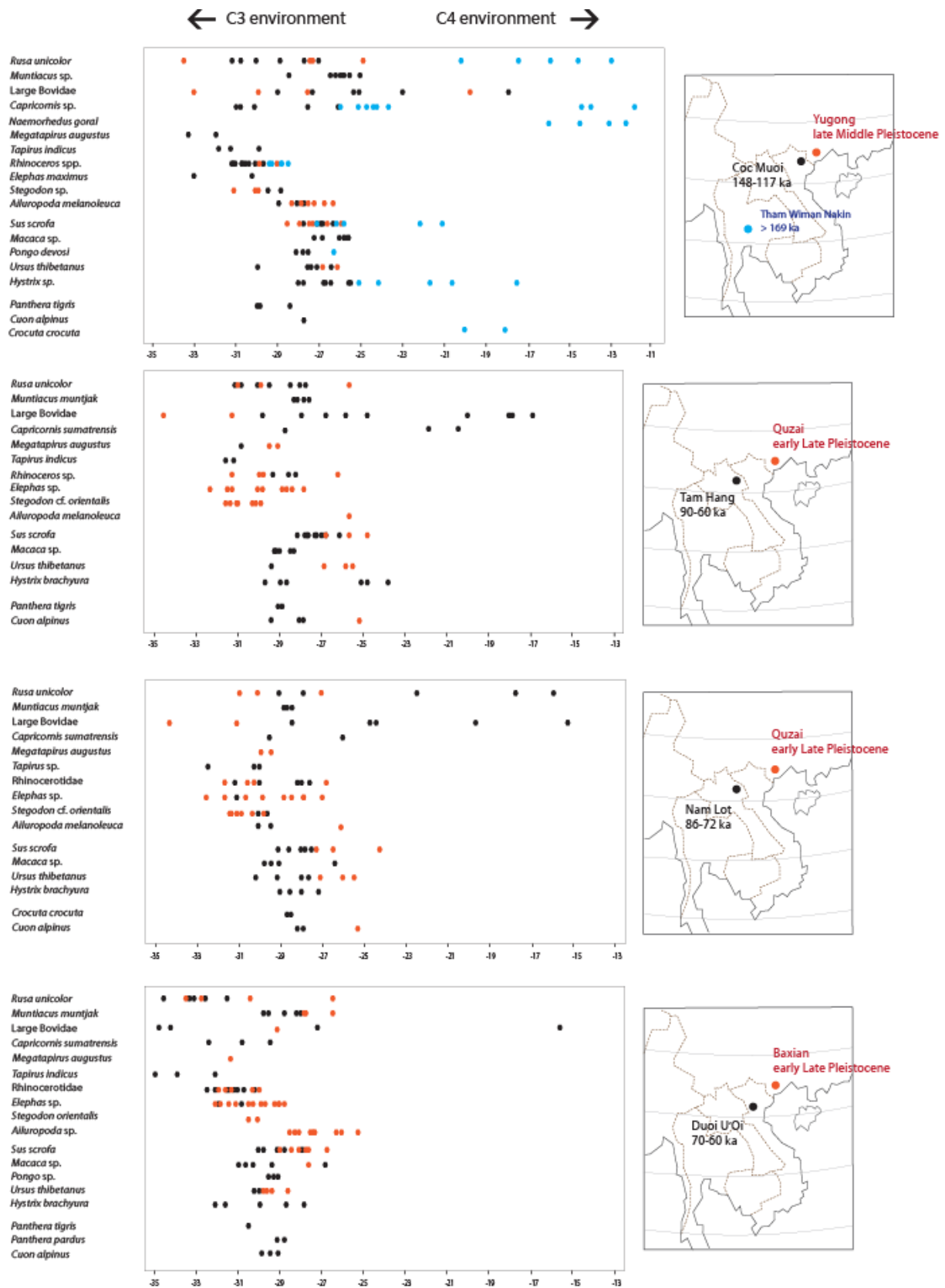
**Figure S4.** Palaeoproteomic analysis reveals the Nam Lot 433 lower incisor to represent a *Pongo* sp. specimen. **a.** Heatmap clustering of amino acid sequence distances between Nam Lot and extant Hominoidea, with *Macaca mulatta* as an outgroup. The phylogenetic tree based on protein distances reflected the commonly proposed hypothesis among Hominidae. **b.** Ion fragment series of a peptide overlapping *Pongo*-derived SAPs at positions 283 and 286 in AMBN. **c.** Ion fragment series of a peptide overlapping *Pongo*-derived SAPs at positions 126 and 137 in ENAM. **d.** Ion fragment series of a peptide overlapping *Pongo*-derived SAP at position 212 in ENAM. Figure S4 was created using the Package ‘pheatmap’ (v.1.0.12), available using this link (<https://mran.microsoft.com/snapshot/2017-09-01/web/packages/pheatmap/pheatmap.pdf>).



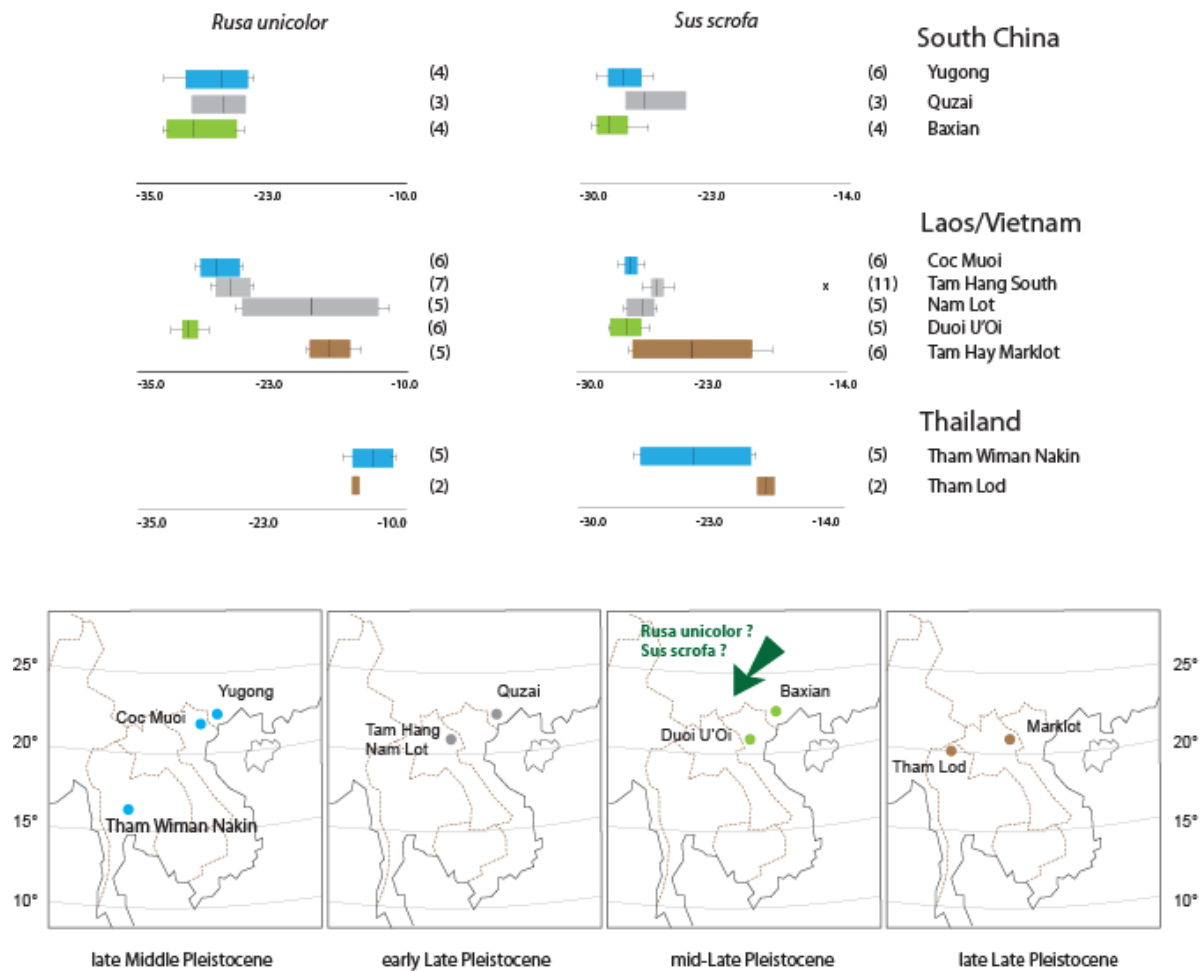
**Figure S5.** Distribution of  $\delta^{13}\text{C}_{\text{carbon source}}$  values (‰ VPDB) in herbivores, omnivores and carnivores from Tham Lod rockshelter, Thailand (33 – 11.5 ka; Suraprasit et al., 2020), Tam Hay Marklot, Laos (38.4 – 13.5 ka; Bourgon et al., 2020), and Boh Dambang, Cambodia (25 -18 ka; Bacon et al., 2018c) (see [Supplementary Annexes S4-S5](#) for  $\delta^{13}\text{C}_{\text{carbon source}}$  values).



**Figure S6.** Ranges of  $\delta^{13}\text{C}_{\text{carbon source}}$  values in *Capricornis sumatraensis* and *Naemorhedus sp.* in sites from Thailand, Laos, and Vietnam. *N. cf. caudatus* at Tam Hay Marklot, Laos (38.4 – 13.5 ka); *N. crispus* and *N. griseus* at Tham Lod, Thailand (33 – 11.5 ka; Suraprasit et al., 2020). Below, location of sites by period, with the hypothesis of movements of populations of *Naemorhedus* species, coming likely from southern latitudes where populations had become over the course of time adapted to living in grasslands (see [Supplementary Annexes S1, S2, S4](#) for  $\delta^{13}\text{C}_{\text{carbon source}}$  values).



**Figure S7.** Distribution of  $\delta^{13}\text{C}_{\text{carbon source}}$  values (‰ VPDB) in herbivores, omnivores and carnivores from Vietnam, Laos, Thailand and southern China sites (see [Supplementary Annexes S1-S3](#) for  $\delta^{13}\text{C}_{\text{carbon source}}$  values).



**Figure S8.** Ranges of  $\delta^{13}\text{C}_{\text{carbon source}}$  values in *Rusa unicolor* and *Sus scrofa* from Thailand, Laos, Vietnam and southern China sites. Below, location of sites by period, with the hypothesis of movements of populations of *R. unicolor* and *S. scrofa* inferred from the present analysis. Other taxa (rhinoceroses, bears, bovids) could have been also concerned by this range shift (see carbon isotope data in Sun et al., 2019).

Vietnam: Coc Muoi (148 – 117 ka); Duoi U’Oi (70 – 60 ka). Laos: Tam Hang South (94 – 60 ka); Nam Lot (86 – 72 ka); Tam Hay Marklot (38.4 – 13.5 ka). Southern China: Yugong cave, Guangxi (late Middle Pleistocene; Sun et al., 2019); Baxian cave, Guangxi (early Late Pleistocene; Ma et al., 2017; Sun et al., 2019); Quzai cave, Guangxi (early Late Pleistocene; Ma et al., 2019). Thailand: Tham Lod (33 – 11.5 ka; Suraprasit et al., 2020); Tham Wiman Nakin (> ~170 ka; (Esposito et al., 1998, 2002; Pushkina et al., 2010; Suraprasit et al., 2020).

## Supplementary Tables

**Table S1**

site	individual	Std $\delta^{13}\text{C}$ (‰)	Std $\delta^{18}\text{O}$ (‰)
Coc Muoi	34890	0.056	0.040
Coc Muoi	34911	0.042	0.036
Coc Muoi	34937	0.136	0.165
Duoi Uoi	34831	0.050	0.151
Duoi Uoi	34845	0.007	0.125
Duoi Uoi	34851	0.044	0.140

Standard deviation of repeated analysis (n=3) for 6 individuals from Coc Muoi and Duoi U’Oi used to estimate the intra-individual heterogeneity and analytical reproductibility of enamel analysis.

**Table S2**

Protein	UniProt <i>Pongo abelii</i> accession	Source	Peptides	Razor+ Unique peptides	Coverage (%)	Deamidation N (% $\pm$ 1 SD)	Deamidation Q (% $\pm$ 1 SD)
AMELX (Isoform 1)	H2PUX0	Enamel proteome	506	506	81.2	92.2 (2.3)	99.3 (0.3)
AMELX (Isoform 3)	H2PUX0	Enamel proteome	500	74	82.4		
AMBN	H2PDI5	Enamel proteome	208	208	353	99.9 (0.0)	98.9 (0.5)
ENAM	H2PDI6	Enamel proteome	376	376	17.9	99.2 (0.3)	96.1 (1.2)
MMP20	H2NF32	Enamel proteome	7	7	8.7	100 (0.0)	NA
AMTN	H2PDI4	Enamel proteome	2	2	8.7	NA	100 (0.0)
ALB	Q5NVH5	Enamel proteome	16	16	17.6	99.0 (1.0)	100 (0.0)
COL17A1	H2NBI5	Enamel proteome	19	19	4.1	NA	84.9 (15.7)
A2VCT4		Contaminant	2	2	3.7	50.8 (28.4)	NA
ENSBTAP00000024146		Contaminant	2	2	1.6		
XP_585019		Contaminant	2	2	3.3		

Protein group LC-MS/MS results and coverage. Deamidation was calculated for the contaminants together. Peptide counts taken from the MaxQuant proteingroups.txt file.

**Table S3**

Order	Common name	Taxon	Coc Muoi	Tam Hang South	Nam Lot	Duoi U'O'i	Tam Hay Marklot	
<b>Artiodactyla</b>	Sambar	<i>Rusa unicolor</i>	X	X	X	X	X	
	-	medium-sized cervid	X	X				
	Thamin	<i>Rucervus eldii</i>					X	
	Hog deer	<i>Axis porcinus</i>					cf.	
	Indian muntjac	<i>Muntiacus muntjak</i>		X	X	X		
	-	<i>Muntiacus sp.</i>	X				X	
	Kouprey	<i>Bos sauveli</i>	cf.	cf.				
	Gayal	<i>Bos frontalis</i>					cf.	
	-	<i>Bos sp.</i>			X			
	Large-sized Bovidae	Bovidae	X	X	X	X	X	
	Water buffalo	<i>Bubalus bubalis</i>		X	X	cf.	X	
	Southern serow	<i>Capricornis sumatraensis</i>		X	X	X	cf.	
	-	<i>Capricornis sp.</i>	X					
	Chinese goral	<i>Naemorhedus caudatus</i>					cf.	
	-	<i>Naemorhedus sp.</i>						
	Wild boar	<i>Sus scrofa</i>	X	X	X	X	X	
	Bearded pig	<i>Sus barbatus</i>		cf.		X	cf.	
	-	<i>Sus sp.</i>						
	<b>Perissodactyla</b>	Giant tapir	<i>Megatapirus augustus</i>	X	X			
		-	<i>Tapirus indicus intermedius</i>		cf.			
Malayan tapir		<i>Tapirus indicus</i>	X			X		
-		<i>Tapirus sp.</i>			X		X	
Indian rhinoceros		<i>Rhinoceros unicornis</i>	cf.	X	X	X		
Javan rhinoceros		<i>Rhinoceros sondaicus</i>	X	X	X	X	X	
Sumatran rhinoceros		<i>Dicerorhinus sumatrensis</i>	X		0	X	X	
-		<i>Rhinoceros sp.</i>		X	X	X		
<b>Proboscidea</b>	Asian elephant	<i>Elephas maximus</i>	X					
	-	<i>Elephas sp.</i>		X	X	X		
	Stegodon	<i>Stegodon orientalis</i>		X	cf.			
	-	<i>Stegodon sp.</i>	X					

Faunal lists of the five sites (Artiodactyla, Perissodactyla, and Proboscidea).

**Table S4**

Order	Common name	Taxon	Coc Muoi	Tam Hang South	Nam Lot	Duoi U'Oi	Tam Hay Marklot
<b>Carnivora</b>	Dhole	<i>Cuon alpinus</i>	X		X	X	?
	Dhole	<i>Cuon alpinus antiquus</i>		cf.			
	-	<i>Cuon sp.</i>					
	Wild dogs	Canidae					X
	Hog-badger	<i>Arctonyx collaris</i>				X	
	Hog-badger	<i>Arctonyx collaris rostratus</i>		cf.			
	Eurasian badger	<i>Meles meles</i>		X	X	X	
	Large tooth ferret-badger	<i>Melogale personata</i>		X			
	Yellow-throated marten	<i>Martes flavigula</i>			cf.	X	
	-	<i>Martes sp.</i>	X				
	Large Indian civet	<i>Viverra zibetha</i>			X	X	X
	Large-spotted civet	<i>Viverra megaspila</i>					cf.
	Common palm civet	<i>Paradoxurus hermaphroditus</i>			X		
	-	<i>Paradoxurus sp.</i>					
	-	large-sized meline	X				
	-	small-sized meline	X				X
	Tiger	<i>Panthera tigris</i>	X	X		X	X
	Leopard	<i>Panthera pardus</i>				X	X
	Leopard cat	<i>Prionailurus bengalensis</i>			cf.		
	Golden cat	<i>Felis temmincki</i>				cf.	
	Clouded leopard	<i>Neofelis nebulosa</i>				X	
	-	small-sized felid	X				X
	Spotted hyena	<i>Crocuta crocuta</i>				X	
	Asiatic black bear	<i>Ursus thibetanus</i>	X			X	X
	Asiatic black bear	<i>Ursus thibetanus kokeni</i>			cf.	cf.	
	Sun bear	<i>Helarctos malayanus</i>	cf.	X		X	X
	Giant Panda	<i>Ailuropoda melanoleuca</i>	X			X	X
-	<i>Ailuropoda sp.</i>						
<b>Primates</b>	Orangutan	<i>Pongo pygmaeus</i>		X	X	X	
	Orangutan	<i>Pongo devosi</i>	X				
	-	<i>Pongo sp.</i>					X
	Macaque	<i>Macaca sp.</i>	X			X	X
	-	Colobine	X	X	X	X	
	Gibbon	<i>Hylobates sp.</i>	X	X		X	
		Hominine				X	
<b>Rodentia</b>	Porcupine	<i>Hystrix brachyura</i>		X	X	X	
	-	<i>Hystrix sp.</i>	X				x
	Brush-tailed porcupine	<i>Atherurus macrourus</i>					cf.
	-	<i>Atherurus sp.</i>	X				

Faunal lists of the five sites (Carnivora, Primates, and large Rodentia).



