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Richa Rathore

Brian A Van Tine

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Review

## Pathogenesis and Current Treatment of Osteosarcoma: Perspectives for Future Therapies

Richa Rathore <sup>1</sup> and Brian A. Van Tine <sup>1,2,3,\*</sup>

- Division of Medical Oncology, Washington University in St. Louis, St. Louis, MO 63110, USA; richarathore@wustl.edu
- Division of Pediatric Hematology and Oncology, St. Louis Children's Hospital, St. Louis, MO 63110, USA
- Siteman Cancer Center, St. Louis, MO 63110, USA
- Correspondence: bvantine@wustl.edu

**Abstract:** Osteosarcoma is the most common primary malignant bone tumor in children and young adults. The standard-of-care curative treatment for osteosarcoma utilizes doxorubicin, cisplatin, and high-dose methotrexate, a standard that has not changed in more than 40 years. The development of patient-specific therapies requires an in-depth understanding of the unique genetics and biology of the tumor. Here, we discuss the role of normal bone biology in osteosarcomagenesis, highlighting the factors that drive normal osteoblast production, as well as abnormal osteosarcoma development. We then describe the pathology and current standard of care of osteosarcoma. Given the complex heterogeneity of osteosarcoma tumors, we explore the development of novel therapeutics for osteosarcoma that encompass a series of molecular targets. This analysis of pathogenic mechanisms will shed light on promising avenues for future therapeutic research in osteosarcoma.

**Keywords:** osteosarcoma; mesenchymal stem cell; osteoblast; sarcoma; methotrexate



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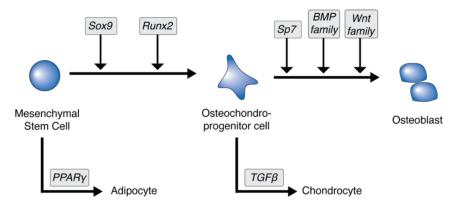
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## 1. Introduction

Osteosarcomas are the most common pediatric and adult bone tumor, with more than 1000 new cases every year in the United States alone. Osteosarcomas arise from mesenchymal cells and are characterized by areas of abnormal bone growth [1]. The various genetic, epigenetic, and environmental factors that drive mesenchymal stem cells to differentiate into bone precursor cells also play a role in the development of osteosarcoma. These molecular pathways can serve as the foundation for the development of new therapies for this tumor [2]. This review describes the basic biology of bone, and how the systems that drive bone development lead to osteosarcomagenesis. Furthermore, the cellular pathways that contribute to the pathogenesis of the tumor are explored, and this information is used to describe the avenues for novel treatment development for osteosarcoma.

incorporate blood vessels. Transforming growth factor-beta (TGF $\beta$ ) expression can further drive differentiation into chondrocytes, but also stimulates alkaline phosphatase (ALP) activity and calcium deposition [1,7].



**Figure 1.** Mesenchymal stem cells differentiate into various cell types based on the expression of different transcription factors and protein families, highlighted in gray. Sox9: SRY-box transcription factor 9; Runx2: Runt-related transcription factor 2; Sp7: osterix; BMP: bone morphogenic protein; Wnt: wingless and int-1; PPAR $\gamma$ : peroxisome proliferator-activated receptor gamma; TGF $\beta$ : transforming growth factor-beta.

Runx2 is a transcription factor that drives the expression of a series of genes related to osteogenesis [8]. Runx2 increases the expression of osterix (Sp7), which is required to commit osteochondroprogenitor cells to osteoblast differentiation, as well as osteocalcin, Type I collagen, and ALP to stimulate osteoblast formation [9–11]. Finally, Runx2 induces the expression of the CDK2 inhibitor p27KIP1, which coordinates G1 cell-cycle arrest in osteoblasts, a process necessary for normal development of bone. Importantly, the expression of Runx2 and ALP decreases as cells differentiate into osteoblasts, and low Runx2 expression is required for normal osteoblast function [8].

Bone morphogenic proteins (BMPs) comprise a family of over 30 different proteins, including TGF $\beta$  family members, that also regulate mesenchymal stem cell differentiation into osteoblasts by activating and inhibiting several genes that affect the expression of Runx2 and Sp7 [9]. Wingless and int-1 (Wnt) signaling proteins and fibroblast growth factors (FGF) further contribute to this regulation [7,9].

Once osteoblasts have been derived, they coordinate with osteoclasts to model and remodel bone, thereby maintaining bone homeostasis. Osteoblasts are the bone-forming cells, which create cartilage using calcium which is then hardened into bone. Osteoclasts, which are derived from hematopoietic stem cells, are the bone-resorbing cells, which break down bone using electrolytes and bone-degrading enzymes. During development, the body models bone by removing bone from some areas and synthesizing bone in others.

#### 3. Bone Transformation to Osteosarcoma

#### 3.1. Bone Cancer and Sarcoma

Cancer is defined as a disease of abnormal cells that acquire certain capabilities that drive unchecked, uncontrolled, and invasive growth and division [13,14]. Cancers are grouped into several categories, including carcinomas, sarcomas, myelomas, leukemias, and lymphomas. Carcinomas, which arise from epithelial cells, comprise approximately 90% of human cancers. Sarcomas, which have a mesenchymal cell of origin, consist of only 1% of adult cancers. Given that the bone consists of a series of cell types that originate from mesenchymal stem cells, the tumors that arise in bone all fall into the sarcoma category [15].

There are a series of other bone sarcomas, including Ewing's sarcoma, chondrosarcoma, hemangiosarcoma, giant cell tumor, chordoma, and the soft tissue sarcomas of bone [15,16]. There are approximately 3600 new bone cancer cases every year [17]. Osteosarcomas are the most common bone tumor, consisting of 40–50% of bone sarcomas.

## 3.2. Osteosarcoma Cell of Origin

Osteosarcomagenesis was originally classified as occurring only from mesenchymal stem cells, though more recent data suggest that osteosarcomas can form at multiple points in bone development, from both mesenchymal stem cells and osteoblasts, as well as dysregulated osteoclasts (Figure 2) [18–20]. Unlike many other sarcomas which are driven by genetic translocations, such as synovial sarcoma or Ewing's sarcoma, osteosarcomas have complex karyotypes [6,21]. Even so, it is widely understood that alterations to TP53 and RB1 tumor suppressor genes play a role in osteosarcoma, as in the development of several other cancers [3,22]. It has also been demonstrated that, once committed to the osteogenic lineage, MSCs with p53 and Rb excised develop into osteosarcoma-like tumors, further demonstrating the oncogenic potential of mutations to these genes [23].

Genes that relate to osteoblast development have also been associated with osteosar-comagenesis (Figure 2). Wnt protein family members have been identified as playing a significant role in the development of osteoblasts from mesenchymal stem cells [9,24]. Aberrant activation of Wnt family members can drive the further progression of osteoblasts into osteosarcoma. In fact,  $\beta$ -catenin, a mediator of Wnt family signaling, has been demonstrated to be expressed in a large percentage of osteosarcoma tumors [25].

BMP/TGF $\beta$  family members that drive osteoblast development can also drive osteosarcoma development. Interestingly, osteosarcoma tumors tend to express higher amounts of TGF $\beta$ 1 and TGF $\beta$ 3, which have been associated with disease progression [6]. TGF $\beta$  also activates SMAD proteins, which can inhibit osteoblast differentiation by decreasing the expression of osteocalcin [26,27]. Smad4 gene mutations have been identified in several cancers, including pancreatic and ovarian cancer, and SMAD proteins have also been identified as being dysregulated in osteosarcoma [28].

