

SUPPLEMENTARY MATERIAL

Focused CRISPR-Cas9 genetic screening reveals USO1 as a vulnerability in B-cell acute lymphoblastic leukemia

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Figure S1

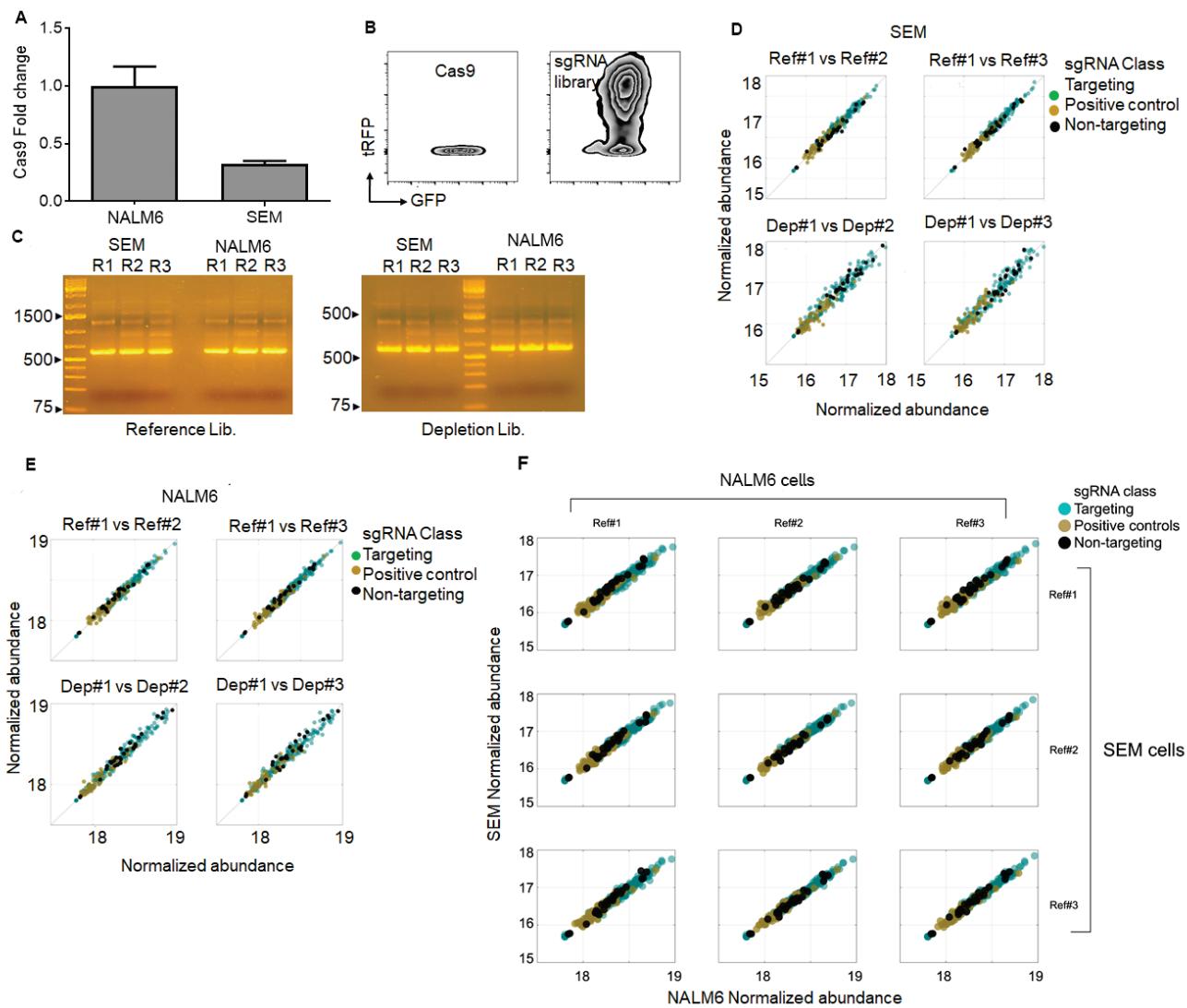


Figure S1. Sub-genomic CRISPR screen preparation. **A.** Cas9 expression in B-ALL cell line NALM6 and SEM prior to CRISPR screen. **B.** FACS plot showing transduction of NALM6 Cas9.EGFP with the undiluted lentiviral vector library, demonstrating effective production of virus with a high titer. **C.** Library preparation from the genomic DNA isolated from the sorted cells as Reference library and from expanded cells for 4 weeks as Depletion library. **D-E.** Intra-cell line Correlation plot of individual sgRNA variance-stabilized abundances in the reference and depletion libraries prepared in SEM and NALM6 cell lines between biological replicates. **F.** Inter-cell line correlation plots of individual sgRNA variance-stabilized abundances in the reference and depletion libraries prepared in SEM and NALM6 cell lines between biological replicates.

