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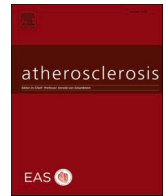
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## Triglyceride-lowering *LPL* alleles combined with LDL-C-lowering alleles are associated with an additively improved lipoprotein profile

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### ABSTRACT

**Background and aims:** Mendelian randomization studies have shown that triglyceride (TG)-lowering lipoprotein lipase (*LPL*) alleles and low-density lipoprotein-cholesterol (LDL-C)-lowering alleles have independent beneficial associations on cardiovascular disease (CVD) risk. We aimed to provide further insight into this observation by applying Mendelian randomization analyses of genetically-influenced TG and LDL-C levels on plasma metabolomic profiles.

**Methods:** We quantified over 100 lipoprotein metabolomic measures in the Netherlands Epidemiology of Obesity (NEO) study (N = 4838) and Oxford Biobank (OBB) (N = 6999) by nuclear magnetic resonance (NMR) spectroscopy. Weighted genetic scores for TG via five *LPL* alleles and LDL-C via 19 alleles were calculated and dichotomized by the median, resulting in four genotype combinations of high/low TG and high/low LDL-C. We performed linear regression analyses using a two × two design with the group with genetically-influenced high TG and LDL-C as a reference.

**Results:** Compared to the individual groups with genetically-influenced lower TG or lower LDL-C only, the group with combined genetically-influenced lower TG and LDL-C showed an overall independent and additive pattern of changes in metabolomic measures. Over 100 measures were different ( $p < 1.35 \times 10^{-3}$ ) compared to the reference, with effect sizes and directionality being similar in NEO and OBB. Most notably, levels of all very-low density lipoprotein (VLDL) and LDL sub-particles were lower.

**Conclusions:** Our findings provide evidence that TG-lowering on top of LDL-C-lowering has additive beneficial effects on the lipoprotein profile compared to TG-lowering or LDL-C-lowering only, which is in accordance with reported additive genetic effects on CVD risk reduction.

### 1. Introduction

Cardiovascular disease (CVD) is the number one cause of death worldwide [1]. Dyslipidemia, characterized by abnormally elevated serum levels of low-density lipoprotein-cholesterol (LDL-C) and triglycerides (TG) and low levels of high-density lipoprotein-cholesterol

(HDL-C), is recognized as one of the main risk factors associated with CVD [2,3]. At present, statins are the first-line therapy for prevention of CVD risk by reducing LDL-C. Statin therapy results in an approximately 30% reduction in primary CVD events [4,5]. To achieve additional reduction of CVD risk, novel lipid-lowering therapies on top of statins are currently being investigated.

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In addition to LDL-C, TG-rich lipoproteins (TRLs) have recently been identified as an independent additional risk factor for CVD [6–8]. Since the enzyme lipoprotein lipase (LPL) is a key player in TRL removal [9], it has gained attention as a druggable target. Several therapies that enhance LPL-mediated clearance of TRL-derived TG are in development for CVD prevention [10–14]. A recent phase 3 trial in which patients with homozygous familial hypercholesterolemia were treated for 24 weeks with the Angiopoietin-like 3 protein inhibitor Evinacumab, to enhance LPL-mediated TRL clearance, on top of classical LDL-C-lowering therapy, showed a 47.1% reduction in LDL-C levels compared with a 1.9% increase in the group who received only lipid-lowering therapy [11]. However, large randomized clinical trials in the general population are needed to show whether these drugs provide cardiovascular benefit in addition to statins.

In addition to randomized controlled clinical trials, Mendelian randomization studies have been exploited to assess whether enhanced LPL-mediated lipolysis has an additional benefit on top of decreased LDL-C in lowering CVD risk. For example, people with both genetically-influenced lower TG levels via *LPL* alleles and genetically-influenced lower LDL-C levels showed an additional 10% lower CVD risk compared to those with genetically-influenced lower LDL-C levels only [15]. This study suggested that drugs that enhance LPL-mediated lipolysis are likely to provide additional cardiovascular benefit on top of LDL-C-lowering agents. However, the association of genotypes with cardiovascular outcomes does not provide insight into the mechanisms behind the beneficial effects of these potential novel drugs. Since metabolites provide a functional read out of the biological processes in the human body [16,17], they can serve as intermediate phenotypes between genetic variation and CVD outcomes [18,19]. Metabolomics analyses [20] may thus provide additional insight into the pathways that mediate the effects of genetically-influenced lower TG levels via *LPL* alleles and genetically-influenced lower LDL-C levels on CVD risk.

In the current study, we assessed the causal associations between lower TG levels via *LPL* alleles and mainly lipoprotein-related metabolomic measures determined with nuclear magnetic resonance (NMR) with and without a background of lower LDL-C levels, through Mendelian randomization in two large population-based cohorts.

## 2. Materials and methods

### 2.1. Study design and population

The Netherlands Epidemiology of Obesity (NEO) study is a population-based prospective cohort study of men and women aged between 45 and 65 years. From the greater area of Leiden, The Netherlands, all inhabitants with a self-reported body mass index (BMI) of 27 kg/m<sup>2</sup> or higher were eligible to participate. In addition, inhabitants from one nearby municipality (Leiderdorp, The Netherlands) in the same age group were invited to participate regardless of their BMI, forming a reference population for BMI distribution. In total, 6671 participants were included from September 2008 until September 2012. Participants visited the NEO study center for extensive physical examination. After an overnight fast of at least 10 h, fasting blood samples were taken at the study center. Research nurses recorded current medication use by means of a medication inventory. Prior to the study visit, participants completed questionnaires at home with respect to demographic, lifestyle, and clinical information. The NEO study design was approved by the medical ethics committee of the Leiden University Medical Center (LUMC), and all participants gave their written informed consent. Detailed information about the study design and data collection has been described elsewhere [20].

For our study, we excluded participants lacking genetic data (N = 927), as described in detail below and elsewhere [21]. Additionally, we excluded participants using lipid-lowering medication (N = 906).

### 2.2. Genotyping and imputation

DNA was isolated from venous blood samples. Genotyping was performed using the Illumina HumanCoreExome-24 BeadChip (Illumina Inc., San Diego, California, United States of America). Participants were excluded in the process of quality control when 1) the sample call rate was <98%, 2) there was a sex mismatch, 3) heterozygosity rate was not within  $\pm 3$  SD of mean heterozygosity rate, 4) participants widely diverged based on the first two principal components (PCs) ( $\pm 3.5$  SD), 5) samples were duplicates, and 6) concordance with another DNA sample was >0.25 (related individuals). Genetic variants were excluded when 1) genotype call rate was <98%, and 2) variants were not in Hardy-Weinberg equilibrium ( $p < 1 \times 10^{-6}$ ). Detailed quality control steps have been described elsewhere [21]. Subsequently, genotypes were imputed to the 1000 Genome Project reference panel [22] (v3 2011) using IMPUTE (v2.2) software [23].

