

## Supplemental Online Content

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This supplemental material has been provided by the authors to give readers additional information about their work.

## **eAppendix.** Supplementary Methods

We utilized serum samples that were prospectively collected from patients who presented to the Barnes Jewish Hospital in Saint Louis, MO with symptoms suggestive of COVID-19 illness and were confirmed to have SARS-CoV-2 infection with nasopharyngeal swabs. Testosterone, estradiol, and IGF-1 were measured by liquid chromatography- mass spectrometry at the time of presentation to the hospital (baseline or day 0) and at days 3, 7, 14 and 28 after admission (if the patient remained hospitalized).

### **Hormone assays:**

Owing to limited sample volumes available, assays for total testosterone, estradiol, and IGF-1 were performed following dilution into charcoal stripped serum as follows:

Testosterone (females): 1 part sample to 2 parts stripped serum

Testosterone (males): 1 part sample to 3 parts stripped serum

IGF-1 (both sexes): 1 part sample to 1 part stripped serum

Estradiol (both sexes): 1 part sample to 1 part stripped serum

Validation experiments have documented that such dilutions yield reproducible, accurate measurements compared to undiluted specimens (Quest Diagnostics Inc., unpublished information).

### **Testosterone**

Total testosterone was measured by liquid chromatography tandem mass spectrometry (LC-MS/MS)<sup>1</sup>. The method is linear from 205 to 2000 ng/dL with an analytical sensitivity (Limit of quantification) set at a coefficient of variation (CV) of  $\leq 20\%$ , is 1.0 ng/dl. Intra-assay and total CVs across four quality control concentrations were less than 10%. The reference intervals for

total testosterone for this assay were 250-1100 ng/dL in adult males <sup>2</sup> and 2-45 ng/dl in adult females. The performing laboratory is certified by Clinical Laboratory Improvement Amendments and is a participant in the Hormone Standardization program of the Centers for Disease Control and Prevention.

### **Estradiol**

LC-MS/MS methodology was also used to measure total estradiol. The method was similar to that previously described <sup>3</sup>, with some improvements. After adding 300  $\mu$ L of ethanol spiked with internal standards ( $17\beta$ -Estradiol-2,3,4-<sup>13</sup>C<sub>3</sub>) to 200  $\mu$ L of patient serum, the mixture was incubated in an ice slurry for 30 minutes before centrifugation. 50  $\mu$ L of the supernatant was injected onto a multiplex High Performance Liquid Chromatography (HPLC) system equipped with an online turbo flow extraction system (Cohesive Technologies Inc.; Franklin, MA, part of Thermo Fisher Scientific) for further clean up and separation from the matrix. The online extraction and HPLC columns were Phenomenex Strata C18-E (70  $\mu$ m, 50x1.0 mm; P/N 00B-S001-A0) and Phenomenex Biphenyl (2.6  $\mu$ m, 100Å, 50x3.0 mm; P/N AF0-5727) respectively. Estradiol and its internal standard were detected by a triple quadrupole mass spectrometer (Sciex 6500+ Framingham, MA) using H-ESI source and operating in Multiple Reaction Monitoring (MRM) mode. The MRM transitions were (m/z) 271.17 to 143.05 and 145.05 for Estradiol and 274.18 to 148.00 and 186.05 for the Internal Standard. The ratio of the analyte response relative to that of the internal standard were used to build a calibration curve and to quantify Estradiol in patient samples. The Inter assay precision (CV) of the assay is 4.06% at 20 pg/mL of, 1.42% at 200 pg/mL, and 1.88% at 800 pg/mL. The LOQ for estradiol in this assay is 2 pg/mL. The

reference range is  $\leq 29$  pg/mL for men, 39-440 pg/mL for premenopausal women and  $\leq 10$  pg/mL for postmenopausal women.

### **IGF-1 measurements**

IGF-1 was measured by highly accurate LC-MS method similar to the method described previously.<sup>4</sup> 100  $\mu$ L aliquots of patient sera were mixed with 10  $\mu$ L of internal standard (15N-labeled IGF-1, ProSpec) and 400  $\mu$ L of acidified ethanol solution (87.5% EtOH, 12.5% 1 N HCl) and mixed vigorously. After incubating at room temperature for 30 min, and centrifugation, 350  $\mu$ L of supernatant was mixed with 60  $\mu$ L of 1.5 M Tris base. The samples were cooled for 60 min at  $-20^{\circ}\text{C}$  and centrifuged again to remove additional protein precipitate. The IGF-1 and its internal standard were separated further from matrix using a multiplex high performance liquid chromatography (HPLC) system (Aria TLX-4) equipped with an on-line extraction (Thermo Scientific, San Jose, CA). The extraction column was a Phenomenex Monolithic Onyx C18 Guard Cartridge; the analytical column was a Phenomenex Kinetex C18  $50 \times 4.6$  mm, 100  $\text{\AA}$  pore size. A Thermo Scientific Q Exactive Focus Hybrid Quadrupole-Orbitrap mass spectrometer (Thermo Scientific, San Jose, CA) was used as the detection system equipped with heated electrospray ionization (HESI) source and operating in positive All Ion Fragmentation (AIF) mode. After IGF-1 quantitation, the chromatograms were then examined for the presence of variants. No variant was found in this study.

The inter assay precision (CV) of the IGF-1 assay is 8.79% at 40 ng/mL, 6.43% at 177 ng/mL, and 5.97% at 358 ng/mL. The LOQ for IGF-1 assay is 8 ng/mL. The reference ranges of IGF-1 vary by age and gender and have been described previously<sup>5</sup>.

**Cytokine Quantification:** Plasma obtained from subjects was frozen at -80°C and subsequently analyzed using a human magnetic cytokine panel providing parallel measurement of 35 cytokines (ThermoFisher). These are Interleukin (IL)-1 $\alpha$ , 1 $\beta$ , 2-10, 12, 13, 15, 17A, 17F and 22, IL-1 receptor antagonist (IL-1RA), IL-2R, Fibroblast growth factor (FGF)-basic, Tumor necrosis factor (TNF)- $\alpha$ , Granulocyte colony-stimulating factor (G-CSF) and Granulocyte-macrophage colony-stimulating factor (GM-CSF), RANTES (Regulated on Activation, Normal T Cell Expressed and Secreted), Eotaxin, Macrophage Inflammatory Proteins (MIP)-1 $\alpha$  and 1 $\beta$ , monocyte chemoattractant protein-1 (MCP-1), Epidermal growth factor (EGF), hepatocyte growth factor (HGF), Vascular endothelial growth factor (VEGF), Interferon (IFN)- $\alpha$  and  $\gamma$ , interferon- $\gamma$ -inducible protein 10 (IP-10) and monokine induced by gamma interferon (MIG) . The assay was performed according to the manufacturer's instructions with each subject sample performed in duplicate and then analyzed on a Luminex FLEXMAP 3D instrument.