Figure S2

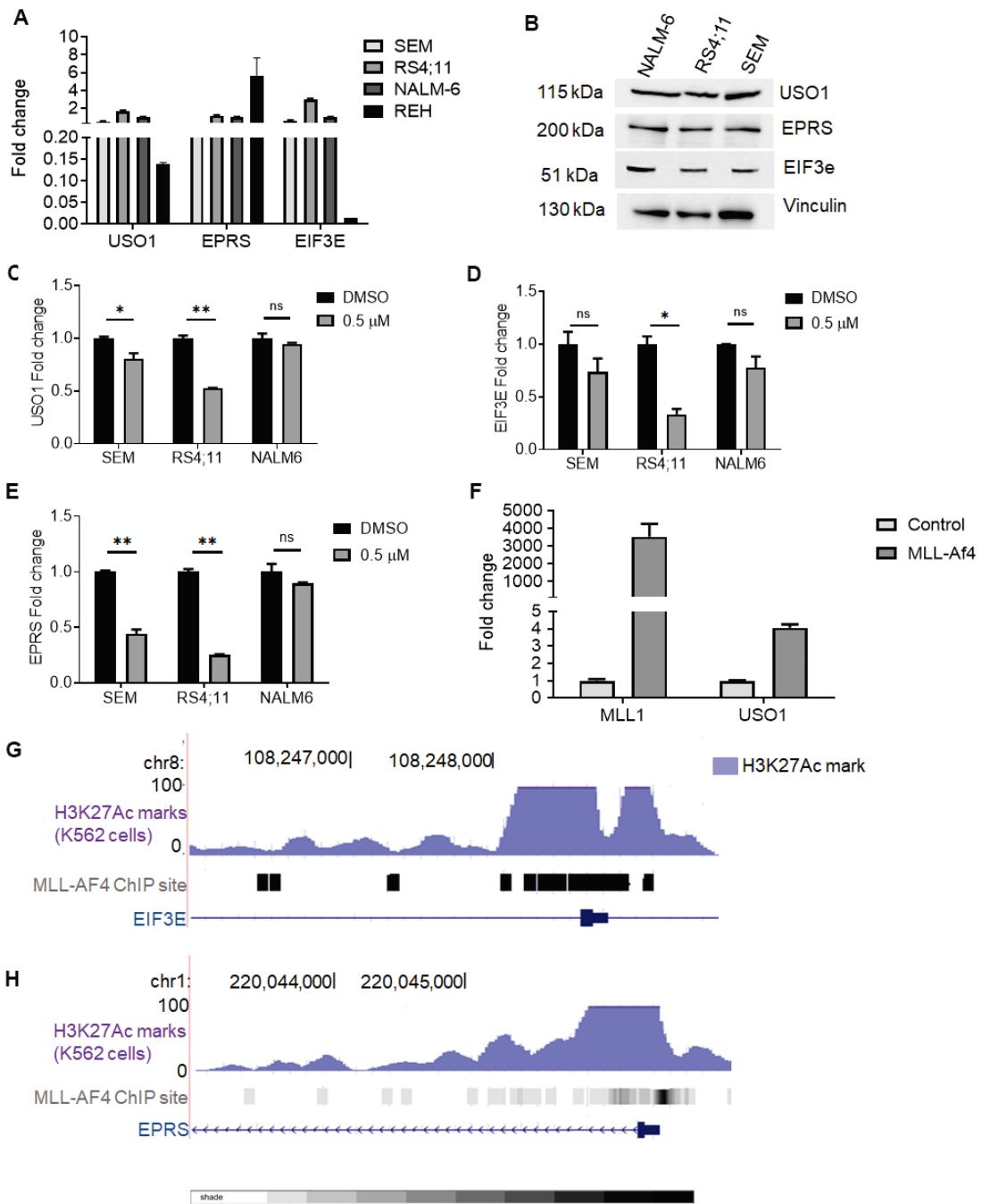


Figure S2. RBP genes expression in B-ALL cell lines. **A.** mRNA expression of RBP genes identified, measured by RT-qPCR (*USO1*, *EIF3E* and *EPERS*). Data was normalized to 18S expression level and is represented as fold-change from SEM cells, which are arbitrarily assigned a fold-change of 1. Cell lines examined include SEM, RS4;11 (carry the MLL-AF4 translocation), and NALM6 and REH (do not carry the MLL-AF4 translocation). **B.** Western blot to study basal level expression of *USO1*, *EIF3E* and *EPERS* in NALM6, SEM and RS4;11 cells. **C-E.** RT-qPCR analysis of expression of *USO1* (C), *EIF3E* (D), and *EPERS* (E) in SEM, RS4;11 and NALM6 cells treated with 0.5 μM EPZ5676. **F.** RT-qPCR analysis of *USO1* in murine bone marrow cells with overexpression of MLL-Af4 (control versus MLL-Af4 bars). **G-H.** UCSC genome browser shots of the *EIF3E* and *EPERS* loci showing the *MLL-AF4* ChIP site(s) in the regulatory region as identified from the ChIP-Seq data from Lin *et. al.*(ref #21); Courtesy: UCSC Genome Browser). Shown are the H3K27Ac track in hematopoietic K562 cells (Blue), and MLL-AF4 binding sites represented as a grayscale score, with black indicating the highest score/highest number of reads from the dataset.

