

WellBeing International
WBI Studies Repository

7-26-2014

Infrared thermal image for assessing animal health and welfare

Irenilza Alencar Nääs
Universidade Federal da Grande Dourados (UFGD)

Rodrigo Garófallo Garcia
Universidade Federal da Grande Dourados (UFGD)

Fabiana Ribeiro Caldara
Universidade Federal da Grande Dourados (UFGD)

Follow this and additional works at: <https://www.wellbeingintludiesrepository.org/assawel>



Part of the [Animal Studies Commons](#), [Other Animal Sciences Commons](#), and the [Other Anthropology Commons](#)

Recommended Citation

Nääs, I. A., Garcia, R. G., & Caldara, F. R. (2014). Infrared thermal image for assessing animal health and welfare. *JABB-Online Submission System*, 2(3), 66-72. DOI: <http://dx.doi.org/10.14269/2318-1265/jabb.v2n3p66-72>

This material is brought to you for free and open access by WellBeing International. It has been accepted for inclusion by an authorized administrator of the WBI Studies Repository. For more information, please contact wbisr-info@wellbeingintl.org.



Infrared thermal image for assessing animal health and welfare

Imagem termográfica infravermelha na avaliação do bem-estar animal

Irenilza Alencar Nääs ▪ Rodrigo Garófallo Garcia ▪ Fabiana Ribeiro Caldara

IA Nääs (Corresponding autor) ▪ RG Garcia ▪ FR Caldara
Universidade Federal da Grande Dourados (UFGD)

email: irenilza@gmail.com

Received: 03 March 2014 ▪ Accepted: 30 June 2014

Abstract Infrared thermal imaging is a non-destructive testing technology that can be used to determine the superficial temperature of objects. This technology has an increasing use in detecting diseases and distress in animal husbandry within the poultry, pig and dairy production. The process can identify changes in peripheral blood flow from the resulting changes in heat loss and; therefore, have been a useful tool for evaluating the presence of disease, edema, and stress in animals. This paper reviews the current literature related to the use of infrared technology and discusses their results and implications in animal welfare issues, poultry, pig and bovine industry.

Keywords emissivity, husbandry, surface temperature, thermal environment

Introduction

All substance emits radiant energy as a consequence of its absolute temperature. The portion of the electromagnetic spectrum extending from approximately 0.1 to 100mm, (the visible and the infrared spectrum) is named thermal radiation (Incropera and DeWitt 2007). Radiation is a form of heat loss through infrared rays involving the transfer of heat from one object to another without physical contact. Skin emissivity is an important factor in determining the true skin temperature, and through the assessment of surface temperature it is possible to acquired knowledge regarding physical and healthy status of humans and other living creatures (Chiu et al 2005; Cook et al 2006; Bouzida et al 2009; Alsaood et al 2014)

Infrared thermography (IRT) is a non-destructive testing technology that can be used to determine the superficial temperature of objects. Thermal cameras collect infrared radiation emitted by the surface, convert it into electrical signals and create a thermal image showing the

Resumo Imagem térmica infravermelha é uma tecnologia experimental não destrutiva, que pode ser utilizado para determinar a temperatura superficial dos objetos. Esta tecnologia tem uma crescente aplicação na detecção de doenças e estresse em criação de animais: aves, suínos e produção de leite. Este processo pode detectar mudanças no fluxo de sangue periférico das alterações resultantes na perda de calor e, portanto, têm sido ferramenta útil para avaliar a presença de doença, edema e estresse nos animais. Este trabalho revisa a literatura atual em relação ao uso da tecnologia de infravermelho e discute seus resultados e implicações em questões de bem-estar animal, e na indústria de produção de aves, suínos e bovinos.

Palavras-chave emissividade, produção animal, temperatura superficial, ambiente térmico

body's superficial temperature distribution (Incropera and DeWitt 2007). In this process, each color expresses a specific temperature range, related to the defined scale. In homeothermic animals, thermoregulation is a key feature in the maintenance of homeostasis. Thermoregulatory abilities are strongly related to energy balance and animals are often seeking to limit the energy costs of normothermia (Donkoh 1989; Cooper and Washburn 1998; Yahav et al 1998; Aksit et al 2006). In the case of thermal changes, physiological mechanisms are enhanced, increasing rates of energy expenditure (Shinder et al 2007; Stewart et al 2008).