### **PBMC sorting and RNA sequencing**

Cryopreserved peripheral blood mononuclear cells (PBMCs) from COVID patients were thawed and washed with HBSS with 2mM EDTA and 0.04% LPS-free BSA. These PBMCs were then blocked with 5% goat serum and stained on ice with anti-CD14-APC/Cy7, anti-CD16-PE/Cy7, anti-CD15 PerCP/Cy5.5, and anti-CD56-Alexa Fluor 700 for 30 minutes and re-suspended in the aforementioned HBSS buffer. Dead cells were excluded based on positive Helix NP<sup>TM</sup> Green staining. The cells were then sorted straight into Trizol by FACS Aria II. Total RNA was isolated as per Trizol protocol. RNA of each sample was sequenced at the Genome Technology Access Center of the McDonnell Genome Institute at Washington University in St. Louis. Total RNA integrity was determined using Agilent Bioanalyzer or 4200 TapeStation. Library preparation

was performed with 10ng of total RNA with a Bioanalyzer RIN score greater than 8.0. ds-cDNA was prepared using the SMARTer Ultra Low RNA kit for Illumina Sequencing (Takara-Clontech) per manufacturer's protocol. cDNA was fragmented using a Covaris E220 sonicator using peak incident power 18, duty factor 20%, cycles per burst 50 for 120 seconds. cDNA was blunt ended, had an A base added to the 3' ends, and then had Illumina sequencing adapters ligated to the ends. Ligated fragments were then amplified for 12-15 cycles using primers incorporating unique dual index tags. Fragments were sequenced on an Illumina NovaSeq-6000 using paired end reads extending 150 bases at the Genome Technology Access Center of the McDonnell Genome Institute at Washington University in St. Louis.

Basecalls and demultiplexing were performed with Illumina's bcl2fastq software and a custom python demultiplexing program with a maximum of one mismatch in the indexing read. RNA-seq reads were then aligned to the Ensembl release 76 top-level assembly of the human genome (GRCh38) along with the SARS-CoV-2 reference genome with STAR version 2.0.4b<sup>6</sup> with the parameters `--outStd BAM_Unsorted --outMultimapperOrder Random --outSAMtype BAM_Unsorted --outSAMunmapped None --outSAMmapqUnique 60 --outFilterScoreMinOverLread 0 --outFilterMatchNmin 20 --outFilterMatchNminOverLread 0 --outFilterMismatchNoverLmax 0.1 --clip3pAdapterSeq`  
AGATCGGAAGAGCGGTTCAGCAGGAATGCCGAG  
AGATCGGAAGAGCGTCGTGTAGGGAAAGAGTGTA. Gene counts were derived from the number of uniquely aligned unambiguous reads by Subread:featureCount version 1.4.5<sup>7</sup> to Ensembl and SARS-CoV-2 gene models. Sequencing performance was assessed for the total number of aligned reads, total number of uniquely aligned reads, and features detected. Differential expression analysis to determine genes that differed with severe disease for each cell

type and both male and females was performed using DESeq2 (v1.24)<sup>8</sup>. Genes without at least 10 read counts across all samples were removed prior to analysis. Gene Set Enrichment Analysis (v4.0.3)<sup>9</sup> was performed using the GSEA Preranked tool against the Hallmark gene set (v7.2). Genes were ranked using the sign of the fold change times the  $-\log_{10}(\text{p-value})$  and otherwise default parameters.

### **Statistical analysis**

We assessed the relation of testosterone, estradiol, and IGF-1 concentrations during the hospitalization with COVID severity, ICU and ventilator use, mortality and inflammatory cytokines using univariate and multivariate regression and mixed model analyses. These models addressed missing data values by maximum likelihood, under the data missing at random assumption. Sensitivity analyses were performed to evaluate this assumption. Based on these analyses, missing at random assumption was deemed reasonable. The comparisons were adjusted for age, BMI, race, smoking and co-morbidities at baseline (Charlson Comorbidity Index, CCI). CCI is a validated method to determine significant medical comorbidity. It considers 17 common medical conditions that predict one-year mortality. These include myocardial infarction, heart failure, peripheral arterial disease, cerebrovascular disease, dementia, chronic pulmonary disease, connective tissue disease, ulcer disease, liver disease, diabetes (including diabetes with end organ damage), hemiplegia, renal disease, leukemia, lymphoma, solid tumors, and AIDS. Medical diagnoses are weighted for severity and summed to provide a weighted index of medical comorbidity with total scores ranging from 0-33.



**eTable 1.** Baseline Characteristics, Hormone Concentrations, and Hospital Outcomes in Men and Women

	Males (N=90)	Females (N=62)	P
Age (years)	64±14	61±18	0.33
BMI (kg/m <sup>2</sup> )	27.7±6.8	33±8.8	<0.001
Charlson comorbidity index	3 [1, 4]	2 [1, 3]	0.046
Severe COVID	71%	60%	0.14
ICU	59%	47%	0.14
ventilator	27%	21%	0.42
Mortality	28%	19%	0.23
Testosterone (ng/dl)	79 [38, 181]	12 [1, 21]	<0.001
Estradiol (pg/ml)	15 [10, 22]	15 [3, 47]	0.34
IGF-1 (ng/ml)	90 [62, 126]	106 [62, 127]	0.90
CRP (mg/L)	122 [58, 214]	96 [48, 174]	0.71

**eTable 2.** Serum Estradiol Concentration by Intensive Care Unit Admission, Ventilator Use, and Mortality in Men

		Day 0	Day 3	Day 7	Day 14	Day 28
ICU	Yes (N=53)	15 [11, 25]	13 [7, 26]	13 [7, 20]	18 [10, 23]	13 [10, 26]
	No (N=37)	13 [10, 20]	13 [9, 26]	12 [10, 13]	No data	15 [9, 20]
Ventilator usage	Yes (N=24)	22 [14, 31]	10 [6, 31]	11 [4, 21]	19 [12, 23]	13 [12, 33]
	No (N=66)	13 [10, 19]	13 [9, 19]	13 [11, 21]	9 [8, 17]	13 [12, 33]
Mortality	Deceased (N= 25)	18 [12, 27]	16 [9, 33]	16 [2, 30]	15 [9, 23]	No data
	Alive (65)	14 [10, 21]	12 [7, 19]	12 [9, 15]	19 [9, 22]	13 [10, 20]

**eTable 3.** Serum Insulinlike Growth Factor 1 Concentration by Intensive Care Unit Admission, Ventilator Use, and Mortality in Men

		Day 0	Day 3	Day 7	Day 14	Day 28
ICU	Yes (N=53)	87 [57, 117]	78 [51, 116]	72 [41, 111]	110 [41, 124]	No data
	No (N=37)	95 [63, 149]	75 [16, 136]	87 [54, 135]	No data	No data
Ventilator usage	Yes (N=24)	77 [53, 113]	88 [44, 115]	75 [40, 138]	112 [41, 131]	No data
	No (N= 66)	90 [63, 134]	74 [39, 123]	79 [55, 110]	101 [42, 173]	No data
Mortality	Deceased (N= 25)	78 [59, 98]	65 [40, 99]	69 [53, 119]	No data	No data
	Alive (65)	98 [62, 137]	85 [46, 120]	82 [40, 117]	122 [79, 151]	79 [58, 147]

**eTable 4.** Serial Hormone Concentration in Women With and Without Severe COVID-19

	Severe COVID	Day 0	Day 3	Day 7
Testosterone (ng/dl)	Yes	10 [1, 21]	9 [3, 32]	15 [5, 24]
	No	14 [1, 24]	10 [5, 13]	No data
Estradiol (pg/ml)	Yes	10 [4, 50]	20 [10, 31]	16 [8, 47]
	No	20 [2, 45]	13 [8, 42]	No data
IGF-1 (ng/ml)	Yes	92 [52, 128]	70 [40, 91]	65 [43, 105]
	No	108 [66, 139]	87 [55, 151]	No data
Estradiol/Testosterone (%)	Yes	16 [6, 58]	23 [11, 120]	19 [5, 16]
	No	29 [21, 40]	20 [18, 80]	No data

#p<0.05 for comparison with day 0

“No data” is indicated if there were insufficient patients in that category.