**Table S5**

Site	Taxon	Crown area	C13 / O18	
<b>Coc Muoi</b>	<i>Rusa unicolor</i>	11 rp3, 5 lp3	3 rm3, 3 lm3	
	<i>Muntiacus</i> sp.	2 lm3, 2 rm3	3 lm3, 2 IP2/P3, 2 IP3/P4	
	Large-sized Bovidae ( <i>Bos</i> cf. <i>sauveli</i> )	-	6 rm3	
	<i>Capricornis</i> sp.	1 lm3, 1 rm3	2 rm1/m2, 3lm1/m2	
	<i>Megatapirus augustus</i>	-	1 m, 1 p3	
	<i>Tapirus indicus</i>	-	1 p3, 2 p4	
	Rhinocerotidae	-	8 p4, 2 M3, 2 m3	
	<i>Elephas maximus</i>	-	2 m	
	<i>Stegodon</i> sp.	-	2 milk teeth	
	<i>Ailuropoda melanoleuca</i>	-	2 IM1, 1 rM1	
	<i>Sus scrofa</i>	8 lp3, 5 rp3	5 lm3, 1 m3	
	<i>Macaca</i> sp.	2 lm3	3 c/C, 1 IM3, 1 lm1/m2	
	<i>Pongo devosi</i>	-	1 II1, 1rM1, 1 IM, 1 lm1	
	<i>Ursus thibetanus/Ursus</i> sp.	-	2 IM1, 1 IM2, 1 rm3, 1 lm3	
	<i>Hystrix</i> sp.	-	8 I/i	
	<i>Atherurus</i> sp.	-	3 I/i	
	<i>Panthera tigris</i>	-	2 IP3, 1 lm1	
	Small-sized Felidae	-	2 rP4	
	<i>Cuon alpinus</i>	-	1 rP4	
	<b>Tam Hang South</b>	<i>Rusa unicolor</i>	3 lp3	2 lp2, 3 m, 1 M, 1 rP3
<i>Muntiacus muntjak</i>		4 lm3, 4 rm3	4 rm3	
Large-sized Bovidae		-	4 rp4, 1 lp4, 1 p4, 1 p3, 1 rm3	
<i>Capricornis sumatraensis</i>		1 m3	1M, 1 m, 1 m3	
<i>Megatapirus augustus</i>		-	1 rm1/m2	
<i>Tapirus indicus</i> cf. <i>intermedius</i>		-	1 rM1/M2, 1 rp2	
<i>Rhinoceros</i> spp.		-	1 rp4/m1, 1 lm3, 1 ld4	
<i>Sus scrofa</i>		9 rp3, 3 lp3	1 IP4, 4 rP4, 5 M	
<i>Macaca</i> sp.		2 rm3, 5 lm3	6 lm3	
<i>Ursus thibetanus</i> cf. <i>kokeni</i>		-	2 rP4	
<i>Helarctos malayanus</i>		-	1 lm2, 1 lm3	
<i>Hystrix brachyura</i>		-	6 I/i	
<i>Panthera tigris</i>		-	2 IP3	
<i>Cuon alpinus</i> cf. <i>antiquus</i>		-	1 rp4, 1 lm2, 1 IM1	
<b>Nam Lot</b>		<i>Rusa unicolor</i>	3 lp3, 5 rp3	2 rp3, 2 lm, 1 rM1
		<i>Muntiacus muntjak</i>	1 rm3	2 rM, 1 IM
	Large-sized Bovidae	-	1 rp4, 1 lp4, 1 lm3, 2 M	
	<i>Capricornis sumatraensis</i>	1 lm3, 2 rm3	2 rm3	
	<i>Tapirus</i> sp.	-	1 lm, 1li	
	<i>Rhinoceros</i> spp.	-	2 p/m, 2 ld, 1d, 1 ld2	
	<i>Elephas</i> sp.	-	1 m (fragment)	
	<i>Stegodon</i> cf. <i>orientalis</i>	-	2 m (fragment)	
	<i>Ailuropoda melanoleuca</i>	-	1 rm1, 1 IM2	
	<i>Sus scrofa</i>	3 rp3, 1 lp3	3 rp4, 1 rm1, 1 fgt	
	<i>Macaca</i> sp.	2 lm3	2 I/I, 2 rm3	
	<i>Pongo</i> sp.	-	1 i	
	<i>Ursus thibetanus</i> cf. <i>kokeni</i>	-	2 rm3, 2 IP4	
	<i>Hystrix brachyura</i>	-	2 I/I, 3 p/m	
	<i>Crocota crocuta</i>	-	1 lp3, 1 pm, 1 rp2, 1 rp3, 2 IP2	
	Small-sized Felidae	-	1 IP3	
	<i>Cuon alpinus</i>	-	1 pm, 1 lp3	

Site, taxon, and tooth type used for the morphometric (crown area dimension) and the stable carbon (C13) and oxygen (O18) isotope analyses.

**Table S6**

Site	Taxon	Crown area	C13 / O18
Duoi U'Oi	<i>Rusa unicolor</i>	3 lp3, 1 rp3	6 lm3
	<i>Muntiacus muntjak</i>	7 lm3, 5 rm3	5 lm3
	Large-sized Bovidae	-	4 m
	<i>Capricornis sumatraensis</i>	2 lm3, 2 rm3	2 rm3, 1 lm3
	<i>Tapirus indicus</i>	-	1 IM1/M2, 2 IP/M
	Rhinocerotidae	-	2 rM2, 5 rM3, 1 rp2
	<i>Elephas sp.</i>	-	2 milk teeth
	<i>Sus scrofa</i>	9 rp3, 5 lp3	5 lp4
	<i>Macaca sp.</i>	7 lm3, 8 rm3	5 i/l
	<i>Pongo pygmaeus</i>	-	2 rp3
	Ursidae	-	2 rI3
	<i>Hystrix brachyura</i>	-	5 I/i
	<i>Panthera tigris</i>	-	1 rm1
	<i>Panthera pardus</i>	-	2 lp4
	<i>Cuon alpinus</i>	-	3 IP4
	Tam Hay Marklot	<i>Rusa unicolor</i>	18 rp3, 16 lp3
<i>Muntiacus sp.</i>		13 lm3, 4 rm3	5 lm3
<i>Axis cf. porcinus</i>		-	2 lm3
<i>Rucervus eldii</i>		-	2 lm3
Large-sized Bovidae		-	5 rp2, 5 lp2
<i>Capricornis cf. sumatraensis</i>		6 lm3, 2 rm3	5 lm3
<i>Naemohedus cf. caudatus</i>		-	4 lm3
Tapiridae/Tapirus sp.		-	1 lc, 1 p/m
<i>Rhinoceros sondaicus</i>		-	1 ld3, 1 rd3, 1 lm2, 2 lm3
<i>Ailuropoda melanoleuca</i>		-	1 rm2
<i>Sus scrofa</i>		10 rp3, 8 lp3	6 lp4
<i>Macaca sp.</i>		1 rm3	2 rp, 3 lm1/m2
<i>Pongo sp.</i>		-	1 c/C, 1 IM
<i>Ursus thibetanus</i>		-	1 rM2, 2 IM2
<i>Helarctos malayanus</i>		-	2 IM2
<i>Hystrix sp.</i>		-	4 I/i
<i>Panthera tigris</i>		-	1 rp4, 2 IP4
<i>Panthera pardus</i>		-	1 rP3, 1 lp4, 1 IP4
<i>Cuon alpinus</i>		-	1 rm1, 1 rP3, 1 IM1
Canidae		-	1 IM1

Site, taxon, and tooth type used for the morphometric (crown area dimension) and the stable carbon (C13) and oxygen (O18) isotope analyses.

**Table S7**

	Artiodactyla		Perissodactyla		Proboscidea		Carnivora		Primates		Rodentia	
<b>Coc Muoi</b>	35.71 %	30/84	20.23 %	17/84	4.76 %	4/84	16.66 %	14/84	10.71 %	9/84	11.90 %	10/84
<b>Tam Hang South</b>	56.45 %	35/62	9.67 %	6/62	-	-	14.51 %	9/62	9.67 %	6/62	9.67 %	6/62
<b>Nam Lot</b>	35.08 %	20/57	15.78 %	9/57	5.26 %	3/57	26.31 %	15/57	10.52 %	6/57	7.01 %	4/57
<b>Duoi U'Oi</b>	38.33 %	23/60	18.33 %	11/60	3.33 %	2/60	13.33 %	8/60	13.33 %	8/60	13.33 %	8/60
<b>Tam Hay Marklot</b>	54.16 %	39/72	9.72 %	7/72	-	-	20.83 %	15/72	9.72 %	7/72	5.55 %	4/72
<b>Boh Dambang</b>	50 %	26/52	5.76 %	3/52	-	-	28.84 %	15/52	9.61 %	5/52	5.76 %	3/52

Percentage (%) and number of specimens (n/N) by taxonomic group used for the carbon and oxygen isotope analyses.

**Table S8**

group1	group2	n (group1)	n (group2)	statistic	p	p.adj
CM	THS	84	62	5.58	0.000	0.000
CM	NL	84	57	3.72	0.000	0.000
CM	DU	84	60	-0.52	0.601	0.601
CM	THM	84	72	7.05	0.000	0.000
THS	NL	62	57	-1.62	0.106	0.133
THS	DU	62	60	-5.65	0.000	0.000
THS	THM	62	72	1.14	0.256	0.284
NL	DU	57	60	-3.93	0.000	0.000
NL	THM	57	72	-3.93	0.000	0.000
DU	THM	60	72	6.98	0.000	0.000

Results of the Post-hoc Dunn's test pair-wise comparisons on  $\delta^{13}\text{C}_{\text{carbon source}}$  values between sites (CM, Coc Muoi; THS, Tam Hang South; NL, Nam Lot; DU, Duoi U'Oi; THM, Tam Hay Marklot).

**Table S9**

group1	group2	n (group1)	n (group2)	statistic	p	p.adj
CM	THS	84	62	-2.83	0.005	0.009
CM	NL	84	57	1.42	0.155	0.221
CM	DU	84	60	-1.99	0.047	0.078
CM	THM	84	72	1.22	0.221	0.277
THS	NL	62	57	3.91	0.000	0.001
THS	DU	62	60	0.76	0.445	0.495
THS	THM	62	72	3.87	0.000	0.001
NL	DU	57	60	-3.14	0.0002	0.006
NL	THM	57	72	-0.27	0.788	0.788
DU	THM	60	72	3.04	0.002	0.006

Results of the Post-hoc Dunn's test pair-wise comparisons on  $\delta^{18}\text{O}$  values between sites (CM, Coc Muoi; THS, Tam Hang South; NL, Nam Lot; DU, Duoi U'Oi; THM, Tam Hay Marklot).

**Table S10**

		18 – 80 kg	80 – 350 kg	350 – 1000 kg	>1000 kg
Coc Muoi (148 – 117 ka)	Ruminant	<i>Muntiacus</i> sp.	<i>Rusa unicolor</i> medium-sized cervid <i>Capricornis</i> sp.	<i>Bos</i> cf. <i>sauveli</i>	
	Non-ruminant		<i>Sus scrofa</i>	<i>Megatapirus augustus</i> <i>Tapirus indicus</i> <i>Dicerorhinus sumatrensis</i>	<i>Rhinoceros sondaicus</i> <i>Rhinoceros</i> cf. <i>unicornis</i> <i>Elephas maximus</i> <i>Stegodon</i> sp.
Tam Hang South (92 – 60 ka)	Ruminant	<i>Muntiacus muntjak</i>	<i>Rusa unicolor</i> medium-sized cervid <i>Capricornis sumatraensis</i>	<i>Bos</i> cf. <i>sauveli</i> <i>Bubalus bubalis</i>	
	Non-ruminant		<i>Sus scrofa</i> <i>Sus</i> cf. <i>barbatus</i>	<i>Megatapirus augustus</i> <i>Tapirus indicus</i> cf. <i>intermedius</i>	<i>Rhinoceros sondaicus</i> <i>Rhinoceros unicornis</i> <i>Rhinoceros</i> sp. <i>Elephas</i> sp. <i>Stegodon orientalis</i>
Nam Lot (86 – 72 ka)	Ruminant	<i>Muntiacus muntjak</i>	<i>Rusa unicolor</i> <i>Capricornis sumatraensis</i>	<i>Bos</i> sp. <i>Bubalus bubalis</i>	
	Non-ruminant		<i>Sus scrofa</i>	<i>Tapirus</i> sp.	<i>Rhinoceros sondaicus</i> <i>Rhinoceros unicornis</i> <i>Rhinoceros</i> sp. <i>Elephas</i> sp. <i>Stegodon</i> cf. <i>orientalis</i>
Duoï U’Oï (70 – 60 ka)	Ruminant	<i>Muntiacus muntjak</i>	<i>Rusa unicolor</i> <i>Capricornis sumatraensis</i>	<i>Bubalus</i> cf. <i>bubalis</i>	
	Non-ruminant		<i>Sus scrofa</i> <i>Sus barbatus</i>	<i>Tapirus indicus</i> <i>Dicerorhinus sumatrensis</i>	<i>Rhinoceros sondaicus</i> <i>Rhinoceros unicornis</i> <i>Elephas</i> sp.
Tam Hay Marklot (38.4 – 13.5 ka)	Ruminant	<i>Muntiacus</i> sp. <i>Axis</i> cf. <i>porcinus</i> <i>Naemorhedus caudatus</i>	<i>Rusa unicolor</i> <i>Rucervus eldii</i> <i>Capricornis</i> cf. <i>sumatraensis</i>	<i>Bos</i> sp. <i>Bubalus bubalis</i>	
	Non-ruminant		<i>Sus</i> sp. <i>Sus</i> cf. <i>barbatus</i>	<i>Tapirus</i> sp. <i>Dicerorhinus sumatrensis</i>	<i>Rhinoceros sondaicus</i> <i>Elephas</i> sp.
Current faunas	Ruminant	<i>Muntiacus muntjak</i> <i>Muntiacus rooseveltorum</i> <i>Naemorhedus caudatus</i> <i>Cervus nippon</i>	<i>Rusa unicolor</i> <i>Rucervus eldii</i> <i>Capricornis sumatraensis</i>	<i>Bos gaurus</i> <i>Bos javanicus</i>	
	Non-ruminant		<i>Sus scrofa</i>	<i>Dicerorhinus sumatrensis</i>	<i>Rhinoceros sondaicus</i> <i>Elephas maximus</i>

Lists of taxa by body mass and dietary strategy. The current faunas are those from the studied latitudinal zone in the pre-industrial period (Corbet and Hill, 1992); body size categories are from Faith et al. (2019). No herbivore species smallest than ~20 kg is recorded in the four fossil assemblages. Only species of Tragulidae constitute the current faunas at these latitudes (Corbet and Hill, 1992).

**Table S11**

group1	group2	n (group1)	n (group2)	statistic	p	p.adj
CM	THS	16	3	0.95	0.341	0.426
CM	NL	16	8	4.11	0.000	0.000
CM	DU	16	4	2.91	0.004	0.012
CM	THM	16	34	1.82	0.069	0.136
THS	NL	3	8	1.74	0.082	0.136
THS	DU	3	4	1.34	0.179	0.255
THS	THM	3	34	-0.08	0.936	0.936
NL	DU	8	4	-0.25	0.804	0.893
NL	THM	8	34	-3.12	0.002	0.009
DU	THM	4	34	-2.03	0.042	0.105

Results of the Post-hoc Dunn's test pair-wise comparisons on crown area dimensions of *Rusa unicolor* between sites (CM, Coc Muoi; THS, Tam Hang South; NL, Nam Lot; DU, Duoi U'Oi; THM, Tam Hay Marklot).

**Table S12**

group1	group2	n (group1)	n (group2)	statistic	p	p.adj
CM	THS	13	12	1.75	0.080	0.133
CM	NL	13	4	2.47	0.014	0.034
CM	DU	13	14	-0.88	0.378	0.431
CM	THM	13	18	-0.09	0.929	0.929
THS	NL	12	4	1.23	0.220	0.314
THS	DU	12	14	-2.65	0.008	0.030
THS	THM	12	18	-1.97	0.049	0.098
NL	DU	4	14	-3.09	0.002	0.020
NL	THM	4	18	-2.61	0.009	0.030
DU	THM	14	18	0.86	0.388	0.431

Results of the Post-hoc Dunn's test pair-wise comparisons on crown area dimensions of *Sus scrofa* between sites (CM, Coc Muoi; THS, Tam Hang South; NL, Nam Lot; DU, Duoi U'Oi; THM, Tam Hay Marklot).

