

# Calcium Phosphate Mineralization on Calcium Carbonate Particle Incorporated Silk-Fibroin Composites

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## Abstract

In this study, three anhydrous forms of calcium carbonate, namely vaterite, aragonite and calcite, with distinct morphologies were incorporated inside silk-fibroin to fabricate composite scaffolds for tissue engineering applications. To assess calcium phosphate mineralization, composite scaffolds were treated with simulated body fluid up to one month. It was observed that composite scaffolds having different calcium carbonate polymorphs expressed different mineralization. Incorporating 25 wt. % of vaterite polymorph, which was the least stable form of calcium carbonate under aqueous conditions, induced the highest calcium phosphate mineralization in silk-fibroin while calcium carbonate-free silk-fibroin scaffolds expressed no calcium phosphate deposition. Results highlighted the importance of calcium carbonate particles in enhancing the bioactivity of silk-fibroin based composite scaffolds.

**Keywords:** Silk-fibroin, CaCO<sub>3</sub>, composite, biomineralization, apatite.

## 1. Introduction

The fibroin core of silk – silk-fibroin (SF) - which is encapsulated inside a sericin outer lining, is a protein based fibrous biopolymer. Owing to its biocompatible nature, its use in various biomedical applications is investigated, i.e. sutures, wound dressings, vascular grafts, cartilage and bone regeneration agents and etc. [1]. Having this said, there are inherent problems associated with the use of SF as a biomaterial. Firstly, SF is not an osteoconductive (permits bone growth) material. It does not stimulate cellular functions of the juxtaposed tissues. Secondly, mechanical properties of SF are not suited for hard tissues. If SF was to be widely used in clinics as a biomaterial, its bioactivity and mechanical properties need to be remedied.

Combining two or more biocompatible materials to match the requirements of a specific application is a promising approach in biomedical research. In fact, composite forming strategy via incorporation of secondary phase particles provides remarkable advantages for SF based biomaterials so that the mechanical strength and bioactivity of SF could be improved simultaneously. For instance, Wang et al. dispersed graphene oxide (GO) sheets via ultrasonication to prepare GO/SF composites by solvent evaporation [2].

GO layers were found to improve thermal stability and mechanical strength simultaneously at low

concentrations. Multiwalled carbon nanotubes (MWCNT) could also be incorporated into SF [3]. When the amount of MWCNT's increased up to 1 wt. %, the beta ( $\beta$ ) sheet content, which was responsible for mechanical strength of the SF, increased from 35 % to 49.4 % with a corresponding 4.4-fold increase in the elastic modulus. In a recent study, up to 20 % TiO<sub>2</sub> particles were incorporated in SF, which increased compressive strength for 15 % [4]. Further incorporation of TiO<sub>2</sub> particles led to agglomeration and failed to disperse homogeneously in the SF matrix and thus strength did not increase any more. When samples were soaked in simulated body fluid (SBF), spherical apatite crystals formed on composite scaffold surfaces up to 21 days.

Apatite is a family of calcium phosphate (CaP) minerals with Ca/P atomic ratio between 1.5-1.67 and apatite constitutes the major inorganic part of the bone [5]. Owing to its osteoinductive potential and bioactive nature, recently it became a significantly promising candidate to reinforce SF [6]. In this regard, synthetic CaP minerals have been utilized to improve bone cell functions on SF scaffolds [7, 8]. Recently, well-known apatite phase, hydroxyapatite (HA; Ca/P: 1.67) was used to reinforce SF. But it was not possible to disperse HA particles homogeneously in aqueous media and this led to agglomeration upon incorporation into SF at high concentrations [9]. To inhibit agglomeration of secondary phase particles and enhance SF biocompatibility, it was recently suggested that SF

could be utilized as a template for the formation of biological apatites [10]. Specifically, the amorphous links in SF  $\beta$ -sheets could act as nucleation sites for HA crystals due to mimicking the anionic structure of non-collagenous proteins present in bone tissue [11].

