

WellBeing International

WBI Studies Repository

2007

Chimpanzee Research: An Examination of Its Contribution to Biomedical Knowledge and Efficacy in Combating Human Diseases

Jarrold Bailey

New England Anti-Vivisection Society

Jonathan Balcombe

Physicians Committee for Responsible Medicine

Theodora Capaldo

New England Anti-Vivisection Society

Follow this and additional works at: https://www.wellbeingintludiesrepository.org/acwp_arte



Part of the [Animal Experimentation and Research Commons](#), [Animal Studies Commons](#), and the [Other Medical Sciences Commons](#)

Recommended Citation

Bailey, J., & Balcombe, J. (2007). Chimpanzee research: an examination of its contribution to biomedical knowledge and efficacy in combating human disease. NEAVS Commissioned report, see www.releasechimps.org.

This material is brought to you for free and open access by WellBeing International. It has been accepted for inclusion by an authorized administrator of the WBI Studies Repository. For more information, please contact wbisr-info@wellbeingintl.org.



**Chimpanzee Research: An Examination of
Its Contribution to Biomedical Knowledge and
Efficacy in Combating Human Diseases**

Jarrod Bailey, Ph.D.

Project R&R: Release and Restitution for Chimpanzees in U.S. Laboratories
Boston, MA

Jonathan Balcombe, Ph.D.

Physicians Committee for Responsible Medicine
Washington, DC

Theodora Capaldo, Ed.D.

Project R&R: Release and Restitution for Chimpanzees in U.S. Laboratories
Boston, MA

Commissioned by **Project R&R: Release and Restitution for Chimpanzees in U.S. Laboratories**
a campaign of the New England Anti-Vivisection Society, Boston, MA

TABLE OF CONTENTS

SUMMARY	3
INTRODUCTION	4
METHODS	6
RESULT	8
Citations of chimpanzee studies	10
Papers describing human prophylactic, diagnostic or therapeutic methods.....	12
DISCUSSION	16
Contributions made by chimpanzee studies.....	16
Sources contributing to human medical papers.....	19
Human utility of medical papers citing chimpanzee studies.....	20
CONCLUSIONS	21
ACKNOWLEDGEMENTS	23
REFERENCES	24

Detailed reviews of all papers involved in this citation analysis can be found in the supplement accompanying this paper.

SUMMARY

Research on captive chimpanzees incurs considerable animal welfare, ethical and financial costs. Advocates of such research claim these costs are outweighed by substantial advancements in biomedical knowledge, and that the genetic similarity of chimpanzees to humans enables the former to make critical contributions to preventing, diagnosing and combating human diseases. To assess these claims, we examined the disciplines investigated in 749 studies of captive chimpanzees published from 1995-2004 inclusive, and subjected 95 randomly selected papers to a detailed citation analysis:

49.5% (47/95) of papers had not been cited at the time of this study; 38.5% (34/95) were cited by 116 papers that did not describe well-developed methods for combating human diseases; 14.7% (14/95) of these chimpanzee studies were cited by (a total of 27) papers describing well-developed prophylactic, diagnostic or therapeutic methods for combating human diseases. Close examination of these 27 human medical papers revealed that *in vitro* research, human clinical and epidemiological investigations, molecular assays and methods, and genomic studies, contributed most to their development. Duplication of human outcomes, inconsistency with other human or primate data, and other causes resulted in the absence of any chimpanzee study demonstrating an essential contribution, or, in most cases, even a significant contribution of any kind, towards the development of the described human treatment.

INTRODUCTION

In a recent plea for increased research on captive chimpanzees, VandeBerg and Zola (VandeBerg and Zola 2005) asserted in *Nature* that, “*Many advances from biomedical research with chimpanzees have been published in the past one to two years, demonstrating that rapid medical progress pertinent to a wide range of human diseases is being made through the use of chimpanzees.*”

Claims such as these, made particularly by many researchers that currently use non-human primates (NHPs) or have used them in the past, are usually based on the ostensible similarities of chimpanzees to humans. Of all NHPs, the common chimpanzee (*Pan troglodytes*) and bonobo (“pygmy chimpanzee,” *Pan paniscus*) are genetically most similar to humans. Though estimates initially suggested this similarity was around 98.5 - 99%, more recent analyses suggest the figure may be in the region of 95 - 96% (Britten 2002; [Varki and Altheide 2005](#)). Either way, these figures suggest that chimpanzees would be the most likely non-human species to be of assistance in the development of diagnostic or therapeutic techniques for combating human diseases. NHPs are used in areas of research as diverse as hematology, immunology, virology, pharmacology ([Herodin et al., 2005](#)), and neurotoxicology (Evans 1990), to investigate potential bio-terrorism agents such as Ebola and Lassa viruses, anthrax and the plague ([Patterson and Carrion 2005](#)), and even to examine the physiological effects of the environment in space ([Pesquies et al., 1978](#)).

Chimpanzees specifically have recently been involved in research in most of these areas, but chiefly in studies into acquired immunodeficiency syndrome (AIDS) and hepatitis C virus ([Herodin et al., 2005](#)).

Research on captive chimpanzees remains controversial, however, with opponents citing numerous and varied animal welfare, ethical, scientific and financial concerns ([Sauer 2000](#); [Thew 2000](#)). Because of these concerns, some form of scientific evaluation and objective appraisal of the necessity and human benefits derived from research on captive chimpanzees is imperative. Notably, captive chimpanzee research must be regarded as a “special case;” most of the public, and indeed much of the scientific community, agree that the associated ethical cost at the very least demands a results-oriented pay-off in the form of substantial, tangible, human medical progress. Determining the scientific justification for chimpanzee research is paramount: if it exists, then concerns can be addressed and weighed against benefits. The argument to end chimpanzee research may become too strong to be effectively countered, if scientific justification is hard to come by other than in the form of speculative claims emanating from individuals closely involved in it.

A first step in assessing the merits of research on captive chimpanzees is to obtain a clear picture of the biomedical disciplines examined by such research. Some work has already been done in this area: based on 184 grant abstracts filed in the Computer Retrieval of Information on Scientific Projects (CRISP) database and 89 journal articles cited in PubMed (the U.S. National Library of Medicine's premier bibliographic database), Conlee et al. (Conlee et al., 2004) provided some initial indications of the disciplines investigated by chimpanzee research within the U.S. In addition, Carlsson et al. (Carlsson et al., 2004) surveyed 2,937 articles published in 2001 describing 4,411 NHP studies using over 41,000 animals worldwide, though only a small minority of these were studies of chimpanzees.

Here we assess the utility of chimpanzee research *in combating human disease*, using a two-phase approach: 1) we determined the frequency with which chimpanzee research published in peer-reviewed journals was cited by papers describing human prophylactic, diagnostic or therapeutic methods (using the Cochrane Collaboration's guidelines for systematic reviews (Higgins and Green 2005) where appropriate), and 2) we quantitatively and qualitatively assessed the nature and importance of the contributions provided by the cited chimpanzee studies. The survey was limited to major biomedical bibliographic databases likely to contain human medical papers, and attention was focused on research on captive chimpanzees because such research has raised the most concerns.

METHODS

We searched three biomedical bibliographic databases for papers describing research involving living chimpanzees or chimpanzee tissue from 1995 to 2004 inclusive: CAB Abstracts, the most comprehensive bibliographic database covering the applied life sciences, containing over 4.5 million records (Anonymous 2006*a,b*); EMBASE, the Excerpta Medica database, which is a biomedical and pharmacological database containing over 10 million records (Anonymous 2006*c*); and Medline, the premier medical and allied health profession database, containing over 12 million records (Anonymous 2006*d*). Jointly these databases included over 6,000 biomedical journals and thousands of other scientific documents sourced from more than 140 countries. Where appropriate, we also applied the Cochrane Collaboration's guidelines for systematic reviews (Higgins and Green 2005), applying many of their recommendations including those regarding protocol development; data collection, analysis and interpretation; and report content and structure.

All titles, abstracts, and associated fields were searched for “chimpanzee,” “bonobo,” “*Pan troglodytes*,” and “*Pan paniscus*,” limiting the search to documents with abstracts. These included:

- Studies of captive chimpanzees;
- Studies of biometric information taken from captive chimpanzees, such as MRI scans;
- Studies of fresh or preserved chimpanzee tissues, other than those specified below.

They excluded:

- Studies of free-living chimpanzees;
- Veterinary case reports of the diagnosis, treatment or post-mortem examination of naturally-ill chimpanzees, whether in captivity or not;
- Genome studies (excluding those of experimentally infected chimpanzees);
- Studies of skeletal anatomy (museum specimens were often used);
- Studies of cell lines (although cell samples obtained from captive chimpanzees were included);
- Studies of chimpanzee blood where the source was not specified;
- Secondary analyses of data obtained in primary studies.

From 749 chimpanzee studies that met the inclusion criteria, we randomly selected a subset of 100 studies using the “Research Randomizer” random number generator (www.randomizer.org). This was done for logistical reasons; performing a detailed citation analysis for 749 studies is a mammoth undertaking. Four of these citing papers were not available through the bibliographic databases used, and one additional study was cited only by a paper for which no abstract was available. Our analysis criteria (including a requirement for readily-accessible papers with published abstracts) led to these studies being excluded from further consideration, leaving 95 chimpanzee studies in the analysis. This sample size represents a number exceeding the sample size required to achieve statistical significance, based on the normal

approximation to the binomial distribution for a 95% confidence level with an error of plus or minus 10%.

These 95 studies were examined to determine the frequency with which they were cited by papers subsequently published in peer-reviewed journals indexed by the named bibliographic databases, as of January 2006. The species and biomedical disciplines that were the focus of these citing papers were also determined.

In particular, where abstracts of citing papers described prophylactic, diagnostic or therapeutic methods with clear potential for combating human diseases, the full-text of the citing articles (human subjects) was reviewed to determine the contribution of the cited chimpanzee study to the development of the method described, in comparison with other cited sources of knowledge. In each case, both the entire citing and cited papers were thoroughly examined, related articles sought (via direct links from literature databases and also via specific new database searches) to determine how the method had progressed since publication of the citing paper, and (where appropriate) clinical trial data obtained to assess the relevance and application of the method to human clinical practice. The contribution of a cited chimpanzee study was assessed by appraising the context of its citation, and by scanning all the references in the citing paper to derive the main contributors to the findings reported by it.

RESULTS

Using the specified search terms, 2400 abstracts were found dating up to 28th August 2005. Of these, 749 were studies of captive chimpanzees or chimpanzee tissue that met our inclusion criteria.

We classified the areas of investigation of all 749 chimpanzee papers into several categories as follows:

48.5% (363/749) of these were biological investigations, and 41.5% (311/749) were virology experiments (**Figure 1**). Biological investigations comprised nine disciplines, of which the most frequent were cognition/neuroanatomy/neurology (36.6%, 133/363) and behavior/communication (20.7%, 75/363) (**Figure 2**). Virological investigations involved 30 different types of virus, of which the most frequent were hepatitis C virus (HCV) and human immunodeficiency virus (HIV), each of which comprised 31.2% (97/311) of all virology experiments (**Figure 3**).

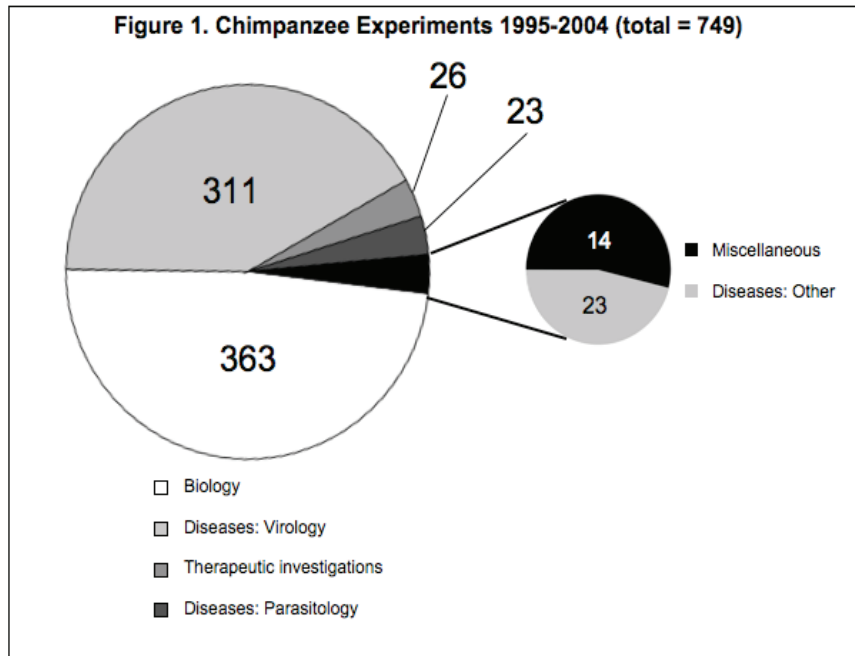


Figure 1: 2400 abstracts describing chimpanzee studies were found, dating from 1995 to 2004. Of these, 749 were studies that met the inclusion criteria. 48.5% (363/749) were biological experiments, and 41.5% (311/749) were virology experiments. A further 10% belonged to a variety of other categories.

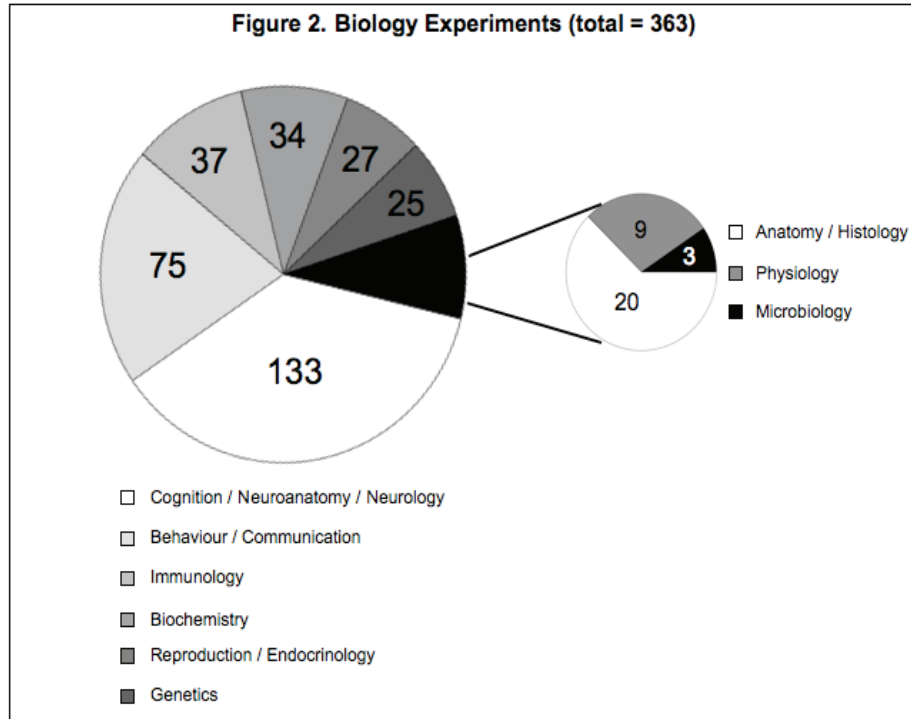


Figure 2: Biological investigations constituted 48.5% of chimpanzee experiments that met the inclusion criteria, performed between 1995 and 2004 inclusive. These were conducted in nine disciplines, of which the most frequent were cognition/neuroanatomy/neurology (36.6%, 133/363) and communication/communication (20.7%, 75/363).

Therapeutic investigations comprised 3.5% (26/749) of all chimpanzee experiments, of which almost two-thirds (16/26) investigated the pharmacological properties of a variety of compounds (**Figure 4**). Other investigations included the testing of surgical techniques or prostheses, anesthesiology, and toxicology of compounds other than pharmaceuticals. Parasitology experiments comprised 3.1% (23/749) of the total (**Figure 5**). Eight parasitic species were examined, of which the most frequent were the malaria protozoa *Plasmodium falciparum* and *P. ovale* (26.1%, 6/23), the roundworm *Onchocerca volvulus* (21.7%, 5/23), and the flatworm *Schistosoma mansoni* (17.4%, 4/23).