**eTable 5.** Serum Estradiol Concentration by Intensive Care Unit Admission, Ventilator Use, and Mortality in Women

		Day 0	Day 3	Day 7
ICU	Yes (N=29)	10 [2, 48]	20 [11, 30]	16 [8, 52]
	No (N=33)	20 [5, 45]	16 [10, 38]	29 [17, 34]
ventilator	Yes (N=13)	47 [5, 57]*	30 [23, 68] <sup>#</sup>	16 [10, 103]
	No (N=49)	10 [4, 28]	14 [9, 25]	21 [8, 32]
Mortality	Deceased (N=12)	8 [2, 56]	22 [15, 24]	16 [11, 108]
	Alive (N=50)	16 [5, 45]	15 [9, 35]	21 [8, 32]

\*p=0.02 as compared to women without ventilator use, adjusted for group differences in age, BMI, CCI, smoking status and race.

<sup>#</sup>p<0.05 for comparison with day 0

**eTable 6:** Serum Testosterone Concentrations by Intensive Care Unit Admission, Ventilator Usage and Mortality in Women

		Day 0	Day 3	Day 7
ICU	Yes (N=29)	10 [4, 21]	8 [3, 22]	15 [4, 24]
	No (N=33)	14 [1, 23]	10 [5, 16]	14 [11, 20]
ventilator	Yes (N=13)	19 [8, 39]	29 [8, 47]	18 [8, 35]
	No (N=49)	10 [1, 20]	7 [3, 14]	14 [8, 17]
Mortality	Deceased (N=12)	15 [4, 39]	9 [1, 45]	35 [4, 39]
	Alive (N=50)	11 [1, 20]	10 [4, 17]	14 [8, 17]

\* $p < 0.05$  as compared to women with comparator group (no ICU stay, no ventilator use or alive), adjusted for group differences in age, BMI, smoking status and race.

**eTable 7.:** Gene Set Enrichment Analyses on Hallmark Gene Sets Demonstrating Pathways That Are Upregulated in 7 Men in Intensive Care Units Compared With 5 Men With Mild Disease in CD14<sup>+</sup>CD16<sup>-</sup> Peripheral Blood Mononuclear Cells

GS follow link to MSigDB	GS DETAILS	SIZE	ES	NES	NOM p-val	FD R q-val	FW ER p-val	RANK AT MAX	LEADING EDGE	
1	<a href="#">HALLMARK TNFA SIGNALING VIA NFKB</a>	<a href="#">Details</a> ...	193	0.59	2.35	0	0	0	3023	tags=35%, list=10%, signal=39%
2	<a href="#">HALLMARK INFLAMMATORY RESPONSE</a>	<a href="#">Details</a> ...	185	0.51	2.04	0	0.001	0.001	4090	tags=32%, list=13%, signal=37%
3	<a href="#">HALLMARK UV RESPONSE DN</a>	<a href="#">Details</a> ...	138	0.51	1.93	0	0.001	0.003	6229	tags=36%, list=21%, signal=45%
4	<a href="#">HALLMARK IL2 STAT5 SIGNALING</a>	<a href="#">Details</a> ...	189	0.48	1.9	0	0.001	0.003	3004	tags=26%, list=10%, signal=29%
5	<a href="#">HALLMARK REACTIVE OXYGEN SPECIES PATHWAY</a>	<a href="#">Details</a> ...	48	0.58	1.86	0	0.001	0.004	4295	tags=38%, list=14%, signal=44%
6	<a href="#">HALLMARK HYPOXIA</a>	<a href="#">Details</a> ...	184	0.47	1.85	0	0.001	0.005	3226	tags=25%, list=11%, signal=28%

7	<a href="#">HALLMARK OXIDATIVE PHOSPHORYLATION</a>	<a href="#">Details</a> ...	199	0.45	1.8	0	0.002	0.009	5685	tags=38%, list=19%, signal=46%
8	<a href="#">HALLMARK COAGULATION</a>	<a href="#">Details</a> ...	115	0.48	1.8	0	0.002	0.001	6073	tags=40%, list=20%, signal=50%
9	<a href="#">HALLMARK ANGIOGENESIS</a>	<a href="#">Details</a> ...	28	0.61	1.72	0.005	0.003	0.024	3456	tags=32%, list=11%, signal=36%
10	<a href="#">HALLMARK XENOBIOTIC METABOLISM</a>	<a href="#">Details</a> ...	183	0.43	1.7	0	0.003	0.026	3941	tags=25%, list=13%, signal=28%
11	<a href="#">HALLMARK KRAS SIGNALING UP</a>	<a href="#">Details</a> ...	178	0.44	1.7	0	0.003	0.027	2937	tags=20%, list=10%, signal=22%
12	<a href="#">HALLMARK TGF BETA SIGNALING</a>	<a href="#">Details</a> ...	52	0.53	1.69	0.004	0.003	0.031	3273	tags=35%, list=11%, signal=39%
13	<a href="#">HALLMARK MTORC1 SIGNALING</a>	<a href="#">Details</a> ...	200	0.41	1.64	0	0.006	0.061	6342	tags=34%, list=21%, signal=43%
14	<a href="#">HALLMARK EPITHELIAL MESENCHYMAL TRANSITION</a>	<a href="#">Details</a> ...	168	0.41	1.6	0	0.008	0.087	6860	tags=37%, list=23%, signal=47%
15	<a href="#">HALLMARK COMPLEMENT</a>	<a href="#">Details</a> ...	189	0.4	1.58	0	0.001	0.107	5096	tags=29%, list=17%, signal=34%



16	<a href="#">HALLMARK HEME METABOLISM</a>	<a href="#">Details</a> ...	188	0.4	1.57	0	0.01	0.18	3315	tags=21%, list=11%, signal=24%
17	<a href="#">HALLMARK INTERFERON GAMMA RESPONSE</a>	<a href="#">Details</a> ...	199	0.39	1.56	0	0.011	0.133	4176	tags=22%, list=14%, signal=25%
18	<a href="#">HALLMARK ADIPOGENESIS</a>	<a href="#">Details</a> ...	195	0.37	1.47	0.003	0.026	0.31	6086	tags=33%, list=20%, signal=41%
19	<a href="#">HALLMARK ESTROGEN RESPONSE EARLY</a>	<a href="#">Details</a> ...	179	0.37	1.46	0.009	0.028	0.339	3231	tags=19%, list=11%, signal=21%
20	<a href="#">HALLMARK ESTROGEN RESPONSE LATE</a>	<a href="#">Details</a> ...	177	0.37	1.45	0.003	0.028	0.366	3167	tags=17%, list=10%, signal=19%
21	<a href="#">HALLMARK WNT BETA CATENIN SIGNALING</a>	<a href="#">Details</a> ...	35	0.46	1.4	0.064	0.045	0.529	5956	tags=43%, list=20%, signal=53%
22	<a href="#">HALLMARK ANDROGEN RESPONSE</a>	<a href="#">Details</a> ...	98	0.38	1.39	0.038	0.048	0.572	3376	tags=20%, list=11%, signal=23%

Pathways shown are based on cutoffs defined as  $q < 0.05$

**eTable 8.** Gene Set Enrichment Analyses on Hallmark Gene Sets Demonstrating Pathways That Are Upregulated in 4 Women in Intensive Care Units Compared With 4 Women With Mild Disease in CD14<sup>+</sup>CD16<sup>-</sup> Peripheral Blood Mononuclear Cells

