

# **Online Research @ Cardiff**

This is an Open Access document downloaded from ORCA, Cardiff University's institutional repository: https://orca.cardiff.ac.uk/id/eprint/145106/

This is the author's version of a work that was submitted to / accepted for publication.

Citation for final published version:

Bandowe, Benjamin A. Musa, Lui, K.H., Jones, Timothy ORCID: https://orcid.org/0000-0002-4466-1260, BeruBe, Kelly ORCID: https://orcid.org/0000-0002-7471-7229, Adams, Rachel, Niu, Xinyi, Wei, Chong, Cao, Jun-Ji, Lee, S.C., Chuang, Hsiao-Chi and Ho, K.F. 2021. The chemical composition and toxicological effects of fine particulate matter (PM2.5) emitted from different cooking styles. Environmental Pollution 288, 117754. 10.1016/j.envpol.2021.117754 filefile

Publishers page: http://dx.doi.org/10.1016/j.envpol.2021.117754 <http://dx.doi.org/10.1016/j.envpol.2021.117754>

## Please note:

Changes made as a result of publishing processes such as copy-editing, formatting and page numbers may not be reflected in this version. For the definitive version of this publication, please refer to the published source. You are advised to consult the publisher's version if you wish to cite this paper.

This version is being made available in accordance with publisher policies.

See

http://orca.cf.ac.uk/policies.html for usage policies. Copyright and moral rights for publications made available in ORCA are retained by the copyright holders.

> information services gwasanaethau gwybodaeth

1	The chemical composition and toxicological effects of fine particulate matter (PM2.5)
2	emitted from different cooking styles
3	Benjamin A. Musa Bandowe <sup>1,2#</sup> , K. H. Lui <sup>3#</sup> , Timothy Jones <sup>4</sup> , Kelly BéruBé <sup>5</sup> , Rachel
4	Adams <sup>6</sup> , Xinyi Niu <sup>3</sup> , Chong Wei <sup>7</sup> , Jun-Ji Cao <sup>8,9</sup> , S. C. Lee <sup>10</sup> , Hsiao-Chi Chuang <sup>11,12</sup> , K. F.
5	Ho <sup>3*</sup>
6 7	<sup>1</sup> Institute of Geography University of Barn Hallerstrasse 12, 3012 Barn Switzerland
,	<sup>2</sup> Max Planck Institute for Chemistry, Multiphase Chemistry, Department, Hahn Meitner Weg, 1, 55128
0	Max Flanck Institute for Chemistry, Multiphase Chemistry Department, Hann-Mether-weg 1, 55128
9	Mainz, Germany
10	<sup>5</sup> The Jockey Club School of Public Health and Primary Care, The Chinese University of Hong Kong,
11	Hong Kong, China
12	<sup>4</sup> School of Earth and Environmental Sciences, Cardiff University, Park Place, Cardiff, U.K.
13	<sup>5</sup> School of Biosciences, Cardiff University, Museum Avenue, Cardiff, U.K.
14	<sup>6</sup> Cardiff School of Health Sciences, Cardiff Metropolitan University, Western Avenue, Cardiff, U.K.
15	<sup>7</sup> Shanghai Carbon Data Research Center (SCDRC) CAS Key Laboratory of Low-Carbon Conversion
16	Science and Engineering, Shanghai Advance Research Institute, Chinese Academy of Sciences,
17	Shanghai 2010210, China
18	<sup>8</sup> Key Laboratory of Aerosol Chemistry and Physics, SKLLQG, Institute of Earth Environment, Chinese
19	Academy of Sciences, Xi'an 710075, China
20	<sup>9</sup> Institute of Global Environmental Change, Xi'an Jiaotong University, Xi'an, China
21	<sup>10</sup> Department of Civil and Structural Engineering, Research Center of Urban Environmental
22	Technology and Management, The Hong Kong Polytechnic University, Hong Kong, China
23	<sup>11</sup> School of Respiratory Therapy, College of Medicine, Taipei Medical University, Taipei, Taiwan
24	<sup>12</sup> Division of Pulmonary Medicine, Department of Internal Medicine, Shuang Ho Hospital, Taipei
25	Medical University, Taipei, Taiwan
26	<sup>#</sup> These authors contributed equally to this work.
27	*Corresponding author. Tel.: +852 22528763; fax: +852 26063500
28	E-mail address: kfho@cuhk.edu.hk

29

## 30 Abstract

The mass, chemical composition and toxicological properties of fine particulates  $(PM_{2.5})$ 31 emitted from cooking activities in three Hong Kong based restaurants and two simulated 32 cooking experiments were characterized. Extracts from the cooking PM<sub>2.5</sub> elicited significant 33 biological activities [cell viability, generation of reactive oxygen species (ROS), DNA damage 34 35 and inflammation effect (TNF- $\alpha$ )] in a dose-dependent manner. The composition of PAHs, oxygenated PAHs (OPAHs) and azaarenes (AZAs) mixtures differed between samples. The 36 37 concentration ranges of the  $\Sigma$ 30PAHs,  $\Sigma$ 170PAHs and  $\Sigma$ 4AZAs and  $\Sigma$ 7 carbonyls in the samples were  $9627 - 23452 \text{ pg m}^{-3}$ ,  $503 - 3700 \text{ pg m}^{-3}$ ,  $33 - 263 \text{ pg m}^{-3}$  and  $158 - 5328 \text{ ng m}^{-3}$ 38 <sup>3</sup>, respectively. Cell viability caused by extracts from the samples was positively correlated to 39 40 the concentration of benzo[a]anthracene, indeno[1,2,3-cd]pyrene and 1,4-naphthoquinone in the PM<sub>2.5</sub> extracts. Cellular ROS production (upon exposure to extracts) was positively 41 correlated with the concentrations of  $PM_{2.5}$ , decaldehyde, acridine,  $\Sigma 17OPAHs$  and 7 42 individual OPAHsTNF- $\alpha$  showed significant positive correlations with the concentrations of 43 most chemical species (elemental carbon, 16 individual PAHs including benzo[a]pyrene, 44  $\Sigma$ 30PAHs, SO<sub>4</sub><sup>2-</sup>, Ca<sup>2+</sup>, Ca, Na, K, Ti, Cr, Mn, Fe, Cu and Zn). The concentrations of Al, Ti, 45 Mn,  $\Sigma$ 30 PAHs and 8 individual PAHs including benzo[a]pyrene in the samples were positively 46 47 correlated with DNA damage caused by extracts from the samples. This study demonstrates 48 that inhalation of PM<sub>2.5</sub> emitted from cooking could result in adverse human health effects.

49

## 50 *Keywords*:

51 Cooking emissions; PAHs; Oxygenated PAHs; Azaarenes; Plasmid scission assay

- 52
- 53

#### 54 1. Introduction

Emissions from cooking constitute an important source of particulate matter (PM) in the indoor 55 and outdoor environment (Abdullahi et al., 2013; Cheng et al., 2016). Emissions from cooking 56 has been identified as an important source of fine particulate matter (aerodynamic diameter < 57 2.5 µm: PM<sub>2.5</sub>) in populated urban areas such as Hong Kong (Allan et al., 2010; Huang et al., 58 2011; Mohr et al., 2012). A previous study showed that commercial cooking restaurants in the 59 60 South Coast Air Basin, USA emitted ~10.4 tons/day of PM<sub>2.5</sub> (Gysel et al., 2018). Such emissions represent a major source of exposure to PM<sub>2.5</sub>, which can adversely affect human 61 health (Chiang et al., 1997; See and Balasubramanian, 2006; Zhong et al., 1999a). 62

Previous review articles have documented the emission of PM of various size ranges and > 63 300 chemicals species [e.g. organic carbon (OC), elemental/black carbon (EC/BC), 64 metals/metalloids, water soluble ions (Cl<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, NO<sub>3</sub>-, K<sup>+</sup>, Ca<sup>2+</sup>, Mg<sup>2+</sup>), volatile organic 65 compounds (VOCs), carbonyls, polycyclic aromatic hydrocarbons (PAHs)] from cooking 66 activities (Abdullahi et al., 2013; Wang et al., 2017; Zhao and Zhao, 2018). The emitted PM, 67 68 chemical species in gaseous phase and bound to PM are formed from a range of reactions (hydrolysis, thermal oxidation, Maillard reaction, recombination) between chemical 69 components of oils, fats, solid food components, water etc. under high temperature (Abdullahi 70 et al., 2013). 71

The amount, size distribution and composition profiles of PM and their bound chemicals emitted from cooking are driven by factors such as cooking styles, cooking methods, types of oil, ingredients, additives, indoor and ambient conditions (Buonanno et al., 2009; He et al., 2004; McDonald et al., 2003; See et al., 2006; Torkmahalleh et al., 2013; Ho et al., 2006; Huang et al., 2011; Nolte et al., 1999; Saito et al., 2014; Zhao et al., 2007). Cooking (particularly in Asian-style) often involves the application of high temperature oil (> 250 °C)

which results in the enhanced formation of various organic compounds such as carbonyls and 78 PAHs (Huang et al., 2011; See et al., 2006). The amount of PAHs released from cooking that 79 80 involve deep frying (oil-based cooking) was found to be higher than boiling and steaming (water-based cooking) (See and Balasubramanian, 2008). In addition, frying and food 81 ingredients (e.g. fat contents) could be important factors in the PAHs formation during cooking 82 (Saito et al., 2014; Tanaka et al., 2012; Zhu and Wang, 2003). Oxygenated PAHs (OPAHs), 83 84 nitrogen heterocyclic PAHs (azaarenes: AZAs) are generated together with PAHs from combustion/pyrolysis process. OPAHs can additionally be formed from the photolysis and 85 86 (photo)chemical oxidation of PAHs (Clergé et al., 2019). The formation of AZAs and OPAHs during cooking have been documented (Blaszczyk and Janoszka, 2008; Szterk, 2015; Li et al., 87 2016) but these compounds are rarely characterized in cooking fumes (Sun et al., 2020). 88 Several OPAHs (quinones) and AZAs are cytotoxic, involved in the generation reactive oxygen 89 species (ROS), genotoxic, mutagenic, and carcinogens (Bolton et al., 2000; Jung et al., 2001; 90 Sovadinová et al., 2006; Clergé et al., 2019). Some OPAHs and AZAs also been classified by 91 the IARC as probable or possible human carcinogens (IARC, 2010; 2011a,b). Despite some 92 OPAHs and AZAs being equally or even more toxic than the parent-PAHs, most previous 93 studies on emissions from cooking did not characterize the OPAHs and AZAs and hence did 94 not elucidate their possible contributions to the adverse effects of PM (Abdullahi et al., 2013; 95 Ding et al., 2012). 96

97 Cooking activities also emit carbonyls and the sources were from cooking fuels combustion
98 and heating of cooking oils (Ho et al., 2006; Lin and Liou, 2000; Zhang and Smith, 1999).
99 Some of the aldehydes that are emitted from cooking activities are toxic themselves and also
100 participate in atmospheric chemical reactions to form secondary pollutants that degrade air
101 quality (Huang et al., 2011; Ho et al., 2006).

Epidemiological studies have demonstrated associations between exposure to cooking fumes 102 and lung cancer risk (Seow et al., 2000; Zhong et al., 1999a). Several studies found that the 103 104 risk of lung cancer in non-smoking women increases with increasing exposure to cooking fumes (Zhong et al., 1999a; Seow et al., 2000; Metayer et al., 2002). Some of the studies linked 105 in lung cancer risk to exposure to fumes from cooking (frying) with oil at high temperature 106 such as in Chinese-style cooking (Metayer et al., 2002; Shields et al., 1995; Zhong et al., 107 108 1999b). PM emitted from cooking was shown to have mutagenic activity (Chiang et al., 1997; Qu et al., 1992; Wu et al., 1998). The International Agency for Research on Cancer (IARC) 109 110 has classified fumes originating from high temperature frying as a probable human carcinogen (Straif et al., 2006). 111

112 Upon inhalation, PM<sub>2.5</sub> and the chemicals bound to it such as quinones and transition metals it can trigger the overproduction of reactive oxygen species (ROS), which counteracts anti-113 oxidative defences (Gao et al., 2020; Charrier et al., 2014). The physiological effects of ROS 114 imbalance can cause oxidative stress, inflammatory response, DNA and cell damage, which is 115 the basis for several diseases (Kelly, 2003; Bitterle et al., 2006). Transition metals, secondary 116 organic aerosols and OPAHs (quinones) were shown to induce ROS formation (Charrier et al., 117 2014; Bates et al., 2019). Polycyclic aromatic compounds (PAHs, OPAHs, AZAs) can cause 118 119 oxidative DNA damage and DNA adduct formation (Clergé et al., 2019; Xue and Warshawsky., 2005; Yamada et al., 2004; Bolton et al., 2000) that can result in 120 carcinogenic/mutagenic effects and cancers. 121

Knowledge of the amounts, chemical composition and toxicity of PM<sub>2.5</sub> emitted from cooking activities remain limited. The aims of this study are to: 1) characterize the chemical composition (including the particularly understudied OPAHs and AZAs) of PM<sub>2.5</sub> samples collected from different cooking operations; 2) determine the bioreactivity (cell viability, ROS production, DNA damage, and inflammation effects) of extracts of these PM<sub>2.5</sub> samples and 3)

determine the relationship between PM<sub>2.5</sub> chemical components and bioreactivity.