**Table S13**

	<i>Rusa unicorn</i>		<i>Muntiacus sp.</i>		<i>Capricornis sumatraensis</i>		<i>Sus scrofa</i>		<i>Macaca sp.</i>	
	p3	$\delta^{13}\text{C}$	m3	$\delta^{13}\text{C}$	m3	$\delta^{13}\text{C}$	p3	$\delta^{13}\text{C}$	m3	$\delta^{13}\text{C}$
<b>Coc Muoi</b>	16	6	4	7	2	5	13	6	2	5
<b>Tam Hang South</b>	3	8	8	4	1	3	12	11	7	6
<b>Nam Lot</b>	8	5	1	3	3	2	4	5	2	4
<b>Duoi U'Oi</b>	4	6	12	5	4	3	14	5	15	5
<b>Tam Hay Marklot</b>	34	5	17	5	8	5	18	6	1	5
<b>Total</b>	65	30	42	24	18	18	61	33	27	25

Taxon, tooth type, and number of specimens used for the crown area measurements, and number of specimens used for the  $\delta^{13}\text{C}$  isotope analysis (data of the Figure 5).

**Table S14**

	$\delta^{13}\text{C}_{\text{carbon source}} < -27.2 \text{‰}$		$\delta^{13}\text{C}_{\text{carbon source}} > -27.2 \text{‰ and } < -21.3 \text{‰}$		$\delta^{13}\text{C}_{\text{carbon source}} > -21.3 \text{‰ and } < -15.3 \text{‰}$		$\delta^{13}\text{C}_{\text{carbon source}} > -15.3 \text{‰}$	
	%	n/N	%	n/N	%	n/N	%	n/N
<b>Coc Muoi</b>	65.48 %	55/84	33.33 %	28/84	1.19 %	1/84	-	0/84
<b>Tam Hang South</b>	27.42 %	17/62	58.06 %	36/62	11.29 %	7/62	3.23 %	2/62
<b>Nam Lot</b>	42.11 %	24/57	49.12 %	28/57	5.26 %	3/57	3.51 %	2/57
<b>Duoi U'Oi</b>	73.33 %	44/60	25.00 %	15/60	-	0/60	1.67 %	1/60
<b>Tam Hay Marklot</b>	26.39 %	19/72	43.06 %	31/72	18.05 %	13/72	12.50 %	9/72
<b>Boh Dambang</b>	9.61 %	5/52	32.69 %	17/52	38.46 %	20/52	19.23 %	10/52

Percentage (%) and number of specimens (n/N) in the five faunas compared with the data from the fauna of Boh Dambang (Bacon et al., 2018c), according to the distribution of  $\delta^{13}\text{C}_{\text{carbon source}}$  values (‰ VPDB).

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**Annex S1.** Faunal lists from main sites investigated in the present study (Coc Muoi, Tam Hang South and Duoi U’Oi) with associated  $\delta^{13}\text{C}_{\text{apatite}}$ ,  $\delta^{13}\text{C}_{\text{carbon source}}$  and  $\delta^{18}\text{O}_{\text{apatite}}$  values (‰ VPDB), as well as body mass and  $\delta^{13}\text{C}$  (‰ VPDB) Enrichment Factor used to obtain  $\delta^{13}\text{C}_{\text{carbon source}}$ .

Number	Country	Site	Taxon	Body Mass (kg)	$\delta^{13}\text{C}_{\text{apatite}}$ (‰ VPDB)	$\delta^{13}\text{C}$ (‰) Enrichment Factor	$\delta^{13}\text{C}_{\text{carbon source}}$ (‰ VPDB)	$\delta^{18}\text{O}_{\text{apatite}}$ (‰ VPDB)
DU876	Vietnam	Duoi U’Oi	<i>Sus scrofa</i>	137	-14.2	13.19	-27.4	-5.1
DU890	Vietnam	Duoi U’Oi	<i>Sus scrofa</i>	137	-12.8	13.19	-26.0	-8.3
DU905	Vietnam	Duoi U’Oi	<i>Sus scrofa</i>	137	-13.7	13.19	-26.9	-7.1
DU906	Vietnam	Duoi U’Oi	<i>Sus scrofa</i>	137	-15	13.19	-28.2	-6
DU913	Vietnam	Duoi U’Oi	<i>Sus scrofa</i>	137	-14.7	13.19	-27.9	-6.2
DU546	Vietnam	Duoi U’Oi	<i>Rusa unicolor</i>	220	-16.6	13.59	-30.2	-7.1
DU557	Vietnam	Duoi U’Oi	<i>Rusa unicolor</i>	220	-19.6	13.59	-33.2	-6.1
DU567	Vietnam	Duoi U’Oi	<i>Rusa unicolor</i>	220	-18.1	13.59	-31.7	-6.1
DU608	Vietnam	Duoi U’Oi	<i>Rusa unicolor</i>	220	-18.1	13.59	-31.7	-7
DU990	Vietnam	Duoi U’Oi	<i>Rusa unicolor</i>	220	-18.3	13.59	-31.9	-5.7
DU1087	Vietnam	Duoi U’Oi	<i>Rusa unicolor</i>	220	-17.7	13.59	-31.3	-8.5
DU437	Vietnam	Duoi U’Oi	<i>Muntiacus muntjak</i>	24	-13.2	12.17	-25.4	-9.3
DU461	Vietnam	Duoi U’Oi	<i>Muntiacus muntjak</i>	24	-14.8	12.17	-27.0	-7.7
DU511	Vietnam	Duoi U’Oi	<i>Muntiacus muntjak</i>	24	-14.6	12.17	-26.8	-5.1
DU433	Vietnam	Duoi U’Oi	<i>Muntiacus muntjak</i>	24	-13.8	12.17	-26.0	-7.8
DU392	Vietnam	Duoi U’Oi	<i>Muntiacus muntjak</i>	24	-13	12.17	-25.2	-6.5
DU574	Vietnam	Duoi U’Oi	<i>Capricornis sumatraensis</i>	112	-15.8	13.14	-28.9	-7.1
DU613	Vietnam	Duoi U’Oi	<i>Capricornis sumatraensis</i>	112	-17.4	13.14	-30.5	-7.5
DU538	Vietnam	Duoi U’Oi	<i>Capricornis sumatraensis</i>	112	-14.5	13.14	-27.6	-6.1
DU992	Vietnam	Duoi U’Oi	Bovidae	875	-19.8	14.57	-34.4	-7.1
DU983	Vietnam	Duoi U’Oi	Bovidae	875	-12.1	14.57	-26.7	-5.2
DU984	Vietnam	Duoi U’Oi	Bovidae	875	-0.6	14.57	-15.2	-4.7
DU560	Vietnam	Duoi U’Oi	Bovidae	875	-19.3	14.57	-33.9	-2.5
	Vietnam	Duoi U’Oi	<i>Hystrix brachyura</i>	12	-16.7	12.18	-28.9	-8
	Vietnam	Duoi U’Oi	<i>Hystrix brachyura</i>	12	-12.6	12.18	-24.8	-6.7
	Vietnam	Duoi U’Oi	<i>Hystrix brachyura</i>	12	-16.26	12.18	-28.4	-9.71
	Vietnam	Duoi U’Oi	<i>Hystrix brachyura</i>	12	-12.8	12.18	-25.0	-6.39
	Vietnam	Duoi U’Oi	<i>Hystrix brachyura</i>	12	-17.08	12.18	-29.3	-8.68
	Vietnam	Duoi U’Oi	<i>Hystrix brachyura</i>	12	-13.7	12.18	-25.9	-6.6
	Vietnam	Duoi U’Oi	<i>Hystrix brachyura</i>	12	-17.3	12.18	-29.5	-5.1
DU7	Vietnam	Duoi U’Oi	<i>Hystrix brachyura</i>	12	-14.7	12.18	-26.9	-8.9
DU728	Vietnam	Duoi U’Oi	<i>Ursus sp.</i>	100	-15	13.3	-28.3	-6.5
DU729	Vietnam	Duoi U’Oi	<i>Ursus sp.</i>	100	-14.9	13.3	-28.2	-9.3
DU1152	Vietnam	Duoi U’Oi	<i>Cuon alpinus</i>	15	-14.2	13.39	-27.6	-7.3
DU1153	Vietnam	Duoi U’Oi	<i>Cuon alpinus</i>	15	-14.9	13.39	-28.3	-6.7

DU77	Vietnam	Duoi U'Oi	<i>Cuon alpinus</i>	15	-14.5	13.39	-27.9	-7.4
DU68	Vietnam	Duoi U'Oi	<i>Panthera pardus</i>	41	-14.2	13.81	-28.0	-5.2
DU86	Vietnam	Duoi U'Oi	<i>Panthera pardus</i>	41	-13.9	13.81	-27.7	-5.2
DU707	Vietnam	Duoi U'Oi	<i>Panthera tigris</i>	212	-15.5	14.53	-30.0	-5.4
DU326	Vietnam	Duoi U'Oi	<i>Macaca sp.</i>	6	-11.9	11.91	-23.8	-6.2
DU331	Vietnam	Duoi U'Oi	<i>Macaca sp.</i>	6	-15.6	11.91	-27.5	-5.1
DU339	Vietnam	Duoi U'Oi	<i>Macaca sp.</i>	6	-14.4	11.91	-26.3	-4.4
DU343	Vietnam	Duoi U'Oi	<i>Macaca sp.</i>	6	-15.4	11.91	-27.3	-5
DU322	Vietnam	Duoi U'Oi	<i>Macaca sp.</i>	6	-16	11.91	-27.9	-6.3
DU32	Vietnam	Duoi U'Oi	<i>Rhinoceros unicornis</i>	2250	-17.1	14.4	-31.5	-7.2
DU26	Vietnam	Duoi U'Oi	<i>Rhinoceros unicornis</i>	2250	-16.1	14.4	-30.5	-6.4
DU28	Vietnam	Duoi U'Oi	<i>Rhinoceros unicornis</i>	2250	-16.2	14.4	-30.6	-6.6
DU30	Vietnam	Duoi U'Oi	<i>Rhinoceros sondaicus</i>	2250	-15.7	14.4	-30.1	-6.4
DU31	Vietnam	Duoi U'Oi	<i>Rhinoceros sondaicus</i>	2250	-15.3	14.4	-29.7	-4.9
DU38	Vietnam	Duoi U'Oi	<i>Rhinoceros sondaicus</i>	2250	-16.6	14.4	-31.0	-7
DU27	Vietnam	Duoi U'Oi	<i>Dicerorhinus sumatrensis</i>	950	-17.5	14	-31.5	-7.9
DU24	Vietnam	Duoi U'Oi	<i>Dicerorhinus sumatrensis</i>	950	-16.4	14	-30.4	-6.6
DU47	Vietnam	Duoi U'Oi	<i>Tapirus indicus</i>	300	-17.3	13.5	-30.8	-7.9
DU43	Vietnam	Duoi U'Oi	<i>Tapirus indicus</i>	300	-20.1	13.5	-33.6	-6.9
DU53	Vietnam	Duoi U'Oi	<i>Tapirus indicus</i>	300	-19.1	13.5	-32.6	-8.3
DU1021	Vietnam	Duoi U'Oi	<i>Pongo pygmaeus</i>	55	-14.6	12.78	-27.4	-6
DU1022	Vietnam	Duoi U'Oi	<i>Pongo pygmaeus</i>	55	-14.5	12.78	-27.3	-5.1
DU1023	Vietnam	Duoi U'Oi	<i>Pongo pygmaeus</i>	55	-14.3	12.17	-26.5	-4.9
DU634	Vietnam	Duoi U'Oi	<i>Elephas sp.</i>	4250	-16.9	14.69	-31.6	-7
DU-	Vietnam	Duoi U'Oi	<i>Elephas sp.</i>	4250	-15.9	14.69	-30.6	-6.8
CM169	Vietnam	Coc Muoi	<i>Muntiacus sp.</i>	24	-14.2	12.17	-26.4	-7.1
CM307	Vietnam	Coc Muoi	<i>Muntiacus sp.</i>	24	-14.5	12.17	-26.7	-6.5
CM357	Vietnam	Coc Muoi	<i>Muntiacus sp.</i>	24	-13.5	12.17	-25.7	-5.7
Cm313	Vietnam	Coc Muoi	<i>Muntiacus sp.</i>	24	-14	12.17	-26.2	-5.5
CM175	Vietnam	Coc Muoi	<i>Muntiacus sp.</i>	24	-13.9	12.17	-26.1	-6.9
CM176	Vietnam	Coc Muoi	<i>Muntiacus sp.</i>	24	-13.2	12.17	-25.4	-4.4
CM324	Vietnam	Coc Muoi	<i>Muntiacus sp.</i>	24	-16.5	12.17	-28.7	-6.2
CM450	Vietnam	Coc Muoi	<i>Sus scrofa</i>	137	-14.5	13.19	-27.7	-6.2
CM676	Vietnam	Coc Muoi	<i>Sus scrofa</i>	137	-13.9	13.19	-27.1	-6.9
CM677	Vietnam	Coc Muoi	<i>Sus scrofa</i>	137	-13.8	13.19	-27.0	-6.5
CM678	Vietnam	Coc Muoi	<i>Sus scrofa</i>	137	-13.1	13.19	-26.3	-6.4
CM748	Vietnam	Coc Muoi	<i>Sus scrofa</i>	137	-13.7	13.19	-26.9	-7.2
CM746	Vietnam	Coc Muoi	<i>Sus scrofa</i>	137	-13.9	13.19	-27.1	-6.2
CM543	Vietnam	Coc Muoi	<i>Ailuropoda melanoleuca</i>	92	-17.4	10.51	-27.9	-4.2
CM544	Vietnam	Coc Muoi	<i>Ailuropoda melanoleuca</i>	92	-17.7	10.51	-28.2	-5.6
CM564	Vietnam	Coc Muoi	<i>Ailuropoda melanoleuca</i>	92	-18.5	10.51	-29.0	-6.7

CM415	Vietnam	Coc Muoi	<i>Ursus sp. (? thibetanus)</i>	100	-14.5	13.3	-27.8	-5.8
CM518	Vietnam	Coc Muoi	<i>Ursus thibetanus</i>	100	-17	13.3	-30.3	-6.9
CM519	Vietnam	Coc Muoi	<i>Ursus thibetanus</i>	100	-13.4	13.3	-26.7	-6.4
CM521	Vietnam	Coc Muoi	<i>Ursus sp. (? thibetanus)</i>	100	-14.4	13.3	-27.7	-8.5
CM525	Vietnam	Coc Muoi	<i>Ursus thibetanus</i>	100	-14.1	13.3	-27.4	-6.8
CM508	Vietnam	Coc Muoi	<i>Pongo devosi</i>	55	-15.3	12.78	-28.1	-4
CM579	Vietnam	Coc Muoi	<i>Pongo devosi</i>	55	-15	12.78	-27.8	-4.7
CM551	Vietnam	Coc Muoi	<i>Pongo devosi</i>	55	-14.8	12.78	-27.6	-4.8
CM553	Vietnam	Coc Muoi	<i>Pongo devosi</i>	55	-15	12.78	-27.8	-5
CM509	Vietnam	Coc Muoi	<i>Macaca sp.</i>	6	-14	11.91	-25.9	-4.7
CM577	Vietnam	Coc Muoi	<i>Macaca sp.</i>	6	-13.8	11.91	-25.7	-7.4
CM614	Vietnam	Coc Muoi	<i>Macaca sp.</i>	6	-14.2	11.91	-26.1	-5.9
CM618	Vietnam	Coc Muoi	<i>Macaca sp.</i>	6	-15.1	11.91	-27.0	-5.2
CM555	Vietnam	Coc Muoi	<i>Macaca sp.</i>	6	-15.4	11.91	-27.3	-4.7
CM534	Vietnam	Coc Muoi	<i>Panthera tigris</i>	212	-16.2	14.53	-30.7	-7.5
CM566	Vietnam	Coc Muoi	<i>Panthera tigris</i>	212	-14.6	14.53	-29.1	-5.4
CM632	Vietnam	Coc Muoi	<i>Panthera tigris</i>	212	-16	14.53	-30.5	-8.6
CM571	Vietnam	Coc Muoi	<i>Cuon alpinus</i>	15	-14.3	13.39	-27.7	-6.7
CM535	Vietnam	Coc Muoi	Small-sized Felidae	18	-14.5	13.46	-28.0	-8.6
CM536	Vietnam	Coc Muoi	Small-sized Felidae	18	-13.2	13.46	-26.7	-6.2
CM421	Vietnam	Coc Muoi	<i>Hystrix sp.</i>	12	-14.8	12.18	-27.0	-8.1
CM472	Vietnam	Coc Muoi	<i>Hystrix sp.</i>	12	-15.8	12.18	-28.0	-3.9
CM902	Vietnam	Coc Muoi	<i>Hystrix sp.</i>	12	-16	12.18	-28.2	-7.2
CM903	Vietnam	Coc Muoi	<i>Hystrix sp.</i>	12	-14.5	12.18	-26.7	-7.5
CM904	Vietnam	Coc Muoi	<i>Hystrix sp.</i>	12	-16	12.18	-28.2	-4.1
CM910	Vietnam	Coc Muoi	<i>Hystrix sp.</i>	12	-13.6	12.18	-25.8	-8.6
CM911	Vietnam	Coc Muoi	<i>Hystrix sp.</i>	12	-13.7	12.18	-25.9	-4.6
CM954	Vietnam	Coc Muoi	<i>Hystrix sp.</i>	12	-14.7	12.18	-26.9	-4.9
CM983	Vietnam	Coc Muoi	<i>Atherurus sp.</i>	12	-14.6	12.18	-26.8	-5.8
CM984	Vietnam	Coc Muoi	<i>Atherurus sp.</i>	12	-14.1	12.18	-26.3	-4.6
CM75	Vietnam	Coc Muoi	<i>Rusa unicolor</i>	220	-17.5	13.59	-31.1	-6.8
CM244	Vietnam	Coc Muoi	<i>Rusa unicolor</i>	220	-13.4	13.59	-27.0	-6.9
CM245	Vietnam	Coc Muoi	<i>Rusa unicolor</i>	220	-15.2	13.59	-28.8	-6.5
CM246	Vietnam	Coc Muoi	<i>Rusa unicolor</i>	220	-14	13.59	-27.6	-5.8
CM247	Vietnam	Coc Muoi	<i>Rusa unicolor</i>	220	-17	13.59	-30.6	-7.8
CM249	Vietnam	Coc Muoi	<i>Rusa unicolor</i>	220	-16.3	13.59	-29.9	-7.3
CM287	Vietnam	Coc Muoi	<i>Capricornis sp.</i>	112	-15	13.14	-28.1	-5.9
CM173	Vietnam	Coc Muoi	<i>Capricornis sp.</i>	112	-17.7	13.14	-30.8	-5.1
CM286	Vietnam	Coc Muoi	<i>Capricornis sp.</i>	112	-17.9	13.14	-31.0	-7.6
CM147	Vietnam	Coc Muoi	<i>Capricornis sp.</i>	112	-14.5	13.14	-27.6	-4.9
CM149	Vietnam	Coc Muoi	<i>Capricornis sp.</i>	112	-17.1	13.14	-30.2	-5.5

