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# Chapter Ciliates as Symbionts

Rosaura Mayén-Estrada, Roberto Júnio Pedroso Dias, Mireya Ramírez-Ballesteros, Mariana Rossi, Margarita Reyes-Santos, Carlos Alberto Durán-Ramírez and Gerardo Cruz-Jiménez

### Abstract

Although many ciliates are free-living, more than 140 families of ciliates (Alveolata, Ciliophora) include symbiotic species of animals. Symbiosis, defined as an interaction between two species, is analyzed in this chapter to show a wide diversity of symbiotic systems in ciliates (epibiosis, commensalism, mutualism, and parasitism), providing some data about ciliate strategies showing their success as symbionts. Some species are free-living as well symbionts, facultative symbionts, and obligate symbionts. Analysis of reconstructions of ancestral state evidence that the parasitism arose numerous times and independently among the lineages of ciliates. At least three evolutionary routes can be traced: (1) transition from free-living to mutualism and parasitism, (2) transition from free-living to parasitism, and (3) regression from parasitism to free-living. The evolution of the symbiosis in ciliates demonstrates a higher diversification rate concerning free-living ciliates. The analysis of the evolution of the life cycles complexity, exploring molecular data of the phases of the ciliate cycle in their hosts is also essential. We propose new approaches for an integrative study of symbiotic ciliates.

**Keywords:** Ciliophora, diversity, ecology, macroevolution, morphology, physiology, symbiosis, taxonomy

# **1. Introduction**

Ciliates (Alveolata: Ciliophora) comprise free-living and symbiotic species. According to Corliss, [1] 2,600 species of ciliates have been described as symbionts, mainly of individuals of metazoan phyla. This is equivalent to 33% of all the known species of the phylum. They belong to eight classes (Armophorea, Heterotrichea, Litostomatea, Nassophorea, Oligohymenophorea, Plagiopylea, Phyllopharyngea and Spirotrichea), 31 orders, 151 families, and almost 700 genera [2]. These symbiotic ciliates have been reported in aerobic and anaerobic environments and from aquatic and terrestrial habitats [2, 3].

The term symbiosis can be defined as a sustained relationship between at least two individuals from different species, either living in direct contact or close enough to each other during a part or the whole life cycles of the partners. This interaction is transmitted vertically (from one generation to the next) or horizontally (acquired *de novo* in each generation). The intricate associations are believed to have an essential driving force in evolutionary biology, as a host and their symbiotic microbiota acclimatize on scales of short time [4]. Due to the diversity of symbioses, a classification system for symbiotic associations has been developed. This classification is based on several features: i) the dependence, where symbionts can be obligate or facultative; ii) specificity of the symbionts; iii) nutrients obtention, then biotrophic and necrotrophic symbionts are distinguished on the basis of whether nutrients are obtained from a living or dead partner, and iv) location of the symbionts, ectosymbionts or endosymbionts [5]. The symbiotic relationships can be categorized into mutualistic, commensalistic, or parasitic [2, 6]. The boundary between these categories sometimes is not clear, and there are frequent transitions between them.

Several papers have been focused on providing taxonomic reports for symbiotic ciliates, some of them as general works, and a few directed to certain groups [7–16], and some were focused on certain geographic areas [17–24]. Critical reviews of some species as *Balantidium coli* were done by Schuster and Ramirez-Avila [25]; for chonotrichs [26]; peritrichs [27] and suctorians [28].

Also, very different topics about ciliates and their hosts have been developed as shown: symbiotic interactions [epibiotic, hyperepibiotic, commensals, parasites (obligates and facultatives)], codiversification: [29–37]. Morphology (variation, molecular characterization): [38], clevellandellid, *Nyctotheroides*; [39], *Dicontophrya*; [40, 41] peritrichs. Taxonomy (new family, genus or species), redescription, revision: Apostomatia: [42]; Apostomatida: [43]; *Trichodina*: [44]; *Epistylis* and *Opercularia*: [45]; *Spirochona*: [46]; *Buetschlia* and *Charonina*: [47, 48, 49–51, 31]. Life cycles, encystment/excystment process: [52–54]. Pathogenicity, damages, infestation degree, virulence: [55–59]. Molecular and phylogeny: [30, 60–68]. Ecological aspects: [69, 70]. Immunity: [71, 72]. Stomatogenesis: [73]. Ultrastructure: [74].