Figure S3

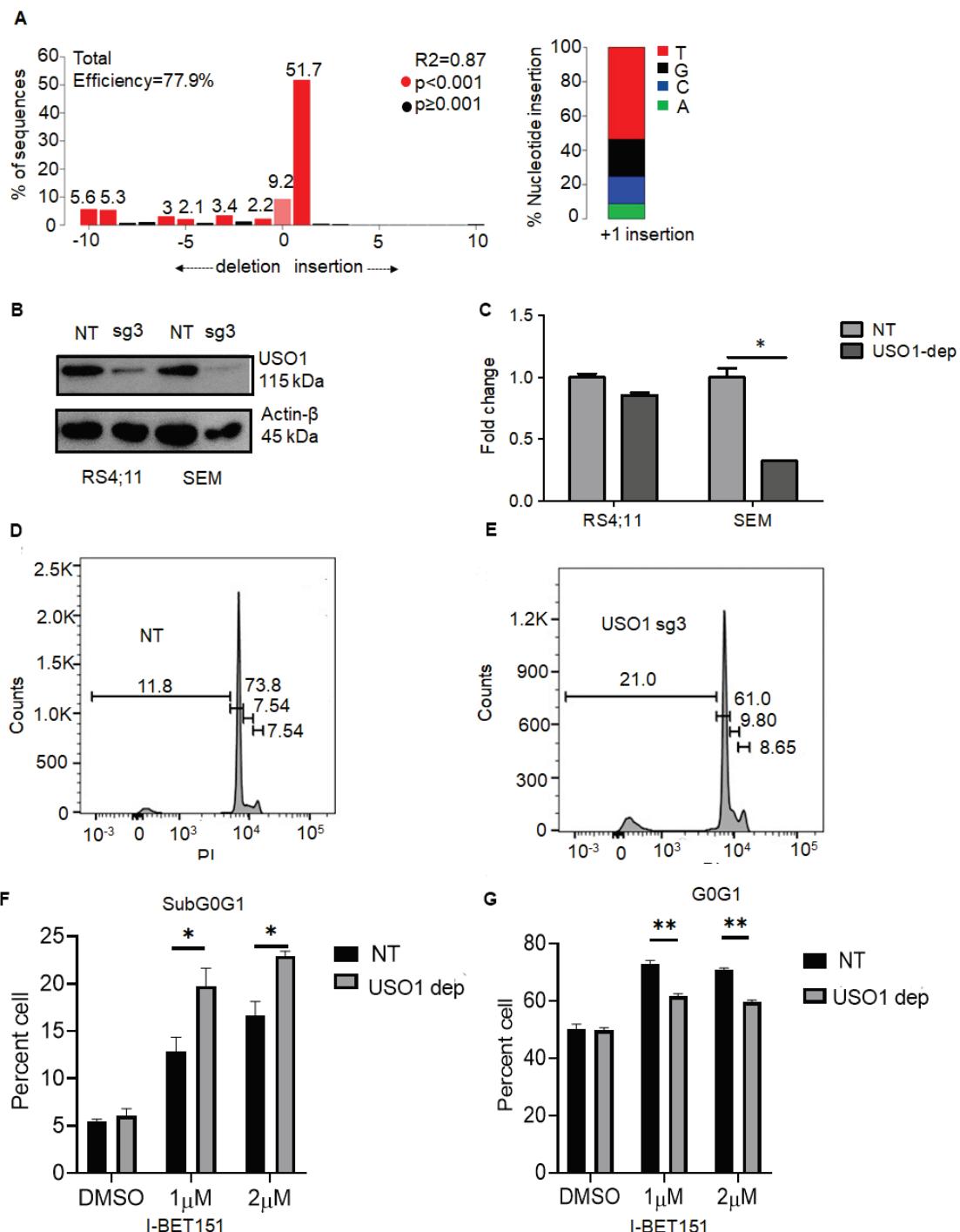


Figure S3. Targeting USO1 in B-ALL cells. **A.** Plot of insertions and deletions by position near the Cas9 cut site in SEM cells targeted with *USO1* sg3; as determined by Tracking of Indels by Decomposition (TIDE). Genomic sequence aberration is highest immediately adjacent to the cut site (designated as 0). **B.** USO1 depletion by transduction with sg3 in RS4;11 and SEM cells was confirmed by Western blot analysis. **C.** RT-qPCR of USO1 mRNA expression level, expressed as fold-change, in RS4;11 and SEM cells (t-test; * $P < 0.05$). **D-G.** Cell cycle analysis using propidium iodide (PI) of control cells (NT) and USO1-depleted cells (D, E, respectively) following treatment with 1mM I-BET151. F-G. Quantitation of cells from cell cycle analysis in Sub-G0 (F) and G0/G1 (G) at 2 different concentrations of I-BET151(t-test; * $P < 0.05$, ** $P < 0.001$).

Figure S4

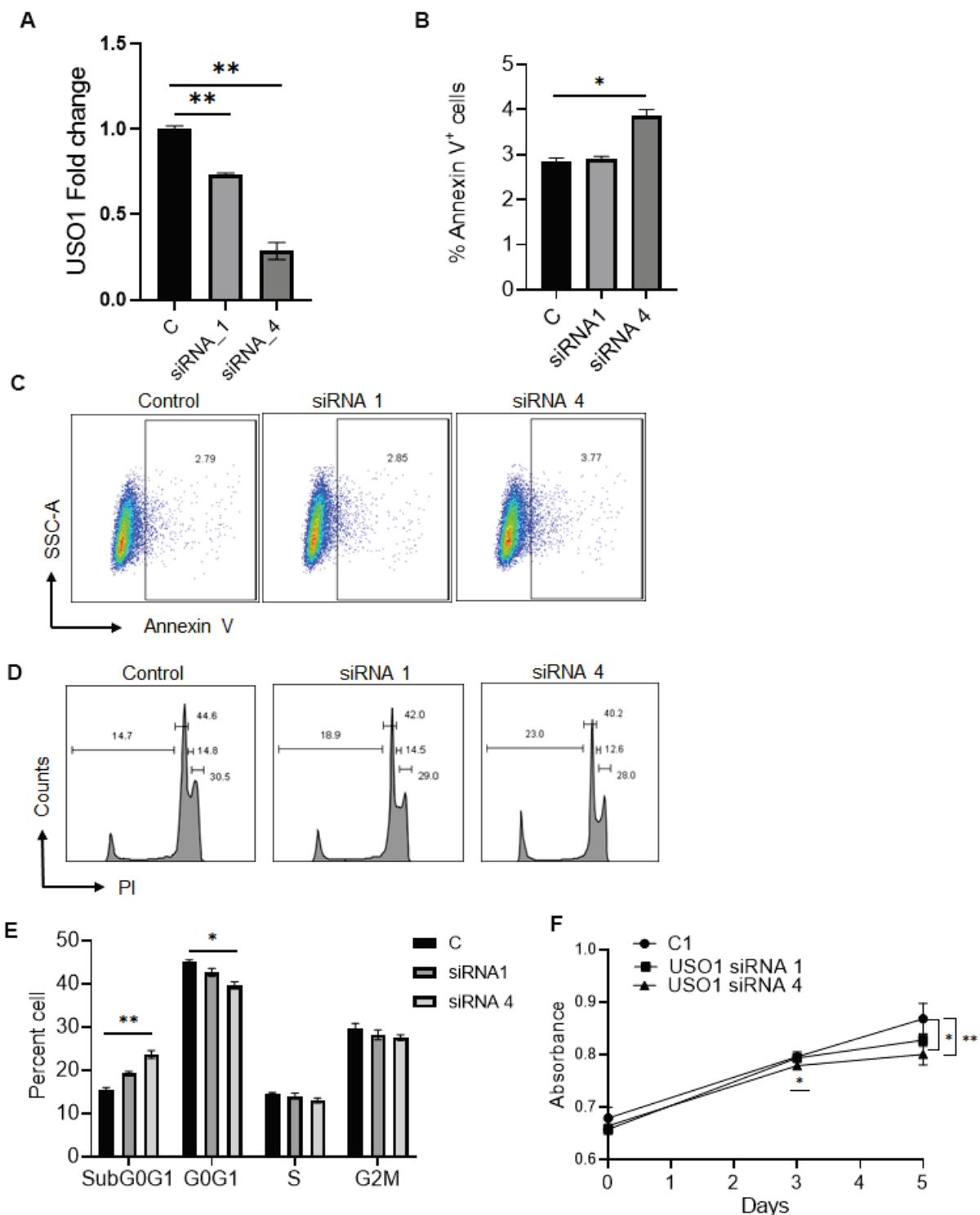
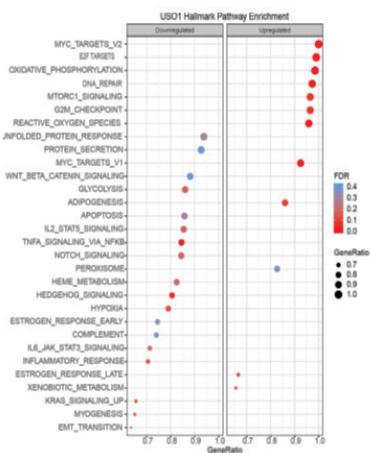