The elevated expression of Runx2, one of the main drivers of osteoblast formation from osteochondroprogenitor cells through the coordinated activation of osteocalcin, Type I collagen, and ALP, has been shown to drive osteosarcomagenesis [10,29]. Runx2 has

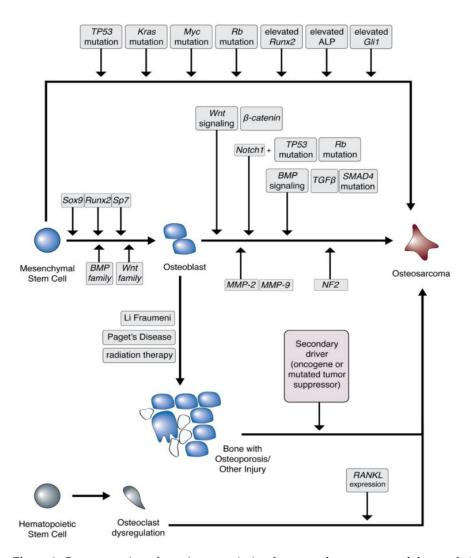


Figure 2. Overexpression of certain transcription factors and oncogenes and dysregulation of tumor suppressor genes can drive osteosarcoma development. TP53: tumor protein p53; Rb: retinoblastoma; Runx2: Runt-related transcription factor 2; ALP: alkaline phosphatase; Gli1: glioma-associated oncogene homolog 1; Sox9: SRY-box transcription factor 9; Sp7: osterix; BMP: bone morphogenic protein; Wnt: wingless and int-1; TGF $\beta$ : transforming growth factor-beta; SMAD4: Mothers against decapentaplegic homolog 4; MMP: matrix metalloproteinase; NF2: neurofibromatosis-2; RANKL: receptor activator of nuclear-factor kappa B ligand.

Gli1, an oncogene that drives the sonic hedgehog (Shh) signaling pathway, has been

neurofibromatosis-2 (NF2) have been correlated with increased incidence of several highly metastatic tumors, including osteosarcoma [6]. The protein encoded for by NF2, Merlin, has been demonstrated to stabilize p53; therefore, in patients with NF2 alterations, p53 is also affected, and can thereby drive incidence and malignancy of osteosarcoma [6,10].

#### 3.3. Bone Microenvironment

The signaling components of the bone microenvironment play a critical role in osteosarcoma development. BMP2 and TGFβ circulate throughout the bone microenvironment, contributing to osteoblast formation but also osteosarcoma differentiation and malignancy [38,39]. Growth-related factors can also contribute to sarcomagenesis as these factors are necessary for osteoblast-driven bone formation [40]. Chondrocytes secrete high-mobility group box 1 protein (HMGB1) that stimulates osteoblast proliferation and can induce osteosarcoma proliferation [3,41].

Factors secreted throughout the bone microenvironment also contribute to abnormal osteoclast activity, which can result in osteosarcoma. As previously noted, osteoclasts are regulated by RANK signaling, which is mediated by RANKL expression on osteoblasts. Dysregulation of RANKL expression and ligand binding by osteoblasts and other cells in the bone microenvironment can limit bone resorption by osteoclasts and allows bone to form unchecked. Factors released by cancer cells including interleukins (IL) such as IL-6 and IL-11, as well as  $TGF\beta$ , can also modulate RANK expression on the osteoclast surface that can further decrease bone resorption and contribute to tumor progression [3].

In addition to being the precursor for osteoblasts, chondrocytes, and osteosarcoma, mesenchymal stem cells themselves also play a role in tumor progression. The cytokines secreted by mesenchymal stem cells in the bone microenvironment, including TGF $\beta$  and tumor necrosis factor  $\alpha$  (TNF $\alpha$ ), can inhibit lymphocyte proliferation and block the response of the immune system, allowing the tumor to escape the inflammatory response [42]. Mesenchymal stem cells can also promote angiogenesis through differentiation into fibroblasts and producing growth factors, thereby improving blood supply to the tumor [43]. Finally, various factors released from mesenchymal stem cells, including TGF $\beta$ , E-cadherin, and micro-RNAs, have been demonstrated to upregulate the epithelial-to-mesenchymal (EMT) transition, resulting in a more invasive phenotype [43,44].

Primary bone cancers are not the only cancers that thrive in the bone microenvironment. Many cancers metastasize to the bone because of its rich tumor-promoting environment, including breast cancer, prostate cancer, and other carcinomas [45]. Though osteosarcoma is a bone-producing tumor, various bone-metastatic breast cancers have been identified as contributing to osteolysis, or the destruction of bone tissue [46]. Enhanced production of the amino acid serine by breast cancer has been attributed to osteoclastogenesis and increased osteolysis due to bone metastases [47].

## 3.4. Osteosarcoma Predisposition

There are several genetic syndromes that predispose patients to developing osteosar-

Various external factors have also been identified as risk factors for osteosarcoma. As early as two and as late as 20 years after radiation therapy exposure, radiation-induced osteosarcomas have been observed; some tumors have arisen at and around radiation sites decades after initial therapy [21]. SV40 viral DNA has also been identified in as much as 50% of osteosarcoma tumors; however, there are no data to verify whether this has any causative role in osteosarcoma development [6,53,54].

## 4. Osteosarcoma Epidemiology and Diagnosis

Osteosarcoma is the most common primary pediatric and adult bone tumor [55]. Over 1000 new cases arise each year in the United States [6,17]. Approximately 80% of osteosarcomas present with a localized, primary tumor, with the other 20% presenting initially with pulmonary metastases [56]. For patients with metastatic disease, the overall survival rate is less than 20% [56].

Osteosarcomas are bone-forming tumors that occur primarily at the metaphysis of the bone, in regions of rapid bone growth [57]. Approximately 80% of osteosarcomas occur in the extremities, primarily in the proximal tibia, distal femur, and proximal humerus [56,58]. Clinically, most osteosarcoma patients present with pain, usually with swelling or a palpable mass identified at the site of the pain [56]. Histologic diagnosis of osteosarcoma is based on morphology as identified by radiograph [16,59]. There are six subtypes of osteosarcoma, including low-grade central osteosarcoma, osteosarcoma not otherwise specified (NOS), parosteal, periosteal, high-grade surface, and secondary osteosarcoma [57]. Within the conventional osteosarcomas are various classes of tumor based on location and originating cell, including osteoblastic, chondroblastic, and fibroblastic osteosarcomas. Osteoblastic osteosarcoma tend to make up the majority of tumors (approximately 70%); however, most osteosarcomas are genetically and morphologically heterogenous, so tumors can contain any combination of these three classes [6,16].

Depending on tumor location and stage, neoadjuvant chemotherapy with the MAP (methotrexate, doxorubicin, and cisplatin) regimen is the initial step of osteosarcoma treatment [56]. After management with resection and adjuvant chemotherapy, the cure rate of osteosarcoma is approximately 60–70% [56,58,60]. There is an association between having greater than 90% tumor necrosis after chemotherapy and overall survival [61]. Approximately 30% of patients do relapse after surgery and chemotherapy, generally within five years, at which point lung and bone metastases are the most common sites of recurrence [6,56].

## 5. Treatment Strategies and Molecular Targets

## 5.1. Current Standard of Care

Current treatment strategies for osteosarcoma are neoadjuvant chemotherapy with cisplatin, doxorubicin, ifosfamide, and high-dose methotrexate with leucovorin rescue, followed by surgical resection and adjuvant chemotherapy [58]. Cisplatin is an antineoplastic alkylating agent that causes DNA damage. The platinum ion in cisplatin forms

with the topoisomerase II enzyme, causing apoptosis. The combination of ifosfamide and etoposide has limited the toxicity of ifosfamide alone [64]. Ifosfamide has been approved for use in testicular cancer, osteosarcoma, soft tissue sarcoma, bladder cancer, non-small-cell lung cancer, cervical cancer, and ovarian cancer, amongst others [64,67].