Symbiotic systems between ciliates/animals are present in a broad spectrum of kingdom Animalia, and some examples are the following (animal group alphabetically arranged, different taxonomic levels): acari: [75]; amphipods: [76]; antilope: [77]; anuran: [78]; Asian elephant: [79]; baboon: [80]; bryozoans: [81]; buffaloes: [82]; capybara: [83–85]; cattle: [86]; chimpanzees: [87]; cirripedians: [88]; crustaceans: [89]; ctenophores: [90]; cuttlefish: [91]; dromedary camels: [92]; elephants: [93]; fishes: [94, 95]; frogs: [96]; great apes: [97]; horses: [98, 99]; humans: [100, 101]; polyps of hydras: [102]; insects: [103]; isopods: [104, 105]; kinorhynchs: [106]; llamas: [107]; maccacus: [108]; mammals: [109]; mollusks: [71, 76]; nematodes: [29, 110]; nemerteans: [13]; oligochaetes: [111, 112]; ostracods: [113]; polychaetes: [114, 115]; rhinoceroses: [116]; sea urchins: [117]; thoroughbreds: [118]; turbellarians: [119]; wood-feeding roaches: [120].

Some examples of ciliate taxa that include symbiotic species are the following: **Heterotrichea:** Folliculinids attach to the integument of various invertebrates as bivalve shells, crustaceans exoskeleton, polychaete tubes, hydroid perisarcs, bryozoan tests, with a widespread occurrence [121], and may cause the skeletal eroding band or brown band diseases of scleractinian corals [2]; their life cycle includes a migratory swimming stage.

**Spirotrichea:** Hypotrichs are known mainly as free-living organisms, but some species such as *Euplotes balteatus* have been recorded in some sea urchins' intestinal tract [122]. Some species of stichotrichids as *Plagiotoma lumbrici* are endosymbionts of oligochaetes [123].

**Armophorea:** Class Armophorea includes clevellandellids as Nyctotheridae, with obligate endosymbionts usually as commensals of invertebrates and vertebrates; life cycles include a phase of the cyst [2].

**Litostomatea:** Trichostomes are symbionts of vertebrates as ruminants and foregut fermenters [2], including the human pathogen, *Balantidium coli*, species that have a life cycle including two phases: trophozoites and cysts [25]. This species

has been considered to be included in a new genus, *Neobalantidium coli* [124]. The genus *Balantidium* has a more significant number of species that have been reported as endocommensals in the digestive tracts of a widely diverse range of metazoan, as mollusks, arthropods, fishes, reptiles, birds, and mammals [124]. In the rumen ecosystem, ciliates can account for up to 50% of the total microbial nitrogen, reaching densities of 10<sup>5</sup> to 10<sup>6</sup> cells/ml rumen fluid, being *Charonina ventriculi* one of the smallest rumen ciliates [125].

Ophryoscolecidae and Cycloposthiidae include species as endosymbionts of ruminants and equids, respectively [126]. Entodiniomorphid ciliates of the genus *Triplumaria* are found in the intestine of elephants and rhinoceroses [60]. Entodiniomorphida do not form cysts, and in non-ruminant mammals, the infections of hosts occur by coprophagy [47].

**Phyllopharyngea:** Chonotrichs live on marine and freshwater hosts and divide by forming external or internal buds [127], with a dimorphism where the adults live attached to several appendages of crustaceans, and the larva is free and swims to reach a new host [128].

Suctorians, as a rule, reproduce by different modes of budding, produce one to several larvae, with a short swimming existence, and then lose their cilia and metamorphose into adults or trophonts [127]. The non-ciliated mature stages of suctorians are usually sessile, attached to the substrate by a non-contractile stalk, and reproduce by ciliary larvae called swarmers or migrators [129].

**Oligohymenophorea:** Yi et al. [130] documented that the life cycle of *Ichthyophthirius multifiliis*, a parasite of fish, consists of three key developmental stages: the infective theront, the parasitic trophont, and the reproductive tomont.

*Mesanophrys pugettensis*, is a scuticociliate thata was observed with a diphasic life history, the larger phase or trophont, and the smaller phase resembling tomites [34], is a facultative parasite of the Dungeness crab. *Conchophthirus* species are generally considered an endocommensal inhabiting the mantle cavity of freshwater clams or mussels [30].

Thigmotrichids from several families were analyzed by Raabe [131–134], where species of Hemispeiridae are symbionts of the mantle cavity and nephridia of molluscan, those of Ancistrocomidae, Sphenopryidae and Thigmophryidae are ectosymbionts of mantle cavity and gills of molluscan, and Hysterocinetidae species were categorized as endoparasites of the gut of prosobranch mollusks; life cycles include tomites.