Other diseases and miscellaneous experiments combined comprised 3.5% (26/749) of all chimpanzee experiments, of which the most frequent were investigations of laboratory/husbandry techniques (42.3%, 11/26) and endotoxemia (30.1%, 8/26) (**Figure 6**). Three radiation studies were performed, and four other diseases were investigated: benign prostatic hyperplasia, Creutzfeldt-Jakob disease, gastrointestinal bacteriology (*Bacillus thuringiensis*), and tuberculosis (*Mycobacterium tuberculosis*).

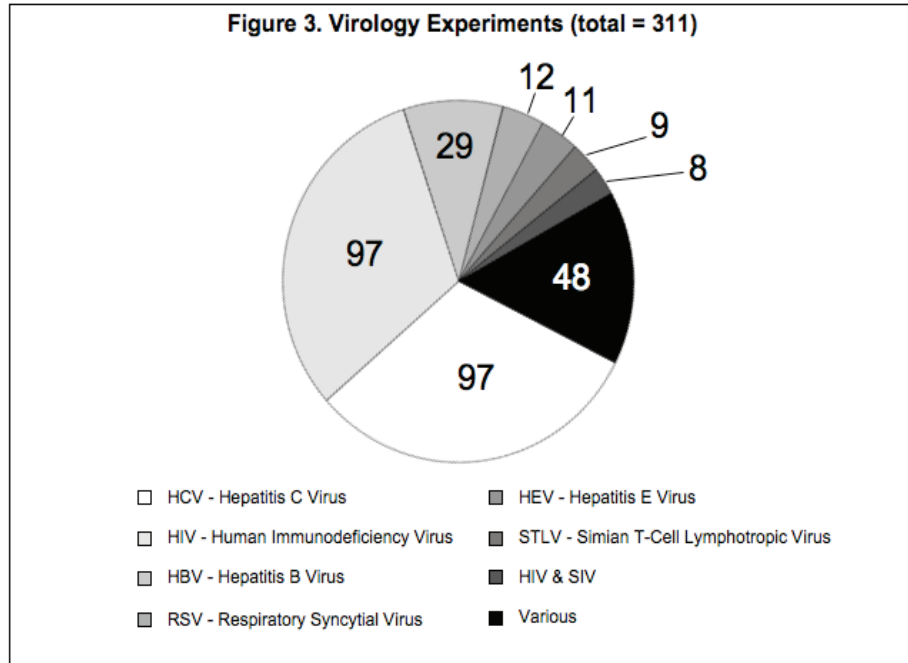


Figure 3: Virological investigations constituted 41.5% of chimpanzee experiments that met the inclusion criteria, performed between 1995 and 2004 inclusive. They were conducted in 30 disciplines, of which the most frequent were hepatitis C virus (HCV) and human immunodeficiency virus (HIV), each of which comprised 31.2% (97/311) of all virology experiments.

HCV = hepatitis C v, HIV = human immunodeficiency v, HBV = hepatitis B v, RSV = respiratory syncytial v, HEV hepatitis E v, STLV = simian T-cell lymphotropic v, SIV = simian immunodeficiency v, TTV = transfusion-transmitted v, FV = foamy v (human and simian FV), HAV = hepatitis A v, GBV-B = a close relative of HCV affecting tamarins, HV = herpes v, IV = influenza v, PIV = parainfluenza v, HCMV = human cytomegalovirus, HGV = hepatitis G v, HMPV = human metapneumovirus, H/S TLV = human/simian T-cell leukemia v, LCV = lymphocryptoviruses, RV2 = rhadinovirus (or gamma-2-herpesvirus) genogroup 2, VZV = varicella-zoster v, WMHBV = woolly monkey hepatitis B v.

Citations of chimpanzee studies

Notably, 49.5% (47/95) of these chimpanzee studies were not cited by any subsequent papers (**Figure 7**) (95% confidence interval for proportion of papers not cited in the original population of 749 papers = [0.395-0.595]. In other words, if this sampling was done repeatedly, one could be 95% certain that the proportion of chimpanzee studies not cited by any subsequent papers would be between these values, i.e. 39.5%-59.5%).

The remaining 48 chimpanzee experiments were cited by 143 papers for which abstracts were available. Some of these citing papers focused on humans alone; others did so in combination with other species, namely: bacteria (*Escherichia coli*); pigeons (*Columba livia*); bottlenose dolphins (*Tursiops truncatus*); dogs (*Canis familiaris*); Asian elephants (*Elephas maximus*); mice (natural or genetically-modified); pigs, and, unsurprisingly, a large variety of primate species: African green monkeys

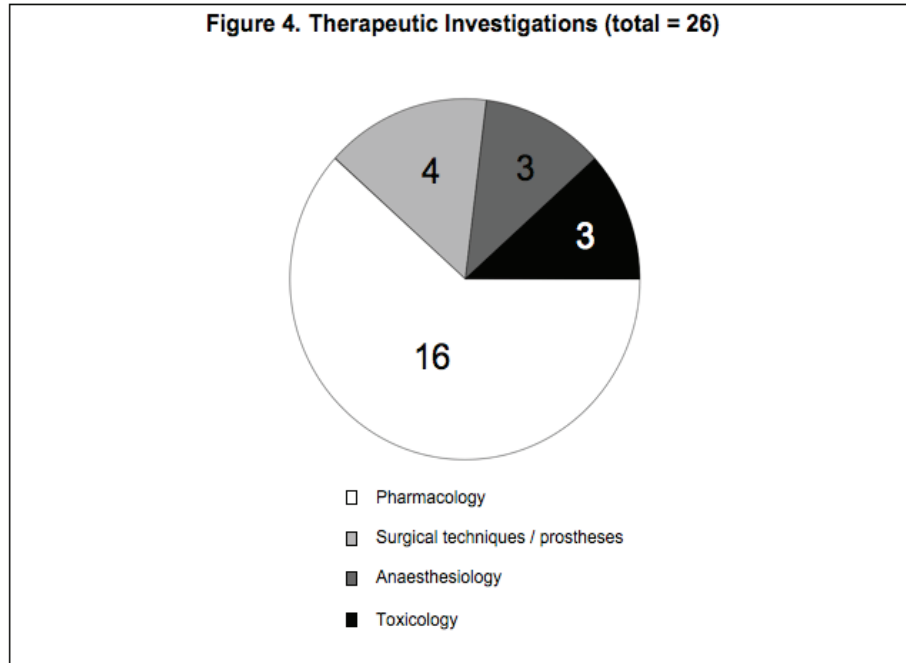


Figure 4: Therapeutic investigations comprised 3.5% (26/749) of all chimpanzee experiments, of which 61.5% (16/26) investigated the pharmacological properties of various compounds. Other investigations included the testing of surgical techniques or prostheses, anesthesiology and toxicology experiments.

(*Chlorocebus aethiops*); chimpanzees (*Pan troglodytes* or *Pan paniscus*); common marmosets (*Callithrix jacchus*); cotton top tamarins (*Saguinus oedipus*); cynomolgus macaques (*Macaca fascicularis*); Japanese macaques (*Macaca fuscata*); rhesus macaques (*Macaca mulatta*); squirrel monkeys (*Saimiri sciureus*); tufted capuchin monkeys (*Cebus apella*); olive baboons (*Papio anubis*); orangutans (*Pongo abelii* and *Pongo pygmaeus*) and western lowland gorillas (*Gorilla gorilla gorilla*).

A variety of biological disciplines were explored in these citing papers, with the hepatitis and human immunodeficiency viruses (HIV) featuring most prominently. Others included (in alphabetical order): asthma, autism, behavior, benign prostatic hyperplasia, cancer, chronic obstructive pulmonary disease (COPD), coxsackievirus B3, Epstein-Barr virus (EBV), genetic studies, human parainfluenza virus type 3, immunology, Kawasaki disease, laboratory techniques (including gene expression profiling and cDNA microarray interpretation), leukemia, malaria, neuroanatomy, neurology, organ transplantation, pathology (clinical), psychology, respiratory syncytial virus (RSV), rheumatoid arthritis (RA), rhinovirus colds, simian immunodeficiency virus (SIV), systemic lupus erythematosus (SLE), surgical techniques (cardiac allografts), toxicity (arsenic, transmissible spongiform encephalopathies (TSE)), and non-specific virology.

In addition to the 49.5% of papers in our random sample that were not cited, 35.8% (34/95) of those chimpanzee studies were cited only by papers (a total of 116) that did

not describe well-developed prophylactic, diagnostic or therapeutic methods for combating human diseases (**Figure 7**).

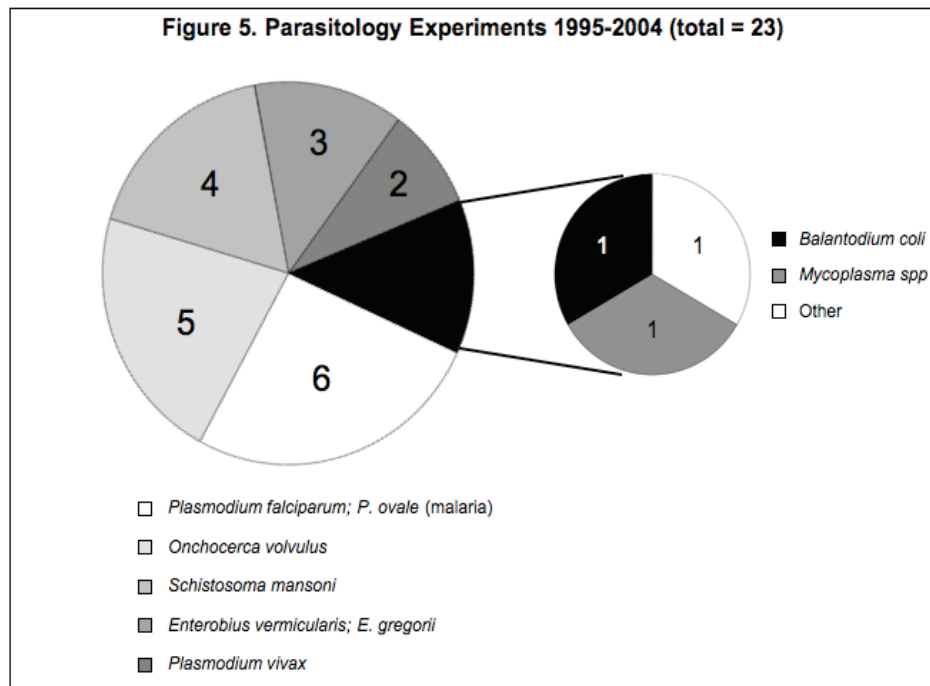


Figure 5: Parasitology experiments comprised 3.1% (23/749) of all chimpanzee experiments. Eight parasitic species were investigated, of which the most frequent were the malaria protozoa *Plasmodium falciparum* and *P. ovale* (26.1%, 6/23), the roundworm *Onchocerca volvulus* (21.7%, 5/23), and the flatworm *Schistosoma mansoni* (17.4%, 4/23).

Papers describing human prophylactic, diagnostic or therapeutic methods

Only 14.7% (14/95) of our random sample of chimpanzee studies were cited by a total of 27 papers that described human prophylactic, diagnostic or therapeutic methods. Of these, 5 described diagnostic methods and 22 described prophylactic and/or therapeutic methods for combating human diseases, which were either fully developed for human use or in the latter stages of development at the time of publication (**Figure 7**). Diseases examined in these citing papers included cancer (non-specific), COPD, EBV, hepatitis viruses A through G (HAV through HGV), hepatocellular carcinoma, HIV, malaria, organ transplant rejection, RSV, RA, rhinovirus colds, SLE and TSE. 63.0% (17/27) of these human medical papers were wide-ranging reviews of up to 300 studies, in which the cited chimpanzee study made only a minor contribution – as discussed later.

Detailed reviews of each of the 27 papers citing the 14 chimpanzee papers from our random sample (that had described human prophylactic, diagnostic or therapeutic methods) were carried out, to determine the contribution of the chimpanzee studies to the human prophylactic, diagnostic or therapeutic method described in the citing paper. These can be found in the supplement to this paper; a summary of the papers

citing the chimpanzee studies from our random sample, and of the cited chimpanzee studies, is provided in **Table 1**.

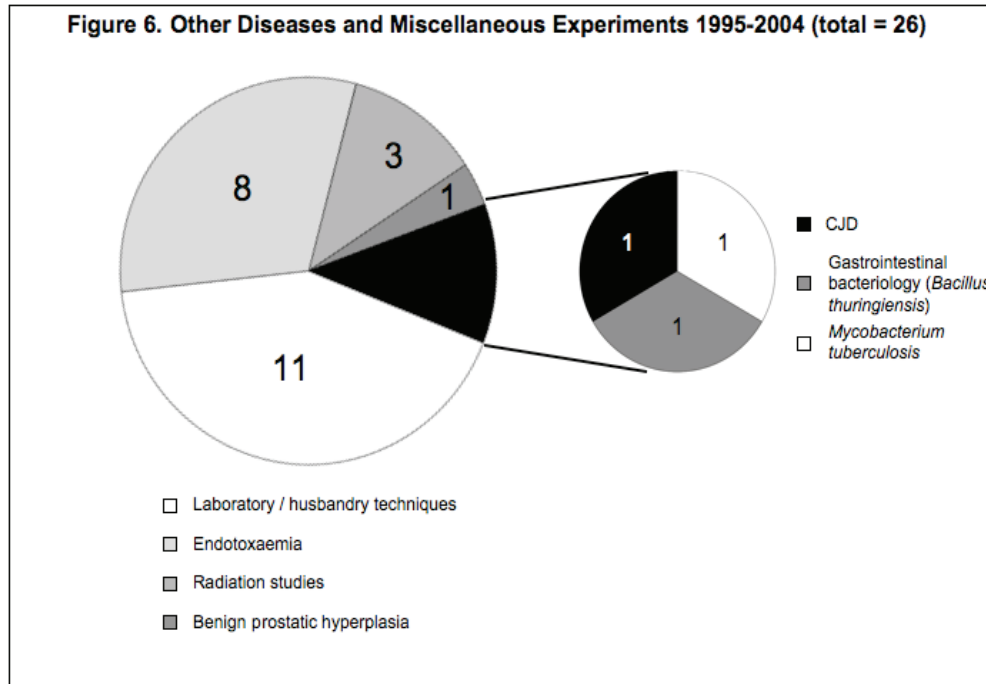


Figure 6: Other diseases and miscellaneous experiments comprised 3.5% (26/749) of all chimpanzee experiments when combined, of which the most frequent were investigations of laboratory/husbandry techniques (42.3%, 11/26) and endotoxemia (30.1%, 8/26). Three radiation studies were also conducted, and four other diseases were investigated, namely benign prostatic hyperplasia, Creutzfeldt-Jakob disease, gastrointestinal bacteriology (*Bacillus thuringiensis*), and tuberculosis (*Mycobacterium tuberculosis*).