CM370	Vietnam	Coc Muoi	<i>Bos cf. sauveli</i>	800	-14.8	14.5	-29.3	-5.8
CM225	Vietnam	Coc Muoi	<i>Bos cf. sauveli</i>	800	-8.9	14.5	-23.4	-6.6
CM226	Vietnam	Coc Muoi	<i>Bos cf. sauveli</i>	800	-11.1	14.5	-25.6	-5.9
CM227	Vietnam	Coc Muoi	<i>Bos cf. sauveli</i>	800	-11.3	14.5	-25.8	-6.9
CM46	Vietnam	Coc Muoi	<i>Bos cf. sauveli</i>	800	-3.6	14.5	-18.1	-4.1
CM152a	Vietnam	Coc Muoi	Bovidae (? <i>Bos sauveli</i> )	875	-13.4	14.57	-28.0	-5.9
CM1067	Vietnam	Coc Muoi	<i>Rhinoceros sondaicus</i>	1750	-17	14.28	-31.3	-5.4
CM1122	Vietnam	Coc Muoi	<i>Rhinoceros sondaicus</i>	1750	-16.6	14.28	-30.9	-4.9
CM1120	Vietnam	Coc Muoi	<i>Rhinoceros sondaicus</i>	1750	-15.6	14.28	-29.9	-5.8
CM1068	Vietnam	Coc Muoi	<i>Rhinoceros sondaicus</i>	1750	-15.5	14.28	-29.8	-5.8
CM1229	Vietnam	Coc Muoi	<i>Rhinoceros sondaicus</i>	1750	-16.3	14.28	-30.6	-4.7
CM1137	Vietnam	Coc Muoi	<i>Rhinoceros sondaicus</i>	1750	-17	14.28	-31.3	-7.8
CM1339	Vietnam	Coc Muoi	<i>Rhinoceros sondaicus</i>	1750	-16.5	14.28	-30.8	-5.4
CM998	Vietnam	Coc Muoi	<i>Rhinoceros sondaicus</i>	1750	-16.4	14.28	-30.7	-7.1
CM1151	Vietnam	Coc Muoi	<i>Rhinoceros unicornis</i>	2250	-16.3	14.4	-30.7	-5.8
CM1278	Vietnam	Coc Muoi	<i>Rhinoceros unicornis</i>	2250	-16.8	14.4	-31.2	-6.2
CM1035	Vietnam	Coc Muoi	<i>Dicerorhinus sumatrensis</i>	950	-15.8	14	-29.8	-6.4
CM1096	Vietnam	Coc Muoi	<i>Dicerorhinus sumatrensis</i>	950	-15.6	14	-29.6	-5
CM1349	Vietnam	Coc Muoi	<i>Megatapirus augustus</i>	500	-20.1	13.72	-33.8	-5.9
CM514	Vietnam	Coc Muoi	<i>Megatapirus augustus</i>	500	-18.7	13.72	-32.4	-5.5
CM515	Vietnam	Coc Muoi	<i>Tapirus indicus</i>	300	-17.8	13.5	-31.3	-6.2
CM516	Vietnam	Coc Muoi	<i>Tapirus indicus</i>	300	-18.3	13.5	-31.8	-6.8
CM1351	Vietnam	Coc Muoi	<i>Tapirus indicus</i>	300	-16.4	13.5	-29.9	-6.6
CM726	Vietnam	Coc Muoi	<i>Stegodon sp.</i>	4000	-14.2	14.66	-28.9	-7.9
CM637	Vietnam	Coc Muoi	<i>Stegodon sp.</i>	4000	-14.7	14.66	-29.4	-9.8
CM728	Vietnam	Coc Muoi	<i>Elephas maximus</i>	4250	-18.3	14.69	-33.0	-5.8
420	Vietnam	Coc Muoi	<i>Elephas maximus</i>	4250	-15.8	14.69	-30.5	-6
TH-860	Laos	Tam Hang South	<i>Cuon alpinus cf. antiquus</i>	15	-12.9	13.39	-26.3	-6.8
TH-861	Laos	Tam Hang South	<i>Cuon alpinus cf. antiquus</i>	15	-14.4	13.39	-27.8	-2.9
TH-870	Laos	Tam Hang South	<i>Cuon alpinus cf. antiquus</i>	15	-13	13.39	-26.4	-6.3
TH-539	Laos	Tam Hang South	<i>Capricornis sumatraensis</i>	112	-6.8	13.14	-19.9	-4.3
TH-767	Laos	Tam Hang South	<i>Capricornis sumatraensis</i>	112	-4.9	13.14	-18.0	-2.8
TH-768	Laos	Tam Hang South	<i>Capricornis sumatraensis</i>	112	-13.7	13.14	-26.8	-5.7
TH-873	Laos	Tam Hang South	<i>Helarctos malayanus</i>	50	-12.6	13.3	-25.9	-5.9
TH-877	Laos	Tam Hang South	<i>Helarctos malayanus</i>	50	-12.5	13.3	-25.8	-7.2
TH-H1	Laos	Tam Hang South	<i>Hystrix brachyura</i>	12	-13.7	12.18	-25.9	-5.3
TH-H2	Laos	Tam Hang South	<i>Hystrix brachyura</i>	12	-9.9	12.18	-22.1	-9.4
TH-H3	Laos	Tam Hang South	<i>Hystrix brachyura</i>	12	-10.2	12.18	-22.4	-5.4
TH-H4	Laos	Tam Hang South	<i>Hystrix brachyura</i>	12	-8.9	12.18	-21.1	-8.9
TH-H5	Laos	Tam Hang South	<i>Hystrix brachyura</i>	12	-14.7	12.18	-26.9	-6.7
TH-H6	Laos	Tam Hang South	<i>Hystrix brachyura</i>	12	-14	12.18	-26.2	-6.6

TH-789	Laos	Tam Hang South	<i>Bubalus bubalis</i>	1000	-2.9	14.66	-17.6	-6.7
TH-791	Laos	Tam Hang South	<i>Bubalus bubalis</i>	1000	-5	14.66	-19.7	-6.8
TH-455	Laos	Tam Hang South	<i>Bos cf. sauveli</i>	800	1.8	14.5	-12.7	-5.7
TH-458	Laos	Tam Hang South	<i>Bos cf. sauveli</i>	800	-10.9	14.5	-25.4	-8.2
TH-459	Laos	Tam Hang South	<i>Bos cf. sauveli</i>	800	-11.8	14.5	-26.3	-7.3
TH-546	Laos	Tam Hang South	<i>Bos cf. sauveli</i>	800	-14.9	14.5	-29.4	-6.4
TH-573	Laos	Tam Hang South	<i>Bos cf. sauveli</i>	800	-9.9	14.5	-24.4	-7.1
TH-790	Laos	Tam Hang South	<i>Bos cf. sauveli</i>	800	-12.9	14.5	-27.4	-8.2
TH-799	Laos	Tam Hang South	<i>Bos cf. sauveli</i>	800	-2.8	14.5	-17.3	-6.3
TH-72	Laos	Tam Hang South	<i>Macaca sp.</i>	6	-14	11.91	-25.9	-5.7
TH-73	Laos	Tam Hang South	<i>Macaca sp.</i>	6	-13.4	11.91	-25.3	-5
TH-74	Laos	Tam Hang South	<i>Macaca sp.</i>	6	-14.1	11.91	-26.0	-5
TH-75	Laos	Tam Hang South	<i>Macaca sp.</i>	6	-14	11.91	-25.9	-5.3
TH-76	Laos	Tam Hang South	<i>Macaca sp.</i>	6	-13.5	11.91	-25.4	-5.1
TH-79	Laos	Tam Hang South	<i>Macaca sp.</i>	6	-14.2	11.91	-26.1	-5.3
TH-593	Laos	Tam Hang South	<i>Megatapirus augustus</i>	500	-15.8	13.72	-29.5	-7.5
TH-213	Laos	Tam Hang South	<i>Muntiacus muntjak</i>	24	-13.2	12.17	-25.4	-6.8
TH-215	Laos	Tam Hang South	<i>Muntiacus muntjak</i>	24	-12.6	12.17	-24.8	-7.5
TH-216	Laos	Tam Hang South	<i>Muntiacus muntjak</i>	24	-12.8	12.17	-25.0	-7.7
TH-219	Laos	Tam Hang South	<i>Muntiacus muntjak</i>	24	-13.3	12.17	-25.5	-8.1
TH-130	Laos	Tam Hang South	<i>Panthera tigris</i>	212	-14.1	14.53	-28.6	-7.8
TH-132	Laos	Tam Hang South	<i>Panthera tigris</i>	212	-14	14.53	-28.5	-7.4
TH-376	Laos	Tam Hang South	<i>Rhinoceros spp.</i>	2000	-13.5	14.34	-27.8	-6.8
TH-379	Laos	Tam Hang South	<i>Rhinoceros spp.</i>	2000	-13.3	14.34	-27.6	-7
TH-576	Laos	Tam Hang South	<i>Rhinoceros spp.</i>	2000	-14.4	14.34	-28.7	-7.7
TH-445	Laos	Tam Hang South	<i>Rusa unicolor</i>	220	-14.5	13.59	-28.1	-5
TH-469	Laos	Tam Hang South	<i>Rusa unicolor</i>	220	-12.7	13.59	-26.3	-6.8
TH-499	Laos	Tam Hang South	<i>Rusa unicolor</i>	220	-4.8	13.59	-18.4	-8.2
TH-549	Laos	Tam Hang South	<i>Rusa unicolor</i>	220	-15.8	13.59	-29.4	-6.1
TH-555	Laos	Tam Hang South	<i>Rusa unicolor</i>	220	-16.2	13.59	-29.8	-6.7
TH-717	Laos	Tam Hang South	<i>Rusa unicolor</i>	220	-15	13.59	-28.6	-8.8
TH-719	Laos	Tam Hang South	<i>Rusa unicolor</i>	220	-13.5	13.59	-27.1	-6.5
TH-748	Laos	Tam Hang South	<i>Rusa unicolor</i>	220	-13	13.59	-26.6	-8.6
TH-390-2	Laos	Tam Hang South	<i>Sus scrofa</i>	137	-12.7	13.19	-25.9	-8.6
TH-410	Laos	Tam Hang South	<i>Sus scrofa</i>	137	-12.6	13.19	-25.8	-8.2
TH-420	Laos	Tam Hang South	<i>Sus scrofa</i>	137	-1.4	13.19	-14.6	-6.3
TH-427	Laos	Tam Hang South	<i>Sus scrofa</i>	137	-12	13.19	-25.2	-6.2
TH-430	Laos	Tam Hang South	<i>Sus scrofa</i>	137	-11.8	13.19	-25.0	-8.5
TH-431	Laos	Tam Hang South	<i>Sus scrofa</i>	137	-12.3	13.19	-25.5	-6.7
TH-433	Laos	Tam Hang South	<i>Sus scrofa</i>	137	-12.7	13.19	-25.9	-8.9
TH-637	Laos	Tam Hang South	<i>Sus scrofa</i>	137	-13.2	13.19	-26.4	-7.8

TH-642	Laos	Tam Hang South	<i>Sus scrofa</i>	137	-11.2	13.19	-24.4	-5.3
TH-645-2	Laos	Tam Hang South	<i>Sus scrofa</i>	137	-12.3	13.19	-25.5	-8.1
TH-646	Laos	Tam Hang South	<i>Sus scrofa</i>	137	-12.7	13.19	-25.9	-8.5
TH-371	Laos	Tam Hang South	<i>Tapirus indicus cf. intermedius</i>	300	-16.5	13.5	-30.0	-6.5
TH-378	Laos	Tam Hang South	<i>Tapirus indicus cf. intermedius</i>	300	-16.3	13.5	-29.8	-5.6
TH-129	Laos	Tam Hang South	<i>Ursus thibetanus cf. kokeni</i>	100	-14.4	13.3	-27.7	-8.5
TH-139	Laos	Tam Hang South	<i>Ursus thibetanus cf. kokeni</i>	100	-14.4	13.3	-27.7	-6.1



**Annex S2.** Faunal lists from already-published sites (Bacon et al., 2018b, Bourgon et al. 2020) of the same latitudinal band used as comparison in the present study (Nam Lot and Tam Hay Marklot), with associated  $\delta^{13}\text{C}_{\text{apatite}}$ ,  $\delta^{13}\text{C}_{\text{carbon source}}$  and  $\delta^{18}\text{O}_{\text{apatite}}$  values, as well as body mass and  $\delta^{13}\text{C}$  (‰) Enrichment Factor used to obtain  $\delta^{13}\text{C}_{\text{carbon source}}$ . (\*) The incisor NL 433 has been identified by using palaeoproteomics.

Number	Country	Site	Taxon	Body Mass (kg)	$\delta^{13}\text{C}_{\text{apatite}}$ (‰ VPDB)	$\delta^{13}\text{C}$ (‰) Enrichment Factor	$\delta^{13}\text{C}_{\text{carbon source}}$ (‰ VPDB)	$\delta^{18}\text{O}_{\text{apatite}}$ (‰ VPDB)
NL-8	Laos	Nam Lot	<i>Capricornis sumatraensis</i>	112	-14.6	13.14	-27.7	-4.1
NL-9	Laos	Nam Lot	<i>Capricornis sumatraensis</i>	112	-13	13.14	-26.1	-3.3
NL-17	Laos	Nam Lot	<i>Rusa unicolor</i>	220	-12.9	13.59	-26.5	-8.7
NL-19	Laos	Nam Lot	<i>Rusa unicolor</i>	220	-7.5	13.59	-21.1	-5.8
NL-29	Laos	Nam Lot	<i>Rusa unicolor</i>	220	-14.1	13.59	-27.7	-6.7
NL-22	Laos	Nam Lot	<i>Rusa unicolor</i>	220	-0.9	13.59	-14.5	-5.5
NL-24	Laos	Nam Lot	<i>Rusa unicolor</i>	220	-2.7	13.59	-16.3	-6.1
NL-63-1	Laos	Nam Lot	<i>Muntiacus muntjak</i>	24	-13.4	12.17	-25.6	-6.4
NL-65-1	Laos	Nam Lot	<i>Muntiacus muntjak</i>	24	-13.8	12.17	-26.0	-7.3
NL-69	Laos	Nam Lot	<i>Muntiacus muntjak</i>	24	-13.9	12.17	-26.1	-3.3
NL-116	Laos	Nam Lot	Bovidae indet.	875	-9.8	14.57	-24.4	-5.9
NL-117	Laos	Nam Lot	Bovidae indet.	875	0.3	14.57	-14.3	-4.9
NL-125	Laos	Nam Lot	Bovidae indet.	875	-13.5	14.57	-28.1	-4.7
NL-161	Laos	Nam Lot	Rhinocerotidae indet.	1633	-13.3	14.25	-27.6	-5.9
NL-162	Laos	Nam Lot	Rhinocerotidae indet.	1633	-13.1	14.25	-27.4	-2.5
NL-254-1-1	Laos	Nam Lot	Rhinocerotidae indet.	1633	-12.7	14.25	-27.0	-6.7
NL-256-1	Laos	Nam Lot	Rhinocerotidae indet.	1633	-15.1	14.25	-29.4	-6.6
NL-256-2	Laos	Nam Lot	Rhinocerotidae indet.	1633	-16.3	14.25	-30.6	-4.9
NL-256-3	Laos	Nam Lot	Rhinocerotidae indet.	1633	-15.1	14.25	-29.4	-6.7
NL-139	Laos	Nam Lot	<i>Bubalus bubalis</i>	1000	-4.7	14.66	-19.4	-6.5
NL-143	Laos	Nam Lot	<i>Bubalus bubalis</i>	1000	-9.5	14.66	-24.2	-4.4
NL-186	Laos	Nam Lot	<i>Ailuropoda melanoleuca</i>	92	-14.5	10.51	-25.0	-6.2
NL-277	Laos	Nam Lot	<i>Ailuropoda melanoleuca</i>	92	-14.9	10.51	-25.4	-4.2
NL-162	Laos	Nam Lot	<i>Sus</i> sp.	137	-13.7	13.19	-26.9	-5.7
NL-208	Laos	Nam Lot	<i>Sus</i> sp.	137	-12.4	13.19	-25.6	-6
NL-216	Laos	Nam Lot	<i>Sus</i> sp.	137	-12.8	13.19	-26.0	-5.3
NL-218	Laos	Nam Lot	<i>Sus</i> sp.	137	-14.1	13.19	-27.3	-6.8
NL-SS-1	Laos	Nam Lot	<i>Sus</i> sp.	137	-13.1	13.19	-26.3	-6
NL-258	Laos	Nam Lot	<i>Tapirus</i> sp.	300	-17.5	13.5	-31.0	-6.4
NL-259	Laos	Nam Lot	<i>Tapirus</i> sp.	300	-14.9	13.5	-28.4	-6.8
NL-260	Laos	Nam Lot	<i>Tapirus</i> sp.	300	-15.3	13.5	-28.8	-5
NL-286	Laos	Nam Lot	<i>Cuon alpinus</i>	15	-13.3	13.39	-26.7	-6.4
NL-368	Laos	Nam Lot	<i>Cuon alpinus</i>	15	-13	13.39	-26.4	-3