The apostomes is a small group of oligohymenophorean ciliates, with four major life histories: 1-exuviotrophic, that remain encysted on the exoskeleton of a crustacean host, and excyst to feed on exuvial fluid, reproducing during the host ecdysis, 2-sanguicolous, penetrate the cuticle of the host, feed on the cells and fluid of the hemocoel and reproduces, 3-chromidinid, found in the renal organs and opalinopsids found in the liver and intestines of cephalopods ingesting fluids and cells, 4-histotrophs, such as *Vampyrophrya* [135]. Apostome ciliates have life cycles typically involving crustaceans, with a non feeding microstome tomite and a macrostomous trophont [127]. Species of apostome of genus *Collinia* are endoparasites able to reproduce rapidly within the host that invariably kill the euphausiid within 40 hours of infection; *Gymnodinioides* genus includes exuviotrophic species that feed on the fluid within the exuviae of crustacean hosts and Landers *et al.*, [136] documented for *Gymnodinioides pacifica* the presence of trophonts, phoronts, tomonts and tomites. For *Synophrya* the phoront, hypertrophont, hypertomont, and hypertomites were observed [137].

Pilisuctorian ciliates spend most of their lives perched on cuticular setae of crustaceans, and complete their life cycle on a single host, having the stages tomite, tomont and trophont [138].

In peritrichs, a significant character is the scopula which is the region that originates the stalk to attach the organism to the substrate and modifies to a highly complicated adhesive apparatus in mobiline [127]; two phases are known, the trophont and the dispersive telotroch.

Species of sessile peritrichs genera such *Ambiphrya*, *Epistylis*, *Heteropolaria*, Rhabdostyla, and Zoothamnium are epibionts of zooplanktonic invertebrates, larval stages of aquatic insects, aquatic mollusks, crustaceans, fish, amphibians, and reptiles as the main groups of organisms [139]. Members of genus *Epistylis* have been reported as epibionts in several metazoans, but also as an important fish ectoparasite being considered an emerging pathogen [140]. Genus Lagenophrys comprises only symbiotic species of freshwater and marine crustaceans [89]. Trichodinids are the most devastating ectoparasites of cultured fish, causing severe damage [141], and for genus *Trichodina* about 300 species have been described, mostly from freshwater environments [142]. Also, there are reports of trichodinids from the gills of limpets [143] and have been documented as symbionts of a broad spectrum of aquatic and terrestrial invertebrates and vertebrates hosts [65]. Trichodinella epizootica is one of the most widely distributed freshwater trichodinids in Europe and Asia, but has also been reported from Africa, the Pacific region and North America [55]. Urceolaria includes species ectosymbionts of freshwater turbellarians, marine polychaetes, and mollusks; Leiotrocha species are ectocommensals and endocommensals of marine molluscans, and species of *Polycycla* are endocommensals of Holothuroidea [144].

#### 2. Ecological relationships: Classical definitions and approaches

#### 2.1 Epibiosis

Epibiosis is a facultative association of two organisms: the epibiont, which colonizes the surface of live substrates, and the basibiont, which hosts the epibionts [145]. Some species of epibiotic communities show preferences for specific location sites on the host [76]. According to Wahl and Mark [146], when the effects associated with epibiosis are neutral or positive for a basibiont species and beneficial for an epibiont species, selection should favor the evolution of the epibiotic relationship, which tends to increase specificity through evolutionary history. Although many epibiont ciliates are not harmful to their basibionts, some studies have shown that the epibionts can cause deleterious effects on their hosts [147–149].

Historically, studies involving epibiont ciliates focus on the following interests: new records and checklists [27, 28], descriptions of new taxa using morphological and molecular data [150], possible deleterious effects on hosts [149, 151], distribution and preferred sites of epibiont populations and communities [152], spatial and temporal distribution of the epibiotic relationship [153], laboratory rearing and experimentation studies [154–156], and even investigations into extrinsic and intrinsic factors involved in the kinetics of epibiont ciliate populations [157, 158].

#### 2.2 Mutualism

Mutualism is a relationship with high metabolic dependence, where both organisms, ciliate and their hosts, obtain benefits [159, 160]. In the phylum Ciliophora, this type of relationship is seen, mainly in the subclass Trichostomatia, which includes the ciliates of the digestive tract of herbivorous mammals [161]. The symbiont ciliates represent approximately 2,600 of the described organisms, of which around 1000 species belong to the subclass Trichostomatia [2]. This subclass comprises ciliated protists, mostly mutualists of the digestive tract of

several vertebrate hosts, with only one species showing parasitism in humans, *Balantidium coli* [2, 162, 163]. The subclass Trichostomatia is divided into three orders: Vestibuliferida, Entodiniomorphida, and Macropodiniida.

