# THE UNIVERSITY OF RHODE ISLAND

University of Rhode Island DigitalCommons@URI

Graduate School of Oceanography Faculty Publications

Graduate School of Oceanography

2021

# The Air that we Breathe: Neutral and volatile PFAS in Indoor Air

Maya E. Morales-McDevitt University of Rhode Island

Jitka Becanova University of Rhode Island

Arlene Blum

Thomas A. Bruton

Simon Vojta University of Rhode Island

Follow this and additional works at: https://digitalcommons.uri.edu/gsofacpubs

The University of Rhode Island Faculty have made this article openly available. Please let us know how Open Access to this research benefits you.

This is a pre-publication author manuscript of the final, published article.

Terms of Use

This article is made available under the terms and conditions applicable towards Open Access Policy Articles, as set forth in our Terms of Use.

#### **Citation/Publisher Attribution**

Morales-McDevitt, Maya; Becanova, Jitka; Blum, Arlene; Bruton, Thomas; Vojta, Simon; Woodward, Melissa; Lohmann, Rainer. The Air that we Breathe: Neutral and volatile PFAS in Indoor Air. *Environmental Science & Technology Letters*, 2021, *accepted*. Doi:10.1021/acs.estlett.1c00481. Available at: https://doi.org/10.1021/acs.estlett.1c00481

This Article is brought to you for free and open access by the Graduate School of Oceanography at DigitalCommons@URI. It has been accepted for inclusion in Graduate School of Oceanography Faculty Publications by an authorized administrator of DigitalCommons@URI. For more information, please contact digitalcommons@etal.uri.edu.

#### Authors

Maya E. Morales-McDevitt, Jitka Becanova, Arlene Blum, Thomas A. Bruton, Simon Vojta, Melissa Woodward, and Rainer Lohmann

The Air that we Breathe: Neutral and volatile PFAS in Indoor Air 1 2 Maya E. Morales-McDevitt<sup>1</sup>; Jitka Becanova<sup>1</sup>, Arlene Blum<sup>2</sup>, Thomas A. Bruton<sup>2</sup>, Simon 3 Vojta<sup>1</sup>, Melissa Woodward<sup>1</sup>, Rainer Lohmann<sup>1,\*</sup> 4 <sup>1</sup> Graduate School of Oceanography, University of Rhode Island, 215 South Ferry Rd, 5 6 Narragansett, 02882 RI, USA 7 <sup>2</sup> Green Science Policy Institute, Berkeley, 94709 CA, USA 8 9 \* Corresponding author: rlohmann@uri.edu; Tel (1) 401-874-6612 10

### 11 Abstract

12 Sources of exposure to per- and polyfluorinated alkyl substances (PFAS) include food, water, and given that humans spend typically 90% of our time indoors, air and dust. 13 14 Quantifying PFAS prevalent indoors, such as neutral, volatile PFAS, and estimating their 15 exposure risk to humans is thus important. To accurately measure these compounds indoors, 16 polyethylene (PE) sheets were employed and validated as passive detection tools, and analyzed 17 by gas chromatography-mass spectrometry. Air concentrations were compared to dust and 18 carpet concentrations reported elsewhere. Partitioning between PE sheets of different 19 thicknesses suggested that interactions of the PEs with the compounds are occurring by 20 absorption. Volatile PFAS, specifically fluorotelomer alcohols (FTOHs), were ubiquitous in 21 indoor environments. For example, in carpeted Californian kindergarten classrooms, 6:2 FTOH 22 dominated with concentrations ranging from 9-600 ng m<sup>-3</sup>, followed by 8:2 FTOH. Concentrations of volatile PFAS from air, carpet and dust were closely related to each other, 23 24 indicating that carpets and dust are major sources of FTOHs in air. Nonetheless, air posed the 25 largest exposure risk of FTOHs and biotransformed perfluorinated alkyl acids (PFAA) in

26	young children.	This	research	highlights	inhalation	of	indoor	air	as	an	important	exposure
27	pathway and the	need	for furthe	er reduction	n of precurs	sors	s to PFA	AA.				

28

29 Passive sampling, Polyethylene sheets, PFAS precursors, gas-phase, Carpet, Dust, Risk
30 Assessment

31

Polyethylene (PE) sheets are effective passive samplers for PFAA precursors which are
ubiquitous in indoor air and dominate indoor exposure.