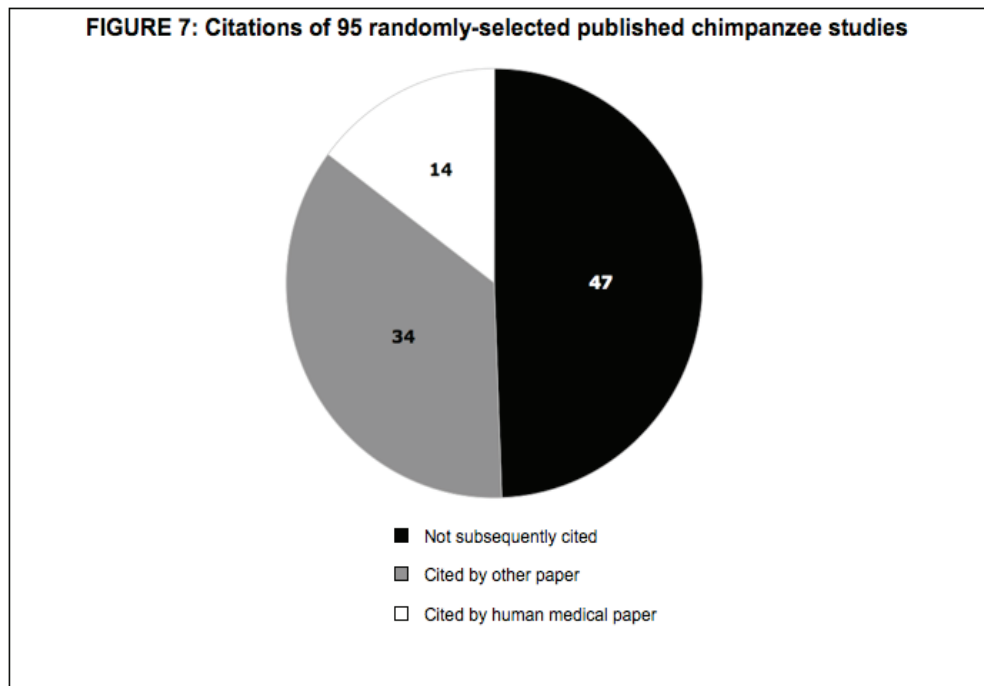


Figure 7: Citations of 95 randomly selected published chimpanzee studies. 49.5% (47/95) of these papers had not been subsequently cited in the scientific literature; (95% confidence interval for proportion of papers not cited in the original population of 749 papers = [0.395-0.595]). The remaining 48 chimpanzee experiments were cited by 143 papers. 35.8% (34/95) of these were cited only by 116 papers that did *not* describe well-developed prophylactic, diagnostic or therapeutic methods for combating human diseases. Just 14.7% (14/95) of our random sample of chimpanzee studies described human prophylactic, diagnostic or therapeutic methods, and these were cited by a total of 27 papers.

TABLE 1

CITING PAPER					CITED PAPER	
No.	Paper	Type	Topic	No. References	Paper	Times Cited
1	Tong & Stone (2003)	Review	Cancer (non-specific): therapeutic	155	Brams <i>et al.</i> (2001)	6
2	Suzuki <i>et al.</i> (2001)	Research paper	COPD: prophylactic	28	Huguenel <i>et al.</i> (1997)	6
3	Khanna <i>et al.</i> (1999)	Review	Epstein-Barr virus: prophylactic	112	Bertoni <i>et al.</i> (1998)	2
4	Koff (2002)	Review	HAV & HBV: prophylactic	85	Ogata <i>et al.</i> (1999)	5
5	Regev & Schiff (1999)	Review	Hepatitis viruses A-G: prophylactic & therapeutic	71	Mast <i>et al.</i> (1998)	13
6	McMahon <i>et al.</i> (2005)	Research paper	HBV: prophylactic	38	Ogata <i>et al.</i> (1999)	5
7	Karayiannis (2003)	Review	HBV: prophylactic & therapeutic	300	Pancholi <i>et al.</i> (2001)	5
8	Hüssy <i>et al.</i> (1997)	Research paper	HCV: diagnostic	24	Wang <i>et al.</i> (1996)	8
9	Feld & Hoofnagle (2005)	Review	HCV: therapeutic	91	Bigger <i>et al.</i> (2001)	10
10	Nakano <i>et al.</i> (1999)	Research paper	HCV: therapeutic	26	Wang <i>et al.</i> (1996)	8
11	Worm & Wirnsberger (2004)	Review	HEV: prophylactic methods	133	Mast <i>et al.</i> (1998)	13
12	Obriadina <i>et al.</i> (2002)	Research paper	HEV: diagnostic	32	Mast <i>et al.</i> (1998)	13
13	Kim & Wang (2003)	Review	Hepatocellular carcinoma: diagnostic	81	Bigger <i>et al.</i> (2001)	10
14	Armbruster <i>et al.</i> (2002)	Research paper	HIV: prophylactic & therapeutic	25	Conley <i>et al.</i> (1996)	10
15	Armbruster <i>et al.</i> (2004)	Research paper	HIV: prophylactic & therapeutic	27	Conley <i>et al.</i> (1996)	10
16	Bardsley-Elliott & Perry (2000)	Review	HIV: prophylactic & therapeutic	130	Grob <i>et al.</i> (1997)	2
17	Hone <i>et al.</i> (2002)	Review	HIV: prophylactic	108	Conley <i>et al.</i> (1996)	10
18	Sleasman & Goodenow (2003)	Review	HIV: prophylactic & therapeutic	93	Conley <i>et al.</i> (1996)	10
19	Yang <i>et al.</i> (1998)	Research paper	HIV: diagnostic	28	Conley <i>et al.</i> (1996)	10
20	Gallo (2002)	Review	HIV: therapeutic	258	Goh <i>et al.</i> (1998)	10
21	Moore & Hill (2004)	Review	Malaria: prophylactic	104	Pancholi <i>et al.</i> (2001)	5
22	Matthews <i>et al.</i> (2003)	Review	Organ transplant rejection: therapeutic	87	Newman <i>et al.</i> (2001)	2
23	Kneyber & Kimpen (2002)	Review	RSV: prophylactic	150	Crowe <i>et al.</i> (1999)	1
24	Hepburn <i>et al.</i> (2003)	Research paper	Rheumatoid arthritis: therapeutic	15	Newman <i>et al.</i> (2001)	2
25	Turner <i>et al.</i> (1999)	Research paper	Rhinoviral colds: prophylactic	34	Huguenel <i>et al.</i> (1997)	6
26	Gescuk & Davis (2002)	Review	SLE: therapeutic	77	Brams <i>et al.</i> (2001)	6
27	Brown (2005)	Review	TSE: diagnostic	26	Cervenakova <i>et al.</i> (2003)	1

Table 1: The twenty-seven medical papers citing publications from our random sample of 95 chimpanzee studies. Relevant details of each citing paper are summarized, along with information concerning the actual chimpanzee study cited in each case and the number of times these chimpanzee studies had been cited in total, at the time of our analysis. COPD = chronic obstructive pulmonary disease; HAV–HCV = hepatitis viruses A–C; HEV = hepatitis E virus; HIV = human immunodeficiency virus; RSV = respiratory syncytial virus; SLE = systemic lupus erythematosus; TSE = transmissible spongiform encephalopathy.

DISCUSSION

Contributions made by chimpanzee studies

For a variety of reasons, the chimpanzee studies from our random sample were found to be incidental to most of these human medical papers (Table 2).

TABLE 2

Reason for lack of contribution of chimpanzee study to human prophylactic, diagnostic or therapeutic method	Total number of reviews in category (of 27)	Specific reviews in category
Redundant (concurrent human experiments/"confirmations" of human data)	14	1, 5, 8-14, 16, 20, 24, 25, 27
Method not developed further (possibly due to chimpanzee/human differences)	7	2, 22-27
Chimpanzee study peripheral to human method described	7	3, 4, 6, 13, 15, 21, 25
Historical citation with no direct relevance	3	2, 17, 19
Inconsistent with data from other NHP studies	3	7, 17, 26
Inconsistent with human data	2	18, 24
Purely speculative in nature	2	3, 14
May have helped establish need for new diagnostic method but did not contribute further to its development.	1	12

Table 2: Reasons why the random sample of chimpanzee studies in our analysis did not contribute to the human prophylactic, diagnostic or therapeutic methods described in papers citing them. Each review detailed in this analysis fits into one or more of the eight categories shown. The total number of reviews in each category is shown, along with the specific reviews as numbered in the results section of this study. It should be noted, however, that due to the complex and overlapping nature of the studies and these categories, this table could not be considered to be exhaustive and all-inclusive.

In fourteen cases (Reviews 1, 5, 8-14, 16, 20, 24, 25 and 27; see Supplement) the chimpanzee studies appeared to be redundant, as humans or human sera were studied concurrently, or because they only served to confirm observations previously made in humans. In seven cases (Reviews 2 and 22-27) the method explored in the cited chimpanzee study was not developed further, sometimes because later clinical trials in humans failed to demonstrate safety or efficacy, contrary to positive results in chimpanzees. In seven cases (Reviews 3, 4, 6, 13, 15, 21 and 25) the chimpanzee study examined a disease or method peripheral to the prophylactic, diagnostic or therapeutic method described. In three cases (Reviews 2, 17 and 19) the chimpanzee study merely illustrated an historical finding, or was cited during historical discussions of attempts to combat the disease in question. In three cases (Reviews 7, 17 and 26) chimpanzee studies yielded results inconsistent with data from other NHP studies, while in two cases (Reviews 18 and 24) they yielded results inconsistent with human data. In two cases (Reviews 3 and 14) the cited chimpanzee studies were purely speculative in nature and had therefore not contributed to the concrete findings of the citing papers. In the remaining case (Review 12) the chimpanzee study may have helped establish the need for a new diagnostic method but did not contribute further to its development.

Five chimpanzee studies were cited by multiple human medical papers. The paper by Bigger *et al.* on the effects of HCV in chimpanzees (Bigger *et al.*, 2001), for example, was peripheral to Kim and Wang's paper (Kim and Wang 2003) describing a diagnostic method for the detection of HCC (Review 13), and served only to confirm what had already been observed in earlier human studies cited by Feld and Hoofnagle's paper (Feld and Hoofnagle 2005) on therapeutic methods for combating HCV (Review 9).

Conley *et al.*'s study (Conley *et al.*, 1996) of the prophylactic use of hMAbs in chimpanzees challenged with HIV-1 was also cited by five medical papers. In two studies of prophylactic and therapeutic methods for combating HIV (Armbruster *et al.*, 2002; 2004), prior positive results in the cited chimpanzee study suggested a potential field of further development with respect to hMAb choices used in combination prophylactic and therapeutic regimes (Reviews 14 and 15). The cited chimpanzee study did not, however, play an integral role in the citing studies, the purpose of which was to examine the safety, immunogenicity and pharmacokinetics of hMAb protocols in clinically healthy HIV-1-infected human volunteers. In papers by Yang *et al.* (Yang *et al.*, 1998) and Hone *et al.* (Hone *et al.*, 2002) describing the development of vaccines and a diagnostic method respectively for combating HIV, the cited chimpanzee study served only to demonstrate that effective HIV antibodies can indeed neutralize HIV in chimpanzees, to varying degrees, although conflicting results were observed in six other NHP studies cited by Hone *et al.* (Reviews 17 and 19). Other than through such peripheral means, the cited chimpanzee study did not contribute to the development of these prophylactic and diagnostic methods. The contribution of this study towards the development of prophylactic and therapeutic methods for combating HIV reviewed by Sleasman *et al.* (Sleasman and Goodenow 2003) was similarly limited by inconsistency with other cited human data (Review 18).

Mast *et al.*'s paper ([Mast *et al.*, 1998](#)) describing the poor concordance, sensitivity and variable efficiency of previously available HEV assays in chimpanzees, particularly against different HEV strains, was cited by Regev *et al.*'s review (Regev and Schiff 1999) of prophylactic and therapeutic options for combating HAV, HBV, HCV, HEV, HGV and TTV (Review 5), as well as by Obriadina *et al.*'s paper ([Obriadina *et al.*, 2002a](#)) describing the development of a diagnostic technique for the detection of HEV (Review 12). The chimpanzees in Mast's study were used as both positive and negative controls, though both were redundant for more relevant positive and negative human controls in the form of human sera were available concurrently. Additionally, one of Mast *et al.*'s key outcomes was the highly discrepant existing HEV assay results from U.S. blood donors that were presumed HEV negative, necessitating great caution when interpreting positive assay results. This outcome relied exclusively on human results.

Newman *et al.*'s paper ([Newman *et al.*, 2001](#)) demonstrating the safety and efficacy of Keliximab, a primatized IgG1 anti-CD4 mAb in modulating T-cell receptor responsiveness in chimpanzees, was cited by two medical papers. In Matthews *et al.*'s review ([Matthews *et al.*, 2003](#)) of therapeutic strategies designed to thwart organ transplant rejection, it was hoped that the targeting of cell-surface receptors might result in T-cell inactivation, thereby delaying allograft rejection because T-cell activation is central to the inflammation and tissue damage that precedes it (Review 22). [Hepburn *et al.*, 2003](#)) also described a clinical trial of Clenoliximab, an anti-CD4 mAb antibody proposed for the treatment of RA (Review 24). However, a later phase II human clinical trial of an analogue drug (Zanolimumab) failed to demonstrate efficacy at combating the disease, resulting in abandonment of the development of this drug for RA patients and casting doubts on the efficacy of anti-CD4 mAbs in decreasing allograft rejection.

A paper describing chimpanzee studies of DNA-based vaccines ([Pancholi *et al.*, 2001](#)) was cited both by Karayiannis' review of the prophylactic and therapeutic options available for combating HBV ([Karayiannis 2003](#)) (Review 7), and by Moore and Hill's review of strategies for the development of a malaria vaccine (Moore and Hill 2004) (Review 21). In the former case, inconsistency with other chimpanzee and NHP results limited the utility of the cited chimpanzee study, while in the latter the disease and vaccination strategies explored were too disparate from those investigated in the cited chimpanzee study to accord it other than peripheral relevance.

Wang *et al.*'s study of the reactivity of humans and chimpanzees to various epitopes of HCV H strain structural proteins ([Wang *et al.*, 1996](#)) was cited by both Hüsey *et al.*'s paper investigating a diagnostic method for HCV ([Hüsey *et al.*, 1997](#)) (Review 8), and by Nakano *et al.*'s paper investigating therapeutic options for HCV ([Nakano *et al.*, 1999](#)) (Review 10). However, in both cases only the human outcomes within this chimpanzee study contributed to the citing medical paper. In fact, the cited chimpanzee study highlighted differences in the immune response of humans and chimpanzees to HCV.

Several authors of human medical papers or cited chimpanzee studies identified potential problems associated with attempts to extrapolate chimpanzee results to human outcomes. For example, in their study of prophylactic methods for combating

EBV, Khanna *et al.* (Khanna *et al.*, 1999a) stated that, “*It would be a mistake to assume that experimental results obtained in these ... primate models had direct relevance to vaccine formulations that might offer protection against infectious mononucleosis [one of the main targets for a vaccine].*” In his review of prophylactic methods for combating HAV and HBV, Koff (Koff 2002) described the cited chimpanzee study as potentially *complicating* our understanding of escape viruses. Wang *et al.* (Wang *et al.*, 1996) identified key differences between the immune response of humans and chimpanzees to HCV infection. When commenting on the failure of a test drug to demonstrate efficacy at combating (T-cell dependent) RA in a phase II clinical trial, despite prior efficacy of an analogue drug in modulating T-cell receptor responsiveness in chimpanzees, Newman *et al.* (Newman *et al.*, 2001) stated that, “*...results to date illustrate the profound difficulties in translating animal model success to the clinical arena.*” Development of this test drug for RA patients was discontinued.

Sources contributing to human medical papers

Techniques featuring most frequently in the development of the prophylactic, diagnostic or therapeutic methods described in the 27 medical papers detailed above included *in vitro* studies, human clinical and epidemiological studies, molecular assays and methods, and genomic studies. Methods featuring particularly prominently included:

- *In vitro* studies in at least 18 papers, utilizing for example: HeLa cells; human adenoid explants; human respiratory epithelium (embryonic lung fibroblasts); human T-cells; LCLs; and rodent cell lines. Several viral studies used *E. coli* and baculoviruses in conjunction with *Sf9* insect cells as vectors for viral delivery and expression.
- Human clinical and epidemiological studies in at least 15 and six papers, respectively.
- Molecular methods in at least eight studies, including: ELISA; immune electron microscopy; PCR; radioimmunoassay; Western blot; and several assays designed for the diagnosis of TSEs: a combination of competitive antibody capture and CE; CDI; SIFT; and an immuno-PCR assay.
- Chimpanzee studies were, of course, cited by all of the medical papers. Other animal models were cited more prominently in five medical papers, including: transgenic and natural mice, rats, hamsters, guinea pigs, goats, sheep, cows, mink, woodchucks and NHPs (orangutans, baboons, cynomolgus macaques and rhesus macaques). Several of these species were cited in only one paper describing diagnostic methods for combating TSEs (Brown 2005), either as sources or recipients of TSE-infected tissues.
- Genomic techniques such as differential display, suppression subtractive hybridization, representational difference analysis, serial analysis of gene expression and microarray analysis (for example of viral genomes) featured prominently in four medical papers.