IRT allows the visualization of temperature distribution, and it can detect changes in peripheral blood flow from the resulting changes in heat loss and; therefore, have been a useful tool for assessing the presence of disease, edema, and stress in animals (Nikkhah et al 2005; Bouzida et al 2009; Alsaood et al 2014). Surface temperature measurements can be made without trouble and with high precision, especially on animal coats that have low heat

capacities (McCafferty et al 1998; Tessier et al 2003; Montanholi et al 2008).

This paper reviews the use of IRT technology in evaluating forms of disease and distress in animal production, and it presents cases reports related to the use of this technology in experimental procedures for assessing thermal stress in pigs and poultry.

Animal welfare assessment

The definition of animal welfare is generally accepted as the balance between the animal and its surrounding environment. In practice, this means to provide them with sufficient health and comfort, as well as avoiding stress of any order. This means that health, welfare, and productivity are intimately connected (Moura et al 2006). Dawkins (1990) support the idea that welfare is mainly dependent on what the animal feels more than its response; however, other authors identified welfare determinants as related to the degree of distress and suffering (Zhou and Yamamoto 1997; Yahav et al 2004; Nääs et al 2010). At the same time, the lack of effective assessments of animal welfare represents a great difficulty for the establishment of welfare regulations and the evolution of animal welfare information.

IRT has been used to evaluate the welfare of livestock under usual managing procedures or health status control in calves (Schaefer et al 2004; Stewart et al 2008), in poultry research (Yahav et al 2001; Yahav et al 2004; Cangar et al 2008), and also in pig production (Warriss et al 2006). It has also been useful for assessing welfare in wild animals (McCafferty et al 1998; Dawson et al 1999; Ward et al 1999). Based on the association between muscle temperature rise and early postmortem pH fall rate (Nicol and Scott 1990) IRT has also been applied to identify the temperature increase in pigs, in response to pre-slaughter handling with the objective of predicting meat quality variation (Weschenfelder et al 2013).

The phenomenon of stress-induced hyperthermia or psychogenic fever occurs in numerous species and is characterized by an increase in body temperature within 10 to 15 min of the onset of an emotional stressor. Studying the handling stress in broiler chickens, Edgar et al (2013) found that the eye temperature changed significantly during handling. The authors concluded that surface temperature changes assessed using IRT are sensitive to routine management such as handling and it represent a potentially useful method for evaluating stress-induced hyperthermia in chickens.

Surface temperature assessment

Several studies have been conducted using as IRT tool for assessment of animal welfare, especially in regard to

issues related to rearing thermal exchanges (Cook et al 2006; Bouzida et al 2009; Ferreira et al 2011). In previous researches, the source for assessing the heat exchange in livestock have been an animal and surroundings surface temperature recorded using IRT (Li et al 1991; Yahav et al 2004; Warriss et al 2006; Shinder et al 2007).

A key issue regarding surface temperature assessment is the material emissivity coefficient (ϵ). In a research related to animal husbandry, there have been various assumptions for establishing the skin temperature emissivity coefficient. For instance, the emissivity coefficient used was 0.94 for the regions with feathers and 0.95 for the featherless regions, which are within the range of emissivity values for biological material (Ferreira et al 2011). There are many suggestions for this subject in the literature. McCafferty et al (1998) indicated the emissivity value of 0.98 for feathers, when calculating the radiant heat loss of a barn owl. Dawson et al (1999) adopted the emissivity value of keratin equal to 0.8, when building a model of radiant heat transfer through the penguin coat. Ward et al (1999) and Cangar et al (2008) suggested the emissivity value of 0.95 for the whole bird. Malheiros et al (2000) used $\epsilon = 0.94$ to define the variation of the surface temperature in post hatch pullets.

However, surface temperatures data do not always explain the amount of heat exchange due to eventual body core temperature increase during metabolization of feed. Case et al (2012) showed that surface temperature traits explained only a small proportion of variation in feed intake, in turkeys. The authors report that the recording of infrared images was efficient and required minimal contact with the caged birds; however, the low correlations indicate that the IRT presented limited advantages for increasing the accuracy of selection for feed efficiency.