Unlike cisplatin, doxorubicin, and ifosfamide, which all function by damaging DNA and inhibiting cellular division, methotrexate targets dihydrofolate reductase (DHFR), an enzyme in the folate cycle and a key metabolic component of nucleotide biosynthesis [68]. DHFR is a cellular source of tetrahydrofolate (THF), recycling THF from dihydrofolate (DHF) [69,70]. THF is required in the biosynthesis of purines and thymidylate from serine. The structure of methotrexate is similar to DHF, allowing the drug to competitively inhibit DHFR and block recycling of THF. In osteosarcoma, methotrexate is given as high doses, defined as >1 g/m² [71]. In order to facilitate relatively safe use of high-dose methotrexate (HD-MTX), leucovorin rescue is used to block import of methotrexate into healthy cells. Leucovorin supplies normal cells with an additional source of THF and can thus counter the activity of methotrexate [72–74]. Cancerous cells lack the leucovorin transporter and are therefore susceptible to inhibition of the folate cycle by HD-MTX [75,76]. This allows the doses of HD-MTX used in osteosarcoma patients to reach doses as high as 8–12 g/m².

Even with leucovorin rescue, HD-MTX treatments still exhibit high rates of toxicity, and can lead to renal and liver failure, particularly in adults, as well as leukoencephalopathy, or damage to the white matter of the brain [68,72,77]. Due to this extremely narrow therapeutic window, an alternative—or ideally, replacement—therapeutic for HD-MTX would be beneficial.

## 5.2. Clinical Trials: The Future of Osteosarcoma Treatment

The treatment landscape for osteosarcoma in the curative setting has not evolved since the introduction of high-dose methotrexate to the standard cytotoxic chemotherapies. The palliative setting is clinical trial-rich, with over 500 clinical trials targeting novel pathways of interest in osteosarcoma. Select small-molecule inhibitor and clinically relevant immunotherapy-based clinical trials in osteosarcoma are highlighted in Table 1. Interestingly, most of the trials currently active and/or recruiting focus on combination therapies in osteosarcoma, highlighting the need for treatment strategies that exploit cellular pathways that drive specific prognostic factors in osteosarcoma, rather than neutral combinations of DNA damage agents that have activity-and side effects—across a number of cancers.

## 5.3. Targeting p53 and RB

Genetic abnormalities in *RB1* have been found in up to 70% of osteosarcoma tumors, and *TP53* mutations have been associated with approximately 90% of osteosarcoma tumors [6,22]. P53 is a tumor suppressor protein that can regulate metabolic reprogramming by acting as a sequence-specific transcription factor and altering transcription of various metabolic enzymes [78]. Mutations to *TP53* can cause structural and functional changes to the resultant mutant p53 protein, resulting in either a tumorigenic gain of function of the

**Table 1.** Current and ongoing clinical trials in osteosarcoma using molecular targeted therapies.

Identifier	Study Title	Status
NCT00470223	Combined chemotherapy with or without zoledronic acid for patients with osteosarcoma	Active, not recruiting
NCT00788125	Dasatinib, ifosfamide, carboplatin, and etoposide in treating young patients with metastatic or recurrent malignant solid tumors	Active, not recruiting
NCT01459484	ABCB1/P-glycoprotein expression as biologic stratification factor for patients with non metastatic osteosarcoma (ISG/OS-2)	Active, not recruiting
NCT01661400	Anti-angiogenic therapy post transplant (ASCR) for pediatric solid tumors	Recruiting
NCT01669369	Clinical trial of lithium carbonate combined with neo-adjuvant chemotherapy to treat osteosarcoma (Li2CO3)	Recruiting
NCT01833520	Phase I dose escalation of monthly intravenous Ra-223 dichloride in osteosarcoma	Active, not recruiting
NCT01953900	iC9-GD2-CAR-VZV-CTLs/refractory or metastatic GD2-positive sarcoma and neuroblastoma	Active, not recruiting
NCT02013336	Phase 1 study of MM-398 plus cyclophosphamide in pediatric solid tumors	Recruiting
NCT02173093	Activated T cells armed with GD2 bispecific antibody in children and young adults with neuroblastoma and osteosarcoma	Recruiting
NCT02243605	Cabozantinib S-malate in treating patients with relapsed osteosarcoma or ewing sarcoma	Active, not recruiting
NCT02357810	Pazopanib hydrochloride and topotecan hydrochloride in treating patients with metastatic soft tissue and bone sarcomas	Active, not recruiting
NCT02389244	A Phase II study evaluating efficacy and safety of regorafenib in patients with metastatic bone sarcomas	Recruiting
NCT02406781	Combination of MK3475 and metronomic cyclophosphamide in patients with advanced sarcomas: multicentre phase II trial	Recruiting
NCT02432274	Study of lenvatinib in children and adolescents with refractory or relapsed solid malignancies and young adults with osteosarcoma	Active, not recruiting
NCT02470091	Denosumab in treating patients with recurrent or refractory osteosarcoma	Active, not recruiting
NCT02484443	Dinutuximab in combination with sargramostim in treating patients with recurrent osteosarcoma	Active, not recruiting
NCT02502786	Humanized monoclonal antibody 3F8 (Hu3F8) with granulocyte-macrophage Colony stimulating factor (GM-CSF) in the treatment of recurrent osteosarcoma	Recruiting
NCT02517918	Metronomic chemotherapy in patients with advanced solid tumor with bone metastasis and advanced pretreated osteosarcoma	Recruiting
NCT02811523	In vivo lung perfusion for pulmonary metastases of sarcoma	Recruiting
NCT02867592	Cabozantinib-S-Malate in treating younger patients with recurrent, refractory, or newly diagnosed sarcomas, wilms tumor, or other rare tumors	Active, not recruiting
NCT02945800 NCT03006848	Nab-paclitaxel and gemcitabine for recurrent/refractory sarcoma A phase II trial of avelumab in patients with recurrent or progressive osteosarcoma	Recruiting Active, not recruiting
NCT03063983	Clinical trial evaluating metronomic chemotherapy in patients with metastatic osteosarcoma (GLATO2017)	Recruiting
NCT03277924	Trial of sunitinib plus nivolumab after standard treatment in advanced soft tissue and bone sarcomas	Recruiting
NCT03449108	LN-145 or LN-145-S1 in treating patients with relapsed or refractory ovarian cancer, anaplastic thyroid cancer, osteosarcoma, or other bone and soft tissue sarcomas	Recruiting
	ADI-PEG 20 in combination with gemcitabine and docetaxel for the treatment of soft	

Table 1. Cont.

Identifier	Study Title	Status
NCT03742193	Pulmonary resectable metastases of osteosarcoma with apatinib and chemotherapy Study of the safety and efficacy of humanized 3F8 bispecific antibody (Hu3F8-BsAb)	Recruiting
NCT03860207	in patients with relapsed/refractory neuroblastoma, osteosarcoma and other solid tumor cancers	Recruiting
NCT03900793	Losartan + sunitinib in treatment of osteosarcoma	Recruiting
NCT03932071	Zoledronic acid in decrease the lung metastatic rate of osteosarcoma	Recruiting
NCT03960177	Glucarpidase after high-dose methotrexate in patients with osteosarcoma	Recruiting
NCT04040205	Abemaciclib for bone and soft tissue sarcoma with cyclin-dependent kinase (CDK) pathway alteration	Recruiting
NCT04055220	Efficacy and safety of regorafenib as maintenance therapy after first-line treatment in patients with bone sarcomas	Recruiting
NCT04154189	A Study to compare the efficacy and safety of ifosfamide and etoposide with or without lenvatinib in children, adolescents and young adults with relapsed and refractory osteosarcoma	Recruiting
NCT04183062	BIO-11006 for osteosarcoma and ewing's sarcoma lung metastases	Recruiting
NCT04294511	Study of camrelizumab in combination with neoadjuvant chemotherapy in the treatment of osteosarcoma	Recruiting
NCT04351308	Comparison of MAPI + camrelizumab versus API + apatinib versus MAPI in patients with a poor response to preoperative chemotherapy for newly diagnosed high-grade osteosarcoma	Recruiting
NCT04383288	ABCB1/P-glycoprotein expression influence on non-metastatic osteosarcoma of the extremities	Recruiting
NCT04433221	Combination immunotherapy targeting sarcomas	Recruiting
NCT04483778	B7H3 CAR T cell immunotherapy for recurrent/refractory solid tumors in children and young adults	Recruiting
NCT04595994	Selinexor plus gemcitabine in selected advanced soft-tissue sarcoma and osteosarcoma	Recruiting

Importantly, while *TP53* and *RB* are the most common alterations in osteosarcoma, these mutations are not directly targetable in the clinic due to the complex nature of their incorporation into normal and cancerous cell biology. Mutant p53-reactivating compounds are being explored as a possible mechanism by which to selectively target mutations to *TP53*, which is encouraging for the field [85]. However, identifying and targeting the pathways driven by altered *TP53* or *RB* would allow for more directed approaches.