To further stimulate formation of uniformly distributed apatite minerals within SF matrix, dispersing calcium sources that can form apatite crystals in-situ in SF would be a promising approach for bone regeneration. Among various calcium sources that can form apatite crystals in-situ, calcium carbonate ( $\text{CaCO}_3$ ) polymorphs drew significant attention owing to their dissolution capability in aqueous media. In fact,  $\text{CaCO}_3$  has three anhydrous polymorphs: i) vaterite, ii) aragonite and iii) calcite.

Vaterite is the least stable and thus most soluble form of  $\text{CaCO}_3$ . When placed in simulated body fluid (SBF) or phosphate buffer solution (PBS), vaterite particles transformed to hydroxyl carbonate apatite [12]. Thus, vaterite has a clear advantage to stimulate bioactivity relative to other synthetic materials and this was specifically important in orthopedics since CaP is present in natural bone [13]. For the case of aragonite, researchers indicated that aragonite particles could dissolve and be replaced with bone tissue [14]. Aragonite particles could be utilized as dual substrates for bone regeneration and drug release and calcite based scaffolds promote adhesion and proliferation of bone cells in vitro [15].

In this study, different anhydrous polymorphs of  $\text{CaCO}_3$  having distinct morphologies were incorporated into SF to fabricate SF/ $\text{CaCO}_3$  composites. To assess CaP mineralization on scaffolds, samples interacted with SBF up to 1 month. This was the first study in literature to focus on the bioactivity enhancement of SF by  $\text{CaCO}_3$  incorporation and to investigate the fabrication and CaP mineral formation on SF/ $\text{CaCO}_3$  composites as per changes in the polymorph of  $\text{CaCO}_3$ .

## 2. Materials and Methods

All chemicals were purchased as analytical grade and used without further purification. Calcium acetate ( $\text{C}_4\text{H}_6\text{O}_4\text{Ca}$ ), sodium bicarbonate ( $\text{NaHCO}_3$ ), ethylene glycol (EG;  $\text{C}_2\text{H}_6\text{O}_2$ ) and sodium hydroxide (NaOH) were purchased from Sigma-Aldrich and water used in these experiments was purified using Millipore Milli-Q purification system.

### 2.1. Synthesis of $\text{CaCO}_3$ Particles

To synthesize  $\text{CaCO}_3$  particles, 4 mL of 1 M  $\text{NaHCO}_3$  and 0.3 M  $\text{C}_4\text{H}_6\text{O}_4\text{Ca}$  solutions were prepared separately and their pH values were adjusted to 12. Afterwards,  $\text{NaHCO}_3$  solution was dispersed in 10, 20, 40 or 50 mL EG to investigate the effect of EG concentration on  $\text{CaCO}_3$  crystallization. Subsequently,  $\text{C}_4\text{H}_6\text{O}_4\text{Ca}$  solution was poured into the EG/ $\text{NaHCO}_3$  solution, followed by stirring for 30 min. Upon precipitation of  $\text{CaCO}_3$  particles, the solution was centrifuged with

water and ethanol three times, respectively, to rinse the particles. The resultant wet powders were dried at  $50^\circ\text{C}$  for 2 h. EG10, EG20, EG40 and EG50 referred to  $\text{CaCO}_3$  particles prepared using  $\text{NaHCO}_3$  solutions containing 10, 20, 40 and 50 mL EG, respectively.

### 2.2. Production of SF/ $\text{CaCO}_3$ Composites

A modified version of Kaplan's protocol was used to prepare SF solutions [16]. SF was extracted from Bombyx Mori cocoons by boiling them in 0.02 M  $\text{Na}_2\text{CO}_3$  solution for 30 min. Afterwards, fibroin was dissolved in 12 M LiBr solution and dialyzed against distilled water for 2 days. Dialyzed fibroin was centrifuged for 30 min, frozen in  $-20^\circ\text{C}$  for 24 h and lyophilized using Christ Alpha 2-4 LD plus freeze dryer to remove water. The lyophilized SF was stored at room temperature. From lyophilized SF, two aqueous SF solutions containing 0.03 g/mL (SF3) and 0.06 g/mL (SF6) fibroin were prepared. Next, 0.025 g of EG10, EG40 and EG50 particles were dispersed manually in 2.5 mL of each SF solution, followed by lyophilizing for 24 h. In this research, six different SF/ $\text{CaCO}_3$  composite samples were prepared, as identified in Table 1.  $\text{CaCO}_3$ -free SF3 and SF6 were used as control samples.