A detailed examination of the level of contribution of the various animal models other than chimpanzees is beyond the remit of this paper. Citation of animal models other than the chimpanzee in this review does not necessarily reflect a tangible contribution to any study: it simply conveys its inclusion in a citing paper, the merit of which must be addressed in the manner of similar citation analyses and systematic reviews that have been conducted recently by various research groups (such as Pound *et al.* (2005), Bailey *et al.* (2005), Knight *et al.* (2006), Perel *et al.* (2006) and Hackam *et al.* (2006)). Our evaluation was confined to the involvement of chimpanzees only.

Human utility of medical papers citing chimpanzee studies

Based purely on their appearance in the bibliographies of biomedical papers, research on captive chimpanzees or chimpanzee tissues appears to have contributed towards a large range of biomedical disciplines. But citations and involvement in research programs are not a definitive indication of the necessity and value of a particular approach or specific study or dataset, for which an in-depth assessment of each citing paper and cited chimpanzee study is required. For example, papers may be cited due to their negative contribution to an area of knowledge, for comparison to other paradigms, and so on. Or, where citations of chimpanzee research arguably indicate a positive contribution to the overall outcome, alternative methods may well have been available and superior.

In our analysis, over three-quarters (21 of 27) of the medical papers did not describe prophylactic, diagnostic or therapeutic methods for combating human diseases that were sufficiently developed for routine human use and efficacious in a sizeable proportion of human patients. Most of the putative vaccination strategies described were far from completion and implementation: for example, one combination chemotherapeutic protocol for HCV did not achieve sustained viral clearance in half of chronically infected patients treated (Review 9), and when describing the development of vaccines against RSV (which have been explored using a substantial number of animal studies over many years), Kneyber *et al.* (Review 23) (Kneyber *et al.*, 2002) stated that “*it will probably be at least another 5 to 10 years before any routine vaccination against RSV becomes daily practice.*”

In at least three cases (Reviews 2, 24 and 25), drugs being tested failed to progress to the market following initial animal or human trials, indicating concerns over human safety and/or efficacy. In one case (Review 8) the sensitivity of the diagnostic assay in question was lower than that of the traditional assay, while in three other cases (Reviews 12, 19 and 27) the diagnostic assays were not ready for routine medical use.

Several reports described vaccines in successful use for some time, such as the combination HAV and HBV vaccine Twinrix (Review 4), and well-established therapeutic methods such as the use of rIFN- α for chronic HCV infection (Review 5), Nevirapine in HIV patients (Review 16), and Cyclophosphamide for the treatment of SLE (Review 26). In one case a single vaccine candidate for the prevention of HEV that had passed a phase I clinical trial was being tested in a field trial in Nepal (Review 11), though further tests were considered necessary to determine its long-term efficacy.

CONCLUSIONS

Advocates of research on captive chimpanzees claim it has been of critical importance during our struggles against major human diseases such as AIDS, hepatitis and cancer (VandeBerg and Zola 2005) – even though it continues in only a small number of nations, and indeed is banned or severely restricted in the UK, the Netherlands, Sweden, New Zealand, Austria and Japan. A ban is also being considered by the Swiss government as well as the European Union. Moreover, proponents insist that it must remain a cornerstone of future biomedical research, and that it is an indispensable facet of research that, if prohibited in the U.S., will see human beings suffer and jeopardize or nullify possible future treatments or cures for human disease.

To be made with such conviction, these claims require strong evidence. It is generally accepted that only a small fraction of scientific results, whatever the discipline and however they were achieved, ever translate to human medical benefit or could ever be remotely applicable to clinical practice. Nevertheless, that research is pursued for the sake of knowledge itself, and also partly because we do not know how applicable any area of investigation might be in the future. However, chimpanzee research is a special case as testified to by not only public and scientific opinion, but also by various policies and laws in the U.S. Research using chimpanzees has an ethical cost way above biomedical research involving bacteria and yeast, for example: passage of the CHIMP (Chimpanzee Health Improvement, Maintenance and Protection) Act in the U.S. in 2000, as well as the many countries limiting the use of great apes, acknowledges a different moral status for chimpanzees and raised the bar regarding the ethical considerations that the scientific community (as well as the general public) confers upon chimpanzees.

Both science and the public acknowledge that chimpanzees have varied and advanced cognitive and emotional abilities; they can be taught human languages such as American Sign Language and demonstrate complex nonverbal communication patterns. They are capable of reasoned thought, abstraction, generalization, symbolic representation and have a concept of self, and they exhibit a broad range of emotions and experience mental as well as physical pain. Chimpanzees (as well as other great apes) in captivity show a range of behavioral abnormalities and measurable signs of distress (psychopathology), which can result from early separation of infants from their mothers, sensory-motor deprivation or social isolation over prolonged periods of time (Brüne *et al.*, 2006).

Quite aside from these characteristics of chimpanzees that arguably make them worthy of a human level of moral consideration, the question remains: Has chimpanzee research made a positive contribution to human health to scientifically justify their continued use in spite of the ethical objections of many? Or has that contribution been limited or even negative? And, overall, has it expedited medical progress or hindered it? If the net contribution of chimpanzee experiments to human medicine has been negligible, or negative, or if there are current and nascent research avenues that are more promising and relevant, then there is a *human*-based ethical argument for it to end. For if species differences have led to confounding data, blind alleys and false dawns for new human therapies, then humans too have suffered as a

result of misleading data from experiments involving chimpanzees – both directly and indirectly.

Proponents of chimpanzee research, however, claim that it is precisely the differences that can make chimpanzees so useful. If biochemical differences can be identified and used to elucidate *why*, for example, chimpanzees respond differently to HIV, hepatitis viruses, the malaria parasite and so on, they argue that these can be used as a foundation on which to build future research and target new drugs. There are two responses: one is that the analysis of such differences does not have to involve invasive research on captive chimpanzees, but can instead use biological samples taken from chimpanzees at sanctuaries, during routine and necessary operative procedures, physical examinations and/or post mortems. The second is that science has the tools to elucidate the molecular differences between *humans* that vary in their responses to infectious agents and diseases. Further, by minimizing extraneous differences and maintaining such investigations in a human context, they promise to deliver tangible results more precisely, accurately quickly and effectively.

Regardless of the findings of analyses such as this study, proponents of chimpanzee research may point to their involvement in, for example, hepatitis B and C research in a bid to demonstrate that not all chimpanzee experimentation has been of little or no use. The argument of the *necessity* of their involvement in hepatitis research and the development of vaccines is a fierce one with strong claims on both sides of the debate, but is not relevant to this review. No matter the veracity of claims on either side; this review seeks to examine the value of chimpanzee experimentation over *recent* years in order to assess its utility prospectively, and determine if there is a scientifically valid reason to support its continuation.

For both ethical and scientific reasons, there is widespread unease about captive-chimpanzee research within the scientific and wider communities. The fact that several countries highly active in sophisticated biomedical research have banned or severely limited research and testing using great apes testifies to this: most recently, in the UK, an independent inspector stated that a new primate research centre in Cambridge should not be built, on the basis that the proposed research to take place there could not be demonstrated to be in the national interest. In the Netherlands, the last research establishment in Europe housing chimpanzees was closed, after the Dutch minister for science announced that it was not scientifically necessary.

In summary, our review identified no studies of captive chimpanzees that made an essential contribution, or, in a large majority of cases, a significant contribution of any kind towards papers describing well-developed prophylactic, diagnostic or therapeutic methods for combating human diseases, including major diseases such as AIDS, cancer and hepatitis. At best, it has on occasion made a minor contribution to a small number of research projects, though an assessment of the crucial nature of this input and possible alternatives to the use of chimpanzees in each instance was not in the scope of our analysis. Additionally, it seems that data from chimpanzee experiments correlate with the human situation in only a small minority of cases. It therefore follows that reliable extrapolation of such data to humans is impossible, even potentially hazardous; statistically, it cannot be considered *predictive* of the human

condition or response. Retrospective correlations are useless, once the human data are known, and “confirmatory” studies in chimpanzees must be deemed similarly futile.

Far from augmenting biomedical research, chimpanzee experimentation appears to have been largely incidental, peripheral, confounding, irrelevant, unreliable and has consumed considerable research funding that would have been better targeted elsewhere. We call upon regulatory and funding bodies to take these data into account when considering the future of chimpanzees in research and testing, and likewise the U.S. government when considering the permanent retirement of chimpanzees currently held in U.S. laboratories and the status of the chimpanzee breeding moratorium.

ACKNOWLEDGEMENTS

This research was funded by NEAVS/Project R&R: Release and Restitution for Chimpanzees in U.S. Laboratories, 333 Washington Street, Suite 850, Boston, MA 02108, U.S. For their various contributions, we thank Theodora Capaldo Ed.D., Lawrence D'Antonio Ph.D., Debra Durham Ph.D., Andrew Knight, B.V.M.S., and Amanda Greenall Ph.D.

REFERENCES

Aberle, J. H., Formann, E., Steindl-Munda, P., Weseslindtner, L., Gurguta, C. (2006). Prospective study of viral clearance and CD4(+) T-cell response in acute hepatitis C primary infection and reinfection. *Journal of Clinical Virology* 36(1): 24-31.

Abraham, G. and Colonno, R. J. (1984). Many rhinovirus serotypes share the same cellular receptor. *Journal of Virology* 51: 340-345.

Aggarwal, R., Shahi, H., Naik, S., Yachha, S. K., Naik, S. R. (1997). Evidence in favour of high infection rate with hepatitis E virus among young children in India. *Journal of Hepatology* 26: 1425-1426.

Anderson, D., Chambers, K., Hanna, N., Leonard, J., Reff, M. *et al.* (1997). A primatized MAb to human CD4 causes receptor modulation, without marked reduction in CD4+ T cells in chimpanzees: in vitro and in vivo characterization of a MAb (IDEC-CE9.1) to human CD4. *Clinical Immunology and Immunopathology* 84(1): 73-84.

Anderson, D. A., Li, F., Riddell, M., Howard, T., Seow, H. F. *et al.* (1999). ELISA for IgG-class antibody to hepatitis virus based on a highly conserved, conformational epitope expressed in Escherichia coli. *Journal of Virological Methods* 81: 131-142.

Anonymous (2006a). CAB Abstracts. Available: <http://www.cabi-publishing.org/pdf/CABAAbstracts/CABAbs.pdf>. Accessed 2006 Aug 11.

Anonymous (2006b). Database coverage. Available: <http://www.cabi-publishing.org/AbstractDatabases.asp?SubjectArea=&Subject=&Section=dc&PID=125>. Accessed 2006 Aug 11.

Anonymous (2006c). About EMBASE: the Excerpta Medica database. Available: http://info.embase.com/embase_suite/index.shtml. Accessed 2006 Aug 11.

Anonymous (2006d). PubMed overview. Available: <http://www.ncbi.nlm.nih.gov/entrez/query/static/overview.html>. Accessed 2006 Aug 11.

Anonymous (1998e). Viramune(R) (nevirapine) tablets and oral suspension. Prescribing information. Columbus, Ohio: Roxane Labs Inc.

Arankalle, V. A., Chadha, M. S., Chobe, L. P., Nair, R., Banerjee, K. (1995). Cross-challenge studies in rhesus monkeys employing different Indian isolates of hepatitis E virus. *Journal of Medical Virology* 46: 358-363.

Armbruster, C., Stiegler, G. M., Vcelar, B. A., Jager, W., Michael, N. L. *et al.* (2002). A phase I trial with two human monoclonal antibodies (hMAb 2F5, 2G12) against HIV-1. *AIDS* 16(2): 227-233.

- Armbruster, C., Stiegler, G. M., Vcelar, B. A., Jager, W., Koller, U. *et al.* (2004). Passive immunization with the anti-HIV-1 human monoclonal antibody (hMAb) 4E10 and the hMAb combination 4E10/2F5/2G12. *Journal of Antimicrobial Chemotherapy* 54(5): 915-920.
- Bailey, J., Knight, A., Balcombe, J. (2005). The future of teratology is *in vitro*. *Biogenic Amines* 19(2): 97-145.
- Balayan, M. S., Andjaparidze, A. G., Savinskaya, S. S., Ketiladze, E. S., Braginsky, D. M. *et al.* (1983). Evidence for a virus in non-A, non-B hepatitis transmitted via the fecal-oral route. *Intervirology* 20: 23-31.
- Balzarini, J., Karlsson, A., Perez-Perez, M. J., Camarasa, M. J., De Clercq, E. (1993). Knocking-out concentrations of HIV-I-specific inhibitors completely suppress HIV-I infection and prevent the emergence of drug-resistant virus. *Virology* 196: 576-585.
- Bardsley-Elliot, A. and Perry, C. M. (2000). Nevirapine: A Review of its Use in the Prevention and Treatment of Paediatric HIV Infection. *Pediatric Drugs* 2(5): 373-407.
- Barletta, J. M., Edelman, D. C., Highsmith, W. E., Constantine, N. T. (2005). Detection of ultra-low levels of pathologic prion protein in scrapie infected hamster brain homogenates using real-time immuno-PCR. *Journal of Virological Methods* 127: 154-164.
- Bertoni, R., Sette, A., Sidney, J., Guidotti, L. G., Shapiro, M. *et al.* (1998). Human class I supertypes and CTL repertoires extend to chimpanzees. *Journal of Immunology* 161(8): 4447-4455.
- Bieschke, J., Giese, A., Schulz-Schaeffer, W., Zerr, I., Poser, S. *et al.* (2000). Ultrasensitive detection of pathological prion protein aggregates by dual-color scanning for intensely fluorescent targets. *Proceedings of the National Academy of Sciences of the USA* 97: 5468-5473.
- Bigger, C. B., Brasky, K. M., Lanford, R. E. (2001). DNA microarray analysis of chimpanzee liver during acute resolving hepatitis C virus infection. *Journal of Virology* 75(15): 7059-7066.
- Bigger, C. B., Guerra, B., Brasky, K. M., Hubbard, G., Beard, M. R. *et al.* (2004). Intrahepatic gene expression during chronic hepatitis C virus infection in chimpanzees. *Journal of Virology* 78: 13779-13792.
- Bogedain, C., Wolf, H., Modrow, S., Stuber, G., Jilg, W. (1995). Specific cytotoxic T lymphocytes recognize the immediate-early transactivator Zta of Epstein-Barr virus. *Journal of Virology* 69(8): 4872-4879.
- Boumpas, D. T., Furie, R., Manzi, S., Illei, G. G., Wallace, D. J. *et al.* (2003). A short course of BG9588 (anti-CD40 ligand antibody) improves serologic activity and

decreases hematuria in patients with proliferative lupus glomerulonephritis. *Arthritis and Rheumatism* 48(3): 719-27.

Bradley, D. W. (1990). Enterically-transmitted non-A, non-B hepatitis. *British Medical Bulletin* 46: 442-461.

Bradley, D. W. (1992). Hepatitis E epidemiology, aetiology and molecular biology. *Reviews in Medical Virology* 2: 19-28.

Brams, P., Black, A., Padlan, E. A., Hariharan, K., Leonard, J. *et al.* (2001). A humanized anti-human CD154 monoclonal antibody blocks CD154-CD40 mediated human B cell activation. *International Immunopharmacology* 1(2):277-294.

Brind, A., Jiang, J., Samuel, D., Gigou, M., Feray, C. *et al.* (1997). Evidence for selection of hepatitis B mutants after liver transplantation through peripheral blood mononuclear cell infection. *Journal of Hepatology* 26: 228-235.