Ruminant ciliates and the host have a fundamental symbiosis relationship for the digestion and absorption of large amounts of plant material by the ruminant [164, 165]. On the one hand, the host provides an ideal environment for the survival of the symbiotic microbiota. The rumen is a strictly anaerobic environment, with temperatures ranging from 38 to 41° C, redox potential around 250 to 450 mV (millivolts), osmolarity ranging from 260 to 340 mOsm (millivolts), and pH levels between 5.0 and 7.5. Maintaining these characteristics is essential for microbial enzymatic activity to occur. In return, symbionts provide energy, protein, and vitamins to the host [166]. In energy terms, about 50–70% of the energy obtained by the host comes from the absorption of volatile fatty acids (VGAs) (eg. acetate, butyrate, and propionate), which are absorbed after the breakdown and fermentation of plant fiber by ruminal microorganisms [165]. Ciliates also represent a great source of protein for the ruminant (about 2 to 5%). Still, the ruminal microbiota also synthesizes B and K vitamins in sufficient quantities for the maintenance and growth of the animal. Due to the important participation in the physiology of the ruminant, the evolutionary dynamics of ruminal ciliates has been suggested as closely associated with the radiation of their hosts [167–169].

#### 2.3 Commensalism and parasitism

Commensalism occurs when the symbiont inhabits in the host with no evident benefit or harm [170].

Parasitism, which is less common in ciliates, involves the parasites that usually cause disease being pathogens. They may be localized or spread throughout a host, defined as the independent and dominant member of the symbiotic pair. Here, the parasite inhabits on or inside the host to obtain resources and to harm it [171].

#### 3. Ecological relationships: evolutionary approach

From an evolutionary point of view, there are species that are entirely free-living, those which can live equally well both free or as symbionts, species that are almost entirely symbiotic with only occasional periods of "free" existence during their life cycles (facultative symbionts), and species which are entirely symbiotic (obligate symbionts). Most of the well documented associations between Ciliophora and Metazoa are the ones leading to a certain degree of metabolic dependence. We will use in this topic the idea of metabolic dependence to define the ecological relationships: "free-living" (no metabolic dependence), "epibiont" (facultative metabolic dependence), "mutualistic" (mutual metabolic dependence) or "parasitic" (unilateral metabolic dependence, including commensalism).

For many years the evolutionary studies for Ciliophora were based only on morphological data, mainly those related to the ultrastructural characterization of its complex infraciliature [2]. However, in recent years this scenario has been modified with the implementation of modern tools that use multidisciplinary methods to integrate morphological, phylogenetic, molecular, and ecological data [161, 172–174]. A reliably dated phylogeny is fundamental to infer a broad macroevolutionary scenario for Ciliophora [172]. The inference of diversification rates from molecular phylogenies has increasingly been used to derive macroevolutionary patterns of lineages. Understanding how the different ecological relationships evolve in Ciliophora along time is a complex task that has been developed for many years. Different hypotheses about the origin and evolution of parasitic life have been proposed. Parasitologists suggest that the symbiotic way of life probably descended from free-living lineages that subsequently adapted to life in special habitats. Besides this, several authors suggest multiple origins of parasitism based on a comparison of morphological and ultrastructural aspects between them and their free life co-specifics [175], however, the processes that lead to its emergence are still imprecise [176–178].

Concerning the phylum Ciliophora, the vast majority of ciliates are categorized as free-living, and studies suggested that symbiosis apparently arising independently among various classes [179]. For genus Tetrahymena (subclass Hymenostomatia, order Hymenostomatida), all gradations of adaptations to symbiosis occur. There are species that live totally free, those that can live equally well both free and as symbionts, species that are almost entirely symbiotic with only occasional periods of "free" existence during their life cycles (optional symbionts), and species that are totally symbiotic (mandatory symbionts) [180]. Different transition routes between ecological associations have also been proposed, based on morphological and ecological characteristics. The first one proposes that free-living organisms assume habits of low metabolic dependence (mutualism, commensalism, among others), and with the strengthening of relationships, where they become parasites [176, 181]. The second hypothesis suggests that a free-living organism, when it comes into contact with a host accidentally, adapts itself to live both freely and within that host (optional parasite) [179], that is, free-living organisms adapt to live inside a host, which becomes something advantageous and increases fitness, making this a favorable way of life for the species.

Previous studies aimed to test these hypotheses based on phylogenetic analyzes of small groups within Ciliophora [174, 182, 183]. The macroevolutionary analyzes from the whole Ciliophora phylogeny presented **Figure 1** suggested that the ancestral way of life of the ciliates originated from a free-living organism and that the parasitic way of life arose numerous times and independently in Ciliophora, which was induced by two types of ancestors, free life and mutualistic (**Figure 1**). The transition to the parasitic way of life was recovered from two different origins: 1) a free-living ancestor evolved into a mutualistic organism and, later, to a parasitic organism, and 2) a free-living ancestor evolved into an organism parasite (highest number of cases). There are also cases where there has been a regression in the ciliate's way of life, where parasite clades evolved to free-living clades (**Figure 1**).