**Table 1.** SF/ $\text{CaCO}_3$  composite formulations.

Composite	SF Concentration, g/mL	Total Volume, mL	$\text{CaCO}_3$ content, wt. %
SF6-EG10			
SF6-EG40	0.06	2.5	15
SF6-EG50			
SF3-EG10			
SF3-EG40	0.03	2.5	25
SF3-EG50			

### 2.3. Characterization

For the structural characterization of  $\text{CaCO}_3$  particles and SF/ $\text{CaCO}_3$  composites, X-ray diffraction (XRD), scanning electron microscopy (SEM) and Fourier-transformed infrared spectroscopy (FTIR) analyses were performed. For polymorphic analysis, Rigaku D-Max-2200 diffractometer with Cu K-alpha radiation was used to scan the samples at  $20$ - $60^\circ$  diffraction angles at a rate of  $2^\circ/\text{min}$ .

For morphological analysis, Nova Nano SEM 430 was used at 20 kV accelerating voltage. Prior to imaging, samples were coated with a thin layer of gold using Quorum SC7640 high-resolution sputter coater. For chemical analysis, FTIR scans were performed to analyze the  $\text{CaCO}_3$  polymorphs chemically. Perkin Elmer Spectrum 100 spectrometer was used in attenuated total reflection (ATR) mode and each sample was scanned in  $4000$ - $400\text{ cm}^{-1}$  range with a

wavenumber resolution of  $4\text{ cm}^{-1}$ . The background spectra were subtracted from the obtained reflectance.

## 2.4. CaP Mineralization

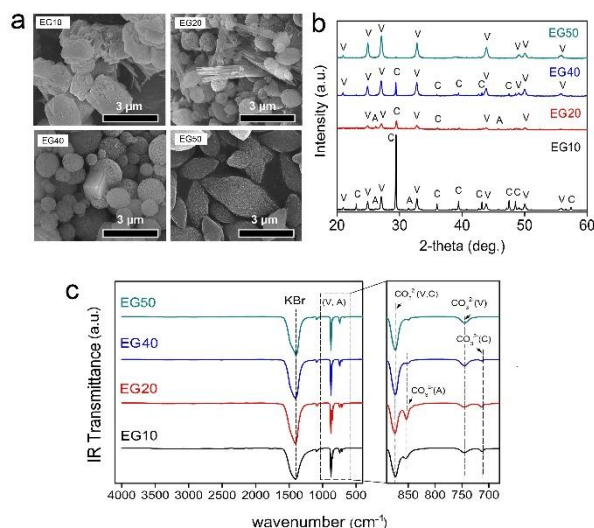
SF, SF3-EG10, SF3-EG40, SF3-EG50, SF6-EG10, SF6-EG40 and SF6-EG50 were immersed in simulated body fluid (SBF) to assess calcium phosphate mineralization. For the preparation of 1xSBF, Kokubo's protocol was used [17]. Briefly, NaCl (8.035 g/L),  $\text{NaHCO}_3$  (0.355 g/L), KCl (0.225 g/L),  $\text{K}_2\text{HPO}_4 \cdot 3\text{H}_2\text{O}$  (0.231 g/L),  $\text{MgCl}_2 \cdot 6\text{H}_2\text{O}$  (0.311 g/L), 1 M HCl (39 ml),  $\text{CaCl}_2$  (0.292 g/L) and  $\text{Na}_2\text{SO}_4$  (0.072 g/L) were dissolved in distilled water, respectively, followed by buffering with Tris (6.118 g/L) and 1 M HCl. Samples were kept in 1xSBF at  $37\text{ }^\circ\text{C}$  for one month. Calcium and phosphate minerals were detected with energy dispersive spectroscopy (EDS) using an EDAX-AMETEK detector attached to SEM.