Britten, R. J. (2002). Divergence between samples of chimpanzee and human DNA sequences is 5%, counting indels. *Proceedings of the National Academy of Sciences of the USA* 99(21): 13633-13635.

Brown, P., Preece, M., Brandel, J-P., Sato, T., McShane, L. *et al.* (2000). Iatrogenic Creutzfeldt-Jakob disease at the Millennium. *Neurology* 55: 1075-1081.

Brown, P. (2005). Blood infectivity, processing and screening tests in transmissible spongiform encephalopathy. *Vox Sanguinis* 89(2): 63-70.

Brüne, M., Brüne-Cohrs, U., McGrew, W. C., Preuschoft, S. (2006). Psychopathology in great apes: Concepts, treatment options and possible homologies to human psychiatric disorders. *Neuroscience and behavioral reviews* doi:10.1016/j.neubiorev.2006.09.002.

Bryan, J. P., Tsarev, S. A., Iqbal, M., Ticehurst, J., Emerson, S. *et al.* (1994). Epidemic hepatitis E in Pakistan: patterns of serologic response and evidence that antibody to hepatitis E virus protects against disease. *Journal of Infectious Diseases* 170(3): 517-21.

Carlsson, H. E., Schapiro, S. J., Farah, I., Hau, J. (2004). Use of primates in research: a global overview. *American Journal of Primatology* 63(4): 225-237.

Carman, W. F., Zanetti, A. R., Karayiannis, P., Waters, J., Manzillo, G. *et al.* (1990). Vaccine-induced escape mutant of hepatitis B virus. *Lancet* 336: 325-329.

Carr, A., Vella, S., de Jong, M. D., Sorice, F., Imrie, A. *et al.* (1996). A controlled trial of nevirapine plus zidovudine versus zidovudine alone in p24 antigenaemic HIV-infected patients. The Dutch-Italian-Australian Nevirapine Study Group. *AIDS* 10(6): 635-641.

Cervenakova, L., Brown, P., Soukharev, S., Yakovleva, O., Diringer, H. *et al.* (2003). Failure of immunocompetitive capillary electrophoresis assay to detect disease-specific prion protein in buffy coat from humans and chimpanzees with Creutzfeldt-Jakob disease. *Electrophoresis* 24(5): 853-859.

Chauhan, A., Dilawari, J. B., Sharma, R., Mukesh, M., Saroa, S. R. (1998). Role of long-persisting human hepatitis E virus antibodies in protection. *Vaccine* 16: 755-756.

Cheeseman, S. H., Havlir, D., McLaughlin, M. M., Greenough, T. C., Sullivan, J. L. *et al.* (1995). Phase I/II evaluation of nevirapine alone and in combination with zidovudine for infection with human immunodeficiency virus. *Journal of Acquired Immune Deficiency Syndromes and Human Retrovirology* 8(2): 141-151.

Chen, L., Borozan, I., Feld, J., Sun, J., Tannis, L. L., Coltescu, C., Heathcote, J., Edwards, A. M., McGilvray, I. D. (2005). Hepatic gene expression discriminates responders and nonresponders in treatment of chronic hepatitis C viral infection. *Gastroenterology* 128: 1437-1444.

Cohen, C. J., Xiang, Q., Gao, G. P., Ertl, H. C., Wilson, J. M. *et al.* (2002). Chimpanzee adenovirus CV-68 adapted as a gene delivery vector interacts with the coxsackievirus and adenovirus receptor. *Journal of General Virology* 83: 151-155.

Conlee, K. M., Hoffeld, E. H., Stephens, M. L. (2004). A demographic analysis of primate research in the United States. *ATLA (Alternatives to Laboratory Animals)* 32 (Supp 1A): 315-322.

Conley, A. J., Kessler, J. A. II., Boots, L. J., McKenna, P. M., Schleif, W. A. *et al.* (1996). The consequence of passive administration of an anti-human immunodeficiency virus type 1 neutralizing monoclonal antibody before challenge of chimpanzees with a primary virus isolate. *Journal of Virology* 70(10): 6751-6758.

Constantine, N. (2006). HIV Antibody Assays. HIV InSite Knowledge Base Chapter. Available: <http://hivinsite.ucsf.edu/InSite?page=kb-02-02-01>. Accessed 2006 Dec 12.

Cosimi, A. (2002). Calcineurin inhibitors are not antagonistic to tolerance induction. *Transplantation Proceedings* 34: 1376-1377.

Crowe, J. E. Jr., Randolph, V., Murphy, B. R. (1999). The live attenuated subgroup B respiratory syncytial virus vaccine candidate RSV 2B33F is attenuated and immunogenic in chimpanzees, but exhibits partial loss of the ts phenotype following replication in vivo. *Virus Research* 59(1): 13-22.

Crump, C. E., Arruda, E., Hayden, F. G. (1993). In vitro inhibitory activity of soluble ICAM-1 for the numbered serotypes of human rhinovirus. *Antiviral Chemistry and Chemotherapy* 4: 323-327.

Cullen, B. R. (1998). HIV-1 auxiliary proteins: making connections in a dying cell. *Cell* 93: 685-692.

Czeschinski, P. A., Binding, N., Witting, U. (2000). Hepatitis A and hepatitis B vaccinations: immunogenicity of combined vaccine and of simultaneously or separately applied single vaccines. *Vaccine 18*: 1074-1080.

Daikh, D. I. and Wofsy, D. (2001). Cutting edge: reversal of murine lupus nephritis with CTLA4 Ig and cyclophosphamide. *Journal of Immunology 166*: 2913-2916.

D'Aquila, R. T., Hughes, M. D., Johnson, V. A., Fischl, M. A., Sommadossi, J. P. *et al.* (1996). Nevirapine, zidovudine, and didanosine compared with zidovudine and didanosine in patients with HIV-1 infection. A randomized, double-blind, placebo-controlled trial. National Institute of Allergy and Infectious Diseases AIDS Clinical Trials Group Protocol 241 Investigators. *Annals of Internal Medicine 124*(12): 1019-1030.

Dawson, G. J., Chau, K. H., Cabal, C. M., Yarbough, P. O., Reyes, G. R. *et al.* (1992). Solid-phase enzyme-linked immunosorbent assay for hepatitis E virus IgG and IgM antibodies utilizing recombinant antigens and synthetic peptides. *Journal of Virological Methods 38*: 175-186.

de Bethune, M. P., Andries, K., Azijn, H., *et al.* (1998). Binding HIV-1 non-nucleoside reverse transcriptase inhibitors to human serum albumin and human reid acid glycoprotein: conflicting results depending on the assay used. [abstract no. 47]. *Antiviral Research 37*: A54.

Delic, D., Nestic, Z., Prostran, M., Simonovic, J., Svirtlih, N. (2005). Treatment of anicteric acute hepatitis C with peginterferon alpha-2a plus ribavirin. *Vojnosanit Pregl 62*(11): 865-868.

Dickson, R. C. (1998). Management of posttransplantation viral hepatitis-hepatitis B. *Liver Transplant Surgery 4*(Suppl 1): S73-S78.

Donohue, J. G., Munoz, A., Ness, P. M., Brown, D. J., Yawn, D. H. (1992). The declining risk of post-transfusion hepatitis C virus infection. *New England Journal of Medicine 327*: 369-373.

el-Amada, Z., Murthy, K. K., Higgins, K., Cobb, E. K., Haigwood, N. L. *et al.* (1995). Resistance of chimpanzees immunized with recombinant gp120SF2 to challenge by HIV-1SF2. *AIDS 9*(12): 1313-1322.

Elliott, S. L., Pye, S. J., Schmidt, C., Cross, S. M., Silins, S. L. *et al.* (1997). Dominant cytotoxic T lymphocyte response to the immediate-early trans-activator protein, BZLF1, in persistent type A or B Epstein-Barr virus infection. *Journal of Infectious Diseases 176*(4): 1068-1072.

Emerman, M. and Malim, M. H. (1998). HIV-1 regulatory accessory genes: keys to unraveling viral and host cell biology. *Science 280*: 1880-1884.

Emini, E. A., Schleif, W. A., Nunberg, J. H., Conley, A. J., Eda, Y. *et al.* (1992). Prevention of HIV-1 infection in chimpanzees by gp120 V3 domain-specific monoclonal antibody. *Nature* 355(6362): 728-730.

Enomoto, N., Sakuma, I., Asahina, Y., Kurosaki, M., Murakami, T. *et al.* (1995). Comparison of full-length sequences of interferon-sensitive and resistant hepatitis C virus 1b. Sensitivity to interferon is conferred by amino acid substitutions in the NS5A region. *Journal of Clinical Investigation* 96(1): 224-230.

Enomoto, N., Sakuma, I., Asahina, Y., Kurosaki, M., Murakami, T. *et al.* (1996). Mutations in the nonstructural protein 5A gene and response to interferon in patients with chronic hepatitis C virus 1b infection. *New England Journal of Medicine* 334: 77-81.

Evans, H. L. (1990). Nonhuman primates in behavioral toxicology: issues of validity, ethics and public health. *Neurotoxicology and Teratology* 12(5): 531-536.

Farina, S. F., Gao, G. P., Xiang, Z. Q., Rux, J. J., Burnett, R. M. *et al.* (2001). Replication-defective vector based on a chimpanzee adenovirus. *Journal of Virology* 75: 11603-11613.

Favorov, M. O., Khudyakov, Y. E., Mast, E. E., Yashina, T. L., Shapiro, C. N. *et al.* (1996). Ig M and IgG antibodies to hepatitis E virus (HEV) detected by enzyme immunoassay based on an HEV specific artificial recombinant mosaic protein. *Journal of Medical Virology* 50: 50-58.

Feld, J. J., and Hoofnagle, J. H. (2005). Mechanism of action of interferon and ribavirin in treatment of hepatitis C. *Nature* 436(7053): 967-972.

Ferguson, M., Walker, D., Mast, E., Fields, H. (2002). Report of a collaborative study to assess the suitability of a reference agent for antibodies to hepatitis E virus. *Biologicals* 30: 43-48.

Ferreira, J. V., Froud, T., Caulfield, A. *et al.* (2002). Can enriched bone marrow infusion induce donor tolerance in solitary islet cell transplantation? [abstract 290] *Transplantation* 74.

Fields, H. A., Khudyakov, Y. E., Favorov, M. O., Khudyakova, N. S., Cong, M. E. *et al.* (1996). Artificial mosaic proteins as new immunodiagnostic reagents: The hepatitis E experience. *Clinical and Diagnostic Virology* 5: 167-179.

Fortuin, M., Karthigesu, V., Allison, L., Howard, C., Hoare, S. *et al.* (1994). Breakthrough infections and identification of a viral variant in Gambian children immunized with hepatitis B vaccine. *Journal of Infectious Diseases* 169: 1374-1376.

Frey, S., Dagan, R., Ashur, Y., Chan, X. Q., Ibarra, J. *et al.* (1999). Interference of antibody production to hepatitis B surface antigen in a combination hepatitis A/hepatitis B vaccine. *Journal of Infectious Diseases* 180: 2018-2022.

Gallo, R. C. (2002). Human retroviruses after 20 years: a perspective from the past and prospects for their future control. *Immunological Reviews* 185: 236-265.

Gao, F., Bailes, E., Robertson, D. L., Chen, Y., Rodenburg, C. M. *et al.* (1999). Origin of HIV-1 in the chimpanzee *Pan troglodytes troglodytes*. *Nature* 397: 436-441.

Gauduin, M. C., Safrit, J. T., Weir, R., Fung, M. S., Koup, R. A. (1995). Pre- and postexposure protection against human immunodeficiency virus type 1 infection mediated by a monoclonal antibody. *Journal of Infectious Diseases* 171(5): 1203-1209.

Genmab A/S (2002). Humax-CD4 in combination therapy not effective in rheumatoid arthritis. 24 Sep. 2002. Copenhagen, Denmark. Available: http://www.genmab.com/html/2002_09_24.shtml. Accessed 2006 Mar 21.

Gern, J. E. and Busse, W. W. (1999). Association of rhinovirus infections with asthma. *Clinical Microbiology Reviews* 12: 9-18.

Gescuk, B. D. and Davis, J. C. (2002). Novel therapeutic agents for systemic lupus erythematosus. *Current Opinion in Rheumatology* 14(5): 515-521.

Ghany, M. G., Ayola, B., Villamil, F. G., Gish, R. G., Rojter, S. *et al.* (1998). Hepatitis B virus S mutants in liver transplant recipients who were reinfected despite hepatitis B immune globulin prophylaxis. *Hepatology* 27: 213-222.

Goh, W. C., Rogel, M. E., Kinsey, C. M., Michael, S. F., Fultz, P. N. (1998). HIV-1 Vpr increases viral expression by manipulation of the cell cycle: a mechanism for selection of Vpr in vivo. *Nature Medicine* 4(1): 65-71.

Goldsmith, R., Yarbough, P. O., Reyes, G. R., Fry, K. E., Gabor, K. A. *et al.* (1992). Enzyme-linked immunosorbent assay for diagnosis of acute sporadic hepatitis E in Egyptian children. *Lancet* 339: 328-331.

Good, M. F. and Doolan, D. L. (1999). Immune effector mechanisms in malaria. *Current Opinion in Immunology* 11: 412-419.

Gramzinski, R. A., Millan, C. L., Obaldia, N., Hoffman, S. L., Davis, H. L. (1998). Immune response to a hepatitis B DNA vaccine in Aotus monkeys: a comparison of vaccine formulation, route, and method of administration. *Molecular Medicine* 4: 109-118.

Grob, P. M., Cao, Y., Muchmore, E., Ho, D. D., Norris, S. *et al.* (1997). Prophylaxis against HIV-1 infection in chimpanzees by nevirapine, a nonnucleoside inhibitor of reverse transcriptase. *Nature Medicine* 3(6): 665-670.

Grunberg, K., Timmers, M. C., Smits, H. H., de Klerk, E. P., Dick, E. C. *et al.* (1997). Effect of experimental rhinovirus 16 colds on airway hyperresponsiveness to histamine and interleukin-8 in nasal lavage in asthmatic subjects in vivo. *Clinical and Experimental Allergy* 27: 36-45.

Hackam, D. G. and Redelmeier, D. A. (2006). Translation of research evidence from animals to humans. *Journal of the American Medical Association (JAMA)* 296(14): 1731-1732.

Hafler, D. A., Ritz, J., Schlossman, S. F., Weiner, H. L. (1988). Anti-CD4 and anti-CD2 monoclonal antibody infusions in subjects with multiple sclerosis. Immunosuppressive effects and human anti-mouse responses. *Journal of Immunology* 141: 131-138.

Halsey, N. A., Markham, R., Wahren, B., Boulos, R., Rossi, P. *et al.* (1992). Lack of association between maternal antibodies to V3 loop peptides and maternal-infant HIV-1 transmission. *Journal of Acquired Immune Deficiency Syndrome* 5(2): 153-157.

Hasan, U. A., Abai, A. M., Harper, D. R., Wren, B. W., Morrow, W. J. (1999). Nucleic acid immunization: concepts and techniques associated with third generation vaccines. *Journal of Immunological Methods* 229: 1-22.

Havlir, D., McLaughlin, M. M., Richman, D. D. (1995a). A pilot study to evaluate the development of resistance to nevirapine in asymptomatic human immunodeficiency virus-infected patients with CD4 cell counts of > 500/mm³: AIDS Clinical Trials Group Protocol 208. *Journal of Infectious Diseases* 172(5): 1379-1383.

Havlir, D., Cheeseman, S. H., McLaughlin, M., Murphy, R., Erice, A. *et al.* (1995b). High-dose nevirapine: safety, pharmacokinetics, and antiviral effect in patients with human immunodeficiency virus infection. *Journal of Infectious Diseases* 171(3): 537-545.

Hayden, F. G., Gwaltney, J. M. Jr., Colonno, R. J. (1988). Modification of experimental rhinovirus colds by receptor blockade. *Antiviral Research* 9: 233-247.

