

We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

5,600

Open access books available

137,000

International authors and editors

170M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.
For more information visit www.intechopen.com



Metabotropic Receptors 4 and the Immune Responses

Zhuoya Wan and Song Li

Abstract

Neurotransmitters (NTs) have recently received increasing appreciation as important immune modulators. The immune cells express receptors for many classes of NTs and the communication between NTs and their receptors establish neuro-immune interactions for regulating effective immune response in both central nervous system (CNS) and peripheral tissues. Metabotropic Glutamate Receptor 4 (mGluR4) is expressed at high level in CNS and plays a role in various physiological and pathophysiological processes in CNS. Recently, mGluR4 has been reported to be expressed on immune cells and have an impact on regulating the immune system. This chapter summarized the works associated with the immunogenic function of mGluR4 and its potential underlying mechanisms.

Keywords: metabotropic glutamate receptor (mGluR4), immune response, peripheral tissues, central nervous system (CNS), cancer, autoimmune diseases

1. Introduction

Neurotransmitters (NTs) have recently received increasing appreciation as important immune modulators. The immune cells express receptors for many classes of NTs and the communication between NTs and their receptors establish neuro-immune interactions for regulating effective immune response in both CNS and peripheral tissues [1]. Interestingly, the role of NTs is very complicated and the same NTs can even exert opposing effects for promoting or inhibiting tissue immunity in different contexts [2–6].

Studies of the NTs and their receptors in modulating immunity are limited and therein are important areas of investigations. L-Glutamate (Glu) is the major excitatory neurotransmitter in the mammalian CNS [7]. It acts via two classes of receptors, ligand gated ion channels (ionotropic receptors (iGluRs))-regulating rapid responses upon activation, and G-protein coupled (metabotropic) receptors-modulating signal transduction cascades. Eight different types of mGluRs, mGluR1 to mGluR8 are divided into groups I, II, and III on the basis of their intracellular signal transduction mechanisms, agonist pharmacology, and sequence homologies (see **Figure 1**) [8]. Group I includes mGluR1 and mGluR5, coupled to Gq protein; group II includes mGluR2 and mGluR3, coupled to Gi and Go proteins; group III includes mGluR4, mGluR6, mGluR7 and mGluR8, coupled to Gi and Go proteins in heterologous expression systems.

mGluR4 is expressed at high levels in CNS and plays a role in various physiological and pathophysiological processes in CNS [9, 10], such as learning, memory, and cognitive impairment. In addition, growing evidence indicates that mGluRs