## 3. Results and Discussion

### 3.1. Synthesis of $\text{CaCO}_3$ particles

SEM images, XRD and FTIR spectra of  $\text{CaCO}_3$  particles were displayed in Figure 1. SEM images (Figure 1a) revealed morphology of the particles. EG-50 samples exhibited ellipsoidal vaterite morphology. As the EG content of the precursor solutions decreased, morphology of the samples was altered. Unlike EG50, EG40 particles were spherical and upon further decreasing the EG content of the precursor to 20 mL, aragonite, which had the appearance of needle-like bundles, began to crystallize and were less apparent at 10 mL EG concentration. SEM micrographs showed EG10 particles also to have rhombohedral (cubic) morphology, which was the hallmark of calcite. XRD spectra confirmed the observations obtained via SEM and identified polymorphs present in the powders (Figure 1b). In the XRD spectra, all peaks were ascribed to the anhydrous forms of  $\text{CaCO}_3$ ; vaterite (V; ICDD: 033-0268), aragonite (A; ICDD: 41-1475) and calcite (C; 005-0586). EG50 particles were phase-pure vaterite, while EG40 had some calcite polymorph. Only peaks belonging to vaterite appeared for EG50 samples, while calcite and vaterite peaks were present in the XRD spectrum of EG40 samples. For the case of EG20 and EG10 samples, XRD spectra had peaks belonging to all three anhydrous polymorphs of  $\text{CaCO}_3$ ; vaterite, calcite and aragonite.

These results were in accord with FTIR spectra of  $\text{CaCO}_3$  particles (Figure 1c) where characteristic absorption bands corresponding to different vibrational modes of  $\text{CO}_3^{2-}$  were investigated. All samples expressed absorption bands of vaterite at  $877\text{ cm}^{-1}$  (v2),  $849\text{ cm}^{-1}$  (v2),  $744\text{ cm}^{-1}$  (v4). Calcite absorption bands at  $877\text{ cm}^{-1}$  (v2),  $712\text{ cm}^{-1}$  (v4) were only present for EG40, EG20 and EG10 samples, while presence of aragonite polymorph was confirmed by absorption bands at  $854\text{ cm}^{-1}$  (v2) and  $700\text{ cm}^{-1}$  (v4) for EG20 and EG10 samples [18].



**Figure 1.** a) SEM images, b) XRD spectra and c) FTIR spectra of  $\text{CaCO}_3$  particles precipitated using 10, 20, 40 and 50 mL EG. “V”, “A” and “C” denote peaks corresponding to vaterite, aragonite and calcite polymorphs, respectively.

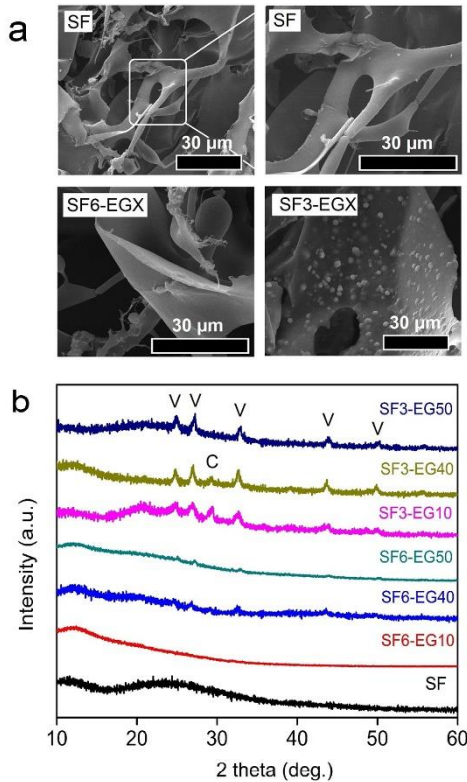
### 3.2. Characterization and bioactivity of SF/ $\text{CaCO}_3$ composites