Heijntink, R. A., Kruining, J., Honkoop, P., Kuhns, M. C., Hop, W. C. J. *et al.* (1997). Serum HBeAg quantitation during antiviral therapy for chronic hepatitis B. *Journal of Medical Virology* 53: 282-287.

Hepburn, T. W., Totoritis, M. C., Davis, C. B. (2003). Antibody-mediated stripping of CD4 from lymphocyte cell surface in patients with rheumatoid arthritis. *Rheumatology* 42(1): 54-61.

Herodin, F., Thullier, P., Garin, D., Drouet, M. (2005). Nonhuman primates are relevant models for research in hematology, immunology and virology. *European Cytokine Network* 16(2): 104-116.

Herzog, C., Walker, C., Pichler, W., Aeschlimann, A., Wassmer, P. (1987). Monoclonal anti-CD4 in arthritis. *Lancet* 2: 1461-1462.

Herzog, C., Walker, C., Muller, W., Rieber, P., Reiter, C. (1989). Anti-CD4 antibody treatment of patients with rheumatoid arthritis: I. Effect on clinical course and circulating T cells. *Journal of Autoimmunity* 2: 627-642.

Hewlett, I. K., Geyer, S. J., Hawthorn, C. A., Ruta, M., Epstein, J. S. (1991). Kinetics of early HIV-1 gene expression in infected H9 cells assessed by PCR. *Oncogene* 6: 491-493.

Higgins, J. P. T., Green, S., Eds. (2005). Cochrane Handbook for Systematic Reviews of Interventions 4.2.5. Available: <http://www.cochrane.org/resources/handbook/hbook.htm>. Accessed 2006 May 27.

Hill, A. V., Allsopp, C. E., Kwiatkowski, D., Anstey, N. M., Twumasi, P. (1991). Common west African HLA antigens are associated with protection from severe malaria. *Nature* 352: 595-600.

Hino, K., Katoh, Y., Vardas, E., Schoub, B., Carman, W. F. et al. (1999). The effect of introduction of universal childhood vaccination on the prevalence of serologically negative hepatitis B virus infection and the selection of immune escape variants in a developing country [Abstract]. *Hepatology* 30: 300A.

Hoffman, S. L., Goh, L. M., Luke, T. C., Schneider, I., Le, T. P. et al. (2002). Protection of humans against malaria by immunization with radiation-attenuated *Plasmodium falciparum* sporozoites. *Journal of Infectious Diseases* 185:1155.

Honda, M., Kaneko, S., Kawai, H., Shirota, Y., Kobayashi, K. (2001). Differential gene expression between chronic hepatitis B and C hepatic lesion. *Gastroenterology* 120: 955-966.

Hone, D. M., DeVico, A. L., Fouts, T. R., Onyabe, D. Y., Agwale, S. M. et al. (2002). Development of vaccination strategies that elicit broadly neutralizing antibodies against human immunodeficiency virus type 1 in both the mucosal and systemic immune compartments. *Journal of Human Virology* 5(1): 17-23.

Horneff, G., Burmester, G. R., Emmrich, F., Kalden, J. R. (1991). Treatment of rheumatoid arthritis with an anti-CD4 monoclonal antibody. *Arthritis and Rheumatism* 34: 129-140.

Horneff, G., Dirksen, U., Schulze-Koops, H., Emmrich, F., Wahn, V. (1995). Treatment of refractory juvenile chronic arthritis by monoclonal CD4 antibodies: A pilot study in two children. *Annals of the Rheumatic Diseases* 54: 846-9.

Hsu, H. H., Donets, M., Greenberg, H. B., Feinstone, S. M. (1993). Characterization of hepatitis C virus structural proteins with a recombinant baculovirus expression system. *Hepatology* 17: 763-771.

Hsu, H. Y., Chang, M. H., Ni, Y. H., Lin, H. H., Wang, S. M. et al. (1997). Surface gene mutants of hepatitis B virus in infants who develop acute or chronic infections despite immunoprophylaxis. *Hepatology* 26: 786-791.

Huguenel, E. D., Cohn, D., Dockum, D. P., Greve, J. M., Fournel, M. A. et al. (1997). Prevention of rhinovirus infection in chimpanzees by soluble intercellular adhesion molecule-1. *American Journal of Respiratory and Critical Care Medicine* 155(4): 1206-1210.

Hunter, N., Foster, J., Chong, A., McCutcheon, S., Parnham, D. et al. (2002). Transmission of prion diseases by blood transfusion. *Journal of General Virology* 83: 2897-2905.

Hüssy, P., Schmid, G., Mous, J., Jacobsen, H. (1996). Purification and in vitro-phospholabeling of secretory envelope proteins E1 and E2 of hepatitis C virus expressed in insect cells. *Virus Research* 45: 45-57.

Hüssy, P., Faust, H., Wagner, J. C., Schmid, G., Mous, J. et al. (1997). Evaluation of hepatitis C virus envelope proteins expressed in E. coli and insect cells for use as tools for antibody screening. *Journal of Hepatology* 26: 1179-1186.

Im, S. W., Zhang, J. Z., Zhuang, H., Che, X. Y., Zhu, W. F. et al. (2001). A bacterially expressed peptide prevents experimental infection of primates by the hepatitis E virus. *Vaccine* 19: 3726-3732.

Ji, X., Cheung, R., Cooper, S., Li, Q., Greenberg, H. B., He, X. S. (2003). Interferon for chronic hepatitis C. *Hepatology* 37: 610–621.

Jorgensen, C., Mason, U., Baton, F., Elliott, M., Jackson, M. et al. (1998). Eleven month clinical safety follow-up in RA patients with CD4 lymphopenia following treatment with the PRIMATIZED anti-CD4 monoclonal antibody Keliximab. In: American College of Rheumatologists Meeting Vol. 41. *Arthritis & Rheumatism*. San Diego, CA. p S56.

Jothikumar, N., Aparna, K., Kamatchiammal, S., Paulmurugan, R., Saravanadevi, S. et al. (1993). Detection of hepatitis E virus in raw and treated wastewater with a polymerase chain reaction. *Applied and Environmental Microbiology* 59: 2558-2562.

Jowett, J. B., Planelles, V., Poon, B., Shah, N. P., Chen, M. L. et al. (1995). The human immunodeficiency virus type 1 vpr gene arrests infected T cells in the G2 π M phase of the cell cycle. *Journal of Virology* 69: 6304–6313.

Kallinowski, B., Knoll, A., Lindner, E., Snager, R., Stremmel, W. et al. (2000). Can monovalent hepatitis A and B vaccines be replaced by a combined hepatitis A/B vaccine during the primary immunization course? *Vaccine* 19: 16-22.

Kamal, S. M., Fouly, A. E., Kamel, R. R., Hockenjos, B., Al Tawil, A. (2006). Peginterferon alfa-2b therapy in acute hepatitis C: impact of onset of therapy on sustained virologic response. *Gastroenterology* 130(3): 632-638.

Kamili, S., Spelbring, J., Krawczynski, K. (2002). DNA vaccination against hepatitis E virus infection in cynomolgus macaques. *Journal of Gastroenterology and Hepatology* 17 Suppl 3: 365-369.

Kamili, S., Spelbring, J., Carson, D., Krawczynski, K. (2004). Protective efficacy of hepatitis E virus DNA vaccine administered by gene gun in the cynomolgus macaque model of infection. *Journal of Infectious Diseases* 189: 258-264.

Karayiannis, P. (2003). Hepatitis B virus: old, new and future approaches to antiviral treatment. *Journal of Antimicrobial Therapy* 51: 761-785.

Karthigesu, V. D., Allison, L. M. C., Fortuin, M., Mendy, M., Whittle, H. C. et al. (1994). A novel hepatitis B virus variant in the sera of immunized children. *Journal of General Virology* 75: 443-448.

Kato, J., Hasegawa, K., Torii, N., Yamauchi, K., Hayashi, N. (1996). A molecular analysis of viral persistence in surface antigen-negative chronic hepatitis B. *Hepatology* 23: 389-395.

Kawai, T., Andrews, D., Colvin, R. B., Sachs, D. H., Cosimi, A. B. (2000). Thromboembolic complications after treatment with monoclonal antibody against CD40 ligand. *Nature Medicine* 6: 114.

Khair, O. A., Devalia, J. L., Abdelaziz, M. M., Sapsford, R. J., Davies, R. J. (1995). Effect of erythromycin on Haemophilus influenzae endotoxin-induced release of IL-6, IL-8, and sICAM-1 by cultured human bronchial epithelial cells. *European Respiratory Journal* 8: 1451-1457.

Khanna, R., Burrows, S. R., Kurilla, M. G., Jacob, C. A., Misko, I. S. et al. (1992). Localisation of Epstein-Barr virus cytotoxic T-cell epitopes using recombinant vaccinia: implications for vaccine development. *Journal of Experimental Medicine* 176: 169-176.

Khanna, R., Moss, D., Burrows, S. R. (1999a). Vaccine strategies against Epstein-Barr virus-associated diseases: lessons from studies on cytotoxic T-cell-mediated immune regulation. *Immunological Reviews* 170: 49-64.

Khanna, R., Sherritt, M., Burrows, S. R. (1999b). EBV structural antigens, gp350 and gp85, as targets for ex vivo virus-specific CTL during acute infectious mononucleosis: potential use of gp350/gp85 CTL epitopes for vaccine design. *Journal of Immunology* 162(5): 3063-3069.

Khorsi, H., Castelain, S., Wyseur, A., Izopet, J., Canva, V. (1997). Mutations of hepatitis C virus 1b NS5A 2209-2248 amino acid sequence do not predict the response to recombinant interferon-alfa therapy in French patients. *Journal of Hepatology* 27: 72-77.

Khudyakov, Y. E., Khudyakova, N. S., Fields, H. A., Jue, D., Starling, C. et al. (1993). Epitope mapping in proteins of hepatitis E virus. *Virology* 194: 89-96.

Khudyakov, Yu. E., Favorov, M. O., Jue, D. L., Hine, T. K., Fields, H. A. (1994). Immunodominant antigenic regions in a structural protein of the hepatitis E virus. *Virology* 198: 390-393.

Khuroo, M. S., Kamili, S., Dar, M. Y., Moecklii, R., Jameel, S. (1993). Hepatitis E and long-term antibody status. *Lancet* 341: 1355.

Kim, J. W. and Wang, X. W. (2003). Gene expression profiling of preneoplastic liver disease and liver cancer: a new era for improved early detection and treatment of these deadly diseases? *Carcinogenesis* 24(3): 363-369.

Kleinman, S., Alter, H., Busch, M., Holland, P., Tegtmeier, G. (1992). Increased detection of hepatitis C virus (HCV)-infected blood donors by a multiple-antigen HCV enzyme immunoassay. *Transfusion* 32: 805-813.

Klöhn, P. C., Stolze, L., Enari, M., Weissmann, C. (2003). A quantitative, highly sensitive cell-based infectivity assay for mouse scrapie prions. *Proceedings of the National Academy of Sciences of the USA* 100: 11666-11671.

Kneyber, M. C and Kimpfen, J. L. (2002). Current concepts on active immunization against respiratory syncytial virus for infants and young children. *Pediatric Infectious Disease Journal* 21(7): 685-696.

Knight, A., Bailey, J., Balcombe, J. (2006). Animal Carcinogenicity Studies: 1. Poor Human Predictivity. *ATLA (Alternatives to Laboratory Animals)* 34: 19-27.

Knoll, A., Hottentrager, B., Kainz, J., Bretschneider, B., Jilg, W. (2000). Immunogenicity of a combined hepatitis A and B vaccine in healthy young adults. *Vaccine* 18: 2029-2032.

Koff, R. S. (2002). Hepatitis A, hepatitis B, and combination hepatitis vaccines for immunoprophylaxis: an update. *Digestive Diseases and Sciences* 47(6): 1183-1194.

Koonin, E. V., Gorbalenya, A. E., Purdy, M. A., Rozanov, M. N., Reyes, G. R. et al. (1992). Computer assisted assignment of functional domains in the nonstructural polyprotein of hepatitis E virus: delineation of an additional group of positive strand RNA plant and animal viruses. *Proceedings of the National Academy of Sciences of the USA* 9: 8259-8263.

Korber, B., Theiler, J., Wolinsky, S. (1998). Limitations of a molecular clock applied to considerations of the origin of HIV-1. *Science* 280: 1868-1871.

Koshy, A., Grover, S., Hyams, K. C., Shabrawy, M. A., Pacsa, A. (1996). Short-term IgM and IgG antibody responses to hepatitis E virus infection. *Scandinavian Journal of Infectious Diseases* 28: 439-441.

Koup, R. A., Merluzzi, V. J., Hargrave, K. D., Adams, J., Grozinger, K. et al. (1991). Inhibition of human immunodeficiency virus type I (HIV-I) replication by the dipyrindodiazepinone BI-RG-587. *Journal of Infectious Diseases* 163: 966-970.

Krawczynski, K. and Bradley, D. W. (1989). Enterically transmitted non-A, non-B hepatitis: identification of virus-associated antigen in experimentally infected cynomolgus macaques. *Journal of Infectious Diseases* 159: 1042-1049.

Kudoh, S. (1998). Erythromycin treatment in diffuse panbronchiolitis. *Current Opinion in Pulmonary Medicine* 4: 116-121.

Kudoh, S., Azuma, A., Yamamoto, M., Izumi, T., Ando, M. (1998). Improvement of survival in patients with diffuse panbronchiolitis treated with low-dose erythromycin. *American Journal of Respiratory and Critical Care Medicine* 157: 1829-1832.

Lai, C. L., Chien, R. N., Leung, N. W. Y., Chang, T. T., Guan, R. et al. (1998). A one year trial of lamivudine for chronic hepatitis B. *New England Journal of Medicine* 339: 61-68.

Lau, D. T. Y., Everhart, J., Kleiner, D. E., Park, Y., Vergalla, J. et al. (1997). Long-term follow-up in patients with chronic hepatitis B treated with interferon alfa. *Gastroenterology* 113: 1660-1667.

Lee, J. S., Chu, I. S., Heo, J., Calvisi, D. F., Sun, Z. et al. (2004). Classification and prediction of survival in hepatocellular carcinoma by gene expression profiling. *Hepatology* 40: 667-676.

Lee, J. S., Thorgeirsson, S. S. (2005). Genetic profiling of human hepatocellular carcinoma. *Seminars in Liver Disease* 25: 125-132.

Lee, S. D., Chan, C. Y., Yu, M. I., Wang, Y. J., Chang, F. Y. et al. (1999). A two doses combined hepatitis A and B vaccine in Chinese youngsters. *Journal of Medical Virology* 59: 1-4.

Lesniewski, R., Okasinski, G., Carrick, R., Vansant, C., Desai, S. et al. (1995). Antibody to hepatitis C virus second envelope (HCV-E2) glycoprotein: a new marker of HCV infection closely associated with viremia. *Journal of Medical Virology* 45: 415-422.

Li, F., Zhuang, H., Kolivas, S., Locarnini, S. A., Anderson, D. A. (1994). Persistent and transient antibody responses to hepatitis E virus detected by Western immunoblot using open reading frame 2 and 3 and glutathione S-transferase fusion proteins. *Journal of Clinical Microbiology* 32: 2060-2066.

Li, F., Torresi, J., Locarnini, S. A., Zhuang, H., Zhu, W. et al. (1997). Amino-terminal epitopes are exposed when full-length open reading frame 2 of hepatitis E virus is expressed in *Escherichia coli*, but carboxy-terminal epitopes are masked. *Journal of Medical Virology* 52: 289-300.

Li, T. C., Zhang, J., Shinzawa, H., Ishibashi, M., Sata, M. et al. (2000). Empty virus-like particle based enzyme-linked immunosorbent assay for antibodies to hepatitis E virus. *Journal of Medical Virology* 62: 327-333.

Lin, C. C., Wu, J. C., Chang, T. T., Chang, W. Y., Yu, M. L. et al. (2000). Diagnostic value of immunoglobulin G (IgG) and IgM anti-hepatitis E virus (HEV) test based on HEV RNA in area where hepatitis E is not endemic. *Journal of Clinical Microbiology* 11: 3915-3918.