#### Figure 1.

Ancestral habit reconstruction for Ciliophora showing the main routes of transitions. Blue: Free-living. Yellow: Mutualism. Red: Parasitism/commensalism.

## 4. Future perspectives

The analytical improvement for morphological, ultrastructural, molecular, and evolutionary characterizations in Ciliophora culminated in an "Age of Integration", which several disciplines interact to infer patterns of biodiversity [184]. Although it is an age in full expansion, several gaps often prevent a study of diversity in its diverse areas in a complete way.

We are in a period of the paradigm shift, where Next Generation Sequencing (NGS) techniques have been applied exponentially, and, therefore, it is expected that new discoveries will emerge and new panoramas will be drawn on the diversity of the strains, as well as their respective ecological interactions. The transition from phylogenetic studies to phylogenomics is based on technological progress combined with exponential sequencing of molecular sequences (DNA, RNA), reduced associated costs, increased computational capacity, and improved analytical protocols. It is important to make efforts in studies to expand such technologies to lineages with little sampling in databases. For example, the classes Prostomatea, Oligohymenophorea, Litostomatea, and Phyllopharyngea, which present several examples of symbiosis, do not have available molecular sequences which prevents the evolutionary inferences of these lineages, requiring in the future more studies to refine the evolutionary hypotheses about the phylum. Efforts to expand metataxonomy using metagenomics and metatranscriptome methods have fed the databases exponentially in several lineages, revolutionized the analysis of environmental microbial diversity [175, 185, 186]. In fact, the generation of data for the target sequencing of phylogenetic, metagenomic, and metatranscriptomic markers is now reasonably well established, and several DNA sequencing platforms based on different technologies are currently available as well as different bioinformatics programs for each level of data extraction. However, due to the limited size of the molecular sequences produced by the platforms (~ 500 bp), phylogenetic estimates may be inadequate. With longer readings comes an improved phylogenetic signal, and we show that it is possible to employ a complete phylogenetic signal approach to taxonomically classify sequences and obtain a robust evolutionary structure of environmental diversity. New sequencing technologies such as nanopore sequencing, which offer long reads, improved the phylogenetic signal and more robust taxonomic patterns, can be an alternative in future studies [187].

With the significant increase in the number of available sequences from NGS sequencing, more effective and less subjective methodologies have been proposed to define the limits and number of independent evolutionary entities, to accelerate the biodiversity assessment process. In the last two decades, the field of species delimitation has intensified in relation to the number of methods available. For this, several methodologies have been proposed, based on biological [188], ecological [189], and molecular data [190], in addition to combining phylogenetic theory and population genetics [191–193]. The use of these methodologies in ciliates performed very recently to delimit organisms of free life, as species of the genus *Frontonia*, using the mitochondrial gene COX1 [194], species of the genus *Spirostomum*, using the ITS spacer region genes [195], and COI and 18S markers of the *Paramecium* genus.

Finally, several authors have emphasized the lack of studies on the distribution and occurrence of ciliates associated with Metazoa in natural conditions and the the lack of information on the ecology and interactions between epibionts and hosts. Few studies are exploring the natural history and complexity of life cycles, which makes it difficult to characterize optional and mandatory relationships. The absence of the characterization of the ciliate at the stage it is in the host, most studies, only in the environment, making it difficult to characterize the life cycle. Relevant information about habitat, life cycle, infection site is rare for Ciliophora [160, 196, 197].

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# **Author details**

Rosaura Mayén-Estrada<sup>1\*</sup>, Roberto Júnio Pedroso Dias<sup>2</sup>, Mireya Ramírez-Ballesteros<sup>1</sup>, Mariana Rossi<sup>3</sup>, Margarita Reyes-Santos<sup>1†</sup>, Carlos Alberto Durán-Ramírez<sup>1</sup> and Gerardo Cruz-Jiménez<sup>1</sup>

1 Laboratorio de Protozoología, Facultad de Ciencias, Universidad Nacional Autónoma de México, Mexico City, Mexico

2 Laboratório de Protozoologia, Programa de Pós-graduação em Biodiversidade e Conservação da Natureza, Universidade Federal de Juiz de Fora, Juiz de Fora, Brazil

3 Laboratório de Protozoologia, Instituto de Ciências Biológicas, Universidade Federal de Juiz de Fora, Juiz de Fora, Brazil

\*Address all correspondence to: rme2@ciencias.unam.mx

† Deceased.