In this research, three formulations of  $\text{CaCO}_3$  particles (EG10, EG40 and EG50), which contained three  $\text{CaCO}_3$  polymorphs (V, C and A), were incorporated in SF. EG10 particles which consisted of calcite, vaterite and aragonite, EG40 particles which consisted mainly of a mixture of spherical vaterite and calcite, and EG50 particles which were almost-pure ellipsoidal vaterite were used to reinforce SF scaffolds at 15 and 25 wt. %. Lyophilized SF/ $\text{CaCO}_3$  composites are displayed in Figure 2a. Results confirmed porous nature of the SF scaffolds where formation of pores upon lyophilization was not affected by the type of  $\text{CaCO}_3$  particles incorporated into the scaffolds [19]. Images revealed that  $\text{CaCO}_3$  particles could be dispersed as a secondary phase within SF scaffolds. SF6/ $\text{CaCO}_3$  composites (SF6-EGX) had 15%  $\text{CaCO}_3$  particles and had smooth surfaces, yet as  $\text{CaCO}_3$  content increased to 25 % (SF3-EGX),  $\text{CaCO}_3$  particles were revealed on SF surfaces. XRD spectra of these composites were given in Figure 2b.

$\text{CaCO}_3$ -free SF has three characteristic peaks centered at  $8.8^\circ$ ,  $20.4^\circ$  and  $24^\circ$  [20]. The spectra revealed tiny peaks at  $21^\circ$  and  $24^\circ$  for  $\text{CaCO}_3$ -free SF and no crystalline peaks, but a large hump maximized around  $24^\circ$  for the composites, indicating that the crystalline orientation between the beta sheets might have been distorted by the interaction with  $\text{CaCO}_3$  particles. XRD analysis confirmed successful incorporation of  $\text{CaCO}_3$  particles inside the SF matrix. Each intense peak observed in the XRD spectra of the composites could be ascribed to  $\text{CaCO}_3$  particles. Specifically, the most intense peaks of the  $\text{CaCO}_3$  polymorphs manifested themselves at the

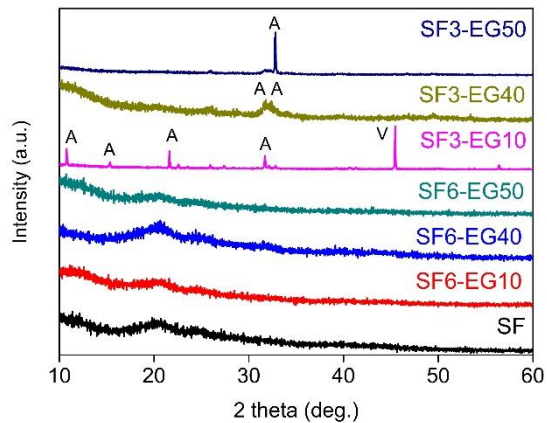


XRD spectra of their corresponding composite formulation. Peaks at  $24.90^\circ$ ,  $27.04^\circ$ ,  $32.65^\circ$ ,  $43.90^\circ$  and  $51.70^\circ$  were present for vaterite and at  $29.45^\circ$  for calcite. Characteristic peaks of vaterite, aragonite and calcite were not evident in SF6 composites – except for the vaterite peak at  $32.65^\circ$  - compared to those of SF3, most probably due to having lower particle concentration inside the composite scaffolds.



**Figure 2.** a) SEM images, b) XRD spectra of SF/CaCO<sub>3</sub> composites (X.10, 40 and 50) compared to CaCO<sub>3</sub>-free SF.