### 2.3. NMR-based metabolomics

A high-throughput proton NMR metabolomics platform [24] (Nightingale Health Ltd., Helsinki, Finland) was used to measure 159 metabolomic markers (excluding ratios) in plasma at the Medical Research Council Integrative Epidemiology Unit (MRC IEU) at the University of Bristol, Bristol, United Kingdom, which were quantified by Nightingale library. This method provides lipoprotein subclass profiling with lipid concentrations within 14 lipoprotein subclasses, defined as a subclass of extremely large very-low density lipoproteins (VLDL) with particle diameters from 75 nm upwards and a possible contribution of chylomicrons, five VLDL subclasses (average particle diameters of 64.0 nm, 53.6 nm, 44.5 nm, 36.8 nm, and 31.3 nm), an intermediate-density lipoprotein (IDL) subclass (28.6 nm), three LDL subclasses (25.5 nm, 23.0 nm, and 18.7 nm), and four HDL subclasses (14.3 nm, 12.1 nm, 10.9 nm, and 8.7 nm). Within the lipoprotein subclasses, the following components were quantified: total cholesterol, total lipids, phospholipids, free cholesterol, cholesteryl esters, and triglycerides. The mean size for VLDL, LDL and HDL sub-particles was calculated by weighting the corresponding subclass diameters with their particle concentrations. Furthermore, 58 metabolomic measures were determined that belong to classes of apolipoproteins, cholesterol, fatty acids (FAs), glycerides, phospholipids, amino acids, fluid balance, glycolysis-related metabolites, inflammation, and ketone bodies. Details of the experimentation and applications of the NMR metabolomics platform [24] as well as representative coefficients of variations (CVs) for the metabolomic biomarkers [25] have been described previously.

In this study, we excluded all reported ratios between metabolites as well as the metabolites that were not measured in the replication cohort (see below), resulting in a final number of 145 NMR-based metabolomic measures. The analyses were performed on ranked-based inverse normally transformed (INT) NMR-metabolites.

### 2.4. Replication dataset

#### 2.4.1. Oxford Biobank (OBB) study design

The OBB is a population-based cohort of randomly selected healthy participants aged 30–50 years from Oxfordshire (UK). Individuals with a history of myocardial infarction, diabetes mellitus, heart failure, untreated malignancy, other ongoing systemic diseases or ongoing pregnancy were not eligible for study inclusion. Participants were included between 1999 and May 2015. The OBB cohort comprises 7185 individuals. A more detailed description of the study recruitment criteria and population characteristics is reported elsewhere [26].

#### 2.4.2. Genotyping

For each OBB participant, 35 mL aliquots of whole blood were collected and frozen at  $-80$  °C for isolation of genomic DNA. Genotyping was performed using the Illumina Infinium Human Exome

Beadchip 12v1 array platform for the first consecutive 5900 DNAs, and Affymetrix UK Biobank Axiom Array chip on the first consecutive 7500 participants [26]. In total 6999 genotyped participants were included in the current study.

#### 2.4.3. Metabolomics

The Nightingale NMR-based metabolomics platform containing >200 metabolomic markers was performed on 7100 Oxford biobank fasting plasma samples. For the replication, we used 145 metabolites overlapping with the NEO cohort.

#### 2.5. Stratified genetic analyses

In this study, we calculated two independent *LPL* and LDL-C genetic scores, similar as described by Lotta et al. [15]. The TG genetic score was constructed using variants associated with TG levels that were mapped to the *LPL* gene, which were weighted by their effect on TG levels in the analyses of the Global Lipids Genetics Consortium [27]. One of the six variants used by Lotta et al. was not measured in the NEO cohort and therefore we constructed the *LPL* genetic score using the other five *LPL* variants (rs268, rs301, rs326, rs328 and rs10096633). All variants were independently and strongly associated with TG levels. More details on the selection of these *LPL* variants are described by Lotta et al. [15]. For the LDL-C score, we added 19 LDL-C-lowering alleles and weighted them by their effect on LDL-C lipid levels in the analyses of the Global Lipids Genetics Consortium [28]. These alleles were genome-wide significantly associated with LDL-C levels without showing associations with the other lipid traits. In addition, all LDL-C variants were over 500 kb away from each other and had no or negligible linkage disequilibrium ( $R^2 < 0.01$ ). The list of variants used for *LPL* and LDL-C genetic scores is given in Supplementary Table 1. The linkage disequilibrium scores between the *LPL* variants are shown in Supplementary Table 2.

Then, we dichotomized each score at their corresponding median value to “naturally randomize” the participants into 4 groups: (1) a reference group, with genetically-influenced higher TG and LDL-C levels, as a proxy for placebo (2) a group with genetically-influenced lower TG levels, as proxy for *LPL* enhancing therapy and (3) a group with genetically-influenced lower LDL-C levels, as proxy for LDL-C-lowering therapy like statins (4) a group with both genetically-influenced lower TG and genetically-influenced lower LDL-C levels, as proxy for *LPL* enhancing therapy on top of LDL-C-lowering therapy. This process of natural allocation is schematically depicted elsewhere [15].

#### 2.6. Statistical analyses

Using the four “naturally randomized” groups constructed as described above, we performed linear regression analyses to estimate the associations with NMR-based metabolomic measures between groups using a two  $\times$  two factorial design. These association analyses were adjusted for age, sex and the first four genomic principal components to correct for possible population stratification. In addition, we performed interaction analyses between the *LPL* and LDL-C genetic scores in order to test whether they had synergetic effects on the NMR-based metabolomic measures.

We corrected for multiple testing using the method described by Li and Ji [29]. Using this method, we were able to correct for the number of independent metabolomic measures, as other correction factors are too stringent given the high intercorrelations between the metabolomic measures, which could give false-negative results. In this study, we corrected for 37 independent tests, and therefore, associations with a  $p < 1.35 \times 10^{-3}$  were considered statistically significant.

In the NEO study, individuals with a BMI of 27 kg/m<sup>2</sup> or higher are oversampled. Therefore, all results were based on analyses weighted towards a normal reference BMI distribution, and thus apply to populations without oversampling of individuals with overweight or obesity. A more detailed description of the weighting can be found

elsewhere [21].

Analyses were performed using STATA Statistical Software version 12.0 (Statacorp, College Station, Texas, USA) and R version 3.6.1 (The R Project, <https://www.r-project.org/>). The circular plots were designed with Python version 2.7.6 (Python Software Foundation, <https://www.python.org/>). The other figures were designed using IBM SPSS Statistics version 25 (SPSS, Inc., Chicago, USA).