GS DETAILS	SIZE	E S	N ES	NO M p- val	FD R q- val	FW ER p- val	RA NK AT MA X	LEAD ING EDGE	
<a href="#">HALLMARK EPITHELIAL MESENCHYMAL TRANSITION</a>	<a href="#">Details</a> ...	1 7 9	0. 46	1.6 9	0	0.0 25	0.0 25	7040	tags=37%, list=21%, signal=46%
<a href="#">HALLMARK COAGULATION</a>	<a href="#">Details</a> ...	1 2 8	0. 48	1.6 8	0.0 01	0.0 13	0.0 26	7771	tags=39%, list=23%, signal=51%
<a href="#">HALLMARK PI3K AKT MTOR SIGNALING</a>	<a href="#">Details</a> ...	9 7	0. 47	1.6 2	0.0 01	0.0 19	0.0 56	7260	tags=36%, list=22%, signal=46%
<a href="#">HALLMARK ANGIOGENESIS</a>	<a href="#">Details</a> ...	3 2	0. 55	1.5 5	0.0 2	0.0 41	0.1 57	3652	tags=31%, list=11%, signal=35%

Pathways shown are based on cutoffs defined as  $q < 0.05$

**eTable 9.** Gene Set Enrichment Analyses on Hallmark Gene Sets Demonstrating Pathways That Are Downregulated in 7 Men in Intensive Care Units Compared With 5 Men With Mild Disease in CD14<sup>+</sup>CD16<sup>-</sup> Peripheral Blood Mononuclear Cells

GS DETAILS	SIZE	ES	NE S	NO M p-val	FD R q-val	FW ER p-val	RA NK AT MAX	LEADING EDGE	
<a href="#">HALLMARK_MYC_TARGETS_V2</a>	<a href="#">Details</a>	58	-0.49	-1.68	0	0.015	0.028	5629	tags=45%, list=19%, signal=55%
<a href="#">HALLMARK_E2F_TARGETS</a>	<a href="#">Details</a>	199	-0.39	-1.63	0	0.011	0.043	4401	tags=32%, list=15%, signal=37%

Pathways shown are based on cutoffs defined by  $q < 0.05$ . No pathways were observed to be significantly downregulated in Female ICU patients as compared to those with mild disease in in CD14<sup>+</sup>CD16<sup>-</sup> peripheral blood mononuclear cells.

**eTable 10.** Gene Set Enrichment Analyses on Hallmark Gene Sets Demonstrating Pathways That Are Upregulated in 7 Men in Intensive Care Units Compared With 5 Men With Mild Disease in CD14<sup>+</sup>CD16<sup>+</sup> Peripheral Blood Mononuclear Cells

	GS follow link to MSigDB	GS DETAILS	SIZ E	ES	NE S	NO M p- val	FDR q- val	FW ER p- val	RA NK AT MA X	LEADIN G EDGE
1	<a href="#">HALLMARK TNFA SIGNALING VIA NFKB</a>	<a href="#">Detail s...</a>	19 4	0.5 9	2.1 7	0	0	0	400 7	tags=34 %, list=12% , signal=3 8%
2	<a href="#">HALLMARK CHOLESTEROL HOMEOSTASIS</a>	<a href="#">Detail s...</a>	72	0.5 7	1.9	0	0	0	379 8	tags=36 %, list=11% , signal=4 1%
3	<a href="#">HALLMARK TGF BETA SIGNALING</a>	<a href="#">Detail s...</a>	51	0.5 8	1.8	0	0.0 02	0.00 5	616 6	tags=41 %, list=18% , signal=5 0%
4	<a href="#">HALLMARK IL2 STATS5 SIGNALING</a>	<a href="#">Detail s...</a>	19 3	0.4 9	1.7 9	0	0.0 01	0.00 5	383 5	tags=25 %, list=11% , signal=2 8%
5	<a href="#">HALLMARK MYOGENESIS</a>	<a href="#">Detail s...</a>	17 8	0.4 9	1.7 8	0	0.0 01	0.00 5	349 6	tags=23 %, list=10% , signal=2 6%
6	<a href="#">HALLMARK COAGULATION</a>	<a href="#">Detail s...</a>	12 7	0.4 9	1.7 3	0	0.0 02	0.01 3	925 8	tags=43 %, list=28% , signal=6 0%
7	<a href="#">HALLMARK UV RESPONSE DN</a>	<a href="#">Detail s...</a>	13 8	0.4 7	1.6 8	0.0 01	0.0 04	0.02 5	570 8	tags=30 %, list=17% ,

										signal=36%
8	<a href="#">HALLMARK_HYPOXIA</a>	<a href="#">Detail</a> <a href="#">s...</a>	186	0.46	1.68	0	0.003	0.026	3420	tags=22%, list=10%, signal=24%
9	<a href="#">HALLMARK_INFLAMMATORY_RESPONSE</a>	<a href="#">Detail</a> <a href="#">s...</a>	192	0.44	1.63	0	0.006	0.055	6182	tags=29%, list=19%, signal=36%
10	<a href="#">HALLMARK_ANGIOGENESIS</a>	<a href="#">Detail</a> <a href="#">s...</a>	30	0.57	1.59	0.012	0.001	0.093	4986	tags=37%, list=15%, signal=43%
11	<a href="#">HALLMARK_REACTIVE_OXYGEN_SPECIES_PATHWAY</a>	<a href="#">Detail</a> <a href="#">s...</a>	49	0.5	1.56	0.015	0.013	0.133	4464	tags=31%, list=13%, signal=35%
12	<a href="#">HALLMARK_XENOBIOTIC_METABOLISM</a>	<a href="#">Detail</a> <a href="#">s...</a>	187	0.42	1.54	0	0.014	0.163	6013	tags=26%, list=18%, signal=32%
13	<a href="#">HALLMARK_KRAS_SIGNALING_UP</a>	<a href="#">Detail</a> <a href="#">s...</a>	184	0.42	1.52	0.003	0.018	0.21	4200	tags=20%, list=13%, signal=23%
14	<a href="#">HALLMARK_COMPLEMENT</a>	<a href="#">Detail</a> <a href="#">s...</a>	195	0.41	1.52	0.001	0.017	0.219	6048	tags=30%, list=18%, signal=36%
15	<a href="#">HALLMARK_EPITHELIAL_MESENCHYMAL_TRANSITION</a>	<a href="#">Detail</a> <a href="#">s...</a>	178	0.41	1.49	0.003	0.021	0.275	6487	tags=28%, list=19%, signal=34%

16	<a href="#">HALLMARK_ESTROGEN_RESPONSE_EARLY</a>	<a href="#">Detail</a> <a href="#">S...</a>	189	0.41	1.49	0	0.021	0.29	3609	tags=22%, list=11%, signal=24%
17	<a href="#">HALLMARK_IL6_JAK_STAT3_SIGNALING</a>	<a href="#">Detail</a> <a href="#">S...</a>	83	0.44	1.49	0.016	0.02	0.295	3311	tags=23%, list=10%, signal=25%
18	<a href="#">HALLMARK_ANDROGEN_RESPONSE</a>	<a href="#">Detail</a> <a href="#">S...</a>	99	0.42	1.46	0.017	0.03	0.428	4057	tags=25%, list=12%, signal=29%
19	<a href="#">HALLMARK_WNT_BETA_CATENIN_SIGNALING</a>	<a href="#">Detail</a> <a href="#">S...</a>	38	0.5	1.44	0.046	0.034	0.488	4427	tags=26%, list=13%, signal=30%