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# References

[1] Corliss JO. Biodiversity, classification, and numbers of species of protists. Nature and human society. The quest for a sustainable world. National Academy Press, Washington. 2000;130-155

[2] Lynn DH. The Ciliated Protozoa.Characterization, classification, and guide to the literature. Ontario: Springer;2008. 605 p

[3] Rosati G. Ectosymbiosis in Ciliated Protozoa. In: Seckbach J, editor. Symbiosis. Mechanisms and Model Systems Cellular Origin, Life in Extreme Habitats and Astrobiology. Dordrecht: Springer; 2004. p. 475-488

[4] Oulhen N, Schulz BJ, Carrier TJ. English translation of Heinrich Anton de Bary's 1878 speech, Die Erscheinung der Symbiose' ('De la symbiose'). 2016;69: 131-139

[5] Paracer S, Ahmadjian V. Symbiosis: An Introduction to Biological Associations. New York: Oxford University Press; 2000. 291 p

[6] Dziallas C, Allgaier M, Monaghan MT, Grossart HP. Act together—implications of symbioses in aquatic ciliates. Frontiers in Microbiology. 2012;3(288):1-17

 [7] Nenninger U. Die Peritrichen der Umgebung von Erlangen mit besonderer Berüksichtigung ihrer Wirtsspezifität.
 Zoologische Jahrbücher (Systematik).
 1948;77:169-266.

[8] Wolska M. Study on the family Blepharocorythidae Hsiung. I. Preliminary remarks. Acta Protozoologica. 1966;4(12):97-103

[9] Sprague V, Couch JA. An annotated list of protozoan parasites, hyperparasites, and commensals of decapod crustaceans. Journal of Protozoology. 1971;18:526-537

[10] Morado JM, Small EB. Ciliate parasites and related diseases of

Crustacea: a review. Reviews in Fisheries Science. 1995;3(4):275-354

[11] Bojko J, Ovcharenko M. Pathogens and other symbionts on the Amphipoda: taxonomic diversity and pathological significance. Diseases of Aquatic Organisms. 2019;136:3-36

[12] Clamp JC, Chatterjee T, Fernandez-Leborans G. Occurrence of *Epistylis anastatica* (Linnaeus, 1767) (Ciliophora: Peritrichia) on *Mesocyclops isabellae* Dussart & Fernando (Crustacea: Copepoda: Cyclopoida) in India, with an annotated checklist of species of *Epistylis* reported as epibionts of cyclopoid copepods and resources for their identification. Zootaxa. 2016;4168(3):525-540

[13] McDermott JJ. Nemerteans as hosts for symbionts: a review. Journal of Natural History. 2006;40(15-16): 1007-1020

[14] Edgerton BF, Evans LH, Stephens FJ,Overstreet RM. Synopsis of freshwater crayfish diseases and commensal organisms. Aquaculture. 2002;296(1-2): 57-135

[15] Matthes D, Guhl W, Haider G. Suctoria und Urceolariidae. Protozoenfauna Band 7/1. Stutgart: Gustav Fischer Verlag; 1988. 309p

[16] de Jager GP, Basson L. Taxonomic assessment of three North American trichodinids by re-evaluating the taxon validity of *Trichodina heterodentata* Duncan, 1977 (Peritrichia). Acta Protozoologica. 2019;58:125-139

[17] Schödel H. A synopsis of the limnetic epizoic Peritrichia (Ciliophora) on invertebrates in Western-, Middle- and Eastern Europe. Denisia. 2018;41:47-294

[18] Chatterjee T, Fernandez-Leborans G, Ramteke D, Ingole B. New records of epibiont ciliates (Ciliophora) from Indian coast with descriptions of six new species. Cahiers de Biologie Marine. 2013;54:143-159

[19] Chatterjee T, Dovgal IV, Nanajkar M, Fernandes V. Report of epibiont ciliates (Ciliophora: Suctorea) on pelagic ostracods (Crustacea: Ostracoda) from the Arabian Sea. Zootaxa. 2019a;4695(4):368-384

[20] Purushothaman A, Dovgal I, Francis SV, Padmakumar KB. Observation of a suctorian ciliate *Ephelota coronata* on the calanoid copepod *Pontella spinipes* in the southeastern Arabian Sea. Symbiosis. 2020;81(3):321-327

[21] Fernandez-Leborans G, Cárdenas CA. Epibiotic protozoan communities on juvenile southern king crabs (*Lithodes santolla*) from subantartic areas. Polar Biology. 2009;32:1693-1703

[22] Viljoen S, van As JG. Sessile peritrichs (Ciliophora: Peritricha) from fresh-water fish in the Transvaal, South Africa. South African Journal of Zoology. 1985;20(3):79-96

[23] O'Donoghue PJ, Adlard RD. Catalogue of protozoan parasites recorded in Australia. Memoirs of the Queensland Museum. 2000;45(1):1-164