### 3. Results

#### 3.1. Population characteristics

Characteristics of the NEO study population (N = 4838) and OBB cohort (N = 6999) are summarized in Table 1. Compared to participants from the NEO cohort, OBB participants had a lower mean age (41.6 vs. 55.5 years, respectively) but a similar mean BMI (25.8 and 26.0 kg/m<sup>2</sup> for OBB and NEO, respectively). Levels of TG, total cholesterol, LDL-C and HDL-C were higher in the NEO cohort compared to the OBB cohort. The median *LPL* and LDL genetic scores (in SD units) were similar between the two cohorts. The characteristics of the NEO and OBB cohorts stratified by the dichotomized *LPL* and LDL-C genetic scores are shown in Supplementary Tables 3 and 4

#### 3.2. Two $\times$ two factorial analyses of genetically-influenced lower TG and genetically-influenced lower LDL-C in the NEO cohort

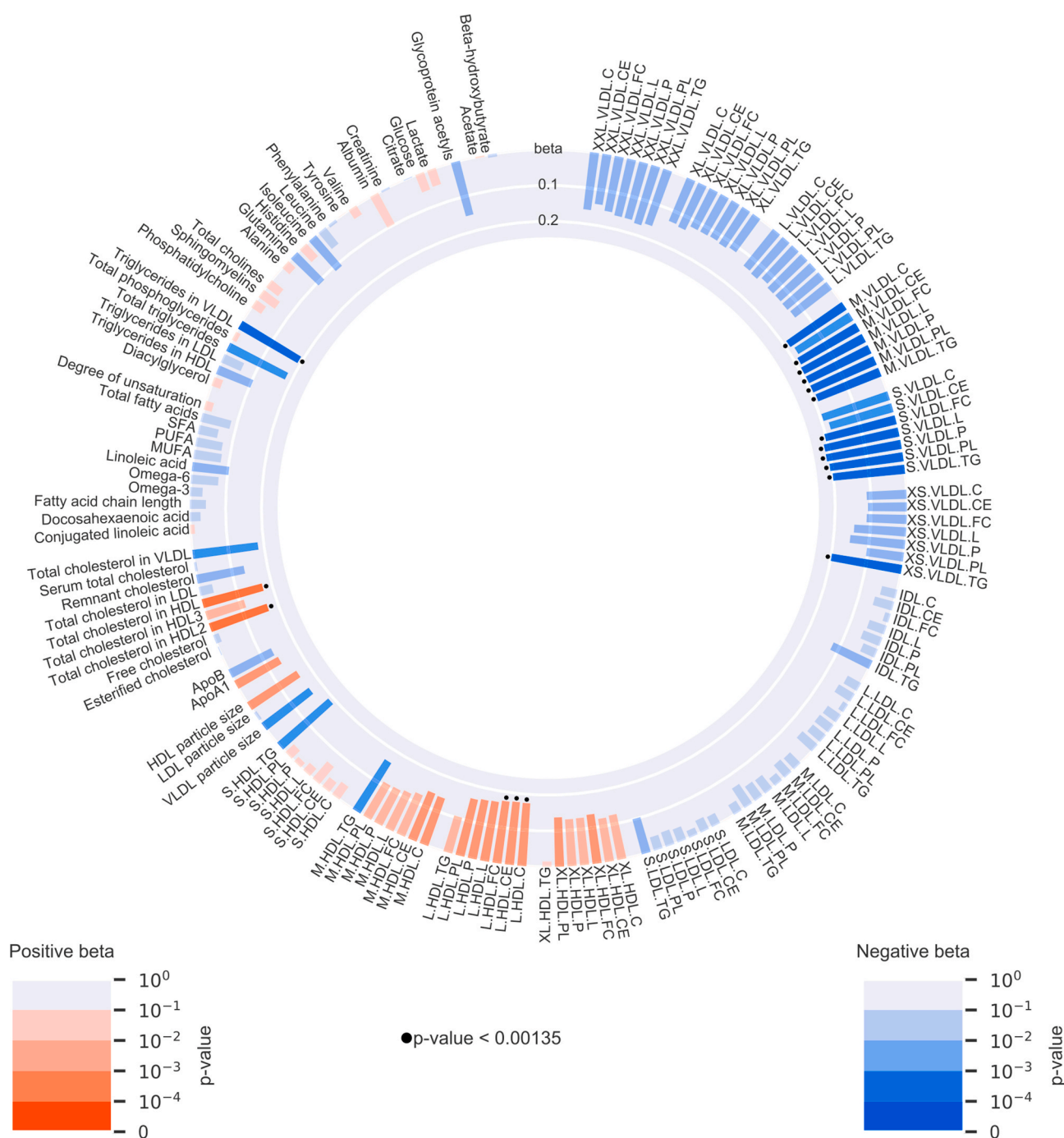
The results of the factorial analyses of groups with genetically-influenced lower TG and/or LDL-C levels on the fasting NMR metabolomic measures are shown in Figs. 1–3 and detailed results are provided in Supplementary Table 5. Compared with the reference group (combined genetically-influenced higher TG and LDL-C levels), the group with genetically-influenced lower TG levels only had lower levels of TG in VLDL and lower numbers of small and medium sized VLDL sub-particles (SVLDL-P: beta (SE) =  $-0.21$  (0.06),  $p = 3.9 \times 10^{-4}$ , MVLDL-P: beta (SE) =  $-0.19$  (0.06),  $p = 7.6 \times 10^{-4}$ ) and higher levels of HDL-C (HDL-C: beta (SE) =  $0.17$  (0.05),  $p = 4.9 \times 10^{-4}$ ) (Fig. 1). The group with genetically-influenced lower LDL-C levels had lower levels of apolipoprotein B (ApoB) and lower levels of medium and large sized LDL sub-particles (strongest association on phospholipid content of medium LDL (MLDL-PL): beta (SE) =  $-0.19$  (0.05),  $p = 6.4 \times 10^{-4}$  and LLDL-PL: beta (SE) =  $-0.18$  (0.05),  $p = 8.1 \times 10^{-4}$ ) (Fig. 2). The group with combined genetically-influenced lower TG and LDL-C levels showed the largest number of measures being different in concentration from the reference group (both genetic exposures: n = 102 vs. genetically-influenced lower LDL-C only: n = 13 vs. genetically-influenced lower

**Table 1**  
Characteristics of the discovery cohort (NEO) and replication cohort (OBB).

Characteristics	Discovery cohort <sup>a</sup> NEO (n = 4838)	Replication cohort OBB (n = 6999)
Age (years)	55.5 (6.0)	41.6 (5.9)
Men	42.0%	43.6%
BMI (kg/m <sup>2</sup> )	26.0 (4.3)	25.8 (4.6)
Fasting serum concentrations (mmol/L)		
TG (median (IQR))	0.99 (0.71; 1.42)	0.93 (0.69; 1.34)
Total cholesterol	5.80 (1.01)	5.18 (1.01)
LDL-cholesterol	3.66 (0.94)	3.22 (1.26)
HDL-cholesterol	1.60 (0.47)	1.38 (0.42)
GRS LDL (median (IQR))	0.88 (0.80; 0.97)	0.90 (0.82; 0.99)
GRS <i>LPL</i> (median (IQR))	0.48 (0.39; 0.63)	0.48 (0.39; 0.63)

Values are mean (SD), unless otherwise specified. GRS unit is in SD. BMI, body mass index; TG, triglycerides; HD, high-density lipoprotein; LDL, low-density lipoprotein; IQR, interquartile range; GRS genetic risk score.