34

# 35 Introduction

36 Human exposure to fluorotelomer alcohols (FTOHs), perfluorooctane sulfonamides (FOSAs) and perfluorooctane sulfonamidoethanols (FOSEs) and other precursors to 37 perfluoroalkyl acids (PFAA) comes primarily from consumer and industrial products readily 38 available in people's homes.<sup>1-4</sup> FTOHs were the dominant polyfluorinated compounds in 39 indoor air <sup>5</sup> where ~60% of detected per- and polyfluorinated alkyl substances (PFAS) were 40 associated with the particle phase.<sup>2</sup> Since most people spend more than 90% of their time 41 indoors,<sup>2</sup> indoor air and dust are important uptake pathways for human PFAS exposure<sup>6</sup> in 42 addition to the widely recognized exposure sources of diet and water.<sup>7–9</sup> Indeed, correlations 43 44 between elevated indoor air exposure to precursors and increased PFAS serum concentrations have been reported.<sup>10,11</sup> 45

The use of passive sampling, which can measure the concentration of freely dissolved or gas-phase trace organic contaminants, has been widely accepted as an effective detection tool.<sup>12,13</sup> Single-phase polymers, such as polyethylene (PE) sheets, have been able to detect a wide range of non-polar and moderately polar contaminants in the gas phase or dissolved in 50 water.<sup>14,15</sup> In addition, PE sheets are inexpensive, easy to handle, and can be easily transported 51 and deployed.<sup>13</sup> Recently, neutral PFAS were successfully measured in outdoor air and water 52 using PE sheets.<sup>16</sup> However, the partitioning of neutral PFAS into or onto the PE sheets indoors 53 is not yet fully understood.

To further assess the role that indoor environments play as an exposure source of airborne PFAS in gas-phase and dust, the main objectives of this research were to (i) derive indoor PE-air partitioning coefficients (K<sub>PE-air</sub>); (ii) compare the volatile PFAS composition in different indoor environments using PE sheets as passive samplers; (iii) evaluate the air-dust partitioning of PFAS in carpeted kindergarten classrooms, and (iv) estimate daily intake (EDI) in children 2-6 years of age.

#### 60 Materials and Methods

Sampling of neutral PFAS was performed in carpeted kindergarten classrooms,
residences, an outdoor gear and apparel store in northern California; university offices,
classrooms, laboratories, and a carpet store in southern Rhode Island between 2018 and 2020.
A total of 90 PE sheets were deployed in the indoor locations, in addition to eight radiello
samplers (Sigma Aldrich) with precleaned XAD-4 as sorbent used for active sampling.

Two types of precleaned PE passive samplers differentiated by thickness (25  $\mu$ m and 50  $\mu$ m) were deployed for 14 days (validation study), 21 days (kinetic study), and 28 days (measurements). Active sampling was performed on days 1, 7 and 14 where the radiello samplers were attached to a QuickTake 30 SKC Pump at a constant flow of 5 L min<sup>-1</sup> for 240 minutes. All samples were kept in a freezer at -20°C until extraction (for details, see SI). 71 Instrumental analysis

Samples were analyzed for nine neutral PFAS on an Agilent 7890B gas chromatograph
 coupled to an Agilent 5977A mass selective detector (MSD) device operating in positive
 chemical ionization mode using selected ion monitoring (for details, see SI).

75 Data interpretation

The partitioning constants of neutral PFAS between PE and air (K<sub>PE-air</sub>) were derived
 in the validation study as:

78  $K_{PE-air} = \frac{c_{PE}}{c_{air}}$ (1),

79 where  $C_{PE}$  is the concentration in PE sheets (ng g<sup>-1</sup><sub>PE</sub>), and

80  $C_{air}$  is the gas-phase concentration (ng m<sup>-3</sup>).

Active sampling was used in the  $K_{PE-air}$  validation study only. For all other campaigns,  $C_{air}$  was calculated using equation (1). Partitioning within the PE sheets was derived as the ratio of the 25 µm passive sampler ( $C_{25}$ ) to the 50 µm passive sampler ( $C_{50}$ ) amounts at equilibrium (for details, see SI).

#### 85 Daily intake

The total estimated daily intake (EDI) of neutral PFAS via air and dust was calculated from PFAS concentrations measured here, and dust concentrations reported elsewhere<sup>17</sup> based on established methods <sup>18,19</sup> (for details, see SI).

89 QA/QC

Field blanks, matrix spikes, matrix blanks, and field duplicate samples were included with each sample batch. Matrix spikes were prepared by spiking 50  $\mu$ L of an 80 pg/ $\mu$ L native standard solution and 50  $\mu$ L of an 80 pg/ $\mu$ L mass labelled standard solution into a clean (unused and never removed from the laboratory) sampler. Method detection limits (MDL) were calculated as the blank average plus three times the standard deviation; however, when a compound was not detected in the blanks, instrumental limits of detection (ILOD) were used. Only values above limits of quantitation (LOQ) were reported (for details, see SI, Table S2).
Recoveries of the matrix spikes ranged between 81% (±35) to 111% (±19) for all compounds
(for details, see SI, Table S2).

## 99 Results and Discussion

#### 100 **PE-air Partitioning Constants**

101 Results from the kinetic study showed that 6:2 FTOH and 8:2 FTOH reached 102 equilibrium after 14 days (see SI Figure S1). Log  $K_{PE-air}$  values were approximately 4 -5 for 103 the FTOHs, ~ 5 for 8:2 FTAcr, and increased with molecular weight. Although 10:2 FTAcr, 104 FOSAs and FOSEs were detected by PE sheets, none were detected by active sampling; 105 calculating their equilibrium partitioning constant was not possible (see SI Table S6). There 106 were only minor differences between the 25  $\mu$ m and 50  $\mu$ m K<sub>PE-air</sub> results, indicating good 107 reproducibility of PE sheets as passive samplers.

Mean log  $K_{PE-air}$  values from the FTOHs of this study were approximately three log units lower than those reported for outdoors (see SI Table S3)<sup>16</sup> where break-through and environmental factors could have affected the partitioning of the compounds. Missing  $K_{PE-air}$ values were derived based on a correlation between previously reported<sup>16</sup> and currently measured PE-air partitioning constants. Further studies are needed to corroborate the partitioning coefficients of the FOSAs and FOSEs.

114

Compound	Molecular weight (g mol <sup>-1</sup> )	Mean log K <sub>PE-air 25</sub> (this study)	Mean log K <sub>PE-air 50</sub> (this study)
6:2 FTOH	364.1	$4.4 \pm 0.1$	$4.3\pm0.0$
8:2 FTOH	464.1	$4.3 \pm 0.1$	$4.5\pm0.0$
10:2 FTOH	564.1	5.0	$5.0\pm0.0$
8:2 FTAcr	518.1	$4.9\pm0.4$	$5.0 \pm 0.2$

115 Table 1. Indoor log K<sub>PE-air</sub> values from the validation study for 25 and 50 um PE sheets.

10:2 FTAcr	618.1	5.2*	5.3*
MeFOSA	527.2	5.1*	5.2*
EtFOSA	513.1	ND	ND
MeFOSE	571.2	5.2*	5.3*
EtFOSE	557.2	5.2*	5.2*

116 ND=not detected,  $K_{PE-air}$  from this study was estimated based on a correlation between those measured here and those 117 reported by Dixon-Anderson and Lohmann, (2018)<sup>16</sup>. Estimated K<sub>PE-air</sub> = 0.44 x K<sub>PE-air</sub>, measured (Dixon-Anderson and Lohmann,

118 2018) + 1.30 (RSQ<sub>PE50</sub>=0.67).

119

#### 120 **PE-air Partitioning ratios**

121 The partitioning ratios of the weight-normalized neutral PFAS between 25 and 50  $\mu$ m 122 thick PE sheets were ~1 (see SI Figure S2) implying absorption as the mechanism of 123 partitioning. The greater mass of the 50  $\mu$ m PE sheets for the same size resulted in easier 124 detection and is thus preferable for future studies.



125

126 Figure 1. Indoor air concentrations measured at California Kindergarten classrooms

127 and an outdoor clothing store, and university classrooms, offices and laboratories, and a

128 **carpet store in southern Rhode Island.** H: home; KG: kindergarten classrooms; Lab: laboratory;

129 Off: office; Elev: elevator; Clrm: classroom; Ctst: carpet store; Strm: storage room. Numbers (i.e.

130 KG7) are indicative of separate/individual samples. Off3 and CtSt2 have concentrations above 1000  $131 \text{ ng m}^{-3}$ .

#### 132 Neutral PFAS Indoor Air Concentrations

133 Indoor air concentrations were derived from the PEs for neutral PFAS in (1) California 134 Kindergarten classrooms (SI Table S4); (2) offices, classrooms and laboratories at a university, 135 and a nearby carpet store in southern Rhode Island (SI Table S5); and (3) a storage room at an 136 outdoor clothing store in California (SI Table S6), see Figure 1. Neutral PFAS were present at all locations, dominated by FTOHs, in-line with previous results.<sup>5,20,21</sup> PFAS profiles and 137 concentrations varied between locations, though, likely driven by the different PFAS-138 139 containing products present. These results indicated that PE-sheets can be used to determine 140 differences in PFAS profiles and concentrations in various indoor air settings.

In the California kindergarten classrooms, 6:2 FTOH dominated with concentrations
ranging from 10-600 ng m<sup>-3</sup> (accounting for 29-96 % of sum of nine PFAS), followed by 8:2
FTOH (2-160 ng m<sup>-3</sup>, 3-54% of total PFAS) (Figure 1; SI Table S2). In three kindergarten
classrooms (KG2, KG3, and KG5), concentrations of 8:2 FTOH exceeded those of 6:2 FTOH.
In all kindergarten classrooms, EtFOSE was present at low concentrations, while MeFOSE was
below method detection limits (MDLs) (Figure 1; SI Table S2). EtFOSA, 8:2 FTAcr, and 10:2
FTAcr were not detected (SI Table 3).

When detected, 6:2 FTOH (with detection frequency of 83%, and ranging from < MDL  $-1900 \text{ ng m}^{-3}$ ), and 8:2 FTOH (17%, < MDL-270 ng m}^{-3}) also dominated total PFAS in the university rooms (Figure 1; SI Table S5). FTOHs were detected only in carpeted rooms and in the analytical laboratory (SI Table S5). The detection of 10:2 FTOH was sporadic, with concentrations up to 33 ng m}^{-3} (Figure 1; SI Table S5). MeFOSA, EtFOSE, and MeFOSE were at or < MDL at all sites, while EtFOSA and FTAcr were rarely above MDLs (SI Table S5).

Volatile PFAS were present in all replicates from the outdoor clothing store (Figure 1).
FTOHs were the most abundant and dominant group; consistent with previous studies on the

156 composition of PFAS in various indoor environments.<sup>5,20,21</sup> The most abundant compound was 157 8:2 FTOH, with an average concentration of ~200 ng m<sup>-3</sup>, followed by 6:2 FTOH and 10:2 158 FTOH with average concentrations of 70 ng m<sup>-3</sup> and 30 ng m<sup>-3</sup> respectively (Figure 1). The 159 dominance of 8:2 FTOH is concerning since this and other longer-chain PFAS have been 160 phased out by PFAS producers in the United States, European Union, and Japan<sup>22</sup>. These 161 results show that these compounds are still being used for textiles, and possibly point to textile 162 imports from other countries where PFAS are poorly regulated.<sup>23</sup>

Fraser et al. (2011)<sup>11</sup> reported concentrations of FTOHs ranging from <MDL to 11 ng 163 m<sup>-3</sup> (6:2 FTOH), 0.3 - 70 ng m<sup>-3</sup> (8:2 FTOH), and 0.14 - 12 ng m<sup>-3</sup> (10:2 FTOH) in multiple 164 office spaces in Boston, Massachusetts, similar to results report here and in other studies.<sup>5,21</sup> A 165 166 study in Ottawa, Canada, in 2005 reported concentrations of MeFOSE, EtFOSE and EtFOSA in indoor air of  $\sim 7$  ng m<sup>-3</sup>, 2 ng m<sup>-3</sup> and 0.1 ng m<sup>-3</sup>, respectively<sup>2</sup>, and even lower in Vancouver, 167 Canada in 2011, at 0.4 ng m<sup>-3</sup>, 0.06 ng m<sup>-3</sup>, 0.03 ng m<sup>-3</sup>, and 0.02 ng m<sup>-3</sup> respectively.<sup>21</sup> In the 168 present study, although present in many locations, FOSEs rarely exceeded concentrations of 169 0.001 ng m<sup>-3</sup>. FOSAs were detected even fewer times. The difference in concentrations of the 170 FOSAs and FOSEs in different locations across North America could reflect geographic 171 172 differences of indoor sources. Additionally, the difference between older and newer data could point to the phase out of PFOS-based chemicals, including FOSAs and FOSEs since 2002, 173 whereas the use of replacement FTOHs in North America has increased since 2000.<sup>21,24,25</sup> 174

175

#### 176 Air-dust-carpet partitioning

177 Concentrations of neutral PFAS in dust and carpet of the same kindergarten classrooms 178 were measured by Wu *et al.*  $(2020)^{17}$  (see SI Table S7). Strong correlations (RSQ>0.7, P $\leq$ 0.05) 179 were observed between different FTOHs in air-dust, and air-carpet (and dust-carpet from Wu *et al.* (2020)<sup>17</sup>, see SI Table S8), except for 6:2 FTOH in air-carpet. On the other hand, FOSEs
were not strongly correlated in air, dust or carpet.

182 Distribution of PFAS between indoor air and floor dust were reported to be controlled by partitioning between the gas phase and PFASs sorbed to the organic phases in the dust.<sup>26</sup> 183 184 Our results corroborated that neutral PFAS were present in air and partitioned to dust. Given 185 that the origin of volatile PFAS in air in the (carpet-free) outdoor clothing storage room was 186 likely to be textiles, it is possible that multiple products in the kindergarten classrooms were in 187 fact the source of these compounds that also partitioned into carpet and dust. Previous work 188 demonstrated that FTOHs, FOSAs, and FOSEs degrade in the atmosphere into more stable PFAA.<sup>27,28</sup> Significant associations between precursors in air and PFCAs in dust have been 189 reported (e.g., 6:2 FTOH and PFHxA).<sup>29</sup> Similarly, significant associations were observed 190 between FOSAs/FOSEs in air and PFOS and PFDS in house dust.<sup>28</sup> 191

In contrast to FTOHs, there were no significant correlations between the 192 193 FOSAs/FOSEs in air, dust and carpet from this study, suggesting that the sources of 194 FOSAs/FOSEs were different and likely not linked to carpets or textiles. A previous study did 195 not find significant correlations between the FOSEs in kindergarten classrooms either, but did however find strong associations in offices<sup>30</sup>, implying that there were common sources of 196 197 these sulfonamidoethanols in items associated with office spaces that perhaps were not usually 198 found in kindergarten classrooms. Additionally, as previously mentioned, the production of FOSAs/FOSEs has been largely phased-out of production since 2002<sup>24</sup>, and thus their low 199 200 concentration or absence is expected<sup>31</sup>.

201

#### 202 Estimated daily intake of volatile PFAS through air and dust

203 To assess the relevance of volatile and neutral PFAS in indoor air for children aged two 204 to six years old, the estimated daily intake (EDI) was calculated (SI Table S8) for three 205 exposure estimates (low, medium, high, see SI). Biotransformation constants for each 206 compound were used to calculate their contribution to the  $\Sigma$ PFAA intake (SI Table S8).

Total EDI (SI Table S9) was 1.5 ng kg<sup>-1</sup> bw day<sup>-1</sup> for low exposure, 14 ng kg<sup>-1</sup> bw day<sup>-</sup> 207 <sup>1</sup> for intermediate exposure, and 150 ng kg<sup>-1</sup> bw day<sup>-1</sup> for high exposure. Compounds that were 208 209 regularly detected in both air and dust were 6:2 FTOH, 8:2 FTOH, and, to a lesser extent, 10:2 210 FTOH; while MeFOSE and EtFOSE appeared to have significantly larger contributions in dust 211 than air (Figure 2, SI Table S9). Volatile and neutral PFAS measured in air contributed 4.9-62 % to  $\Sigma$ PFAA intake, while ionic PFAS measured in dust contributed 34-95 % (SI Table S9). 212 These results are similar to other studies that found precursors contributing 41–68 % to  $\Sigma$ PFOS 213 uptake via all investigated exposure pathways, <sup>32</sup> and precursors responsible for 90 % to the 214  $\Sigma$ PFOS intake in air (Figure 2).<sup>26,33</sup> Our results imply that air inhalation was a major exposure 215 216 pathway for FTOHs, while dust ingestion was dominant for FOSEs in children, similar to prior results.30 217





Figure 2. Percent of volatile and neutral PFAS (top panel) and indirect or
biotransformed PFAA (bottom panel) intake via air inhalation (pink) and dust ingestion
(blue) for children at ages 2 through 6. Bars represent the relative contribution of

individual precursors to total PFAS (left axis); bars are differentiated by color for both

224 matrices. Lines represent the percent estimated contribution for each compound in air and 225 dust (right axis). MeFOSE was detected at low concentrations in dust and <MDL in air.

226

Given the potential for precursors to be biotransformed into more stable PFAA, estimations of PFAA indirect exposure were also calculated as 1.2 ng kg<sup>-1</sup> bw day<sup>-1</sup>, 75 ng kg<sup>-1</sup> bw day<sup>-1</sup>, 2800 ng kg<sup>-1</sup> bw day<sup>-1</sup> for the low, intermediate, and high exposure scenario respectively (SI Table S9). The major contributors to indirect PFAA exposure were 6:2 FTOH and 8:2 FTOH in air, and MeFOSE in dust (Figure 2). This study demonstrated that volatile neutral PFAS, such as FTOHs, are major contributors to exposure in air.

218

# 233 Supporting Information

- 234 The Supporting Information contains additional details on the analysis, data interpretation
- and EDI calculation, and is available free of charge at <u>https://pubs.acs.org/doi/XXX</u>.
- 236

# 237 Acknowledgements

- 238 The authors acknowledge funding from NIEHS (P42ES027706). The analysis was conducted
- at a Rhode Island NSF EPSCoR research facility, Molecular Characterization Facility,
- supported in part by the EPSCoR Cooperative Agreement # OIA-1655221. We thank Marta
- 241 Venier (Indiana U) for comments on a previous version of this manuscript.
- 242

# 243 Conflict of interest

244 The authors declare no competing financial interests.

245

# 246 References247

248 (1)Prevedouros, K.; Cousins, I. T.; Buck, R.; Korzeniowski, S. H. Critical Review 249 Sources, Fate and Transport of Perfluorocarboxylates. Environ. Sci. Technol. 2006. 250 https://doi.org/10.1021/es0512475. Shoeib, M.; Harner, T.; Wilford, B. H.; Jones, K. C.; Zhu, J. Perfluorinated 251 (2)252 Sulfonamides in Indoor and Outdoor Air and Indoor Dust: Occurrence, Partitioning, 253 and Human Exposure. Environ. Sci. Technol. 2005. https://doi.org/10.1021/es048340y. 254 Langer, V.; Dreyer, A.; Ebinghaus, R. Polyfluorinated Compounds in Residential and (3) 255 Nonresidential Indoor Air. Environ. Sci. Technol. 2010. 256 https://doi.org/10.1021/es102384z. Gremmel, C.; Fr, T.; Omel, €; Knepper, T. P. Systematic Determination of 257 (4) 258 Perfluoroalkyl and Polyfluoroalkyl Substances (PFASs) in Outdoor Jackets. 2016. 259 https://doi.org/10.1016/j.chemosphere.2016.06.043. 260 Schlummer, M.; Gruber, L.; Fiedler, D.; Kizlauskas, M.; Müller, J. Detection of (5) 261 Fluorotelomer Alcohols in Indoor Environments and Their Relevance for Human 262 Exposure. Environ. Int. 2013, 57-58, 42-49. 263 https://doi.org/10.1016/j.envint.2013.03.010. 264 Ericson Jogsten, I.; Nadal, M.; Van Bavel, B.; Lindström, G.; Domingo, J. L. Per- and (6) 265 Polyfluorinated Compounds (PFCs) in House Dust and Indoor Air in Catalonia, Spain: Implications for Human Exposure. Environ. Int. 2012, 39, 172–180. 266 267 https://doi.org/10.1016/j.envint.2011.09.004. 268 Sunderland, E. M.; Hu, X. C.; Dassuncao, C.; Tokranov, A. K.; Wagner, C. C.; Allen, (7)269 J. G. A Review of the Pathways of Human Exposure to Poly- and Perfluoroalkyl 270 Substances (PFASs) and Present Understanding of Health Effects. J. Expo. Sci. 271 Environ. Epidemiol. 2019, 29 (2), 131–147. https://doi.org/10.1038/s41370-018-0094272

1.

- 273 (8) EFSA Panel on Contaminants in the Food Chain (EFSA CONTAM Panel); Schrenk,
- D.; Bignami, M.; Bodin, L.; Chipman, J. K.; del Mazo, J.; Grasl-Kraupp, B.;
- 275 Hogstrand, C.; Hoogenboom, L. (Ron); Leblanc, J.-C.; Nebbia, C.S.; Nielsen, E.;
- 276 Ntzani, E.; Petersen, A.; Sand, S.; Vleminckx, C.; Wallace, H.; Barregård, L.;
- 277 Ceccatelli, S.; Cravedi, J.P.; Halldorsson, T.I.; et al. Risk to Human Health Related to
- the Presence of Perfluoroalkyl Substances in Food. *EFSA J.* **2020**, *18* (9), e06223.
- 279 https://doi.org/https://doi.org/10.2903/j.efsa.2020.6223.
- 280 (9) ATSDR. *Toxicological Profile for Perfluoroalkyls*. (*Draft for Public Comment*);
  281 Atlanta, GA, 2018.
- 282 (10) Makey, C. M.; Webster, T. F.; Martin, J. W.; Shoeib, M.; Harner, T.; Dix-cooper, L.;
- Webster, G. M. Airborne Precursors Predict Maternal Serum Perfluoroalkyl Acid
  Concentrations. *Env. Sci Technol* 2017, *51*, 7697–7675.
- 285 https://doi.org/10.1021/acs.est.7b00615.
- 286 (11) Fraser, A. J.; Webster, T. F.; Watkins, D. J.; Nelson, J. W.; Stapleton, H. M.; Calafat,
- A. M.; Kato, K.; Shoeib, M.; Vieira, V. M.; McClean, M. D. Polyfluorinated
- 288 Compounds in Serum Linked to Indoor Air in Office Environments. *Env. Sci Technol*
- 289 **2011**, No. 46, 1209–1215. https://doi.org/10.1021/es2038257.
- 290 (12) Lohmann, R. Critical Review of Low-Density Polyethylene's Partitioning and
- 291 Diffusion Coefficients for Trace Organic Contaminants and Implications for Its Use
- 292 As a Passive Sampler. **2011**. https://doi.org/10.1021/es202702y.
- 293 (13) Lohmann, R.; Booij, K.; Smedes, F.; Vrana, B. POPs Workshop, Ten Years after the
- 294 Signature of the Stockholm Convention. Use of Passive Sampling Devices for
- 295 Monitoring and Compliance Checking of POP Concentrations in Water. *Environ. Sci.*
- 296 *Pollut. Res.* **2012**, No. 19, 1885–1895. https://doi.org/10.1007/s11356-012-0748-9.

- 297 (14) Booij, K.; Sleiderink, H. M.; Smedes, F. Calibrating the Uptake Kinetics of
- 298 Semipermeable Membrane Devices Using Exposure Standards. *Environ. Toxicol.*
- 299 *Chem.* **1998**, *17* (7), 1236–1245. https://doi.org/10.1897/1551-5028.
- 300 (15) Adams, R.; Lohmann, R.; Fernandez, L.; MacFarlane, J.; Gschwend, P. Polyethylene
- 301 Devices: Passive Samplers for Measuring Dissolved Hydrophobic Organic
- 302 Compounds in Aquatic Environments. *Environ. Sci. Technol* 2007.
- 303 https://doi.org/10.1021/es0621593.
- 304 (16) Dixon-Anderson, E.; Lohmann, R. Field-Testing Polyethylene Passive Samplers for
- 305 the Detection of Neutral Polyfluorinated Alkyl Substances in Air and Water. *Environ*.
- 306 *Toxicol. Chem.* **2018**, *37* (12), 3002–2010. https://doi.org/10.1002/etc.4264.
- 307 (17) Wu, Y.; Romanak, K.; Bruton, T.; Blum, A.; Venier, M. Per-and Polyfluoroalkyl
- 308 Substances in Paired Dust and Carpets from Childcare Centers. *Chemosphere* 2020.
  309 https://doi.org/10.1016/j.chemosphere.2020.126771.
- 310 (18) USEPA. *Exposure Factors Handbook (EFH)*; Washington, DC, 2008.
- 311 https://doi.org/EPA/600/R-06/096F.
- 312 (19) Gebbink, W.; Berger, U.; Cousins, I. Estimating Human Exposure to PFOS Isomers
- 313 and PFCA Homologues: The Relative Importance of Direct and Indirect (Precursor)
- 314 Exposure. *Environ. Int.* **2015**, *74*, 160–169.
- 315 https://doi.org/10.1016/j.envint.2014.10.013.
- 316 (20) Liu, W.; Takahashi, S.; Sakuramachi, Y.; Harada, K. H.; Koizumi, A. Polyfluorinated
- 317 Telomers in Indoor Air of Japanese Houses. **2012**.
- 318 https://doi.org/10.1016/j.chemosphere.2012.09.062.
- 319 (21) Shoeib, M.; Harner, T.; Webster, G. M.; Lee, S. C. Indoor Sources of Poly-and
- 320 Perfluorinated Compounds (PFCS) in Vancouver, Canada: Implications for Human
- 321 Exposure. *Environ. Sci. Technol* **2011**, *45*, 7999–8005.

322

https://doi.org/10.1021/es103562v.

- 323 (22) Buck, R. C.; Franklin, J.; Berger, U.; Conder, J. M.; Cousins, I. T.; Voogt, P. De;
- Jensen, A. A.; Kannan, K.; Mabury, S. A.; van Leeuwen, S. P. J. Perfluoroalkyl and
- 325 Polyfluoroalkyl Substances in the Environment: Terminology, Classification, and
- 326 Origins. Integr. Environ. Assess. Manag. 2011. https://doi.org/10.1002/ieam.258.
- 327 (23) Vestergren, R.; Herzke, D.; Wang, T.; Cousins, I. T. Are Imported Consumer Products
- 328 an Important Diffuse Source of PFASs to the Norwegian Environment? **2015**.
- 329 https://doi.org/10.1016/j.envpol.2014.12.034.
- 330 (24) Barber, J. L.; Berger, U.; Chaemfa, C.; Huber, S.; Jahnke, A.; Temme, C.; Jones, K. C.
- 331 Analysis of Per- and Polyfluorinated Alkyl Substances in Air Samples from Northwest
- 332 Europe. J. Environ. Monit. 2007, 9 (6), 530–541. https://doi.org/10.1039/b701417a.
- 333 (25) OECD. RESULTS OF THE 2006 SURVEY ON PRODUCTION AND USE OF PFOS,

334 PFAS, PFOA, PFCA, THEIR RELATED SUBSTANCES AND

- 335 PRODUCTS/MIXTURES CONTAINING THESE SUBSTANCES; 2006.
- 336 https://doi.org/ENV/JM/MONO(2006)36.
- 337 (26) Winkens, K.; Giovanoulis, G.; Koponen, J.; Vestergren, R.; Berger, U.; Karvonen, A.;
- 338 Pekkanen, J.; Kiviranta, H.; Cousins, I. Perfluoroalkyl Acids and Their Precursors in
- 339 Floor Dust of Children's Bedrooms Implications for Indoor Exposure. *Environ. Int.*
- 340 **2018**, *119* (June), 493–502. https://doi.org/10.1016/j.envint.2018.06.009.
- 341 (27) Ellis, D. A.; Martin, J. W.; De Silva, A. O.; Mabury, S. A.; Hurley, M. D.; Sulbaek
- 342 Andersen, M. P.; Wallington, T. J. Degradation of Fluorotelomer Alcohols: A Likely
- 343 Atmospheric Source of Perfluorinated Carboxylic Acids. *Environ. Sci. Technol.* 2004,
- 344 38 (12), 3316–3321. https://doi.org/10.1021/es049860w.
- 345 (28) Haug, L.; Huber, S.; Schlabach, M.; Becher, G.; Thomsen, C. Investigation on Per-and
- 346 Polyfluorinated Compounds in Paired Samples of House Dust and Indoor Air from

- 347 Norwegian Homes. *Environ. Sci. Technol* **2011**, *45*, 7991–7998.
- 348 https://doi.org/10.1021/es103456h.
- 349 (29) Huber, S.; Haug, L.; Schlabach, M. Per-and Polyfluorinated Compounds in House
- 350 Dust and Indoor Air from Northern Norway-A Pilot Study. 2011.
- 351 https://doi.org/10.1016/j.chemosphere.2011.04.075.
- 352 (30) Goosey, E.; Harrad, S. Perfluoroalkyl Substances in UK Indoor and Outdoor Air:
- 353 Spatial and Seasonal Variation, and Implications for Human Exposure. *Environ. Int.*
- **2012**, *45* (1), 86–90. https://doi.org/10.1016/j.envint.2012.04.007.
- 355 (31) Karásková, P.; Venier, M.; Melymuk, L.; Bečanová, J.; Vojta, Š.; Prokeš, R.;
- 356 Diamond, M. L.; Klánová, J. Perfluorinated Alkyl Substances (PFASs) in Household
- 357 Dust in Central Europe and North America. 2016.
- 358 https://doi.org/10.1016/j.envint.2016.05.031.
- 359 (32) Vestergren, R.; Cousins, I.; Trudel, D.; Wormuth, M.; Scheringer, M. Estimating the
- 360 Contribution of Precursor Compounds in Consumer Exposure to PFOS and PFOA.
- 361 *Chemosphere* **2008**, *73*, 1617–1624.
- 362 https://doi.org/10.1016/j.chemosphere.2008.08.011.
- 363 (33) Winkens, K.; Koponen, J.; Schuster, J.; Shoeib, M.; Vestergren, R.; Berger, U.;
- 364 Karvonen, A. M.; Pekkanen, J.; Kiviranta, H.; Cousins, I. T. Perfluoroalkyl Acids and
- 365 Their Precursors in Indoor Air Sampled in Children's Bedrooms. *Environ. Pollut.*
- 366 **2017**, 222, 423–432. https://doi.org/10.1016/J.ENVPOL.2016.12.010.
- 367
- 368

369 Graphical TOC

# 

