

The interaction between the renin-angiotensin system and peroxisome proliferator activated receptors: a hypothesis including the participation of mitochondria in aging

Elena M. V. de Cavanagh¹, Barbara Piotrkowski¹, and Cesar G. Fraga^{1,2}

¹ Physical Chemistry-PRALIB, School of Pharmacy and Biochemistry, University of Buenos Aires, Argentina, ² Department of Nutrition, University of California, Davis, California, USA

TABLE OF CONTENTS

1. Abstract
2. Introduction
3. Oxidants and mitochondria in the aging process
 - 3.1. Oxidant species, antioxidants, and oxidative stress
 - 3.2. Mitochondria as sources and targets of reactive oxygen species and reactive nitrogen species
 - 3.3. Mitochondria and peroxisome proliferator activated receptors
 - 3.4. Aging, mitochondria, and peroxisome proliferator activated receptors
4. The renin-angiotensin system, mitochondria, and aging
 - 4.1. The renin-angiotensin system and the cellular generation of ROS and NO
 - 4.2. The renin-angiotensin system and mitochondria
 - 4.3. The renin-angiotensin system and peroxisome proliferator activated receptors
 - 4.4. The renin-angiotensin system and aging
 - 4.5. Renin-angiotensin system inhibition in hypertension and diabetes
5. Caloric restriction, aging, oxidative stress, mitochondria and peroxisome proliferators activated receptors
6. Renin-angiotensin system inhibition and caloric restriction, two strategies converging at biological effects that include peroxisome proliferator activated receptors
7. Conclusion and perspectives
8. References

1. ABSTRACT

The objective of improving health is intimately associated with preventing and delaying age-related diseases. Nutritional and pharmacological approaches aimed at retarding aging are uncovering mechanisms, whose definitive roles in cell and tissue physiology need to be defined. In this article we hypothesize that peroxisome proliferator activated receptor (PPAR)-modulation is a pivotal process that underlies the association between mitochondria and the renin-angiotensin system (RAS) in aging. This hypothesis is based on several lines of evidence suggesting that: a) mitochondrial function and oxidant production are active participants in the aging process; b) PPARs, by regulating mitochondrial function and uncoupling proteins (UCP), seem to play a major role in the age-retarding effects of caloric restriction; c) RAS inhibition delays the deleterious effects of aging and also upregulates PPARs; and d) a number of physiological and molecular events that occur in experimental caloric restriction, and experimental and clinical RAS inhibition, involve changes in mitochondrial functions.

2. INTRODUCTION

The inhibition of the renin-angiotensin system (RAS), a broadly used therapeutic strategy to counteract hypertension, provides ancillary health benefits apparently

unrelated to the lowering of blood pressure (1-4). The mechanisms responsible for those beneficial effects are not completely characterized. We have extensively investigated the effects of RAS inhibition on mitochondrial function, mitochondrial oxidant production and oxidative damage to mitochondria (5-8). Based on both our findings and published evidence, in this review we will discuss the hypothesis of a pivotal role of peroxisome proliferator activated receptor (PPAR)-modulation in the preservation of mitochondrial function to explain the beneficial health effects of RAS inhibition. Considering the relevant participation of mitochondria (9-12) and the RAS (13-15) in both, aging and aging-associated diseases, we will focus most of our discussion on aging.

3. OXIDANTS AND MITOCHONDRIA IN THE AGING PROCESS

3.1. Oxidant species, antioxidants, and oxidative stress

Reactive oxygen species (ROS) and reactive nitrogen species (RNS) are generic terms that include byproducts of oxygen metabolism continuously generated in aerobic organisms (10, 16). ROS and RNS can react with other biologically relevant molecules (e.g. lipids, proteins, and nucleic acids) leading to cell and tissue damage (17). Aerobic organisms are endowed with an assorted group of

Mitochondria, renin-angiotensin system and aging

molecules, referred to as antioxidants, to control oxidant production and prevent cell and tissue damage (18). Antioxidants essentially include both, low molecular weight compounds able to scavenge or deactivate oxidants, and enzymes that catalyze the decomposition of oxidants. In addition, non-enzyme proteins (uncoupling proteins, thioredoxin, histones, etc.) as well as other biomolecules (plant polyphenols, etc.) can afford antioxidant protection to biological systems by mechanisms that do not involve direct oxidant reduction (18). However, under certain circumstances, control over ROS and RNS levels is lost, resulting in oxidative stress. Oxidative stress is a concept defined by Sies (19) as “an imbalance between oxidants and antioxidants in favor of the oxidants, potentially leading to damage”.

3.2. Mitochondria as sources and targets of reactive oxygen species and reactive nitrogen species

Mitochondria are essential for the maintenance of aerobic life not only as sources of energy, but also by regulating Ca^{2+} homeostasis (20), tissue O_2 gradients (21), cell apoptosis (22), and intracellular signaling (23). However, the partial reduction of O_2 by the mitochondrial respiratory chain components results in an undesirable production of oxidants. Thus, mitochondria are a major source of ROS, and are themselves targets of ROS-mediated damage. Mitochondria produce superoxide anion (O_2^-) by univalent reduction of O_2 (10) at two sites of the electron transport chain (24, 25), and hydrogen peroxide (H_2O_2) as the product of the enzymatic reduction of O_2^- by superoxide dismutase. Both, O_2^- and H_2O_2 , are ROS that can initiate free radical chain reactions leading to the generation of other oxidants, and potentially to a situation of oxidative stress. Further reduction of H_2O_2 to water is catalyzed by the enzymes catalase and glutathione peroxidase. If the targets of ROS are molecules that can propagate free radical reactions (e.g. lipids), damage can extend inside the cells leading to the oxidation of other relevant molecules (proteins, nucleic acids, etc.). Nitric oxide (NO) is an essential signaling molecule that is additionally involved in ROS-mediated oxidations. Nitric oxide is produced by the oxidation of L-arginine to L-citrulline, a reaction catalyzed by several nitric oxide synthase (NOS) isoforms (26). By diffusion-limited reaction with O_2^- , NO yields peroxynitrite (ONOO^-), a very strong oxidant. Nitric oxide, peroxynitrite, and other oxidant species derived from NO are termed RNS. The existence of a constitutive mitochondrial activity of NOS (mtNOS) (27, 28) underlies the biological relevance of NO generation in mitochondria.

Current evidence supports a prominent role for ROS and RNS in both, the decline of mitochondrial function and the increase in mitochondrial DNA oxidation that occurs in various tissues upon aging (29-33).

To minimize the production of ROS and RNS, mitochondria drive O_2 reduction to water with very high efficiency (accounting for about 97-98% of the O_2 respired) (10). Additionally, to control the damage inflicted by the partial reduction of the remaining 2-3% of respired O_2 , mitochondria are well equipped with a battery of enzymes that metabolize ROS, including manganese superoxide

dismutase (Mn-SOD), catalase, and glutathione peroxidase. Other enzymes, such as glutathione reductase and thioredoxin reductase, that catalyze the NADPH-dependent reduction of disulfides, can be also considered part of the mitochondrial antioxidant defenses. Finally, there are other mitochondrial proteins that can limit the production of oxidants, e.g. uncoupling proteins (UCP).

Recently, the concept has emerged that mitochondria not only receive signals from elsewhere in the cell, but they also generate signaling molecules (23). For example, it has been demonstrated that H_2O_2 and NO diffuse from the mitochondria into the cytosol modulating redox-sensitive signaling pathways (34, 35).

In summary, mitochondria are not only recognized as the main sources of cellular energy, but also as organelles that by driving ROS and RNS generation play other key roles in the regulation of cell functions and survival.

3.3. Mitochondria and peroxisome proliferator activated receptors

PPARs are nuclear transcription factors that regulate the expression of a number of genes related to lipid metabolism and energy homeostasis (36). PPAR isoforms are members of the nuclear hormone receptor superfamily of transcription factors, and include PPAR-alpha, PPAR-beta, and PPAR-gamma. Upon ligand-binding, PPARs heterodimerize with 9-cis-retinoic acid receptor (RXR), thereby acquiring the ability to recognize and bind peroxisome proliferator-responsive elements (PPRE) in target genes. All three PPAR subtypes bind the same PPRE (37) and can interact with either RXR-alpha, beta, or gamma. Most of the PPAR ligands that have been studied are synthetic, and a limited number are of endogenous origin (38).

PPAR-alpha is expressed highly in liver, heart and skeletal muscle (39). At the mitochondrial level, activation of PPAR-alpha results in an increased expression of many nuclear genes associated with mitochondrial function, including those involved in fatty acid beta-oxidation (38), mitochondrial proton leak (40, 41), and those encoding antioxidant enzymes, i.e. Mn-SOD and catalase (42). PPAR-gamma, which is highly expressed in adipocytes, is involved in adipocyte differentiation and controls the expression of lipid storage genes. Another relevant function of PPAR-gamma is the promotion of insulin sensitivity (43). PPAR-delta is involved in the regulation of fatty acid catabolism, metabolic rate and proliferation of mitochondria mainly in skeletal muscles (44).

3.4. Aging, mitochondria, and peroxisome proliferator activated receptors

The participation of mitochondria in the continuous production of ROS and RNS supports the mitochondrial free radical theory of aging as formulated in 1979 by Chance *et al.* (10) as an extension of the more general free radical theory of aging (9, 12, 45). Concerning aging and mitochondrial ROS generation, it was recently reported that the mitochondria targeted overexpression of catalase, extends the median and maximum lifespan in

Mitochondria, renin-angiotensin system and aging

mice (46). This life extension concurred with decreases in oxidant damage to DNA, mitochondrial H₂O₂ production, aconitase inactivation, and in the accumulation of mitochondrial DNA deletions (46), suggesting a link between mitochondrial ROS generation, mitochondrial damage, and aging.