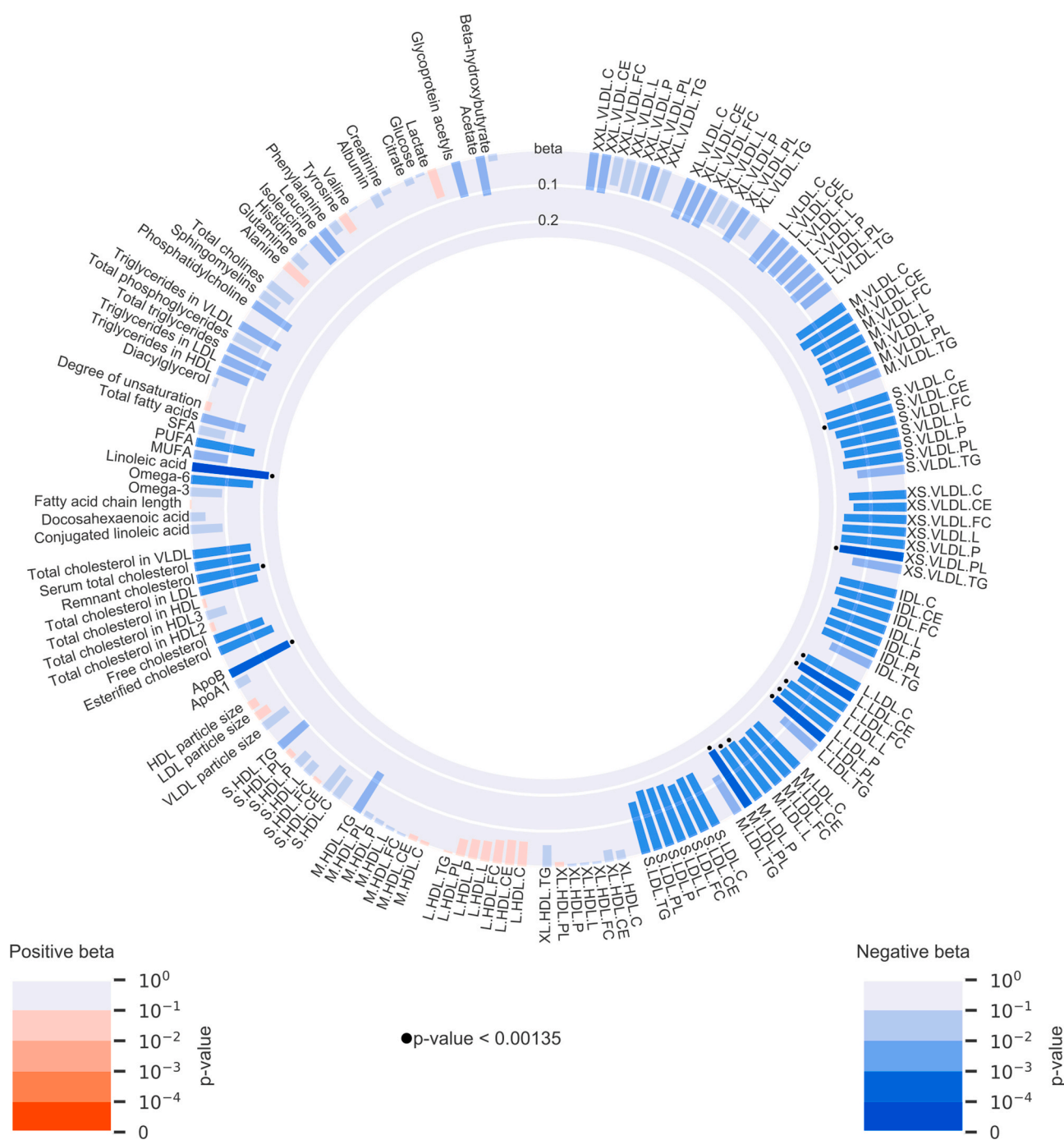
<sup>a</sup> In NEO, results are based on analyses weighted towards the reference BMI distribution of the general Dutch population.



**Fig. 1. Associations of the group with genetically-influenced lower TG levels with 145 NMR-based metabolomic measures in two × two factorial analyses.** Group with genetically-influenced lower TG levels compared with the reference group in the NEO cohort. Bar heights represent the magnitude of the beta coefficient from linear regression, which is expressed in standard deviation (SD) units. Red bars indicate positive betas and blue bars indicate negative betas. The transparency of the bars indicates the level of statistical significance. A  $p < 1.35 \times 10^{-3}$  is regarded statistically significant, as represented by the black dots.

TG only:  $n = 18$ ) and the effect sizes were substantially larger compared with the other groups (Fig. 3). Overall, these effects showed an additive pattern between the *LPL* and *LDL-C* genetic scores, but no evidence for an interaction between these scores ( $p$  for interaction  $> 1.35 \times 10^{-3}$ ). All components of all VLDL sub-particles and LDL sub-particles were significantly lower, with the exception of the TG component of medium LDL (MLDL-TG). In this combination group, ApoB, remnant cholesterol, total serum cholesterol, VLDL-cholesterol (VLDL-C) and LDL-C were significantly lower, whereas levels of large HDL sub-particles and HDL-C were higher. Furthermore, the combination group had a lower average VLDL particle size (VLDLD: beta (SE) =  $-0.27$  (0.05),  $p = 3.83 \times 10^{-7}$ )

and higher average HDL particle size (HDL: beta (SE) =  $0.22$  (0.05),  $p = 3.26 \times 10^{-5}$ ), but no differences in LDL particle size compared to the reference group. Levels of total FAs (beta (SE) =  $-0.29$  (0.06),  $p = 2.46 \times 10^{-7}$ ) and several free FAs including omega-3, omega-6, mono-unsaturated FAs, polyunsaturated FAs and linoleic acid were also lower in the combination group. TG content in almost all lipoprotein sub-particles was lower, as well as total TG (beta (SE) =  $-0.35$  (0.05),  $p = 9.8 \times 10^{-11}$ ). Despite not being significantly different in either the genetically lower TG or LDL-C group only vs. the reference group, the inflammation marker glycoprotein acetyls was significantly lower in the combination group (glycoprotein acetyls: beta (SE) =  $-0.31$  (0.06),  $p =$