Pathways shown are based on cutoffs defined by  $q < 0.05$

**eTable 11.** Gene Set Enrichment Analyses on Hallmark Gene Sets Demonstrating Pathways That Are Upregulated in 4 Women in Intensive Care Units Compared With 4 Women With Mild Disease in CD14<sup>+</sup>CD16<sup>+</sup> Peripheral Blood Mononuclear Cells

	GS DETAILS follow link to MSigDB	SIZE	ES	NE S	NO M p- val	FDR q- val	FW ER p- val	RA NK AT MA X	LEADI NG EDGE	
1	<a href="#">HALLMARK ANGIOGENESIS</a>	<a href="#">Details ...</a>	30	0.68	1.92	0.002	0.002	0.002	1610	tags=27%, list=5%, signal=28%
2	<a href="#">HALLMARK COAGULATION</a>	<a href="#">Details ...</a>	122	0.52	1.86	0	0.002	0.005	5784	tags=35%, list=19%, signal=43%
3	<a href="#">HALLMARK IL6 JAK STAT3 SIGNALING</a>	<a href="#">Details ...</a>	83	0.54	1.82	0	0.002	0.007	4873	tags=34%, list=16%, signal=40%
4	<a href="#">HALLMARK INFLAMMATORY RESPONSE</a>	<a href="#">Details ...</a>	187	0.46	1.75	0	0.005	0.02	3113	tags=25%, list=10%, signal=28%
5	<a href="#">HALLMARK EPITHELIAL MESENCHYMAL TRANSITION</a>	<a href="#">Details ...</a>	176	0.44	1.66	0	0.009	0.049	3591	tags=22%, list=12%, signal=25%
6	<a href="#">HALLMARK IL2 STAT5 SIGNALING</a>	<a href="#">Details ...</a>	192	0.42	1.6	0.001	0.019	0.116	3312	tags=20%, list=11%, signal=23%
7	<a href="#">HALLMARK REACTIVE OXYGEN SPECIES PATHWAY</a>	<a href="#">Details ...</a>	49	0.51	1.59	0.011	0.018	0.123	2649	tags=22%, list=9%, signal=25%

8	<a href="#">HALLMARK_ADIPOGENESIS</a>	<a href="#">Details ...</a>	19 5	0.4 1	1.5 6	0.0 05	0.02 2	0.16 9	8251	tags=44 %, list=27% , signal=6 0%
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Pathways shown are based on cutoffs defined as  $q < 0.05$



**eTable 12.** Gene Set Enrichment Analyses on Hallmark Gene Sets Demonstrating Pathways That Are Downregulated in 7 Men in Intensive Care Units Compared With 5 Men With Mild Disease in CD14<sup>+</sup>CD16<sup>+</sup> Peripheral Blood Mononuclear Cells

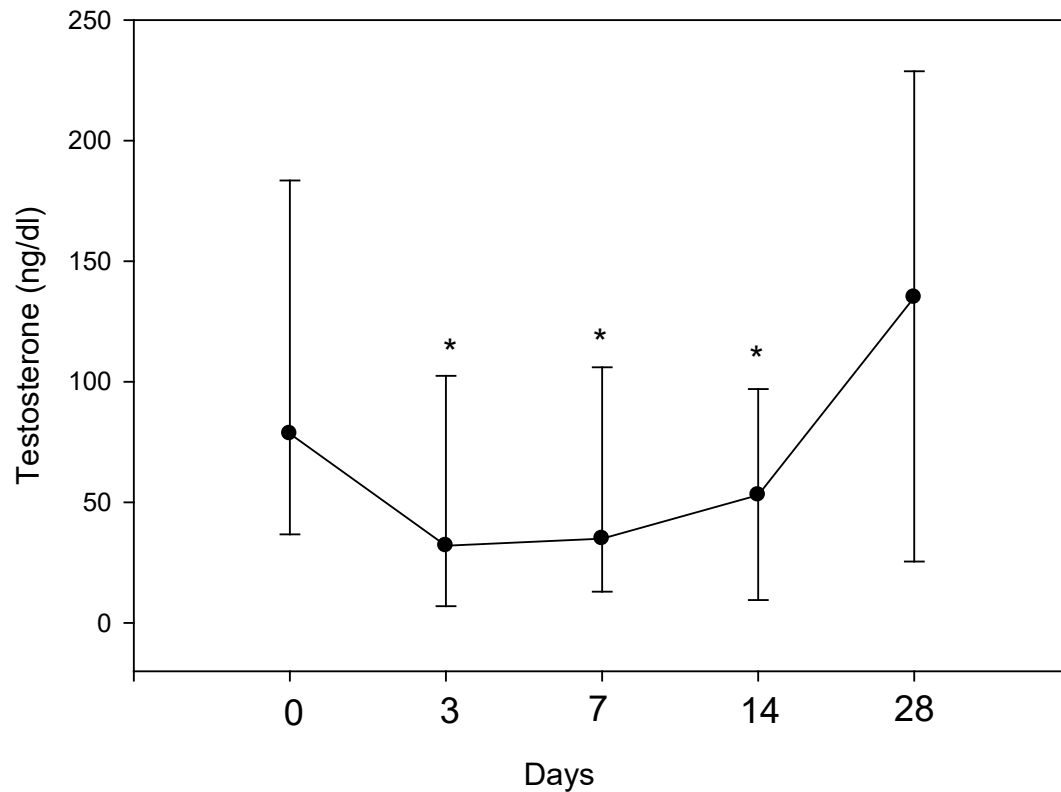
GS	GS DETAILS	SIZE	ES	NE S	NO M p-val	FDR q-val	FWE R p-val	RANK AT MAX	LEADING EDGE	
1	<a href="#">HALLMARK MYC TARGETS V1</a>	<a href="#">Details...</a>	200	-0.37	-1.67	0	0.019	0.017	4454	tags=29%, list=13%, signal=34%
2	<a href="#">HALLMARK MYC TARGETS V2</a>	<a href="#">Details...</a>	58	-0.44	-1.64	0.012	0.012	0.023	4074	tags=31%, list=12%, signal=35%
3	<a href="#">HALLMARK E2F TARGETS</a>	<a href="#">Details...</a>	198	-0.33	-1.53	0	0.02	0.056	3927	tags=28%, list=12%, signal=

Pathways shown are based on cutoffs defined as  $q < 0.05$

**eTable 13.** Gene Set Enrichment Analyses on Hallmark Gene Sets Demonstrating Pathways That Are Downregulated in 4 Women in Intensive Care Units Compared With 4 Women With Mild Disease in CD14<sup>-</sup>CD16<sup>+</sup> Peripheral Blood Mononuclear Cells