Using data on surface temperature measurements from IRT have also been successfully used to recognize orthopedic injuries in horses (Eddy et al 2001),

Pig production

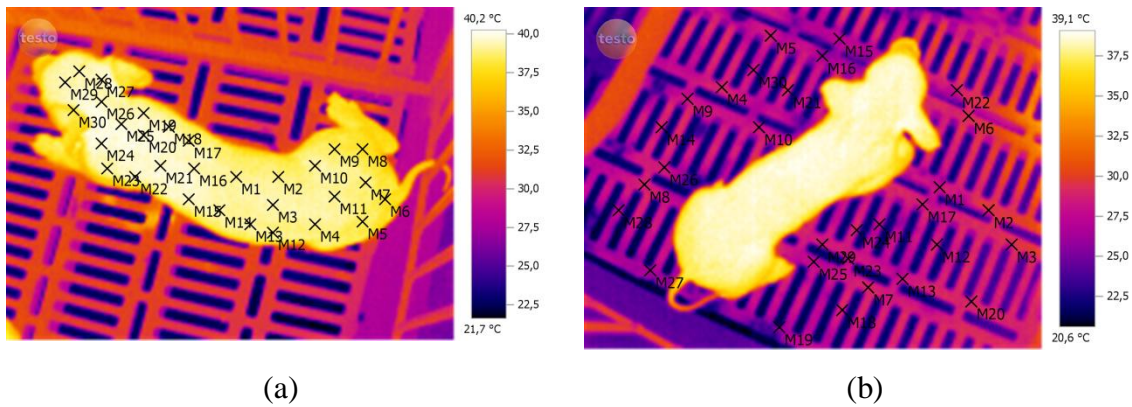
Warriss et al (2006) studied the variation of body temperature of groups of pigs using IRT, in order to evaluate the housing conditions, and found that IRT was an interesting and non-invasive way of assessing surface temperature for determining heat exchange. Other research in pig production is reported by Kastberger and Stachl (2003).

Sykes et al (2012) used IRT to discriminate between estrus and diestrus phases of the porcine estrous cycle. The authors found that vulva thermal images were positively correlated with ambient temperature, and vulva thermal temperatures were greater at estrus than at diestrus (36.6 ± 0.2 °C and 33.4 ± 0.3 °C vs. 35.6 ± 0.3 °C and 31.8 ± 0.6 °C, respectively), whereas they did not differ between stages of the cycle.

Graciano et al (2014) used IRT images to detect edema in pigs. The authors showed the efficiency of image analysis in early detection of arthritis.

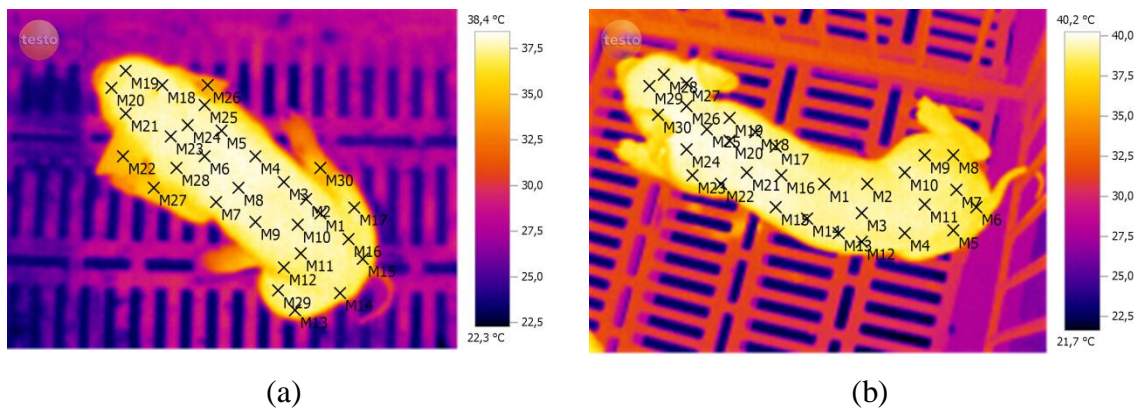
Caldara et al (2014) evaluated by means of IRT images of the rearing environment and the effect of it in the weight of piglets at birth. The authors calculated piglets heat loss in the first hours after birth and reported the value of proper temperature in the nursing rooms. Thermal exchanges by conduction between newborn piglets and the farrowing