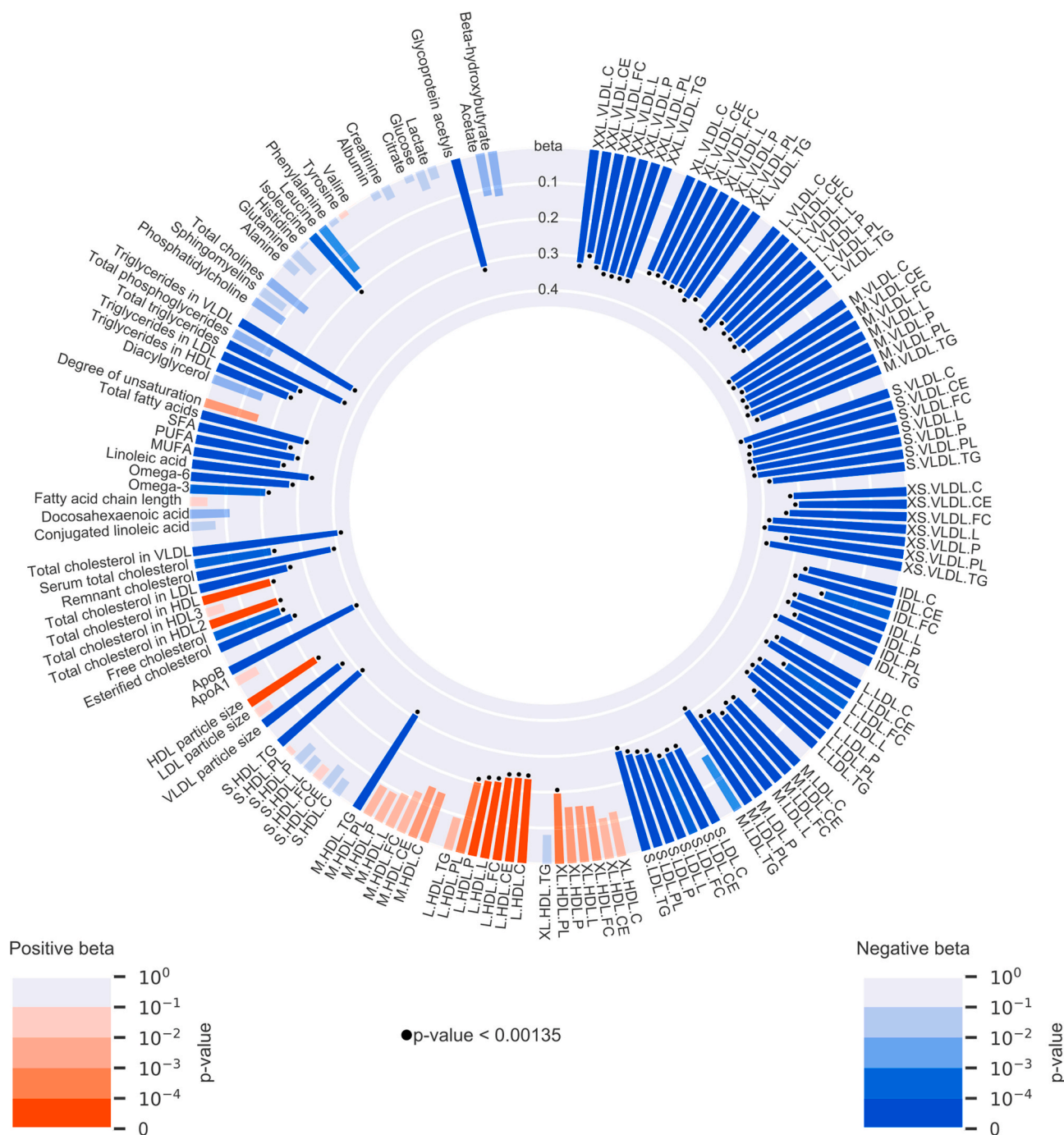
**Fig. 2.** Associations of the group with genetically-influenced lower LDL-C levels with 145 NMR-based metabolomic measures in two × two factorial analyses. Group with genetically-influenced lower LDL-C levels compared with the reference group in the NEO cohort. Bar heights represent the magnitude of the beta coefficient from linear regression, which is expressed in standard deviation (SD) units. Red bars indicate positive betas and blue bars indicate negative betas. The transparency of the bars indicates the level of statistical significance. A  $p < 1.35 \times 10^{-3}$  is regarded statistical significant, as represented by the black dots.

$1.9 \times 10^{-7}$ ).

**3.3. Two × two factorial analyses of genetically-influenced lower TG and genetically-influenced lower LDL-C in the OBB cohort**

In OBB, the group with genetically-influenced lower TG levels only, did not exhibit any differences in NMR-based total TG or lipoprotein sub-particle TG levels, contrasting the findings in NEO. However, genetically-influenced lower TG levels were associated with higher levels of apolipoprotein A1 and HDL sub-particles (Supplementary Fig. 1A and Supplementary Table 6), similar to the findings in NEO. In the

OBB, the group with genetically-influenced lower LDL-C levels had lower levels of ApoB and lower numbers of medium and large sized LDL sub-particles (strongest association on MLDL-P: beta (SE) = -0.20 (0.03),  $p = 1.8 \times 10^{-10}$ ). These observations were consistent with the findings in NEO (Supplementary Fig. 1B and Supplementary Table 6). Also similar to NEO, the group of combined genetically-influenced lower TG and LDL-C levels had the largest number of significant differences with the largest effect sizes compared with the reference group (both genetic exposures:  $n = 106$  vs. genetically-influenced lower LDL-C only:  $n = 65$  vs. genetically-influenced lower TG only:  $n = 16$ ) (Supplementary Fig. 1C and Supplementary Table 6). With the exception of glycoprotein



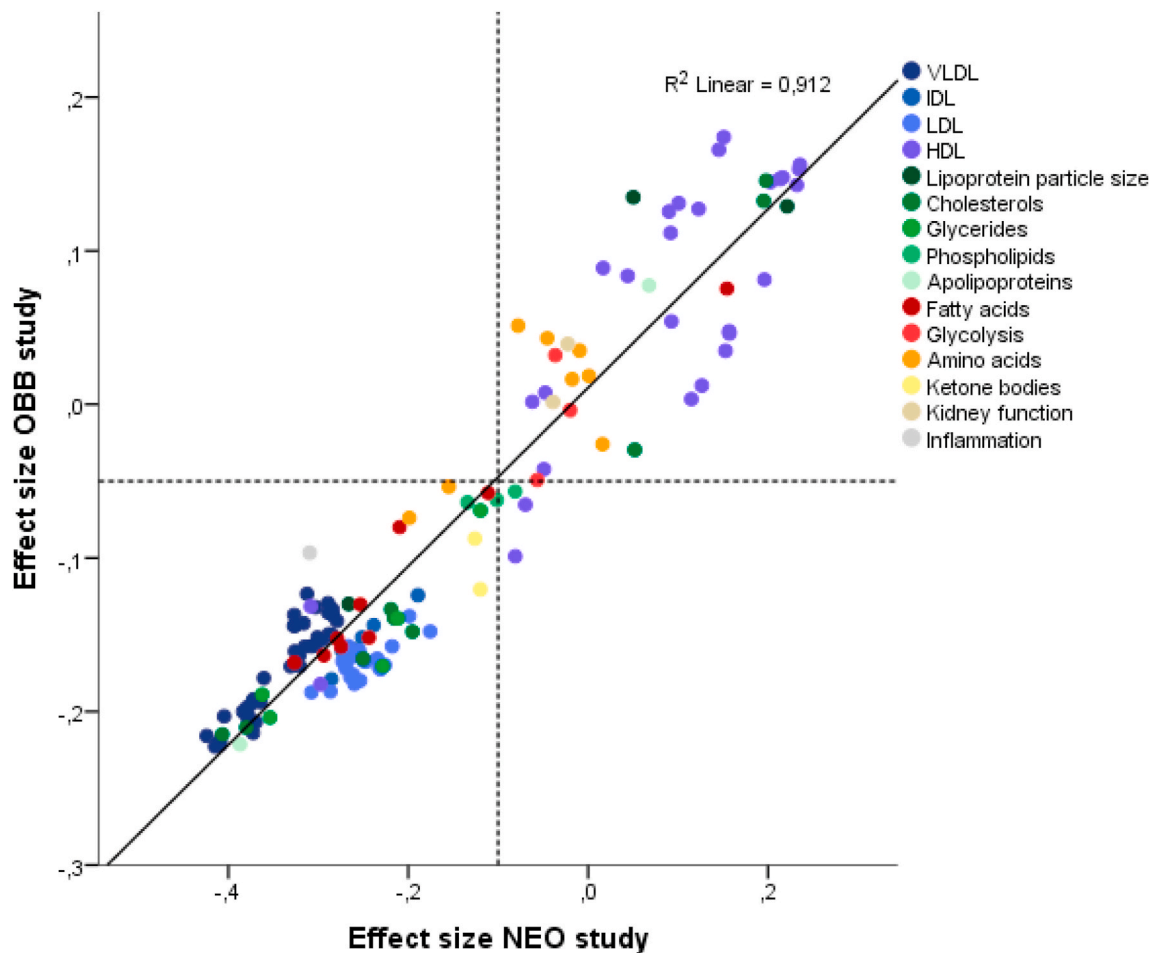
**Fig. 3. Associations of the group with both genetically-influenced lower TG and LDL-C levels with 145 NMR-based metabolomic measures in two × two factorial analyses.** Group with combined genetically-influenced lower TG and LDL-C levels compared with the reference group in the NEO cohort. Bar heights represent the magnitude of the beta coefficient from linear regression, which is expressed in standard deviation (SD) units. Red bars indicate positive betas and blue bars indicate negative betas. The transparency of the bars indicates the level of statistical significance. A  $p < 1.35 \times 10^{-3}$  is regarded statistical significant, as represented by the black dots.