[24] Dovgal I, Chatterjee T, Subba Rao DV, Chan BKK, De Troch M. New records of *Praethecacineta halacari* (Schulz) (Suctorea: Ciliophora) from Taiwan, Tanzania and Canada. Marine Biodiversity Records. 2009;2:e136, 1-3

[25] Schuster FL, Ramirez-Avila L. Current world status of *Balantidium coli*. Clinical Microbiology Reviews. 2008;21:626-638

[26] Fernandez-Leborans G. A review of the species of protozoan epibionts on crustaceans. III. Chonotrich ciliates. Crustaceana. 2001;74(6):581-607 [27] Fernandez-Leborans G, Tato-Porto ML. A review of the species of protozoan epibionts of crustaceans. I. Peritrich ciliates. Crustaceana. 2000a; 73:643-683

[28] Fernandez-Leborans G,
Tato-Porto ML. A review of the species of protozoan epibionts of crustaceans.
II. Suctorian ciliates. Crustaceana.
2000b;73(10):1205-1237

[29] Baldrighi E, Dovgal I, Zeppilli D, Abibulaeva A, Michelet C, Michaud E, Franzo A, Grassi E, Cesaroni L, Guidi L, Balsamo M, Sandulli, R, Semprucci, F. The cost for biodiversity: records of ciliate-nematode epibiosis with the description of three new suctorian species. Diversity. 2020;12:(224)1-25.

[30] Antipa GA, Strüder-Kypke MC, Lynn DH. Molecular phylogeny, taxonomic relationships and North American distribution of *Conchophthirus* (Conchophthiridae, Scuticociliatia). Aquatic Ecosystem Health and Management. 2020;23(1):58-68.

[31] Li M, Wang J, Zhu D, Gu Z, Zhang J, Gong X. Study of *Apiosoma piscicola* (Blanchard 1885) occurring on fry of freshwater fishes in Hongze, China with consideration of the genus *Apiosoma*. Parasitology Research. 2008;102:931-937

[32] Kozloff EN, Norenberg JP. *Raabella concinna* sp. nov. and *Hypocomina obstipa* sp. nov., ciliates of the family Ancistrocomidae parasitizing marine mollusks on the Pacific coast of North America. Cahiers de Biologie Marine. 2013;54:175-180

[33] Xu K, Lei, Y, Al-Rasheid KAS, Song W. Two new ectoparasitic ciliates, *Sphenophrya solinis* sp. nov. and *Planeticovorticella paradoxa* sp. nov. (Protozoa: Ciliophora), from marine molluscs. Journal of the Marine Biological Association of the United Kingdom. 2011;91(2):265-274

[34] Morado JM, Small EB. Morphology and stomatogenesis of *Mesanophrys pugettensis* n. sp. (Scuticociliatida: Orchitophryidae), a facultative parasite ciliate of the dungeness crab, *Cancer magister* (Crustacea: Decapoda). Transactions of the American microscopical Society. 1994;113(3):343-364

[35] Fernandez-Leborans G, Dávila P, Cerezo E, Contreras C. Epibiosis and hyperepibiosis on *Pagurus bernhardus* (Crustacea: Decapoda) from the west coast of Scotland. Journal of the Marine Biological Association of the United Kingdom. 2013;93(5):1351-1362

[36] Van As JG, Viljoen S.A. Taxonomic study of sessile peritrichs (Ciliophora: Peritricha) associated with crustacean fish ectoparasites in South Africa. South African Journal of Zoology. 1984;19(4):275-279

[37] Lynn DH, Gómez-Gutiérrez J, Strüder-Kypke MC, Shaw CT. Ciliate species diversity and host-parasitoid codiversification in *Pseudocollinia* infecting krill, with description of *Pseudocollinia similis* sp. nov. Diseases of Aquatic Organisms. 2014;112:89-102

[38] Li M, Sun ZY, Grim JN, Ponce-Gordo F, Wang G, Zou H, Li W, Wu S. Morphology of *Nyctotheroides hubeiensis* Li et al. 1998 from frog hosts with molecular phylogenetic study of clevelandellid ciliates (Armophorea, Clevelandellida). Journal of Eukaryotic Microbiology. 2016;63(6):751-759

[39] Fokam Z, Nana PA, Ngassam P, Bricheux G, Bouchard P, Viguès B, Sime-Ngando T. Morphological characterization of two new species of *Dicontophrya* (Ciliophora: Astomatia: Contophryidae) comensal of earthworms (Oligochaeta: Annelida) of Edeba and Nkolbikogo (Cameroon). International Journal of Biosciences. 2015b;7(1):49-57