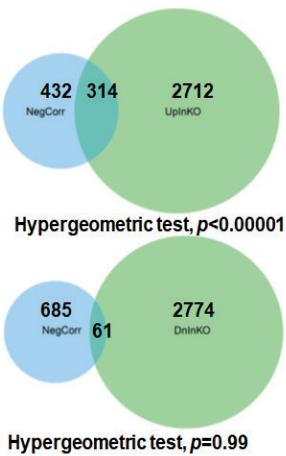
Figure S4. Modulation of USO1 expression by siRNA mediated knockdown. **A.** RT-qPCR analysis of USO1 expression levels, expressed as fold-change, in SEM cells that were subjected to nucleofection with a control siRNA or siRNA_1 or siRNA_4. **B-C.** Apoptosis induction, as measured by Annexin V staining, in SEM cells that were transduced with USO1-siRNAs. **D-E.** Cell cycle analysis, using propidium iodide staining, of USO1-siRNA-treated cells. **F.** MTS assays (Absorbance at 490nm) measuring the growth over time (days) of USO1-siRNA-treated SEM cells. (T-test; *P < 0.05, **P< 0.001).

Figure S5

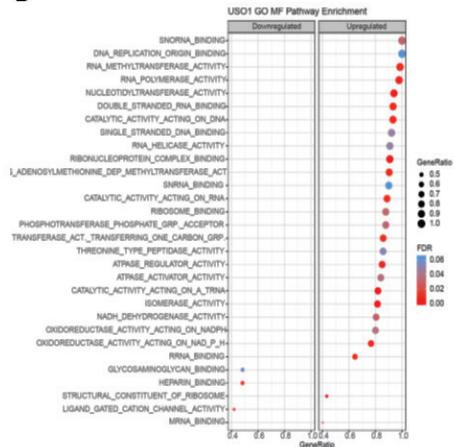
A



C



B



D

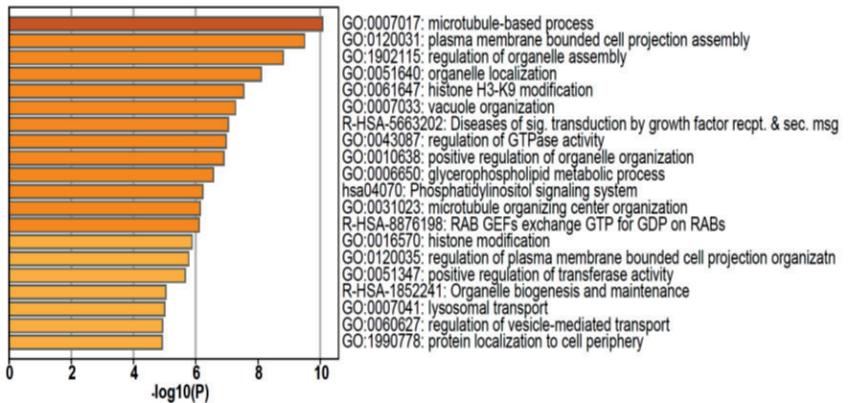


Figure S5. GO Molecular function analysis from RNA-Seq data. **A-B.** Differential expression analysis of *USO1* depleted SEM cells analyzed by GSEA with msigDB Hallmark gene sets and the GO Molecular function dataset, show enrichment of multiple RNA homeostasis related pathway. **C.** Venn diagrams showing overlap of *USO1*-negatively correlated genes in Target-Phase II ALL dataset with the genes that are significantly downregulated or upregulated in *USO1* depleted SEM cells. See Figure 4 for assumptions utilized in the Hypergeometric test. **D.** Enrichment analysis of the genes downregulated in *USO1* depleted cells that also have a high positive correlation with *USO1* using Metascape. The data shows enrichment of several cancer-relevant pathways.