The SF/CaCO<sub>3</sub> composite scaffolds were soaked in 1xSBF for 1 month to assess their bioactivity in biological fluids. XRD scans taken after 1xSBF treatment are displayed in Figure 3 and compared with CaCO<sub>3</sub>-free SF. XRD spectra indicated that CaCO<sub>3</sub>-free SF surfaces preserved their amorphous-like structure; no significant CaP formation was observed. Similarly, 15 % CaCO<sub>3</sub> incorporation did not significantly induce CaP formation (SF6-EGX). However, increasing CaCO<sub>3</sub> content to 25 % (SF3-EGX) led to formation of apatite (A, JCPDS: 74-0566) after soaking in 1xSBF for one month. Apatite mineralization was confirmed also via SEM/EDS analysis. As shown in Figure 2a, CaCO<sub>3</sub>-free scaffold surfaces were smooth before 1xSBF treatment. In Figure 4, the change in scaffold surfaces are shown after 1xSBF treatment for 1 month.

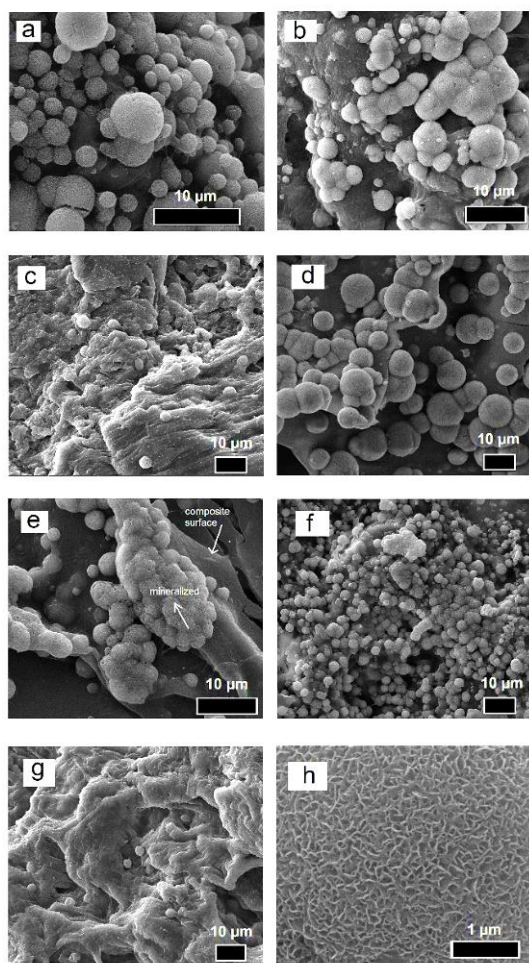


**Figure 3.** XRD spectra of SF/CaCO<sub>3</sub> composites (X.10, 40 and 50) compared to CaCO<sub>3</sub>-free SF after soaking in 1xSBF for 1 month.

SEM images of composite scaffold surfaces in Figure 4a to 4f showed that SF scaffolds stimulated minimal CaP mineralization and incorporation of CaCO<sub>3</sub> significantly influenced mineralization. In Figure 4e, deposition of mineralized material on the smooth scaffold surface is most differentiable. A closer view of the CaCO<sub>3</sub>-free SF indicated mineral deposition on SF surface but not with a definite morphology (Figure 4g).

In the SEM micrographs of SF3/CaCO<sub>3</sub> composites treated with SBF, crystallization of a new morphology (apatite) on each SF/CaCO<sub>3</sub> scaffold surfaces was evident (Figure 4h). EDS spectra exhibited calcium (Ca) and phosphorous (P) peaks for all the SF3 composite formulations, while P peak was not apparent for CaCO<sub>3</sub>-free SF (Table 2). SF6-EG10 and SF6-EG40 composites contained Ca and P, however SF6-EG50 contained only Ca. It can be speculated that Ca present in the CaCO<sub>3</sub> partially contributed to the intensity of calcium peak obtained with the EDS analysis, yet phosphorous was only present at the CaP mineral forming on the scaffolds.

It could be speculated that these particles acted as templates for CaP nucleation and growth, and thus promoted mineralization via favoring an alternate route. The other elements observed in the EDS spectra (Na, Mg and Cl) were most probably the ions adsorbed on the composite surfaces from the unreacted Kokubo's solution.