Llewelyn, C. A., Hewitt, P. E., Knight, R. S., Amar, K., Cousens, S. et al. (2004). Possible transmission of variant Creutzfeldt-Jakob disease by blood transfusion. *Lancet* 363: 417-421.

Luzuriaga, K., Bryson, Y., McSherry, G., Robinson, J., Stechenberg, B. et al. (1996). Pharmacokinetics, safety, and activity of nevirapine in human immunodeficiency virus type 1-infected children. *Journal of Infectious Diseases* 174(4): 713-721.

Ma, Y., Lin, S. Q., Gao, Y., Li, M., Luo, W. X. et al. (2003). Expression of ORF2 partial gene of hepatitis E virus in tomatoes and immunoactivity of expression products. *World Journal of Gastroenterology* 9: 2211-2215.

Mancini-Bourguine, M., Fontaine, H., Scott-Algara, D., Pol, S., Brechot, C. et al. (2004). Induction or expansion of T-cell responses by a hepatitis B DNA vaccine administered to chronic HBV carriers. *Hepatology* 40(4): 874-882.

Marlin, S. D., Staunton, D. E., Springer, T. A., Stratowa, C., Sommergruber, W. et al. (1990). A soluble form of intercellular adhesion molecule-1 inhibits rhinovirus infection. *Nature* 344: 70-72.

Mason, U., Aldrich, J., Breedveld, F., Davis, C. B., Elliott, M. et al. (2002). CD4 Coating, but not CD4 depletion is a predictor of efficacy with primatized monoclonal anti-CD4 treatment of active rheumatoid arthritis. *Journal of Rheumatology* 29: 220-229.

Mast, E. E., Alter, M. J., Holland, P. V., Purcell, R. H. (1998). Evaluation of assays for antibody to hepatitis E virus by a serum panel. *Hepatology* 27(3): 857-861.

Mathieson, P. W., Cobbold, S. P., Hale, P., Clark, M. R., Oliveira, D. B. et al. (1990). Monoclonal-antibody therapy in systemic vasculitis. *New England Journal of Medicine* 323: 250-254.

Matthews, J. B., Ramos, E., Bluestone, J. A. (2003). Clinical trials of transplant tolerance: Slow but steady progress. *American Journal of Transplantation* 3: 794-803.

Mazzulli, T., Rusconi, S., Merrill, D. P., D'Aquila, R. T., Moonis, M. (1994). Alternating versus continuous drug regimens in combination chemotherapy of human immunodeficiency virus type I infection in vitro. *Antimicrobial Agents and Chemotherapy* 38(4): 656-661.

McAtee, C. P., Zhang, Y., Yarbough, P. O., Bird, T., Fuerst, T. R. (1996a). Purification of soluble hepatitis E open reading frame 2- derived protein with unique antigenic properties. *Protein Expression and Purification* 8: 262-270.

McAtee, C. P., Zhang, Y., Yarbough, P. O., Fuerst, T. R., Stone, K. L. *et al.* (1996b). Purification and characterization of a recombinant hepatitis E protein vaccine candidate by liquid chromatography-mass spectrometry. *Journal of Chromatography B - Biomedical Applications* 685: 91-104.

McConkey, S. J., Reece, W. H., Moorthy, V. S., Webster, D., Dunachie, S. *et al.* (2003). Enhanced T-cell immunogenicity of plasmid DNA vaccines boosted by recombinant modified vaccinia virus Ankara in humans. *Nature Medicine* 9: 729-735.

McMahon, B. J., Bruden, D. L., Petersen, K. M., Bulkow, L. R., Parkinson, A. J. *et al.* (2005). Antibody levels and protection after hepatitis B vaccination: results of a 15-year follow-up. *Annals of Internal Medicine* 142(5): 333-341.

McNamara, M. J., Phillips, I. A., Williams, O. B. (1969). Viral and Mycoplasma pneumoniae infections in exacerbations of chronic lung disease. *American Review of Respiratory Disease* 100: 19-24.

Meng, J., Dai, X., Chang, J. C., Lopareva, E., Pillot, J. *et al.* (2001). Identification and characterization of the neutralization epitope(s) of the hepatitis E virus. *Virology* 288(2): 203-211.

Merluzzi, V. J., Hargrave, K. D., Labadia, M., Grozinger, K., Skoog, M. *et al.* (1990). Inhibition of HIV-I replication by a nonnucleoside reverse transcriptase inhibitor. *Science* 250(4986): 1411-1413.

Merrill, D. P., Moonis, M., Chou, T. C., Hirsch, M. S. (1996). Lamivudine or stavudine in two- and three-drug combinations against human immunodeficiency virus type I replication in vitro. *Journal of Infectious Diseases* 173: 355-364.

Micallef, J. M., Kaldor, J. M., Dore, G. J. (2006). Spontaneous viral clearance following acute hepatitis C infection: a systematic review of longitudinal studies. *Journal of Viral Hepatology* 13(1): 34-41.

Millan, M. T., Shizuru, J. A., Hoffmann, P., Dejbakhsh-Jones, S., Scandling, J. D. *et al.* (2002a). Mixed chimerism and immunosuppressive drug withdrawal after HLA-mismatched kidney and hematopoietic progenitor transplantation. *Transplantation* 73: 1386-1391.

Millan, M. T., Shizuru, J. A., Hoffmann, P., Dejbakhsh-Jones, S., Scandling, J. D. *et al.* (2002b). Mixed chimerism and donor-specific unresponsiveness without graft-versus-host disease after MHC-mismatched hematopoietic stem cell infusion and kidney transplantation [abstract 59]. *Transplantation* 74: 37.

Misko, I. S., Kane, R. G., Pope, J. H. (1981). Generation in vitro of EBV-induced specific cytotoxic T cells in autologous serum avoids complications due to self-preferred foetal calf serum-specific T-cell cytotoxicity. *International Journal of Cancer* 27(4): 513-519.

Misko, I. S., Soszynski, T. D., Kane, R. G., Pope, J. H. (1984). Factors influencing the human cytotoxic T cell response to autologous lymphoblastoid cell lines in vitro. *Clinical Immunology and Immunopathology* 32(3): 285-297.

Monto, A. S. (1995). Epidemiology of respiratory viruses in person with and without asthma and COPD. *American Journal of Respiratory and Critical Care Medicine* 151: 1653-1658.

Moore, A. C. and Hill, A. V. S. (2004). Progress in DNA-based heterologous prime-boost immunization strategies for malaria. *Immunological Reviews* 199: 126-143.

Morel, P., Vincent, C., Cordier, G., Panaye, G., Carosella, E. et al. (1990). Anti-CD4 monoclonal antibody administration in renal transplanted patients. *Clinical Immunology and Immunopathology* 56: 311-322

Moss, D. J., Rickinson, A. B., Pope, J. H. (1978). Long-term T-cell-mediated immunity to Epstein-Barr virus in man. I. Complete regression of virus-induced transformation in cultures of seropositive donor leukocytes. *International Journal of Cancer* 22: 662-668.

Moss, D. J., Rickinson, A. B., Pope, J. H. (1979). Long-term T-cell-mediated immunity to Epstein-Barr virus in man. III. Activation of cytotoxic T-cells in virus-infected leukocyte cultures. *International Journal of Cancer* 23: 618-625.

Mould, D. R., Davis, C. B., Minthorn, E. A., Kwok, D. C., Elliott, M. J. et al. (1999). A population pharmacokinetic-pharmacodynamic analysis of single doses of clenoliximab in rheumatoid arthritis patients. *Clinical Pharmacology and Therapeutics* 66: 246-257.

Murray, R. J., Kurilla, M. G., Brooks, J. M., Thomas, W. A., Rowe, M. et al. (1992). Identification of target antigens for the human cytotoxic T-cell response to Epstein-Barr virus (EBV): implications for the immune control of EBV-positive malignancies. *Journal of Experimental Medicine* 176:157-168.

Nainan, O. V., Stevens, C. E., Taylor, P. E., Margolis, H. S. (1997). Hepatitis B virus (HBV) antibody resistant mutants among mothers and infants with chronic HBV infection. In: Rizzetto M, Purcell RH, Gerin JL, Verme G, eds. *Viral Hepatitis and Liver Disease*. Turin: Edizioni Minerva Medica. pp132-134.

Nakano, I., Fukuda, Y., Katano, Y., Nakano, S., Kumada, T. et al. (1999). Why is the interferon sensitivity-determining region (ISDR) system useful in Japan? *Journal of Hepatology* 30(6): 1014-1022.

Newman, R., Hariharan, K., Reff, M., Anderson, D. R., Braslawsky, G. et al. (2001). Modification of the Fc region of a primatized IgG antibody to human CD4 retains its ability to modulate CD4 receptors but does not deplete CD4(+) T cells in chimpanzees. *Clinical Immunology* 98(2): 164-174.

Ngui, S. L., O'Connell, S., Eglin, R. P., Heptonstall, J., Teo, C. G. (1997). Low detection rate and maternal provenance of hepatitis B virus S gene mutants in cases of failed postnatal immunoprophylaxis in England and Wales. *Journal of Infectious Diseases* 176: 1360-1365.

Nishida, N., Nishimura, T., Ito, T., Komeda, T., Fukuda, Y. et al. (2003). Chromosomal instability and human hepatocarcinogenesis. *Histology and Histopathology* 18: 897-909.

Nishihara, T., Nozaki, C., Nakatake, H., Hoshiko, K., Esumi, M. et al. (1993). Secretion and purification of hepatitis C virus NS1 glycoprotein produced by recombinant baculovirus-infected insect cells. *Gene* 129: 207-214.

Nussenweig, V., and Nussenweig, R. S. (1989). Rationale for the development of an engineered sporozoite malaria vaccine. *Advances in Immunology* 45: 193-205.

Obriadina, A., Meng, J. H., Ulanova, T., Trinta, K., Burkov, A. et al. (2002a). A new enzyme immunoassay for the detection of antibody to hepatitis E virus. *Journal of Gastroenterology & Hepatology* 17 Suppl 3: S360-S364.

Obriadina, A., Meng, J. H., Lopareva, E. et al. (2002b). *Proceedings of 10th International Symposium on Viral Hepatitis and Liver Disease*. London: International Medical Press. pp117-121.

Ogata, N., Zanetti, A. R., Yu, M., Miller, R. H., Purcell, R. H. (1997). Infectivity and pathogenicity in chimpanzees of a surface gene mutant of hepatitis B virus that emerged in a vaccinated infant. *Journal of Infectious Diseases* 175: 511-523.

Ogata, N., Cote, P. J., Zanetti, A. R., Miller, R. H., Shapiro, M. et al. (1999). Licensed recombinant hepatitis B vaccines protect chimpanzees against infection with the prototype surface gene mutant of hepatitis B virus. *Hepatology* 30(3): 779-786.

Okamoto, H., Yano, K., Nozaki, Y., Matsui, A., Miyazaki, H. et al. (1992). Mutations within the S gene of hepatitis B virus transmitted from mothers to babies immunized with hepatitis B immune globulin and vaccine. *Pediatric Research* 32: 264-268.

Pancholi, P., Lee, D. H., Liu, Q., Tackney, C., Taylor, P. (2001). DNA prime/canarypox boost-based immunotherapy of chronic hepatitis B virus infection in a chimpanzee. *Hepatology* 33(2): 448-454.

Patel, S. S., and Benfield, P. (1996). Nevirapine. *Clin Immunother* 6: 307-317.

Patterson, J. L. and Carrion, R. (2005). Demand for nonhuman primate resources in the age of biodefense. *Institute for Laboratory Animal Research Journal* 46(1): 15-22.

Paul, D. A., Knigge, M. F., Ritter, A., Gutierrez, R., Pilot-Matias, T. et al. (1994). Determination of hepatitis E virus seroprevalence by using recombinant fusion proteins and synthetic peptides. *Journal of Infectious Diseases* 169: 801-806.

Peden, A. H., Head, M. W., Ritchie, D. L., Bell, J. E., Ironside, J. W. (2004). Preclinical vCJD after blood transfusion in a PRNP codon 129 heterozygous patient. *Lancet* 364: 529-531.

Pepperl, S., Benninger-Doring, G., Modrow, S., Wolf, H., Jilg, W. (1998). Immediate-early transactivator Rta of Epstein-Barr virus (EBV) shows multiple epitopes recognized by EBV-specific cytotoxic T lymphocytes. *Journal of Virology* 72(11): 8644-8649.

Perel, P., Roberts, I., Sena, E., Wheble, P., Sandercock, P., Macleod, M., Mignini, L. E., Jayaram, P., Khan, K. S. (2006). Available at:
http://www.pcpoh.bham.ac.uk/publichealth/nccrm/PDFs%20and%20documents/Publications/JH18_Final_Report_May_2006.pdf
Accessed December 13th 2006

Pesquies, P. C., Milhaud, C. L., Cailler, B. G., Kaplan, D., Nogues, C. F. (1978). Use of a primate as an experimental model on board SPACELAB: definition of a two-primate biological facility. *Life Sciences and Space Research* 16: 127-130.

Pillot, J., Turkoglu, S., Dubreuil, P., Cosson, A., Lemaigre, G. et al. (1995). Cross reactive immunity against different strains of the hepatitis E virus transferable by simian and human sera. *Comptes Rendus de l'Academie des Sciences serie III-sciences de la vie-life sciences* 318: 1059-1064.

Pound, P., Ebrahim, S., Sandercock, P., Bracken, M. B., Roberts, I. (2004). Where is the evidence that animal research benefits humans? *British Medical Journal* 328: 514-517.

Prince, A. M., Whalen, R., Brotman, B. (1997). Successful nucleic acid based immunization of newborn chimpanzees against hepatitis B virus. *Vaccine* 15(8): 916-919.

Protzer-Knolle, U., Naumann, U., Bartenschlager, R., Berg, T., Hopf, U. et al. (1998). Hepatitis B virus with antigenically altered hepatitis B surface antigen is selected by high-dose hepatitis B immune globulin after liver transplantation. *Hepatology* 27: 254-263.

Purdy, M. A., McCaustland, K. A., Krawczynski, K., Tam, A., Beach, M. J. (1992). Expression of hepatitis E virus (HEV)-trpE fusion protein containing epitopes recognized by antibodies in sera from human cases and experimentally infected primates. *Archives of Virology* 123: 335-349.

Purdy, M. A., McCaustland, K. A., Krawczynski, K., Spelbring, J., Reyes, G. R. et al. (1993). Preliminary evidence that a trpE-HEV fusion protein protects cynomolgus

macaques against challenge with the wild-type hepatitis E virus (HEV). *Journal of Medical Virology* 41: 90-94.

Ralston, R., Thudium, K., Berger, K., Kuo, C., Gervase, B. (1993). Characterization of hepatitis C virus envelope glycoprotein complexes expressed by recombinant vaccinia viruses. *Journal of Virology* 67: 6753-6761.

Reddy, M. P., Kinney, C. A., Chaikin, M. A., Payne, A., Fishman-Lobell, J. *et al.* (2000). Elimination of Fc receptor-dependent effector functions of a modified IgG4 monoclonal antibody to human CD4. *Journal of Immunology* 164(4): 1925-1933.

Regev, A. and Schiff, E. R. (1999). Viral hepatitis. *Current Opinion in Gastroenterology* 15(3): 234-239.

Reyes, G. R., Purdy, M. A., Kim, J. P., Luk, K. C., Young, L. M. *et al.* (1990). Isolation of a cDNA from the virus responsible for enterically transmitted non-A, non-B hepatitis. *Science* 247: 1335-1339.

Richman, D., Rosenthal, A. S., Skoog, M., Eckner, R. J., Chou, T. C. *et al.* (1991). BI-RG-587 is active against zidovudine-resistant human immunodeficiency virus type I and synergistic with zidovudine. *Antimicrobial Agents and Chemotherapy* 35(2): 305-308.