acetyls, there was large consistency in the findings between NEO and OBB (Fig. 4). Also, consistent with findings in NEO, formal interaction analysis of genetically lower TG and genetically lower LDL-C showed that there was no interaction between these genetic scores ( $p$  for interaction  $> 1.35 \times 10^{-3}$ ) for any of the metabolomic measures.

**4. Discussion**

In this study, we assessed the effects of genetically-influenced lower TG levels via five *LPL* alleles and genetically-influenced lower LDL-C

levels via 19 LDL-C-lowering genes separately and in combination, on NMR-based metabolomic measures, including detailed measures of lipoprotein levels and composition. Our results showed that in the NEO study, genetically-influenced lower TG levels were mainly associated with lower levels of small and medium sized VLDL sub-particles. Although these effects were most apparent in the NEO study, direction and pattern of the effects were mostly overlapping in the OBB. The genetically-influenced lower LDL-C levels were associated with lower levels of ApoB and lower levels of medium and large sized LDL sub-particles in both NEO and OBB, and the effect sizes of these changes were similar



**Fig. 4. Correlation between beta coefficients of the discovery and replication cohorts.** The effect sizes (i.e. beta coefficients) for the associations between genetically-influenced lower TG and LDL-C levels and 145 NMR-based metabolomic measures in the Netherlands Epidemiology of Obesity (NEO) study are partially replicated in Oxford Biobank (OBB) cohort, as shown by the high correlation between the beta coefficients from both cohorts.

between the two cohorts. In the group with both genetically-influenced lower TG and genetically-influenced lower LDL-C levels, the most interesting observations were that concomitant with a lower level of ApoB, the vast majority of the number and sub-particles of LDL, IDL and VLDL lipoproteins were lower. The effect sizes, direction and pattern of these changes were highly overlapping between NEO and OBB studies. Importantly, the effect sizes of the associations observed in the combination group were independent and additive, indicating that pharmacological TG-lowering therapy on top of LDL-C-lowering therapy will have additional beneficial effects on the lipoprotein profile.

Our results show that the genetic effects of lower TG and lower LDL-C levels on the metabolomic profile are independent and additive. These findings further support the previously reported independent and additive effects of genetically-influenced lower TG and LDL-C levels on CVD risk [15,30]. The study from Lotta et al. [15], that also used a two  $\times$  two factorial design, showed that people in the group with both genetically-influenced lower TG levels and lower LDL-C levels had the largest reduction in CVD, compared to the reference group. This was 7% more than expected based on the separate associations of the two genetic exposures ( $p$  for interaction = 0.02). However, Lotta et al. further reported that interaction analyses using a continuous score of *LPL* and stratifying above or below the median or by quintiles of distribution of LDL-C-lowering alleles were not consistent with an interaction between the two genetic scores. These results indicated that the effects of TG-lowering via *LPL* and LDL-C-lowering on CVD are independent. This finding is in line with the absence of an interaction between

genetically-influenced lower TG and LDL-C as observed in our study. A recent study from Ference et al. [30], that used similar *LPL* variants to the ones we used for the TG genetic score and *LDLR* variants for the LDL-C genetic score, also showed that these scores were associated with lower CVD risk. They further concluded that the individual associations of the *LPL* and LDL-C genetic scores with CVD appeared to be independent, additive, and proportional to the absolute change in ApoB. In our study, the effect size of the combination of genetic exposures on ApoB was close to the sum of the effect sizes on ApoB for each of the *LPL* and LDL-C genetic scores separately (combined: beta (SE) =  $-0.39$  (0.06); *LPL*: beta (SE) =  $-0.13$  (0.06); LDL-C: beta (SE) =  $-0.18$  (0.06), respectively), which is thus in concordance with the paper of Ference et al. The independent and additive effects of genetically-influenced lower TG levels via *LPL* and lower LDL-C levels on the lipoprotein profile is further evidence for an expected additional effect of pharmacologically enhanced LPL activity on top of LDL-C-lowering therapy on reduction of CVD risk.

The effects of genetically-influenced lower TG levels on the lipoprotein profile are fully in line with the current understanding of the role of LPL in lipid metabolism, and they confirm the previously reported associations of increased LPL activity with decreased TG and VLDL-C levels [31]. In addition to changes in TRLs, we also observed that genetically-influenced lower TG levels were associated with increased HDL-C levels. This is in line with the previously reported inverse association between TG levels and HDL-C levels and HDL particle size [32, 33]. Furthermore, our study showed that levels of total FAs and several



specific free FAs were significantly lower in the combined genetically-influenced lower TG and LDL-C group both in the NEO and OBB cohort (Fig. 3 and Supplementary Fig. 1C). High levels of circulating free FAs have been associated with increased oxidative stress and inflammation, which, in turn, may lead to formation of atherosclerotic plaques [34,35]. Therefore, the lower levels of free FAs observed in the group with both genetically-influenced lower TG levels and lower LDL-C levels may play an additional role in reducing CVD risk beyond lowering of atherogenic lipoproteins.