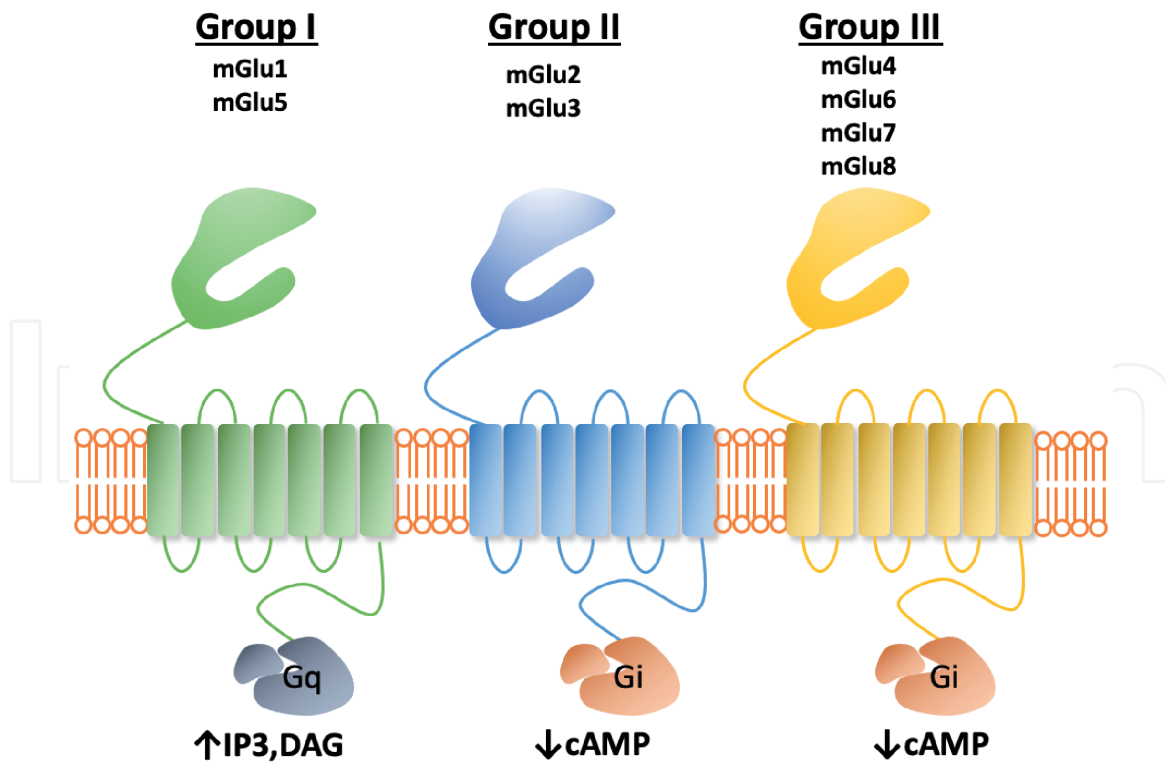


Figure 1. The summary of mGluRs families. mGluRs are classified into three families: group I, group II, and group III. In the CNS, activation of mGluRs from group I leads to the induction of phosphoinositide hydrolysis with formation of inositol 1,4,5-triphosphate (IP₃) and diacylglycerol (DAG). The activation of groups II and III receptors induce a decrease on the intracellular levels of cyclic adenosine monophosphate (cAMP) [7].

are expressed in the peripheral such as thymus and lymphocytes [11]. These results suggest a potential role of mGluR4 in immune regulation. In this chapter, we summarized the association of mGluR4 with immune responses and its role in different diseases. The potential of mGluR4 as a novel therapeutic target in immune-related diseases was also discussed.

2. Expression of mGluR4 on immune cells

Clinical data indicated that elevated plasma concentrations of Glu are associated with immune deficiency [12, 13]. In addition, in vitro assays showed that high concentration of Glu (>100 uM) can inhibit mitogen-induced T-cell proliferation [12, 13]. Therefore, it is not surprising that immune cells express mGluRs. It has been proposed that mGluRs can mediate an emergency mechanism once high levels of Glu are reached.

Using immunostaining and Western blot analysis, Rezzani et al. observed the expression of mGluR4 in rat thymic cells [14]. The expression of mGluR4 was abundant in dendritic cells (DCs) and lymphocytes of the thymic medulla but was weak in lymphocytes of the cortex. It is interesting to note that a rapid inhibition on the expression of mGluR4 was induced in the rat thymus after treatment with cyclosporine (an immunosuppressant). The mGluR4 expression reached undetectable levels after a longer treatment regimen of cyclosporine.

Other evidence also pointed out that the expression of mGluRs is not exclusive to young immune cells because mature lymphocytes are activated by selective mGluRs ligands. In addition, rat peripheral lymphocytes responded by producing reactive oxygen species (ROS) when they were exposed to the group III mGluRs

agonist L-2-amino-4-phosphonobutyric acid (L-AP4) [15]. ROS play important roles in T-cell biology and participate in activation-induced T cell apoptosis and hence in the termination of the immune response [16]. Moreover, DCs are capable of secreting glutamate when interacting with T lymphocytes, a process that might be essential for the function of lymphocyte. This hypothesis is based on the fact that the absence of glutamate led to impaired Th1 (Interleukin-2 (IL-2) and interferon- γ) and proinflammatory (IL-6 and tumor necrosis factor-alpha) cytokine production. However, these changes were not correlated with a decrease in T-cell proliferation.