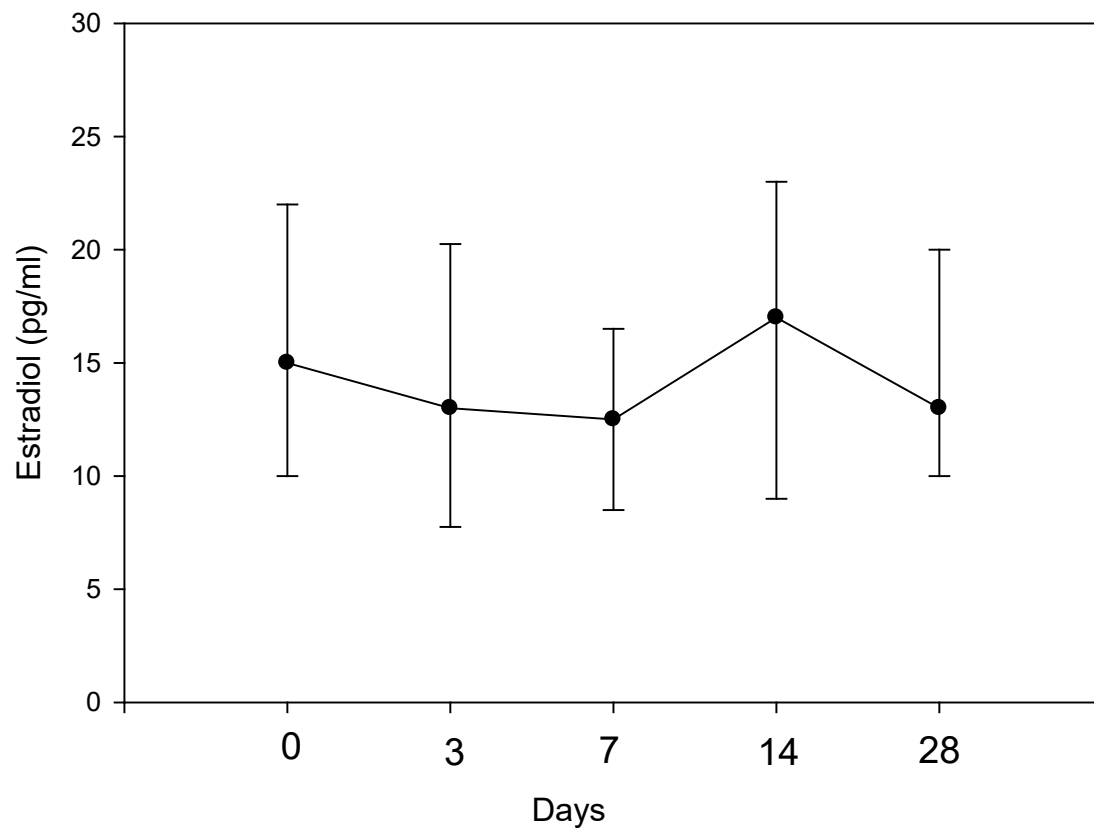
GS DETAILS	SIZE	ES	NE S	NO M p-val	FDR q-val	FWE R p-val	RAN K AT MA X	LEADIN G EDGE	
<a href="#">HALLMARK MITOTIC SPINDLE</a>	<a href="#">Detail</a> <a href="#">Is ...</a>	198	-0.37	-1.6	0	0.072	0.064	5838	tags=35%, list=19%, signal=43%
<a href="#">HALLMARK E2F TARGETS</a>	<a href="#">Detail</a> <a href="#">Is ...</a>	198	-0.36	-1.57	0	0.042	0.073	5859	tags=31%, list=19%, signal=38%
<a href="#">HALLMARK P53 PATHWAY</a>	<a href="#">Detail</a> <a href="#">Is ...</a>	193	-0.37	-1.57	0	0.028	0.073	3916	tags=23%, list=13%, signal=27%
<a href="#">HALLMARK TGF BETA SIGNALING</a>	<a href="#">Detail</a> <a href="#">Is ...</a>	534	-0.44	-1.56	0.018	0.022	0.078	2192	tags=23%, list=7%, signal=24%
<a href="#">HALLMARK G2M CHECKPOINT</a>	<a href="#">Detail</a> <a href="#">Is ...</a>	198	-0.3	-1.31	0.026	0.146	0.481	4658	tags=26%, list=15%, signal=31%

Pathways shown are based on cutoffs defined as  $q < 0.05$



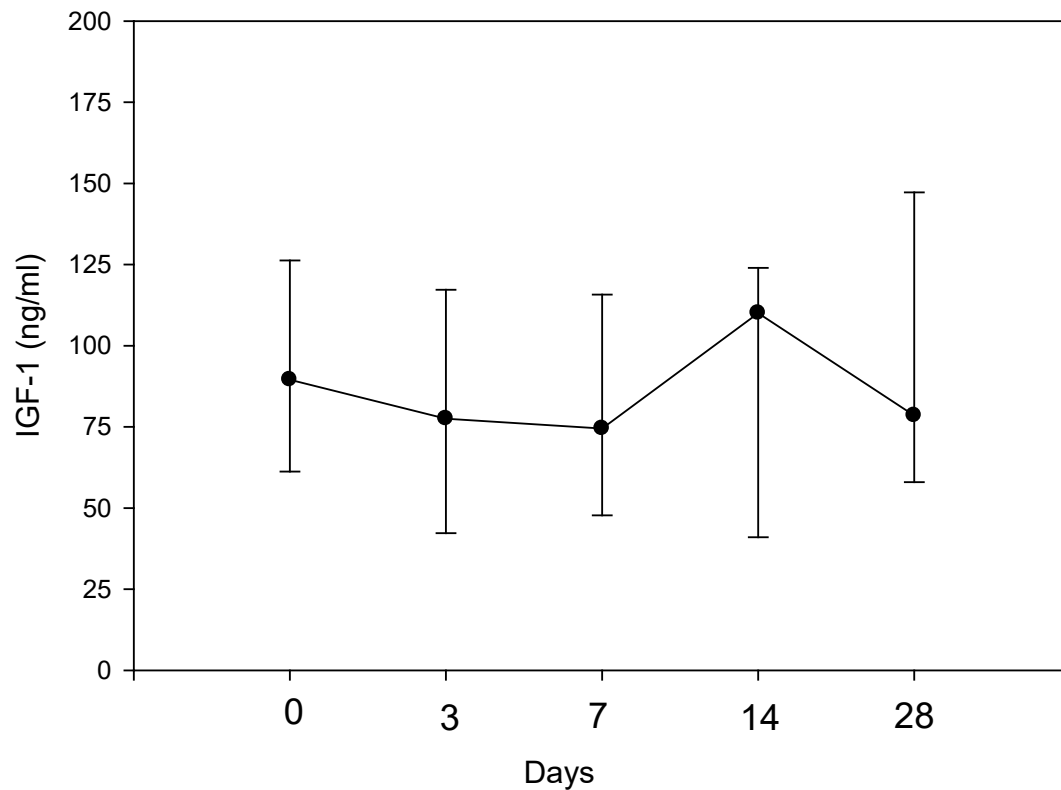
**eFigure 1.** Testosterone Concentrations During Hospital Stay in Male Patients

Values are median [25<sup>th</sup>, 75<sup>th</sup> percentile]. Day 0 includes the 6 patients who were not hospitalized. Serum testosterone concentrations were available on days 0, 3, 7, 14 and 28 in 76, 53, 41, 21 and 12 patients, respectively. \*  $p < 0.05$  as compared to day 0.