When interpreting the results of our study, several assumptions and limitations of the Mendelian randomization approach should be taken into consideration. First, when translating genetic findings into pharmacological strategies, it should be realized that the consequences of lifelong exposure to genetically-influenced lower TG and LDL-C determined by Mendelian randomization may differ from the relatively short-term pharmacological (combined) effects of TG and LDL-C-lowering agents. Second, Mendelian randomization assumes that genetic variants are associated with the end point of interest only via the pathway of the exposure of interest and thus pleiotropic effects could invalidate the results. For this study, we attempted to minimize possible pleiotropic effects of the LDL-C genetic score by including variants associated with LDL-C only, hence without associations to the other lipid traits. The results of the group with genetically-influenced lower LDL-C levels (Fig. 1B), which were predominantly lower levels of LDL sub-particles and ApoB, confirm that the effects are most likely exerted via lowering LDL-C only. Furthermore, the *LPL* genetic score comprised variants that were in or within 10 kb of the *LPL* gene itself. Two of the *LPL* variants were intronic variants (rs326 and rs301) that were significant eQTLs in adipose tissue, one intronic (rs10096633) located in a regulatory region and two coding variants (rs268 and rs328) associated with *LPL* function [36,37]. This makes it likely that the effects of genetically-influenced lower TG resulted through *LPL*. Another limitation of our study is that our data are pertinent only to European populations, given that both the NEO and the OBB are European cohorts.

In conclusion, our study showed that genetically-influenced lower TG levels via enhanced *LPL*-mediated lipolysis and genetically-influenced lower LDL-C levels have independent and additive effects on the lipoprotein profile, providing insight on how these genetic exposures might reduce CVD risk. Altogether, these findings provide further evidence for an additional clinical benefit of pharmacologically enhancing *LPL* activity on top of LDL-C-lowering to further improve cardiovascular outcomes of patients at risk.

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### Author contributions

Study design: D.I., L.B., R.N., P.R., K.W.v.D.; Statistical analyses: D.I., L.B., R.N.; Drafting of the manuscript: D.I., K.W.v.D., R.N.; Project supervision: K.W.v.D., R.N. Results interpretation and discussion: All co-authors. Critical commenting and final approval of the manuscript: All co-authors.

### Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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### Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.atherosclerosis.2021.04.015>.

### References

- [1] K. Mc Namara, H. Alzubaidi, J.K. Jackson, Cardiovascular disease as a leading cause of death: how are pharmacists getting involved? *Integrated Pharm. Res. Pract.* 8 (2019) 1–11, <https://doi.org/10.2147/IPRP.S133088>. Published 2019 Feb 4.
- [2] V.J. Ama Moor, S. Ndongo Amougou, S. Ombotto, F. Ntone, D.E. Woumba, B. Ngo Nonga, Dyslipidemia in patients with a cardiovascular risk and disease at the university teaching hospital of yaoundé, Cameroon, *Int J Vasc Med* 2017 (2017) 6061306, <https://doi.org/10.1155/2017/6061306>.
- [3] R. Stein, F. Ferrari, F. Scolari, Genetics, dyslipidemia, and cardiovascular disease: new insights, *Curr. Cardiol. Rep.* 21 (8) (2019 Jun 21) 68, <https://doi.org/10.1007/s11886-019-1161-5>. PMID: 31227920.
- [4] B.M. Cheung, I.J. Lauder, C.P. Lau, C.R. Kumana, Meta-analysis of large randomized controlled trials to evaluate the impact of statins on cardiovascular outcomes, *Br. J. Clin. Pharmacol.* 57 (5) (2004) 640–651, <https://doi.org/10.1111/j.1365-2125.2003.02060.x>.
- [5] H.G. Yebo, H.E. Aschmann, M. Kaufmann, M.A. Puhon, Comparative effectiveness and safety of statins as a class and of specific statins for primary prevention of cardiovascular disease: a systematic review, meta-analysis, and network meta-analysis of randomized trials with 94,283 participants, *Am. Heart J.* 210 (2019) 18–28, <https://doi.org/10.1016/j.ahj.2018.12.007>.
- [6] C. Xiao, S. Dash, C. Morgantini, R.A. Hegele, G.F. Lewis, Pharmacological targeting of the atherogenic dyslipidemia complex: the next frontier in CVD prevention beyond lowering LDL cholesterol, *Diabetes* 65 (7) (2016) 1767–1778, <https://doi.org/10.2337/db16-0046>.
- [7] P.B. Sandesara, S.S. Virani, S. Fazio, M.D. Shapiro, The forgotten lipids: triglycerides, remnant cholesterol, and atherosclerotic cardiovascular disease risk, *Endocr. Rev.* 40 (2) (2019) 537–557, <https://doi.org/10.1210/er.2018-00184>.
- [8] P.P. Toth, Triglyceride-rich lipoproteins as a causal factor for cardiovascular disease, *Vasc. Health Risk Manag.* 12 (2016) 171–183, <https://doi.org/10.2147/VHRM.S104369>. Published 2016 May 6.
- [9] K. Preiss-Landl, R. Zimmermann, G. Hämmerle, R. Zechner, Lipoprotein lipase: the regulation of tissue specific expression and its role in lipid and energy metabolism, *Curr. Opin. Lipidol.* 13 (5) (2002) 471–481, <https://doi.org/10.1097/00041433-200210000-00002>.
- [10] D. Gaudet, D.A. Gipe, R. Pordy, Z. Ahmad, M. Cuchel, et al., ANGPTL3 inhibition in homozygous familial hypercholesterolemia, *N. Engl. J. Med.* 377 (3) (2017) 296–297, <https://doi.org/10.1056/NEJMc1705994>.
- [11] F.J. Raal, R.S. Rosenson, L.F. Reeskamp, G.K. Hovingh, J.J.P. Kastelein, et al., Evinacumab for homozygous familial hypercholesterolemia, *N. Engl. J. Med.* 383 (8) (2020) 711–720, <https://doi.org/10.1056/NEJMoa2004215>.
- [12] M.J. Graham, R.G. Lee, T.A. Brandt, L.J. Tai, W. Fu, et al., Cardiovascular and metabolic effects of ANGPTL3 antisense oligonucleotides, *N. Engl. J. Med.* 377 (3) (2017) 222–232, <https://doi.org/10.1056/NEJMoa1701329>.
- [13] D. Gaudet, D. Brisson, K. Tremblay, V.J. Alexander, W. Singleton, et al., Targeting APOC3 in the familial chylomicronemia syndrome, *N. Engl. J. Med.* 371 (23) (2014) 2200–2206, <https://doi.org/10.1056/NEJMoa1400284>.
- [14] W.J. Geldenhuys, D. Aring, P. Sadana, A novel Lipoprotein lipase (LPL) agonist rescues the enzyme from inhibition by angiopoietin-like 4 (ANGPTL4), *Bioorg. Med. Chem. Lett* 24 (9) (2014) 2163–2167, <https://doi.org/10.1016/j.bmcl.2014.03.021>.
- [15] L.A. Lotta, I.D. Stewart, S.J. Sharp, F.R. Day, S. Burgess, et al., Association of genetically enhanced lipoprotein lipase-mediated lipolysis and low-density lipoprotein cholesterol-lowering alleles with risk of coronary disease and type 2 diabetes, *JAMA Cardiol* 3 (10) (2018 Oct 1) 957–966, <https://doi.org/10.1001/jamacardio.2018.2866>. PMID: 30326043; PMCID: PMC6217943.