crates' floor are high, and it may affect piglets' body heat loss decreasing their performance (Figure 1). The authors also observed by recording IRT images that piglets with lower birth weight tend to have higher drop in body temperature in the first hour of life than others, a fact related to their smaller body energy, making it more disposed to hypothermia, thus suggesting particular attention to newborns of low birth weight (Figure 2).



(a) (b)
Source: Caldara et al (2014)

Figure 1 Thermography images in selected points to present the difference on the surface temperature of the piglet's skin (a), and the floor of rear part of farrowing crates (b)



(a) (b)
Source: Caldara et al (2014)

Figure 2 Thermography images and selected points to present the difference on the surface temperature of the piglet's skin reared in cold stress (a) and at the thermal neutral zone (b), 15 min after birth

IR images were also used successfully to verify the importance of the presence and proper functioning of cooling water sprinkler systems in piggeries rest before slaughter, on the maintenance of body temperature of the animals and consequently their thermal comfort (Caldara et al 2014).

IRT images were also used successfully to verify the quality of the presence and proper functioning of cooling water sprinkler systems in piggeries rest before slaughter, on the maintenance of body temperature of the animals and

consequently their thermal comfort (Caldara et al 2014). Analyzing the IRT images obtained by comparing the use of intermittent water sprinkler systems during the management of pigs unloading and waiting for slaughter, at the slaughterhouse, the authors showed that intermittent water sprinkler systems at intervals 30 minutes during this period are effective in reducing the surface temperature (Figure 3). However, they point out that planning the proper operation of sprinklers (interval between sprays and duration of

spraying) are conditioned to both ambient temperature and relative humidity.