128

## 129 2. Materials and methods

## 130 2.1 Sampling locations

Cooking fumes were collected from the exhausts of three commercial restaurants operating in 131 Hong Kong and two simulated cooking experiments conducted in environmental chambers. 132 The characteristics of the different sampling sites are detailed in Table 1. The studied 133 restaurants cooked and served common cuisines in Hong Kong. The restaurants were selected 134 based on several criteria. Factors such as roof top access availability, electricity supply during 135 sampling, sampling space (minimum 2 m<sup>2</sup>), floor plans, information of exhaust system and 136 sampling safety were carefully considered prior to the sampling campaign. Each of the selected 137 restaurants had to possess an independent exhaust system. This was to ensure that the collected 138 samples were only generated from the target restaurant. 139

140 The simulated cooking by two common Chinese-style techniques (namely as stir-frying and 141 deep-frying) were conducted in stainless steel environmental chamber of 19.1 m<sup>3</sup> (3.05 m  $\times$ 142 3.05 m  $\times$  2.05 m) that was designed for measuring emissions from indoor sources.

143

#### 144 2.2 Experimental procedures

145 2.2.1 Sampling from restaurant exhaust system

Sampling duration was synchronised to the peak hours of the restaurants. The peak hours in each restaurant were based on information provided by the owners. A particle sampler (DRI MEDVOL) was used for sampling in this study. The device consisted of a PM2.5 cyclone, an inlet stilling chamber, a conical plenum, open-faced filter packs and differential pressure flow 150 control together with a pump. The PM<sub>2.5</sub> cyclone (Bendix 240) was operated at 113 L/min, 151 which removed particles (aerodynamic diameter >  $2.5 \,\mu$ m) in the air stream. The air was purged 152 through the cyclone and further diffused inside to the plenum. The plenum was coated with 153 perfluoro alkoxyalkane (PFA) teflon. The PFA teflon open faced filter holder (Savillex 47-mm 154 injection-moulded) was used throughout the sampling section. A Teflon-membrane (47mm), 155 Nuclepore polycarbonate membrane and quartz-fiber filter (47 mm) were positioned in the 156 filter holders separately.

Sampling was not performed in the kitchen area but at the rooftop, where sufficient space and access to exhaust for sampling were assured. The PM<sub>2.5</sub> samples were collected simultaneously during the cooking process. The sampling duration was set as 1.5 hours. Background samples from the kitchens were also collected when the kitchens were not in operation. The background samples were typically collected 1.5-3 hours after the restaurants have closed their operations for the day. Four cooking emission samples, in addition to background and field blank samples were collected at each restaurant for four consecutive sampling days.

164

165 2.2.2 Sampling from simulated cooking in an environmental test chamber

Induction hot plates were used for the cooking. Each cooking experiment was completed in 1.5 166 hours. The dishes were representative of typical Hong Kong cuisine including fried "Choy 167 Sum" (a leafy vegetable commonly used in Chinese cuisine) and deep fried chicken breast. The 168 portion of all dishes was enough for 4-5 members of a family. The same chef was responsible 169 for the cooking experiments in order to ensure sample harmony. PM2.5 emitted from these 170 simulated cooking experiments (sample D\*-E\* from test chamber and sample A-C from 171 commercial restaurant) were sampled with an identical sampling system as described in section 172 2.2.1. After each sampling, the filters were sealed in Petri dishes and frozen (-20 °C) until 173 chemical and biological analysis. 174

The quartz filters were heated at 900 °C for 3 hours in order to remove any organic vapours on filters before being used to sample. All filters were pre-conditioned at  $23\pm0.5$  °C and  $50\pm5\%$ relative humidity (RH) for 48 hours before and after weighing. Each filter was weighed on a microbalance (±1 µg precision, Sartorius AG MC5, Germany) before and after PM<sub>2.5</sub> sample collection.

180

181 2.3 Analytical methods

182 2.3.1 Chemical components analysis

183 Detailed description of the analytical methods applied for the determination of the chemical components [OC, EC, inorganic elements, water-soluble ions (Cl<sup>-</sup>, NO<sub>3</sub>-, SO<sub>4</sub><sup>2-</sup>, Na<sup>+</sup>, Ca<sup>2+</sup> and 184 NH4<sup>+</sup>), PAHs, OPAHs, AZAs and carbonyls] in the sampled PM<sub>2.5</sub> can be found in 185 Supplementary Material (Text S1-S4). In summary, the OC and EC were analysed by the 186 thermal/optical carbon analyser. The inorganic elements and water soluble ions were analysed 187 by the Energy Dispersive X-ray Fluorescence Analyser (ED-XRF) and ion chromatography 188 system, respectively. The polycyclic aromatic compounds (PACs) in the filters were extracted 189 with organic solvents and determined by gas chromatography-mass spectrometry (GC-MS) 190 (Bandowe et al., 2016; Bandowe et al., 2014). The carbonyls were extracted from filters with 191 methanol, derivatized using pentafluorobenzylhydroxyl amine (PFBHA) and analyzed by GC-192 MS. A list of the chemical species determined in the samples are shown in Table S1. 193

- 194
- 195 2.3.2 Cell culture and bioreactivity analysis

The human alveolar epithelial cells (A549) were obtained from American Type Culture Collection (ATCC, Rockville, MD, USA) and were cultured in RPMI medium containing 10% fetal bovine serum, penicillin, and streptomycin at 37 °C with 95% humidity and 5% CO<sub>2</sub>. PM<sub>2.5</sub> samples were from Teflon filters removed according to previous reports (Wang et al., 200 2021), followed by resuspended in 0.01% dimethyl sulfoxide (DMSO) in serum-free minimum 201 essential RPMI medium. Cells were exposed to the PM<sub>2.5</sub> samples at 0 (from blank filter), 100 202 and 200  $\mu$ g/mL for 24 hrs. Cells were examined for cell viability and ROS activity, whereas 203 the supernatant was collected for cytokine analyses.

The cell viability was identified by MTT (3-[4, 5-dimethylthiazol-2-yl]-2, 5 diphenyl tetrazolium bromide) assay. Cellular ROS production was determined by fluorogenic cell based method using 2',7'-dichlorodihydrofluorescein diacetate (DCFH-DA) as a probe. Inflammation marker tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) was detected through enzyme-linked immunosorbent assay (ELISA) kits (R&D systems, Inc., MN, USA) as per manufacturer guidelines. Details of the cell experiments can be found in Supplementary Material (Text S5-6).

210

211

## 212 2.3.3 Oxidative DNA damage

213 The plasmid scission assay (PSA) was used to determine the capability of each sample to induce oxidative DNA damage. The level of particle–DNA interaction and subsequent damage 214 were measured by the three conformations of plasmid DNA topological states namely: 215 supercoiled (no damage), relaxed (minor damage), and linear (severe damage) as shown in 216 Figure S1-2 (Supplementary Material). Due to the amount of sample required for the analysis, 217 PM<sub>2.5</sub> samples were pooled together for PSA analysis. Additional information about the 218 procedure can be referred to in previous studies (Chuang et al., 2013; Shao et al., 2006). The 219 PM<sub>2.5</sub> samples were run in suspension using molecular grade water over a range of 220 221 concentrations. Twenty nanograms (20 ng) of  $\Phi$ X174 RF DNA was added to the liquid and incubated for the analysis. The samples were conducted in triplicate. The final gel results were 222 captured in images and determined by densitometric analysis (Genetools; Syngene system, 223

UK). Molecular grade water and restriction enzyme PstI were used as control and positivecontrol in this study, which caused 4.1% and 95.9% DNA damage, respectively.

226

227 2.4 Calculations and statistical analysis

The sum of the concentration of all analysed PAHs, parent-PAHs, 16 US EPA PAH, OPAHs, 228 AZAs, carbonyls are referred to as  $\Sigma$ 30PAHs,  $\Sigma$ 21 parent-PAHs,  $\Sigma$ US-EPA PAHs,  $\Sigma$ 170PAHs, 229 230  $\Sigma$ 4AZAs and  $\Sigma$ Carbonyls, respectively. The sum of the concentration of parent-PAHs with 2-3 and 4-7 benzene rings are referred to as  $\Sigma$ LMW-PAHs and  $\Sigma$ HMW-PAHs, respectively. 231 232  $\Sigma$ Carci-PAHs refers to the sum of the concentration of eight carcinogenic-PAHs (benzo [a]anthracene, chrysene, benzo[b]fluoranthene, benzo[k]fluoranthene, benzo[a]pyrene, 233 indeno[1,2,3-cd]pyrene, dibenzo[a,h]anthracene, benzo[ghi]perylene). Statistical analysis was 234 performed using SPSS 21.0 software. The significance level was set at p < 0.05. Due to the small 235 sample size and non-parametric nature of the dataset, comparisons of the means of all samples types 236 237 were done with Games-Howell test. Spearman's rank correlation was applied to identify relationships 238 between chemical species and also between chemical species and biological end points [cell viability, ROS formation, TNF- $\alpha$ , oxidative DNA damage]. The DNA damage value at 1000  $\mu$ g ml<sup>-1</sup> dosage and 239 240 ROS production and TNF- $\alpha$  values at extract concentration of 200 µg/ml were chosen for the correlation analysis. 241

242

## 243 **3.** Results and discussion

244 3.1 Concentration of PM<sub>2.5</sub>, OC and EC

Mass concentrations of PM<sub>2.5</sub> in all samples are shown in Table 2. The highest and lowest mass concentrations of PM<sub>2.5</sub> were in samples C (711.5  $\pm$  257.2 µg m<sup>-3</sup>) and B (177.4  $\pm$  58.4 µg m<sup>-</sup> <sup>3</sup>). The concentration of PM<sub>2.5</sub> in sample C was ~1.45–4.01 times higher than in other samples. The high concentration of PM<sub>2.5</sub> in sample C could be attributed to cooking methods (stir-frying and deep-frying) in this restaurant which is supported by similar findings from other studies (Abdullahi et al., 2013; See and Balasubramanian, 2006). The concentrations of  $PM_{2.5}$  in all samples are comparable to a recent study focused on outdoor char broiling and conventional Chinese cooking (Li et al., 2018). The concentrations of  $PM_{2.5}$  in sample D\* and E\* (chamber) are also comparable to other Chinese cooking styles (Shandong and Hunan) as reported in a previous study (Wang et al., 2015).

255 The OC was the most abundant chemical component in the PM<sub>2.5</sub> (samples A-E\*). The average OC concentration is in a range of 99.9–338.8  $\mu$ g/m<sup>3</sup>. The OC concentration in sample C 256 257 (highest concentration) is ~3.39 times higher than in sample B (lowest concentration). The composition of the PM<sub>2.5</sub> mass was predominantly comprised of carbonaceous particles, 258 particularly in OC at a proportion, which are higher than or comparable to previous studies (Li 259 260 et al., 2018; Wang et al., 2015). The results are consistent with other findings that PM emissions from cooking operations were primarily organic in nature (Gysel et al., 2017; Li et al., 2015; 261 Wang et al., 2015; Zhang et al., 2017). The lower OC composition (<90%) compared to another 262 study on emissions from charbroiling/grilling of chicken and beef could be due to relatively 263 higher use of vegetables in Chinese cuisine (McDonald et al., 2003). A previous study also 264 reported that OC constituted the highest fraction of the mass PM emissions from different 265 cooking processes (See and Balasubramanian, 2008). The highest carbon fractions (> 60%) in 266 sample D\* and E\* could possibly be due to high-fat content of the cooking materials and high 267 amount of oils used in the cooking processes (Zhang et al., 2017). The EC fractions contribute 268 < 5% of the total PM<sub>2.5</sub> mass in each of the five samples and is consistent with a previous study 269 (Wang et al., 2015). The variations in the average concentrations of PM<sub>2.5</sub>, OC and EC in the 270 five samples could be due to the differences in cooking ingredients, cooking conditions and 271 methods (Gysel et al., 2018) some of which are outlined in Table 1. 272

#### 274 3.2 Concentration of inorganic elements and water extractable ions

The concentrations of elements and ions can be found in Table 2. The two most abundant 275 elements in each sample were S and Cl. Other major elements found in the samples were Na, 276 K, Fe, Mg, Al, Ca, Zn and Ba. Elements with medium range concentrations in the samples 277 were Mn, Sb, Pb and Cu while four other elements (i.e. Ti, V, Cr, Co and Ni) were in low 278 abundance (< 10 ng m-3) in all samples. The highest concentration of 16 of the 20 elements 279 280 studied was in sample C. Most of the elements are components of food ingredients (vegetables, meat, cooking oils, salt, spices, water (Butnariu and Butu; Cobos and Diaz, 2014; Epstein, 1999) 281 282 and can therefore be emitted as part of the PM in cooking fumes. Previous studies have also detected these elements in fumes from restaurants with different cooking styles and cooking of 283 meat in simulated facilities (Gysel et al., 2018; Abdullahi et al., 2013; See and Balasubramian, 284 2008; McDonald et al., 2003). The presence of Fe, Ni, Cr and Cu in the samples could also be 285 due to the release from cooking utensils (See et al., 2006; Taner et al., 2013; Gysel et al., 2018). 286 A previous study showed that the Cr composition of stainless steel materials could vary from 287 11 to 30% (Kuligowski and Halperin, 1992). Some of the elements could have contaminated 288 the food materials during their growing or processing because these elements are present in 289 environmental compartments (air, soil) from anthropogenic (traffic) and natural sources crustal 290 materials, dust, rocks, soils (Kebata-Pendias and Mukherjee, 2007; Louie et al., 2005). Some 291 of the elements (e.g. Na, Mg, Ca, Fe, Zn, Cu, Co, Mn) that have been detected in the PM 292 293 samples are essential for humans and are therefore harmless at the required concentrations, but others (e.g. Pb) have no known biological functions in humans and are only harmful to human 294 health (Kebata-Pendias and Mukherjee, 2007). Upon inhalation, some of the transition metals 295 (e.g. Fe, Cu, Mn) can catalyse ROS production leading to oxidative stress, inflammatory effects 296 and oxidative DNA damage, which results in diseases such as cancers, respiratory and 297

cardiovascular diseases (Gao et al., 2020; Pardo et al., 2015; Danielson et al., 2011; GerlofsNijland et al., 2009).