3. mGluR4 and Autoimmunity in CNS

A role of mGluR4 in immune modulation was first described in an autoimmune disease model [17]. Fallarino et al. [17] reported that mGlu4 knockout mice (*Grm4*^{-/-}) were highly susceptible to experimental autoimmune encephalomyelitis (EAE), a model of multiple sclerosis. More specifically, *Grm4*^{-/-} mice and their wild-type (WT) counterparts were immunized with myelin oligodendrocyte glycoprotein (MOG35–55), which can induce EAE in C57BL/6 mice. The EAE clinical scores were recorded periodically and a lack of mGluR4 was found to be associated with earlier onset, more severe, and ultimately fatal EAE in >40% of the hosts. Along with these changes, white matter demyelination and inflammatory infiltrates were more prevalent in the spinal cord of MOG-vaccinated *Grm4*^{-/-} mice in comparison to their WT counterparts, according to the morphological changes. The phenotype has also been characterized in littermates as well (heterozygote breeding—with cohorts of mice being matched for gender and age) and the disease indications were also more severe in *Grm4*^{-/-} and *Grm4*^{+/-} mice than in WT mice. In contrast, treatment of N-Phenyl-7-(hydroxyimino) cyclopropa [b] chromen-1a-carboxamide (PHCCC), an *Grm4*-positive allosteric modulator led to increased resistance to EAE. This was in agreement with previous reports demonstrating that long-term treatment of L-AP4 can increase the recovery rate from EAE in Lewis rats [17, 18].

There was significant infiltration of CD4⁺ T cells, CD8⁺ T cells and B220⁺ B cells in both peripheral lymphoid organs and the CNS in both *Grm4*^{-/-} and WT mice, but the percentages of CD4⁺ and CD8⁺ T cells as well as CD11b⁺ and CD11c⁺ cells were significantly higher in the CNS of *Grm4*^{-/-} mice at the peak of disease [17]. Extended studies using littermates from heterozygote breeding further showed that the disease course was more severe in *Grm4*^{-/-} and *Grm4*^{+/-} mice than in WT mice. The cytokine profiling of sorted CD4⁺ T cells from brain-infiltrating leukocytes (BILs) and pooled lymph nodes demonstrated a significant increase in *Rorc* transcripts (encoding the T_H17 specification factor), a reduction in *Foxp3* (Treg) transcripts, and no change in *Tbx21* (coding for Tbet; a T_H1 maker) in *Grm4*^{-/-} mice during the neurologic signs. No changes were observed in *Gata3* (a T_H2 marker) in both groups. These data suggested that *Grm4*^{-/-} tipped the balance of transcriptional activation in favor of inflammatory genes in response to MOG vaccination. In particular, *Grm4*^{-/-} favored the emergence of T_H17 over Treg cells, which would sustain inflammation and exacerbate EAE [18].

Expression of mGluR4 was confirmed in several immune subpopulations, such as CD4⁺ T cells, CD8⁺ T cells, $\gamma\delta$ T cells, B220⁺ B cells, CD11b⁺ and CD11c⁺ cells, particularly in CD4⁺ T and CD11c⁺ cells, suggesting those cells as potential targets for *Grm4* mediated effects. The expression of mGluR4 was also confirmed in DC subsets in splenocytes, including conventional DCs (cDCs; CD11b⁺CD11c^{high}) and plasmacytoid DCs (pDCs; mPDCA1⁺ CD11c^{low}). They have further shown that

treatment with toll-like receptor ligands such as lipopolysaccharide (LPS) and CpG-oligonucleotide (CpG-ODN) led to increased Grm4 expression. Modulation of mGluR4 expression in activated nTreg cells (CD4⁺ CD25⁺) and LPS-stimulated cDCs was also confirmed, further supporting that mGluR4 activation within an immunologic synapse contributes to the crosstalk and reciprocal influence between T and accessory cells [17].

IL-17-producing T helper (Th17) cells are considered mediators of autoimmunity in multiple sclerosis and EAE. The accumulation of Th17 cells in the CNS as well as in the periphery is also associated with the development of demyelinating plaques of multiple sclerosis [19]. Fallarino et al. [17] also pointed out that the absence of mGlu4 in dendritic cells is key to inducing a differentiation of T helper cells toward the Th17 phenotype. More specifically, possible regulatory function of mGluR4 in the interaction between CD4⁺ T cells and DCs has been examined. Both cDCs and pDCs from Grm4^{-/-} mice produced higher amounts of IL-6 and IL-23, but less IL-12 and IL-27, compared to their WT counterparts in response to LPS or CpG-ODN, respectively [17].