Riddell, M. A., Li, F., Anderson, D. A. (2000). Identification of immunodominant and conformational epitopes in the capsid protein of hepatitis E virus by using monoclonal antibodies. *Journal of Virology* 74: 8011-8017.

Robb, M. L., Polonis, V., Vahey, M., Gartner, S., Michael, N. *et al.* (1992). HIV neutralization assay using polymerase chain reaction derived molecular signals. *Journal of Acquired Immune Deficiency Syndromes and Human Retrovirology* 5: 1224-1229.

Robertson, C. A., Mok, J. Y., Froebel, K. S., Simmonds, P., Burns, S. M. *et al.* (1992). Maternal antibodies to gp120 V3 sequence do not correlate with protection against vertical transmission of human immunodeficiency virus. *Journal of Infectious Diseases* 166(4): 704-709.

Robinson, R. A., Burgess, W. H., Emerson, S. U., Leibowitz, R. S., Sosnovtseva, S. A. *et al.* (1998). Structural characterization of recombinant hepatitis E virus ORF2 proteins in baculovirus-infected insect cells. *Protein Expression and Purification* 12: 75-84.

Rogel, M. E., Wu, L. I., Emerman, M. (1995). The human immunodeficiency virus type 1 vpr gene prevents cell proliferation during chronic infection. *Journal of Virology* 69: 882-888.

Rottinghaus, S. T., Poland, G. A., Jacobson, R. M., Barr, L. J., Roy, M. J. (2003). Hepatitis B DNA vaccine induces protective antibody responses in human non-responders to conventional vaccination. *Vaccine* 21(31): 4604-4608.

Roy, M. J., Wu, M. S., Barr, L. J., Fuller, J. T., Tussey, L. G. et al. (2000). Induction of antigen-specific CD8+ cells, T helper cells, and protective levels of antibody in humans by particle-mediated administration of a hepatitis B virus DNA vaccine. *Vaccine 19*: 764-778.

Rusconi, S., Merrill, D. P., Hirsch, M. S. (1994). Inhibition of human immunodeficiency virus type I replication in cytokine-stimulated monocytes/macrophages by combination therapy. *Journal of Infectious Diseases 170*: 1361-1366.

Safar, J., Wille, H., Itri, V., Groth, D., Serban, H. et al. (1998). Eight prion strains have PrPsc molecules with different conformations. *Nature Medicine 4*: 1157-1165.

Safary, A. (2001). Perspectives of vaccination against hepatitis E. *Intervirology 44*: 162-166.

Sallberg, M., Hughes, J., Javadian, A., Ronlov, G., Hultgren, C. et al. (1998). Genetic immunization of chimpanzees chronically infected with the hepatitis B virus, using a recombinant retroviral vector encoding the hepatitis B virus core antigen. *Human Gene Therapy 9*: 1719-1729.

Sato, K., Suga, M., Akaike, T., Fujii, S., Muranaka, H. et al. (1998). Therapeutic effects of erythromycin on influenza virus-induced lung injury in mice. *American Journal of Respiratory and Critical Care Medicine 157*: 853-857.

Sauer, U. G. (2000). Reasons for not using primates in research [Article in German]. *ALTEX (Alternativen zu Tierexperimenten) 17(4)*: 217-220.

Schmerr, M. J., Jenny, A. L., Bulgin, M. S., Miller, J. M., Hamir, A. N. et al. (1999). Use of capillary electrophoresis and fluorescent labeled peptides to detect the abnormal prion protein in the blood of animals that are infected with a transmissible spongiform encephalopathy. *Journal of Chromatography A 853*: 207-214.

Schneider, J., Langermans, J. A., Gilbert, S. C., Blanchard, T. J., Twigg, S. et al. (2001). A prime-boost immunisation regimen using DNA followed by recombinant modified vaccinia virus Ankara induces strong cellular immune responses against the Plasmodium falciparum TRAP antigen in chimpanzees. *Vaccine 19*: 4595-4602.

Schofield, D. J., Purcell, R. H., Nguyen, H. T., Emerson, S. U. (2003). Monoclonal antibodies that neutralize HEV recognize an antigenic site at the carboxyterminus of an ORF2 protein vaccine. *Vaccine 22*: 257-267.

Schrezenmeier, H. and Fleischer, B. (1988). A regulatory role for the CD4 and CD8 molecules in T cell activation. *Journal of Immunology 141(2)*: 398-403.

Sleasman, J. W. and Goodenow, M. M. (2003). HIV-1 infection. *Journal of Allergy and Clinical Immunology 111(2 Suppl)*: S582-S592.

Smith, C. B., Golden, C. A., Kanner, R. E., Renzetti, A. D. Jnr. (1980). Association of viral and Mycoplasma pneumoniae infections with acute respiratory illness in patients with chronic obstructive pulmonary diseases. *American Review of Respiratory Disease* 121: 225-232.

Sokal, E. M., Conjeevaram, H. S., Roberts, E. A., Alvarez, F., Bern, E. M. et al. (1998). Interferon alfa therapy for chronic hepatitis B in children: a multinational randomized controlled trial. *Gastroenterology* 114: 988-995.

Spaete, R. R., Alexander, D., Rugroden, M. E., Choo, Q. L., Berger, K. et al. (1992). Characterization of the hepatitis C virus E2/NS1 gene product expressed in mammalian cells. *Virology* 188: 819-830.

Stenhouse, A. C. (1967). Rhinovirus infection in acute exacerbation of chronic bronchitis: a controlled prospective study. *British Medical Journal* 3: 461-463.

Steven, N. M., Annels, N. E., Kumar, A., Leese, A. M., Kurilla, M. G. et al. (1997). Immediate early and early lytic cycle proteins are frequent targets of the Epstein-Barr virus-induced cytotoxic T cell response. *Journal of Experimental Medicine* 185(9): 1605-1617.

Stevenson, M., Stanwick, T. L., Dempsey, M. P., Lamonica, C. A. (1990). HIV-1 replication is controlled at the level of T cell activation and proviral integration. *EMBO Journal* 9: 1551-1560.

Su, A. I., Pezacki, J. P., Wodicka, L., Brideau, A. D., Supekova, L. et al. (2002). Genomic analysis of the host response to hepatitis C virus infection. *Proceedings of the National Academy of Sciences of the USA* 99: 15669-15674.

Suzuki, T., Yanai, M., Yamaya, M., Satoh-Nakagawa, T., Sekizawa, K. et al. (2001). Erythromycin and common cold in COPD. *Chest* 120(3): 730-733.

Takeuchi, T., Schlossman, S. F., Morimoto, C. (1987). The T4 molecule differentially regulating the activation of subpopulations of T4+ cells. *Journal of Immunology* 139(3): 665-671.

Tam, A. W., Smith, M. M., Guerra, M. E., Huang, C. C., Bradley, D. W. et al. (1991). Hepatitis E virus (HEV): molecular cloning and sequencing of the full-length viral genome. *Virology* 185: 120-31.

Tan, L. C., Gudgeon, N., Annels, N. E., Hansasuta, P., O'Callaghan, C. A. et al. (1999). A re-evaluation of the frequency of CD8+ T cells specific for EBV in healthy virus carriers. *Journal of Immunology* 162(3): 1827-1835.

Tassopoulos, N. C. (1998). Chronic hepatitis B: interferon-monotherapy or combination with nucleoside analogues? *Liver* 18: 71-72.

Teran, L. M., Johnston, S. L., Schroder, J-M., Church, M. K., Holgate, S. T. (1997). Role of nasal interleukin-8 in neutrophil recruitment and activation in children with

virus-induced asthma. *American Journal of Respiratory and Critical Care Medicine* 155: 1362-1366.

[Thew, M. \(2002\). Are results of primate research worth the suffering it causes? *Nature* 418\(6895\): 273.](#)

[Thoelen, S., Van Damme, P., Leentvaar-Kuypers, A., Leroux-Roels, G., Bruguera, M. et al. \(1999\). The first combined vaccine against hepatitis A and B: An overview. *Vaccine* 17: 1657-1662.](#)

[Tong, A. W. and Stone, M. J. \(2003\). Prospects for CD40-directed experimental therapy of human cancer. *Cancer Gene Therapy* 10\(1\): 1-13.](#)

Trkola, A., Kuster, H., Rusert, P., Joos, B., Fischer, M. et al. (2005). Delay of HIV-1 rebound after cessation of antiretroviral therapy through passive transfer of human neutralizing antibodies. *Nature Medicine* 11(6): 615-622.

[Trono, D. \(1992\). Partial reverse transcripts in virions from human immunodeficiency and murine leukaemia viruses. *Journal of Virology* 66: 4893-4900.](#)

[Tsarev, S. A., Tsareva, T. S., Emerson, S. U., Kapikian, A. Z., Ticehurst, J. et al. \(1993\). ELISA for antibody to hepatitis E virus \(HEV\) based on complete open-reading frame-2 protein expressed in insect cells: Identification of HEV infection in primates. *Journal of Infectious Diseases* 168: 369-378.](#)

[Tsarev, S. A., Tsareva, T. S., Emerson, S. U., Govindarajan, S., Shapiro, M. et al. \(1994\). Successful passive and active immunization of cynomolgus monkeys against hepatitis E. *Proceedings of the National Academy of Sciences of the USA* 91: 10198-10202](#)

Turner, R. B., Weingand, K. W., Yeh, C-H., Leedy, D. (1998) .Association between nasal secretion interleukin-8 concentration and symptom severity in experimental rhinovirus colds. *Clinical Infectious Diseases* 26: 840-846.

Turner, R. B., Wecker, M. T., Pohl, G., Witek, T. J., McNally, E. et al. (1999). Efficacy of tremacamra, a soluble intercellular adhesion molecule 1, for experimental rhinovirus infection: a randomized clinical trial. *Journal of the American Medical Association (JAMA)* 281(19): 1797-1804.

Ulmer, J. B., Donnelly, J. J., Parker, S. E., Rhodes, G. H., Felgner, P. L. et al. (1993). Heterologous protection against influenza by injection of DNA encoding a viral protein. *Science* 258: 1745-1849.

[VandeBerg, J. L. and Zola, S. M. \(2005\). A unique biomedical resource at risk. *Nature* 437: 30-32.](#)

[Van Der Wielen, M., van Damme, P., Collard, F. \(2000\). A two dose schedule for combined hepatitis A and hepatitis B vaccination in children ages one to eleven years. *Pediatric Infectious Disease Journal* 19: 848-853.](#)

- Varki, A. and Altheide, T. K. (2005). Comparing the human and chimpanzee genomes: searching for needles in a haystack. *Genome Research* 15(12): 1746-1758.
- Veale, D. J., Reece, R. J., Parsons, W., Radjenovic, A., O'Connor, P. J. *et al.* (1999). Intra-articular primatised anti-CD4: Efficacy in resistant rheumatoid knees. A study of combined arthroscopy, magnetic resonance imaging and histology. *Annals of the Rheumatic Diseases* 58: 342-349.
- Vento, S., Garofano, T., Renzini, C., Cainelli, F., Casali, F. *et al.* (1998). Fulminant hepatitis associated with hepatitis A virus superinfection in patients with chronic hepatitis C. *New England Journal of Medicine* 338: 286-290.
- Wang, Y. F., Brotman, B., Andrus, L., Prince, A. M. (1996). Immune response to epitopes of hepatitis C virus (HCV) structural proteins in HCV-infected humans and chimpanzees. *Journal of Infectious Diseases* 173(4): 808-821.
- Waters, J. A., Bailey, C., Love, C., Thomas, H. C. (1998). A study of the antigenicity and immunogenicity of a new hepatitis B vaccine using a panel of monoclonal antibodies. *Journal of Medical Virology* 54: 1-6.
- WHO (World Health Organisation: Vaccine Assessment and Monitoring Team, Department of Immunization, Vaccines and Biologicals) (2005) *WHO Vaccine Preventable Diseases: Monitoring System 2005 Global Summary*. Available: <http://www.who.int/vaccines-documents/GlobalSummary/GlobalSummary.pdf>. Accessed 2006 May 11.
- Wolff, J. A., Malone, R. W., Williams, P., Chong, W., Acsadi, G. *et al.* (1990). Direct gene transfer into mouse muscle in vivo. *Science* 247: 1465-1468.
- Worm, H. C., Wurzer, H., Frösner, G. (1998). Sporadic hepatitis E in Austria. *New England Journal of Medicine* 339: 1554-1555.
- Worm, H. C. and Wirnsberger, G. (2004). Hepatitis E Vaccines: Progress and Prospects. *Drugs* 64(14): 1517-1531.
- Wu, J. C., Sheen, I. J., Chiang, T. Y., Sheng, W. Y., Wang, Y. J. *et al.* (1998). The impact of traveling to endemic areas on the spread of hepatitis E virus infection: epidemiological and molecular analysis. *Hepatology* 27: 1415-1420.
- Yamashita, T., Kaneko, S., Hashimoto, S., Sato, T., Nagai, S. *et al.* (2001). Serial analysis of gene expression in chronic hepatitis C and hepatocellular carcinoma. *Biochemical and Biophysical Research Communications* 282: 647-654.
- Yang, G., D'Souza, M. P., Vyas, G. N. (1998). Neutralizing Antibodies Against HIV Determined by Amplification of Viral Long Terminal Repeat Sequences From Cells Infected in vitro by Nonneutralized Virions. *Journal of Acquired Immune Deficiency Syndromes and Human Retrovirology* 17(1): 27-34.

Yang, W. C., Yeung, E. S., Schmerr, M. J. (2005). Detection of prion protein using a capillary electrophoresis-based competitive immunoassay with laser-induced fluorescence detection and cyclodextrin-aided separation. *Electrophoresis* 26: 1751-1759.

Yarborough, P. O., Tam, A. W., Fry, K. E., Krawczynski, K., McCaustland, K. A. et al. (1991). Hepatitis E virus: identification of type-common epitopes. *Journal of Virology* 65: 5790-5797.

Ye, Q. H., Qin, L. X., Forgues, M., He, P., Kim, J. W. et al. (2003). Predicting hepatitis B virus-positive metastatic hepatocellular carcinomas using gene expression profiling and supervised machine learning. *Nature Medicine* 9: 416-423.

Yocum, D. E., Solinger, A. M., Tesser, J., Gluck, O., Cornett, M. et al. (1998). Clinical and immunologic effects of a PRIMATIZED anti-CD4 monoclonal antibody in active rheumatoid arthritis: results of a phase I, single dose, dose escalating trial. *Journal of Rheumatology* 25(7): 1257-1262.

Zeuzem, S., Lee, J. H., Roth, W. K. (1997). Mutations in the nonstructural 5A gene of European hepatitis C virus isolates and response to interferon alfa. *Hepatology* 25: 740-744.

Zhang, H., Dornadula, G., Pomerantz, R. J. (1996a). Endogenous reverse transcription of human immunodeficiency virus type I in physiological microenvironments: an important stage for viral infection of nondividing cells. *Journal of Virology* 70(5): 2809-2824.

Zhang, H., Dornadula, G., Wu, Y., Havlir, D., Richman, D. D. et al. (1996b). Kinetic analysis of intravirion reverse transcription in the blood plasma of human immunodeficiency virus type I-infected individuals: direct assessment of resistance to reverse transcriptase inhibitors in vivo. *Journal of Virology* 70(1): 628-634.

Zhang, H., Dornadula, G., Pomerantz, R. J. (1998). Natural endogenous reverse transcription of HIV type I. *AIDS Research and Human Retroviruses* 14 Suppl 1: S93-S95.

Zhu, Z., Tang, W., Ray, A., Wu, Y., Einarsson, O. et al. (1996). Rhinovirus stimulation of interleukin-6 in vivo and in vitro: evidence for nuclear factor kB-dependent transcriptional activation. *Journal of Clinical Investigation* 97: 421-430.

Zhu, Z., Tang, W. L., Gwaltney, J. M., Elias, J. A. (1997). Rhinovirus stimulation of interleukin-8 in vivo and in vitro: role of NF-Kappa-beta. *American Journal of Physiology* 17: L814-L824.