Figure S6

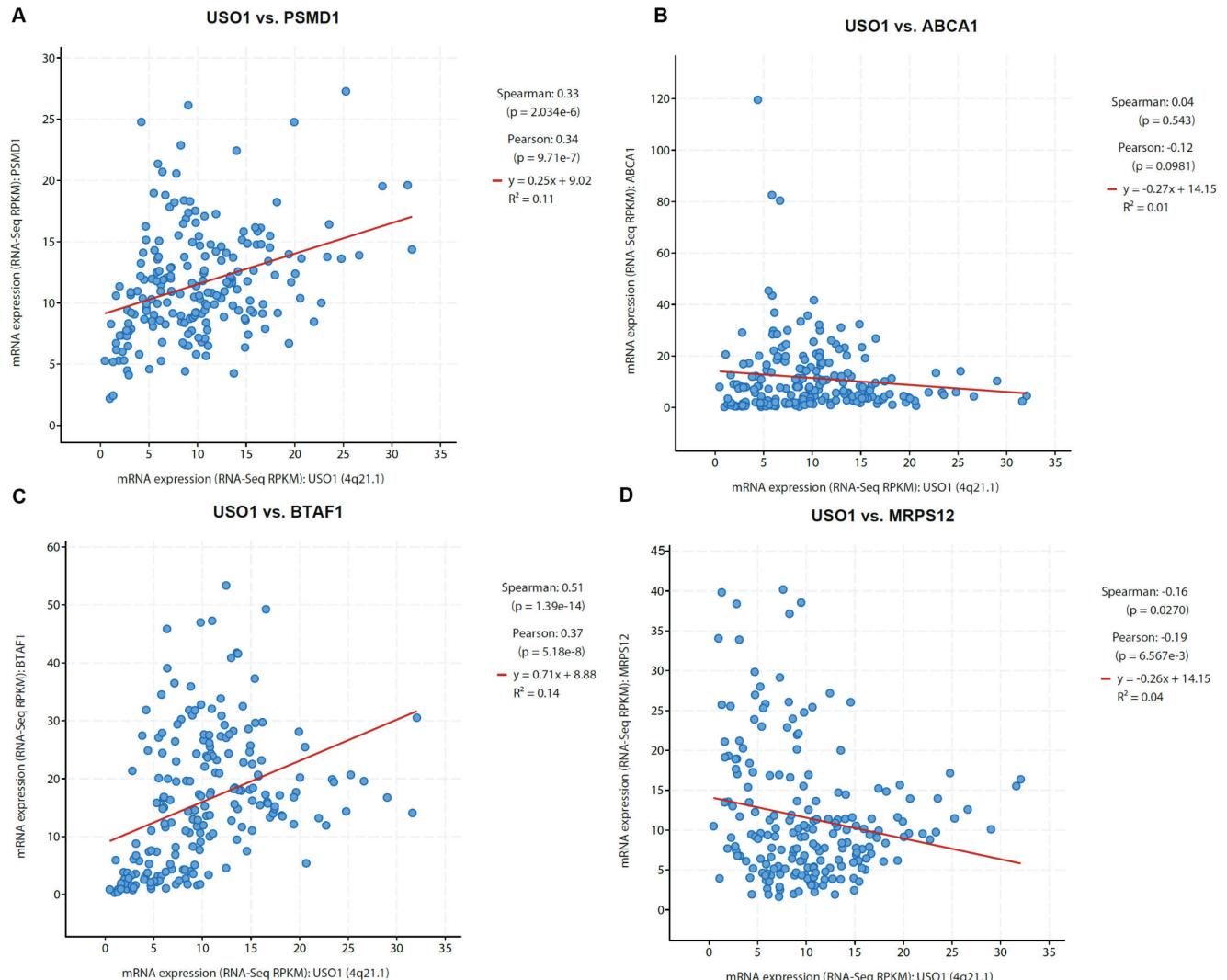


Figure S6. Correlation of USO1 with experimentally downregulated genes following USO1 knockdown. A-D. Genes downregulated in *USO1* depleted SEM, including *PSMD1*, *ABCA1*, *BTAF1* and *MRPS12* show a strong positive correlation with USO1 expression in clinical B-ALL samples (*Courtesy: cBioPortal*).

Figure S7a

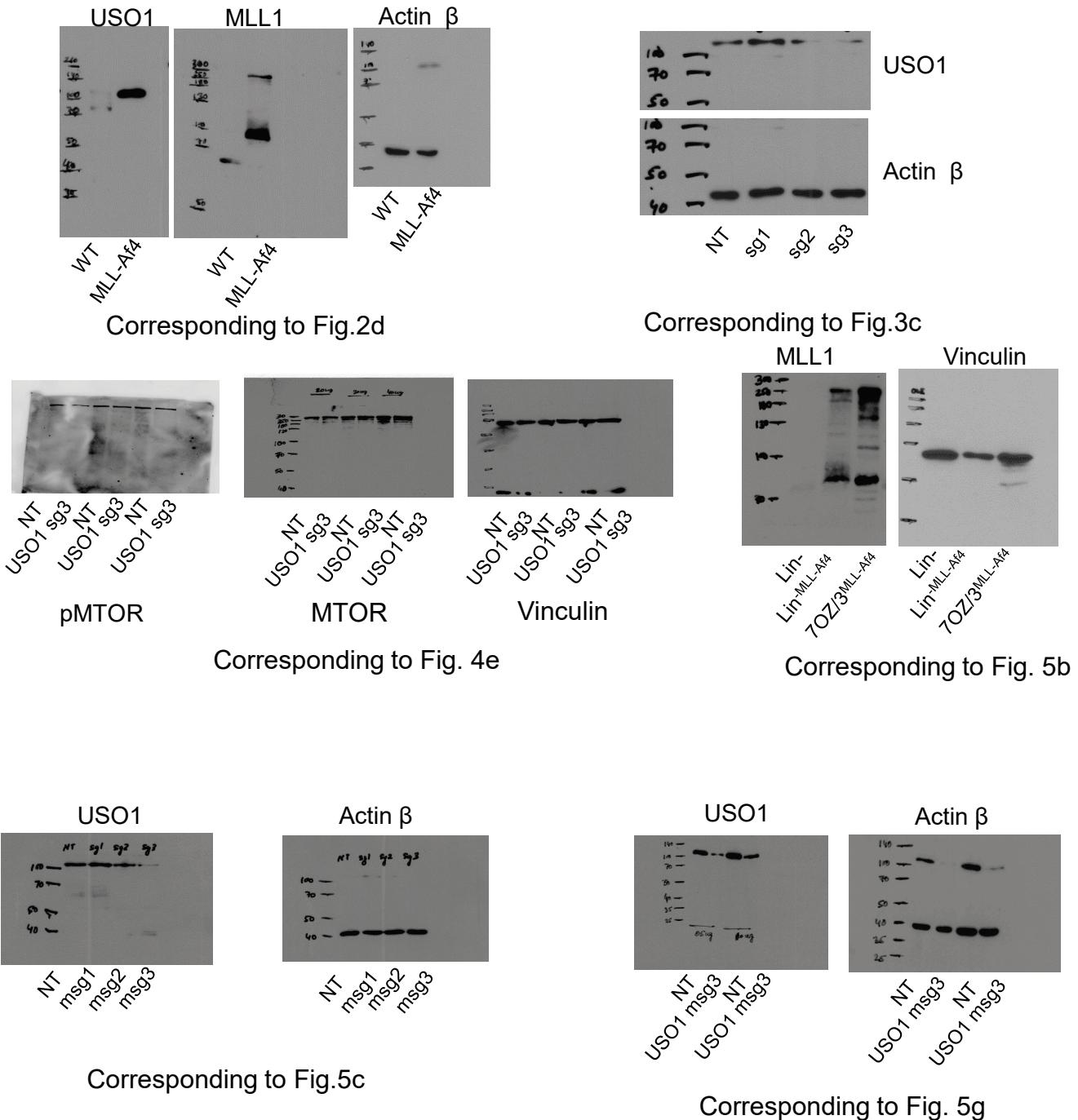
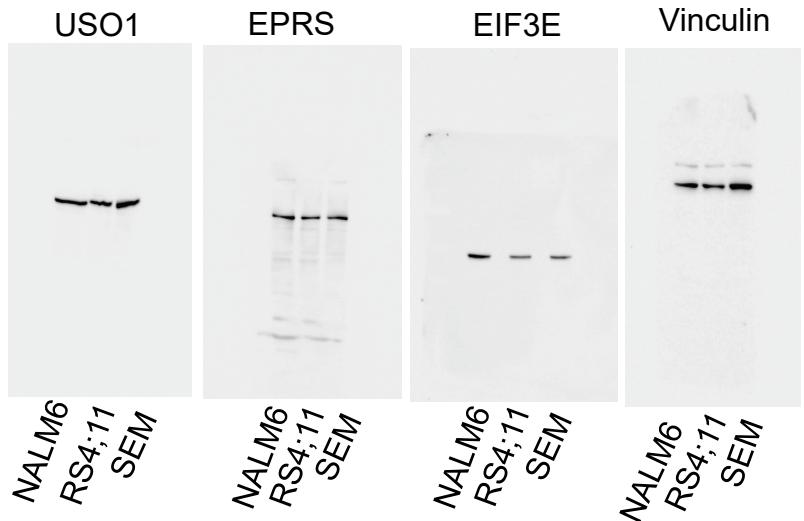
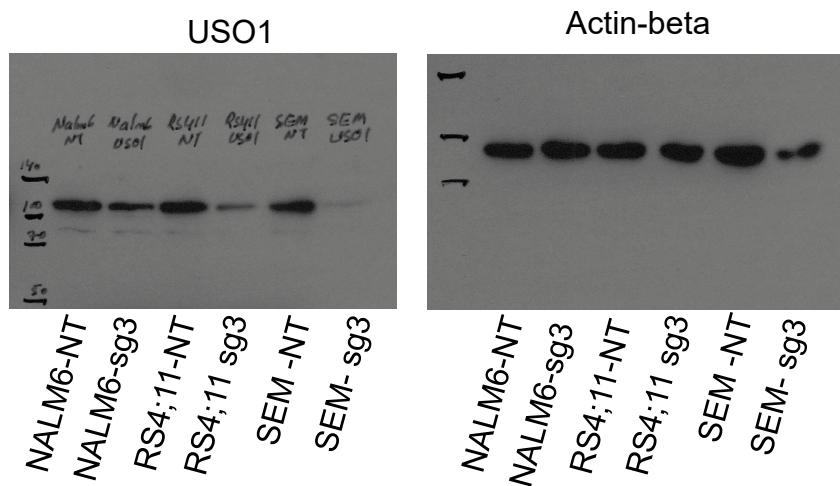