NL-269	Laos	Nam Lot	<i>Ursus thibetanus cf. kokeni</i>	100	-15.3	13.3	-28.6	-9
NL-271	Laos	Nam Lot	<i>Ursus thibetanus cf. kokeni</i>	100	-14.1	13.3	-27.4	-8.3
NL-275	Laos	Nam Lot	<i>Ursus thibetanus cf. kokeni</i>	100	-12.6	13.3	-25.9	-6.5
NL-310	Laos	Nam Lot	<i>Ursus thibetanus cf. kokeni</i>	100	-13.4	13.3	-26.7	-3.1
NL-288	Laos	Nam Lot	<i>Crocota crocuta</i>	70	-13.7	14.04	-27.7	-6.4
NL-295	Laos	Nam Lot	<i>Crocota crocuta</i>	70	-13.6	14.04	-27.6	-6.1
NL-433*	Laos	Nam Lot	<i>Pongo sp.</i>	55	-14.5	12.78	-27.3	-3.3
NL-302	Laos	Nam Lot	<i>Pongo sp.</i>	55	-14.5	12.78	-27.3	-6.1
NL-297	Laos	Nam Lot	<i>Macaca sp.</i>	6	-14.1	11.91	-26.0	-4.8
NL-314	Laos	Nam Lot	<i>Macaca sp.</i>	6	-11.5	11.91	-23.4	-3.7
NL-323	Laos	Nam Lot	<i>Macaca sp.</i>	6	-14.8	11.91	-26.7	-6.1
NL-357	Laos	Nam Lot	<i>Macaca sp.</i>	6	-14.6	11.91	-26.5	-3.6
NL-362	Laos	Nam Lot	<i>Elephas sp.</i>	4250	-16.1	14.69	-30.8	-6.2
NL-365	Laos	Nam Lot	<i>Stegodon orientalis</i>	4000	-18.2	14.66	-32.9	-4.3
NL-367	Laos	Nam Lot	<i>Stegodon orientalis</i>	4000	-15.4	14.66	-30.1	-6.5
NL-369	Laos	Nam Lot	<i>Hystrix sp.</i>	12	-13.6	12.18	-25.8	-6.5
NL-385	Laos	Nam Lot	<i>Hystrix sp.</i>	12	-12.1	12.18	-24.3	-5.7
NL-392	Laos	Nam Lot	<i>Hystrix sp.</i>	12	-14.1	12.18	-26.3	-5.2
NL-397	Laos	Nam Lot	<i>Hystrix sp.</i>	12	-13.3	12.18	-25.5	-5.7
NLII-1	Laos	Nam Lot	<i>Crocota crocuta</i>	70	-14.3	14.04	-28.3	-6.6
NLII-2	Laos	Nam Lot	<i>Crocota crocuta</i>	70	-12.5	14.04	-26.5	-6.8
NLII-3	Laos	Nam Lot	<i>Crocota crocuta</i>	70	-15.1	14.04	-29.1	-6.2
NLII-4	Laos	Nam Lot	<i>Crocota crocuta</i>	70	-11.8	14.04	-25.8	-7.2
NLII-5	Laos	Nam Lot	Felidae ( <i>Neofelis nebulosa</i> ?)	18	-15	13.46	-28.5	-6.6
MI-20	Laos	Tam Hay Marklot	<i>Capricornis cf. sumatraensis</i>	112	-13.4	13.14	-26.5	-2.2
MI-21	Laos	Tam Hay Marklot	<i>Capricornis cf. sumatraensis</i>	112	-14.7	13.14	-27.8	-5.2
MI-22	Laos	Tam Hay Marklot	<i>Capricornis cf. sumatraensis</i>	112	-15.2	13.14	-28.3	-5.2
MI-23	Laos	Tam Hay Marklot	<i>Capricornis cf. sumatraensis</i>	112	-13.5	13.14	-26.6	-7
MI-24	Laos	Tam Hay Marklot	<i>Capricornis cf. sumatraensis</i>	112	-14.6	13.14	-27.7	-7.6
MI-25	Laos	Tam Hay Marklot	<i>Naemorhedus cf. caudatus</i>	27	-2.4	12.24	-14.6	0.2
MI-26	Laos	Tam Hay Marklot	<i>Naemorhedus cf. caudatus</i>	27	-3.9	12.24	-16.1	-2.5
MI-27	Laos	Tam Hay Marklot	<i>Naemorhedus cf. caudatus</i>	27	-3.7	12.24	-15.9	-1.8
MI-28	Laos	Tam Hay Marklot	<i>Naemorhedus cf. caudatus</i>	27	-2.5	12.24	-14.7	-1.6
MI-103	Laos	Tam Hay Marklot	<i>Helarctos malayanus</i>	50	-14.7	12.59	-27.3	-3.9
MI-121	Laos	Tam Hay Marklot	<i>Helarctos malayanus</i>	50	-14.9	12.59	-27.5	-5.5
MI-117	Laos	Tam Hay Marklot	<i>Ursus thibetanus</i>	100	-15.4	13.3	-28.7	-7.4
MI-119	Laos	Tam Hay Marklot	<i>Ursus thibetanus</i>	100	-13.3	13.3	-26.6	-6.6
MI-122	Laos	Tam Hay Marklot	<i>Ursus thibetanus</i>	100	-14.4	13.3	-27.7	-6.3
MI-134	Laos	Tam Hay Marklot	<i>Panthera pardus</i>	41	-7.9	13.81	-21.7	-7.3
MI-135	Laos	Tam Hay Marklot	<i>Panthera pardus</i>	41	-4	13.81	-17.8	-7.3
MI-136	Laos	Tam Hay Marklot	<i>Panthera pardus</i>	41	-13.8	13.81	-27.6	-6.8

MI-166	Laos	Tam Hay Marklot	<i>Rusa unicolor</i>	220	-7.5	13.59	-21.1	-5.7
MI-180	Laos	Tam Hay Marklot	<i>Rusa unicolor</i>	220	-5.3	13.59	-18.9	-4.4
MI-185	Laos	Tam Hay Marklot	<i>Rusa unicolor</i>	220	-6	13.59	-19.6	-4.9
MI-187	Laos	Tam Hay Marklot	<i>Rusa unicolor</i>	220	-3.4	13.59	-17.0	-4.9
MI-191	Laos	Tam Hay Marklot	<i>Rusa unicolor</i>	220	-7.9	13.59	-21.5	-6.1
MI-512	Laos	Tam Hay Marklot	<i>Rucervus eldii</i>	123	2.4	13.19	-10.8	-5.3
MI-595	Laos	Tam Hay Marklot	<i>Rucervus eldii</i>	123	1.8	13.19	-11.4	-3.1
MI-556	Laos	Tam Hay Marklot	<i>Axis cf. porcinus</i>	43	-0.8	12.53	-13.3	-5.8
MI-557	Laos	Tam Hay Marklot	<i>Axis cf. porcinus</i>	43	-0.6	12.53	-13.1	-5.3
MI-627	Laos	Tam Hay Marklot	<i>Muntiacus sp.</i>	24	-13.8	12.17	-26.0	-7.4
MI-628	Laos	Tam Hay Marklot	<i>Muntiacus sp.</i>	24	-14.8	12.17	-27.0	-8.1
MI-629	Laos	Tam Hay Marklot	<i>Muntiacus sp.</i>	24	-12.7	12.17	-24.9	-8.1
MI-630	Laos	Tam Hay Marklot	<i>Muntiacus sp.</i>	24	-14.4	12.17	-26.6	-5
MI-631	Laos	Tam Hay Marklot	<i>Muntiacus sp.</i>	24	-14.5	12.17	-26.7	-7.5
MI-650	Laos	Tam Hay Marklot	<i>Bubalus bubalis</i>	1000	-10.3	14.66	-25.0	-6
MI-651	Laos	Tam Hay Marklot	<i>Bubalus bubalis</i>	1000	-4	14.66	-18.7	-6.6
MI-652	Laos	Tam Hay Marklot	<i>Bubalus bubalis</i>	1000	1	14.66	-13.7	-4.7
MI-653	Laos	Tam Hay Marklot	<i>Bubalus bubalis</i>	1000	-10.9	14.66	-25.6	-6.3
MI-654	Laos	Tam Hay Marklot	<i>Bubalus bubalis</i>	1000	0.1	14.66	-14.6	-5.9
MI-655	Laos	Tam Hay Marklot	<i>Bos sp.</i>	800	-2.5	14.5	-17.0	-4.2
MI-656	Laos	Tam Hay Marklot	<i>Bos sp.</i>	800	-14.4	14.5	-28.9	-6.1
MI-657	Laos	Tam Hay Marklot	<i>Bos sp.</i>	800	-0.3	14.5	-14.8	-4.6
MI-658	Laos	Tam Hay Marklot	<i>Bos sp.</i>	800	-10	14.5	-24.5	-7.6
MI-659	Laos	Tam Hay Marklot	<i>Bos sp.</i>	800	-13.1	14.5	-27.6	-5.9
MI-130	Laos	Tam Hay Marklot	<i>Panthera tigris</i>	212	-4.3	14.53	-18.8	-3.2
MI-693	Laos	Tam Hay Marklot	<i>Panthera tigris</i>	212	-6.9	14.53	-21.4	-3.5
MI-694	Laos	Tam Hay Marklot	<i>Panthera tigris</i>	212	-10	14.53	-24.5	-6.8
MI-662	Laos	Tam Hay Marklot	<i>Sus sp.</i>	137	-7.6	13.19	-20.8	-7.5
MI-663	Laos	Tam Hay Marklot	<i>Sus sp.</i>	137	-8.6	13.19	-21.8	-10
MI-664	Laos	Tam Hay Marklot	<i>Sus sp.</i>	137	-6	13.19	-19.2	-5.9
MI-665	Laos	Tam Hay Marklot	<i>Sus sp.</i>	137	-13.5	13.19	-26.7	-5.8
MI-666	Laos	Tam Hay Marklot	<i>Sus sp.</i>	137	-14	13.19	-27.2	-7.6
MI-667	Laos	Tam Hay Marklot	<i>Sus sp.</i>	137	-13.2	13.19	-26.4	-5.4
MI-131	Laos	Tam Hay Marklot	<i>Cuon alpinus</i>	15	-16	13.39	-29.4	-7.3
MI-681	Laos	Tam Hay Marklot	<i>Cuon alpinus</i>	15	-11.4	13.39	-24.8	-3.2
MI-683	Laos	Tam Hay Marklot	<i>Pongo sp.</i>	55	-14.8	12.78	-27.6	-4.1
MI-685	Laos	Tam Hay Marklot	<i>Pongo sp.</i>	55	-13.5	12.78	-26.3	-4.6
MI-682	Laos	Tam Hay Marklot	Canidae	15	-13.2	13.39	-26.6	-6
MI-684	Laos	Tam Hay Marklot	<i>Ailuropoda melanoleuca</i>	92	-16.7	10.51	-27.2	-6.3
MI-691	Laos	Tam Hay Marklot	Tapiridae	300	-15.5	13.5	-29.0	-5.9
MI-692	Laos	Tam Hay Marklot	<i>Tapirus sp.</i>	300	-11.3	13.5	-24.8	-7.9

MI-695	Laos	Tam Hay Marklot	<i>Macaca sp.</i>	6	-13.9	11.91	-25.8	-5.3
MI-696	Laos	Tam Hay Marklot	<i>Macaca sp.</i>	6	-14.2	11.91	-26.1	-5.3
MI-697	Laos	Tam Hay Marklot	<i>Macaca sp.</i>	6	-15.1	11.91	-27.0	-5.6
MI-698	Laos	Tam Hay Marklot	<i>Macaca sp.</i>	6	-13.3	11.91	-25.2	-4.7
MI-699	Laos	Tam Hay Marklot	<i>Macaca sp.</i>	6	-12.9	11.91	-24.8	-4.6
MI-700	Laos	Tam Hay Marklot	<i>Hystrix sp.</i>	12	-11.5	12.18	-23.7	-7.6
MI-701	Laos	Tam Hay Marklot	<i>Hystrix sp.</i>	12	-11.5	12.18	-23.7	-5.5
MI-702	Laos	Tam Hay Marklot	<i>Hystrix sp.</i>	12	-7.9	12.18	-20.1	-5.2
MI-703	Laos	Tam Hay Marklot	<i>Hystrix sp.</i>	12	-11.6	12.18	-23.8	-8.5
MI-686	Laos	Tam Hay Marklot	<i>Rhinoceros sondaicus</i>	1750	-16.2	14.28	-30.5	-6.5
MI-687	Laos	Tam Hay Marklot	<i>Rhinoceros sondaicus</i>	1750	-13.3	14.28	-27.6	-5.5
MI-688	Laos	Tam Hay Marklot	<i>Rhinoceros sondaicus</i>	1750	-15.6	14.28	-29.9	-6.3
MI-689	Laos	Tam Hay Marklot	<i>Rhinoceros sondaicus</i>	1750	-15.3	14.28	-29.6	-7.1
MI-690	Laos	Tam Hay Marklot	<i>Rhinoceros sondaicus</i>	1750	-14.6	14.28	-28.9	-7.5

**Annex S3.** Data from Pleistocene Chinese sites ( $\delta^{13}\text{C}_{\text{apatite}}$ ,  $\delta^{13}\text{C}_{\text{carbon source}}$  and  $\delta^{18}\text{O}_{\text{apatite}}$  values (‰ VPDB), body mass and  $\delta^{13}\text{C}$  (‰ VPDB) Enrichment Factor used to obtain  $\delta^{13}\text{C}_{\text{carbon source}}$ ).