- [16] A. Ganna, S. Salihovic, J. Sundström, C.D. Broeckling, A.K. Hedman, et al., Large-scale metabolomic profiling identifies novel biomarkers for incident coronary heart disease, *PLoS Genet.* 10 (12) (2014), e1004801, <https://doi.org/10.1371/journal.pgen.1004801>. Published 2014 Dec 11.
- [17] T. Illig, C. Gieger, G. Zhai, W. Römisch-Margl, R. Wang-Sattler, et al., A genome-wide perspective of genetic variation in human metabolism, *Nat. Genet.* 42 (2) (2010) 137–141, <https://doi.org/10.1038/ng.507>.
- [18] S.H. Shah, J.R. Bain, M.J. Muehlbauer, R.D. Stevens, D.R. Crosslin, et al., Association of a peripheral blood metabolic profile with coronary artery disease and risk of subsequent cardiovascular events, *Circ Cardiovasc Genet* 3 (2) (2010) 207–214, <https://doi.org/10.1161/CIRCGENETICS.109.852814>.
- [19] G.D. Lewis, R.E. Gerszten, Toward metabolomic signatures of cardiovascular disease, *Circ Cardiovasc Genet* 3 (2) (2010) 119–121, <https://doi.org/10.1161/CIRCGENETICS.110.954941>.
- [20] R. de Mutsert, M. den Heijer, T.J. Rabelink, J.W. Smit, J.A. Romijn, et al., The Netherlands Epidemiology of Obesity (NEO) study: study design and data collection, *Eur. J. Epidemiol.* 28 (6) (2013) 513–523, <https://doi.org/10.1007/s10654-013-9801-3>.
- [21] L.L. Blauw, R. Li-Gao, R. Noordam, R. de Mutsert, S. Trompet, et al., CETP (cholesteryl ester transfer protein) concentration: a genome-wide association study followed by mendelian randomization on coronary artery disease, *Circ Genom Precis Med* 11 (5) (2018), e002034, <https://doi.org/10.1161/CIRCGEN.117.002034>.
- [22] 1000 Genomes Project Consortium, A. Auton, L.D. Brooks, R.M. Durbin, E. P. Garrison, et al., A global reference for human genetic variation, *Nature* 526 (7571) (2015) 68–74, <https://doi.org/10.1038/nature15393>.
- [23] B. Howie, C. Fuchsberger, M. Stephens, J. Marchini, G.R. Abecasis, Fast and accurate genotype imputation in genome-wide association studies through pre-phasing, *Nat. Genet.* 44 (8) (2012) 955–959, <https://doi.org/10.1038/ng.2354>. Published 2012 Jul 22.
- [24] P. Soininen, A.J. Kangas, P. Würtz, T. Suna, M. Ala-Korpela, Quantitative serum nuclear magnetic resonance metabolomics in cardiovascular epidemiology and genetics, *Circ Cardiovasc Genet* 8 (1) (2015) 192–206, <https://doi.org/10.1161/CIRCGENETICS.114.000216>.
- [25] J. Kettunen, A. Demirkan, P. Würtz, H.H. Draisma, T. Haller, et al., Genome-wide study for circulating metabolites identifies 62 loci and reveals novel systemic effects of LPA, *Nat. Commun.* 7 (2016) 11122, <https://doi.org/10.1038/ncomms11122>. Published 2016 Mar 23.
- [26] F. Karpe, S.K. Vasan, S.M. Humphreys, J. Miller, J. Cheeseman, et al., Cohort profile: the Oxford biobank, *Int. J. Epidemiol.* 47 (1) (2018), <https://doi.org/10.1093/ije/dyx132>, 21–21g.
- [27] A.V. Khera, H.H. Won, G.M. Peloso, C. O'Dushlaine, D. Liu, et al., Association of rare and common variation in the lipoprotein lipase gene with coronary artery disease, *J. Am. Med. Assoc.* 317 (9) (2017) 937–946, <https://doi.org/10.1001/jama.2017.0972>.
- [28] C.J. Willer, E.M. Schmidt, S. Sengupta, G.M. Peloso, S. Gustafsson, et al., Discovery and refinement of loci associated with lipid levels, *Nat. Genet.* 45 (11) (2013) 1274–1283, <https://doi.org/10.1038/ng.2797>.
- [29] J. Li, L. Ji, Adjusting multiple testing in multilocus analyses using the eigenvalues of a correlation matrix, *Hereditas* 95 (3) (2005) 221–227, <https://doi.org/10.1038/sj.hdy.6800717>.
- [30] B.A. Ference, J.J.P. Kastelein, K.K. Ray, H.N. Ginsberg, M.J. Chapman, et al., Association of triglyceride-lowering LPL variants and LDL-C-lowering LDLR variants with risk of coronary heart disease, *J. Am. Med. Assoc.* 321 (4) (2019) 364–373, <https://doi.org/10.1001/jama.2018.20045>.
- [31] F. Drenos, G. Davey Smith, M. Ala-Korpela, J. Kettunen, P. Würtz, et al., Metabolic characterization of a rare genetic variation within APOC3 and its lipoprotein lipase-independent effects, *Circ Cardiovasc Genet* 9 (3) (2016) 231–239, <https://doi.org/10.1161/CIRCGENETICS.115.001302>.
- [32] H.B. Brewer Jr., Hypertriglyceridemia: changes in the plasma lipoproteins associated with an increased risk of cardiovascular disease, *Am. J. Cardiol.* 83 (9B) (1999) 3F–12F, [https://doi.org/10.1016/s0002-9149\(99\)00308-2](https://doi.org/10.1016/s0002-9149(99)00308-2).
- [33] M. Miller, P. Langenberg, S. Havas, Impact of lowering triglycerides on raising HDL-C in hypertriglyceridemic and non-hypertriglyceridemic subjects, *Int. J. Cardiol.* 119 (2) (2007) 192–195, <https://doi.org/10.1016/j.ijcard.2006.07.132>.
- [34] N.R. Madamanchi, A. Vendrov, M.S. Runge, Oxidative stress and vascular disease, *Arterioscler. Thromb. Vasc. Biol.* 25 (1) (2005) 29–38, <https://doi.org/10.1161/01.ATV.0000150649.39934.13>.
- [35] K. Cervantes Gracia, D. Llanas-Cornejo, H. Husi, CVD and oxidative stress, *J. Clin. Med.* 6 (2) (2017) 22, <https://doi.org/10.3390/jcm6020022>. Published 2017 Feb 20.
- [36] K. D Bruce, M. Tang, P. Reigan, R. H Eckel, Genetic variants of lipoprotein lipase and regulatory factors associated with alzheimer's disease risk, *Int. J. Mol. Sci.* 21 (21) (2020) 8338, <https://doi.org/10.3390/ijms21218338>. Published 2020 Nov 6.
- [37] M. Nejati, M.A. Atlasi, M. Karimian, H. Nikzad, A. Azami Tameh, Lipoprotein lipase gene polymorphisms as risk factors for stroke: a computational and meta-analysis, *Iran J Basic Med Sci* 21 (7) (2018) 701–708, <https://doi.org/10.22038/IJBMS.2018.29009.7001>.