Figure S7a. Primary Western Blot Data. Each sub-panel lists the corresponding main panel in the main figures. These panels are the portion of the Western Blot scanned image that corresponds with the experiment being performed. In other cases, panels are showing the portion of a scanned image obtained by imaging on a ChemiDoc Imaging system from Biorad.

Figure S7b



Corresponding to Fig. S2b



Corresponding to Fig. S3b

Figure S7b. Primary Western Blot Data. Each sub-panel lists the corresponding main panel in the main figures. These panels are the portion of the Western Blot scanned image that corresponds with the experiment being performed. In other cases, panels are showing the portion of a scanned image obtained by imaging on a ChemiDoc Imaging system from Biorad.

Gene Name	Guide RNA sequence
IGF2BP3	AGCTTGGTCCTACTGGAAT
IGF2BP3	AAGCACCAAGATGCTAAAGTG
IGF2BP3	C CAC CGT AAA GAA AAT GCG G
IGF2BP3	A GGC GCA GAG GCA AAT CAC A
IGF2BP3	G CTG GGC GGC CAT TTC ATC A
BAZ1A	C AGT GCA TAA GCC AAA TCG G
BAZ1A	T GGT GTC AAA TCA TAC ACT G
BAZ1A	T CCA GTA TCG TCT ATA CAT G
BAZ1A	G TAA GCG CGA ACG ATG GGT A
BAZ1A	T TTG CTG ATG GAG TAA CCC T
USO1	A GGG TGA TAA GAT CGA CAG A
USO1	G GAC TTA CTA GCG GAT TCC A
USO1	A GCT ACT GGT AGC ACC AGG A
USO1	T CAG CTG GCC AGT TAT TAT G
DDX3Y	TATGAACACCACTACAAGGG
DDX3Y	C AGA TTC GGG ACT TAG AAC G
DDX3Y	C AGT TTA GCG ATA TTG ACA T
DDX3Y	A TGA TAA AGA CAG TTC AGG T
DDX3Y	G TAT ATA TCT GAC TCA GTA T
MBNL1	G TAT GTC GAG AGT ACC AAC G
MBNL1	CAACGTGGCAATTGCAACCG
MBNL1	C TTG GTG CAA CTG AAA ACA T
MBNL1	G TAA CCA ACA TTG GTG CAG T
MBNL1	T GCC CAA TAC CAG GTC AAC C
TUBB6	G CTG ACA ACG CCC ACC TAC G
TUBB6	C AGC TGG TGC ACC GAC AGT G
TUBB6	C CAT GGC CGC TAC CTG ACC G
TUBB6	C TGG CCC GGG AAG CGC AGC G
TUBB6	GTTGCAGGCCAGACGGGTGC
DDX21	C CTA CAT TCA TCG ATC CGG G
DDX21	G CAA CAT TAA ATA CCC AAT G
DDX21	G ATG TCA TCC GAG TAT ATA G
DDX21	C ATC ACA AAA AAG CTG TCA G
DDX21	CTAGTTGCCATTCTGTATG
LUC7L3	G TAG AAC GTA GGA TCA GAC G
LUC7L3	C ATC AAA TGG TCA TCT ACC C
LUC7L3	G TAT TTG TGA ACA ATT CCG C
LUC7L3	T TTA GAC GCT CAT CAC GAT C
LUC7L3	A ACT AAT TTC ATC ATC CCC T
LARP7	T GGA AGG CAC CAG AAT CCG G