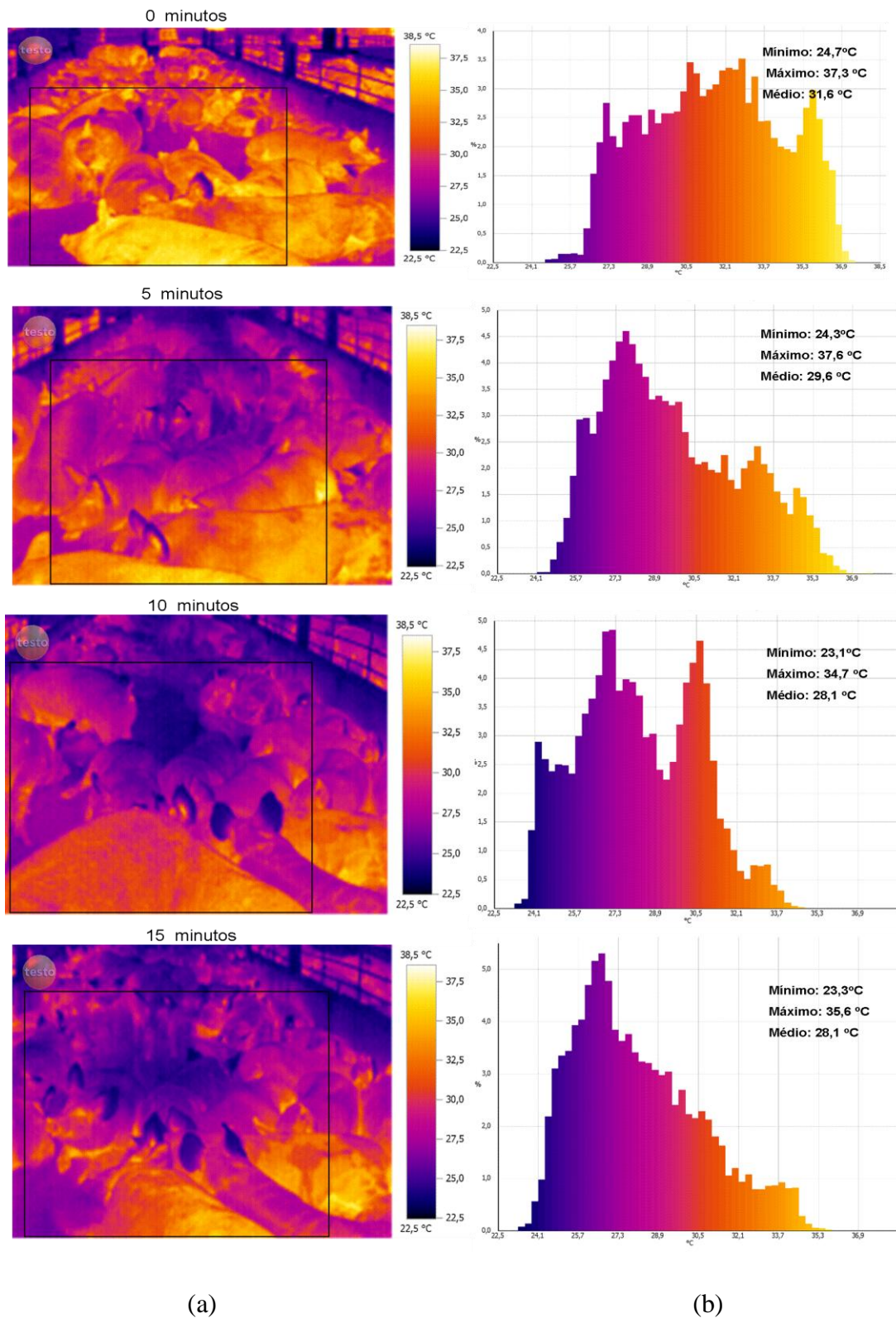


Figure 3 Thermal images of the pigs recorded in rest pens, 180 min after the animals arrive in the slaughterhouse, in time equivalent 0, 5, 10 and 15, after activation of the sprinklers (a), and histogram of the surface temperature calculated based on the area marked on the image (b).

Poultry production

IRT images were used by Ferreira et al (2011) to estimate the effectiveness of using images of infrared thermography in assessing the loss of sensible heat in pullets fed different dietary energy levels. Results indicated that IRT was efficient to predict the excess of metabolic heat loss due to a high energy diet. Average surface temperature of the body area and the flock were calculated using the surface temperature recorded at 100 spots within the image (Figure 4). Total radiant heat loss was calculated based on the average surface temperature data, and results showed that pullets fed the high-energy diet presented higher metabolic energy loss than the baseline.

Several studies on poultry production used IRT to predict heat exchange between the bird and the environment (Malheiros et al 2000; Yahav et al 2001; Tessier et al 2003;

Cook et al 2006), while others estimated feed consumption decrease during heat stress (Zhou and Yamamoto 1997; Cooper and Washburn 1998). Malheiros et al (2000) found that body weight declined in chicks reared at 20°C, and radiant heat loss was nine times higher than for the birds kept at 35°C at 7 days of age.

Using IRT images to assess the effects of exposure of commercial laying hens to cold, Alves et al (2012) found that birds under cold stress conditions spent about four times more energy trying to maintain body temperature. Due to its limited capacity to consume food they have been unable to generate enough metabolic heat to balance these losses and maintain their body temperature, thereby causing a reduction in egg production (Figure 5).