**Yugong** cave, Guangxi Zhuang Autonomous region, southern China (~22° 14" N, 107° 23" E). Dated to the late Middle Pleistocene on the composition of the faunal assemblage, in comparison with those of other sites from south China (Sanhe, Boyueshan, Juyuan, Mohui) (Sun et al., 2019). Faunal list (non- exhaustive list): *Sus xiaozhu wenzhongii*, *Sus cf. pei*, *Muntiacus sp.*, *Cervus (Rusa) cf. unicolor*, Caprinae gen. et sp. indet., *Leptobos sp.*, *Bos (Bibos) sp.*, *Ailuropoda baconi*, *Ursus thibetanus*, *Stegodon sp.*

**Baxian** cave, Guangxi Zhuang Autonomous region, southern China (22° 34" N, 107° 21" E). Dated to the early Late Pleistocene on the similarity of the faunal assemblage with those from Zhiren, Fuyan, Daoxian, and Hunan sites (Ma et al., 2017; Sun et al., 2019). Faunal list: *Macaca sp.*, *Namascus sp.*, *Rhinopithecus sp.*, *Pongo sp.*, *Ailuropoda baconi*, *Ursus thibetanus*, *Arctonyx collaris*, *Panthera tigris*, *Elephas maximus*, *Stegodon orientalis*, *Rhinoceros sondaicus*, *Megatapirus augustus*, *Sus scrofa*, *Muntiacus sp.*, *Cervus (Rusa) sp.*, *Bos (Bibos) sp.*

**Quzai** cave, Guangxi Zhuang Autonomous region, southern China (22° 27" N, 107° 46" E). Dated to the early Late Pleistocene on the similarity of the faunal assemblage with those from Zhiren and Fuyan (Ma et al., 2019). Faunal list: *Macaca sp.*, *Namascus sp.*, *Rhinopithecus sp.*, *Pongo sp.*, *Ailuropoda baconi*, *Ursus thibetanus*, *Arctonyx collaris*, *Felis sp.*, *Rhinoceros sondaicus*, *Megatapirus augustus*, *Elephas maximus*, *Stegodon orientalis*, *Sus scrofa*, *Cervus unicolor*, *Bos (Bibos) sp.*

Country	Site	Taxon	Body Mass (kg)	$\delta^{13}\text{C}$ (‰) Enrichment Factor	$\delta^{13}\text{C}_{\text{carbon source}}$ (‰ VPDB)	$\delta^{13}\text{C}_{\text{apatite}}$ (‰ VPDB)	$\delta^{18}\text{O}_{\text{apatite}}$ (‰ VPDB)
China	Yugong	<i>Cervus sp.</i>	220	13.59	-27.59	-14	-6.7
China	Yugong	<i>Cervus sp.</i>	220	13.59	-25.49	-11.9	-7.9
China	Yugong	<i>Cervus sp.</i>	220	13.59	-27.49	-13.9	-5.9
China	Yugong	<i>Cervus sp.</i>	220	13.59	-33.79	-20.2	-6.2
China	Yugong	<i>Leptobos sp.</i>	320	13.85	-33.05	-19.2	-5.7
China	Yugong	<i>Leptobos sp.</i>	320	13.85	-30.85	-17	-6.5
China	Yugong	<i>Leptobos sp.</i>	320	13.85	-27.65	-13.8	-5.9
China	Yugong	<i>Leptobos sp.</i>	320	13.85	-20.05	-6.2	-6.5
China	Yugong	<i>Rhinoceros sondaicus</i>	1750	14.28	-30.48	-16.2	-6.6
China	Yugong	<i>Rhinoceros sondaicus</i>	1750	14.28	-30.48	-16.2	-7
China	Yugong	<i>Rhinoceros sondaicus</i>	1750	14.28	-29.28	-15	-8.2

China	Yugong	<i>Rhinoceros sondaicus</i>	1750	14.28	-29.08	-14.8	-6.4
China	Yugong	<i>Stegodon</i> sp.	4000	14.66	-30.16	-15.5	-7.1
China	Yugong	<i>Stegodon</i> sp.	4000	14.66	-30.36	-15.7	-8.3
China	Yugong	<i>Stegodon</i> sp.	4000	14.66	-31.26	-16.6	-8.2
China	Yugong	<i>Ailuropoda baconi</i>	92	10.51	-26.51	-16	-6.2
China	Yugong	<i>Ailuropoda baconi</i>	92	10.51	-26.91	-16.4	-5.7
China	Yugong	<i>Ailuropoda baconi</i>	92	10.51	-28.01	-17.5	-4.1
China	Yugong	<i>Ailuropoda baconi</i>	92	10.51	-27.61	-17.1	-6.2
China	Yugong	<i>Ailuropoda baconi</i>	92	10.51	-28.81	-18.3	-6.3
China	Yugong	<i>Ailuropoda baconi</i>	92	10.51	-27.91	-17.4	-5.3
China	Yugong	<i>Ailuropoda baconi</i>	100	13.3	-26.9	-13.6	-5.7
China	Yugong	<i>Ailuropoda baconi</i>	100	13.3	-26.7	-13.4	-5.6
China	Yugong	Suidae	137	13.19	-28.79	-15.6	-6.9
China	Yugong	Suidae	137	13.19	-27.49	-14.3	-6.4
China	Yugong	Suidae	137	13.19	-27.79	-14.6	-6.5
China	Yugong	Suidae	137	13.19	-25.59	-12.4	-7.7
China	Yugong	Suidae	137	13.19	-26.59	-13.4	-7.7
China	Yugong	Suidae	137	13,19	-27,99	-14,8	-8,7
China	Baxian	<i>Elephas maximus</i>	4250	14,69	-29,19	-14,5	-9
China	Baxian	<i>Elephas maximus</i>	4250	14,69	-31,29	-16,6	-6,8
China	Baxian	<i>Elephas maximus</i>	4250	14,69	-30,79	-16,1	-6,3
China	Baxian	<i>Elephas maximus</i>	4250	14,69	-31,49	-16,8	-6,6
China	Baxian	<i>Elephas maximus</i>	4250	14,69	-30,09	-15,4	-8,4
China	Baxian	<i>Elephas maximus</i>	4250	14,69	-32,29	-17,6	-6,8
China	Baxian	<i>Elephas maximus</i>	4250	14,69	-30,49	-15,8	-5,9
China	Baxian	<i>Elephas maximus</i>	4250	14,69	-28,99	-14,3	-8,5
China	Baxian	<i>Elephas maximus</i>	4250	14,69	-31,89	-17,2	-7,2
China	Baxian	<i>Elephas maximus</i>	4250	14.69	-29.79	-15.1	-6.4
China	Baxian	<i>Elephas maximus</i>	4250	14.69	-29.19	-14.5	-8.2
China	Baxian	<i>Cervus</i> sp.	220	13.59	-32.89	-19.3	-7.3
China	Baxian	<i>Cervus</i> sp.	220	13.59	-30.59	-17	-7.1
China	Baxian	<i>Muntiacus</i> sp.	24	12.17	-26.57	-14.4	-7.1
China	Baxian	<i>Muntiacus</i> sp.	24	12.17	-27.77	-15.6	-4.6
China	Baxian	<i>Muntiacus</i> sp.	24	12.17	-27.67	-15.5	-5.8
China	Baxian	<i>Sus scrofa</i>	137	13.19	-28.89	-15.7	-7.8

China	Baxian	<i>Sus scrofa</i>	137	13.19	-27.79	-14.6	-6.9
China	Baxian	<i>Sus scrofa</i>	137	13.19	-26.09	-12.9	-6.7
China	Baxian	<i>Rhinoceros sondaicus</i>	1750	14.28	-31.38	-17.1	-7.3
China	Baxian	<i>Rhinoceros sondaicus</i>	1750	14.28	-30.58	-16.3	-5.5
China	Baxian	<i>Rhinoceros sondaicus</i>	1750	14.28	-31.48	-17.2	-6.8
China	Baxian	<i>Rhinoceros sondaicus</i>	1750	14.28	-32.38	-18.1	-9.2
China	Baxian	<i>Rhinoceros sondaicus</i>	1750	14.28	-30.58	-16.3	-8.5
China	Baxian	<i>Megatapirus augustus</i>	500	13.72	-31.22	-17.5	-6.1
China	Baxian	<i>Stegodon orientalis</i>	4000	14.66	-30.56	-15.9	-8.4
China	Baxian	<i>Stegodon orientalis</i>	4000	14.66	-30.16	-15.5	-8.2
China	Baxian	<i>Bos (Bibos) sp.</i>	800	14.5	-29.2	-14.7	-6.6
China	Baxian	<i>Macaca sp.</i>	6	11.91	-27.81	-15.9	-7.1
China	Baxian	<i>Ailuropoda baconi</i>	92	10.51	-26.41	-15.9	-8
China	Baxian	<i>Ailuropoda baconi</i>	92	10.51	-28.51	-18	-7.1
China	Baxian	<i>Ailuropoda baconi</i>	92	10.51	-27.81	-17.3	-6.4
China	Baxian	<i>Cervus sp.</i>	220	13.59	-33.59	-20	-4.7
China	Baxian	<i>Cervus sp.</i>	220	13.59	-26.69	-13.1	-5.9
China	Baxian	Suidae	137	13.19	-27.99	-14.8	-9.1
China	Baxian	Suidae	137	13.19	-28.29	-15.1	-6.9
China	Baxian	Suidae	137	13.19	-27.89	-14.7	-7.1
China	Baxian	<i>Ursus thibetanus</i>	100	13.3	-30	-16.7	-6.9
China	Baxian	<i>Ursus thibetanus</i>	100	13.3	-29.8	-16.5	-8.1
China	Baxian	<i>Ursus thibetanus</i>	100	13.3	-28.4	-15.1	-9.8
China	Baxian	<i>Ursus thibetanus</i>	100	13.3	-28.7	-15.4	-7.5
China	Baxian	<i>Ursus thibetanus</i>	100	13.3	-28.8	-15.5	-7.2
China	Baxian	<i>Ailuropoda melanoleuca</i>	92	10.51	-25.41	-14.9	-3
China	Baxian	<i>Ailuropoda melanoleuca</i>	92	10.51	-28.71	-18.2	-2
China	Baxian	<i>Ailuropoda melanoleuca</i>	92	10.51	-28.11	-17.6	-4.2
China	Baxian	<i>Ailuropoda melanoleuca</i>	92	10.51	-28.11	-17.6	-2.7
China	Baxian	<i>Ailuropoda melanoleuca</i>	92	10.51	-28.11	-17.6	-4.6
China	Baxian	<i>Ailuropoda melanoleuca</i>	92	10.51	-27.81	-17.3	-5.3
China	Baxian	<i>Ailuropoda melanoleuca</i>	92	10.51	-28.81	-18.3	-4.7
China	Baxian	<i>Ailuropoda melanoleuca</i>	92	10.51	-27.81	-17.3	-4.7
China	Baxian	<i>Ailuropoda melanoleuca</i>	92	10.51	-27.81	-17.3	-5.8
China	Baxian	<i>Ailuropoda melanoleuca</i>	92	10.51	-26.91	-16.4	-6.5

China	Ouzai	<i>Elephas maximus</i>	4250	14.69	-27.79	-13.1	-4.7
China	Ouzai	<i>Elephas maximus</i>	4250	14.69	-30.19	-15.5	-5
China	Ouzai	<i>Elephas maximus</i>	4250	14.69	-31.49	-16.8	-5.6
China	Ouzai	<i>Elephas maximus</i>	4250	14.69	-32.59	-17.9	-7.5
China	Ouzai	<i>Elephas maximus</i>	4250	14.69	-30.59	-15.9	-6.2
China	Ouzai	<i>Elephas maximus</i>	4250	14.69	-26.59	-11.9	-6
China	Ouzai	<i>Elephas maximus</i>	4250	14.69	-28.69	-14	-5.3
China	Ouzai	<i>Elephas maximus</i>	4250	14.69	-31.19	-16.5	-5.7
China	Ouzai	<i>Elephas maximus</i>	4250	14.69	-28.59	-13.9	-4.6
China	Ouzai	<i>Elephas maximus</i>	4250	14.69	-28.79	-14.1	-5.1
China	Ouzai	<i>Stegodon orientalis</i>	4000	14.66	-31.26	-16.6	-7.1
China	Ouzai	<i>Stegodon orientalis</i>	4000	14.66	-29.36	-14.7	-4.1
China	Ouzai	<i>Stegodon orientalis</i>	4000	14.66	-29.96	-15.3	-7.2
China	Ouzai	<i>Stegodon orientalis</i>	4000	14.66	-31.06	-16.4	-7.9
China	Ouzai	<i>Stegodon orientalis</i>	4000	14.66	-30.96	-16.3	-7.2
China	Ouzai	<i>Stegodon orientalis</i>	4000	14.66	-31.36	-16.7	-5
China	Ouzai	<i>Stegodon orientalis</i>	4000	14.66	-31.06	-16.4	-5.7
China	Ouzai	<i>Bos (Bibos) sp.</i>	800	14.5	-34.4	-19.9	-5.4
China	Ouzai	<i>Bos (Bibos) sp.</i>	800	14.5	-31.1	-16.6	-5.9
China	Ouzai	<i>Rusa unicolor</i>	220	13.59	-29.89	-16.3	-4.3
China	Ouzai	<i>Rusa unicolor</i>	220	13.59	-25.89	-12.3	-5.7
China	Ouzai	<i>Rusa unicolor</i>	220	13.59	-31.09	-17.5	-3.2
China	Ouzai	<i>Rhinoceros sondaicus</i>	1750	14.28	-26.28	-12	-5.5
China	Ouzai	<i>Rhinoceros sondaicus</i>	1750	14.28	-29.98	-15.7	-6.5
China	Ouzai	<i>Rhinoceros sondaicus</i>	1750	14.28	-31.68	-17.4	-5.5
China	Ouzai	<i>Rhinoceros sondaicus</i>	1750	14.28	-30.18	-15.9	-6.6
China	Ouzai	<i>Megatapirus augustus</i>	500	13.72	-29.02	-15.3	-6.4
China	Ouzai	<i>Megatapirus augustus</i>	500	13.72	-29.42	-15.7	-7.1
China	Ouzai	<i>Ursus thibetanus</i>	100	13.3	-27	-13.7	-8.2
China	Ouzai	<i>Ursus thibetanus</i>	100	13.3	-25.3	-12	-7.8
China	Ouzai	<i>Ursus thibetanus</i>	100	13.3	-25.6	-12.3	-7.9
China	Ouzai	<i>Sus scrofa</i>	137	13.19	-26.49	-13.3	-6.1
China	Ouzai	<i>Sus scrofa</i>	137	13.19	-27.19	-14	-7.9
China	Ouzai	<i>Sus scrofa</i>	137	13.19	-24.29	-11.1	-7.2
China	Ouzai	<i>Ailuropoda baconi</i>	92	10.51	-25.71	-15.2	-6.3



China	Ouzai	<i>Cuon</i> sp.	15	13.39	-25.29	-11.9	-7.3
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**Annex S4.** Data from Pleistocene sites of Thailand ( $\delta^{13}\text{C}_{\text{apatite}}$ ,  $\delta^{13}\text{C}_{\text{carbon source}}$  and  $\delta^{18}\text{O}_{\text{apatite}}$  values (‰ VPDB), body mass and  $\delta^{13}\text{C}$  (‰ VPDB) Enrichment Factor used to obtain  $\delta^{13}\text{C}_{\text{carbon source}}$ ).

**Tham Lod rockshelter**, Pang Mapha District, Mae Hong Son, Northwest Thailand. The estimated age interval of Tham Lod (radiocarbon  $^{14}\text{C}$  dates on organic materials and on freshwater bivalves *Margaritanopsis laosensis*, and thermoluminescence on sediments gave a 33,000 – 11,500 BP age interval; Marwick and Gagan, 2011). Isotopic records published in Suraprasit et al. (2020).

Faunal list: *Macaca arctoides*, *M. mulatta*, *Ursus thibetanus*, *Arctonyx collaris*, *Panthera tigris*, *Elephas maximus*, *Rhinoceros* sp., *Sus scrofa*, *Muntiacus muntjac*, *Rusa unicolor*, *Rucervus eldii*, *Axis porcinus*, *Bos* sp., *Bos javanicus*, *Bos gaurus*, *Bubalus arnee*, *Naemoredus goral*, *N. griseus*, *Capricornis sumatraensis* (Wattanapitaksakul et al., 2018).

**Tham Wiman Nakin**, Khon San District, Chaiyaphum, northeast Thailand, dated to  $>\sim 170$  ka (U-series dating on calcite) (Esposito et al., 1998, 2002). Isotopic records published in Pushkina et al., (2010) and Suraprasit et al. (2020).

Faunal list: *Macaca* cf. *nemestrina*, *Trachypithecus* sp., *Pongo pygmaeus*, *Crocota Crocota ultima*, *Ailuropoda melanoleuca baconi*, *Ursus thibetanus*, *Martes flavigula*, *Arctonyx collaris rostratus*, *Paguma larvata*, *Paradoxurus hermaphroditus*, *Elephas* cf. *maximus*, *Rhinoceros* cf. *unicornis*, *Rhinoceros sondaicus*, *Tapirus indicus* cf. *intermedius*, *Sus scrofa*, *Sus* cf. *barbatus*, *Bos javanicus*, *Bos sauveli*, *Bos frontalis*, *Bubalus bubalis*, *Naemoredus sumatraensis* cf. *kanjereus*, *Muntiacus muntjac*, *Cervus unicolor*, *Cervus eldii*, *Axis porcinus*, *Cervinae* indet. (Tougaard et al., 1998), *Meles* cf. *leucurus*, *Panthera* cf. *tigris*, *Naemoredus goral* (Suraprasit et al., in press).