LARP7	A TCA CTC CAC TCA CGA ACT G
LARP7	A GAC TAT CAG GAT CCG GAG A
LARP7	T GCT AGA TTT AAA ACT CCT G
LARP7	GAGAAGTTCAGCTGTTGTAG
IQGAP1	C TAC GCG TCC AAC CAG CGA G
IQGAP1	ATCAGCGGAGGTACCGACAG
IQGAP1	C CTG TCG AAC TAA GTA TCC A
IQGAP1	G AAT GAT ATC CAA GCT TGC G
IQGAP1	CCGTGGATACTTAGTTCGAC
CSTF3	C TCC TTC CTC ACC TGC TCC G
CSTF3	G CTG AGG CCA TGT CAG GAG A
CSTF3	C CCA GAG AAG GTG AAG AAA G
CSTF3	A GCT GAG TAT GTC CCA GAG A
CSTF3	GGTTTACACCCTGTACCTGG
EPRS	A ATG TAT ATC AAA ACA CAC G
EPRS	C ATG TTC ACG ACT GAA CGT G
EPRS	C TCC AGT TCA TGT AAA ACG T
EPRS	A GAA TGC TTA CTG TCC CTG A
EPRS	A GTA GAG TAT AAG CCT GTG T
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ANXA1	AGACCTTCGACGTGAATCGC
ANXA1	C CTT ACA GGT CAC CTT GAG G
ANXA1	A AAA TCT CCA GAT GTG TCT G
EIF3E	T ACA GCA GTC ATA ACA AAC A
EIF3E	T AAT ATT ATC GCG ACC TTT G
EIF3E	CTT CTT AGT GAT ACC AAC A
EIF3E	A ATG CTT TAA GTT CAC TCT G
EIF3E	C AAA ATT CCA GTA CGA ATG T
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CAMK2B	G TGC ATC ATG GAT GCT ACC G
CAMK2B	C TCT TTG CGA AGG ACC TCA G
PAICS	A TTG GAA TCA TTT CAC ACT G
PAICS	T ACG AAT TGT TAG ACA GTC C

PAICS	G CTA GAA AAA ACC ACC TGG A
PAICS	A TGC CAA TAA TGA CCC ACA G
PAICS	C TTA CGA TTT GAC TTA TAT G
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MSH2	C TGT CTC TGG CCA TCA ACT G
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RAD50	A AGC GGC GTG ATG AAA TGC T
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SKIV2L2	T TTGT CTT GTT ATG ACC ACA G
SKIV2L2	T TTC CAT TTG ACC TAC AAC A
SKIV2L2	A CCA AAT TAG ATT TCA ACA C
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SMARCAD1	T GCA TCA CCA AAC ATC AGC A
SMARCAD1	A CCT CCA TTT CCA GTA AGC A
OGT	T GGC AAT AGA CAC CTA CAG G
OGT	G CCA ATA TCA AAC GAG AAC A
OGT	T GGC ACA AAC TTC CGA GTG A
OGT	G GCA CGC GTA TAA CAC TGC A
OGT	C TAT ACA CTT CCA GTG TCG A
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C1QBP	GGACAGAAGCGAAATTAGTG
HSPD1P6	G AAA GCT TGA AGT TTG ATC G
HSPD1P6	G GTC TTA TCT AAA GAC AGT G
HSPD1P6	T AGC TAT GCC AAA GGT TGG T
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HSPD1P6	A TAA TCA CTG AAG ATG TTG G

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PSDM14	A TCA AAC CCA TAA CTT CCA T
PSDM14	A GAC AAA CAA CTT CGA ATC T
PSDM14	A GCC TTG TCG GAG AGA GCT G
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MYC	AACGTTGAGGGCATCGTCG
MYC	GCCGTATTCCTACTGCGACG
MYC	CGAGGAGAGCAGAGAACCG
MYC	TGCGTAGTTGTGCTGATGTG
HOXA7	CTCACAGAGAACGGCTACG
HOXA7	GCCTGCGACAAGACGGACGA
HOXA7	CAAAGGGCTCTGATAAAGG
HOXA7	TACCTGAAGACCGCATCCAG
HOXA7	GGAGCAAAGGAGCAAGAAGT
MEIS1	GGTGGCCACACGTACACAG
MEIS1	ATGCGGGTCCCCATACATCG
MEIS1	CACAGCTCATACCAACGCCA
MEIS1	ATCATGATCTCTGTTCCAAG
MEIS1	CTTTAAAGCGTCATTGACAG
CDK6	GCCC CGCGACTTGAAGAACGG
CDK6	AACACTCCAGAGATCCACGG
CDK6	GGAAACTATAGATGCGGGCA
CDK6	TGGCTCACCTGACCACGTTG
CDK6	CCAGCAGTACGAATGCGTGG
HOXA9	CAAAGTGTGAGTGTCAAGCG
HOXA9	GCGTTGGCCGCTATGCGCCG
HOXA9	CGACGGTGTGGCGCCCTCG
HOXA9	ATGGTGGTGGTACACCGCAG
HOXA9	CAGGTTTAATGCCATAAGGC
CoA5	CATAGGAAGGAAAATCACCT
CoA5	TATTATGAGGACAAGCCGCA
CoA5	CGTCATGCCTAAGTATTATG
CoA5	TCTGCTGCAGTCGGACTGTG
CoA5	CGGCGCGTGCACGGGGCCTGA
RPL11	TGTCCACTGCACAGTTCGAG
RPL11	GGATGCGAAGTCCCCGCATG
RPL11	CTCACCTTGAAAACACAG
RPL11	AACTCATACTCCCCGACACTG
RPL11	TTCATTCTCCGGATGCCAA
SRSF2	AACGACTCCGACTCCGGGAT