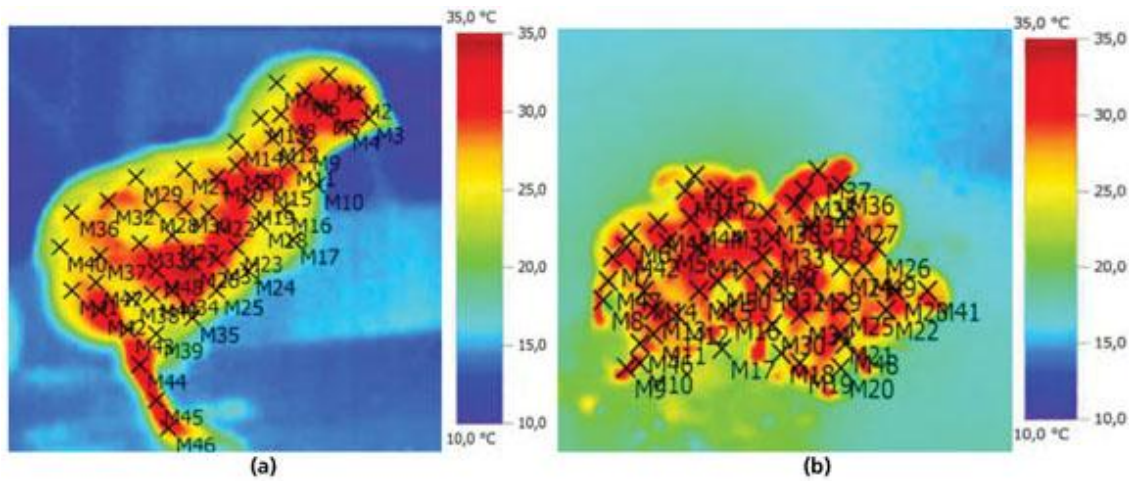
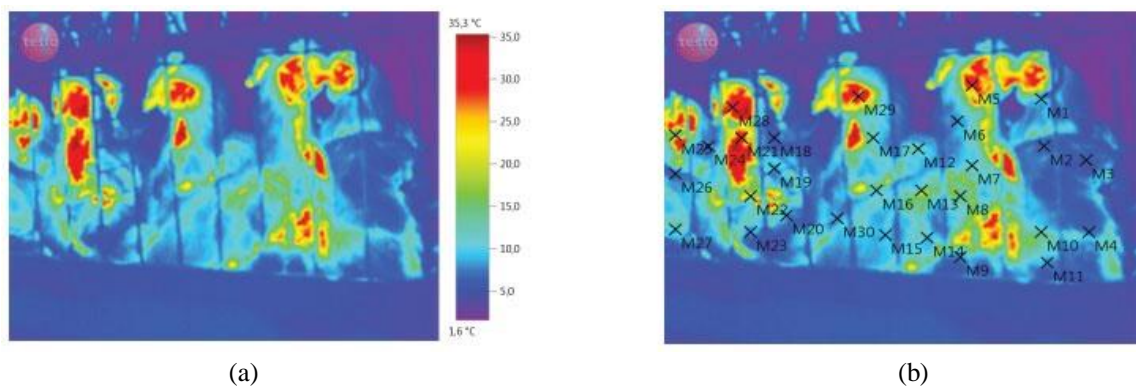


Figure 4 Infrared thermal image of a pullet (a) and the flock (b) with the spots where the surface temperature was marked and registered for calculating the total heat loss



Source: Alves et al (2012)

Figure 5 Infrared thermal image of caged laying hens (a), and the spots where the surface temperature was assessed (b)

Dairy and cattle production

In bovine medicine, IRT is used primarily for diagnostic purposes, but also for assessment of animal welfare and even feed utilization efficiency. Uses of IRT in the dairy industry include early detection of estrus, mastitis and lameness. Previous studies have focused on the use of infrared thermography to detect mastitis much earlier than previously possible (Stewart et al 2005).

Previous studies used IRT analysis as a method for early detection of animals infected with bovine viral diarrhoea virus or bovine respiratory disease using facial scans (Schaefer et al 2012). IRT has been also identified as a possible detection method for laminitis in lactating dairy cattle (Nikkhah et al 2005). These studies have concluded that while IRT provides an additional perspective on disease and injury, it should complement traditional diagnostics methods (Schaeffer et al 2004; Alsaad et al 2014).

The possibility that changes in eye temperature, measured using IRT, can detect stress in dairy cattle was examined by Schaefer et al (2012). Despite the findings using IRT, the increases in eye temperature following catheterization might need a cognitive component for assessing an appropriate animal response.