## 5.4. Gemcitabine and Docetaxel

Gemcitabine is a fluorinated version of the nucleoside deoxycytidine that is taken up by the nucleoside transporter SLC29A1, or human equilibrative nucleoside transporter 1 (hENT1) [86,87]. Gemcitabine incorporates into DNA as a fraudulent base pair, resulting in premature DNA strand termination [86]. Docetaxel is a mitosis inhibitor that functions by stabilizing tubulin [86]. Gemcitabine and docetaxel have been combined in multiple

ABCB1 expression to characterize the role of ABCB1 expression in MDR and osteosar-coma patient outcomes (NCT identifier: NCT01459484 and NCT04383288). CRISPR/Cas9 mediated inhibition of ABCB1 may also help to combat MDR in osteosarcoma [91].

## 5.6. RANK Ligand Antibodies

RANK signaling has been demonstrated to be important in osteosarcoma development, growth, and motility, and the overexpression of RANK and RANKL has been correlated with poorer outcomes [58]. Osteoblast-secreted RANKL has therefore been explored as a possible target for antibody-based therapies, as inhibiting RANK signaling on osteoclasts could decrease osteosarcoma cell migration and invasion abilities. Denosumab, an antibody to RANKL, is an anti-resorptive agent that has been used in patients with osteoporosis and is being explored in patients with refractory or relapsed osteosarcoma [9,58,93].

### 5.7. Tyrosine Kinase Inhibitors

Tyrosine kinases are enzymes such as tyrosine-protein kinase Met (c-Met) and vascular endothelial growth factor receptor 2 (VEGFR2), that phosphorylate tyrosine residues on proteins using ATP as a phosphate donor [94]. Tyrosine kinases are activated by ligand binding and have been implicated in various roles that drive the aggressive growth, migration, and invasion of osteosarcoma. As such, tyrosine kinase inhibitors have been approved in several cancers, including renal cell carcinoma, hepatocellular carcinoma, and soft tissue sarcomas. Tyrosine kinases are possible therapeutic options, though drugs that target this class of enzymes tend to be promiscuous.

One such tyrosine kinase inhibitor, cabozantinib, inhibits c-Met and VEGFR2, in addition to other tyrosine kinases, and has been explored as a therapeutic option in osteosarcoma in multiple ongoing trials with positive results [95]. A recently completed phase II trial in Ewing's sarcoma and osteosarcoma patients (CABONE) found that five of 42 osteosarcoma patients (12%) demonstrated a partial response to cabozantinib treatment [96].

Sorafenib, another tyrosine kinase inhibitor, targets extracellular signal-related kinase (ERK), VEGFR, and platelet-derived growth factor receptor (PDGFR)- $\alpha$  and - $\beta$ . Use of single-agent sorafenib in osteosarcoma has demonstrated anti-tumor activity, but osteosarcoma has been shown to progress through sorafenib treatment for several reasons [93,97,98]. Tyrosine kinase inhibitors such as sorafenib have therefore been explored in greater detail in combination with other therapies, such as everolimus (SERIO; NCT identifier: NCT01804374) [97,99].

Regorafenib targets VEGFR2 and PDGFR-β, and tunica interna endothelial cell kinase 2 (TIE2). Regorafenib has been demonstrated to have less severe side effects than sorafenib [100]. A phase II trial in metastatic osteosarcoma (SARC024) found that regorafenib treatment had activity in progressive metastatic osteosarcoma, where progression-free survival (PFS) was doubled from 1.7 months in the placebo group to 3.6 months in the treatment group [101]. Additional phase II trials are currently recruiting to explore the efficacy

Programmed cell death 1 (PD-1) is a cell surface protein that is expressed on activated immune cells, including CD8+ T lymphocytes, B cells, and natural killer cells, that can also be expressed in tumor cells [102]. The ligand of PD-1, PD-L1, has also been demonstrated to be overexpressed in cancer cells and associated with poorer prognosis [104]. Camrelizumab and pembrolizumab are humanized antibodies against PD-1 and used in several cancers as immunotherapies. In a trial of pembrolizumab in patients with advanced sarcoma (SARC028; NCT identifier: NCT02301039), one of the 22 patients in the osteosarcoma arm that were given pembrolizumab demonstrated an objective response [105]. A trial of camrelizumab in combination with apatinib in osteosarcoma also seemed to slightly prolong progression-free survival, though the response rate to these therapies is as of yet not optimized [106].

#### 5.9. mTOR Inhibitors

There are many alterations in the phosphoinositide 3-kinase (PI3K)/mammalian target of rapamycin (mTOR) pathway that are present in osteosarcoma, thereby suggesting mTOR as a valid target for therapy [107]. The mTORC1 signaling pathways are also activated by several oncogenes, indicating baseline activation in osteosarcoma [108]. The small molecule drug rapamycin binds to FKBP12, a receptor for immunosuppressant molecules that was found to also be a component of the protein complex that makes up mTOR complex 1 (mTORC1) [109]. Rapamycin has been utilized in osteosarcoma, but has only shown cytostatic, rather than cytotoxic, effects [108]. This is in part because rapamycin preferentially targets mTORC1, allowing mTOR complex 2 (mTORC2) to activate protein kinase B (PKB)/AKT signaling and enhance mTOR activity [110]. Various analogs of rapamycin, including everolimus and temsirolimus, have been explored in the clinic with some anti-tumoral activity (Table 1) [111]. Importantly, the adaptability of mTOR signaling in the cell allows the cell to counteract and develop resistance to mTOR inhibitors [112].

## 5.10. Combination Metabolic Therapies

As is the case with many cancers, common therapeutics tend to target known oncogenes, which are upregulated in many cancers. Several of the aforementioned therapeutics, conversely, are known metabolic inhibitors. Metabolism plays a key role in cancer therapeutic development as metabolic processes drive biomass production, redox homeostasis, and energy production [13]. Elevated glycolysis is a known attribute of many tumors, including osteosarcoma, as tumors require increased glucose uptake to facilitate rapid proliferation. This attribute is exploited in a diagnostic sense using 18F-FDG PET/CT [113,114].

The overexpression of several additional metabolic genes has also been correlated with poor survival in osteosarcoma, including X-Box Binding Protein 1 (XBP1), monocarboxylate transporter 4 (MCT4), and 3-phosphoglycerate dehydrogenase (PHGDH) [37,115,116]. Targeting these tumor biomarkers by inhibiting the overexpressed metabolic genes typically results in decreased proliferation and cytostasis, but the innate adaptability of metabolic pathways leads to limited clinical applicability of metabolic inhibitors as single agents [117,118]. As such,

## 5.11. HER2-Targeted Therapies

Osteosarcoma cell lines have been demonstrated to have high levels of HER2 cell surface expression, and HER2 expression has been correlated with poorer outcomes and decreased response to neoadjuvant chemotherapy in osteosarcoma patients [121,122]. A recent phase I trial directing chimeric antigen receptor T (CAR T) cells, or T cells that have been specifically engineered to target a particular protein and produce an immune response, at surface-expressing HER2 showed some efficacy in osteosarcoma.