300 Water extractable ions were also found in all sample extracts (Table 2), which is consistent with previous studies that also detected these substances in cooking fumes both from cooking 301 test chambers and restaurants (Abdullahi et al., 2013; Schauer et al., 2002; See and 302 Balasubramian, 2008). The highest concentrations of each of the ions can be found in Sample 303 304 C. The concentration of the water soluble ions in sample C showed the following trend: NO<sub>3</sub><sup>-</sup> >  $SO_4^{2-}$  >  $NH_4^+$  >  $Cl^-$  >  $Na^+$  >  $Ca^{2+}$ . The trends in other samples are slightly different.  $NO_3^-$ , 305 SO<sub>4</sub><sup>2-</sup> and NH<sub>4</sub><sup>+</sup> could be components of water and other cooking ingredients but could also be 306 secondary products formed from SO<sub>2</sub>, NOx and NH<sub>3</sub> emitted from heating of cooking 307 ingredients (See and Balasubramanian, 2008; Schauer etal., 2002). 308

309

310 3.3 PAHs, OPAHs and AZAs in samples

The concentrations of PACs measured in the samples are shown in Table 3 and Figure S3. 311 Samples C shows the highest concentrations of  $\Sigma$ 30PAHs,  $\Sigma$ 170PAHs and  $\Sigma$ 4AZAs. Sample 312 C also showed the highest concentrations of 27 of 30 PAHs measured in the samples and the 313 sums of PAHs sub-groups (216 US-EPA PAHs, 2Carci PAHs, 2LMW-PAHs and 2HMW-314 315 PAHs). Out of the 17 individual OPAHs measured, 13 had the highest average concentrations in Sample C, while the highest concentration of each of the individual AZAs was also in 316 Sample C (Table 3). Benzo[a]pyrene is often used as a main indicator or marker of carcinogenic 317 PAHs (Boström et al., 2002). The concentration (mean±standard deviation in pg/m<sup>3</sup>) of 318 benzo[a]pyrene in the samples increased in the order: D (135 $\pm$ 32) > C (134.5 $\pm$ 82.4) > B 319  $(124\pm67) > E (114\pm35) > A (94\pm34)$ . The comparatively large amount of food processed and 320 cooked in the kitchen of this commercial restaurant (Sample C) could be a reason for PM<sub>2.5</sub> 321 samples to have the highest concentration of PAHs. Another unique feature of this restaurant 322

is that it cooks and serves Cantonese and Hong Kong local cuisine that are prepared with 323 methods such as deep frying, stir frying, pan frying and steaming (Table 1). Since the cooking 324 325 methods and size of restaurant C is comparable to restaurant B, the main reason for the high concentration of the chemical species in C can be explained by higher emissions during 326 cooking of Cantonese dishes than during the cooking of mixed cuisines and Western dishes 327 prepared in restaurants B and A, respectively (Table 1). Such influence of various cultural 328 329 cooking styles on the amount of emitted PAHs, as well as the fact that Asian cooking style emits higher PAHs than Western cooking has been reported in previous studies (See and 330 331 Balasubramanian, 2008; Abdullahi et al., 2013). Our study reveals that different cultural cooking styles will also result in different concentrations of the PAH derivatives (AZAs and 332 OPAHs) in cooking fumes. Specific studies have shown that cooking can result in the formation 333 of AZAs and OPAHs (Blaszczyk and Janoszka, 2008; Li et al., 2016; Szterk, 2015). A 334 combination of factors such as food components, relatively high usage of oil, type of oils, fat 335 content of meat, spices and cooking methods applied (frying at high temperatures) during the 336 cooking of Cantonese and local dishes results in the highest emitted concentrations of PAHs, 337 OPAHs and AZAs (Li et al., 2018; See and Balasubramanian, 2008). Oil-based cooking 338 methods could generally release more PAHs and their derivatives due to direct evaporation, 339 oxidation, pyrolysis, and/or degradation of organic compounds from oils at higher temperature 340 (Abdullahi et al., 2013; Moret and Conte, 2000). The higher concentration of  $\Sigma$ 30PAHs, 341 342  $\Sigma$ 170PAHs and  $\Sigma$ 4AZAs in sample E\* compared to sample D\* demonstrates that meat cooking by pan frying not only generates higher concentrations of PAHs (which has also been 343 previously reported) but also OPAHs and AZAs than cooking of plant based food by stir frying 344 (Abdullahi et al., 2013; Schauer et al., 1999a). The pyrolysis of animal fats and oils during the 345 cooking of meat could be an explanation for the formation of PAHs and OPAHs. Azaarenes 346 can be formed during cooking as a result of the pyrolysis of nitrogen containing organic 347

compound components (e.g. aromatic amino acids) of food ingredients (Blaszczyk and 348 Janoszka, 2008). The formed PACs could then subsequently be carried with the cooking fumes 349 (Rogge et al., 1991). Food ingredients may also be contaminated with PAHs, which can 350 351 subsequently be released with the cooking fumes (Martorell et al., 2010). The composition pattern of the PAHs were slightly different for the different samples (Figure 1). The most 352 abundant PAHs in samples A were naphthalene (10.9%), phenanthrene (10.7%), acenaphthene 353 354 (9.2%) with  $\Sigma$ LMW-PAHs/ $\Sigma$ HMW-PAHs of 1.4. Sample B was dominated by pyrene (10.6%), naphthalene (10.2%) and benzo[b+k+k]fluoranthene (8.1%) with  $\Sigma$ LMW-PAHs/ $\Sigma$ HMW-355 356 PAHs of 0.65. The most abundant PAHs in sample C were naphthalene (13%), cyclopenta[def]phenanthrene (11%) and pyrene (8%) with  $\Sigma LMW-PAHs/\Sigma HMW-PAHs$  of 357 1.08. Sample D\* were dominated by acenaphthene (11.2%), phenanthrene (9.9%) and retene 358 (9.8%) while sample E\* was dominated by naphthalene (14.4%), retene (12.6%) and 359 phenanthrene (9.6%). The *SLMW*-PAHs/*SHMW*-PAHs were 1.85 and 2.30 for samples D and 360 E, respectively (Table S3). A previous study showed that the concentration of pyrene emitted 361 from Chinese style cooking was higher than emitted from other cooking styles (e.g. Japanese) 362 (He et al., 2004). 363

The composition pattern of OPAHs mixtures differed in the various samples (Figure 1). Sample 364 A was dominated by 1,4-chysenequinone (21%), 1-indanone (11.4%) and 9-fluorenone (9.5%), 365 Sample B on the other hand was dominated by 1-indanone (14.4%), 1,4-chysenequinone 366 367 (13.8%) and 6H-benzo(cd)pyren-6-one (8.3%). 9-fluorenone (14%), 9,10-anthraquinone (12%) and 2-methylanthracene-9,10-dione (9.9%) were the dominant OPAHs in sample C. 368 Sample D\* was dominated by 1-indanone (19%), 1,4-naphthoquinone (9.9%), 2-methyl-369 anthracene-9,10-dione (14%) while sample E\* was dominated by 1-indanone (20.2%), 9-370 fluorenone (13.2%) and 1,4-anthraquinone (12.2%). Many of these found can also be found in 371 other environmental compartments such as ambient air and soil (Clergé et al., 2019). The 372

individual OPAH/parent-PAH ratio was in most cases highest in sample C. The 9,10anthraquinone/anthracene ratio was >1 for some samples (Table S3). The concentrations of  $\Sigma$ 170PAHs was significantly correlated with  $\Sigma$ 30PAHs (r = 0.91, p < 0.05). Many individual OPAHs were also significantly correlated with their related parent-PAHs. This can be explained by their similar sources and fate.

The four individual AZAs targeted in this study were identified and quantified in each of the 378 379 five samples (Table 3). Sample C had the highest average concentration of the  $\Sigma$ 4AZAs, which is similar to all the other PAC groups (Table 3). The contribution of individual AZAs to the 380  $\Sigma$ 4AZAs concentration was slightly different for each of the samples (Figure 1). For sample A, 381 382 the highest contributions were by carbazole (48%) and quinoline (32%). Quinoline (45%) and 383 benzo[h]quinoline (24%) were the dominant AZAs in sample B. Sample C was dominated by quinoline (38.4%) and carbazole (32.4%). This same trend was in sample D\* and E\* in which 384 385 the highest contribution was from quinoline, followed by carbazole (Figure 1). The substances are toxicologically relevant with carbazole being classified as possible human carcinogen 386 (IARC, 2011a; Yamada et al., 2004). None of the samples had individual AZA/parent-PAH 387 concentration ratio >1. The concentration ratios of individual AZA/parent-PAH were mostly < 388 10%, except in a few samples in which the acridine/anthracene and carbazole/fluorene 389 390 concentration ratio were  $\geq 10\%$  (Table S3). These ratios were thus generally lower than the individual OPAH/parent-PAH and individual alkylated PAH/parent-PAH concentration ratios 391 (Table S3). The  $\Sigma$ 4AZAs were also significantly correlated with  $\Sigma$ 30PAHs (r=0.83, p < 0.05). 392 393 This can be explained by the sources and fate of AZAs are similar to those of the PAHs and OPAHs (Bandowe et al., 2016). 394

395

## **396** 3.4 Concentration of carbonyls

The average concentrations of high-molecular-weight (HMW) mono-carbonyl ( $C \ge 6$ ) and di-397 carbonyl compounds (glyoxal and methylglyoxal) in PM<sub>2.5</sub> are shown in Table S4. Sample C 398 (5327.6±1974.1 ng/m<sup>3</sup>) showed the highest total concentration of HMW mono-carbonyl and 399 di-carbonyl compounds, whereas sample B recorded the lowest (159.2±48.7 ng/m<sup>3</sup>). 400 Nonanaldehyde was the most abundant component in all samples, accounting for  $\sim$ 31-81% of 401 total carbonyl compounds (Figure S4). The contribution of nonanaldehyde to the carbonyl 402 403 mixtures was > 80% in samples A and C. The contribution of each of the other carbonyl compounds to the total mixture was < 10% in sample A and C. Nonaldehyde was also the most 404 405 dominant contributor to the total carbonyl compounds in the other three sampling locations (B, D\*, E\*) but at these sites methylgloxal also made a high contribution (> 10%). Nonanaldehyde 406 was typically identified as a dominant carbonyl component in cooking that involves usage of 407 edible oils (Ho et al., 2006). The presence of nonanaldehyde could be due to the decomposition 408 of 9-octadecenoic acid (oleic acid), a known fatty acid produced from cooking oil thermal 409 410 decomposition (Schauer et al., 2002). A previous study showed that kitchens involved with frequent frying activities (e.g. western fast-food chain shops and Korean barbecue restaurant) 411 could be more abundant in nonanaldehyde (Ho et al., 2006). The two dicarbonyl compounds 412 413 were detected in all sampling locations, accounting for  $\sim 0.6-26.1\%$  of total carbonyl compounds. Sample A and C showed similar glyoxal (0.7% and 0.6%) and methylglyoxal 414 (3.3% and 2.9%) in their compositions. The highest contribution of decaldehyde to the carbonyl 415 mixtures was observed in sample  $D^*$  (10.5%) with much lower contributions (0.7-4.1%) in the 416 other samples (Figure S4). This observation could be attributed to the types of oils usage in the 417 cooking processes. A previous study showed that different types of seed oils (e.g. soybean and 418 canola oil) could generate ~4.77 times difference of decaldehyde in emissions (Schauer et al., 419 2002), although further study is necessary. The results show that cooking activities are 420 significant anthropogenic source of semi-volatile aldehydes. 421