Country	Site	Taxon	Body Mass (kg)	$\delta^{13}\text{C}$ (‰) Enrichment Factor	$\delta^{13}\text{C}_{\text{carbon source}}$ (‰ VPDB)	$\delta^{13}\text{C}_{\text{apatite}}$ (‰ VPDB)	$\delta^{18}\text{O}_{\text{apatite}}$ (‰ VPDB)
Thailand	Tham Lod	<i>Naemoredus goral</i>	28	12,26	-13.26	-1	-3.7
Thailand	Tham Lod	<i>Naemoredus goral</i>	28	12.26	-16.26	-4	-1.8
Thailand	Tham Lod	<i>Naemoredus goral</i>	28	12.26	-12.86	-0.6	-7.3
Thailand	Tham Lod	<i>Naemoredus goral</i>	28	12.26	-14.76	-2.5	-5.5
Thailand	Tham Lod	<i>Naemoredus goral</i>	28	12.26	-12.36	-0.1	-11.4
Thailand	Tham Lod	<i>Naemoredus goral</i>	28	12.26	-17.56	-5.3	-7.5
Thailand	Tham Lod	<i>Naemoredus goral</i>	28	12.26	-16.86	-4.6	-2.8
Thailand	Tham Lod	<i>Naemoredus goral</i>	28	12.26	-16.76	-4.5	-7.4
Thailand	Tham Lod	<i>Naemoredus griseus</i>	27	12.24	-12.64	-0.4	-2.8

Thailand	Tham Lod	<i>Naemorhedus griseus</i>	27	12.24	-16.54	-4.3	-5
Thailand	Tham Lod	<i>Naemorhedus griseus</i>	27	12.24	-13.04	-0.8	-6.9
Thailand	Tham Lod	<i>Naemorhedus griseus</i>	27	12.24	-12.04	0.2	-5.6
Thailand	Tham Lod	<i>Naemorhedus griseus</i>	27	12.24	-11.64	0.6	-5.4
Thailand	Tham Lod	<i>Naemorhedus griseus</i>	27	12.24	-11.84	0.4	-8.9
Thailand	Tham Lod	<i>Naemorhedus griseus</i>	27	12.24	-15.44	-3.2	-4.6
Thailand	Tham Lod	<i>Naemorhedus griseus</i>	27	12.24	-19.34	-7.1	-0.3
Thailand	Tham Lod	<i>Naemorhedus griseus</i>	27	12.24	-12.04	0.2	-6.1
Thailand	Tham Lod	<i>Naemorhedus griseus</i>	27	12.24	-15.24	-3	-3.5
Thailand	Tham Lod	<i>Naemorhedus griseus</i>	27	12.24	-11.34	0.9	-6.7
Thailand	Tham Lod	<i>Naemorhedus griseus</i>	27	12.24	-10.34	1.9	-3.90
Thailand	Tham Lod	<i>Naemorhedus griseus</i>	27	12.24	-12.54	-0.3	-7.60
Thailand	Tham Lod	<i>Naemorhedus griseus</i>	27	12.24	-15.24	-3	-6.50
Thailand	Tham Lod	<i>Naemorhedus griseus</i>	27	12.24	-15.14	-2.9	-7.80
Thailand	Tham Lod	<i>Naemorhedus griseus</i>	27	12.24	-12.24	0	-4.1
Thailand	Tham Lod	<i>Naemorhedus griseus</i>	27	12.24	-11.74	0.5	-4.4
Thailand	Tham Lod	<i>Naemorhedus griseus</i>	27	12.24	-12.84	-0.6	-6.7
Thailand	Tham Lod	<i>Naemorhedus griseus</i>	27	12.24	-15.94	-3.7	-7.2
Thailand	Tham Lod	<i>Capricornis sumatraensis</i>	112	13.14	-15.84	-2.7	0.1
Thailand	Tham Lod	<i>Capricornis sumatraensis</i>	112	13.14	-25.44	-12.3	-7.9
Thailand	Tham Lod	<i>Capricornis sumatraensis</i>	112	13.14	-25.94	-12.8	-5.6
Thailand	Tham Lod	<i>Capricornis sumatraensis</i>	112	13.14	-24.64	-11.5	-6.9
Thailand	Tham Lod	<i>Capricornis sumatraensis</i>	112	13.14	-25.44	-12.3	-8.1
Thailand	Tham Lod	<i>Capricornis sumatraensis</i>	112	13.14	-26.34	-13.2	-7.2
Thailand	Tham Lod	<i>Capricornis sumatraensis</i>	112	13.14	-27.14	-14	-3.9
Thailand	Tham Lod	<i>Capricornis sumatraensis</i>	112	13.14	-16.74	-3.6	-4.4
Thailand	Tham Lod	<i>Capricornis sumatraensis</i>	112	13.14	-25.14	-12	-6.3
Thailand	Tham Lod	<i>Capricornis sumatraensis</i>	112	13.14	-17.74	-4.6	-6.4
Thailand	Tham Lod	<i>Capricornis sumatraensis</i>	112	13.14	-11.24	1.9	-7.8
Thailand	Tham Lod	<i>Capricornis sumatraensis</i>	112	13.14	-27.44	-14.3	-4.1
Thailand	Tham Lod	<i>Ursus thibetanus</i>	100	13.3	-28.60	-15.3	-5.8
Thailand	Tham Lod	<i>Ursus thibetanus</i>	100	13.3	-27.10	-13.8	-5.5
Thailand	Tham Lod	<i>Ursus thibetanus</i>	100	13.3	-26.40	-13.1	-6.2
Thailand	Tham Lod	<i>Panthera tigris</i>	212	14.53	-27.53	-13	-7.4
Thailand	Tham Lod	<i>Sus scrofa</i>	137	13.19	-21.09	-7.9	-9.7

Thailand	Tham Lod	<i>Sus scrofa</i>	137	13.19	-20.09	-6.9	-10.1
Thailand	Tham Lod	<i>Rucervus eldii</i>	123	13.19	-10.89	2.3	-5.5
Thailand	Tham Lod	<i>Rucervus eldii</i>	123	13.19	-11.69	1.5	-6.1
Thailand	Tham Lod	<i>Rusa unicolor</i>	220	13.59	-18.79	-5.2	-8.6
Thailand	Tham Lod	<i>Rusa unicolor</i>	220	13.59	-18.49	-4.9	-8
Thailand	Tham Lod	<i>Bubalus arnee</i>	1000	14.66	-12.26	2.4	-6.4
Thailand	Tham Lod	<i>Bubalus arnee</i>	1000	14.66	-11.66	3	-7.5
Thailand	Tham Wiman Nakin	<i>Naemorhedus goral</i>	28	12.26	-15.76	-3.5	-2.7
Thailand	Tham Wiman Nakin	<i>Naemorhedus goral</i>	28	12.26	-13.86	-1.6	-6.5
Thailand	Tham Wiman Nakin	<i>Naemorhedus goral</i>	28	12.26	-11.86	0.4	-5.7
Thailand	Tham Wiman Nakin	<i>Naemorhedus goral</i>	28	12.26	-13.06	-0.8	-1.7
Thailand	Tham Wiman Nakin	<i>Capricornis sumatraensis</i>	112	13.14	-11.44	1.7	-3.6
Thailand	Tham Wiman Nakin	<i>Capricornis sumatraensis</i>	112	13.14	-14.44	-1.3	-4.9
Thailand	Tham Wiman Nakin	<i>Capricornis sumatraensis</i>	112	13.14	-24.84	-11.7	-5.5
Thailand	Tham Wiman Nakin	<i>Capricornis sumatraensis</i>	112	13.14	-25.94	-12.8	-8.9
Thailand	Tham Wiman Nakin	<i>Capricornis sumatraensis</i>	112	13.14	-14.74	-1.6	-6
Thailand	Tham Wiman Nakin	<i>Capricornis sumatraensis</i>	112	13.14	-23.34	-10.2	-5.7
Thailand	Tham Wiman Nakin	<i>Capricornis sumatraensis</i>	112	13.14	-24.14	-11	-5.2
Thailand	Tham Wiman Nakin	<i>Capricornis sumatraensis</i>	112	13.14	-24.54	-11.4	-7
Thailand	Tham Wiman Nakin	<i>Capricornis sumatraensis</i>	112	13.14	-25.14	-12	-6
Thailand	Tham Wiman Nakin	<i>Rhinoceros sp.</i>	2000	14.34	-29.54	-15.2	-2.9
Thailand	Tham Wiman Nakin	<i>Rhinoceros sp.</i>	2000	14.34	-29.04	-14.7	-7.3
Thailand	Tham Wiman Nakin	<i>Rhinoceros sp.</i>	2000	14.34	-29.44	-15.1	-3.7
Thailand	Tham Wiman Nakin	<i>Rhinoceros sp.</i>	2000	14.34	-29.24	-14.9	-7.8
Thailand	Tham Wiman Nakin	<i>Rhinoceros sp.</i>	2000	14.34	-28.74	-14.4	-8.5
Thailand	Tham Wiman Nakin	<i>Sus scrofa</i>	137	13.19	-26.19	-13	-6.6
Thailand	Tham Wiman Nakin	<i>Sus scrofa</i>	137	13.19	-21.79	-8.6	-4.5
Thailand	Tham Wiman Nakin	<i>Sus scrofa</i>	137	13.19	-21.09	-7.9	-7.4
Thailand	Tham Wiman Nakin	<i>Sus scrofa</i>	137	13.19	-26.29	-13.1	-9.7
Thailand	Tham Wiman Nakin	<i>Sus scrofa</i>	137	13.19	-26.99	-13.8	-8.8
Thailand	Tham Wiman Nakin	Cervidae	220	13.59	-15.79	-2.2	-8.7
Thailand	Tham Wiman Nakin	Cervidae	220	13.59	-12.99	0.6	-8
Thailand	Tham Wiman Nakin	Cervidae	220	13.59	-17.29	-3.7	-4
Thailand	Tham Wiman Nakin	Cervidae	220	13.59	-14.49	-0.9	-6.2
Thailand	Tham Wiman Nakin	Cervidae	220	13.59	-20.39	-6.8	-6.3

Thailand	Tham Wiman Nakin	<i>Crocota crocuta</i>	70	14.04	-18.14	-4.1	-4.2
Thailand	Tham Wiman Nakin	<i>Crocota crocuta</i>	70	14.04	-20.14	-6.1	-7
Thailand	Tham Wiman Nakin	<i>Hystrix sp.</i>	12	12.18	-21.58	-9.4	-5
Thailand	Tham Wiman Nakin	<i>Hystrix sp.</i>	12	12.18	-23.88	-11.7	-6.9
Thailand	Tham Wiman Nakin	<i>Hystrix sp.</i>	12	12.18	-20.48	-8.3	-7.9
Thailand	Tham Wiman Nakin	<i>Hystrix sp.</i>	12	12.18	-17.68	-5.5	-5.5
Thailand	Tham Wiman Nakin	<i>Hystrix sp.</i>	12	12.18	-25.08	-12.9	-7
Thailand	Tham Wiman Nakin	<i>Pongo sp.</i>	55	12.78	-25.88	-13.1	-4

**Annex S5.** Data from Boh Dambang, Cambodia (25 -18 ka; Bacon et al., 2018c) ( $\delta^{13}\text{C}_{\text{apatite}}$ ,  $\delta^{13}\text{C}_{\text{carbon source}}$  (‰ VPDB), body mass and  $\delta^{13}\text{C}$  (‰ VPDB) Enrichment Factor used to obtain  $\delta^{13}\text{C}_{\text{carbon source}}$ ).

Number	Country	Site	Taxon	Body Mass (kg)	$\delta^{13}\text{C}_{\text{apatite}}$ (‰ VPDB)	$\delta^{13}\text{C}$ (‰) Enrichment Factor	$\delta^{13}\text{C}_{\text{carbon source}}$ (‰ VPDB)
BD-477	Cambodia	Boh Dambang	<i>Bos cf. frontalis</i>	800	0.6	14.5	-15.1
BD-85	Cambodia	Boh Dambang		800	-2.7	14.5	-17.2
BD-95	Cambodia	Boh Dambang		800	-0.3	14.5	-14.8
BD-528	Cambodia	Boh Dambang		800	0.9	14.5	-15.4
BD-163	Cambodia	Boh Dambang	<i>Capricornis sumatraensis</i>	112	-11.9	13.14	-25.04
BD-363	Cambodia	Boh Dambang		112	-13.2	13.14	-26.34
BD-166	Cambodia	Boh Dambang		112	-11.0	13.14	-24.14
BD-443	Cambodia	Boh Dambang		112	-1.6	13.14	-14.74
BD-410	Cambodia	Boh Dambang	<i>Rucervus eldii</i>	123	1.6	13.19	-14.79
BD-134	Cambodia	Boh Dambang		123	-0.4	13.19	-13.59
BD-385	Cambodia	Boh Dambang		123	0.8	13.19	-13.99
BD-146	Cambodia	Boh Dambang		123	0.0	13.19	-13.19
BD-533	Cambodia	Boh Dambang	<i>Bubalus bubalis</i>	1000	-2.2	14.66	-16.86
BD-122	Cambodia	Boh Dambang		1000	-1.5	14.66	-16.16
BD-129	Cambodia	Boh Dambang		1000	2.8	14.66	-17.46
BD-388	Cambodia	Boh Dambang	<i>Rusa unicolor</i>	220	-14.6	13.59	-28.19
BD-179	Cambodia	Boh Dambang		220	-1.4	13.59	-14.99
BD-133	Cambodia	Boh Dambang		220	-0.7	13.59	-14.29
BD-138	Cambodia	Boh Dambang		220	-2.1	13.59	-15.69
BD-375	Cambodia	Boh Dambang		220	-12.3	13.59	-25.89
BD-381	Cambodia	Boh Dambang		220	-0.1	13.59	-13.69
BD-50	Cambodia	Boh Dambang	<i>Rhinoceros sp.</i>	1633	1.3	14.25	-15.55
BD-514	Cambodia	Boh Dambang		1633	-14.3	14.25	-28.55
BD-557	Cambodia	Boh Dambang		1633	-14.0	14.25	-28.25
BD-21b	Cambodia	Boh Dambang	<i>Sus cf. scrofa</i>	137	-6.3	13.19	-19.49
BD-20	Cambodia	Boh Dambang		137	-11.6	13.19	-24.79
BD-19	Cambodia	Boh Dambang		137	-7.0	13.19	-20.19
BD-184	Cambodia	Boh Dambang	<i>Muntiacus muntjak</i>	24	-13.8	12.17	-25.97

BD-438	Cambodia	Boh Dambang		24	-13.3	12.17	-25.47
BD-238	Cambodia	Boh Dambang	<i>Hystrix brachyura</i>	12	-13.5	12.18	-25.68
BD-240	Cambodia	Boh Dambang		12	-12.8	12.18	-24.98
BD-251	Cambodia	Boh Dambang		12	-14.1	12.18	-26.28
BD-41	Cambodia	Boh Dambang		<i>Pongo pygmaeus</i>	55	-14.3	12.78
BD-45	Cambodia	Boh Dambang	55		-12.1	12.78	-24.88
BD-47	Cambodia	Boh Dambang	55		-13.2	12.78	-25.98
BD-331	Cambodia	Boh Dambang	<i>Macaca sp.</i>	6	-12.8	11.91	-24.71
BD-333	Cambodia	Boh Dambang		6	-12.5	11.91	-24.41
BD-257	Cambodia	Boh Dambang	<i>Crocota crocuta ultima</i>	70	-3.5	14.04	-17.54
BD-204	Cambodia	Boh Dambang		70	-3.9	14.04	-17.94
BD-203	Cambodia	Boh Dambang		70	-4.1	14.04	-18.14
BD-252	Cambodia	Boh Dambang		70	-1.4	14.04	-15.44
BD-200	Cambodia	Boh Dambang		70	-4.1	14.04	-18.14
BD-263	Cambodia	Boh Dambang		70	-6.1	14.04	-20.14
BD-267	Cambodia	Boh Dambang		70	-4.3	14.04	-28.34
BD-285	Cambodia	Boh Dambang		70	-3.8	14.04	-17.84
BD-16	Cambodia	Boh Dambang	<i>Canis aureus</i>	8	-7.8	13.13	-20.98
BD-304	Cambodia	Boh Dambang		8	-5.4	13.13	-18.53
BD-278	Cambodia	Boh Dambang	<i>Panthera pardus</i>	41	-6.5	13.81	-20.31
BD-286	Cambodia	Boh Dambang		41	-14	13.81	-27.81
BD-291	Cambodia	Boh Dambang		41	-7.7	13.81	-21.51
BD-51	Cambodia	Boh Dambang	<i>Ursus thibetanus</i>	100	-2.6	13.3	-15.9
BD-52	Cambodia	Boh Dambang		100	-12.6	13.3	-25.9

**Annex S6.** Crown area dimensions of teeth (r, right; l, left; m, molar; p, premolar).

n°	Tooth type	Taxon	Length	Width	Crown area
CM127	rm3	<i>Muntiacus</i> sp.	18,7	9,58	179,15
CM168	rm3	<i>Muntiacus</i> sp.	18,27	9,79	178,86
CM307	lm3	<i>Muntiacus</i> sp.	17,05	9,03	153,96
CM357	lm3	<i>Muntiacus</i> sp.	18,61	9,99	185,91
THS214	lm3	<i>Muntiacus muntjak</i>	19,32	8,96	173,11
THS217	lm3	<i>Muntiacus muntjak</i>	18,15	8,81	159,90
THS218	lm3	<i>Muntiacus muntjak</i>	18,2	9,25	168,35
THS893	lm3	<i>Muntiacus muntjak</i>	20,77	9,15	190,05
THS213	rm3	<i>Muntiacus muntjak</i>	17,99	9,33	167,85
THS216	rm3	<i>Muntiacus muntjak</i>	18,34	8,54	156,62
THS219	rm3	<i>Muntiacus muntjak</i>	17,84	8,69	155,03
THS895	rm3	<i>Muntiacus muntjak</i>	18,59	9,15	170,10
NL67b	rm3	<i>Muntiacus muntjak</i>	16,5	7,7	127,05
DU392	lm3	<i>Muntiacus muntjak</i>	17,72	9,32	165,15
DU426	lm3	<i>Muntiacus muntjak</i>	14,13	7,3	103,15
DU433	lm3	<i>Muntiacus muntjak</i>	15,8	8,45	133,51
DU437	lm3	<i>Muntiacus muntjak</i>	16,34	8,78	143,47
DU461	lm3	<i>Muntiacus muntjak</i>	17,98	9,05	162,72
DU465	lm3	<i>Muntiacus muntjak</i>	18,88	9,18	173,32
DU511	lm3	<i>Muntiacus muntjak</i>	17,8	9,61	171,06
DU394	rm3	<i>Muntiacus muntjak</i>	16,94	8,75	148,23
DU429	rm3	<i>Muntiacus muntjak</i>	17,12	9,58	164,01
DU436	rm3	<i>Muntiacus muntjak</i>	17,43	9,07	158,09
DU475	rm3	<i>Muntiacus muntjak</i>	17,15	8,97	153,84
DU1134	rm3	<i>Muntiacus muntjak</i>	16,08	8,05	129,44
THM627	lm3	<i>Muntiacus</i> sp.	16,79	9,37	157,32
THM628	lm3	<i>Muntiacus</i> sp.	17,45	8,92	155,65
THM629	lm3	<i>Muntiacus</i> sp.	18,56	9,22	171,12
THM630	lm3	<i>Muntiacus</i> sp.	18,6	9,17	170,56
THM631	lm3	<i>Muntiacus</i> sp.	18,84	9,6	180,86
THM632	lm3	<i>Muntiacus</i> sp.	15,55	8,76	136,22
THM633	lm3	<i>Muntiacus</i> sp.	18,7	9,58	179,15
THM634	lm3	<i>Muntiacus</i> sp.	15,57	8,17	127,21
THM635	lm3	<i>Muntiacus</i> sp.	17,73	9,29	164,71
THM636	lm3	<i>Muntiacus</i> sp.	17,04	9,19	156,60
THM637	lm3	<i>Muntiacus</i> sp.	16,16	8,4	135,74
THM638	lm3	<i>Muntiacus</i> sp.	16,29	8,94	145,63
THM639	lm3	<i>Muntiacus</i> sp.	18,1	9,23	167,06
THM640	rm3	<i>Muntiacus</i> sp.	17,88	9,4	168,07
THM642	rm3	<i>Muntiacus</i> sp.	17,28	9,41	162,60
THM643	rm3	<i>Muntiacus</i> sp.	18,76	9,51	178,41
THM644	rm3	<i>Muntiacus</i> sp.	19	9,53	181,07