Cell surface HER2 can also be targeted using monoclonal antibodies. Presently, a phase II trial is exploring the effects of trastuzumab deruxtecan on the treatment of HER2-positive osteosarcoma. Trastuzumab is a monoclonal antibody for HER2 that has been previously tested in combination with chemotherapy in osteosarcoma with no significant effects [123]. Trastuzumab was therefore linked to the chemotherapy drug deruxtecan, and is formulated in such a way that the chemotherapy can be directly delivered to the HER2-positive cancer cells (NCT identifier: NCT04616560).

## 5.12. Engineered Mesenchymal Stem Cells

In addition to contributing to osteosarcomagenesis and osteosarcoma progression, mesenchymal stem cells have been identified as a possible delivery system for therapeutic agents, making them uniquely exploitable for osteosarcoma treatment. Transduction of mesenchymal stem cells to express interferons (cytokine proteins that induce antiangiogenic and anti-tumor immune activity) and interleukins allows an immune response that can be directed to the tumor site [43,103].

Mesenchymal stem cells can also be engineered to overexpress ligands and antibodies against the tumor that have short half-lives in the body and cannot reach the tumor site independently. For example, mesenchymal stem cells can be engineered to express tumor necrosis factor related apoptosis-inducing ligand (TRAIL), which has limited use systemically because of its short half-life, and have more significant effects on apoptosis than administering TRAIL alone [124]. Studies have also been conducted to explore the efficacy of mesenchymal stem cell-delivered TRAIL in combination with chemotherapy to further enhance these apoptotic effects [43].

Finally, mesenchymal stem cells can be used as a biological method to improve bone health after surgical treatment for osteosarcoma management. As mesenchymal stem cells can differentiate into the various cell types of the bone microenvironment, application of these cells to damaged bone areas has been demonstrated in several cases to be effective in filling bone defects [43,124]. Further exploration into the genetic programs that can guide mesenchymal stem cells towards one cell type over another will allow for even more targeted defect filling as an option for post-surgical therapy in osteosarcoma.

#### 6. Conclusions

Though rare, osteosarcomas are the most common primary bone tumor in children and young adults. The transformation of normal functioning bone cells into osteosarcoma

development of a class of novel therapies. Elucidating the role of tumor metabolism in the progression of osteosarcoma is therefore necessary.

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#### References

- 1. Broadhead, M.L.; Sivaji, S.; Balogh, Z.; Choong, P.F.M. Osteosarcoma: From Molecular Biology to Mesenchymal Stem Cells. Osteosarcoma Biol. Behav. Mech. 2017. [CrossRef]
- 2. Abarrategi, A.; Tornin, J.; Martinez-Cruzado, L.; Hamilton, A.; Martinez-Campos, E.; Rodrigo, J.P.; González, M.V.; Baldini, N.; Garcia-Castro, J.; Rodriguez, R.; et al. Osteosarcoma: Cells-of-Origin, Cancer Stem Cells, and Targeted Therapies. *Stem Cells Int.* **2016**, 2016, 1–13. [CrossRef] [PubMed]
- 3. Alfranca, A.; Martinez-Cruzado, L.; Tornin, J.; Abarrategi, A.; Amaral, T.; De Alava, E.; Menendez, P.; Garcia-Castro, J.; Rodriguez, R. Bone microenvironment signals in osteosarcoma development. *Cell. Mol. Life Sci.* **2015**, *72*, 3097–3113. [CrossRef]
- 4. Katagiri, T.; Takahashi, N. Regulatory mechanisms of osteoblast and osteoclast differentiation. *Oral Dis.* **2002**, *8*, 147–159. [CrossRef] [PubMed]
- 5. Lu, Z.F.; Kleine-Nulend, J.; Li, B. Bone Microenvironment, Stem Cells, and Bone Tissue Regeneration. *Stem Cells Int.* **2017**, 2017, 2–4. [CrossRef] [PubMed]
- 6. Tang, N.; Song, W.X.; Luo, J.; Haydon, R.C.; He, T.C. Osteosarcoma development and stem cell differentiation. *Clin. Orthop. Relat. Res.* **2008**, 466, 2114–2130. [CrossRef]
- 7. Tam, W.L.; Luyten, F.P.; Roberts, S.J. From skeletal development to the creation of pluripotent stem cell-derived bone-forming progenitors. *Philos. Trans. R. Soc. B Biol. Sci.* **2018**, *373*, 1–11. [CrossRef] [PubMed]
- 8. Xu, J.; Li, Z.; Hou, Y.; Fang, W. Potential mechanisms underlying the Runx2 induced osteogenesis of bone marrow mesenchymal stem cells. *Am. J. Transl. Res.* **2015**, *7*, 2527–2535.
- 9. Hu, L.; Yin, C.; Zhao, F.; Ali, A.; Ma, J.; Qian, A. Mesenchymal stem cells: Cell fate decision to osteoblast or adipocyte and application in osteoporosis treatment. *Int. J. Mol. Sci.* **2018**, *19*, 360. [CrossRef]
- 10. Kansara, M.; Thomas, D.M. Molecular pathogenesis of osteosarcoma. DNA Cell Biol. 2007, 26, 1–18. [CrossRef]
- 11. Kim, E.-K.; Lim, S.; Park, J.-M.; Seo, J.K.; Kim, J.H.; Kim, K.T.; Ryu, S.H.; Suh, P.-G. Human mesenchymal stem cell differentiation to the osteogenic or adipogenic lineage is regulated by AMP-activated protein kinase. *J. Cell. Physiol.* **2012**, 227, 1680–1687. [CrossRef]
- 12 Mohamed A M.E.S. An overview of hone cells and their regulating factors of differentiation. Malays. I. Med. Sci. 2008, 15, 4–12.

23. Rubio, R.; Gutierrez-Aranda, I.; Sáez-Castillo, A.I.; Labarga, A.; Rosu-Myles, M.; Gonzalez-Garcia, S.; Toribio, M.L.; Menendez, P.; Rodriguez, R. The differentiation stage of p53-Rb-deficient bone marrow mesenchymal stem cells imposes the phenotype of in vivo sarcoma development. *Oncogene* **2013**, *32*, 4970–4980. [CrossRef]