422

## 423 3.5 Relationships between chemical species

There were significant correlations between the concentrations of many individual and sums 424 of chemical species (Table S5). For example, the significant correlations between the  $\Sigma$ 30PAHs, 425  $\Sigma$ 170PAHs and  $\Sigma$ 4AZAs and the PM<sub>2.5</sub> mass, TC, EC, OC and  $\Sigma$ carbonyls can be attributed to 426 their similar sources, the fact that they are co-sorbed with each other and hence have similar 427 428 fates. Some ions like Cl<sup>-</sup> and several metals were also strongly correlated with PAHs, OPAHs and AZAs, which might also be strong indication of their similarity in sources but especially 429 similarity in their fates. Unlike Cl<sup>-</sup> the correlations between the PACs and SO<sub>4</sub><sup>2-</sup>, NO<sub>3</sub><sup>-</sup>, NH<sub>4</sub><sup>+</sup>, 430 and Ca<sup>2+</sup> were not statistically significant indicating that the sources and fate of these ions 431 might be much more different to that of the PACs. 432

433

## 434 3.6 Bioreactivity of PM<sub>2.5</sub>

The cell viability, oxidative potential (ROS generation), inflammatory reactions (TNF- $\alpha$ ) and 435 oxidative DNA damage elicited by PM<sub>2.5</sub> collected from different cooking sites are shown in 436 Figures 2 and Table S2 (i.e. oxidative DNA damage under particle concentrations of 50, 100, 437 500 and 1000 µg/ml dosage). The cell viability of A549 cells demonstrates negative dose-438 response from all samples (Figure 2). Under the particle concentration of 200 µg/ml, it can be 439 observed that sample C and D\* further showed lowest cell viabilities. Positive dose-response 440 441 was nevertheless identified in ROS generation and TNF-a (Figure 2) suggesting that oxidative and inflammatory reactions could be enhanced by the increase of particle concentration. 442 Sample C shows the highest ROS generation followed by sample B, whereas sample D\* and 443 E\* demonstrated higher oxidative potential (under 200 µg/ml dosage) in comparison with other 444 samples. 445

A general increasing trend between particle dose concentration and DNA damage was observed in most sub-samples (except for the sub-sample 1-4 of sample A, Table S2 and Figure S6). The amount of damage to the plasmid DNA induced by  $PM_{2.5}$  varied over the range of 2.3-65.8% (Figure S5) in the samples under 1000 µg/ml dosage. These findings are relatively low compared to the result from a study on  $PM_{10}$  derived from coal burning (0-55% under 500 µg/ml) (Shao et al., 2016).

452 Median lethal dose (LD<sub>50</sub>) of the samples (Figure S5) were determined by dosage response 453 analysis in Figure S6 (Supplementary Material). Sample A showed highest LD<sub>50</sub> and the lowest 454 LD<sub>50</sub> was in sample E\*. The results indicate that lower PM<sub>2.5</sub> concentration was required to 455 cause 50% DNA damage for sample E\* compared to the other samples. Both sample B and C 456 show comparable DNA damage in this study. This observation could be due to similar cooking 457 characteristics.

Chemical species and elements some of which have been quantified (e.g. heterocyclic amines, metals, and PACs) in emissions from cooking could cause oxidative stress, cell injury, DNA damage and mutations, which could result in the above observed effects and risk of diseases such as lung cancer, cardiovascular and pulmonary diseases (Pardo et al., 2015; Gerlofs-Nijland et al., 2009; Wei et al., 2009; Xue and Warshawsky, 2005; Bolton et al., 2000; Seow et al., 2000).

464

465 3.7 Correlation between chemical components and bioreactivities

466 Cell viability showed significant negative correlations with the concentrations of
467 benzo[a]anthracene, indeno [1,2,3-cd]pyrene and 1,4-naphthoquinone in the samples (Table
468 4). These relationships suggest that these compounds might contribute to the toxicity of extracts
469 of the PM<sub>2.5</sub> samples emitted from cooking activities.

Significant positive correlations were found between the ROS levels and the concentrations 470  $\Sigma$ 170PAHs in addition to several individual OPAHs (2-biphenlcarboxaldehyde, 1-471 472 acenaphthenone, 9-fluorenone, 9,10-anthraquinone, 2-methylanthraquinone, benzo[a]fluorenone), acridine and decaldehyde (Table 4 and Figure S7). The results of our 473 study are consistent with other studies that show that OPAHs (and other quinones) are redox 474 active chemical species in air PM that are associated with particle induced oxidative potential 475 476 (Gao et al., 2020; Tuet et al., 2019; Sheng and Lu, 2017; Shang et al., 2013; Bolton et al., 2000).

None of the PAHs or alkyl-PAHs were significantly positively correlated to ROS generation 477 (oxidative potential), in this study (Table 4). Positive correlations between the concentrations 478 479 of PAHs in PM and cellular ROS activity (after exposure to extracts of the PM) has been 480 reported (Tuet et al., 2019; Daher et al., 2012; Hu et al., 2008). PAHs and alkyl PAHs can only participate in reactions leading to ROS production after their biological transformation to redox 481 482 active compounds (Verma et al., 2011; Ntziachristos et al., 2007; Bolton et al., 2000). The lack of significant positive correlation between ROS generation and the concentration of PAHs in 483 our study might be explained by the mismatches between the PAHs extracted from the filters 484 and the fraction of PAHs that is bioavailable/bioaccessible to the human alveolar epithelial 485 cells (A549) for bioreactivity (Li et al., 2019; Baulig et al., 2004). 486

None of the (transition) metals which have also been identified as strong inducers of ROS formation were significantly positively correlated with ROS generation in this study (Gao et al., 2020; Bates et al., 2019; Daher et al., 2014; Verma et al., 2009; Hu et al., 2008). It is important to note that the method for determining the concentration of metals in our study (ED-XRF, see Text S2) measures the total concentration and not the water-soluble (bioavailable/bioaccessible fractions). It is the water-soluble concentrations (not the total concentrations) of some transition metals in fine particulate matter that are found to have 494 positive correlation with ROS generation in cellular assays (Shafer et al., 2016; Daher et al.,
495 2014; Verma et al., 2009; Hu et al., 2008).

Strong positive correlations were also found between TNF- $\alpha$  and the concentration of several 496 chemical species (EC, Ca<sup>2+</sup>, SO<sub>4</sub><sup>2-</sup>, K, Ca, Ti, Cr, Mn, Fe, Cu, Zn, Ni, 14 individual PAHs and 497  $\Sigma$ 30PAHs; Table 4 and Figure S7). The TNF- $\alpha$  was however significantly negatively correlated 498 with the concentration of more polar PAH derivatives ( $\Sigma$ 4AZAs and  $\Sigma$ 30OPAHs). This is 499 500 despite the fact that the concentrations of PAH derivatives were positively correlated with the concentrations of PAHs. This suggests that the impact of PAHs extracted from the PM<sub>2.5</sub> on 501 the generation of pro-inflammatory cytokines was opposite to the impact of PAH derivatives 502 503 (OPAHs and AZAs). Most of the PAHs showing strongest positive correlations with TNF-a 504 are those with 4-6 ring sizes (HMW-PAHs) with several of them classified as probable human carcinogens. Our study suggests that inflammation is a strong pathway for the toxicity of  $PM_{2.5}$ 505 506 emitted from cooking emissions and that several chemical species including PAHs, transition metals might be the triggers for the inflammatory response. Relationships between 507 inflammation and metal/PAH contents of PM has been reported in other studies (Pardo et al., 508 2015; Gerlofs-Nijland et al., 2009). 509

Significant positive correlation was observed between DNA damage and concentrations of several elements (Al, Ti, Cr and Mn),  $\Sigma$ 30PAHs and individual PAHs (1,2,3,4tetrahydronaphthalene, naphthalene, acenaphthene, dibenzo[a,h]anthracene, perylene, benzo[a]pyrene, benzo[b+j+k]fluoranthene and benzo[ghi]perylene) (Table 4). PAHs are known mutagens and carcinogens and damage to DNA is a known mechanism for their carcinogenic effect (Wei et al., 2009; IARC 2005; Bolton et al., 2000). A previous study showed cytotoxicity of Cr in Chinese commercial cooking PM<sub>2.5</sub> emissions (Sun et al., 2020).

517

## 518 **4.** Conclusions

This study investigated PM<sub>2.5</sub> emissions generated from different cooking conditions. The concentrations of nearly all chemical species were highest in one restaurant that served Cantonese dishes and applied oil frying methods, suggesting the cooking ingredients and conditions could be the determining factors for higher emissions. The extracts from the samples elicited toxicologically relevant responses (cell death, ROS generation, inflammation activity and DNA damage). The responses were in dose dependent manner and demonstrated higher

525 responses under higher  $PM_{2.5}$  concentrations. These responses were correlated to

526 concentrations of specific chemical species in the  $PM_{2.5}$  suggesting that some of the determined

- 527 chemical species might play roles in the toxicity of PM<sub>2.5</sub> emitted from cooking activities.
- 528

## 529 Acknowledgments

530 This study was supported by the Research Grants Council of the Hong Kong Special

Administrative Region China (General Research Fund, Project number: 14205318).

532

## 533 **References**

- Abdullahi, K.L., Delgado-Saborit, J.M., Harrison, R.M., 2013. Emissions and indoor concentrations of
   particulate matter and its specific chemical components from cooking: a review. Atmospheric
   Environment 71: 260-294. https://doi.org/10.1016/j.atmosenv.2013.01.061.
- Allan, J.D., Williams, P.I., Morgan, W.T., Martin, C.L., Flynn, M.J., Lee, J., Nemitz, E., Phillips, G.J.,
  Gallagher, M.W., Coe, H., 2010. Contributions from transport, solid fuel burning and cooking
  to primary organic aerosols in two UK cities. Atmospheric Chemistry and Physics 10: 647-668.
  https://doi.org/10.5194/acp-10-647-2010.
- Bandowe, B.A.M., Meusel, H., Huang, R., Ho, K., Cao, J., Hoffman, T., Wilcke, W., 2014. PM2.5bound oxygenated PAHs, 2014. PM2.5-bound oxygenated PAHs, nitro-PAHs and parent-PAHs
  from the atmosphere of a Chinese megacity:seasonal variation, sources and cancer risk
  assessment. Science of the Total Environment 473-474, 77-87
- Bandowe, B.A.M., Meusel, H., Huang, R., Hoffmann, T., Cao, J., Ho, K., 2016. Azaarenes in fine
  particulate matter from the atmosphere of a Chinese megacity. Environmental Science and
  Pollution Research 23, 16025-16036 https://doi.org/10.1007/s11356-016-6740-z.
- Baulig, A., Poirault, J.J., Ausset, P., Schins, R., Shi, T., Baralle, D., Dorlhene, P., Meyer, M., Lefevre,
  R., Baeza-Squiban A., Marano, F., 2004. Physicochemical characteristics and biological
  activities of seasonal atmospheric particulate matter sampling in two locations of Paris.
  Environmental Science and Technology 38: 5985-5992. https://doi.org/10.1021/es049476z.
- Bein, K.J., Wexler, A.S., 2015. Compositional variance in extracted particulate matter using different
   filter extraction techniques. Atmospheric Environment 107: 24-34.
   https://doi.org/10.1016/j.atmosenv.2015.02.026.