**Figure 4.** SEM images of a) SF6-EG10, b) SF6-EG40, c) SF6-EG50, d) SF3-EG10, e) SF3-EG40, f) SF3-EG50, g) SF, h) detailed, representative image of mineralized apatite morphology on composites after immersion in 1xSBF for one month.

**Table 2.** Elemental compositions of the composite surfaces (average of 3 measurements) compared to SF after immersion in 1xSBF for one month.

Composite	Na	Mg	P	Cl	Ca
SF6-EG10	4.81	4.48	35.68	6.62	48.41
SF6-EG40	2.01	-	36.22	1.62	59.72
SF6-EG50	29.42	5.44	-	57.34	4.11
SF3-EG10	7.04	2.39	31.52	8.66	50.39
SF3-EG40	7.87	-	36.08	5.23	50.82
SF3-EG50	3.19	2.89	38.39	-	54.74
SF	24.79	12.66	-	33.69	19.32

#### 4. Conclusions

In this work,  $\text{CaCO}_3$  polymorphs were synthesized at different EG concentrations. Three  $\text{CaCO}_3$  formulations were chosen and SF/ $\text{CaCO}_3$  scaffolds having different SF concentrations were prepared. Composites were

characterized and tested for CaP mineralization upon interaction with 1xSBF.

XRD, SEM and FTIR analyses of  $\text{CaCO}_3$  particles indicated that EG concentration directly influenced the polymorphic growth in solution. As the EG concentration decreased, phase-pure vaterite (EG50) transformed to vaterite-aragonite-calcite mixtures, in which almost phase-pure calcite could be obtained at the lowest EG concentration (EG10).

In the second part,  $\text{CaCO}_3$  incorporated SF scaffolds induced higher mineralization compared to SF-only scaffolds in 1xSBF at 1 month. By the XRD and SEM-EDS analyses, it was shown that scaffolds containing 25%  $\text{CaCO}_3$  (EG50) displayed the highest CaP mineralization on the particle surfaces (SF3-EG50). Vaterite polymorph was the most effective  $\text{CaCO}_3$  form for the in-situ formation of CaP in SF matrix. This was a promising finding for the use of SF/ $\text{CaCO}_3$  scaffolds in the biomaterials field and open the way for biological tests to assess the biocompatibility of these scaffolds. For future work, authors continue their studies to further assess the structural interaction at the SF- $\text{CaCO}_3$  interface and enhance the mechanical properties of the composites for use in orthopedic applications.

#### Author's Contributions

**Derya Kapusuz:** Contributed equally in drafting and writing the manuscript, performed the experiments, analyzed results.

**Batur Ercan:** Contributed equally in drafting and writing the manuscript, helped interpret the results.

#### Ethics

There are no ethical issues after the publication of this manuscript.

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#### References

- Murphy, A, R, Kaplan, D, L. 2010. Biomedical applications of chemically-modified silk fibroin. *Journal of Materials Chemistry*; 19(36): 6443-6450.
- Wang, L, Chunxiang, L, Zhang, B, Zhao, B, Wu, F, Guan, S. 2014. Fabrication and characterization of flexible silk fibroin films reinforced with graphene oxide for biomedical applications. *RSC Advances*; 4: 40312-40320.
- Pan, C, Xie, Q, Hu, Z, Yang, M, Zhu, L. 2015. Mechanical and biological properties of silk fibroin/carbon nanotube nanocomposites films. *Fibers and Polymers*; 16(8): 1781-1787.
- Johari, N, Hosseini, M, Taromi, N, Arasteh, S, Kazemnejad, S, Samadikuchaksaraci, A. 2018. Evaluation of bioactivity and biocompatibility of silk fibroin/ $\text{TiO}_2$  nanocomposite. *Journal of Medical and Biological Engineering*; 38: 99-105.
- Martin, R, B. 1999. Bone as a ceramic composite material. *Materials Science Forum*; 293: 5-16.