SRSF2	TCGAGCGGCTGTAGCGAGAT
SRSF2	CTTGTGCGTAAAGCGAACGA
SRSF2	GGAGGGTATGACCTCCCTCA
SRSF2	CGGGTGCAAATGGCGCGCTA
RPS27	TTGTGCATGGCTAAAGACCG
RPS27	CTGTCCCTCTTAGCTCGCAA
RPS27	TTCCTCCTGAAGGAACATCC
RPS27	TTCCTCCAGGATGTTCCCTTC
RPS27	CTCCAGGATGTTCCCTTCAGG
HEATR1	ATTGAACGTGTTAGACAG
HEATR1	ACAACAAAGGAACCTTACGC
HEATR1	CAGGAACATTACATAAGTGA
HEATR1	ATACAACATCTAATGTTCTG
HEATR1	GCCCCGTAAGAAGTCGT
HIST1H2AH	GCTCGGCATAATTACCCTTG
HIST1H2AH	GAAGCCCAGCCGAGAAGAG
HIST1H2AH	TCTGGACGTGGCAAGCAAGG
HIST1H2AH	GGCTGGCTTCAGTCCCCG
HIST1H2AH	GAGGTGACGCCGGATGATAC
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Atr(LOC648152)	CTGTGTGAGATGGTCAAGCA
Atr(LOC648152)	AGTCAATTAGATGAACACAT
Atr(LOC648152)	AGAACCTATTATCTCACAGT
Atr(LOC648152)	TAATTATAGAATTAAACATG
STAT5A_1	GCACTACGCCACGCAGCTCC
STAT5A_2	TACACCCACCTGGAGCTGCG
STAT5A_3	ACCAGGACTGTAGCACGTCC
STAT5A_4	CATTGACTTGGACAATCCCC
STAT5A_5	CGTGCACATGAATCCCCCC
BCL-2	GGCCTCTTGAGTTGGTG
BCL-2	TGGACATCTCGCGAAGTCG
BCL-2	TGTCGCAGAGGGGCTACGGAG
BCL-2	CTGACGCCCTCACCGCGCG
BCL-2	GCCCCATCCAGCCGCATCCC
NT1	ACGGAGGGCTAACGCGTCGCAA
NT2	CGCTTCCGCGGGCCGTTCAA
NT3	ATCGTTCCGCTAACGGCG
NT4	GTAGGGCGCGCCGCTCTAC
NT5	CCATATCGGGCGAGACATG
NT6	TACTAACGCCGCTCCTACAG

NT7	TCATCTTACATCTGGGAGAC
NT8	GGGCCCGCATAGGATATCGC
NT9	TAGACAACCGCGGAGAATGC
NT10	ACGGGCGGCTATCGCTGACT
NT11	CGCGGAAATTTACCGACGA
NT12	CTTACAATCGTCGGTCCAAT
NT13	GCGTGCCTCCGGGTTACCC
NT14	CGGAGTAACAAGCGGACGGA
NT15	CGAGTGTATACGCACCGTT
NT16	CGACTAACCGGAAACTTTTT
NT17	CAAGTTTCTGAAAGGCAAT
NT18	AAAGATATAGCAAATTATGG
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NT20	GTGTCGGATTCCGCCGCTTA
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NT22	ACTTCCCACTTCTTAGGTTG
NT23	CCATTACAATCCCACTACA
NT24	TGTTTGATGTTGCATAGG
NT25	TAAGCCTCATGAAGGAGGGG
NT26	GGTACTGGAAGTCCGAAACC
NT27	AAATAATATGCATCTCTCGA
NT28	AGCTGGACTCTGTAGAAATC

Table S1. Short guide RNA (sgRNA) sequences.

Genes	Primer Sequence
USO1-Forward Primer	TACCGCTTGGAAAGTGGGTAT
USO1-Reverse Primer	AAATCTCACTCTGTCTTGTGGAAT
EPRS- Forward Primer	GGG AAG GCT TAT GTG GAT GA
EPRS- Reverse Primer	CTC GCA AAC AAC AGG ACT
EIF3E-Forward Primer	CCCTCACCTCCTTACAGAG
EIF3E-Reverse Primer	GCACAGACTCCCTTCTTT
MRSP12-Forward Primer	CCTCTTCATCAGGACCCT
MRSP12-Reverse Primer	CAGTGTCTTGTCCCAGCAG
TRFC-Forward Primer	TGGACAGCACAGACTTCACC
TRFC-Reverse Primer	CCAGGATTCTCCACCAAGG
NOP16-Forward Primer	AGGGTGTAAGGCAAGGAGGT
NOP16-Reverse Primer	ATCACCCACCCCTGAAAACA
DDX21-Forward Primer	GCAGCAGTTATTGGGGATGT
DDX21-Reverse Primer	TGAATTCTGGGACAGCTCCT
PSMD1-Forward Primer	CTGTGGTTGGCGTCCTGTAT
PSMD1-Reverse Primer	CAAATGTGGATGGTTACAG
Human 18s Forward Primer	GTAACCCGTTGAACCCCATT
Human 18s Reverse Primer	CCATCCAATCGGTAGTAGCG
Uso1-Forward Primer	CCAGGGAAGCAAAATACAGA
Uso1-Reverse Primer	GCACATAAGCCTGGACC
L-32 Forward Primer	AAG CGA AAC TGG CGG AAA C
L-32 Reverse Primer	TAA CCG ATG TTG GGC ATC AG

Genes	ChIP primer sequence
USO1-Fw	TGGCTAACGGCAAGATGAAT
USO1- Rev	CTGGGATTCAGGAGCGGA

Genes	sgRNA sequence
Ms-NT	GCGAGGTATTGGCTCCGCG
Uso1-1	GGTAACGTAGAGGAAGCTACG
Uso1-2	GTCGGTCATCCAGTAACGTAG
Uso1-3	GACTGCTGGACATTATTACAG

CRISPR Lib. primers	Sequence
Part I Illumina Universal Adaptor with variable region and vector seq.	ACACTCTTCCCTACACGACGCTCTCCGATCTT AA TGGAAAGGACGAAACACCGA
PartII Illumina Index adaptor	GACTGGAGTTCAGACGTGTGCTCTCCGATCTT TTAGTTGTATGTCTGTTG
Rest Illumina universal adaptor	AATGATAACGGCGACCACCGAGATCTACACTCTT TCCCTACACGACGC
Rest Illumina Index adaptor	CAAGCAGAAGACGGCATACGAGATCGTGATGT GACTGGAGTTCAGACGTGTGC

Table S2. Primer sequences. Please see text for details.