- Bitterle, E., Karg, E., Schroeppel, A., Kreyling, W.G., Tippe, A., Ferron, G.A., Schmid, O., Heyder, J.,
  Maier, K.L., Hofer, T., 2006. Dose-controlled exposure of A549 epithelial cells at the air–liquid
  interface to airborne ultrafine carbonaceous particles. Chemosphere 65: 1784-1790.
  https://doi.org/10.1016/j.chemosphere.2006.04.035.
- Blaszczyk, U., Janoszka, B., 2008. Analysis of azaarenes in pan fried meat and its gravy by liquid
  chromatography with fluorescence. Food Chemistry 109, 235-242.
  https://doi.org/10.1016/j.foodchem.2007.12.038.
- Bolton, J.L, Trush, M.A., Penning, T.M., Dryhurst, G., Monks, T.J., 2000. Role of quinones in toxicology. Chemical Research in Toxicology 13, 135-160. https://doi.org/10.1021/tx9902082.
- Boström, C-E., Gerde, P., Hanberg, A., Jernström, B., Johansson, C., Kyrklund, T., Rannug, A.,
  Törnqvist, M., Victorin, K., Westerholm, R., 2002. Cancer risk assessment, indicators, and
  guidelines for polycyclic aromatic hydrocarbons in the ambient air. Environmental health
  perspectives 110: 451. https://doi.org/10.1289/ehp.110-1241197.
- Buonanno, G., Morawska, L., Stabile, L., 2009. Particle emission factors during cooking activities.
   Atmospheric Environment 43: 3235-3242. https://doi.org/10.1016/j.atmosenv.2009.03.044.
- Butnariu, M., Butu, A, 2014. Chemical composition of vegetables and their products. Handbook of
   Food Chemistry. Springer-Verlag, Berlin Heidelberg. pp. 1-49.
   https://link.springer.com/content/pdf/10.1007%2F978-3-642-41609-5 17-1.pdf
- 573 Charrier, J.G., McFall, A.S., Richards-Henderson, N.K., Anastasio, C., 2014. Hydrogen peroxide
  574 formation in a surrogate lung fluid by transition metals and quinones present in particulate
  575 matter. Environmental Science & Technology 48: 7010-7017.
  576 https://doi.org/10.1021/es501011w.
- 577 Cheng, S., Wang, G., Lang, J., Wen, W., Wang, X., Yao, S., 2016. Characterization of volatile organic
  578 compounds from different cooking emissions. Atmospheric Environment 145: 299-307.
  579 https://doi.org/10.1016/j.atmosenv.2016.09.037.
- Chiang, T-A., Wu, P-F., Wang, L-F., Lee, H., Lee, C-H., Ko, Y-C., 1997. Mutagenicity and polycyclic aromatic hydrocarbon content of fumes from heated cooking oils produced in Taiwan. Mutation
  Research/Fundamental and Molecular Mechanisms of Mutagenesis 381: 157-161. https://doi.org/10.1016/S0027-5107(97)00163-2.
- Chuang, H-C., BéruBé, K., Lung, S-CC., Bai, K-J., Jones, T., 2013. Investigation into the oxidative
   potential generated by the formation of particulate matter from incense combustion. Journal of
   hazardous materials 244: 142-150. https://doi.org/10.1016/j.jhazmat.2012.11.034.
- 587 Clergé, A., Le Goff, J., Lopez, C., Ledauphin, J., Delépée, R., 2019. Oxy-PAHs: occurrence in the
  588 environment and potential genotoxic/mutagenic risk assessment for human health. Critical
  589 Reviews in Toxicology 49, 302-328. https://doi.org/10.1080/10408444.2019.1605333.
- Cobos, A., Diaz, O, 2014. Chemical composition of meat and meat products. Handbook of Food
   Chemistry. Springer-Verlag, Berlin-Heldelberg pp 1-22.
   https://link.springer.com/content/pdf/10.1007%2F978-3-642-41609-5\_6-1.pdf.
- Daher, N., Ruprecht, A., Invernizzi, G., De Marco, C., Miller-Schulze, J., Heo, J.B., Shafer, M.M.,
  Shelton, B.R., Schauer, J.J., Sioutas. C., 2012. Characterization, sources and redox activity of
  fine and coarse particulate matter in Milan, Italy. Atmospheric Environment 49: 130-141.
  https://doi.org/10.1016/j.atmosenv.2011.12.011.
- 597 Daher, N., Saliba, N.A., Shihadeh, A.L., Jaafar, M., Baalbaki, R., Shafer, M.M., Schauer, J.J., Sioutas.
  598 C., 2014. Oxidative potential and chemical speciation of size-resolved particulate matter (PM)
  599 at near-freeway and urban background sites in the greater Beirut area. Science of the Total
  600 Environment 470-471: 417-426. https://doi.org/10.1016/j.scitotenv.2013.09.104.
- Danielsen, P.G., Møller P., Jensen, K.A., Sharma, A.K., Wallin, H., Bossi, R., Autrup, H., Mølhave, L,.
  Ravanat, J.L, Briedé, J.J., de Kok, T.M., Loft, S., 2011. Oxidative stress, DNA Damage, and
  inflammation induced by Air and wood smoke particulate matter in Human A549 and THP-1
  Cell lines. Chemical Research in Toxicology 24, 168-184. https://doi.org/10.1021/tx100407m.
- De Oliveira, L.C.C., Vieira, M.A., Ribeiro, A.S., Lisboa, M.T., Gonçalves, R.A., De Campos, R.C.,
  2012. Determination of silicon in vegetable oil and biodiesel by high-resolution continuum
  source flame atomic absorption spectrometry using sample dilution with xylene. Energy &
  Fuels 26: 7041-7044. https://doi.org/10.1021/ef3012867.

- 609 Ding, J., Zhong, J., Yang, Y., Li, B., Shen, G., Su, Y., Wang, C., Li, W., Shen, H. Wang, B., Wang, R., Huang, Y., Zhang, Y., Cao, H., Zhu, Y., Simonich, S.L.M., Tao, S., 2012. Occurrence and 610 exposure to polycyclic aromatic hydrocarbons and their derivatives in a rural Chinese home 611 612 through biomass fuelled cooking. Environmental Pollution 169: 160-166. https://doi.org/10.1016/j.envpol.2011.10.008. 613
- Epstein, E., 1999. Silicon. Annual Review of Plant Physiology and Plant Biology 50, 641-664.
   https://doi.org/10.1146/annurev.arplant.50.1.641.
- Fortmann, R., Kariher, P., Clayton, R., 2001. Indoor air quality: residential cooking exposures final
   report. https://hero.epa.gov/hero/index.cfm/reference/details/reference\_id/90689.
- Fullana, A., Carbonell-Barrachina, Á.A., Sidhu, S., 2004. Comparison of volatile aldehydes present in
   the cooking fumes of extra virgin olive, olive, and canola oils. Journal of Agricultural and Food
   Chemistry 52: 5207-5214. https://doi.org/10.1021/jf035241f.
- Fullana, A., Carbonell Barrachina, Á.A., Sidhu S., 2004. Volatile aldehyde emissions from heated
  cooking oils. Journal of the Science of Food and Agriculture 84: 2015-2021.
  https://doi.org/10.1002/jsfa.1904.
- Gao, D., Ripleyb, S., Weichenthalb, S., Godri Pollitt, J.K., 2020. Ambient particulate matter oxidative
  potential: chemical determinants, associated health effects, and strategies for risk management.
  Free Radical Biology and Medicine 151, 7-25.
  https://doi.org/10.1016/j.freeradbiomed.2020.04.028.
- Gerlofs-Nijland, M.E., Rummelhard, M., Boere, A.J.F., Leseman, D.L.A.C., Duffin, R., Schins, R.P.F.,
  Borm, P.J.A., Sillanpää, M., Salonen, R.O., Cassee, F.R., 2009. Particle induced toxicity in
  relation to transition metal and polycyclic aromatic hydrocarbon content. Environmental
  Science and Technology 43, 4729-4736. https://doi.org/10.1021/es803176k.
- Gysel, N., Dixit, P., Schmitz, D.A., Engling, G., Cho, A.K., Cocker, D.R., Karavalakis, G., 2018.
  Chemical speciation, including polycyclic aromatic hydrocarbons (PAHs), and toxicity of
  particles emitted from meat cooking operations. Science of The Total Environment 633: 14291436. https://doi.org/10.1016/j.scitotenv.2018.03.318.
- Gysel, N., Welch, W.A., Chen, C-L., Dixit, P., Cocker III, D.R., Karavalakis, G., 2017. Particulate
  matter emissions and gaseous air toxic pollutants from commercial meat cooking operations.
  Journal of Environmental Sciences 65: 162-170. https://doi.org/10.1016/j.jes.2017.03.022.
- He, L-Y., Hu, M., Huang, X-F., Yu, B-D., Zhang, Y-H., Liu, D-Q., 2004. Measurement of emissions
  of fine particulate organic matter from Chinese cooking. Atmospheric Environment 38: 65576564. https://doi.org/10.1016/j.atmosenv.2004.08.034.
- Ho, S.S.H., Yu, J.Z., Chu, K.W., Yeung, L.L., 2006. Carbonyl emissions from commercial cooking sources in Hong Kong. Journal of the Air & Waste Management Association 56: 1091-1098. https://doi.org/10.1080/10473289.2006.10464532.
- Hu, S., Polidori, A., Arhami, M., Shafer, M.M., Schauer, J.J., Cho, A., Sioutas, C., 2008. Redox activity
  and chemical speciation of size fractionated PM in the communities of the Los Angeles-Long
  Beach harbor. Atmospheric Chemistry and Physics 8: 6439-6451. https://doi.org/10.5194/acp8-6439-2008.
- Huang, Y., Ho, S.S.H., Ho, K.F., Lee, S.C., Yu, J.Z., Louie, P.K.K., 2011. Characteristics and health
  impacts of VOCs and carbonyls associated with residential cooking activities in Hong Kong.
  Journal of hazardous materials 186: 344-351. https://doi.org/10.1016/j.jhazmat.2010.11.003.
- IARC, 2010. Some non-heterocyclic polycyclic aromatic hydrocarbons and some related exposures.
   IARC Monographs on the Evaluation of Carcinogenic Risks to Humans, Volume 92, Lyon
   France. https://monographs.iarc.who.int/wp-content/uploads/2018/06/mono92.pdf
- IARC, 2011a. Bitumens and bitumen emissions, and some N- and S-heterocyclic aromatic
   hydrocarbons. IARC Monographs on the Evaluation of Carcinogenic Risks to Humans 103, 1 311.https://publications.iarc.fr/Book-And-Report-Series/Iarc-Monographs-On-The-
- Identification-Of-Carcinogenic-Hazards-To-Humans/Bitumens-And-Bitumen-Emissions And-Some-Em-N-Em---And-Em-S-Em--Heterocyclic-Polycyclic-Aromatic-Hydrocarbons 2013
- IARC, 2011b. Some chemicals present in industrial and consumer products, food and drinking water.
   IARC Monographs on the Evaluation of Carcinogenic Risks to Humans, Volume 101, Lyon
   France. https://monographs.iarc.who.int/wp-content/uploads/2018/06/mono101.pdf

- Jung, D.K., Klaus, T., Fent, K., 2001. Cytochrome P450 induction by nitrated polycyclic aromatic
  hydrocarbons, azaarenes, and binary mixtures in fish hepatoma cell line PLHC-1.
  Environmental Toxicology and Chemistry 20, 149-159.
  https://doi.org/10.1002/etc.5620200117.
- Kebata-Pendias, A., Mukherjee, A.B., 2007. Trace elements from Soil to Humans. Springer-Verlag
   Berlin Heildelberg, https://link.springer.com/content/pdf/10.1007%2F978-3-540-32714-1.pdf.
- Kelly, F.J., 2003. Oxidative stress: its role in air pollution and adverse health effects. Occupational and
   Environmental Medicine 60: 612-616. http://dx.doi.org/10.1136/oem.60.8.612.
- Kuligowski, J., Halperin, K.M., 1992. Stainless steel cookware as a significant source of nickel,
  chromium, and iron. Archives of environmental contamination and toxicology 23: 211-215.
  https://doi.org/10.1007/BF00212277
- Li, C-T., Lin, Y-C., Lee, W-J., Tsai, P-J., 2003. Emission of polycyclic aromatic hydrocarbons and their carcinogenic potencies from cooking sources to the urban atmosphere. Environmental Health Perspectives 111: 483. https://doi.org/10.1289/ehp.5518.
- Li, Y-C., Qiu, J-Q., Shu, M., Ho, S.S.H., Cao, J-J., Wang, G-H., Wang, X-X., Zhao, X-Q., 2018.
  Characteristics of polycyclic aromatic hydrocarbons in PM 2.5 emitted from different cooking
  activities in China. Environmental Science and Pollution Research 25: 4750-4760.
  https://doi.org/10.1007/s11356-017-0603-0.
- Li Y-C., Shu, M., Ho, S.S.H., Wang, C., Cao, J-J., Wang, G-H., Wang, X-X. Wang, K., Zhao, XQ.,2015 Characteristics of PM2. 5 emitted from different cooking activities in China.
  Atmospheric Research 166: 83-91. https://doi.org/10.1016/j.atmosres.2015.06.010.
- Li, G., Wu, S., Zeng, J., Wang, L., Yu, W., 2016. Effect of frying and aluminium on the levels and migration of parent and oxygenated PAH in a popular Chinese fried bread youtiao. Food Chemistry 209, 123-130. https://doi.org/10.1016/j.foodchem.2016.04.036.
- Lin, J-M., Liou, S-J., 2000. Aliphatic aldehydes produced by heating Chinese cooking oils. Bulletin of
   environmental contamination and toxicology 64: 817-824.
   https://doi.org/10.1007/s0012800076.
- Louie, P.K.K., Chow, J.C., Antony Chen, L.-W., Watson, J.G., Leung, G., Sin, D.W.M, 2005. PM2.5
   chemical composition in Hong Kong: urban and regional variations. Science of the Total
   Environment 338, 267-281. https://doi.org/10.1016/j.scitotenv.2004.07.021.
- Lui, K.H., Bandowe, B.A.M et al., 2016. Characterization of the chemical components and bioreactivity
   of fine particulate matter (PM<sub>2.5</sub>) during incense burning. Environmental Pollution 213, 524 533.
- Martorelli, I., Perello, G., Martí-Cid, R., Castell, V., Llobet, J.M., Domingo, J.L., 2010. Polycyclic
  aromatic hydrocarbons (PAHs) in foods and estimated intake by the population of Catalonia,
  Spain Temporal trend. Environment International 36, 424-432.
  https://doi.org/10.1016/j.envint.2010.03.003
- McDonald, J.D., Zielinska, B., Fujita, E.M., Sagebiel, J.C., Chow, J.C., Watson, J.G.,2003. Emissions
   from charbroiling and grilling of chicken and beef. Journal of the Air & Waste Management
   Association 53: 185-194. https://doi.org/10.1080/10473289.2003.10466141.
- Metayer, C., Wang, Z., Kleinerman, R.A., Wang, L., Brenner, A.V., Cui, H., Cao J., Lubin, J.H.,2002.
   Cooking oil fumes and risk of lung cancer in women in rural Gansu, China. Lung Cancer 35:
   111-117. https://doi.org/10.1016/S0169-5002(01)00412-3.
- Mohr, C., DeCarlo, P.F., Heringa, M.F., Chirico, R., Slowik, J.G., Richter, R., Reche, C., Alastuey, A.,
  Querol, X., Seco, R., Peñuelas, J., Jiménez, J.L., Crippa, M., Zimmermann, R., Baltensperge,
  U., Prévôt, A.S.H., 2012. Identification and quantification of organic aerosol from cooking and
  other sources in Barcelona using aerosol mass spectrometer data. Atmospheric Chemistry and
  Physics 12: 1649-1665. https://doi.org/10.5194/acp-12-1649-2012
- Moret, S., Conte, L.S., 2000. Polycyclic aromatic hydrocarbons in edible fats and oils: occurrence and analytical methods. Journal of chromatography A 882: 245-253.
  https://doi.org/10.1016/S0021-9673(00)00079-0.
- Ntziachristos, L., Froines, J.R., Cho, A.K., Sioutas, C., 2007. Relationship between redox activity and chemical composition of size-fractionated particulate matter. Particle and Fibre Toxicology 4:5.

- Nolte, C.G., Schauer, J.J., Cass, G.R., Simoneit, B.R.T., 1999. Highly polar organic compounds present
  in meat smoke. Environmental Science & Technology 33: 3313-3316.
  https://doi.org/10.1021/es990122v.
- Pardo, M., Shafer, M.M., Rudich, A., Schauer, J.J., Rudich, Y, 2015. Single exposure to near Roadway
   particulate matter leads to confined inflammatory and defence responses: possible role of
   metals. Environmental Science and Technology 49, 8777-8785.
   https://doi.org/10.1021/acs.est.5b01449
- Qu, Y.H., Xu, G.X., Zhou, J.Z., Chen, T.D., Zhu, L.F., Shields, P.G., Wang, H.W., Gao, Y.T., 1992.
   Genotoxicity of heated cooking oil vapors. Mutation Research/Genetic Toxicology 298: 105-111. https://doi.org/10.1016/0165-1218(92)90035-X.
- Robberect, H., Van Dyck, K., Bosscher, D., Van Cauwenbergh, R., 2008. Silicon in Foods: content and bioavailability. International Journal of Food Properties 11, 638-645.
  https://www.tandfonline.com/doi/full/10.1080/10942910701584252.
- Rogge, W.F., Hildemann, L.M., Mazurek, M.A., Cass, G.R., Simoneit, B.R.T., 1991. Sources of fine
  organic aerosol. 1. Charbroilers and meat cooking operations. Environmental Science &
  Technology 25: 1112-1125. https://doi.org/10.1021/es00018a015.
- Roper, C., Chubb, L.G., Cambal, L., Tunno, B., Clougherty, J.E., Fattman, C., Mischler, S.E., 2017.
  Association of IL-6 with PM 2.5 Components: Importance of Characterizing Filter-Based PM
  2.5 Following Extraction. Water, Air, & Soil Pollution 228: 43. https://doi.org/10.1007/s11270016-3219-y.
- Roper, C., Delgado, L.S., Barrett, D., Massey Simonich, S.L., Tanguay, R.L., 2018. PM2. 5 Filter
   Extraction Methods: Implications for Chemical and Toxicological Analyses. Environmental
   science & technology 53: 434-442. https://doi.org/10.1021/acs.est.8b04308.
- Saito, E., Tanaka, N., Miyazaki, A., Tsuzaki, M., 2014. Concentration and particle size distribution of
   polycyclic aromatic hydrocarbons formed by thermal cooking. Food chemistry 153: 285-291.
   https://doi.org/10.1016/j.foodchem.2013.12.055.
- Schauer, J.J., Kleeman, M.J., Cass, G.R., Simoneit, B.R.T., 1999a. Measurement of emissions from air pollution sources. 1. C1 through C29 organic compounds from meat charbroiling.
  Environmental Science & Technology 33: 1566-1577. https://doi.org/10.1021/es980076j
- Schauer, J.J., Kleeman, M.J., Cass, G.R., Simoneit, B.R.T., 2002. Measurement of emissions from air pollution sources. 4. C1– C27 organic compounds from cooking with seed oils. Environmental Science & Technology 2002; 36: 567-575. https://doi.org/10.1021/es002053m.
- See, S.W., Balasubramanian, R., 2006. Risk assessment of exposure to indoor aerosols associated with
   Chinese cooking. Environmental research 102: 197-204.
   https://doi.org/10.1016/j.envres.2005.12.013.
- See, S.W., Balasubramanian, R., 2008. Chemical characteristics of fine particles emitted from different
   gas cooking methods. Atmospheric Environment 42: 8852-8862.
   https://doi.org/10.1016/j.atmosenv.2008.09.011.
- See, S.W., Karthikeyan, S., Balasubramanian, R., 2006. Health risk assessment of occupational
  exposure to particulate-phase polycyclic aromatic hydrocarbons associated with Chinese,
  Malay and Indian cooking. Journal of Environmental Monitoring 8: 369-376.
  https://doi.org/10.1039/B516173H
- Seow, A., Poh, W.T., Teh, M., Eng, P., Wang, Y.T., Tan, W.C., Yu, M.C. Lee, H.P., 2000. Fumes from meat cooking and lung cancer risk in Chinese women. Cancer Epidemiology and Prevention Biomarkers 2000; 9: 1215-1221. https://cebp.aacrjournals.org/content/9/11/1215.
- Shafer, M.M., Hemming, J.D.C., Antkiewicz, D.S., Schauer, J.J., 2016. Oxidative potential of size fractionated atmospheric aerosol in urban and rural sites across Europe. Faraday Discussions
   189: 381-405. https://doi.org/10.1039/C5FD00196J.
- Shang, Y., Fan, L., Feng, J., Lv, S., Wu, M., Li, B., Zang, Y.S., 2013. Genotoxic and inflammatory 765 effects of organic extracts from traffic-related particulate matter in human lung epithelial A549 766 767 cells. The role quinones. Toxicology in Vitro 27: 922-931. of https://doi.org/10.1016/j.tiv.2013.01.008. 768
- Shao, L., Hou, C., Geng, C., Liu, J., Hu, Y., Wang, J., Jones, T., Zhao, C., BéruBé, K., 2016. The oxidative potential of PM10 from coal, briquettes and wood charcoal burnt in an experimental

- 771domesticstove.AtmosphericEnvironment127:372-381.772https://doi.org/10.1016/j.atmosenv.2015.12.007.
- Shao, L., Hu, Y., Shen, R., Schäfer, K., Wang, J., Wang, J., Schnelle-Kreis, J., Zimmermann, R.,
  BéruBé, K., Suppan, P., 2017. Seasonal variation of particle-induced oxidative potential of
  airborne particulate matter in Beijing. Science of The Total Environment 579: 1152-1160.
  https://doi.org/10.1016/j.scitotenv.2016.11.094.
- Shao, L., Shi, Z., Jones, T.P., Li, J., Whittaker, A.G., Berube, K.A., 2006. Bioreactivity of particulate
   matter in Beijing air: results from plasmid DNA assay. Science of the Total Environment 367:
   261-272. https://doi.org/10.1016/j.scitotenv.2005.10.009.
- Sheng, K., Lu, J., 2017. Typical airborne quinones modulate oxidative stress and cytokine expression
  in lung epithelial A549 cells. Journal of Environmental Science and Health, Part A 52: 127134. https://doi.org/10.1080/10934529.2016.1237127.
- Shields, P.G., Xu, G.X., Blot, W.J., Fraumeni Jr, J.F., Trivers, G.E., Pellizzari, E.D., Qu, Y.H., Gao,
  Y.T., Harris, C.C., 1995. Mutagens from heated Chinese and US cooking oils. Journal of the
  National Cancer Institute 87: 836-841. https://doi.org/10.1093/jnci/87.11.836.
- Sovadinová, I., Bláha, L., Janošek, J., Hilcherová, K., Giessy, J.P., Jones, P.D., Holoubek, I., 2006.
   Cytotoxicity and aryl hydrocarbon receptor-mediated activity of N-heterocyclic polycyclic aromatic hydrocarbons: structure-activity relationships. Environmental Toxicology and Chemistry 25, 1291-1297. https://doi.org/10.1897/05-388R.1.
- Sun, J., Shen, Z., Niu, X., Yu, J., Zhang, Y., Liu, S., Niu, X., Zhang, Y., Xu, H., Li, X.J.A.E., 2020.
  PM2. 5 source profiles from typical Chinese commercial cooking activities in northern China and its influences on bioreactivity of vascular smooth muscle cells (VSMCs). Environmental Pollution 239: 117750.
- Svendsen, K., Jensen, H.N., Sivertsen, I., Sjaastad, A.K., 2002. Exposure to cooking fumes in restaurant
   kitchens in Norway. Annals of Occupational Hygiene 46: 395-400.
- Szterk, A., 2015. Acridine derivatives (PANH, azaarenes) in raw, fried or grilled pork from different
   origins, and PANH formation during pork thermal processing. Journal of Food Composition
   and Analysis 43, 18-24. https://doi.org/10.1016/j.jfca.2015.04.011.
- Tanaka, N., Ohtake, K., Tsuzaki, M., Miyazaki, A., 2012. Analysis of polycyclic aromatic hydrocarbons
  in oil-mist emitted from food grilling. Bunseki Kagaku 2012; 61: 77-86.
  https://doi.org/10.2116/bunsekikagaku.61.77.
- Taner, S., Pekey, B., Pekey, H., 2013. Fine particulate matter in the indoor air of barbeque restaurants:
   Elemental compositions, sources and health risks. Science of the Total Environment 454: 79 804 87. https://doi.org/10.1016/j.scitotenv.2013.03.018
- Torkmahalleh, M.A., Zhao, Y., Hopke, P.K., Rossner, A., Ferro, A.R., 2013. Additive impacts on particle emissions from heating low emitting cooking oils. Atmospheric environment 74: 194-198. https://doi.org/10.1016/j.atmosenv.2013.03.038.
- Tuet, W.Y., Liu, F., de Oliveira Alves, N., Fok, S., Artaxo, P., Vasconcellos, P., Champion, J.A., Ng,
  N.L., 2019. Chemical oxidative potential and cellular oxidative stress from open biomass
  burning aerosol. Environmental Science and Technology Letters 6: 126-132.
  https://doi.org/10.1021/acs.estlett.9b00060.
- Van Winkle, L.S., Bein, K., Anderson, D., Pinkerton, K.E., Tablin, F., Wilson, D., Wilson, D., Wexler,
  A.S., 2014. Biological dose response to PM2. 5: effect of particle extraction method on platelet
  and lung responses. Toxicological Sciences 143: 349-359.
  https://doi.org/10.1093/toxsci/kfu230.
- Verma, V., Pakbin, P., Cheung, K.L., Cho, A.K., Schauer, J.J., Shafer, M.M., Kleinman, M.T., Sioutas,
  C., 2011. Physicochemical and oxidative characteristics of semi-volatile components of quasiultrafine particles in an urban atmosphere. Atmospheric Environment 45: 1025-1033.
  https://doi.org/10.1016/j.atmosenv.2010.10.044.
- Wang, G., Cheng, S., Wei, W., Wen, W., Wang, X., Yao, S., 2015. Chemical characteristics of fine particles emitted from different Chinese cooking styles. Aerosol Air Qual. Res 15: 2357-2366.
  https://doi.org/10.4209/aaqr.2015.02.0079.
- Wang, L., Xiang, Z., Stevanovic, S., Ristovski, Z., Salimi, F., Gao, J., Wang, H., Li, L., 2017. Role of Chinese cooking emissions on ambient air quality and human health. Science of the Total Environment 589: 173-181. https://doi.org/10.1016/j.scitotenv.2017.02.124.

- Wei, Y., Han, I.-K., Shao, M., Hu, M., Zhang, J., Tang, X., 2009. PM2.5 constituents and oxidative
  DNA damage in Humans. Environmental Science and Technology 43, 4757-4762. https://doi.org/10.1021/es803337c.
- Wu, P.F., Chiang, T.A., Wang, L.F., Chang, C.S., Ko, Y.C., 1998. Nitro-polycyclic aromatic
  hydrocarbon contents of fumes from heated cooking oils and prevention of mutagenicity by
  catechin. Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis 403: 2934. https://doi.org/10.1016/S0027-5107(98)00015-3.
- Xue, W., Warshawsky, D., 2005. Metabolic activation of polycyclic and heterocyclic aromatic
  hydrocarbons and DNA damage: A review. Toxicology and Applied Pharmacology 206, 7393. https://doi.org/10.1016/j.taap.2004.11.006.
- Yamada, K., Suzuki, T., Kohara, A., Hayashi, M., Mizutani, T., Saeki, K-I., 2004. In vivo mutagenicity
   of benzo[f]quinoline, benzo[h]quinoline, and 1,7-phenanthroline using *lacZ* transgenic mice.
   Mutation Research 559, 83-95 https://doi.org/10.1016/j.mrgentox.2003.12.012.
- Yang, A., Jedynska, A., Hellack, B., Kooter, I., Hoek, G., Brunekreef, B., Kuhlbusch, T.A.J., Cassee,
  F.R., Janssen, N.A.H., 2014. Measurement of the oxidative potential of PM2. 5 and its
  constituents: The effect of extraction solvent and filter type. Atmospheric environment 83: 3542. https://doi.org/10.1016/j.atmosenv.2013.10.049.
- Zhang, J., Smith, K.R., 1999. Emissions of carbonyl compounds from various cookstoves in China.
   Environmental Science & Technology 33: 2311-2320. https://doi.org/10.1021/es9812406.
- Zhang, N., Han, B., He, F., Xu, J., Zhao, R., Zhang, Y., Bai, Z., 2017. Chemical characteristic of PM2.
  5 emission and inhalational carcinogenic risk of domestic Chinese cooking. Environmental
  pollution 227: 24-30. https://doi.org/10.1016/j.envpol.2017.04.033
- Zhao, Y., Hu, M., Slanina, S., Zhang, Y., 2007. Chemical compositions of fine particulate organic
  matter emitted from Chinese cooking. Environmental Science & Technology 41: 99-105.
  https://doi.org/10.1021/es0614518.
- Zhong, L., Goldberg, M.S., Gao, Y-T., Jin, F., 1999a. Lung cancer and indoor air pollution arising from
   Chinese-style cooking among nonsmoking women living in Shanghai, China. Epidemiology
   488-494. https://pubmed.ncbi.nlm.nih.gov/10468420/.
- Zhong, L., Goldberg, M.S., Parent, M-É., Hanley, J.A., 1999b. Risk of developing lung cancer in relation to exposure to fumes from Chinese-style cooking. Scandinavian journal of work, environment & health 309-316. https://www.sjweh.fi/show\_abstract\_php?abstract\_id=440.
- Zhu, L., Wang, J., 2003. Sources and patterns of polycyclic aromatic hydrocarbons pollution in kitchen
   air, China. Chemosphere 50: 611-618. https://doi.org/10.1016/S0045-6535(02)00668-9.
- 859

861

- 863
- 864
- 865
- 866
- 867
- 868

## 869 List of Tables

- Table 1: Characteristic of cooking operations (n = 4 for each type of operation).
- Table 2: Average concentrations  $\pm$  standard deviations of PM<sub>2.5</sub> ( $\mu$ g/m<sup>3</sup>), TC ( $\mu$ g/m<sup>3</sup>), OC
- 872 ( $\mu$ g/m<sup>3</sup>) and EC ( $\mu$ g/m<sup>3</sup>) of inorganic elements (ng/m<sup>3</sup>), water extractable ions ( $\mu$ g/m<sup>3</sup>) in
- 873 cooking emission samples.
- Table 3: The average concentrations  $\pm$  standard deviations (pg/m<sup>3</sup>) of PAHs, OPAHs and AZAs in five sampling locations
- 876

Table 4: Spearman's rank correlation coefficients (R) of the cell viability, ROS generated (DCFH) and inflammatory activity (TNF- $\alpha$  & IL-6), and oxidative DNA damage elicited by extracts and the concentration chemical species

880

## 881 List of Figures and Figure Legends

Figure 1: Mean contributions of individual PAHs, OPAHs, and AZAs to the  $\Sigma$ 30PAHs, 217OPAHs, and  $\Sigma$ 4AZAs concentrations in different sampling locations. The y-axis was broken at 15% to enlarge the scale before the break. Error bars indicate standard deviations for each sample.

- Figure 2: Cell viability, ROS generation (fluorescence intensity) and TNF-α induced by
- extracts of PM<sub>2.5</sub> from five sampling locations (\*p < 0.05). Bars are the mean ±standard
- 888 deviation.

Sample	Sampling location name	Seating capacity (seats)	Peak hour	Fuel type	Cooking - style	Cooking method	Relative humidity <sup>d</sup> (%)	Temperature <sup>d</sup> (°C)
A	A core (theatre lounge)	80	12:30-14:30	Town gas, Electricity	Western cuisine (e.g. pasta, salad and rice)	Baking, Frying (pan frying, stir frying), Grilling, Steaming	77.1	21.6
В	Student canteen (communal student canteen)	520	12:00-14:00	Town gas, Electricity	Mixed cuisine (e.g. siu mei <sup>b</sup> , hamburger and vegetarian diet)	Frying (pan frying, stir frying, deep frying), Roasting, Steaming	63.9	28.4
С	Chinese restaurant (communal student & staff restaurant)	450	12:00-13:00	Town gas, Electricity	Cantonese and local cuisine (noodles, congee, steamed rice and Chinese dim sum <sup>c</sup>	Frying (pan frying, stir frying, deep frying), Steaming	83.2	29.1
D*a	Environmental test chamber	Not available	Not available	Electricity	Stir-fried rice noodle	Stir frying	60.0	27.0
E*a	Environmental test chamber	Not available	Not available	Electricity	Fried chicken	Pan frying	60.0	27.0

#### Table 1: Characteristic of cooking operations (n = 4 for each type of operation). 890

891 892 <sup>a</sup> represents samples collected from stainless steel environmental constant temperature and humidity test chamber that mimic residential kitchen hood condition. <sup>b</sup> Siu mei is the generic name in Cantonese cuisine given to meats roasted on spits over an open fire or a huge wood burning rotisserie oven.

<sup>c</sup> Dim sum is a style of Chinese cuisine prepared as small bite-sized portions of food served in small steamer baskets or on small plates.

893 894 <sup>d</sup> represents the sampling condition of exhaust system in restaurant and the test chamber.

Table 2: Average concentrations  $\pm$  standard deviations of PM<sub>2.5</sub> (µg/m<sup>3</sup>), TC (µg/m<sup>3</sup>), OC (µg/m<sup>3</sup>) and EC (µg/m<sup>3</sup>) of inorganic elements (ng/m<sup>3</sup>), water extractable ions (µg/m<sup>3</sup>) in cooking emission samples. 

Component <sup>a</sup>	Sample A	Sample B	Sample C	Sample D*	Sample E*
PM <sub>2.5</sub>	234.5±22.1	177.4±50.6	711.5±222.6	354.1±60.8	492.0±257.9
TC	113.8±4.2	105.2±32.6	353.1±124.1	217.9±25.5	319.3±164.9
OC	$108.3 \pm 4.2$	99.9±31.5	338.8±121.1	215.0±25.1	315.5±164.4
EC	$5.5 \pm 0.7$	5.3±1.5	$14.4 \pm 4.0$	2.9±0.5	3.7±0.9
Elements <sup>a</sup>					
Na	181.9±21.8	349.8±106.2	1046.7±315.1	189.0±14.4	191.7±33.5
Mg	93.7±76.0	44.6±78.1	156.8±147.5	64.2±111.2	49.0±84.9
Al	73.5±49.5	89.7±65.1	171.7±80.3	84.4±73.5	45.2±48.2
Si	169.9±65.2	163.1±94.3	279.9±60.5	267.4±89.4	312.2±123.1
S	1276.1±676.8	1100.5±262.5	3459.0±796.3	1575.7±915.0	476.5±293.6
Cl	329.9±55.6	616.3±210.4	2882.9±855.0	1078.6±401.9	3497.1±2024.4
Κ	224.5±99.5	258.7±131.4	467.1±139.1	107.4±42.9	47.9±22.4
Ca	88.9±19.9	114.0±74.6	$129.0\pm 50.4$	67.5±40.0	31.7±13.6
Ti	8.0±1.6	$7.2\pm5.8$	5.3±4.6	4.8±3.0	$2.8 \pm 2.8$
V	2.9±2.9	1.7±0.7	$10.3 \pm 10.5$	3.8±3.1	5.3±7.0
Cr	4.0±0.3	4.1±1.1	7.6±1.0	5.4±0.6	$6.9 \pm 4.0$
Mn	14.8±3.6	22.2±9.6	27.4±14.6	13.7±3.5	21.1±5.2
Fe	147.2±20.5	203.4±127.4	330.0±191.5	94.6±27.1	91.4±22.1
Co	$1.6 \pm 1.8$	$1.5 \pm 1.8$	0.3±0.6	$1.4 \pm 1.4$	$0.4\pm0.7$
Ni	$1.6 \pm 1.1$	3.0±2.1	6.6±5.1	3.6±0.7	3.2±2.2
Cu	14.7±4.6	12.3±5.0	28.1±11.4	11.3±3.4	11.4±3.1
Zn	87.1±21.7	99.4±68.9	132.8±60.6	45.0±6.3	74.3±16.7
Sb	16.6±6.1	22.9±7.8	39.4±12.1	33.4±12.5	48.7±30.3
Ba	48.5±8.9	38.8±16.2	144.3±37.3	72.6±18.0	107.9±62.0
Pb	28.5±14.3	20.3±10.5	38.6±4.8	30.3±11.1	36.5±5.2
Ions <sup>a</sup>					
Cl	0.2±0.0	0.4±0.1	3.0±0.8	0.5±0.3	2.7±1.4
NO <sub>3</sub> -	4.8±0.9	2.4±0.7	23.9±8.5	0.3±0.0	$0.4 \pm 0.1$
$SO_4^{2-}$	3.7±1.6	2.2±0.6	$10.0 \pm 2.6$	2.5±1.1	$0.5 \pm 0.5$
Na+	0.3±0.3	0.4±0.1	1.1±0.7	0.6±0.5	0.2±0.2
$\mathrm{NH_4^+}$	1.9±0.5	1.1±0.3	8.0±2.4	0.7±0.4	$0.5 \pm 0.1$
Ca <sup>2+</sup>	0.3±0.0	0.1±0.0	0.3±0.1	0.1±0.1	0.1±0.1

<sup>a</sup>Name of the individual component can be referred to Table S1 (Supplementary Material).

Compound	Sample A	Sample B	Sample C	Sample D*b	Sample E*
PAHs (ng/m <sup>3</sup> )	Sumple II	Sumple B	bumple c	bumpie D	Sumple E
1 2 3 4-Tetrahydronanhthalene	224 4+43 8	135 8+50 6	135 1+158 6	2/12 0+38 0	330 8+1/19 7
Nanhthalene	1049 9+147 9	10217+3171	2983 5+923 7	1694 9+289 8	2363 3+736 2
2-Methylpaphthalene	$212.6 \pm 48.5$	1021.7±517.1	542 7+194 0	3/2 8+55 1	409 1+135 8
1-Methylnaphthalene	212.0140.5	218 1+53 7	617 6+216 5	$412.3 \pm 24.1$	921 2+432 8
Binhanyl	242 3+55 1	201 0+58 4	$681 1 \pm 208 0$	333 3±30 6	477 8+146 0
1.3 Dimethylpophthalene	$527.1 \pm 120.1$	201.01010.4	$0128 \pm 3484$	400 0±30 1	571 4+210 2
A cenanbthylene	1785+421	02 5+53 5	364 2+110 2	$205.0\pm 30.1$	2/13 1+139 2
Acenaphthylene	883 1+204 0	630 5+237 4	966.0+398.4	1335 8+264 2	245.12155.2
Fluorene	217 8+20 3	102 8+55 1	734 5+311 9	346 3+101 4	481 7+290 0
Phenanthrene	$10286 \pm 1440$	603 4+102 9	2150 5+871 9	1187 9+131 1	1615 2+625 7
Anthracene	127 9+23 6	82 0+20 7	273 8+123 6	190 6+99 0	129 8+93 8
4H-Cyclopenta(d e f)phenanthrene	692 6+56 7	550 8+189 7	2662 8+1126 2	847 5+258 8	1125 7+437 3
1-Methylphenanthrene	179 1+56 4	99.0+32.1	481 3+206 6	192 1+32 9	315 9+89 2
3 6-Dimethylnaphthalene	94 3+19 3	63 7+21 5	246 5+100 5	136 9+4 4	251 6+105 8
Fluoranthene	249 4+49 2	285 3+155 6	888 7+271 1	273 9+47 4	296 7+126 8
Pyrene	475 1+114 0	$1190.9 \pm 1105.9$	1917 8+696 3	617 4+86 8	814 9+341 6
Retene	638 3+148 9	439 5+132 3	1185 8+445 5	1175 2+118 9	2210 6+1465 4
Benzo[a]anthracene	66 3+17 7	174 8+68 6	275 2+162 1	248 7+6 8	210.8+70.0
Chrysene <sup>a</sup>	324.9±73.2	212.6±81.3	551.1±176.5	210.1±11.9	238.9±80.5
Benzo[b+i+k]fluoranthenes <sup>b</sup>	625.7+199.0	823 7+363 5	1404 4+541 3	472.1+104.8	611.9+208.3
Benzo[e]pyrene	230.1+78.0	387.0+243.0	645 6+346 5	139.9+46.6	177 4+60 1
Benzo[a]pyrene	94 1+33.8	123 8+67.2	134 5+82.4	135.0+32.0	114 1+35.3
Pervlene	$23.7 \pm 11.8$	$42.1\pm29.0$	$1.8\pm0.7$	$19.4 \pm 12.0$	6.7±2.1
Indeno[1.2.3-cd]pyrene	255.2+43.0	664.0±171.7	972.4±920.1	456.6+253.2	89.8±36.4
Dibenzo[a,h]anthracene	88.4±45.1	98.8±67.8	127.2±62.6	97.7±18.3	$31.2\pm10.3$
Benzo[g.h.i]pervlene	332.0±26.0	832.1±477.7	712.3±355.7	145.5±127.4	655.1±538.9
Coronene	309.0±114.1	675.5±419.6	582.8±373.9	81.7±44.1	81.2±22.3
ΣLMW-PAHs <sup>c</sup>	3485.8±341.3	2622.8±632.7	7472.6±2320.5	4960.8±653.4	6929.3±2174.9
ΣHMW-PAHs <sup>d</sup>	2511.0±435.2	4406.1±1580.9	6983.6±2056.0	2657.1±76.4	3063.4±1079.1
ΣCarci-PAHs <sup>e</sup>	1786.5±297.0	2929.9±930.3	4177.1±1245.3	1765.8±37.1	1951.8±724.5
ΣUS-EPA PAHs	5996.8±752.3	7029.0±2100.8	14456.2±4183.6	7617.9±713.6	9992.7±3253.2
Σ21 Parent-PAHs	6559.6±888.1	8133.6±2608.7	15686.4±4242.0	7858.9±775.0	10258.1±3283.9
Σ30PAHs	9626.7±1078.7	10329.7±3166.4	23452.4±6498.8	12031.9±1219.4	16872.1±5753.7
OPAHs (pg/m <sup>3</sup> )					
1-Indanone	65.3±12.9	68.1±32.8	264.5±136.8	226.9±51.3	642.4±428.9
1,4-Naphthoquinone	23.7±5.9	28.2±14.0	305.9±257.3	116.2±27.4	45.6±26.1
1-Naphthaldehyde	22.1±10.6	$17.5 \pm 8.0$	$95.8 \pm 68.8$	51.8±16.6	$78.9 \pm 40.0$
2-Biphenylcarboxaldehyde	8.1±1.6	7.8±3.4	71.7±39.1	23.9±5.0	236.2±126.3
1-Acenaphthenone	23.5±4.1	$14.0\pm5.7$	$168.1 \pm 84.4$	38.2±5.9	91.7±55.2
9-Fluorenone	$54.6 \pm 7.8$	35.7±14.9	524.7±243.3	112.6±5.1	423.1±264.4
9,10-Anthraquinone	41.5±7.0	38.4±17.8	413.1±165.9	$103.6 \pm 29.7$	261.6±185.6
1,8-Naphtalic anhydride	41.9±2.3	25.9±14.9	216.8±82.6	23.8±6.2	$105.0\pm 66.9$
1,4-Anthraquinone	25.5±5.3	42.2±24.6	217.4±111.4	96.3±28.4	359.6±116.3
4H-Cyclopenta[d,e,f]phenanthrenone	$7.8 \pm 1.5$	$10.2\pm5.9$	$101.3 \pm 46.9$	$12.6\pm2.1$	$38.5 \pm 29.5$
2-Meth-9,10-anthraquinone	47.7±13.5	$26.4 \pm 16.1$	317.1±133.1	$165.7 \pm 18.6$	355.8±243.3
Benzo[a]florenone	$22.5 \pm 4.2$	$16.8 \pm 7.8$	$216.1 \pm 98.8$	$38.0\pm7.1$	84.9±36.2
7H-Benzo[d,e]anthracene-7-one	$17.8 \pm 2.3$	32.1±14.2	259.9±121.6	$22.8 \pm 7.0$	52.4±27.3
Benzo[a]anthracene-7,12-dione	$15.2\pm2.6$	$10.1 \pm 4.2$	112.4±45.5	$12.2\pm2.1$	$25.2\pm8.2$
1,4-Chrysenequinone	140.7±106.6	80.1±60.9	291.1±152.6	$114.9\pm 28.0$	149.3±6.3
5,12-naphthacenequinone	14.7±3.2	8.4±5.3	57.1±24.9	18.6±1.7	105.3±108.8
6H-Benzo[c,d]pyran-6-one	12.5±3.1	40.7±21.7	66.8±42.1	13.2±6.2	58.6±65.1
$\Sigma \Gamma/OPAHs$	585.2±135.6	502.7±249.4	3700.1±1735.3	1191.1±190.8	2944.5±1494.4
AZAs (pg/m <sup>3</sup> )	150.05	141.70	0.0.0.0	#1.0.14 <b>*</b>	00.0.11.0
Quinoline	15.3±3.5	14.1±5.8	97.9±62.9	51.9±14.2	83.0±44.8
Benzo(h)quinoline	5.9±0.7	7.0±2.4	41.9±28.0	28.8±14.5	33.6±21.7
Acridine	3.3±1.5	4.4±3.2	36.0±20.6	18.2±6.0	35.3±24.5
Carbazole	26.8±17.4	7.2±5.8	87.5±52.3	38.9±24.7	49.0±45.5
24AZAS	51.3±19.3	32./±15./	263.3±160.0	137.8±39.3	200.8±133.0

909	Table 3: The average concentrations $\pm$ standard deviations (pg/m <sup>3</sup>	) of PAHs,	OPAHs and AZAs
910	and carbonyls in five sampling locations		

911 <sup>a</sup> represents chrysene + triphenylene; <sup>b</sup> represents benzo[b]fluoranthene + benzo[j]fluoranthene 912 + benzo[k]fluoranthene;. <sup>c</sup> $\Sigma$ LMW-PAHs: low molecular weight PAHs with 2-3 aromatic rings; 913 <sup>d</sup> $\Sigma$ HMW-PAHs: high molecular weight PAHs with 4-7 aromatic rings; <sup>e</sup> $\Sigma$ Carci-PAHs: 914 benzo[a]anthracene + chrysene +benzo[b+j+k]fluoranthenes + benzo[a]pyrene + indeno[1,2,3-

915 cd]pyrene + dibenzo[a,h]anthracene + benzo[g,h,i]perylene.

916 Table 4: Spearman's rank correlation coefficients (R) of the biological responses [cell viability,

917 ROS generated (DCFH), inflammatory activity (TNF- $\alpha$ ), and oxidative DNA damage] elicited

918 by extracts with the concentrations of chemical species.919

		ROS			
Chemical Species	Cell viability	generated (DCFH)	TNF-α	DNA damage	
ТС	-0.102	-0.239	-0.336	0.307	
OC	-0.101	-0.158	-0.423	0.233	
EC	-0.028	-0.502*b	0.824**	0.325	
Cl-	0.327	0.367	-0.386	-0.292	
NO <sub>3</sub> -	0.216	0.151	0.437	-0.239	
SO4 <sup>2-</sup>	-0.148	-0.176	0.671**	-0.046	
Na <sup>+</sup>	-0.346	-0.238	0.389	0.393	
NH4 <sup>+</sup>	0.165	0.158	0.431	-0.345	
Ca <sup>2+</sup>	0.122	-0.427	0.676**	0.279	
Na	-0.332	-0.287	0.799**	0.137	
Mg	-0.056	-0.156	0.231	-0.164	
Al	-0.112	-0.689**	0.624*	0.643**	
Si	-0.150	-0.590*	0.286	0.357	
S	-0.360	-0.227	0.644**	-0.056	
Cl	0.214	0.300	-0.479*	-0.185	
К	-0.252	-0.582*	0.930**	0.274	
Ca	-0.275	-0.711**	0.906**	0.434	
Ti	0.087	-0.742**	0.748**	0.540*	
V	0.191	-0.014	0.035	-0.024	
Cr	-0.250	-0.622**	0.565*	0.570*	
Mn	-0.140	-0.715**	0.641**	0.543*	
Fe	-0.240	-0.592**	0.912**	0.298	
Со	-0.315	-0.202	0.118	-0.068	
Ni	-0.334	-0.359	0.444	0.203	
Cu	-0.082	-0.543*	0.720**	0.287	
Zn	0.076	-0.646**	0.693**	0.430	
Sb	-0.198	-0.575*	0.255	0.427	
Ba	-0.057	0.008	-0.004	0.317	
Pb	-0.017	-0.491*	0.360	0.316	
Hexaldehyde	0.277	0.393	0.106	-0.405	
Heptaldehyde	0.307	0.398	0.116	-0.334	
Octaldehyde	0.186	0.399	0.197	-0.384	
Nonaldehyde	0.342	0.285	0.268	-0.256	
Decaldehyde	0.285	0.595**	-0.329	-0.432	
Glyoxal	-0.085	0.414	-0.382	-0.306	
Methylglyoxal	-0.061	0.379	-0.387	-0.319	
1,2,3,4-Tetrahydronaphthalene	0.177	-0.470 <sup>*</sup>	0.301	0.472*	
Naphthalene	-0.117	-0.397	0.096	0.485*	
2-Methylnaphthalene	-0.293	-0.421	0.244	0.261	
1-Methylnaphthalene	0.327	-0.374	-0.231	0.359	
Biphenyl	0.082	-0.164	0.101	0.025	
1,3-Dimethylnaphthalene	0.051	-0.449	0.550*	0.335	
Acenaphthylene	0.300	-0.102	0.100	0.189	

Acenaphthene	0.193	-0.445	-0.117	0.533*
Fluorene	-0.267	0.034	0.105	0.046
Phenanthrene	0.364	-0.396	0.244	0.366
Anthracene	-0.343	-0.392	0.602**	0.361
Cyclopenta[def]phenanthrene	0.000	0.069	0.297	-0.015
1-Methylphenanthrene	0.463	0.082	-0.091	0.112
3,6-Dimethylphenanthrene	0.451	-0.074	-0.360	0.367
Fluoranthene	-0.212	-0.176	0.683**	0.089
Pyrene	-0.195	-0.041	0.511*	0.060
Retene	0.297	-0.257	-0.446	0.457
Benzo[a]anthracene	-0.602**	-0.368	0.234	0.285
Chrysene + Triphenylene	0.054	-0.593**	0.822**	0.447
Benzo [b+j+k] fluoranthene	-0.153	-0.660**	0.873**	0.474*
Benzo[e]pyrene	-0.210	-0.549*	0.896**	0.346
Benzo[a]pyrene	-0.336	-0.789**	0.580*	0.763**
Perylene	-0.308	-0.888**	0.688**	0.740**
Indeno [1,2,3-cd] pyrene	-0.615**	-0.478*	0.743**	0.248
Dibenzo[ah]anthracene	-0.456	-0.677**	0.818**	0.599**
Benzo[ghi]perylene	0.081	-0.696**	0.660**	0.690**
Coronene	-0.152	-0.587*	0.853**	0.465
Quinoline	0.005	0.378	-0.603**	-0.247
1-Indanone	0.104	0.178	-0.631**	0.024
1,4-Naphthoquinone	-0.665**	0.187	0.082	-0.315
1-Naphthaldehyde	0.073	0.243	-0.426	-0.291
2-Biphenylcarboxaldehyde	0.234	0.594**	-0.796**	-0.339
1-Acenaphthenone	0.435	0.711**	-0.637**	-0.516*
9-Fluorenone	0.409	0.739**	-0.756**	-0.495*
Benzo[h]quinoline	-0.144	0.266	-0.445	-0.219
Acridine	0.044	0.581*	<i>-0.733</i> **	-0.358
Carbazole	0.174	0.424	-0.283	-0.196
9,10-Anthraquinone	0.234	0.661**	-0.577*	<i>-0.471</i> *
1,8-Naphthalic anhydride	$0.524^{*}$	0.446	-0.072	-0.382
1,4-Anthraquinone	0.173	0.335	-0.621**	-0.114
4H-Cyclopenta[def]phenanthrenone	0.046	0.444	-0.133	-0.314
2-Methylanthracene-9,10-dione	0.187	0.686**	-0.884**	-0.418
Benzo[a]fluorenone	0.359	0.719**	-0.591**	-0.432
7H-Benz[de]anthracene-7-one	0.089	0.281	0.075	-0.184
Benz[a]anthracene-7,12-dione	0.347	0.328	0.164	-0.280
1,4-Chrysenequinone	0.320	0.011	-0.208	0.149
Naphthacene-5,12-dione	$0.665^{**}$	0.415	<i>-0.541</i> *	-0.232
6H-benzo[cd]pyrene-6-one	-0.136	-0.256	0.439	0.171
Σ7Carbonyls	0.233	0.348	0.239	-0.293
Σ30PAHs	-0.048	-0.876**	0.745**	0.763**
Σ17OPAHs	0.371	0.474*	-0.617**	-0.223
Σ4AZAs	0.067	0.419	-0.558*	-0.124

920 <sup>a\*\*</sup>Correlation is significant at the 0.01 level (2-tailed).
 921 <sup>b\*</sup> Correlation is significant at the 0.05 level (2-tailed).



924 925

- Figure each sample. broken at 15% to enlarge the scale before the break. Error bars indicate standard deviations for  $\Sigma 15 OPAHs$ , and 1: Mean contributions Σ4AZAs concentrations in different sampling locations. of individual PAHs, OPAHs, and AZAs 5 The y-axis the Σ30PAHs, was
- 928



929

930 Figure 2: Cell viability, ROS generation (fluorescence intensity) and TNF- $\alpha$  induced by

931 extracts of  $PM_{2.5}$  from five sampling locations (\*p < 0.05). Bars are the mean ±standard 932 deviation

933