6. Li, C, Jin, H, J, Botsaris, G, D, Kaplan, D, L. 2005. Silk apatite composites from electrospun fibers. *Journal of Materials Research*; 20(12): 3374e84.
7. Bhumiratana, S, Grayson, W, L, Castaneda, A, Rockwood, D, N, Gil, E, S, Kaplan, D, L, Novakovic, G. 2011. Nucleation and growth of mineralized bone matrix on silk-hydroxyapatite composite scaffolds. *Biomaterials*; 32: 2812-20.
8. Eliaz, N, Metoki, N. 2017. Calcium Phosphate Bioceramics: A Review of Their History, Structure, Properties, Coating Technologies and Biomedical Applications. *Materials (Basel)*; 10: 334.
9. Farokhi, M, Mottaghitalab, F, Samani, S, Shokrgozar, M, A, Kundu, S, C, Reis, R, L, Fatahi, Y, Kaplan, D, L. 2018. Silk fibroin/hydroxyapatite composites for bone tissue Engineering. *Biotechnology Advances*; 36(1): 68-91.
10. Wang, J, Yu, F, Qu, L, Meng, X, Wen, G. 2010. Study of synthesis of nano-hydroxyapatite using a silk fibroin template. *Biomedical Materials*; 5: 041002-041007.
11. Marelli, B, Ghezzi, C, E, Alessandrino, A, Barralet, J, E, Freddi, G, Nazhat, S, N. 2012. Silk fibroin derived polypeptide-induced biomineralization of collagen. *Biomaterials*; 33: 102-108.
12. Schröder, R, Besch, L, Pohlitz, H, Panthöfer, M, Roth, W, Frey, H, Tremel, W, Unger, R, E. 2018. Particles of vaterite, a metastable  $\text{CaCO}_3$  polymorph, exhibit high biocompatibility for human osteoblasts and endothelial cells and may serve as a biomaterial for rapid bone regeneration. *Journal of Tissue Engineering and Regenerative Medicine*; 12: 1754-1768.
13. Trushina, D, B, Bukreeva, T, V, Kovalchuk, M, V, Antipina, M, N. 2014.  $\text{CaCO}_3$  vaterite microparticles for biomedical and personal care applications. *Materials Science and Engineering C*; 45: 644-658.
14. Akilal, N, Lemarie, F, Bercu, N, B, Sayen, S, Gangloff, S, C, Khelfaoui, Y, Rammal, H, Kedjoudj, H. 2019. Cowries derived aragonite as raw Materials for bone regenerative medicine. *Materials Science and Engineering C*; 94, 894-900.
15. Wolsetsadik, A, D, Sharma, S, K, Khapli, S, Jagannathan, R. 2017. Hierarchically porous calcium carbonate scaffolds for bone tissue engineering. *ACS Biomaterials Science and Engineering*; 3: 2457-2469.
16. Rockwood, D, N, Preda, R, C, Yücel, T, Wang, X, Lovett, M, L, Kaplan, D, L. 2011. Materials fabrication from Bombyx mori silk fibroin, *Nature Protocols*; 6(10): 1612-1631.
17. Kokubo, T, Takadama, H. 2006. How useful is SBF in predicting in vivo bone bioactivity? *Biomaterials*; 27(15): 2907-2915.
18. Ševčík, R, Pérez-Estébanez, M, Viani, A, Šašek, P, Mácová, P. 2015. Characterization of vaterite synthesized at various temperatures and stirring velocities without use of additives. *Powder Technology*; 284: 265-271.
19. Lu, Y, Zhang, S, Liu, X, Ye, S, Zhou, X, Huang, Q, Ren, L. 2017. Silk/agarose scaffolds with tunable properties via SDS assisted rapid gelation. *RSC Advances*; 7: 21740-21748.
20. Asakura, T, Kuzuhara, A, Tabeta, R, Saito, H. 1985. Conformation Characterization of Bombyx mori silk fibroin in the solid state by high frequency  $^{13}\text{C}$  cross-polarization-magic Angle spinning NMR, X-ray diffraction, and infrared spectroscopy. *Macromolecules*; 18: 1841-1845.