References

- Aksit M, Yalçın S, Özkan S et al (2006) Effects of temperature during rearing and crating on stress parameters and meat quality of broilers. *Poultry Science* 85:1867-1874.
- Alves FMS, Felix GA, Almeida Paz ICL et al (2012) Impact of exposure to cold on layer production. *Brazilian Journal of Poultry Science* 14:223-226.
- Alsaad C, Syring J, Dietrich MG et al (2014) A field trial of infrared thermography as a non-invasive diagnostic tool for early detection of digital dermatitis in dairy cows. *The Veterinary Journal* 199:281-285.
- Bouzida N, Bendada A, Maldague XP (2009) Visualization of body thermoregulation by infrared imaging. *Journal of Thermal Biology* 34:120-126.
- Caldara FR, dos Santos LS, Machado ST, Moi M et al (2014) Piglets' surface temperature change at different weights at birth. *Asian Australasian Journal of Animal Science* 27: 431-438.
- Cangar Ö, Aerts J-M, Buyse J et al (2008) Quantification of the spatial distribution of surface temperatures of broilers. *Poultry Science* 87: 2493-2499.
- Case LA, Wood BJ, Miller SP (2012) Investigation of body surface temperature measured with infrared imaging and its correlation with feed efficiency in the turkey (*Meleagris gallopavo*). *Journal of Thermal Biology* 37:397-401.
- Chiu WT, Lin PW, Chiou HY et al (2005) Infrared thermography to mass-screen suspected SARS patients with fever. *Asia Pacific Journal of Public Health* 17:26-28.

Final remarks

Surface temperature is an important indicator of animals' illnesses and for estimation of their physiological status; therefore, surface temperature estimation should be fast and accurate. In practice, different methods may be used for recording skin temperature; however, with IRT images analysis for estimating animals' body temperature and forecasting its implication is becoming more popular.

The surface temperature distribution is shown by the regions of different colors, and the image might be analyzed by scanning the surface by points, or focusing the thermal radiation of the surface with a number of pixels sufficient for getting precise images.

Nowadays most affordable cameras with adequate measurement accuracy are developed to have the thermal accuracy up to $\pm 1^{\circ}\text{C}$ and opportunity to save both thermal and visual images. Thermographic equipment has found increasing applicability in human and veterinary medicine, as infrared thermography is a non-invasive safe method for evaluating surface temperature and its derivative implications.

- Cook NJ, Smykot AB, Holm DE et al (2006) Assessing feather cover of laying hens by infrared thermography. *Journal of Applied Poultry Research* 15: 274-279.
- Cooper MA, Washburn KW (1998) The relationships of body temperature to weight gain, feed consumption, and feed utilization in broilers under heat stress. *Poultry Science* 77: 237-242.
- Dawkins MS (1990) From an animal's point of view: Motivation, fitness and animal welfare. *Behavioral and Brain Sciences* 13:1-9.
- Dawson C, Vincent JFV, Jeronimidis G et al (1999) Heat transfer through penguin feathers. *Journal of Theoretical Biology* 199: 291-295.
- Donkoh A (1989) Ambient temperature: a factor affecting performance and physiological response of broiler chickens. *International Journal of Biometeorology* 33: 259-265.
- Eddy AL, Van Hoogmoed LM, Snyder JR (2001) The role of thermography in the management of equine lameness. *The Veterinary Journal* 162: 172-181.
- Edgar JL, Nicol CJ, Pugh CA et al (2013) Surface temperature changes in response to handling in domestic chickens. *Physiology & Behavior*, Volume 119, 2 July 2013, Pages 195-200
- Ferreira VMOS, Francisco NSI, Belloni MI et al (2011) Infrared thermography applied to the evaluation of metabolic heat loss of chicks fed with different energy densities *Brazilian Journal of Poultry Science* 13: 113-118.