[40] Zhou T, Wang Z, Yang H, Gu Z. Morphological and molecular identification of epibiontic sessilid *Epistylis semiciculus* n. sp. (Ciliophora, Peritrichia) from *Procambarus clarki* (Crustacea, Decapoda) in China. LJP: Parasites and Wildlife. 2019a;10:289-298

[41] Clamp JC. The occurrence of Lagenophrys aselli (Ciliophora: Peritricha: Lagenophryidae) in North America and a description of environmentally-induced morphological variation in th especies. Transactions of the American microscopical Society.
1988;107(1):17-27

[42] Chantangsi C, Lynn DH, Rueckert S, Prokopowicz AJ, Panha S, Lenader BS. *Fusiforma themisticola* n. gen. n. sp., a new genus and species of apostome ciliate infecting the hyperiid amphipod *Themisto libellula* in the Canadian Beaufort Sea (Arctic Ocean), and establishment of the Pseudocolliniidae (Ciliophora, Apostomatia). Protist. 2013;164:793-810

#### [43] Gómez-Gutiérrez J,

Strüder-Kypke MC, Lynn DH, Shaw TC, Aguilar-Méndez MJ, López-Cortés A, Martínez-Gómez S, Robinson CJ. *Pseudocollinia brintoni* gen. nov., sp. nov. (Apostomatida: Colliniidae), a parasitoid ciliate infecting the euphausiid *Nyctiphanes simplex*. Diseases of Aquatic Organisms. 2012;99:57-78

[44] Pinto HA, Wieloch AH, de Melo AL. Uma nova espécie de *Trichodina* Ehrenberg, 1838 (Ciliophora: Trichodinidae) em *Biomphalaria schrammi* (Crosse, 1864) (Mollusca: Planorbidae). Lundiana. 2006;7(2):121-124.

[45] Zhou T, Wang Z, Yang H, Gu Z. Two new colonial peritrich ciliates (Ciliophora, Peritrichia, Sessilida) from China: with a note on taxonomic distinction between Epistylididae and Operculariidae. European Journal of Protistology. 2019b;70:17-31

[46] Dovgal IV, Grigorovich IA. *Spirochona americana* sp. n. (Ciliophora,

Chonotrichia)- a new North American freshwater chonotrich species from the amphipod (*Gammarus lacustris*). Vestnik Zoologii. 2001;35(5):77-80

[47] Gürelli G. New entodiniomorphid ciliates, *Buetschlia minuta* n. sp., *B. cirrata* n. sp., *Charonina elephanti* n. sp., from Asian elephants of Turkey. Zootaxa. 2019;4545(3):419-433

[48] Grim JN, Buonanno F. A re-description of the ciliate genus ans type species, *Balantidium entozoon*. European Journal of Protistology. 2009;45:174-182

[49] van Hoven W, Gilchrist FMC, Hamilton-Attwell VL. A new family, genus, and seven new species of Entodiniomorphida (Protozoa) from the gut of African rhinoceros. Journal of Protozoology. 1988;35(1):92-97

[50] Bradbury PC, Clamp JC. *Hyalophysa lwoffi* sp. n. from the freswater shrimp *Palaemonetes paludosus* and revision of the genus *Hyalophysa*. Journal of Protozoology. 1973;20(2):210-213

[51] Van As JG, Basson L. A further contribution to the taxonomy of the Trichodinidae (Ciliophora: Peritrichia) and a review of the taxonomic status of some fish ectoparasitic trichodinids. Systematic Parasitology. 1989;14:157-179

[52] Bassham RD, Browning JS, Landers SC. The complete life cycle of the inusual apostome *Hyalophysa clampi* (Ciliophora, Apostomatida), a symbiont of crayfish in Alabama (USA). European Journal of Protistology. 2020;72:125654

[53] Lucas, C. A study of excystation in *Nyctotherus ovalis* with notes on other intestinal protozoa of the cockroach. The Journal of Parasitology. 1928;14(3): 161-175

[54] Segade P, García N, García-Estevez JM, Arias C, Iglesias R. Encystment/excystment response and serotypic variation in the gastropod parasite *Tetrahymena rostrata* (Ciliophora, Tetrahymenidae). Parasitology Research. 2016;115:771-777

[55] Basson, L. First records of trichodinid ectoparasites (Ciliophora: Peritrichia) from introduced freshwater fishes in Tasmania, Australia, with comment son pathogenicity. Acta Protozoologica. 2010;49:253-265

[56] Small HJ, Neil DM, Taylor AC, Bateman K, Coombs GH. A parasitic scuticociliate infection in the Norway lobster (*Nephrops norvegicus*). Journal of Invertebrate Pathology. 2005;90:108-117

[57] Xie P, Sanderson B, