The remote assessment of parkinsonism supporting ongoing development of interventions in Gaucher disease

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Practice points

- The therapeutic focus for Parkinson's disease (PD) is currently symptomatic, rather than disease modifying or neuroprotective.
- Nonmotor prodromal features of PD may precede the motor symptoms required for clinical diagnosis by several years.
- *GBA* mutations, which are associated with Gaucher disease in their biallelic form, have emerged as the most common genetic risk factor for PD.
- *GBA*-associated PD displays a slightly exaggerated phenotype, including earlier age of onset, enhanced cognitive decline, more severe affective disturbances and a greater likelihood of manifesting REM sleep behavior disorder, hyposmia and autonomic dysfunction.
- The RAPSODI GD study is an annual remote assessment which aims to define the clinical prodrome of PD in a large cohort of *GBA* mutation carriers, to accurately predict clinical diagnosis.
- The secondary objectives of RAPSODI GD are to risk stratify homozygote and heterozygote *GBA* mutation carriers for PD risk, understand the variable penetrance of the *GBA*-associated PD phenotype and create a platform for the future evaluation of biomarkers of disease.
- The putative implication of the RAPSODI GD study is to contribute toward earlier diagnosis of *GBA*-associated PD, to provide a timeframe for delivering neuroprotective interventions.
- Defining prodromal PD in *GBA* carriers may have wider implications for sporadic PD.

Mutations in *GBA* which are causative of Gaucher disease in their biallelic form, are the most common genetic risk factor for Parkinson's disease (PD). The diagnosis of PD relies upon clinically defined motor features which appear after irreversible neurodegeneration. Prodromal symptoms of PD may provide a means to predict latent pathology, years before the onset of motor features. Previous work has reported prodromal features of PD in *GBA* mutation carriers, however this has been insufficiently sensitive to identify those that will develop PD. The Remote Assessment of Parkinsonism Supporting Ongoing Development of Interventions in Gaucher Disease (RAPSODI GD) study assesses a large cohort of *GBA* mutation carriers, to aid development of procedures for earlier diagnosis of PD.

Lay abstract: Changes in a gene called *GBA* cause a rare condition called Gaucher disease and are the most common genetic risk factor for Parkinson's disease (PD). To diagnose PD, patients must show symptoms of disordered movement which only occur after irreversible brain cell loss. Earlier symptoms may allow for the prediction of PD, years before movement symptoms occur. Previous work has reported earlier symptoms of PD occurring in people with *GBA* changes, however these studies have not been able to identify those at risk of developing PD. The Remote Assessment of Parkinsonism Supporting Ongoing Development of Interventions in Gaucher Disease (RAPSODI GD) study assesses a large group of people with *GBA* changes, to help develop a way to diagnose PD earlier.

Future Medicine

Background
Challenges facing early diagnosis & intervention in Parkinson's disease
Parkinson's disease (PD) is the second most common neurodegenerative disease, with increasing global prevalence which doubled between 1990 and 2016 [1,2]. The clinicopathology of PD is characterized by motor symptoms including bradykinesia, tremor and rigidity, attributed to the progressive degeneration of dopaminergic neurons in the substantia nigra pars compacta [3,4]. An important neuropathological hallmark of PD is the presence of filamentous intracellular inclusions of α-synuclein, known as Lewy bodies [5].

Current treatments of PD focus primarily on symptom management, rather than disease modification [6]. With an aging population, the putative continuation of the upward trend in cases of PD calls for the development of preventative therapies, of which there currently are none [7,8]. One of the main challenges for developing neuroprotective therapies for PD is that clinical diagnosis relies upon the presentation of defined motor deficits, which manifest after the onset of neurodegeneration [9]. As such, there is increasing interest in characterizing the prodromal features of PD, which may precede motor symptoms by several years [10]. Several latent features of PD's clinical prodrome have been validated thus far, including REM sleep behaviour disorder (RBD), hyposmia, depression and anxiety, global cognitive impairment, constipation and autonomic dysfunction [11]. In recent years, studies such as PREDICT-PD, have optimized the validation of prodromal features of PD through remote assessment methods, enhancing recruitment and power, and allowing for repeated measures without inter-observer variation [12].

Gaucher disease & *GBA*
The *GBA* gene located on chromosome 1q21, encodes for the lysosomal hydrolase glucocerebrosidase (GCase), and has 12 exons (exon 1 is noncoding) [13,14]. Gaucher disease (GD) is a rare autosomal recessive disorder caused by biallelic homozygous or compound heterozygous *GBA* mutations, of which there have been over 400 pathogenic mutations identified [15–17]. In GD, a loss of function of *GBA* results in deficient GCase activity, and the subsequent accumulation of its substrate, a sphingolipid called glucosylceramide primarily in macrophages [18]. Literature concerning the epidemiology of GD is highly variable, with birth incidence of GD in the general population falling between 0.39 and 5.80 per 100,000, and prevalence ranging between 0.70 and 1.75 per 100,000 [19]. The prevalence of GD varies with ethnicity and is markedly higher in Ashkenazi Jewish populations, who also have a higher *GBA* mutation carrier frequency [19]. GD has three clinical subtypes: 1, 2 and 3 [20]. Type 1 GD is the mildest and most common phenotype, accounting for 94% of cases in western populations [19]. Type 1 is characterized by clinicopathological features including bone lesions, hepatosplenomegaly, thrombocytopenia and anemia, in the absence of primary neurological disease [18]. Types 2 and 3 are neuronopathic forms, with mean lethality at <4 years old and <40 years old, respectively [21]. *GBA* mutations vary in pathogenicity therefore those that cause type 1 GD (such as N370S) are considered mild, whereas those that also cause type 2 and 3 GD (such as L444P) are denoted as severe [22,23].

The link between *GBA* mutations & PD
Within the last three decades, clinicians have reported the presentation of parkinsonian motor features in a subset of patients with GD [24,25]. An increased incidence of PD within GD kindreds was observed, and PD patients with *GBA* mutations were more likely to report a first or second degree relative with PD, than those without *GBA* mutations [26–28]. Parkinsonism has presented in GD patients with both neuronopathic and non-neuronopathic *GBA* variants [29]. Subsequently, in an international multi-centre analysis a strong association was detected between *GBA* mutations and PD, with an odds ratio of 5.43 for carrying heterozygous *GBA* mutations in PD patients versus controls [28]. This work established heterozygous *GBA* mutations as the most common genetic risk factor for PD. While an increased lifetime risk for developing PD has been established for both biallelic and monoallelic carriers of *GBA* mutations, it is still only a minority of individuals with these mutations that will go on to develop PD [30,31]. Heterozygous *GBA* mutation carrier status confers a 3–15% risk of PD by age 80, contingent upon the severity of the variant, however the literature concerning the penetrance of these mutations in PD is variable [23,30,32,33].

Penetrance of *GBA* mutations in PD has been estimated between 10 and 30% and can be stratified by genetic and nongenetic factors, including age-specificity and severity of mutation [33,34].

The prodrome of GBA-associated PD

There is an overrepresentation of literature concerning the natural history of PD associated with the two most frequent *GBA* pathogenic variants, L444P and N370S, compared with the role of many of the other hundreds of variants [13,22,35–37]. The phenotype of *GBA* associated PD (*GBA* PD), differs from sporadic PD (sPD), with a slightly earlier age of onset of approximately 5 years, with a dose effect for severe versus mild mutations [23,38]. It has also emerged that *GBA* PD is associated with more nonmotor symptoms than its idiopathic counterpart, including a greater likelihood of manifesting cognitive dysfunction concomitant with more aggressive cognitive decline, more severe affective disorder, and an increased risk of experiencing RBD, hyposmia and autonomic dysfunction [30,39–42].

GCase in sporadic PD

While the molecular mechanisms of pathogenesis have not been fully elucidated for *GBA* PD, much like in GD there is a significant reduction in GCase activity [43]. Histopathological analyses of post-mortem PD brain tissue have identified reduced GCase activity and protein levels in regions with elevated expression of α-synuclein [43], and this was subsequently reproduced [44,45]. The most significant reduction in GCase activity was noted in the substantia nigra, and notably these observations were found in both *GBA* PD tissue and tissue from individuals with sPD [43]. Work in iPSC models has suggested that the involvement of GCase in both *GBA* PD and sPD, may result from a bidirectional relationship between reduced lysosomal GCase activity and cytotoxic α-synuclein aggregation [46–48]. Due to the implication of deficient GCase activity in both *GBA* PD and sPD, GCase has become an attractive therapeutic target for PD [49–51]. Murine and iPSC models of *GBA* PD and sPD have demonstrated the promise of gene therapies and small-molecule chaperone modulation, to increase GCase activity and subsequently reduce α-synuclein accumulation with neuroprotective outcomes [52,53]. Understanding the mechanisms of GCase-mediated PD pathogenesis is therefore a salient research focus for not only *GBA* PD but more widely sPD.

Objectives & rationale

The increasing global burden of PD highlights the relevance of utilizing PD's clinical prodrome for earlier diagnosis, and intervention [2]. As the most common genetic risk factor for PD, with premotor features and putative implications for sPD, defining prodromal PD features associated with *GBA* is an important target for enhancing detection of disease and earlier intervention timepoints [3,28,39,49].

The RAPSODI GD study aims to define prodromal symptoms of PD in individuals who carry *GBA* mutations, both homozygote and heterozygote, and to further elucidate the natural history of *GBA* PD. It will test the hypothesis that the presentation of prodromal features of PD for *GBA* can predict the onset of clinical diagnosis. In doing so, it may be possible to develop models for the accurate prediction of individuals at-risk of developing *GBA* PD, affording earlier opportunities for intervention before irreversible neurodegeneration. Secondary objectives of the study include:

- Undertaking PD risk stratification for individuals with biallelic versus monoallelic *GBA* mutations;
- Serving as a platform for future research to identify at-risk individuals for *GBA* PD, to explore biomarkers of *GBA* PD;
- Probing the limited and variable penetrance of the PD phenotype for individuals with *GBA* mutations.

Design & methodology

RAPSODI GD is a remote, longitudinal, descriptive, natural history, registry-based study. Repeated measures are delivered on an annual or biennial basis following baseline assessment. Assessment is undertaken via the online study portal (www.rapsodistudy.com), and by post. Saliva sample collection and genotyping for *GBA* and *LRRK2* mutations is an independent measure, occurring once at baseline. The study aims to assess participants for the presence and progression of six prodromal features of PD: olfactory dysfunction, motor dysfunction, cognitive decline, anxiety and depression, motor symptoms, REM sleep dysfunction. RAPSODI GD has 25 years of ethical approval from London - Queen Square Research Ethics Committee (REC reference: 15/LO/1155).

Sampling

Purposive sampling has been used to recruit participants from the following strata: heterozygous *GBA* mutation carriers, homozygous *GBA* mutation carriers, heterozygote *GBA* mutation carriers with a confirmed clinical diagnosis of PD, idiopathic PD cases with unknown *GBA* status, controls. These sampling methods were chosen with the aim of optimizing recruitment for each stratum, to develop a large registry of *GBA* mutation homozygotes and heterozygotes. This method seeks to enhance the statistical power necessary for the detection of subtle prodromal features of PD, allowing for future analysis and comparison of the distribution of premotor symptoms of PD between groups.

RAPSODI GD has eight Patient Identification Centres (PICs), where we are partnered with lysosomal storage disorder units, (Royal Free Hospital, Addenbrooke's Hospital, Queen Elizabeth Hospital Birmingham, Salford Royal Hospital, Cardiff and Vale University Health Board, University College Hospital, Great Ormond Street Hospital and Manchester Children's Hospital). From these PICs GD patients with homozygous *GBA* mutations are recruited. These participants act as index cases, to recruit first-degree relatives from. These relatives once consented are subsequently screened and determined to be homozygous or heterozygous for *GBA* mutations, or controls. Our sister study, PD FRONTLINE, is a newly established offshoot which screens idiopathic PD cases for *GBA* mutations. All idiopathic PD cases will initially be recruited and screened via the PD FRONTLINE platform to ensure *GBA* mutation status. These *GBA* mutation positive individuals will then be asked to reconsent and reassign from PD FRONTLINE to RAPSODI GD and will act as index cases to contact first degree relatives through. Relatives will be determined to be homozygous or heterozygous for *GBA* mutations, or controls. Family tracing from index cases is only pursued with permission from the participant. Due to the descriptive nature of RAPSODI GD, we envision that recruitment will remain active throughout the full course of the study period, up to the limit of 10,000 participants.

The inclusion criteria for RAPSODI GD are as follows: homozygous *GBA* mutation carriers, heterozygous *GBA* mutation carriers, first degree relatives of heterozygous or homozygous *GBA* mutation carriers, spouses of individuals matching one of inclusion criteria 1–3, heterozygous or homozygous *GBA* mutation carriers with a confirmed clinical diagnosis of PD, idiopathic PD cases with unknown *GBA* status, age 18–90, informed consent.

The exclusion criteria for RAPSODI GD are as follows: *GBA* mutation negative individuals with a confirmed clinical diagnosis of PD, a history of parkinsonism, individuals with other neurological disorders, individuals on drugs known to be associated with parkinsonism.

Remote annual & biennial assessments

Annual remote assessments within RAPSODI GD take place online, via the RAPSODI GD portal (www.rapsodistudy.com). The battery of assessments takes approximately 1 h to complete.

Informed consent

Participant consent is gained upon enrolment within the online RAPSODI GD portal (www.rapsodistudy.com), or the PD FRONTLINE portal for participants with idiopathic PD (www.pdfrontline.com).

Self-report rating scales

The RAPSODI GD study uses three self-administered scales to assess: anxiety and depression, motor symptoms of PD, REM sleep disturbance. Anxiety and depression are examined through the Hospital Anxiety and Depression Scale (HADS) [54]. This 14-item scale determines separate scores for anxiety and depression, between 0 and 21, distinguishing between mild, moderate, severe and noncases. Motor symptoms of PD are assessed via the Movement Disorder Society-Unified Parkinson's Disease Rating Scale (MDS-UPDRS) part II [55]. This 13-item section of the MDS-UPDRS, rates motor experiences in daily living such as eating and walking, on a scale of 0–4 from normal to severely dysfunctional. REM sleep disturbance is measured via the REM Sleep behaviour Disorder Screening Questionnaire (RBDSQ) [56]. This 10-item scale assesses clinical features of REM sleep behaviour disorder, such as dream content and sleep movement.

Bradykinesia akinesia incoordination test

Motor dysfunction is assessed via the Bradykinesia Akinesia Incoordination (BRAIN) Test [57]. On a standard physical computer/laptop keyboard, participants must alternate between tapping the 's' key and ';' key for 30 s per hand. Incoordination, akinesia, dysmetria and kinesia are assigned scores based on the variance in time travelling

between the keys, the time spent on keys, the accuracy of pressing the correct keys and the total number of key presses, respectively.

Cognitive testing

Several aspects of cognitive function are assessed via a battery of precision cognitive tests. This was historically delivered via the online platform 'CogTrack™' [58], until the platform expired, and cognitive testing moved to a new platform 'NeurOn' in November 2020 (www.neuropsychology.online). The battery includes tasks of image encoding, simple reaction time, digit span backwards, spatial working memory and image recall. Within this battery participants are scored on dysfunction in working and episodic memory, disturbances in attention focus and sustainment and memory retrieval rate.

Hyposmia assessment

Olfactory function is assessed on a biennial basis via the University of Pennsylvania Smell Identification Test (UPSIT) [59]. The UPSIT is delivered remotely through the post, and scores are uploaded directly onto the RAPSODI GD portal by participants. The UPSIT utilizes a series of booklets, each containing ten microencapsulated odorant panels. Participants apply mechanical pressure to the panels to release the odorants, and then select which of four options they believe each odorant to represent. The participants are assigned a score of olfaction sensitivity from 0 to 40 depending on the accuracy of their responses.

Genotyping

In their baseline year with RAPSODI GD, participants are posted a saliva sample collection kit (DNA Genotek Oragene.DNA OG-500), with biological substance category B compliant return packaging. Genomic DNA is extracted via the protocol provided in the DNA purification handbook published by DNA Genotek, for use with the Oragene.DNA OG-500 kits [60]. The full 8.9 kb amplicon of the *GBA* gene is sequenced for all known deleterious variants, on the Oxford Nanopore MinION, via a protocol validated by Leija-Salazar *et al.* [61]. Due to RAPSODI GD's active selection of individuals with *GBA* mutations, there is a likelihood that individuals from an Ashkenazi Jewish population will be overrepresented within the cohort [23]. As such, genomic DNA samples are also MinION sequenced for the most common *LRRK2* mutation, G2019S, as this is a common, salient and potentially confounding PD associated risk factor for Ashkenazi Jewish individuals [62].

Upon detection of mutation positive cases, DNA samples will be sent for external confirmatory Sanger sequencing in an NHS accredited lab.

Follow-up

The self-reporting scales, Bradykinesia akinesia incoordination test, and cognitive testing are repeated once a year from baseline. The UPSIT is repeated once every other year from baseline.

Conclusion

The RAPSODI GD study described here aims to define prodromal features of PD in a large cohort of *GBA* mutation carriers, to accurately predict clinical diagnosis. The study also looks to stratify the risk of phenoconversion to PD in homozygote and heterozygote *GBA* mutation carriers, and to clarify the variable penetrance of the *GBA*-associated PD phenotype, while developing a platform that will allow for the assessment of biomarkers of disease. The postulated outcome of the RAPSODI GD study is to allow for the identification and earlier diagnosis of individuals at risk of developing PD, so that neuroprotective interventions can be administered within an efficacious therapeutic timeframe.

Future perspective

The development and validation of clearly defined features of prodromal PD for *GBA* mutation carriers, could eventually allow for the accurate detection of at-risk individuals. Clinical detection of disease in this latent phase, would lend itself to the identification of earlier therapeutic timeframes to deliver neuroprotective interventions before irreversible neurodegeneration can take place. As the mechanisms of pathogenesis in the GCase-mediated pathway of PD are elucidated, biomarkers and therapeutic targets characterized for *GBA* PD may also prove salient in developing preventative measures within sPD as well.

Ethical conduct of research

RAPSODI has been approved by London - Queen Square Research Ethics Committee (REC reference: 15/LO/1155). Participant consent is gained upon enrolment within the online RAPSODI GD portal (www.rapsodistudy.com), or the PD FRONTLINE portal for participants with idiopathic Parkinson's disease (www.pdfrontline.com).

References

Papers of special note have been highlighted as: ● of interest; ●● of considerable interest

1. Elbaz A, Carcaillon L, Kab S, Moisan F. Epidemiology of Parkinson's disease. *Rev. Neurol.* 172(1), 14–26 (2016).

2. Dorsey ER, Elbaz A, Nichols E *et al.* Global, regional, and national burden of Parkinson's disease, 1990–2016: a systematic analysis for the Global Burden of Disease Study 2016. *The Lancet Neurol.* 17(11), 939–953 (2018).

●   **Highlights the upward trend in Parkinson's disease (PD) global burden.**

3. Shulman JM, De Jager PL, Feany MB. Parkinson's disease: genetics and pathogenesis. *Annu. Rev. Pathol. Mech. Dis.* 6(1), 193–222 (2011).

4. Schapira AHV, Chaudhuri KR, Jenner P. Non-motor features of Parkinson's disease. *Nat. Rev. Neurosci.* 18(7), 435–450 (2017).

5. Wakabayashi K, Tanji K, Odagiri S, Miki Y, Mori F, Takahashi H. The Lewy body in Parkinson's disease and related neurodegenerative disorders. *Mol. Neurobiol.* 47(2), 495–508 (2013).

6. Ellis JM, Fell MJ. Current approaches to the treatment of Parkinson's disease. *Bioorg. Med. Chem. Lett.* 27(18), 4247–4255 (2017).

7. Rocca WA. The burden of Parkinson's disease: a worldwide perspective. *The Lancet Neurol.* 17(11), 928–929 (2018).

8. Paolini Paoletti F, Gaetani L, Parnetti L. The challenge of disease-modifying therapies in Parkinson's disease: role of CSF biomarkers. *Biomolecules* 10(2), 335 (2020).

9. Mahlknecht P, Seppi K, Poewe W. The concept of prodromal Parkinson's disease. *J. Parkinsons. Dis.* 5(4), 681–697 (2015).

10. Schapira AHV, Tolosa E. Molecular and clinical prodrome of Parkinson's disease: implications for treatment. *Nat. Rev Neurol.* 6(6), 309–317 (2010).

●   **Postulates that nonmotor features of PD can emerge years before clinically defined motor deficits**

11. Heinzel S, Berg D, Gasser T *et al.* Update of the MDS research criteria for prodromal Parkinson's disease. *Mov. Disord.* 34(10), 1464–1470 (2019).

●● **Discusses validated features of the clinical prodrome of PD.**

12. Noyce AJ, Bestwick JP, Silveira-Moriyama L *et al.* PREDICT-PD: identifying risk of Parkinson's disease in the community: methods and baseline results. *J. Neurol. Neurosur. Ps.* 85(1), 31–37 (2014).

13. Riboldi GM, Di Fonzo AB. GBA, Gaucher disease, and Parkinson's disease: from genetic to clinic to new therapeutic approaches. *Cells* 8(4), 364 (2019).

14. National Center for Biotechnology Information. GBA glucosylceramidase beta [*Homo sapiens* (human)] (2004). https://www.ncbi.nlm.nih.gov/gene/2629

15. Stirnemann J, Belmatoug N, Camou F *et al.* A review of Gaucher disease pathophysiology, clinical presentation and treatments. *Int. J. Mol. Sci.* 18(2), 441 (2017).

16.   Hruska KS, LaMarca ME, Scott CR, Sidransky E. Gaucher disease: mutation and polymorphism spectrum in the glucocerebrosidase gene (GBA). *Hum. Mutat.* 29(5), 567–583 (2008).

17.   Stenson *et al.* The Human Gene Mutation Database (HGMD®): 2003 update. *Hum. Mutat.* 21, 577–581 (2003).

18.   Hughes D, Mikosch P, Belmatoug N *et al.* Gaucher disease in bone: from pathophysiology to practice. *J. Bone Miner. Res.* 34(6), 996–1013 (2019).

19.   Nalysnyk L, Rotella P, Simeone JC, Hamed A, Weinreb N. Gaucher disease epidemiology and natural history: a comprehensive review of the literature. *Hematology* 22(2), 65–73 (2017).

20.   Lee J-Y, Lee BH, Kim G-H *et al.* Clinical and genetic characteristics of Gaucher disease according to phenotypic subgroups. *Korean J. Pediatr.* 55(2), 48 (2012).

21.   Pastores GM, Hughes DA. GeneReviews®: Gaucher disease (2000). https://www.ncbi.nlm.nih.gov/books/NBK1269/

22.   Beutler E, Gelbart T, Scott CR. Hematologically important mutations: Gaucher disease. *Blood Cells Mol. Dis.* 35(3), 355–364 (2005).

23.   Gan-Or Z, Amshalom I, Kilarski LL *et al.* Differential effects of severe vs mild GBA mutations on Parkinson's disease. *Neurology* 84(9), 880–887 (2015a).

24.   Neudorfer O, Giladi N, Elstein D *et al.* Occurrence of Parkinson's syndrome in type 1 Gaucher disease. *QJM* 89(9), 691–694 (1996).

25.   Tayebi N, Callahan M, Madike V *et al.* Gaucher disease and parkinsonism: a phenotypic and genotypic characterization. *Mol. Genet. Metab.* 73(4), 313–321 (2001).

26.   Halperin A, Elstein D, Zimran A. Increased incidence of Parkinson's disease among relatives of patients with Gaucher disease. *Blood Cells Mol. Dis.* (3), 426–428 (2006).

27.   Goker-Alpan O, Schiffmann R, LaMarca M, Nussbaum R, McInerey-Leo A, Sidransky E. Parkinsonism among Gaucher disease carriers. *J. Med. Genet.* 41(12), 937–940 (2004).

28.   Sidransky E, Nalls MA, Aasly JO *et al.* Multicenter analysis of glucocerebrosidase mutations in Parkinson's disease. *N. Engl. J. Med.* 361(17), 1651–1661 (2009).
••   **Identified heterozygous GBA as the most common risk factor for PD.**

29.   Tayebi N, Walker J, Stubblefield B *et al.* Gaucher disease with parkinsonian manifestations: does glucocerebrosidase deficiency contribute to a vulnerability to parkinsonism? *Mol. Genet. Metab.* 79(2), 104–109 (2003).

30.   McNeill A, Duran R, Hughes DA, Mehta A, Schapira AHV. A clinical and family history study of Parkinson's disease in heterozygous glucocerebrosidase mutation carriers. *J. Neurol. Neurosur. Ps.* 83(8), 853–854 (2012a).

31.   Bultron G, Kacena K, Pearson D *et al.* The risk of Parkinson's disease in type 1 Gaucher disease. *J. Inherit. Metab. Dis.* 33(2), 167–173 (2010).

32.   Rana HQ, Balwani M, Bier L, Alcalay RN. Age-specific Parkinson's disease risk in GBA mutation carriers: information for genetic counselling. *Genet. Med.* 15(2), 146–149 (2013).

33.   Anheim M, Elbaz A, Lesage S *et al.* Penetrance of Parkinson's disease in glucocerebrosidase gene mutation carriers. *Neurology* 78(6), 417–420 (2012).

34.   Balestrino R, Tunesi S, Tesei S, Lopiano L, Zecchinelli AL, Goldwurm S. Penetrance of glucocerebrosidase (GBA) mutations in Parkinson's disease: a kin cohort study. *Mov. Disord.* 35(11), 2111–2114 (2020).

35.   Neumann J, Bras J, Deas E *et al.* Glucocerebrosidase mutations in clinical and pathologically proven Parkinson's disease. *Brain* 132(7), 1783–1794 (2009).

36.   Winder-Rhodes SE, Evans JR, Ban M *et al.* Glucocerebrosidase mutations influence the natural history of Parkinson's disease in a community-based incident cohort. *Brain* 136(2), 392–399 (2013).

37.   Malek N, Weil RS, Bresner C *et al.* Features of GBA – associated Parkinson's disease at presentation in the UK Tracking Parkinson's study. *J. Neurol. Neurosur. Ps.* 89(7), 702–709 (2018).

38.   Zhang Y, Sun Q, Zhao Y *et al.* Effect of GBA mutations on phenotype of Parkinson's disease: a study on Chinese population and a meta-analysis. *Parkinsons Dis.* 2015, 1–10 (2015).

39.   Beavan M, McNeill A, Proukakis C, Hughes DA, Mehta A, Schapira AHV. Evolution of prodromal clinical markers of Parkinson's disease in a GBA mutation-positive cohort. *JAMA Neurol.* 72(2), 201 (2015).
•   **Identified enhanced prodromal featured of GBA-associated PD.**

40.   Brockmann K, Srulijes K, Hauser A-K *et al.* GBA-associated PD presents with nonmotor characteristics. *Neurology* 77(3), 276–280 (2011).

41.   Gan-Or Z, Mirelman A, Postuma RB *et al.* GBA mutations are associated with Rapid Eye Movement Sleep Behavior Disorder. *Ann. Clin. Transl. Neurol.* 2(9), 941–945 (2015b).

42.   McNeill A, Duran R, Proukakis C *et al.* Hyposmia and cognitive impairment in Gaucher disease patients and carriers. *Mov. Disord.* 27(4), 526–532 (2012b).

43. Gegg ME, Burke D, Heales SJR *et al.* Glucocerebrosidase deficiency in substantia nigra of Parkinson's disease brains. *Ann. Neurol.* 72(3), 455–463 (2012).

• **Linked the GCase deficiency to both GBA associated PD and sporadic PD.**

44. Do J, McKinney C, Sharma P, Sidransky E. Glucocerebrosidase and its relevance to Parkinson's disease. *Mol. Neurodegener.* 14(1), 36 (2019).

45. Murphy KE, Gysbers AM, Abbott SK *et al.* Reduced glucocerebrosidase is associated with increased α-synuclein in sporadic Parkinson's disease. *Brain* 137(3), 834–848 (2014).

46. Mazzulli JR, Xu Y-H, Sun Y *et al.* Gaucher disease glucocerebrosidase and α-synuclein form a bidirectional pathogenic loop in synucleinopathies. *Cell* 146(1), 37–52 (2011).

47. Schöndorf DC, Aureli M, McAllister FE *et al.* iPSC-derived neurons from GBA1-associated Parkinson's disease patients show autophagic defects and impaired calcium homeostasis. *Nat. Commun.* 5, 4028 (2014).

48. Yang SY, Beavan M, Chau KY, Taanman JW, Schapira AHV. A human neural crest stem cell-derived dopaminergic neuronal model recapitulates biochemical abnormalities in GBA1 mutation carriers. *Stem Cell Rep.* 8(3), 728–742 (2017).

49. Chen Y, Sam R, Sharma P, Chen L, Do J, Sidransky E. Glucocerebrosidase as a therapeutic target for Parkinson's disease. *Expert Opin. Ther.* 24(4), 287–294 (2020).

50. Menozzi E, Schapira AHV. Enhancing the activity of glucocerebrosidase as a treatment for Parkinson's disease. *CNS Drugs* 34(9), 915–923 (2020).

51. Mullin S, Smith L, Lee K *et al.* Ambroxol for the treatment of patients with Parkinson's disease with and without glucocerebrosidase gene mutations: a nonrandomized, noncontrolled trial. *JAMA Neurol.* 77(4), 427–434 (2020).

52. Burbulla LF, Jeon S, Zheng J, Song P, Silverman RB, Krainc D. A modulator of wild-type glucocerebrosidase improves pathogenic phenotypes in dopaminergic neuronal models of Parkinson's disease. *Sci. Transl. Med.* 11(514), eaau6870 (2019).

53. Rocha EM, Smith GA, Park E *et al.* Glucocerebrosidase gene therapy prevents α-synucleinopathy of midbrain dopamine neurons. *Neurobiol. Dis.* 82, 495–503 (2015).

54. Zigmond AS, Snaith RP. The hospital anxiety and depression scale. *Acta. Psychiatr. Scand.* 67(6), 361–370 (1983).

55. Goetz CG, Tilley BC, Shaftman SR *et al.* Movement Disorder Society-sponsored revision of the Unified Parkinson's Disease Rating Scale (MDS-UPDRS): scale presentation and clinimetric testing results: MDS-UPDRS: clinimetric assessment. *Mov. Disord.* 23(15), 2129–2170 (2008).

56. Stiasny-Kolster K, Mayer G, Schäfer S, Möller JC, Heinzel-Gutenbrunner M, Oertel WH. The REM sleep behavior disorder screening questionnaire – a new diagnostic instrument. *Mov. Disord.* 22(16), 2386–2393 (2007).

57. Homann CN, Suppan K, Wenzel K *et al.* The Bradykinesia Akinesia Incoordination Test (BRAIN TEST), an objective and user-friendly means to evaluate patients with parkinsonism. *Mov. Disord.* 15(4), 641–647 (2000).

58. Wesnes KA, Brooker H, Ballard C, McCambridge L, Stenton R, Corbett A. Utility, reliability, sensitivity and validity of an online test system designed to monitor changes in cognitive function in clinical trials: online cognitive testing in clinical trials. *Int. J. Geriatr. Psychiatry* 32(12), e83–e92 (2017).

59. Doty RL, Shaman P, Dann M. Development of the university of pennsylvania smell identification test: a standardized microencapsulated test of olfactory function. *Physiol. Behav.* 32(3), 489–502 (1984).

60. DNA Genotek. Laboratory protocol for manual purification of DNA from 0.5 ml of sample (2018). https://www.dnagenotek.com/ROW/pdf/PD-PR-006.pdf

61. Leija-Salazar M, Sedlazeck FJ, Toffoli M *et al.* Evaluation of the detection of GBA missense mutations and other variants using the Oxford Nanopore MinION. *Mol. Genet. Genom. Med.* 7(3), e564 (2019).

62. Ozelius LJ, Senthil G, Saunders-Pullman R *et al.* LRRK2 G2019S as a cause of Parkinson's disease in Ashkenazi Jews. *N. Engl. J. Med.* 354(4), 424–425 (2006).