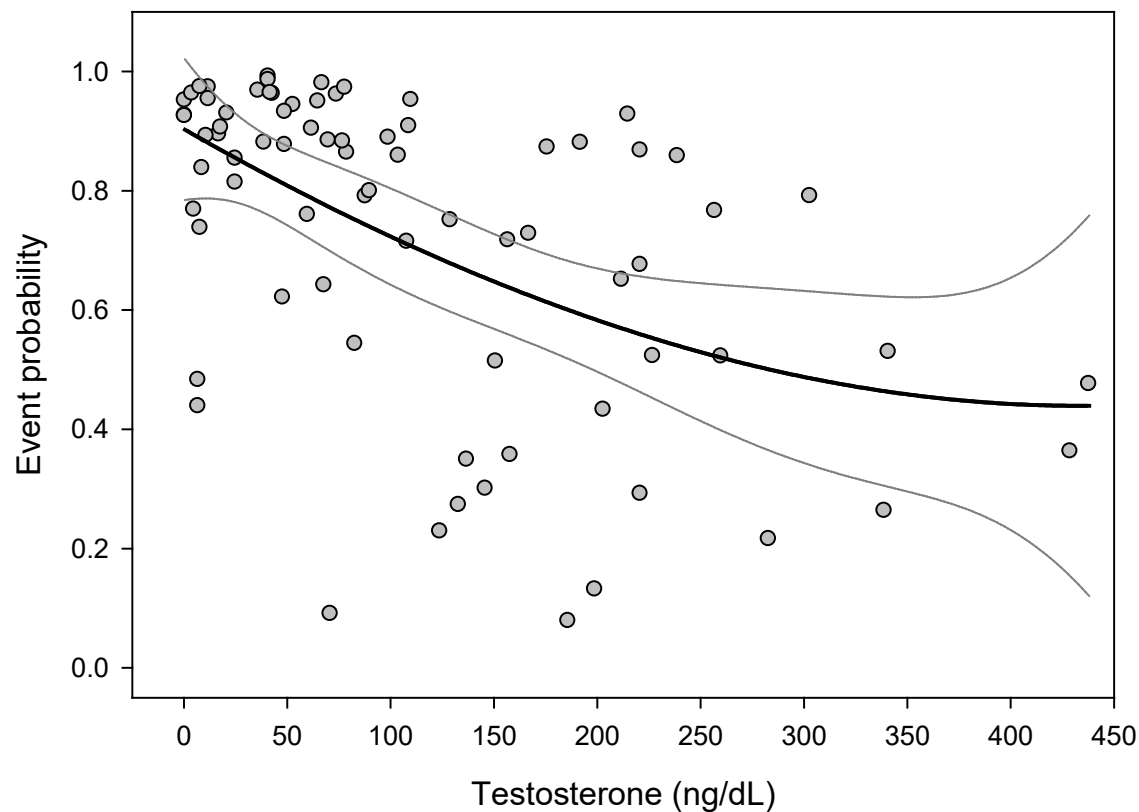
**eFigure 2.** Estradiol Concentrations During Hospital Stay in Male Patients

Values are median [25<sup>th</sup>, 75<sup>th</sup> percentile]. Day 0 includes the 6 patients who were not hospitalized. Serum estradiol concentrations were available on days 0, 3, 7, 14 and 28 in 55, 42, 30, 19 and 11 patients, respectively.



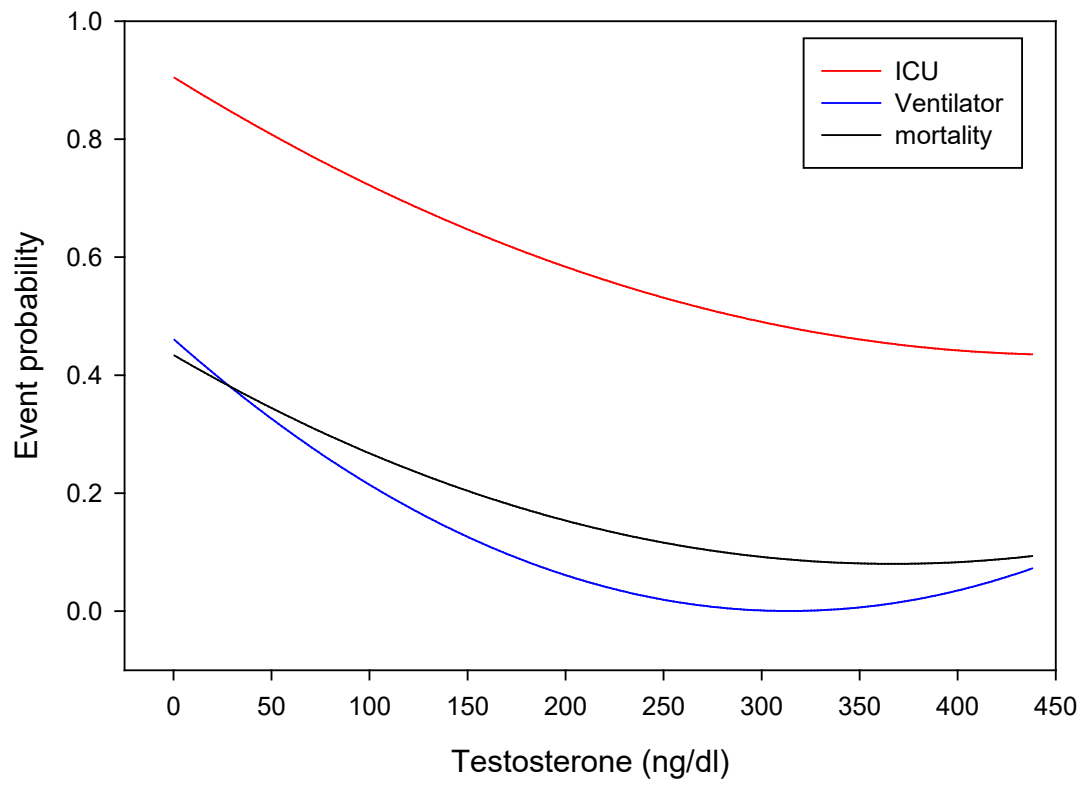
**eFigure 3.** Insulinlike Growth Factor 1 Concentrations During Hospital Stay in Male Patients

Values are median [25<sup>th</sup>, 75<sup>th</sup> percentile]. Day 0 includes the 6 patients who were not hospitalized. Serum IGF-1 concentrations were available on days 0, 3, 7, 14 and 28 in 64, 36, 30, 15 and 10 patients, respectively.



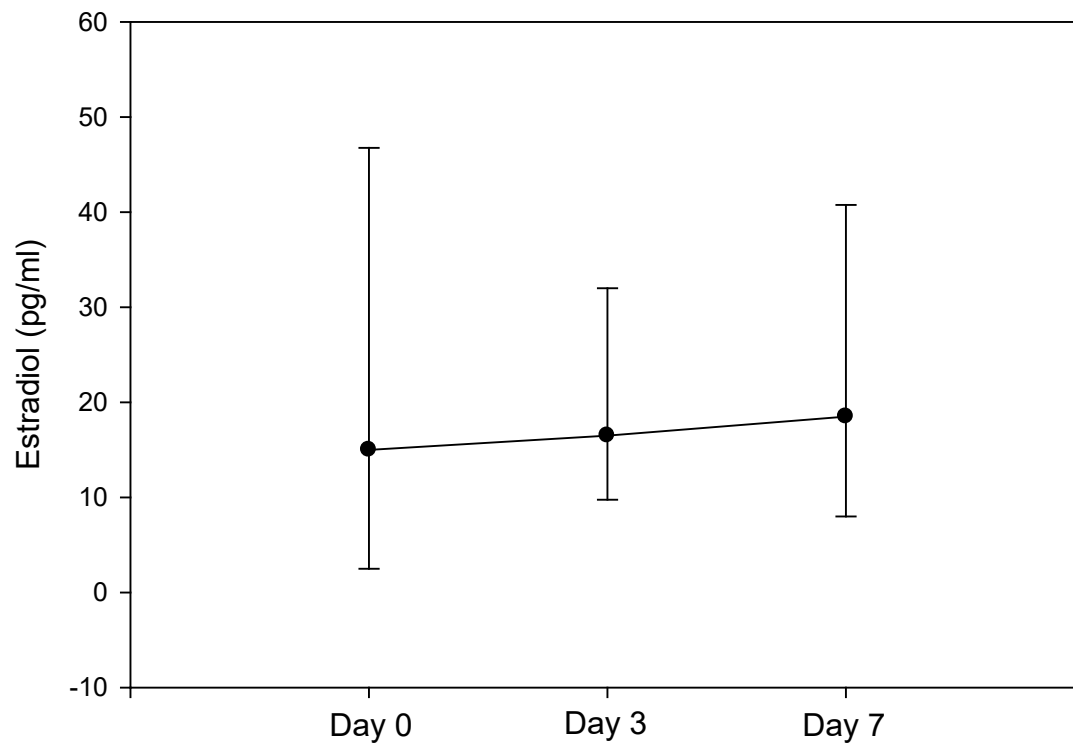
**eFigure 4.** Regression Curve Demonstrating Probability of COVID-19 Severity as Predicted by Testosterone

Includes 95% confidence intervals. After multivariate adjustment for age, BMI, CCI, smoking and race.