In addition, age-associated diseases, including hypertension, diabetes, cancer, and cardiovascular pathologies, are often accompanied by alterations in lipid metabolism, which is largely modulated by mitochondrial activity. The involvement of PPAR- α and PPAR- γ in the above mentioned diseases is underscored by studies showing the beneficial effects of PPAR- α and PPAR- γ agonist administration (47-51). Aging is associated with a decline in the expression of PPAR- α and γ (52). In addition, in skeletal muscle cells, increased ROS production was shown to downregulate PPAR- α mRNA (53). In this setting, a body of evidence support a strong association between PPAR activities and the aging process (54).

Relationships among PPARs, oxidative stress, and aging are supported by studies showing that ligand activated PPAR- α can suppress the age-dependent augmentation of both, oxidative stress and oxidant-dependent NF- κ B activation (55). These results suggested that the prooxidant state displayed by cells of aged animals may be related to the age-related decline in PPAR- α expression (34). Furthermore, PPAR- α activators upregulate uncoupling protein-2 (UCP-2) expression (41), and treatment with PPAR- γ agonists increases UCP-2 mRNA levels in adipose and skeletal muscle cells (56, 57).

UCPs are proton gradient-dissipating mitochondrial proteins that seem to act in both, the regulation of fatty acid and glucose oxidation (58, 59), and the control of O₂⁻ generation by mitochondria (58-60). PPAR- γ activation increased endothelial NO production in bovine aortic endothelial cells (61). The participation of NO as a downstream effector of PPARs activation has also been observed in animal models. In rat kidney, activation of PPAR- α promoted Na⁺ excretion by increasing NO generation (62). In hypertensive rats, the cardioprotective effects afforded by PPAR- α activation were mediated by an increase in NO production and/or an inhibition of NADPH oxidase activity (63).

The above evidence suggests that PPAR- α , through mechanisms involving UCP-2, may lower mitochondrial oxidant production, and, consequently, retard the aging process as well the development of age-associated diseases.

4. THE RENIN-ANGIOTENSIN SYSTEM, MITOCHONDRIA, AND AGING

4.1. The renin-angiotensin system and the cellular generation of ROS and NO

In the classical view, the RAS was exclusively recognized by its circulating actions as a regulator of

systemic blood pressure and renal electrolyte balance. After the discovery of RAS gene expression and function in a variety of tissues, a role for RAS as regulator of organ functions (autocrine) was acknowledged. These local tissue effects of RAS are a subject of current research, and appear to be distinct from circulating RAS actions (64). The major components of both, circulating and tissue RAS, include the polypeptides angiotensinogen, angiotensin I (Ang-I) and angiotensin II (Ang-II). Ang-I is the product of enzymatic angiotensinogen cleavage by renin, and Ang-II is the product of Ang-I cleavage by Ang-I-converting enzyme (ACE) (65). Ang-II, the main effector of the RAS, is responsible for vasoconstriction and Na⁺ retention (66). Ang-II is also a pro-inflammatory and a pro-fibrotic agent (67, 68).

Abundant evidence supports the notion that both, the increased generation of cellular ROS, and the activation of redox-sensitive signaling cascades are critical events involved in Ang-II actions (69). Many of the functional effects of Ang-II are mediated via the angiotensin-II-receptor 1 (AT1). Ang-II binding to AT1 triggers intracellular O₂⁻ production by NAD(P)H oxidase activation (70, 71), and also as a result of endothelial NOS (eNOS) uncoupling (72). On account of O₂⁻ production, cellular NO steady state level may be compromised due to the reaction of NO with O₂⁻ to generate peroxynitrite. As a final consequence of this series of events, and as it has been observed in a variety of cell types (72, 73), Ang-II promotes the production of both, ROS and RNS, and reduces NO availability. Relevant to endothelial cell NO production, it has been demonstrated that bradykinin, another ACE substrate, binds to BK1 and BK2 plasma membrane receptors, thereby activating endothelial NOS (eNOS) (74).

Under normal physiological conditions, Ang-II-mediated ROS and RNS production, and the resulting stimulation of redox-sensitive signaling pathways, are closely regulated (73). However, under conditions leading to overactivation of the RAS, such as hypertension, diabetes (75, 76) and normal aging (77-80), Ang-II-dependent oxidant generation may become a significant contributor to cell oxidation and tissue damage. In vascular smooth muscle cells from spontaneously hypertensive rats (SHR), Ang-II was shown to enhance the activation of NF- κ B and AP-1, two transcription factors modulated by the cellular redox status (81). In addition, it was shown that antioxidants and antioxidant enzymes inhibit the regulatory effects of Ang-II on Ras/Raf/ERK and AP-1 signaling pathways (82, 83).

4.2. The renin-angiotensin system and mitochondria

The relationships between RAS and mitochondria started to gain attention when it became evident that, by prompting ROS formation, RAS effectors can activate redox-sensitive transcription factors involved in the modulation of mitochondrial function.

A causative link between RAS activation and alteration of mitochondrial function is supported by the observation that in mice, acute (24 h) and long-term (14 d)

Mitochondria, renin-angiotensin system and aging

Ang-II infusion leads to decreased cardiac expression of mitochondrial electron transfer chain and Krebs-TCA cycle genes (84). In the short term treatment, Ang-II also promoted the depression of mitochondrial metabolism and Mn-SOD gene, and induced the expression of genes for proteins that protect against oxidative stress, such as thioredoxin, glutaredoxin and transferring receptor 1 (84). These effects are in line with previous observations that indicated the participation of Ang-II in the depression of mitochondrial energy metabolism (85-87). In addition, recent findings show that Ang-II stimulates mitochondrial ROS production associated with the reduction of mitochondrial membrane potential (88).

In addition, there is some evidence supporting a direct interaction between Ang-II and nuclear and mitochondrial components. A number of studies have produced evidence for the existence of nuclear AT1-like AngII receptors in rat liver and spleen cells. In studies using ¹²⁵I-labeled Ang-II, the presence of Ang-II was shown in nuclei and mitochondria of heart, brain and smooth muscle cells (89, 90). Treatment of isolated rat liver nuclei with Ang-II stimulated the transcription of specific genes (91, 92). In addition AT1 receptor blockers (losartan) inhibit Ang-II binding to nuclear receptors in rat hepatocytes (91, 93). The inhibition of Ang-II binding to nuclear receptors by losartan might suggest this possibility. In rat adrenal cortex, immunocytochemical and biochemical evidence indicates the presence of renin, angiotensinogen and ACE within intramitochondrial dense bodies of the zona glomerulosa (94).

Concerning cell signaling modulation by mitochondria, c-Jun NH₂-terminal kinase (JNK) is both, a downstream target of AT1 dependent signaling (95) and a regulator of AP-1 activity. Since AP-1 regulates cytochrome c expression (96), it was suggested that JNK may facilitate changes in the mitochondrial content of cytochrome c in response to Ang-II (97). Adding to previous evidence that supports the participation of the RAS in mitochondrial function responses, the modulation of heart and liver mitochondrial NOS activity and H₂O₂ production by the ACE inhibitor enalapril has been observed in rats (98, 99). Moreover, a recent study showed that the expression of genes related to fatty acid beta-oxidation, mitochondrial proton-electron coupling, and oxidative phosphorylation were up-regulated in captopril-treated diabetic animals, suggesting that RAS inhibition with ACE inhibitors may protect the myocardium by enhancing energy supply (100).

4.3. The renin-angiotensin system and peroxisome proliferator activated receptors

Ang-II infusion was shown to downregulate PPAR-alpha and PPAR-gamma mRNA and protein in apolipoprotein E-deficient (apoE-KO) mice (101). This downregulation occurred in parallel with the activation of NF-kappaB and NF-kappaB-mediated proinflammatory genes (101). Conversely, activators of PPAR-alpha and PPAR-gamma antagonize the proliferative, inflammatory and oxidant-generating actions of Ang-II both, *in vivo* and *in vitro* (43, 102, 103). In this context, it is possible to

hypothesize that RAS inhibition lowers oxidant production not only by blocking Ang-II activation of NADPH-oxidase, but also by regulating the expression of PPARs. In this regard, enalapril was shown to upregulate PPAR-alpha and PPAR-gamma while displaying antiatherogenic and anti-inflammatory effects in mice (104). Two AT1-receptor blockers, irbesartan and telmisartan, were identified as activators of PPAR-gamma (105, 106). Recently, a product of the hepatic metabolism of losartan (EXP3179) was identified as a partial PPAR-gamma agonist, suggesting that some of the AT1-receptor blockers can mediate AT1-receptor independent effects (107).

4.4. The renin-angiotensin system and aging

Based on evidence suggesting the involvement of mitochondria in the aging process, we have investigated how the modulation of the RAS can affect mitochondrial dysfunction associated with aging and aging-associated diseases. In one study, we set forth to investigate whether long-term RAS inhibition, either by treatment with an ACE inhibitor (enalapril) or with an AT1 receptor blocker (losartan) might attenuate structural and functional changes that occur in mitochondria upon aging. Supporting our hypothesis, kidney mitochondria from old rats (22 month-old) that were treated for 8 months with enalapril or losartan, showed an improved capacity for energy production, a lower rate of H₂O₂ production, a higher activity of mtNOS, and a higher content of UCP-2, compared to mitochondria isolated from untreated old rats (8) (Figure 1). In the same study we observed a general improvement in mitochondrial number and structure. Proximal tubular epithelial cells from the enalapril- or losartan-treated old rats showed a higher number of mitochondria, a better definition of mitochondrial cristae, and a lower number of osmiophilic bodies (probably derived from lipid oxidation). Furthermore, both treatments (enalapril or losartan) prevented the alteration of mitochondrial distribution inside proximal tubular cells that was observed in untreated old rats, and attenuated glutathione oxidation in renal tissue (8). These results indicate that RAS inhibition, regardless of how it is implemented, protects mitochondrial components and function from certain effects of aging.

A body of evidence supports the concept that vascular Ang-II presence and responsiveness increase with aging (78-81), and that the RAS may contribute to the development of age-related tissue damage (108). Inhibition of the RAS, either with ACE inhibitors or AT1 receptor blockers, can attenuate several effects of aging in rodents (109, 110). Numerous studies have shown that RAS inhibition prevents different aspects of renal, cardiac, vascular, and behavioral changes that are associated with the aging process in rodents (111).