<b>CM157</b>	lm3	<i>Capricornis</i> sp.	26,11	11,26	294,00
<b>CM284</b>	rm3	<i>Capricornis</i> sp.	24,79	11,49	284,84
<b>THS570</b>	m3	<i>Capricornis sumatraensis</i>	24,3	11,3	274,59
<b>NL7</b>	lm3	<i>Capricornis sumatraensis</i>	25,7	9,9	254,43
<b>NL8</b>	rm3	<i>Capricornis sumatraensis</i>	24,9	10,6	263,94
<b>NL9</b>	rm3	<i>Capricornis sumatraensis</i>	24,8	10,3	255,44
<b>DU538</b>	lm3	<i>Capricornis sumatraensis</i>	26,84	11,41	306,24
<b>DU569</b>	lm3	<i>Capricornis sumatraensis</i>	26,24	11,69	306,75
<b>DU574</b>	rm3	<i>Capricornis sumatraensis</i>	26,33	11,49	302,53
<b>DU613</b>	rm3	<i>Capricornis sumatraensis</i>	25,47	10,56	268,96
<b>THM19</b>	lm3	<i>Capricornis</i> cf. <i>sumatraensis</i>	30,43	13,4	407,76
<b>THM20</b>	lm3	<i>Capricornis</i> cf. <i>sumatraensis</i>	25,28	10,81	273,28
<b>THM21</b>	lm3	<i>Capricornis</i> cf. <i>sumatraensis</i>	31,81	11,82	375,99
<b>THM22</b>	lm3	<i>Capricornis</i> cf. <i>sumatraensis</i>	24,94	12,66	315,74
<b>THM23</b>	lm3	<i>Capricornis</i> cf. <i>sumatraensis</i>	26,32	11,45	301,36
<b>THM24</b>	lm3	<i>Capricornis</i> cf. <i>sumatraensis</i>	29,02	10,88	315,74
<b>THM44</b>	rm3	<i>Capricornis</i> cf. <i>sumatraensis</i>	27,3	11,84	323,23
<b>THM45</b>	rm3	<i>Capricornis</i> cf. <i>sumatraensis</i>	24,27	11,36	275,71
<b>CM78</b>	rp3	<i>Rusa unicolor</i>	15,35	9,09	139,53
<b>CM93</b>	rp3	<i>Rusa unicolor</i>	17,6	9,63	169,49
<b>CM140</b>	rp3	<i>Rusa unicolor</i>	17,92	11,66	208,95
<b>CM183</b>	rp3	<i>Rusa unicolor</i>	14,32	8,4	120,29
<b>CM184</b>	rp3	<i>Rusa unicolor</i>	16,74	10	167,40
<b>CM217a</b>	rp3	<i>Rusa unicolor</i>	16,83	9,61	161,74
<b>CM270</b>	rp3	<i>Rusa unicolor</i>	13,18	8,65	114,01
<b>CM347</b>	rp3	<i>Rusa unicolor</i>	14,86	9,36	139,09
<b>CM379</b>	rp3	<i>Rusa unicolor</i>	13,35	8,05	107,47
<b>CM491</b>	rp3	<i>Rusa unicolor</i>	15,75	10,51	165,53
<b>CM862</b>	rp3	<i>Rusa unicolor</i>	16,96	10,91	185,03
<b>CM141</b>	lp3	<i>Rusa unicolor</i>	14,66	9,39	137,66
<b>CM142</b>	lp3	<i>Rusa unicolor</i>	16,64	8,45	140,61
<b>CM220</b>	lp3	<i>Rusa unicolor</i>	15,15	8,74	132,41
<b>CM269</b>	lp3	<i>Rusa unicolor</i>	16,69	9,74	162,56
<b>CM863</b>	lp3	<i>Rusa unicolor</i>	14,79	8,73	129,12
<b>THS495</b>	lp3	<i>Rusa unicolor</i>	17,95	9,82	176,27
<b>THS739</b>	lp3	<i>Rusa unicolor</i>	16,94	10,84	183,63
<b>THS744</b>	lp3	<i>Rusa unicolor</i>	13,42	8,97	120,38
<b>NL22</b>	rp3	<i>Rusa unicolor</i>	20,33	12,85	261,24
<b>NL23</b>	rp3	<i>Rusa unicolor</i>	21,7	12,85	278,85
<b>NL27</b>	rp3	<i>Rusa unicolor</i>	18,64	9,95	185,47
<b>NL29</b>	rp3	<i>Rusa unicolor</i>	20,08	11,27	226,30
<b>NL34</b>	rp3	<i>Rusa unicolor</i>	15,82	10,29	162,79
<b>NL25</b>	lp3	<i>Rusa unicolor</i>	22,88	11,08	253,51
<b>NL103</b>	lp3	<i>Rusa unicolor</i>	18,26	9,45	172,56
<b>NL104</b>	lp3	<i>Rusa unicolor</i>	23,31	13,22	308,16

THM239	rp3	<i>Rusa unicolor</i>	17,14	8,8	150,83
THM241	rp3	<i>Rusa unicolor</i>	16,34	9,49	155,07
THM242	rp3	<i>Rusa unicolor</i>	16,42	8,55	140,39
THM243	rp3	<i>Rusa unicolor</i>	17,7	9,83	173,99
THM244	rp3	<i>Rusa unicolor</i>	17,43	10,61	184,93
THM245	rp3	<i>Rusa unicolor</i>	16,59	8,81	146,16
THM246	rp3	<i>Rusa unicolor</i>	16,37	9,36	153,22
THM247	rp3	<i>Rusa unicolor</i>	18,27	10,31	188,36
THM248	rp3	<i>Rusa unicolor</i>	16,27	9,7	157,82
THM249	rp3	<i>Rusa unicolor</i>	19,48	9,63	187,59
THM250	rp3	<i>Rusa unicolor</i>	16,52	10,2	168,50
THM251	rp3	<i>Rusa unicolor</i>	17,53	9	157,77
THM252	rp3	<i>Rusa unicolor</i>	15,87	9,42	149,50
THM252b	rp3	<i>Rusa unicolor</i>	17,42	9,49	165,32
THM253	rp3	<i>Rusa unicolor</i>	17,28	10,55	182,30
THM254	rp3	<i>Rusa unicolor</i>	20,01	9,35	187,09
THM255	rp3	<i>Rusa unicolor</i>	16,68	8,36	139,44
THM256	rp3	<i>Rusa unicolor</i>	17,38	9,39	163,20
THM198	lp3	<i>Rusa unicolor</i>	16,68	8,25	137,61
THM200	lp3	<i>Rusa unicolor</i>	15,97	9,31	148,68
THM201	lp3	<i>Rusa unicolor</i>	17,26	8,98	154,99
THM202	lp3	<i>Rusa unicolor</i>	17,07	9,5	162,17
THM203	lp3	<i>Rusa unicolor</i>	16,99	10,19	173,13
THM204	lp3	<i>Rusa unicolor</i>	18,36	9,73	178,64
THM205	lp3	<i>Rusa unicolor</i>	17,67	10,38	183,41
THM206	lp3	<i>Rusa unicolor</i>	16,44	9,86	162,10
THM207	lp3	<i>Rusa unicolor</i>	18,33	9,21	168,82
THM208	lp3	<i>Rusa unicolor</i>	15,6	8,88	138,53
THM209	lp3	<i>Rusa unicolor</i>	16,53	10,11	167,12
THM210	lp3	<i>Rusa unicolor</i>	16,47	10,51	173,10
THM211	lp3	<i>Rusa unicolor</i>	17,52	10,51	184,14
THM212	lp3	<i>Rusa unicolor</i>	17,88	10,38	185,59
THM213	lp3	<i>Rusa unicolor</i>	15,44	8,99	138,81
THM214	lp3	<i>Rusa unicolor</i>	16,52	9,24	152,64
CM495	lp3	<i>Sus scrofa</i>	11,68	6,56	76,62
CM687	lp3	<i>Sus scrofa</i>	13,98	7,18	100,38
CM688	lp3	<i>Sus scrofa</i>	12,59	7,42	93,42
CM689	lp3	<i>Sus scrofa</i>	11,89	7,14	84,89
CM690	lp3	<i>Sus scrofa</i>	13	7,08	92,04
CM791	lp3	<i>Sus scrofa</i>	13,64	7,57	103,25
CM891	lp3	<i>Sus scrofa</i>	12,05	6,16	74,23
CM892	lp3	<i>Sus scrofa</i>	14,28	7,7	109,96
CM460	rp3	<i>Sus scrofa</i>	14,79	8,79	130,00
CM788	rp3	<i>Sus scrofa</i>	12,66	7,25	91,79
CM789	rp3	<i>Sus scrofa</i>	13,35	8,29	110,67

<b>CM790</b>	rp3	<i>Sus scrofa</i>	13,6	7,27	98,87
<b>CM792</b>	rp3	<i>Sus scrofa</i>	12,03	6,58	79,16
<b>THS492</b>	rp3	<i>Sus scrofa</i>	12,72	7,79	99,09
<b>THS461</b>	rp3	<i>Sus scrofa</i>	10,16	8,06	81,89
<b>THS500</b>	rp3	<i>Sus scrofa</i>	11,6	6,99	81,08
<b>THS501</b>	rp3	<i>Sus scrofa</i>	12,96	9,52	123,38
<b>THS673</b>	rp3	<i>Sus scrofa</i>	13,51	7,08	95,65
<b>THS703</b>	rp3	<i>Sus scrofa</i>	14,02	7,98	111,88
<b>THS708</b>	rp3	<i>Sus scrofa</i>	13,51	9,07	122,54
<b>THS709</b>	rp3	<i>Sus scrofa</i>	14,07	8,9	125,22
<b>THS710</b>	rp3	<i>Sus scrofa</i>	14,11	7,97	112,46
<b>THS676</b>	lp3	<i>Sus scrofa</i>	13,67	8,69	118,79
<b>THS702</b>	lp3	<i>Sus scrofa</i>	13,03	8,44	109,97
<b>THS704</b>	lp3	<i>Sus scrofa</i>	13,9	7,98	110,92
<b>NL209</b>	lp3	<i>Sus scrofa</i>	14,62	7,88	115,21
<b>NL213</b>	rp3	<i>Sus scrofa</i>	14,95	8,95	133,80
<b>NL219</b>	rp3	<i>Sus scrofa</i>	14,05	7,83	110,01
<b>NL220</b>	rp3	<i>Sus scrofa</i>	14,18	8,88	125,92
<b>DU866</b>	rp3	<i>Sus scrofa</i>	12,61	6,62	83,48
<b>DU867</b>	rp3	<i>Sus scrofa</i>	12,1	5,55	67,16
<b>DU868</b>	rp3	<i>Sus scrofa</i>	13,91	7,71	107,25
<b>DU869</b>	rp3	<i>Sus scrofa</i>	11,59	6,12	70,93
<b>DU883</b>	rp3	<i>Sus scrofa</i>	14,05	8,37	117,60
<b>DU884</b>	rp3	<i>Sus scrofa</i>	14,41	7,25	104,47
<b>DU886</b>	rp3	<i>Sus scrofa</i>	12,8	6,41	82,05
<b>DU891</b>	rp3	<i>Sus scrofa</i>	14,13	7,96	112,47
<b>DU1129</b>	rp3	<i>Sus scrofa</i>	12,38	5,94	73,54
<b>DU865</b>	lp3	<i>Sus scrofa</i>	12,5	6,6	82,50
<b>DU870</b>	lp3	<i>Sus scrofa</i>	12,88	6,83	87,97
<b>DU878</b>	lp3	<i>Sus scrofa</i>	13,53	6,25	84,56
<b>DU885</b>	lp3	<i>Sus scrofa</i>	12,63	7,01	88,54
<b>DU887</b>	lp3	<i>Sus scrofa</i>	12,25	6,33	77,54
<b>THM823</b>	rp3	<i>Sus scrofa</i>	12,8	6,47	82,82
<b>THM858</b>	rp3	<i>Sus scrofa</i>	14,16	8,02	113,56
<b>THM859</b>	rp3	<i>Sus scrofa</i>	13,37	7,76	103,75
<b>THM860</b>	rp3	<i>Sus scrofa</i>	13,21	6,58	86,92
<b>THM862</b>	rp3	<i>Sus scrofa</i>	13,56	8,65	117,29
<b>THM864</b>	rp3	<i>Sus scrofa</i>	12,79	7,84	100,27
<b>THM865</b>	rp3	<i>Sus scrofa</i>	11,7	5,86	68,56
<b>THM866</b>	rp3	<i>Sus scrofa</i>	11,58	6,12	70,87
<b>THM879b</b>	rp3	<i>Sus scrofa</i>	15,01	7,89	118,43
<b>THM880</b>	rp3	<i>Sus scrofa</i>	11,5	5,73	65,90
<b>THM867</b>	lp3	<i>Sus scrofa</i>	13,05	6,39	83,39
<b>THM868</b>	lp3	<i>Sus scrofa</i>	11,5	5,55	63,83
<b>THM873</b>	lp3	<i>Sus scrofa</i>	13,19	8,17	107,76
<b>THM874</b>	lp3	<i>Sus scrofa</i>	14,74	7,7	113,50

<b>THM875</b>	lp3	<i>Sus scrofa</i>	13,07	8,03	104,95
<b>THM876</b>	lp3	<i>Sus scrofa</i>	13,32	7,51	100,03
<b>THM877</b>	lp3	<i>Sus scrofa</i>	13,2	6,83	90,16
<b>THM878</b>	lp3	<i>Sus scrofa</i>	12,63	7,52	94,98
<b>CM473</b>	lm3	<i>Macaca sp.</i>	11,28	6,94	78,28
<b>CM613</b>	lm3	<i>Macaca sp.</i>	9,39	6,17	57,93
<b>THS71</b>	rm3	<i>Macaca sp.</i>	12,68	7,84	99,41
<b>THS73</b>	lm3	<i>Macaca sp.</i>	11,65	7,01	81,67
<b>THS74</b>	lm3	<i>Macaca sp.</i>	11,76	7,64	89,85
<b>THS75</b>	lm3	<i>Macaca sp.</i>	13,73	9,01	123,71
<b>THS76</b>	lm3	<i>Macaca sp.</i>	12,31	7,85	96,63
<b>THS855</b>	lm3	<i>Macaca sp.</i>	11,53	7,76	89,47
<b>THS856</b>	rm3	<i>Macaca sp.</i>	10,17	7,02	71,39
<b>NL314</b>	rm3	<i>Macaca sp.</i>	11,8	7,2	84,96
<b>NL323</b>	rm3	<i>Macaca sp.</i>	13	8,2	106,6
<b>DU186</b>	lm3	<i>Macaca sp.</i>	10,23	7,55	77,24
<b>DU187</b>	lm3	<i>Macaca sp.</i>	12,3	7,32	90,04
<b>DU188</b>	lm3	<i>Macaca sp.</i>	10,71	6,86	73,47
<b>DU189</b>	lm3	<i>Macaca sp.</i>	10,98	7,44	81,69
<b>DU190</b>	rm3	<i>Macaca sp.</i>	11,45	7,09	81,18
<b>DU191</b>	rm3	<i>Macaca sp.</i>	10,38	6,59	68,40
<b>DU192</b>	rm3	<i>Macaca sp.</i>	10,37	6,46	66,99
<b>DU193</b>	lm3	<i>Macaca sp.</i>	10,42	6,44	67,10
<b>DU194</b>	lm3	<i>Macaca sp.</i>	11,27	7,6	85,65
<b>DU195d</b>	rm3	<i>Macaca sp.</i>	12,32	7,56	93,14
<b>DU196</b>	rm3	<i>Macaca sp.</i>	12,41	7,54	93,57
<b>DU1130</b>	rm3	<i>Macaca sp.</i>	12,18	7,89	96,10
<b>DU1160</b>	lm3	<i>Macaca sp.</i>	11,68	6,92	80,83
<b>DU1161</b>	rm3	<i>Macaca sp.</i>	11,55	6,91	79,81
<b>DU1162</b>	rm3	<i>Macaca sp.</i>	11,75	7,28	85,54
<b>THM1193</b>	rm3	<i>Macaca sp.</i>	11,55	7,54	87,08