- Graciano DE, Nääs IA, Caldara FR et al (2014) Identificação de artrite em suíno utilizando imagem termográfica. *Boletim de Indústria Animal (Online)* 71: 58-62.
- IUPS Thermal Commission. 2001. Glossary of terms for thermal physiology. *The Japanese Journal of Physiology* 51: 245-280.
- Kastberger G, Stachl R (2003) Infrared imaging technology and biological applications. *Behavior Research. Methods, Instruments and Computers* 35: 429-439.
- Li Y, Ito T, Yamamoto S (1991) Diurnal variation in heat production related to some physical activities in laying hens. *British Poultry Science* 32: 821-827.
- Malheiros RD, Moraes VMB, Bruno LDG et al (2000) Environmental temperature and cloacal and surface temperatures of broilers chicks in first week post hatch. *Journal of Applied Poultry Research* 9: 111-117.
- McCafferty DJ, Moncrieff JB, Taylor R et al (1998) The use of IR thermography to measure the radiative temperature and heat loss of a barn owl (*Tyto alba*). *Journal of Thermal Biology* 23: 311-318.
- Montanholi YR, Odongo NE, Swanson KC et al (2008) Application of infrared thermography as an indicator of heat and methane production and its use in the study of skin temperature in response to physiological events in dairy cattle (*Bos taurus*). *Journal of Thermal Biology* 33: 468-475.
- Moura DJ, Nääs IA, Pereira DF et al (2006) Animal welfare concepts and strategy for poultry production: a review. *Revista Brasileira de Ciência Avícola* 8: 137-147.
- Nääs IA, Romanini C E B, Neves DP et al (2010) Broiler surface temperature distribution of 42 day old chickens. *Scientia Agricola*, 67(5), 497-502.
- Nicol CJ, Scott GB (1990) Pre-slaughter handling and transport of broiler chickens. *Applied Animal Behaviour Science* 28: 57-73.
- Nikkhah A, Plaizier JC, Einarson MS et al (2005) Infrared thermography and visual examination of hooves of dairy cows in two stages of lactation. *Journal of Dairy Science* 88: 2749-2753.
- Schaefer AL, Cook N, Tessaro SV et al (2004) Early detection and prediction of infection using infrared thermography. *Canadian Journal of Animal Science* 84: 73-80.
- Schaefer AL, Cook NJ, Bench C et al (2012) The non-invasive and automated detection of bovine respiratory disease onset on receiver calves using infrared thermography. *Research in Veterinary Science* 93: 928-935.
- Shinder D, Rusal M, Tanny J (2007) Thermoregulatory response of chicks (*Gallus domesticus*) to low ambient temperatures at an early age. *Poultry Science* 86: 2200-2209.
- Stewart M, Webster JR, Schaefer AL et al (2005) Infrared thermography as a non-invasive tool to study animal welfare. *Animal Welfare* 14: 319-325.
- Stewart M, Stafford KJ, Dowling SK et al (2008) Eye temperature and heart rate variability of calves disbudded with or without local anaesthetic. *Physiology & Behavior* 93:789-797.
- Sykes DJ, Couvillion JS, Cromiak A et al (2012) The use of digital infrared thermal imaging to detect estrus in gilts. *Theriogenology* 78:147-152.
- Tessier M, Du Tremblay D, Klopfenstein C et al (2003) Abdominal skin temperature variation in healthy broiler chickens as determined by thermography. *Poultry Science* 82: 846-849.
- Ward S, Rayner JMV, Möller U et al (1999) Heat transfer from starling *Sturnus vulgaris* during flight. *Journal of Experimental Biology* 202: 1589-1602.
- Warriss PD, Pope SJ, Brown SN et al (2006) Estimating the body temperature of groups of pigs by thermal imaging. *Veterinary Record* 158:331-334.
- Weschenfelder AV, Saucier L, Maldague X et al (2013) Use of infrared ocular thermography to assess physiological conditions of pigs prior to slaughter and predict pork quality variation. *Meat Science* 95: 616-620
- Yahav S, Luger D, Cahaner A et al (1998) Thermoregulation in naked neck chickens subjected to different ambient temperatures. *British Poultry Science* 39: 133-138.
- Yahav S, Straschnow A, Vax E et al (2001) Air velocity alters broiler performance under harsh environmental conditions. *Poultry Science* 80: 724-726.
- Yahav S, Straschnow A, Luger D et al (2004) Ventilation, sensible heat loss, broiler energy, and water balance under harsh environmental conditions. *Poultry Science* 83:253-258.
- Zhou WT, Yamamoto S (1997) Effects of environmental temperature and heat production due to food intake on abdominal temperature, shank skin temperature and respiration rate of broilers. *British Poultry Science* 38: 107-114.