The potential associations between RAS inhibition and PPAR modulation in aging are congruent with: i) the relatively high levels of Ang-II present in aged organisms (77-80); and ii) the prevention of age-dependent decreases in UCP content that occur secondary to RAS inhibition (8). Furthermore, the stimulation of PPAR-alpha dependent transcription of nuclear genes involved in

Mitochondria, renin-angiotensin system and aging

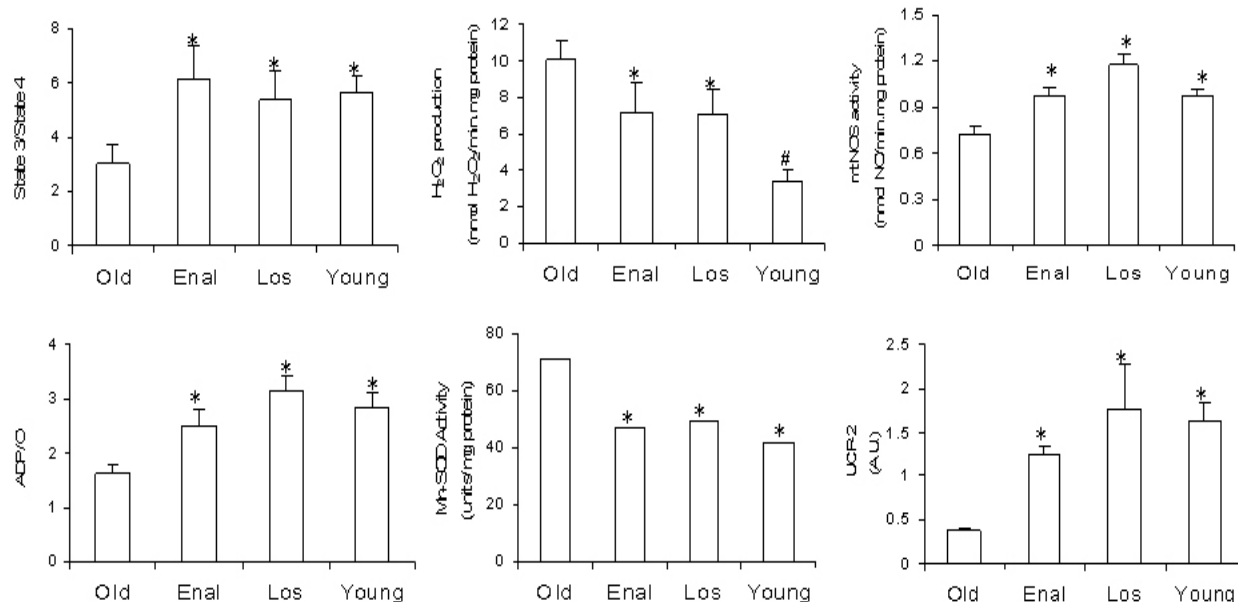


Figure 1. Enalapril and losartan treatments make mitochondrial functions in old rats resemble those of young rats. Respiratory control (respiratory state 3/ respiratory state 4), ADP/O, H₂O₂ production, Mn-SOD activity, nitric oxide synthase (mtNOS) activity, and UCP-2 content, in kidney mitochondria from 22-month old untreated rats (Old); 22-month old rats treated with enalapril (Enal) or losartan (Los) during 8 months; and 4-month old untreated rats (Young). *p< 0.05 vs. Old; #p< 0.01 vs Old, Enal, and Los. Data taken from (8).

mitochondrial fatty acid oxidation would enhance the generation of electron donors for the respiratory chain, thus suppressing the age-dependent deficit of ATP production. This is supported by our observation of an increase in mitochondrial ADP/O ratios following RAS inhibition in aging rats (8).

4.5. Renin-angiotensin system inhibition in hypertension and diabetes

Pharmacological RAS inhibition, either with ACE inhibitors or AT1 receptor blockers, is widely used in patients with hypertension, cardiovascular disease and diabetes (112-114). In these pathological states, the cardiac and renal benefits of RAS inhibition go beyond its blood pressure lowering effects (112, 113, 115) suggesting that ACE inhibitors and AT1 receptor blockers can exert tissue actions that are not associated with their hemodynamic effects.

Incidentally, both, mitochondrial dysfunction and RAS have been independently implicated in hypertension (116) and diabetes (117). These findings led us to investigate whether RAS inhibition might protect mitochondria from damage related to both pathological conditions. In spontaneously hypertensive rats (SHR), treatment with losartan prevented the alterations in kidney mitochondrial membrane potential, UCP-2 content, H₂O₂ production rate, and in the activities of mtNOS, Mn-SOD, and cytochrome oxidase that occurred in untreated SHR (118). In rats rendered diabetic by streptozotocin injection, losartan protected kidney mitochondria against changes in membrane potential, H₂O₂ production rate, and pyruvate content, without lowering plasma glucose content (119). In

both studies, the administration of amlodipine, a Ca²⁺ channel blocker, reduced blood pressure to an extent similar to losartan, but showed no beneficial effects on kidney mitochondria alterations. The results obtained in these two rat models of hypertension and diabetes, indicate: a) a dissociation between blood pressure lowering and the improvement of mitochondrial function; and b) the involvement of Ang-II-AT1 receptor interaction as a relevant step for mitochondrial dysfunction.

5. CALORIC RESTRICTION, AGING, OXIDATIVE STRESS, MITOCHONDRIA, AND PEROXISOME PROLIFERATOR ACTIVATED RECEPTORS

Caloric restriction, without malnutrition, is one of the most consistent interventions to reduce the rate of aging in different species. Caloric restriction increases mean and maximum lifespan, and retards both, the decay of physiological functions, and the appearance of diseases associated with aging, such as hypertension, diabetes, nephropathy, cardiovascular disease, and cancer (120-124).

Reduction in metabolic rate represents one explanation for the beneficial effects of caloric restriction observed in different animal species. This reduction leads to lower O₂ consumption, ROS generation (125), and oxidatively damaged proteins (126-128), lipids (129-131), and DNA (32, 33, 132, 133) than do *ad libitum* fed animals. With regard to its effects on mitochondria, long-term caloric restriction lowers the rate of mitochondrial H₂O₂ production and decreases the levels of mitochondrial DNA oxidative damage in rat liver, heart, skeletal muscle and brain (32, 33, 133, 134). In addition, caloric restriction was

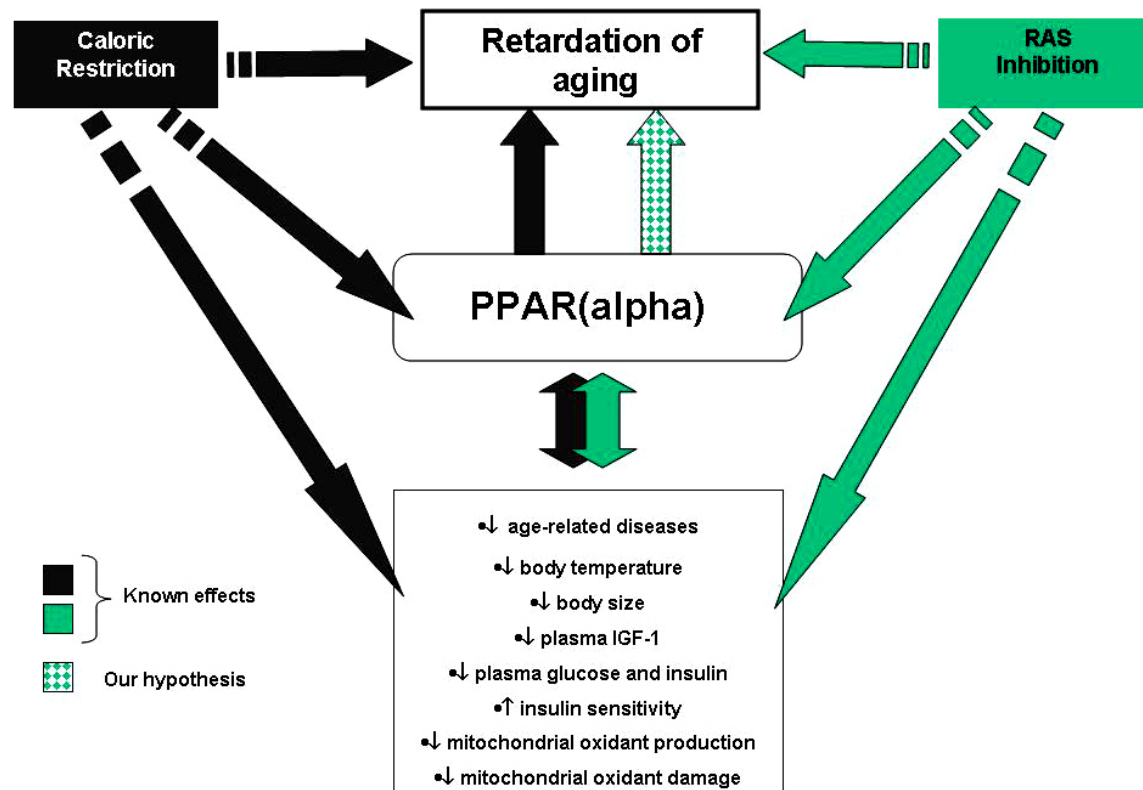


Figure 2. Scheme of the physiological actions elicited by caloric restriction and RAS inhibition and their impact on aging, having peroxisome proliferator activated receptors (PPARs) as central players. Our hypothesis of PPAR participation in the RAS-mediated retardation of aging is developed based on the converging effects of caloric restriction and RAS inhibition, and on the effects of RAS inhibition on mitochondrial function and PPAR modulation.

shown to increase the expression of UCP-2 in mice and in humans (135, 136), which may explain the effects of dietary manipulation on the reduction of mitochondrial H₂O₂ production. Analogous antioxidant and protective effects were observed in different animal species as well as in different tissues and cells (31). These observations emphasize the relevance of mitochondria and oxidative stress in the aging process.

Recent evidence suggests that PPARs may play an important role in the delay of aging caused by dietary restriction (54). In this context, PPAR nuclear protein, mRNA level, and DNA binding activity were shown to decrease with age, whereas caloric restriction blunted these reductions (52). Alternatively, using microarray technology, PPAR target genes were shown to be upregulated early and intensely in the livers of calorie-restricted mice (137).

6. RENIN-ANGIOTENSIN SYSTEM INHIBITION AND CALORIC RESTRICTION, TWO STRATEGIES CONVERGING AT BIOLOGICAL EFFECTS THAT INCLUDE PEROXISOME PROLIFERATOR ACTIVATED RECEPTORS

Considering that PPARs seem to play a major role in the age-retarding effects of caloric restriction, and

that RAS inhibition has been shown to retard aging and to upregulate PPARs, we hypothesize that the converging effects displayed by both interventions are mediated by the maintenance of adequate levels of PPAR protein/activity (Figure 2). This concurrence is supported by a number of physiological and pathological conditions that are affected in a similar manner when RAS inhibition or caloric restriction, are studied in both humans and animal models. The converging effects include: i) retarding the manifestations of hypertension (121), diabetes (123, 138), nephropathy (120, 139), cardiovascular disease (112, 122), and cancer (124, 140, 141); ii) increasing body temperature (142, 143) and loss of body weight (144, 145); iii) lowering of insulin-like growth factor-I (IGF-1) plasma levels (146-148); iv) reduction of plasma glucose and insulin levels in hypertensive patients (149), and rats (143, 150-152); v) improvement of insulin sensitivity in hypertensive patients (153-156), which is in agreement with several lines of evidence that point to a role for Ang-II in the development of insulin resistance (153, 157-159); vi) diminution of protein oxidation (126-128), lipid oxidation (129-131), and DNA oxidation (32, 33, 132, 133); vii) diminution of the rate of mitochondrial H₂O₂ production in concurrence with a decrease of mitochondrial DNA oxidative damage in rat liver, heart, skeletal muscle, and brain (32, 33, 133, 134) and increased expression of UCP-2 in mice and in humans (8, 135, 136).

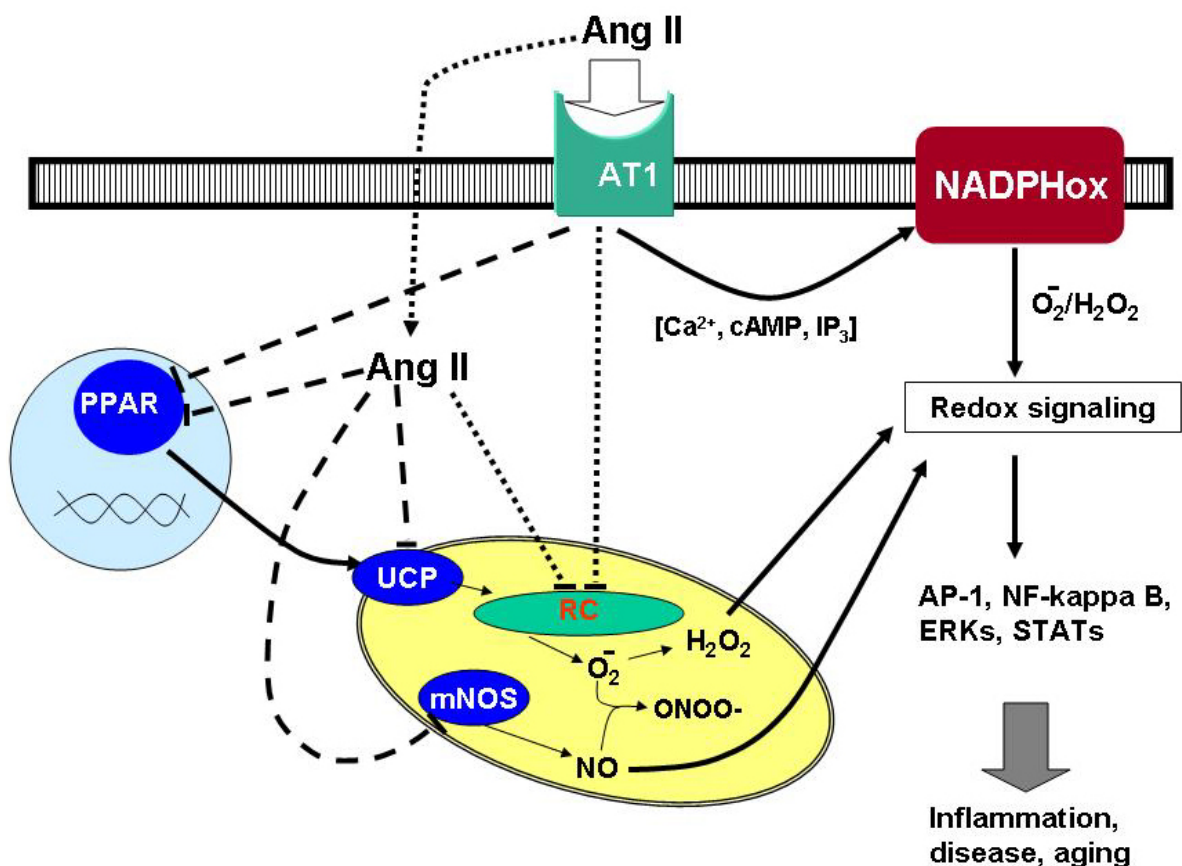


Figure 3. Scheme of the molecular pathways relating Ang-II with aging, having PPARs and mitochondria as central players. Solid arrows indicate accepted molecular pathways; dashed lines, pathways with experimental support; and dotted lines, hypothetical pathways. The mitochondrion is shown in yellow and the nucleus in light blue. Ang-II, angiotensin II; PPAR, peroxisome proliferator activated receptor; UCP, uncoupling proteins; mtNOS, mitochondrial nitric oxide synthase; RC, respiratory chain; AT1, angiotensin II-receptor 1; NADPHox, NADPH oxidase; TF, transcription factors. For the sake of clarity oxidant damage by NO, O₂⁻, H₂O₂, and ONOO⁻, as well as other agents in addition to Ca²⁺, cAMP, and IP₃, that are generated downstream of AT1 activation are not described in the scheme.

Regarding the effects of RAS inhibition and caloric restriction on weight loss, we must add that a positive correlation between plasma angiotensinogen levels and body mass index was reported in humans (160). However, the fact that Ang-II can produce anorexia (145) may afford an alternative interpretation for the Ang-II-mediated weight loss.

7. CONCLUSIONS AND PERSPECTIVES

By integrating current knowledge in the areas of mitochondrial metabolism, RAS, caloric restriction, and aging, we have developed a hypothesis in which PPARs serve an integrative role as central players. Figure 3 illustrates the potential molecular mechanisms underlying this hypothesis. The proposed mechanisms are based on the known activation of the AT1 receptor by Ang-II that leads to changes in Ca²⁺ homeostasis, and cAMP- and inositol triphosphate-mediated events, and the subsequent activation of NADPH-oxidase which, by generating O₂⁻, activates redox-sensitive signaling pathways. The changes driven by the activation of the AT1 receptor could also lead

to the uncoupling of mitochondrial respiration with the subsequent increase in O₂⁻ generation, and/or to the downregulation of PPARs activity, which could result in UCP deactivation and increased O₂⁻ generation. If Ang-II or any of its metabolites could reach the cytosol and the mitochondria they might interact with: a) PPARs or UCPs, leading to their inactivation; b) mitochondrial components, resulting in an increased production of O₂⁻ and a decreased of NO bioavailability; and/or c) NOS leading to a decreased NO generation.

The present hypothesis relating PPARs, mitochondria, and the RAS, provides a possible explanation for the molecular events that could be involved in the well documented effects ascribed to RAS inhibition in delaying the onset of age-associated pathologies.

8. REFERENCES

- Gerstein H C, J. Pogue, J. F. Mann, E. Lonn, G. R. Dagenais, M. McQueen & S. Yusuf: The relationship between dysglycaemia and cardiovascular and renal risk in

Mitochondria, renin-angiotensin system and aging

- diabetic and non-diabetic participants in the HOPE study: a prospective epidemiological analysis. *Diabetologia* 48, 1749-1755 (2005)
2. Poulter N R, H. Wedel, B. Dahlof, P. S. Sever, D. G. Beevers, M. Caulfield, S. E. Kjeldsen, A. Kristinsson, G. T. McInnes, J. Mehlsen, M. Nieminen, E. O'Brien, J. Ostergren & S. Pocock: Role of blood pressure and other variables in the differential cardiovascular event rates noted in the Anglo-Scandinavian Cardiac Outcomes Trial-Blood Pressure Lowering Arm (ASCOT-BPLA). *Lancet* 366, 907-913 (2005)
 3. Williams B: Recent hypertension trials: implications and controversies. *J Am Coll Cardiol* 45, 813-827 (2005)
 4. Yusuf S, P. Sleight, J. Pogue, J. Bosch, R. Davies & G. Dagenais: Effects of an angiotensin-converting-enzyme inhibitor, ramipril, on cardiovascular events in high-risk patients. The Heart Outcomes Prevention Evaluation Study Investigators. *N Engl J Med* 342, 145-153 (2000)
 5. de Cavanagh E M V, C. G. Fraga, L. Ferder & F. Inserra: Enalapril and captopril enhance antioxidant defenses in mouse tissues. *Am J Physiol* 272, R514-518 (1997)
 6. de Cavanagh E M V, F. Inserra, L. Ferder, L. Romano, L. Ercole & C. G. Fraga: Superoxide dismutase and glutathione peroxidase activities are increased by enalapril and captopril in mouse liver. *FEBS Lett* 361, 22-24 (1995)
 7. de Cavanagh E M V, F. Inserra, J. Toblli, I. Stella, C. G. Fraga & L. Ferder: Enalapril attenuates oxidative stress in diabetic rats. *Hypertension* 38, 1130-1136 (2001)
 8. de Cavanagh E M V, B. Piotrkowski, N. Basso, I. Stella, F. Inserra, L. Ferder & C. G. Fraga: Enalapril and losartan attenuate mitochondrial dysfunction in aged rats. *Faseb J* 17, 1096-1098 (2003)
 9. Beckman K B & B. N. Ames: The free radical theory of aging matures. *Physiol Rev* 78, 547-581 (1998)
 10. Chance B, H. Sies & A. Boveris: Hydroperoxide metabolism in mammalian organs. *Physiol Rev* 59, 527-605 (1979)
 11. DiMauro S & E. A. Schon: Mitochondrial respiratory-chain diseases. *N Engl J Med* 348, 2656-2668 (2003)
 12. Miquel J: An update on the oxygen stress-mitochondrial mutation theory of aging: genetic and evolutionary implications. *Exp Gerontol* 33, 113-126 (1998)
 13. de Cavanagh E M V, B. Piotrkowski & C. G. Fraga: Concerted action of the renin-angiotensin system, mitochondria, and antioxidant defenses in aging. *Mol Aspects Med* 25, 27-36 (2004)
 14. Long D A, W. Mu, K. L. Price & R. J. Johnson: Blood vessels and the aging kidney. *Nephron Exp Nephrol* 101, e95-99 (2005)
 15. Valdez L B, T. Zaobornyj, S. Alvarez, J. Bustamante, L. E. Costa & A. Boveris: Heart mitochondrial nitric oxide synthase. Effects of hypoxia and aging. *Mol Aspects Med* 25, 49-59 (2004)
 16. Beckman J S, T. W. Beckman, J. Chen, P. A. Marshall & B. A. Freeman: Apparent hydroxyl radical production by peroxynitrite: implications for endothelial injury from nitric oxide and superoxide. *Proc Natl Acad Sci U S A* 87, 1620-1624 (1990)
 17. Castro L & B. A. Freeman: Reactive oxygen species in human health and disease. *Nutrition* 17, 161, 163-165 (2001)
 18. Azzi A, K. J. Davies & F. Kelly: Free radical biology - terminology and critical thinking. *FEBS Lett* 558, 3-6 (2004)
 19. Sies H: Oxidative stress: from basic research to clinical application. *Am J Med* 91, 31S-38S (1991)
 20. Nicholls D G: Mitochondria and calcium signaling. *Cell Calcium* 38, 311-317 (2005)
 21. Thomas D D, X. Liu, S. P. Kantrow & J. R. Lancaster, Jr.: The biological lifetime of nitric oxide: implications for the perivascular dynamics of NO and O₂. *Proc Natl Acad Sci U S A* 98, 355-360 (2001)
 22. Brookes P S, E. P. Salinas, K. Darley-USmar, J. P. Eiserich, B. A. Freeman, V. M. Darley-USmar & P. G. Anderson: Concentration-dependent effects of nitric oxide on mitochondrial permeability transition and cytochrome c release. *J Biol Chem* 275, 20474-20479 (2000)
 23. Cadenas E: Mitochondrial free radical production and cell signaling. *Mol Aspects Med* 25, 17-26 (2004)
 24. Boveris A & E. Cadenas: Mitochondrial production of superoxide anions and its relationship to the antimycin insensitive respiration. *FEBS Lett* 54, 311-314 (1975)
 25. Turrens J F & A. Boveris: Generation of superoxide anion by the NADH dehydrogenase of bovine heart mitochondria. *Biochem J* 191, 421-427 (1980)
 26. Ignarro L J: Biosynthesis and metabolism of endothelium-derived nitric oxide. *Annu Rev Pharmacol Toxicol* 30, 535-560 (1990)
 27. Ghafourifar P & C. Richter: Nitric oxide synthase activity in mitochondria. *FEBS Lett* 418, 291-296 (1997)
 28. Giulivi C, J. J. Poderoso & A. Boveris: Production of nitric oxide by mitochondria. *J Biol Chem* 273, 11038-11043 (1998)

Mitochondria, renin-angiotensin system and aging

29. Cadenas E & K. J. Davies: Mitochondrial free radical generation, oxidative stress, and aging. *Free Radic Biol Med* 29, 222-230 (2000)
30. Gadaleta M N, A. Cormio, V. Pesce, A. M. Lezza & P. Cantatore: Aging and mitochondria. *Biochimie* 80, 863-870 (1998)
31. Gredilla R & G. Barja: Minireview: the role of oxidative stress in relation to caloric restriction and longevity. *Endocrinology* 146, 3713-3717 (2005)
32. Gredilla R, A. Sanz, M. Lopez-Torres & G. Barja: Caloric restriction decreases mitochondrial free radical generation at complex I and lowers oxidative damage to mitochondrial DNA in the rat heart. *Faseb J* 15, 1589-1591 (2001)
33. Lopez-Torres M, R. Gredilla, A. Sanz & G. Barja: Influence of aging and long-term caloric restriction on oxygen radical generation and oxidative DNA damage in rat liver mitochondria. *Free Radic Biol Med* 32, 882-889 (2002)
34. Brookes P S, A. L. Levonen, S. Shiva, P. Sarti & V. M. Darley-Usmar: Mitochondria: regulators of signal transduction by reactive oxygen and nitrogen species. *Free Radic Biol Med* 33, 755-764 (2002)
35. Darley-Usmar V: The powerhouse takes control of the cell; the role of mitochondria in signal transduction. *Free Radic Biol Med* 37, 753-754 (2004)
36. Berger, J P T. E. Akiyama & P. T. Meinke: PPARs: therapeutic targets for metabolic disease. *Trends Pharmacol Sci* 26, 244-251 (2005)
37. Schoonjans K, B. Staels & J. Auwerx: The peroxisome proliferator activated receptors (PPARs) and their effects on lipid metabolism and adipocyte differentiation. *Biochim Biophys Acta* 1302, 93-109 (1996)
38. Scarpulla R C: Transcriptional activators and coactivators in the nuclear control of mitochondrial function in mammalian cells. *Gene* 286, 81-89 (2002)
39. Reddy J K & T. Hashimoto: Peroxisomal beta-oxidation and peroxisome proliferator-activated receptor alpha: an adaptive metabolic system. *Annu Rev Nutr* 21, 193-230 (2001)
40. Kelly L J, P. P. Vicario, G. M. Thompson, M. R. Candelore, T. W. Doebber, J. Ventre, M. S. Wu, R. Meurer, M. J. Forrest, M. W. Conner, M. A. Cascieri & D. E. Moller: Peroxisome proliferator-activated receptors gamma and alpha mediate in vivo regulation of uncoupling protein (UCP-1, UCP-2, UCP-3) gene expression. *Endocrinology* 139, 4920-4927 (1998)
41. Nakatani T, N. Tsuboyama-Kasaoka, M. Takahashi, S. Miura & O. Ezaki: Mechanism for peroxisome proliferator-activated receptor-alpha activator-induced up-regulation of UCP2 mRNA in rodent hepatocytes. *J Biol Chem* 277, 9562-9569 (2002)
42. Takahashi M, N. Tsuboyama-Kasaoka, T. Nakatani, M. Ishii, S. Tsutsumi, H. Aburatani & O. Ezaki: Fish oil feeding alters liver gene expressions to defend against PPARalpha activation and ROS production. *Am J Physiol Gastrointest Liver Physiol* 282, G338-348 (2002)
43. Schiffrin E L, F. Amiri, K. Benkirane, M. Iglarz & Q. N. Diep: Peroxisome proliferator-activated receptors: vascular and cardiac effects in hypertension. *Hypertension* 42, 664-668 (2003)
44. Tanaka T, J. Yamamoto, S. Iwasaki, H. Asaba, H. Hamura, Y. Ikeda, M. Watanabe, K. Magoori, R. X. Ioka, K. Tachibana, Y. Watanabe, Y. Uchiyama, K. Sumi, H. Iguchi, S. Ito, T. Doi, T. Hamakubo, M. Naito, J. Auwerx, M. Yanagisawa, T. Kodama & J. Sakai: Activation of peroxisome proliferator-activated receptor delta induces fatty acid beta-oxidation in skeletal muscle and attenuates metabolic syndrome. *Proc Natl Acad Sci U S A* 100, 15924-15929 (2003)
45. Harman D: Aging: a theory based on free radical and radiation chemistry. *J Gerontol* 11, 298-300 (1956)
46. Schriener S E, N. J. Linford, G. M. Martin, P. Treuting, C. E. Ogburn, M. Emond, P. E. Coskun, W. Ladiges, N. Wolf, H. Van Remmen, D. C. Wallace & P. S. Rabinovitch: Extension of murine life span by overexpression of catalase targeted to mitochondria. *Science* 308, 1909-1911 (2005)
47. Celi F S & A. R. Shuldiner: The role of peroxisome proliferator-activated receptor gamma in diabetes and obesity. *Curr Diab Rep* 2, 179-185 (2002)
48. Chawla A, J. J. Repa, R. M. Evans & D. J. Mangelsdorf: Nuclear receptors and lipid physiology: opening the X-files. *Science* 294, 1866-1870 (2001)
49. Evans R M, G. D. Barish & Y. X. Wang: PPARs and the complex journey to obesity. *Nat Med* 10, 355-361 (2004)
50. Kintscher U, C. J. Lyon & R. E. Law: Angiotensin II, PPAR-gamma and atherosclerosis. *Front Biosci* 9, 359-369 (2004)
51. Schiffrin E L: Peroxisome proliferator-activated receptors and cardiovascular remodeling. *Am J Physiol Heart Circ Physiol* 288, H1037-1043 (2005)
52. Sung B, S. Park, B. P. Yu & H. Y. Chung: Modulation of PPAR in aging, inflammation, and calorie restriction. *J Gerontol A Biol Sci Med Sci* 59, 997-1006 (2004)
53. Cabrero A, M. Alegret, R. M. Sanchez, T. Adzet, J. C. Laguna & M. V. Carrera: Increased reactive oxygen species production down-regulates peroxisome proliferator-

Mitochondria, renin-angiotensin system and aging

- activated alpha pathway in C2C12 skeletal muscle cells. *J Biol Chem* 277, 10100-10107 (2002)
54. Pardee K, J. Reinking & H. Krause: Nuclear hormone receptors, metabolism, and aging: what goes around comes around. Transcription factors link lipid metabolism and aging-related processes. *Sci Aging Knowledge Environ* 2004, re8 (2004)
55. Poynter M E & R. A. Daynes: Peroxisome proliferator-activated receptor alpha activation modulates cellular redox status, represses nuclear factor-kappaB signaling, and reduces inflammatory cytokine production in aging. *J Biol Chem* 273, 32833-32841 (1998)
56. Aubert J, O. Champigny, P. Saint-Marc, R. Negrel, S. Collins, D. Ricquier & G. Ailhaud: Up-regulation of UCP-2 gene expression by PPAR agonists in preadipose and adipose cells. *Biochem Biophys Res Commun* 238, 606-611 (1997)
57. Camirand A, V. Marie, R. Rabelo & J. E. Silva: Thiazolidinediones stimulate uncoupling protein-2 expression in cell lines representing white and brown adipose tissues and skeletal muscle. *Endocrinology* 139, 428-431 (1998)
58. Fleury C, M. Neverova, S. Collins, S. Raimbault, O. Champigny, C. Levi-Meyrueis, F. Bouillaud, M. F. Seldin, R. S. Surwit, D. Ricquier & C. H. Warden: Uncoupling protein-2: a novel gene linked to obesity and hyperinsulinemia. *Nat Genet* 15, 269-272 (1997)
59. Gimeno R E, M. Dembski, X. Weng, N. Deng, A. W. Shyjan, C. J. Gimeno, F. Iris, S. J. Ellis, E. A. Woolf & L. A. Tartaglia: Cloning and characterization of an uncoupling protein homolog: a potential molecular mediator of human thermogenesis. *Diabetes* 46, 900-906 (1997)
60. Negre-Salvayre A, C. Hirtz, G. Carrera, R. Cazenave, M. Trolly, R. Salvayre, L. Penicaud & L. Casteilla: A role for uncoupling protein-2 as a regulator of mitochondrial hydrogen peroxide generation. *Faseb J* 11, 809-815 (1997)
61. Cho D H, Y. J. Choi, S. A. Jo & I. Jo: Nitric oxide production and regulation of endothelial nitric-oxide synthase phosphorylation by prolonged treatment with troglitazone: evidence for involvement of peroxisome proliferator-activated receptor (PPAR) gamma-dependent and PPARgamma-independent signaling pathways. *J Biol Chem* 279, 2499-2506 (2004)
62. Newaz M A, K. Ranganna & A. O. Oyekan: Relationship between PPARalpha activation and NO on proximal tubular Na⁺ transport in the rat. *BMC Pharmacol* 4, 1 (2004)
63. Newaz M, A. Blanton, P. Fidelis & A. Oyekan: NAD(P)H oxidase/nitric oxide interactions in peroxisome proliferator activated receptor (PPAR)alpha-mediated cardiovascular effects. *Mutat Res* 579, 163-171 (2005)
64. Lee M A, M. Bohm, M. Paul & D. Ganten: Tissue renin-angiotensin systems. Their role in cardiovascular disease. *Circulation* 87, IV7-13 (1993)
65. Skeggs L T, K. E. Lentz, J. R. Kahn & H. Hochstrasser: Kinetics of the reaction of renin with nine synthetic peptide substrates. *J Exp Med* 128, 13-34 (1968)
66. Brewster U C, J. F. Setaro & M. A. Perazella: The renin-angiotensin-aldosterone system: cardiorenal effects and implications for renal and cardiovascular disease states. *Am J Med Sci* 326, 15-24 (2003)
67. Omura T, S. Kim, K. Takeuchi, H. Iwao & T. Takeda: Transforming growth factor beta 1 and extracellular matrix gene expression in isoprenaline induced cardiac hypertrophy: effects of inhibition of the renin-angiotensin system. *Cardiovasc Res* 28, 1835-1842 (1994)
68. Tummala P E, X. L. Chen, C. L. Sundell, J. B. Laursen, C. P. Hammes, R. W. Alexander, D. G. Harrison & R. M. Medford: Angiotensin II induces vascular cell adhesion molecule-1 expression in rat vasculature: A potential link between the renin-angiotensin system and atherosclerosis. *Circulation* 100, 1223-1229 (1999)
69. Touyz R M, F. Tabet & E. L. Schiffrin: Redox-dependent signalling by angiotensin II and vascular remodelling in hypertension. *Clin Exp Pharmacol Physiol* 30, 860-866 (2003)
70. Griendling K K & M. Ushio-Fukai: Reactive oxygen species as mediators of angiotensin II signaling. *Regul Pept* 91, 21-27 (2000)
71. Kimura S., G. X. Zhang, A. Nishiyama, T. Shokoji, L. Yao, Y. Y. Fan, M. Rahman, T. Suzuki, H. Maeta & Y. Abe: Role of NAD(P)H oxidase- and mitochondria-derived reactive oxygen species in cardioprotection of ischemic reperfusion injury by angiotensin II. *Hypertension* 45, 860-866 (2005)
72. Mollnau H, M. Wendt, K. Szocs, B. Lassegue, E. Schulz, M. Oelze, H. Li, M. Bodenschatz, M. August, A. L. Kleschyov, N. Tsilimingas, U. Walter, U. Forstermann, T. Meinertz, K. Griendling & T. Munzel: Effects of angiotensin II infusion on the expression and function of NAD(P)H oxidase and components of nitric oxide/cGMP signaling. *Circ Res* 90, E58-65 (2002)
73. Touyz R M: Activated oxygen metabolites: do they really play a role in angiotensin II-regulated vascular tone? *J Hypertens* 21, 2235-2238 (2003)
74. Henriksen E J & S. Jacob: Modulation of metabolic control by angiotensin converting enzyme (ACE) inhibition. *J Cell Physiol* 196, 171-179 (2003)
75. Rincon-Choles H, B. S. Kasinath, Y. Gorin & H. E. Abboud: Angiotensin II and growth factors in the pathogenesis of diabetic nephropathy. *Kidney Int Suppl* 8-11 (2002)

Mitochondria, renin-angiotensin system and aging

76. Touyz R M & E. L. Schiffrin: Signal transduction mechanisms mediating the physiological and pathophysiological actions of angiotensin II in vascular smooth muscle cells. *Pharmacol Rev* 52, 639-672 (2000)
77. Baylis C, K. Engels, A. Hymel & L. G. Navar: Plasma renin activity and metabolic clearance rate of angiotensin II in the unstressed aging rat. *Mech Ageing Dev* 97, 163-172 (1997)
78. Harada K, M. Ohmori & A. Fujimura: Vasoconstricting effect of angiotensin II in human hand veins: influence of aging, diabetes mellitus and hypertension. *Hypertens Res* 25, 683-688 (2002)
79. Thompson M M, T. T. Oyama, F. J. Kelly, T. M. Kennefick & S. Anderson: Activity and responsiveness of the renin-angiotensin system in the aging rat. *Am J Physiol Regul Integr Comp Physiol* 279, R1787-1794 (2000)
80. Wang M, G. Takagi, K. Asai, R. G. Resuello, F. F. Natividad, D. E. Vatner, S. F. Vatner & E. G. Lakatta: Aging increases aortic MMP-2 activity and angiotensin II in nonhuman primates. *Hypertension* 41, 1308-1316 (2003)
81. Ruiz-Ortega M, M. Ruperez, V. Esteban & J. Egido: Molecular mechanisms of angiotensin II-induced vascular injury. *Curr Hypertens Rep* 5, 73-79 (2003)
82. Puri P L, M. L. Avantiaggiati, V. L. Burgio, P. Chirillo, D. Collepardo, G. Natoli, C. Balsano & M. Levrero: Reactive oxygen intermediates mediate angiotensin II-induced c-Jun/c-Fos heterodimer DNA binding activity and proliferative hypertrophic responses in myogenic cells. *J Biol Chem* 270, 22129-22134 (1995)
83. Shih N L, T. H. Cheng, S. H. Loh, P. Y. Cheng, D. L. Wang, Y. S. Chen, S. H. Liu, C. C. Liew & J. J. Chen: Reactive oxygen species modulate angiotensin II-induced beta-myosin heavy chain gene expression via Ras/Raf/extracellular signal-regulated kinase pathway in neonatal rat cardiomyocytes. *Biochem Biophys Res Commun* 283, 143-148 (2001)
84. Larkin J E, B. C. Frank, R. M. Gaspard, I. Duka, H. Gavras & J. Quackenbush: Cardiac transcriptional response to acute and chronic angiotensin II treatments. *Physiol Genomics* 18, 152-166 (2004)
85. Casademont J & O. Miro: Electron transport chain defects in heart failure. *Heart Fail Rev* 7, 131-139 (2002)
86. Sanbe A, K. Tanonaka, R. Kobayasi & S. Takeo: Effects of long-term therapy with ACE inhibitors, captopril, enalapril and trandolapril, on myocardial energy metabolism in rats with heart failure following myocardial infarction. *J Mol Cell Cardiol* 27, 2209-2222 (1995)
87. Sorescu D & K. K. Griendling: Reactive oxygen species, mitochondria, and NAD(P)H oxidases in the development and progression of heart failure. *Congest Heart Fail* 8, 132-140 (2002)
88. Kimura S, G. X. Zhang, A. Nishiyama, T. Shokoji, L. Yao, Y. Y. Fan, M. Rahman & Y. Abe: Mitochondria-derived reactive oxygen species and vascular MAP kinases: comparison of angiotensin II and diazoxide. *Hypertension* 45, 438-444 (2005)
89. Robertson A L, Jr. & P. A. Khairallah: Angiotensin II: rapid localization in nuclei of smooth and cardiac muscle. *Science* 172, 1138-1139 (1971)
90. Sirett N E, A. S. McLean, J. J. Bray & J. I. Hubbard: Distribution of angiotensin II receptors in rat brain. *Brain Res* 122, 299-312 (1977)
91. Eggena P, J. H. Zhu, K. Clegg & J. D. Barrett: Nuclear angiotensin receptors induce transcription of renin and angiotensinogen mRNA. *Hypertension* 22, 496-501 (1993)
92. Eggena P, J. H. Zhu, S. Serevinyayut, M. Giordani, K. Clegg, P. C. Andersen, P. Hyun & J. D. Barrett: Hepatic angiotensin II nuclear receptors and transcription of growth-related factors. *J Hypertens* 14, 961-968 (1996)
93. Booz G W, K. M. Conrad, A. L. Hess, H. A. Singer & K. M. Baker: Angiotensin-II-binding sites on hepatocyte nuclei. *Endocrinology* 130, 3641-3649 (1992)
94. Peters J, B. Kranzlin, S. Schaeffer, J. Zimmer, S. Resch, S. Bachmann, N. Gretz & E. Hackenthal: Presence of renin within intramitochondrial dense bodies of the rat adrenal cortex. *Am J Physiol* 271, E439-450 (1996)
95. Kudoh S, I. Komuro, T. Mizuno, T. Yamazaki, Y. Zou, I. Shiojima, N. Takekoshi & Y. Yazaki: Angiotensin II stimulates c-Jun NH2-terminal kinase in cultured cardiac myocytes of neonatal rats. *Circ Res* 80, 139-146 (1997)
96. Xia Y, L. M. Buja & J. B. McMillin: Activation of the cytochrome c gene by electrical stimulation in neonatal rat cardiac myocytes. Role of NRF-1 and c-Jun. *J Biol Chem* 273, 12593-12598 (1998)
97. Leary S C, D. Michaud, C. N. Lyons, T. M. Hale, T. L. Bushfield, M. A. Adams & C. D. Moyes: Bioenergetic remodeling of heart during treatment of spontaneously hypertensive rats with enalapril. *Am J Physiol Heart Circ Physiol* 283, H540-548 (2002)
98. Boveris A, G. D'Amico, S. Lores-Arnaiz & L. E. Costa: Enalapril increases mitochondrial nitric oxide synthase activity in heart and liver. *Antioxid Redox Signal* 5, 691-697 (2003)
99. Costa L E, P. La-Padula, S. Lores-Arnaiz, G. D'Amico, A. Boveris, M. L. Kurnjek & N. Basso: Long-term angiotensin II inhibition increases mitochondrial nitric oxide synthase and not antioxidant enzyme activities in rat heart. *J Hypertens* 20, 2487-2494 (2002)
100. Chen G, L. X. Lin, W. T. Zhuang, J. Yao, H. B. Huang, J. X. Liang, F. L. Zhang, J. P. Wen, L. T. Li, M. Lin & Q. M. Lin: [Effects of captopril on myocardial tissue

Mitochondria, renin-angiotensin system and aging

- energy metabolism and inflammation in rats with diabetic cardiomyopathy]. *Di Yi Jun Yi Da Xue Xue Bao* 24, 827-828, 831 (2004)
101. Tham D M, B. Martin-McNulty, Y. X. Wang, D. W. Wilson, R. Vergona, M. E. Sullivan, W. Dole & J. C. Rutledge: Angiotensin II is associated with activation of NF-kappaB-mediated genes and downregulation of PPARs. *Physiol Genomics* 11, 21-30 (2002)
102. Benkirane K, F. Amiri, Q. N. Diep, M. El Mabrouk & E. L. Schiffrin: PPAR-gamma inhibits ANG II-induced cell growth via SHIP2 and 4E-BP1. *Am J Physiol Heart Circ Physiol* 290, H390-397 (2006)
103. Muller D N, J. Theuer, E. Shagdarsuren, E. Kaergel, H. Honeck, J. K. Park, M. Markovic, E. Barbosa-Sicard, R. Dechend, M. Wellner, T. Kirsch, A. Fiebeler, M. Rothe, H. Haller, F. C. Luft & W. H. Schunck: A peroxisome proliferator-activated receptor-alpha activator induces renal CYP2C23 activity and protects from angiotensin II-induced renal injury. *Am J Pathol* 164, 521-532 (2004)
104. da Cunha V, D. M. Tham, B. Martin-McNulty, G. Deng, J. J. Ho, D. W. Wilson, J. C. Rutledge, R. Vergona, M. E. Sullivan & Y. X. Wang: Enalapril attenuates angiotensin II-induced atherosclerosis and vascular inflammation. *Atherosclerosis* 178, 9-17 (2005)
105. Clasen R, M. Schupp, A. Foryst-Ludwig, C. Sprang, M. Clemenz, M. Krikov, C. Thone-Reineke, T. Unger & U. Kintscher: PPARgamma-activating angiotensin type-1 receptor blockers induce adiponectin. *Hypertension* 46, 137-143 (2005)
106. Tuck M L: Angiotensin-receptor blocking agents and the peroxisome proliferator-activated receptor-gamma system. *Curr Hypertens Rep* 7, 240-243 (2005)
107. Schupp M, L. D. Lee, N. Frost, S. Umbreen, B. Schmidt, T. Unger & U. Kintscher: Regulation of Peroxisome Proliferator-Activated Receptor {gamma} Activity by Losartan Metabolites. *Hypertension* (2005)
108. Heudes D, O. Michel, J. Chevalier, E. Scalbert, E. Ezan, J. Bariety, A. Zimmerman & B. Corman: Effect of chronic ANG I-converting enzyme inhibition on aging processes. I. Kidney structure and function. *Am J Physiol* 266, R1038-1051 (1994)
109. Ferder L, F. Inserra, L. Romano, L. Ercole & V. Pszeny: Decreased glomerulosclerosis in aging by angiotensin-converting enzyme inhibitors. *J Am Soc Nephrol* 5, 1147-1152 (1994)
110. Ma L J, S. Nakamura, J. S. Whitsitt, C. Marcantoni, J. M. Davidson & A. B. Fogo: Regression of sclerosis in aging by an angiotensin inhibition-induced decrease in PAI-1. *Kidney Int* 58, 2425-2436 (2000)
111. Ferder L F, F. Inserra & N. Basso: Advances in our understanding of aging: role of the renin-angiotensin system. *Curr Opin Pharmacol* 2, 189-194 (2002)
112. Bosch J, E. Lonn, J. Pogue, J. M. Arnold, G. R. Dagenais & S. Yusuf: Long-term effects of ramipril on cardiovascular events and on diabetes: results of the HOPE study extension. *Circulation* 112, 1339-1346 (2005)
113. Dahlof B, R. B. Devereux, S. Julius, S. E. Kjeldsen, G. Beevers, U. de Faire, F. Fyhrquist, T. Hedner, H. Ibsen, K. Kristianson, O. Lederballe-Pedersen, L. H. Lindholm, M. S. Nieminen, P. Omvik, S. Oparil & H. Wedel: Characteristics of 9194 patients with left ventricular hypertrophy: the LIFE study. Losartan Intervention For Endpoint Reduction in Hypertension. *Hypertension* 32, 989-997 (1998)
114. Pfeffer M A, E. Braunwald, L. A. Moye, L. Basta, E. J. Brown, Jr., T. E. Cuddy, B. R. Davis, E. M. Geltman, S. Goldman, G. C. Flaker & et al.: Effect of captopril on mortality and morbidity in patients with left ventricular dysfunction after myocardial infarction. Results of the survival and ventricular enlargement trial. The SAVE Investigators. *N Engl J Med* 327, 669-677 (1992)
115. Park J. B., H. D. Intengan & E. L. Schiffrin: Reduction of resistance artery stiffness by treatment with the AT(1)-receptor antagonist losartan in essential hypertension. *J Renin Angiotensin Aldosterone Syst* 1, 40-45 (2000)
116. Ramachandran A, A. L. Levonen, P. S. Brookes, E. Ceaser, S. Shiva, M. C. Barone & V. Darley-USmar: Mitochondria, nitric oxide, and cardiovascular dysfunction. *Free Radic Biol Med* 33, 1465-1474 (2002)
117. Schrauwen P & M. K. Hesselink: Oxidative capacity, lipotoxicity, and mitochondrial damage in type 2 diabetes. *Diabetes* 53, 1412-1417 (2004)
118. de Cavanagh E M V, J. E. Toblli, L. Ferder, B. Piotrkowski, I. Stella & F. Inserra: Renal Mitochondrial Dysfunction In Spontaneously Hypertensive Rats Is Attenuated By Losartan But Not By Amlodipine. *Am J Physiol Regul Integr Comp Physiol* (2006)
119. de Cavanagh E M V, B. Piotrkowski, J. Toblli, L. Ferder, F. Inserra & C. G. Fraga: Losartan ameliorates renal mitochondrial dysfunction in streptozotocin-induced diabetes and aging (abstract). *Free Rad Biol Med* 37 (suppl 1), s36 (2004)
120. Cefalu W T, Z. Q. Wang, A. D. Bell-Farrow, J. Collins, T. Morgan & J. D. Wagner: Caloric restriction and cardiovascular aging in cynomolgus monkeys (Macaca fascicularis): metabolic, physiologic, and atherosclerotic measures from a 4-year intervention trial. *J Gerontol A Biol Sci Med Sci* 59, 1007-1014 (2004)

Mitochondria, renin-angiotensin system and aging

121. Das M, I. Gabriely & N. Barzilai: Caloric restriction, body fat and ageing in experimental models. *Obes Rev* 5, 13-19 (2004)
122. Loupal G, A. Url, M. Skalicky & A. Viidik: Physical exercise retards the development of chronic nephropathy in the ageing rat as efficiently as food restriction does. *Gerontology* 51, 83-93 (2005)
123. Mattson M P, S. L. Chan & W. Duan: Modification of brain aging and neurodegenerative disorders by genes, diet, and behavior. *Physiol Rev* 82, 637-672 (2002)
124. Spindler S R: Rapid and reversible induction of the longevity, anticancer and genomic effects of caloric restriction. *Mech Ageing Dev* 126, 960-966 (2005)
125. Adelman, R, R. L. Saul & B. N. Ames: Oxidative damage to DNA: relation to species metabolic rate and life span. *Proc Natl Acad Sci U S A* 85, 2706-2708 (1988)
126. Forster M J, B. H. Sohal & R. S. Sohal: Reversible effects of long-term caloric restriction on protein oxidative damage. *J Gerontol A Biol Sci Med Sci* 55, B522-529 (2000)
127. Leeuwenburgh C, P. Wagner, J. O. Holloszy, R. S. Sohal & J. W. Heinecke: Caloric restriction attenuates dityrosine cross-linking of cardiac and skeletal muscle proteins in aging mice. *Arch Biochem Biophys* 346, 74-80 (1997)
128. Zainal T A, T. D. Oberley, D. B. Allison, L. I. Szweda & R. Weindruch: Caloric restriction of rhesus monkeys lowers oxidative damage in skeletal muscle. *Faseb J* 14, 1825-1836 (2000)
129. Lambert A J, M. Portero-Otin, R. Pamplona & B. J. Merry: Effect of ageing and caloric restriction on specific markers of protein oxidative damage and membrane peroxidizability in rat liver mitochondria. *Mech Ageing Dev* 125, 529-538 (2004)
130. Lee J, B. P. Yu & J. T. Herlihy: Modulation of cardiac mitochondrial membrane fluidity by age and calorie intake. *Free Radic Biol Med* 26, 260-265 (1999)
131. Pamplona R, M. Portero-Otin, J. Requena, R. Gredilla & G. Barja: Oxidative, glycoxidative and lipoxidative damage to rat heart mitochondrial proteins is lower after 4 months of caloric restriction than in age-matched controls. *Mech Ageing Dev* 123, 1437-1446 (2002)
132. Chung M H, H. Kasai, S. Nishimura & B. P. Yu: Protection of DNA damage by dietary restriction. *Free Radic Biol Med* 12, 523-525 (1992)
133. Drew B, S. Phaneuf, A. Dirks, C. Selman, R. Gredilla, A. Lezza, G. Barja & C. Leeuwenburgh: Effects of aging and caloric restriction on mitochondrial energy production in gastrocnemius muscle and heart. *Am J Physiol Regul Integr Comp Physiol* 284, R474-480 (2003)
134. Sanz A, P. Caro, J. Ibanez, J. Gomez, R. Gredilla & G. Barja: Dietary restriction at old age lowers mitochondrial oxygen radical production and leak at complex I and oxidative DNA damage in rat brain. *J Bioenerg Biomembr* 37, 83-90 (2005)
135. Millet L, H. Vidal, F. Andreelli, D. Larrouy, J. P. Riou, D. Ricquier, M. Laville & D. Langin: Increased uncoupling protein-2 and -3 mRNA expression during fasting in obese and lean humans. *J Clin Invest* 100, 2665-2670 (1997)
136. Xiao H, D. Massaro, G. D. Massaro & L. B. Clerch: Expression of lung uncoupling protein-2 mRNA is developmentally and by caloric intake. *Exp Biol Med (Maywood)* 229, 479-485 (2004)
137. Bauer M, A. C. Hamm, M. Bonaus, A. Jacob, J. Jaekel, H. Schorle, M. J. Pankratz & J. D. Katzenberger: Starvation response in mouse liver shows strong correlation with life-span-prolonging processes. *Physiol Genomics* 17, 230-244 (2004)
138. Lindholm L H, H. Ibsen, K. Borch-Johnsen, M. H. Olsen, K. Wachtell, B. Dahlöf, R. B. Devereux, G. Beevers, U. de Faire, F. Fyhrquist, S. Julius, S. E. Kjeldsen, K. Kristianson, O. Lederballe-Pedersen, M. S. Nieminen, P. Omvik, S. Oparil, H. Wedel, P. Aurup, J. M. Edelman & S. Snapinn: Risk of new-onset diabetes in the Losartan Intervention For Endpoint reduction in hypertension study. *J Hypertens* 20, 1879-1886 (2002)
139. Parving H H, H. Lehnert, J. Brochner-Mortensen, R. Gomis, S. Andersen & P. Arner: The effect of irbesartan on the development of diabetic nephropathy in patients with type 2 diabetes. *N Engl J Med* 345, 870-878 (2001)
140. Deshayes F & C. Nahmias: Angiotensin receptors: a new role in cancer? *Trends Endocrinol Metab* 16, 293-299 (2005)
141. Uemura H, N. Nakaigawa, H. Ishiguro & Y. Kubota: Antiproliferative efficacy of angiotensin II receptor blockers in prostate cancer. *Curr Cancer Drug Targets* 5, 307-323 (2005)
142. Cassis L A, D. E. Marshall, M. J. Fetting, B. Rosenbluth & R. A. Lodder: Mechanisms contributing to angiotensin II regulation of body weight. *Am J Physiol* 274, E867-876 (1998)
143. Roth G S, D. K. Ingram & M. A. Lane: Caloric restriction in primates and relevance to humans. *Ann N Y Acad Sci* 928, 305-315 (2001)
144. DeLany J P, B. C. Hansen, N. L. Bodkin, J. Hannah & G. A. Bray: Long-term calorie restriction reduces energy expenditure in aging monkeys. *J Gerontol A Biol Sci Med Sci* 54, B5-11; discussion B12-13 (1999)

Mitochondria, renin-angiotensin system and aging

145. Engeli S, R. Negrel & A. M. Sharma: Physiology and pathophysiology of the adipose tissue renin-angiotensin system. *Hypertension* 35, 1270-1277 (2000)
146. Jalil J E, R. Ebensperger, J. Melendez, E. Acevedo, M. Sapag-Hagar, F. Gonzalez-Jara, A. Galvez, V. Perez-Montes & S. Lavandero: Effects of antihypertensive treatment on cardiac IGF-1 during prevention of ventricular hypertrophy in the rat. *Life Sci* 64, 1603-1612 (1999)
147. Katic M & C. R. Kahn: The role of insulin and IGF-1 signaling in longevity. *Cell Mol Life Sci* 62, 320-343 (2005)
148. Wang A Y, A. W. Yu, C. W. Lam, L. M. Yu, P. K. Li, J. Goh & S. F. Lui: Effects of losartan or enalapril on hemoglobin, circulating erythropoietin, and insulin-like growth factor-1 in patients with and without posttransplant erythrocytosis. *Am J Kidney Dis* 39, 600-608 (2002)
149. Vitale C, G. Mercurio, C. Castiglioni, A. Cornoldi, A. Tulli, M. Fini, M. Volterrani & G. M. Rosano: Metabolic effect of telmisartan and losartan in hypertensive patients with metabolic syndrome. *Cardiovasc Diabetol* 4, 6 (2005)
150. Chen S, Y. Noguchi, T. Izumida, J. Tatebe & S. Katayama: A comparison of the hypotensive and hypoglycaemic actions of an angiotensin converting enzyme inhibitor, an AT1a antagonist and troglitazone. *J Hypertens* 14, 1325-1330 (1996)
151. Uresin Y, B. Erbas, M. Ozek, E. Ozkok & A. O. Gurol: Losartan may prevent the elevation of plasma glucose, corticosterone and catecholamine levels induced by chronic stress. *J Renin Angiotensin Aldosterone Syst* 5, 93-96 (2004)
152. Wu Y, J. P. Ouyang, Y. F. Zhou, K. Wu, D. H. Zhao & C. Y. Wen: Mechanism of improving effect of losartan on insulin sensitivity of non-insulin-dependent diabetes mellitus rats. *Sheng Li Xue Bao* 56, 539-549 (2004)
153. Coimbra C C, M. A. Garofalo, D. R. Foscolo, A. R. Xavier & R. H. Migliorini: Gluconeogenesis activation after intravenous angiotensin II in freely moving rats. *Peptides* 20, 823-827 (1999)
154. Furuhashi M, N. Ura, K. Higashiura, H. Murakami, M. Tanaka, N. Moniwa, D. Yoshida & K. Shimamoto: Blockade of the renin-angiotensin system increases adiponectin concentrations in patients with essential hypertension. *Hypertension* 42, 76-81 (2003)
155. Galletti F, P. Strazzullo, B. Capaldo, R. Carretta, F. Fabris, L. A. Ferrara, N. Glorioso, A. Semplicini & M. Mancini: Controlled study of the effect of angiotensin converting enzyme inhibition versus calcium-entry blockade on insulin sensitivity in overweight hypertensive patients: Trandolapril Italian Study (TRIS). *J Hypertens* 17, 439-445 (1999)
156. Park S Y, G. H. Choi, H. I. Choi, J. Ryu, C. Y. Jung & W. Lee: Calorie restriction improves whole-body glucose disposal and insulin resistance in association with the increased adipocyte-specific GLUT4 expression in Otsuka Long-Evans Tokushima fatty rats. *Arch Biochem Biophys* 436, 276-284 (2005)
157. Boschmann M, J. Ringel, S. Klaus & A. M. Sharma: Metabolic and hemodynamic response of adipose tissue to angiotensin II. *Obes Res* 9, 486-491 (2001)
158. Folli F, C. R. Kahn, H. Hansen, J. L. Bouchie & E. P. Feener: Angiotensin II inhibits insulin signaling in aortic smooth muscle cells at multiple levels. A potential role for serine phosphorylation in insulin/angiotensin II crosstalk. *J Clin Invest* 100, 2158-2169 (1997)
159. Velloso L A, F. Folli, X. J. Sun, M. F. White, M. J. Saad & C. R. Kahn: Cross-talk between the insulin and angiotensin signaling systems. *Proc Natl Acad Sci U S A* 93, 12490-12495 (1996)
160. Umemura S, N. Nyui, K. Tamura, K. Hibi, S. Yamaguchi, M. Nakamaru, T. Ishigami, M. Yabana, M. Kihara, S. Inoue & M. Ishii: Plasma angiotensinogen concentrations in obese patients. *Am J Hypertens* 10, 629-633 (1997)

Key Words: Angiotensin, Mitochondria, Respiratory Chain, Oxidative Damage, Review

Send correspondence to: Cesar G. Fraga, Department of Nutrition, University of California, One Shields Ave., Davis, CA 95616, USA, Tel: 530-754-667, Fax: 530-752-8966, E-mail: cgfraga@ucdavis.edu

<http://www.bioscience.org/current/vol12.htm>