The notable results of coculturing of WT CD4⁺ T cell and Grm4^{-/-} DCs demonstrated an increase of IL-17A⁺ CD4⁺ T cells, along with a significantly reduction of IFN- γ producing CD4⁺ T cells (a portion of which also expressed IL-17A). However, they failed to see this effect when the coculture consisted of WT DCs and Grm4^{-/-} T cells, suggesting that the effect of mGluR4 depletion was largely dependent on DCs in this *in vitro* system. The cytokine production in culture supernatants has been examined and there are decreased amounts of T_H1-associated IL-2 in coculture system involving Grm4^{-/-} cDCs. IL-27 is known to counter the effect of IL-6 in directing TH17 cell development, which can limit the EAE progression. The decrease in IL-27 during activation of naïve CD4⁺ T cells might be another reason for favoring the emergence of Th17 cells [17].

They also suggest that activation of mGlu4 (as a result of elevated levels of glutamate during the neuroinflammation) might exert a protective effect by preventing the unbalance in T helper cells. Such mechanism presents a clear therapeutic potential for treating autoimmune related disorders.

The underlying mechanism for Grm4-mediated immune regulation is not clear at present. However, there appears to be a cross-talk and reciprocal influences between Grm4 and indoleamine 2,3-dioxygenase 1 (IDO-1) pathways [20]. IDO1 has been well known to be involved in generating an immunosuppressive environment through catalyzing the metabolism of tryptophan, resulting in tryptophan depletion and accumulation of kynurenine [21]. A protective role of IDO-1 has been shown in mice with different forms of EAE including acute, relapsing–remitting, and adoptively transferred disease [22]. Interestingly, in addition to the direct immunosuppressive effect of kynurenine through inhibition of CD8⁺ T cells and activation of Treg cells, kynurenine metabolites such as cinnabarinic acid (CA) act as selective, although weak, orthosteric agonists of mGluR4 [23]. The therapeutic effect of CA in acute EAE was attenuated in Grm4^{-/-} mice [24]. On the other hand, activation of Grm4 could positively impact the IDO1 pathway. Treatment of DCs with ADX88178, a positive allosteric modulator (PAM) of Grm4, led to both increased expression levels of IDO-1 and phosphorylation of IDO-1 [20]. These effects require a Gi-independent, alternative signaling pathway that involves phosphatidylinositol-3-kinase (PI3K), Src kinase, and the signaling activity of IDO1. Moreover, the effect of ADX88178 on the expression of several cytokines was impaired in IDO1^{-/-} DCs [20]. Therefore, Grm4 and IDO1 constitute a loop that provides a positive feedback mechanism to amplify the immune-protective effect in EAE and possibly other immune-related diseases [20].

4. mGluR4 and cancers

Most studies on the role of glutamate receptor in cancers have been focused on iGluRs [25, 26]. Tumor cells originated from neuronal tissues express iGluRs subunits and iGluRs antagonists have shown inhibitory effect on the proliferation of the tumor cells. Similarly, iGluRs subunits have been shown to be expressed in several peripheral cancers and blockade of the N-methyl-D-aspartate (NMDA) and α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) ionotropic glutamate receptor subtypes leads to decreased cell proliferation and migration [26].

mGluRs are also expressed in several cell lines derived from human tumors, including neuroblastoma, thyroid carcinoma, rhabdomyosarcoma/medulloblastoma, lung carcinoma, multiple myeloma, glioma, colon adenocarcinoma, astrocytoma, T cell leukemia, and breast carcinoma [27]. In particular, mGluR1 has been shown to be expressed in subsets of human melanomas [28]. Ectopic expression of mGluR1 in melanocytes drives the development of melanoma in mouse models. Pharmacological inhibition of mGluR1 led to inhibition of tumor cell growth both in vitro and in vivo [28]. Riluzole, an antagonist of mGluR1 signaling has advanced to phase II clinical trial in patients with advanced melanoma [29, 30].

The studies on the roles of mGluR4 in cancers are very limited and controversial. Change et al. studied the expression pattern of mGluR4 in several healthy and diseases-derived human tissues [31]. mGluR4 receptor expression was identified in 68% of colorectal carcinomas, 50% of laryngeal carcinoma, and 46% breast carcinomas. In the case of colorectal carcinoma, overexpression of mGluR4 was correlated with poor prognosis, and cell lines derived from human colorectal carcinomas showed increased cell invasiveness when treated with L-AP4. In another study, comparative proteomics was used to characterize a human colon cancer cell line that was resistant to 5-fluorouracil (5-FU, a common chemotherapy agent). Interestingly, 5-FU resistant cells were found to overexpress mGluR4 in comparison with parental cancer cells. It has been demonstrated that cell survival was increased by the group III mGlu receptor agonist L-AP4 in the nonresistant parent cancer cells; conversely, survival was synergistically decreased by 5-FU and the group III receptor antagonist MAP4 in 5-FU-resistant cells. It is noteworthy to mention that 5-FU downregulated mGluR4 expression, and MAP4 has a dose dependent cytotoxic effect in both cell lines [32].

In contrast to the above reports, mGluR4 agonists are shown to inhibit the proliferation of human breast and bladder cancer cells in a GRM4-dependent manner [33]. In the study by Lasek et al., the expression of mGlu4 was shown to be inversely correlated with the severity of human medulloblastoma [34]. After scoring the extent of immunoreactivity for mGlu4 in human biopsies of medulloblastoma, the absence of spinal metastases, cerebrospinal fluid spread, and tumor recurrence as well as the survival of patients were all shown to be associated with high levels of mGlu4 immunoreactivity. Treatment with PHCCC (which is considered a group I mGlu receptor antagonist but can also act as a positive allosteric modulator of mGlu4 receptor) reduced the proliferation of cultured medulloblastoma cells and inhibited the growth of medulloblastoma implants in mice. In addition, subcutaneous or intracranial injections of PHCCC during the first week of life reduced the incidence of medulloblastoma from 85 to 28% in a mutant mouse model known to develop the disease upon X-ray irradiation. This indicates that activation of mGlu4 receptors also affects early events in tumorigenesis [35].

The above studies focus on the role of tumor cell-derived Grm4. It has been reported that the plasma levels of Glu are generally elevated in patients with carcinoma and seem to correlate with an impairment in immune function [36]. However,

the role of immune cell-derived mGluR including mGluR4 has hardly been studied. Kansara et al. reported that Grm4^{-/-} mice showed accelerated radiation-induced tumor development in an irradiation-induced osteosarcoma model [37]. Outside the CNS, mGluR4 is highly expressed by DCs, as well as CD4⁺ T cells [17]. In the mouse osteosarcomas, they found that mGluR4 is predominantly expressed by CD45⁺CD11c⁺MHC⁺ myeloid cells within the tumor microenvironment (TME) instead of tumor cells. Few CD4⁺ T cells were detectable to characterize mGluR4 expression. In consistent with the study by Fallarino et al. [17] in an EAE model, Grm4^{-/-} DCs isolated from the tumors showed increased expression of IL-23. Interestingly, high expression of IL-23 has been observed in primary osteosarcomas and allografted cell lines relative to normal bone, while ex-vivo cultured osteosarcoma cell lines and primary tumor cells did not express IL23. A role of IL-23 in tumorigenesis has been well established from previous studies [38]. Indeed, IL23^{-/-} mice were resistant to the irradiation-induced osteosarcoma. They hypothesized that knockout of Grm4 in DCs facilitates the oncogenesis of osteosarcoma through increased production of IL-23 [37].

We have recently shown in three murine syngeneic tumor models (B16, MC38, and 3LL) that either genetic knockout (Grm4^{-/-}) or pharmacological inhibition led to significant delay in tumor growth (Wan et al., unpublished data). Mechanistically, perturbation of GRM4 resulted in a strong anti-tumor immunity by promoting nature killer (NK), CD4⁺ and CD8⁺ T cells toward an activated, proliferative, and functional phenotype. We have further shown that the antitumor activity of Grm4 antagonists can be further improved through combination with anti-PD-1 antibody. The differing role of Grm4 in different tumor models may reflect the complex functions of Grm4 in different tumor environment. More studies are needed to further define the roles of immune cells-derived Grm4 and its potential as a novel therapeutic target for cancer immunotherapy.

5. Conclusions

Although the neurological function of GRM4 in CNS has been well established, its role in modulating immune response just began to be appreciated. GRM4 is expressed in various immune cells and loss of GRM4 function in immune cells led to sensitization to EAE. GRM4 selective agonists may hold potential as a novel therapy for autoimmune disorders of CNS. GRM4 is also expressed in various cancer cells, however, conflicting results have been reported regarding whether GRM4 promotes or inhibits tumor cell proliferation. The role of immune cells-derived GRM4 in antitumor immunity is also controversial and may reflect the complex function of GRM4 in different tumor microenvironment. Further studies using more defined animal models and selective GRM4 modulators may not only advance our understanding of the complex immunobiology of GRM4 but also lead to the development of a new immunotherapy for the treatment of cancer.

Acknowledgements

This work was supported, in part, by NIH grant RO1 CA219399.

Conflict of interest

The authors declare no conflict of interest.

IntechOpen

Author details


Zhuoya Wan^{1,2} and Song Li^{1,2*}

1 Center for Pharmacogenetics, School of Pharmacy, University of Pittsburgh, Pittsburgh, PA, USA

2 Department of Pharmaceutical Sciences, School of Pharmacy, University of Pittsburgh, Pittsburgh, PA, USA

*Address all correspondence to: sol4@pitt.edu

IntechOpen

© 2021 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/3.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. 

References

- [1] C. Chu, D. Artis, I.M. Chiu, Neuro-immune interactions in the tissues, *Immunity* 52(3) (2020) 464-474.
- [2] C. Godinho-Silva, R.G. Domingues, M. Rendas, B. Raposo, H. Ribeiro, J.A. da Silva, A. Vieira, R.M. Costa, N.L. Barbosa-Morais, T. Carvalho, Light-entrained and brain-tuned circadian circuits regulate ILC3s and gut homeostasis, *Nature* 574(7777) (2019) 254-258.
- [3] P. Baral, B.D. Umans, L. Li, A. Wallrapp, M. Bist, T. Kirschbaum, Y. Wei, Y. Zhou, V.K. Kuchroo, P.R. Burkett, Nociceptor sensory neurons suppress neutrophil and $\gamma\delta$ T cell responses in bacterial lung infections and lethal pneumonia, *Nature medicine* 24(4) (2018) 417.
- [4] J.R. Huh, H. Veiga-Fernandes, Neuroimmune circuits in inter-organ communication, *Nature Reviews Immunology* 20(4) (2020) 217-228.
- [5] C.S. Klose, D. Artis, Neuronal regulation of innate lymphoid cells, *Current opinion in immunology* 56 (2019) 94-99.
- [6] H. Veiga-Fernandes, D. Mucida, Neuro-immune interactions at barrier surfaces, *Cell* 165(4) (2016) 801-811.
- [7] M. Julio-Pieper, P.J. Flor, T.G. Dinan, J.F. Cryan, Exciting times beyond the brain: metabotropic glutamate receptors in peripheral and non-neural tissues, *Pharmacological reviews* 63(1) (2011) 35-58.
- [8] J.F. Cryan, K.K. Dev, The glutamatergic system as a potential therapeutic target for the treatment of anxiety disorders, *Handbook of behavioral neuroscience* 17 (2008) 269-301.
- [9] P.J. Conn, Physiological roles and therapeutic potential of metabotropic glutamate receptors, *Annals of the New York Academy of Sciences* 1003(1) (2003) 12-21.
- [10] R. Gerlai, J.C. Roder, D.R. Hampson, Altered spatial learning and memory in mice lacking the mGluR4 subtype of metabotropic glutamate receptor, *Behavioral neuroscience* 112(3) (1998) 525.
- [11] R. Pacheco, H. Oliva, J.M. Martinez-Navío, N. Climent, F. Ciruela, J.M. Gatell, T. Gallart, J. Mallol, C. Lluís, R. Franco, Glutamate released by dendritic cells as a novel modulator of T cell activation, *The Journal of Immunology* 177(10) (2006) 6695-6704.
- [12] C. Ferrarese, A. Aliprandi, L. Tremolizzo, L. Stanzani, A. De Micheli, A. Dolara, L. Frattola, Increased glutamate in CSF and plasma of patients with HIV dementia, *Neurology* 57(4) (2001) 671-675.
- [13] G. Lombardi, C. Dianzani, G. Miglio, P.L. Canonico, R. Fantozzi, Characterization of ionotropic glutamate receptors in human lymphocytes, *British journal of pharmacology* 133(6) (2001) 936-944.
- [14] R. Rezzani, G. Corsetti, L. Rodella, P. Angoscini, C. Lonati, R. Bianchi, Cyclosporine-A treatment inhibits the expression of metabotropic glutamate receptors in rat thymus, *Acta histochemica* 105(1) (2003) 81-87.
- [15] A.A. Boldyrev, V.I. Kazey, T.A. Leinsoo, A.P. Mashkina, O.V. Tyulina, P. Johnson, J.O. Tuneva, S. Chittur, D.O. Carpenter, Rodent lymphocytes express functionally active glutamate receptors, *Biochemical and biophysical research communications* 324(1) (2004) 133-139.
- [16] D.A. Hildeman, T. Mitchell, T.K. Teague, P. Henson, B.J. Day, J. Kappler, P.C. Marrack, Reactive oxygen species

regulate activation-induced T cell apoptosis, *Immunity* 10(6) (1999) 735-744.

[17] F. Fallarino, C. Volpi, F. Fazio, S. Notartomaso, C. Vacca, C. Busceti, S. Bicciato, G. Battaglia, V. Bruno, P. Puccetti, Metabotropic glutamate receptor-4 modulates adaptive immunity and restrains neuroinflammation, *Nature medicine* 16(8) (2010) 897-902.

[18] G. Besong, G. Battaglia, M. D'Onofrio, R. Di Marco, R.T. Ngomba, M. Storto, M. Castiglione, K. Mangano, C.L. Busceti, F.R. Nicoletti, Activation of group III metabotropic glutamate receptors inhibits the production of RANTES in glial cell cultures, *Journal of Neuroscience* 22(13) (2002) 5403-5411.

[19] J. Milovanovic, A. Arsenijevic, B. Stojanovic, T. Kanjevac, D. Arsenijevic, G. Radosavljevic, M. Milovanovic, N. Arsenijevic, Interleukin-17 in chronic inflammatory neurological diseases, *Frontiers in Immunology* 11 (2020) 947.

[20] C. Volpi, G. Mondanelli, M.T. Pallotta, C. Vacca, A. Iacono, M. Gargaro, E. Albini, R. Bianchi, M.L. Belladonna, S. Celanire, Allosteric modulation of metabotropic glutamate receptor 4 activates IDO1-dependent, immunoregulatory signaling in dendritic cells, *Neuropharmacology* 102 (2016) 59-71.

[21] Z. Wan, J. Sun, J. Xu, P. Moharil, J. Chen, J. Xu, J. Zhu, J. Li, Y. Huang, P. Xu, Dual functional immunostimulatory polymeric prodrug carrier with pendent indoximod for enhanced cancer immunochemotherapy, *Acta biomaterialia* 90 (2019) 300-313.

[22] Y. Yan, G.-X. Zhang, B. Gran, F. Fallarino, S. Yu, H. Li, M.L. Cullimore, A. Rostami, H. Xu, IDO upregulates regulatory T cells via tryptophan catabolite and suppresses encephalitogenic T cell responses in experimental autoimmune

encephalomyelitis, *The Journal of Immunology* 185(10) (2010) 5953-5961.

[23] F. Fazio, C. Zappulla, S. Notartomaso, C. Busceti, A. Bessedè, P. Scarselli, C. Vacca, M. Gargaro, C. Volpi, M. Allegrucci, Cinnabarinic acid, an endogenous agonist of type-4 metabotropic glutamate receptor, suppresses experimental autoimmune encephalomyelitis in mice, *Neuropharmacology* 81 (2014) 237-243.

[24] F. Fazio, L. Lionetto, G. Molinaro, H.-O. Bertrand, F. Acher, R.T. Ngomba, S. Notartomaso, M. Curini, O. Rosati, P. Scarselli, Cinnabarinic acid, an endogenous metabolite of the kynurenine pathway, activates type 4 metabotropic glutamate receptors, *Molecular pharmacology* 81(5) (2012) 643-656.

[25] M.P. Ribeiro, J.B. Custódio, A.E. Santos, Ionotropic glutamate receptor antagonists and cancer therapy: time to think out of the box?, *Cancer chemotherapy and pharmacology* 79(2) (2017) 219-225.

[26] A. Stepulak, R. Rola, K. Polberg, C. Ikonomidou, Glutamate and its receptors in cancer, *Journal of neural transmission* 121(8) (2014) 933-944.