- 24. Adamopoulos, C.; Gargalionis, A.N.; Basdra, E.K.; Papavassiliou, A.G. Deciphering signaling networks in osteosarcoma pathobiology. *Exp. Biol. Med.* **2016**, 241, 1296–1305. [CrossRef] [PubMed]
- 25. Haydon, R.C.; Deyrup, A.; Ishikawa, A.; Heck, R.; Jiang, W.; Zhou, L.; Feng, T.; King, D.; Cheng, H.; Breyer, B.; et al. Cytoplasmic and/or Nuclear Accumulation of the β-Catenin Protein is a Frequent Event in Human Osteosarcoma. *Int. J. Cancer* **2002**, *102*, 338–342. [CrossRef]
- 26. Alliston, T.; Choy, L.; Ducy, P.; Karsenty, G.; Derynck, R. TGF-β-induced repression of CBFA1 by Smad3 decreases cbfa1 and osteocalcin expression and inhibits osteoblast differentiation. *EMBO J.* **2001**, *20*, 2254–2272. [CrossRef] [PubMed]
- 27. Chen, G.; Deng, C.; Li, Y.P. TGF-β and BMP signaling in osteoblast differentiation and bone formation. *Int. J. Biol. Sci.* **2012**, *8*, 272–288. [CrossRef]
- 28. Lamora, A.; Talbot, J.; Mullard, M.; Brounais-Le Royer, B.; Redini, F.; Verrecchia, F. TGF-β Signaling in Bone Remodeling and Osteosarcoma Progression. *J. Clin. Med.* **2016**, *5*, 96. [CrossRef] [PubMed]
- 29. Nathan, S.S.; Pereira, B.P.; Zhou, Y.F.; Gupta, A.; Dombrowski, C.; Soong, R.; Pho, R.W.H.; Stein, G.S.; Salto-Tellez, M.; Cool, S.M.; et al. Elevated expression of Runx2 as a key parameter in the etiology of osteosarcoma. *Mol. Biol. Rep.* **2009**, *36*, 153–158. [CrossRef] [PubMed]
- 30. Ren, H.Y.; Sun, L.L.; Li, H.Y.; Ye, Z.M. Prognostic significance of serum alkaline phosphatase level in osteosarcoma: A meta-analysis of published data. *Biomed. Res. Int.* **2015**, *2015*. [CrossRef]
- 31. Wang, J.J.; Ye, F.; Cheng, L.J.; Shi, Y.J.; Bao, J.; Sun, H.Q.; Wang, W.; Zhang, P.; Bu, H. Osteogenic differentiation of mesenchymal stem cells promoted by overexpression of connective tissue growth factor. *J. Zhejiang Univ. Sci. B* **2009**, *10*, 355–367. [CrossRef] [PubMed]
- 32. Sadikovic, B.; Thorner, P.; Chilton-MacNeill, S.; Martin, J.W.; Cervigne, N.K.; Squire, J.; Zielenska, M. Expression analysis of genes associated with human osteosarcoma tumors shows correlation of RUNX2 overexpression with poor response to chemotherapy. *BMC Cancer* **2010**, *10*. [CrossRef] [PubMed]
- 33. Shi, Y.; He, G.; Lee, W.C.; McKenzie, J.A.; Silva, M.J.; Long, F. Gli1 identifies osteogenic progenitors for bone formation and fracture repair. *Nat. Commun.* **2017**, *8*, 1–12. [CrossRef]
- 34. Angulo, P.; Kaushik, G.; Subramaniam, D.; Dandawate, P.; Neville, K.; Chastain, K.; Anant, S. Natural compounds targeting major cell signaling pathways: A novel paradigm for osteosarcoma therapy. *J. Hematol. Oncol.* 2017, 10, 1–13. [CrossRef] [PubMed]
- 35. Beristain, A.G.; Narala, S.R.; Di Grappa, M.A.; Khokha, R. Homotypic RANK signaling differentially regulates proliferation, motility and cell survival in osteosarcoma and mammary epithelial cells. *J. Cell Sci.* **2012**, 125, 943–955. [CrossRef] [PubMed]
- 36. Song, Z.; Feng, C.; Lu, Y.; Lin, Y.; Dong, C. PHGDH is an independent prognosis marker and contributes cell proliferation, migration and invasion in human pancreatic cancer. *Gene* **2018**, *642*, 43–50. [CrossRef]
- 37. Rathore, R.; Caldwell, K.E.; Schutt, C.; Brashears, C.B.; Prudner, B.C.; Ehrhardt, W.R.; Leung, C.H.; Lin, H.; Daw, N.C.; Beird, H.C.; et al. Metabolic compensation activates pro-survival mTORC1 signaling upon 3-phosphoglycerate dehydrogenase inhibition in osteosarcoma. *Cell Rep.* **2021**, *34*, 108678. [CrossRef] [PubMed]
- 38. Rubio, R.; Abarrategi, A.; Garcia-Castro, J.; Martinez-Cruzado, L.; Suarez, C.; Tornin, J.; Santos, L.; Astudillo, A.; Colmenero, I.; Mulero, F.; et al. Bone environment is essential for osteosarcoma development from transformed mesenchymal stem cells. *Stem Cells* **2014**, *32*, 1136–1148. [CrossRef]
- 39. Wang, L.; Park, P.; La Marca, F.; Than, K.; Rahman, S.; Lin, C.Y. Bone formation induced by BMP-2 in human osteosarcoma cells. *Int. J. Oncol.* **2013**, *43*, 1095–1102. [CrossRef] [PubMed]
- 40. De Azevedo, J.W.V.; de Medeiros Fernandes, T.A.A.; Fernandes, J.V.; de Azevedo, J.C.V.; Lanza, D.C.F.; Bezerra, C.M.; Andrade, V.S.; de Araújo, J.M.G.; Fernandes, J.V. Biology and pathogenesis of human osteosarcoma (Review). *Oncol. Lett.* **2020**, *19*, 1099–1116. [CrossRef]
- 41. Huang, I.; Ni, I.; Liu, K.; Yu, Y.; Xie, M.; Kang, R.; Vernon, P.; Cao, L.; Tang, D. HMGB1 promotes drug resistance in osteosarcoma.

48. Olivier, M.; Hollstein, M.; Hainaut, P. TP53 Mutations in Human Cancers: Origins, Consequences, and Clinical Use. *Cold Spring Harb. Perspect. Biol.* **2010**, 2, 1–17. [CrossRef]

- 49. Iurlaro, R.; León-Annicchiarico, C.L.; Muñoz-Pinedo, C. Regulation of cancer metabolism by oncogenes and tumor suppressors. Methods Enzymol. 2014, 542, 59–80. [CrossRef]
- 50. Hansen, M.F.; Seton, M.; Merchant, A. Osteosarcoma in Paget's Disease of Bone. *J. Bone Miner. Res.* **2006**, *21*, P58–P63. [CrossRef] [PubMed]
- 51. Nellissery, M.J.; Padalecki, S.S.; Brkanac, Z.; Singer, F.R.; Roodman, G.D.; Unni, K.K.; Leach, R.J.; Hansen, M.F. Evidence for a novel osteosarcoma tumor-suppressor gene in the chromosome 18 region genetically linked with Paget disease of bone. *Am. J. Hum. Genet.* **1998**, *63*, 817–824. [CrossRef]
- 52. Ricafort, R.; Gorlick, R. Molecularly Targeted Therapy for Osteosarcoma: Where Do We Go from Here? *Mol. Target. Ther. Child. Cancer* **2010**, 459–498. [CrossRef]
- 53. Fuchs, B.; Pritchard, D.J. Etiology of osteosarcoma. Clin. Orthop. Relat. Res. 2002, 40–52. [CrossRef]
- 54. Mendoza, S.M.; Konishi, T.; Miller, C.W. Integration of SV40 in human osteosarcoma DNA. *Oncogene* **1998**, *17*, 2457–2462. [CrossRef]
- 55. Geller, D.S.; Gorlick, R. Osteosarcoma: A Review of Diagnosis, Management, and Treatment Strategies. *Clin. Adv. Hematol. Oncol.* **2010**, *8*, 705–718. [CrossRef] [PubMed]
- 56. Heare, T.; Hensley, M.A.; Dell'Orfano, S. Bone tumors: Osteosarcoma and Ewing's sarcoma. *Curr. Opin. Pediatr.* **2009**, *21*, 365–372. [CrossRef]
- 57. Choi, J.H.; Ro, J.Y. The 2020 WHO Classification of Tumors of Bone: An Updated Review. Adv. Anat. Pathol. 2021, 1–20. [CrossRef]
- 58. Isakoff, M.S.; Bielack, S.S.; Meltzer, P.; Gorlick, R. Osteosarcoma: Current Treatment and a Collaborative Pathway to Success. *J. Clin. Oncol.* **2015**, *33*, 3029–3035. [CrossRef] [PubMed]
- 59. Whelan, J.S.; Davis, L.E. Osteosarcoma, chondrosarcoma, and chordoma. J. Clin. Oncol. 2018, 36, 188–193. [CrossRef] [PubMed]
- 60. Bielack, S.S.; Werner, M.; Tunn, P.U.; Helmke, K.; Jürgens, H.; Calaminus, G.; Gerss, J.; Butterfass-Bahloul, T.; Reichardt, P.; Smeland, S.; et al. Methotrexate, doxorubicin, and cisplatin (MAP) plus maintenance pegylated interferon alfa-2b versus MAP alone in patients with resectable high-grade osteosarcoma and good histologic response to preoperative MAP: First results of the EURAMOS-1 good respons. *J. Clin. Oncol.* 2015, 33, 2279–2287. [CrossRef] [PubMed]
- 61. Wu, C.; Wang, Q.; Li, Y. Prediction and evaluation of neoadjuvant chemotherapy using the dual mechanisms of 99m-Tc-MIBI scintigraphy in patients with osteosarcoma. *J. Bone Oncol.* **2019**, 17. [CrossRef]
- 62. Dasari, S.; Bernard Tchounwou, P. Cisplatin in cancer therapy: Molecular mechanisms of action. *Eur. J. Pharmacol.* **2014**, 740, 364–378. [CrossRef] [PubMed]
- 63. Meredith, A.M.; Dass, C.R. Increasing role of the cancer chemotherapeutic doxorubicin in cellular metabolism. *J. Pharm. Pharmacol.* **2016**, *68*, 729–741. [CrossRef]
- 64. Dechant, K.L.; Brogden, R.N.; Pilkington, T.; Faulds, D. Ifosfamide/Mesna: A Review of its Antineoplastic Activity, Pharmacokinetic Properties and Therapeutic Efficacy in Cancer. *Drugs* 1991, 42, 428–467. [CrossRef] [PubMed]
- 65. Jaffe, N.; Puri, A.; Gelderblom, H. Osteosarcoma: Evolution of Treatment Paradigms. *Sarcoma* 2013, 203531. [CrossRef]
- 66. Balamuth, N.J.; Womer, R.B. Ewing's sarcoma. Lancet Oncol. 2010, 11, 184-192. [CrossRef]
- 67. Liu, Y.; Xu, Y.; Lin, N.; Jiang, S.; Wang, Y.; Ye, Z. High-Dose Methotrexate (HD-MTX) Used as an Adjunct with Other Chemotherapeutics for the Treatment of Osteosarcoma. *Cell Biochem. Biophys.* **2015**, *71*, 1097–1104. [CrossRef] [PubMed]
- 68. Patiño-García, A.; Zalacaín, M.; Marrodán, L.; San-Julián, M.; Sierrasesúmaga, L. Methotrexate in Pediatric Osteosarcoma: Response and Toxicity in Relation to Genetic Polymorphisms and Dihydrofolate Reductase and Reduced Folate Carrier 1 Expression. J. Pediatr. 2009, 154, 688–693. [CrossRef] [PubMed]
- 69. Rajagopalan, P.T.R.; Zhang, Z.; McCourt, L.; Dwyer, M.; Benkovic, S.J.; Hammes, G.G. Interaction of dihydrofolate reductase with methotrexate: Ensemble and single-molecule kinetics. *Proc. Natl. Acad. Sci. USA* **2002**, *99*, 13481–13486. [CrossRef]
- 70. Serra, M.; Reverter-Branchat, G.; Maurici, D.; Benini, S.; Shen, J.N.; Chano, T.; Hattinger, C.M.; Manara, M.C.; Pasello, M.; Scotlandi, K.; et al. Analysis of dihydrofolate reductase and reduced folate carrier gene status in relation to methotrexate

77. Grem, J.L.; King, S.A.; Wittes, R.E.; Leyland-Jones, B. The role of methotrexate in osteosarcoma. *J. Natl. Cancer Inst.* **1988**, *80*, 626–655. [CrossRef] [PubMed]

- 78. Freed-Pastor, W.A.; Prives, C. Mutant p53: One name, many proteins. Genes Dev. 2012, 26, 1268–1286. [CrossRef] [PubMed]
- 79. Wang, X.; Kua, H.Y.; Hu, Y.; Guo, K.; Zeng, Q.; Wu, Q.; Ng, H.H.; Karsenty, G.; De Crombrugghe, B.; Yeh, J.; et al. P53 Functions As a Negative Regulator of Osteoblastogenesis, Osteoblast-Dependent Osteoclastogenesis, and Bone Remodeling. *J. Cell Biol.* 2006, 172, 115–125. [CrossRef] [PubMed]
- 80. Ladanyi, M.; Cha, C.; Lewis, R.; Jhanwar, S.C.; Huvos, A.G.; Healey, J.H. MDM2 Gene Amplification in Metastatic Osteosarcoma. *Cancer Res.* **1993**, *53*, 16–18.
- 81. Lonardo, F.; Ueda, T.; Huvos, A.G.; Healey, J.; Ladanyi, M. p53 and MDM2 Alterations in Osteosarcomas. *Cancer* 1997, 79, 1541–1547. [CrossRef]
- 82. Manfredi, J.J. The Mdm2–p53 relationship evolves: Mdm2 swings both ways as an oncogene and a tumor suppressor. *Genes Dev.* **2010**, *24*, 1580–1589. [CrossRef] [PubMed]
- 83. Yoshida, A.; Ushiku, T.; Motoi, T.; Beppu, Y.; Fukayama, M.; Tsuda, H.; Shibata, T. MDM2 and CDK4 Immunohistochemical Coexpression in High-grade Osteosarcoma: Correlation With a Dedifferentiated Subtype. *Am. J. Surg. Pathol.* **2012**, *36*, 423–431. [CrossRef] [PubMed]
- 84. Ou, Y.; Wang, S.J.; Jiang, L.; Zheng, B.; Gu, W. p53 Protein-mediated regulation of phosphoglycerate dehydrogenase (PHGDH) is crucial for the apoptotic response upon serine starvation. *J. Biol. Chem.* **2015**, 290, 457–466. [CrossRef]
- 85. Bykov, V.J.N.; Eriksson, S.E.; Bianchi, J.; Wiman, K.G. Targeting mutant p53 for efficient cancer therapy. *Nat. Rev. Cancer* **2018**, *18*, 89–102. [CrossRef]
- 86. Maki, R.G. Gemcitabine and Docetaxel in Metastatic Sarcoma: Past, Present, and Future. Oncologist 2007, 12, 999–1006. [CrossRef]
- 87. Zakeri-Milani, P.; Farkhani, S.M.; Shirani, A.; Mohammadi, S.; Mojarrad, J.S.; Akbari, J.; Valizadeh, H. Cellular Uptake and Anti-Tumor Activity of Gemcitabine Conjugated with New Amphiphilic Cell Penetrating Peptides. *EXCLI J.* **2017**, *16*, 650–662. [PubMed]
- 88. Prudner, B.C.; Rathore, R.; Robinson, A.M.; Godec, A.; Chang, S.F.; Hawkins, W.G.; Hirbe, A.C.; Van Tine, B.A. Arginine starvation and docetaxel induce c-Myc-driven HENT1 surface expression to overcome gemcitabine resistance in ASS1-negative tumors. *Clin. Cancer Res.* **2019**, *25*, 5122–5134. [CrossRef] [PubMed]
- 89. Bush, J.A.; Li, G. Cancer chemoresistance: The relationship between p53 and multidrug transporters. *Int. J. Cancer* **2002**, *98*, 323–330. [CrossRef] [PubMed]
- 90. Zawadzka, I.; Jeleń, A.; Pietrzak, J.; Żebrowska-Nawrocka, M.; Michalska, K.; Szmajda-Krygier, D.; Mirowski, M.; Łochowski, M.; Kozak, J.; Balcerczak, E. The impact of ABCB1 gene polymorphism and its expression on non-small-cell lung cancer development, progression and therapy—Preliminary report. *Sci. Rep.* **2020**, *10*, 1–10. [CrossRef] [PubMed]
- 91. Liu, T.; Li, Z.; Zhang, Q.; Bernstein, K.D.A.; Lozano-Calderon, S.; Choy, E.; Hornicek, F.J.; Duan, Z. Targeting ABCB1 (MDR1) in multi-drug resistant osteosarcoma cells using the CRISPR-Cas9 system to reverse drug resistance. *Oncotarget* **2016**, *7*, 83502–83513. [CrossRef]
- 92. Xiaohui, S.; Aiguo, L.; Xiaolin, G.; Ying, L.; Hongxing, Z.; Yilei, Z. Effect of ABCB1 polymorphism on the clinical outcome of osteosarcoma patients after receiving chemotherapy. *Pakistan J. Med. Sci.* **2014**, *30*, 886–890. [CrossRef]
- 93. Cathomas, R.; Rothermundt, C.; Bode, B.; Fuchs, B.; Von Moos, R.; Schwitter, M. RANK ligand blockade with denosumab in combination with sorafenib in chemorefractory osteosarcoma: A possible step forward? *Oncology* **2015**, *88*, 257–260. [CrossRef] [PubMed]
- 94. Paul, M.K.; Mukhopadhyay, A.K. Tyrosine kinase—Role and significance in Cancer. *Int. J. Med. Sci.* **2004**, *1*, 101–115. [CrossRef] [PubMed]
- 95. Schöffski, P.; Blay, J.Y.; Ray-Coquard, I. Cabozantinib as an emerging treatment for sarcoma. *Curr. Opin. Oncol.* **2020**, 32, 321–331. [CrossRef]
- 96. Italiano, A.; Mir, O.; Mathoulin-Pelissier, S.; Penel, N.; Piperno-Neumann, S.; Bompas, E.; Chevreau, C.; Duffaud, F.; Entz-Werlé, N.; Saada, E.; et al. Cabozantinib in patients with advanced Ewing sarcoma or osteosarcoma (CABONE): A multicentre,

103. Shaikh, A.B.; Li, F.; Li, M.; He, B.; He, X.; Chen, G.; Guo, B.; Li, D.; Jiang, F.; Dang, L.; et al. Present advances and future perspectives of molecular targeted therapy for osteosarcoma. *Int. J. Mol. Sci.* **2016**, *17*, 506. [CrossRef] [PubMed]

- 104. Brahmer, J.R.; Tykodi, S.S.; Chow, L.Q.M.; Hwu, W.-J.; Topalian, S.L.; Hwu, P.; Drake, C.G.; Camacho, L.H.; Kauh, J.; Odunsi, K.; et al. Safety and Activity of Anti-PD-L1 Antibody in Patients with Advanced Cancer. *N. Engl. J. Med.* 2012, 366, 2455–2465. [CrossRef] [PubMed]
- 105. Tawbi, H.A.; Burgess, M.; Bolejack, V.; Van Tine, B.A.; Schuetze, S.M.; Hu, J.; D'Angelo, S.; Attia, S.; Riedel, R.F.; Priebat, D.A.; et al. Pembrolizumab in advanced soft-tissue sarcoma and bone sarcoma (SARC028): A multicentre, two-cohort, single-arm, open-label, phase 2 trial. *Lancet Oncol.* **2017**, *18*, 1493–1501. [CrossRef]
- 106. Xie, L.; Xu, J.; Sun, X.; Guo, W.; Gu, J.; Liu, K.; Zheng, B.; Ren, T.; Huang, Y.; Tang, X.; et al. Apatinib plus camrelizumab (anti-PD1 therapy, SHR-1210) for advanced osteosarcoma (APFAO) progressing after chemotherapy: A single-arm, open-label, phase 2 trial. *J. Immunother. Cancer* **2020**, *8*, 1–9. [CrossRef]
- 107. Perry, J.A.; Kiezun, A.; Tonzi, P.; Van Allen, E.M.; Carter, S.L.; Baca, S.C.; Cowley, G.S.; Bhatt, A.S.; Rheinbay, E.; Pedamallu, C.S.; et al. Complementary genomic approaches highlight the PI3K/mTOR pathway as a common vulnerability in osteosarcoma. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, E5564–E5573. [CrossRef]
- 108. Xie, J.; Wang, X.; Proud, C.G. mTOR inhibitors in cancer therapy [version 1; referees: 3 approved]. F1000Research 2016, 5. [CrossRef]
- 109. Leone, M.; Crowell, K.J.; Chen, J.; Jung, D.; Chiang, G.G.; Sareth, S.; Abraham, R.T.; Pellecchia, M. The FRB domain of mTOR: NMR solution structure and inhibitor design. *Biochemistry* **2006**, *45*, 10294–10302. [CrossRef] [PubMed]
- 110. Rodrik-Outmezguine, V.S.; Chandarlapaty, S.; Pagano, N.C.; Poulikakos, P.I.; Scaltriti, M.; Moskatel, E.; Baselga, J.; Guichard, S.; Rosen, N. mTOR kinase inhibition causes feedback-dependent biphasic regulation of AKT signaling. *Cancer Discov.* **2011**, *1*, 248–259. [CrossRef] [PubMed]
- 111. Mita, M.M.; Tolcher, A.W. The role of mTOR inhibitors for treatment of sarcomas. *Curr. Oncol. Rep.* **2007**, *9*, 316–322. [CrossRef] [PubMed]
- 112. Ding, L.; Congwei, L.; Bei, Q.; Tao, Y.; Ruiguo, W.; Heze, Y.; Bo, D.; Zhihong, L. mTOR: An attractive therapeutic target for osteosarcoma? *Oncotarget* **2016**, *7*, 50805–50813. [CrossRef]
- 113. Charest, M.; Hickeson, M.; Lisbona, R.; Novales-Diaz, J.A.; Derbekyan, V.; Turcotte, R.E. FDG PET/CT imaging in primary osseous and soft tissue sarcomas: A retrospective review of 212 cases. *Eur. J. Nucl. Med. Mol. Imaging* **2009**, *36*, 1944–1951. [CrossRef]
- 114. Tsiambas, E.; Fotiades, P.P.; Sioka, C.; Kotrotsios, D.; Gkika, E.; Fotopoulos, A.; Mastronikolis, S.N.; Armata, I.E.; Giotakis, E.; Ragos, V. Novel molecular and metabolic aspects in osteosarcoma. *J. BUON* **2017**, *22*, 1595–1598.
- 115. Yang, J.; Cheng, D.; Zhou, S.; Zhu, B.; Hu, T.; Yang, Q. Overexpression of X-Box binding protein 1 (XBP1) correlates to poor prognosis and up-regulation of PI3K/mTOR in human osteosarcoma. *Int. J. Mol. Sci.* **2015**, *16*, 28635–28646. [CrossRef] [PubMed]
- 116. Liu, Y.; Sun, X.; Huo, C.; Sun, C.; Zhu, J. Monocarboxylate Transporter 4 (MCT4) Overexpression Is Correlated with Poor Prognosis of Osteosarcoma. *Med. Sci. Monit.* **2019**, 25, 4278–4284. [CrossRef] [PubMed]
- 117. Amorim, R.; Pinheiro, C.; Miranda-Gonçalves, V.; Pereira, H.; Moyer, M.P.; Preto, A.; Baltazar, F. Monocarboxylate transport inhibition potentiates the cytotoxic effect of 5-fluorouracil in colorectal cancer cells. *Cancer Lett.* **2015**, *365*, *68–78*. [CrossRef] [PubMed]
- 118. Pacold, M.E.; Brimacombe, K.R.; Chan, S.H.; Rohde, J.M.; Lewis, C.A.; Swier, L.J.Y.M.; Possemato, R.; Chen, W.W.; Sullivan, L.B.; Fiske, B.P.; et al. A PHGDH inhibitor reveals coordination of serine synthesis and one-carbon unit fate. *Nat. Chem. Biol.* **2016**, 12, 452–458. [CrossRef] [PubMed]
- 119. Issaq, S.H.; Mendoza, A.; Kidner, R.; Rosales, T.; Duveau, D.Y.; Heske, C.M.; Rohde, J.M.; Boxer, M.B.; Thomas, C.J.; DeBerardinis, R.J.; et al. EWS-FLI1-regulated serine synthesis and exogenous serine are necessary for Ewing sarcoma cellular proliferation and tumor growth. *Mol. Cancer Ther.* 2020. [CrossRef] [PubMed]
- 120. Newman, A.C.; Maddocks, O.D.K. Serine and Functional Metabolites in Cancer. *Trends Cell Biol.* **2017**, 27, 645–657. [CrossRef] [PubMed]