**eFigure 5.** Regression Curves Demonstrating Probability of Intensive Care Unit Admission, Ventilator Usage, or Mortality as Predicted by Testosterone

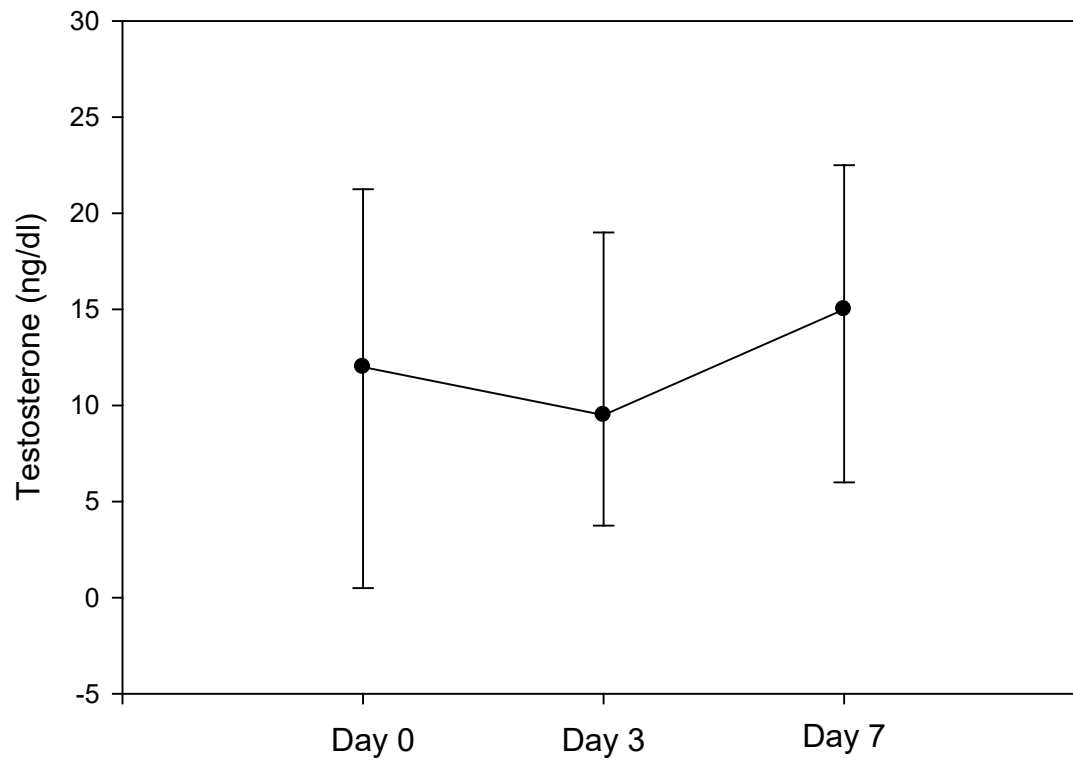
After multivariate adjustment for age, BMI, CCI, smoking and race.



**eFigure 6.** Estradiol Concentrations in Women at Days 0, 3, and 7

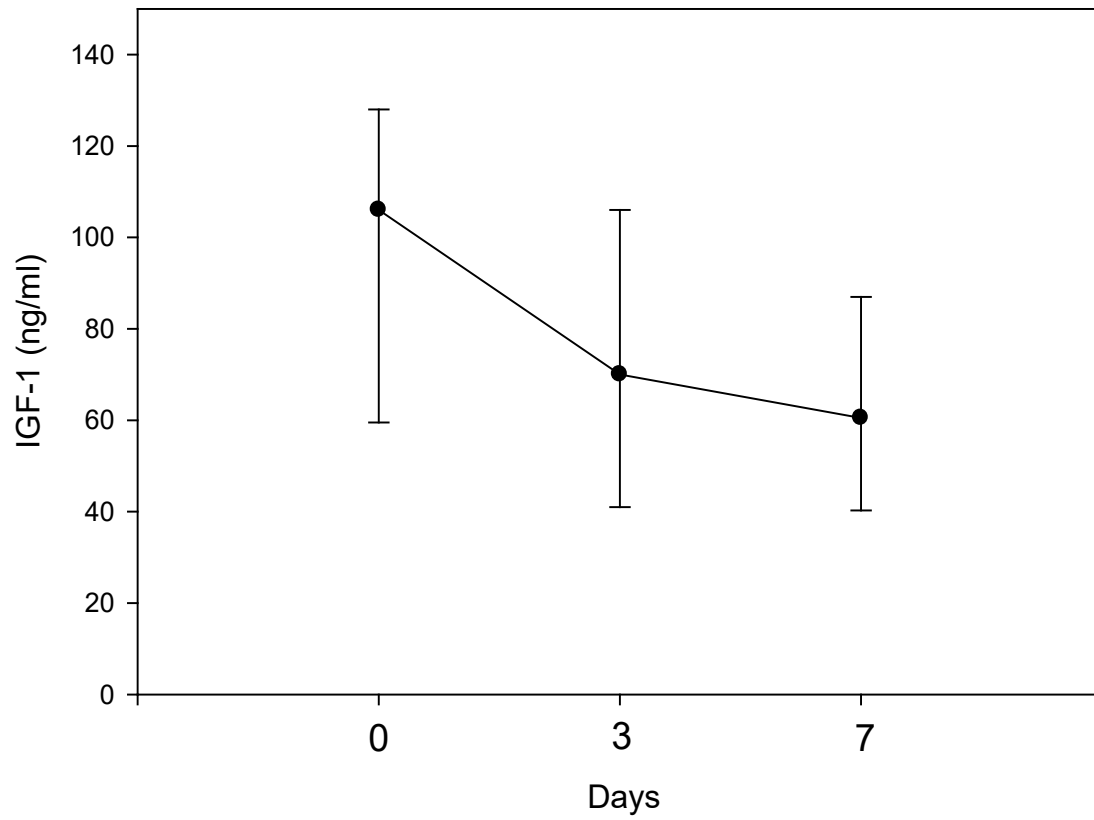
Values are median [25<sup>th</sup>, 75<sup>th</sup> percentile]. Estradiol concentrations were available on days 0, 3 and 7 in 44, 26 and 18 patients, respectively.





**eFigure 7.** Testosterone Concentrations in Women at Days 0, 3, and 7

Values are median [25<sup>th</sup>, 75<sup>th</sup> percentile]. Testosterone concentrations were available on days 0, 3 and 7 in 56, 30 and 21 patients, respectively.



**eFigure 8.** Insulinlike Growth Factor 1 Concentrations in Women at Days 0, 3, and 7

Values are median [25<sup>th</sup>, 75<sup>th</sup> percentile]. IGF-1 concentrations were available on days 0, 3 and 7 in 49, 23 and 14 patients, respectively.

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