[27] A. Stepulak, H. Luksch, C. Gebhardt, O. Uckermann, J. Marzahn, M. Sifringer, W. Rzeski, C. Staufner, K.S. Brocke, L. Turski, Expression of glutamate receptor subunits in human cancers, *Histochemistry and cell biology* 132(4) (2009) 435-445.

[28] Y. Ohtani, T. Harada, Y. Funasaka, K. Nakao, C. Takahara, M. Abdel-Daim, N. Sakai, N. Saito, C. Nishigori, A. Aiba, Metabotropic glutamate receptor subtype-1 is essential for in vivo growth of melanoma, *Oncogene* 27(57) (2008) 7162-7170.

[29] C.L. Speyer, M.A. Nassar, A.H. Hachem, M.A. Bukhsh, W.S. Jafry, R.M.

Khansa, D.H. Gorski, Riluzole mediates anti-tumor properties in breast cancer cells independent of metabotropic glutamate receptor-1, *Breast cancer research and treatment* 157(2) (2016) 217-228.

[30] J.M. Mehnert, A.W. Silk, J. Lee, L. Dudek, B.S. Jeong, J. Li, J.M. Schenkel, E. Sadimin, M. Kane, H. Lin, A phase II trial of riluzole, an antagonist of metabotropic glutamate receptor 1 (GRM 1) signaling, in patients with advanced melanoma, *Pigment cell & melanoma research* 31(4) (2018) 534-540.

[31] H.J. Chang, B.C. Yoo, S.-B. Lim, S.-Y. Jeong, W.H. Kim, J.-G. Park, Metabotropic glutamate receptor 4 expression in colorectal carcinoma and its prognostic significance, *Clinical cancer research* 11(9) (2005) 3288-3295.

[32] B.C. Yoo, E. Jeon, S.-H. Hong, Y.-K. Shin, H.J. Chang, J.-G. Park, Metabotropic glutamate receptor 4-mediated 5-Fluorouracil resistance in a human colon cancer cell line, *Clinical cancer research* 10(12) (2004) 4176-4184.

[33] Z. Zhang, Y. Liu, K. Wang, K. Zhu, X. Zheng, L. Wang, Y. Luan, X. Wang, H. Lu, K. Wu, Activation of type 4 metabotropic glutamate receptor promotes cell apoptosis and inhibits proliferation in bladder cancer, *Journal of cellular physiology* 234(3) (2019) 2741-2755.

[34] S.-Y. Park, S. Lee, I.-H. Han, B.-C. Yoo, S.-H. Lee, J.-Y. Park, I.-H. Cha, J. Kim, S.-W. Choi, Clinical significance of metabotropic glutamate receptor 5 expression in oral squamous cell carcinoma, *Oncology reports* 17(1) (2007) 81-87.

[35] G. Battaglia, C.L. Busceti, G. Molinaro, F. Biagioni, A. Traficante, F. Nicoletti, V. Bruno, Pharmacological activation of mGlu4 metabotropic

glutamate receptors reduces nigrostriatal degeneration in mice treated with 1-methyl-4-phenyl-1, 2, 3, 6-tetrahydropyridine, *Journal of Neuroscience* 26(27) (2006) 7222-7229.

[36] W. Dröge, H.-P. Eck, M. Betzler, P. Schlag, P. Drings, W. Ebert, Plasma glutamate concentration and lymphocyte activity, *Journal of cancer research and clinical oncology* 114(2) (1988) 124-128.

[37] M. Kansara, K. Thomson, P. Pang, A. Dutour, L. Mirabello, F. Acher, J.-P. Pin, E.G. Demicco, J. Yan, M.W. Teng, Infiltrating myeloid cells drive osteosarcoma progression via GRM4 regulation of IL23, *Cancer discovery* 9(11) (2019) 1511-1519.

[38] I. Chan, R. Jain, M. Tessmer, D. Gorman, R. Mangadu, M. Sathe, F. Vives, C. Moon, E. Penaflo, S. Turner, Interleukin-23 is sufficient to induce rapid de novo gut tumorigenesis, independent of carcinogens, through activation of innate lymphoid cells, *Mucosal immunology* 7(4) (2014) 842-856.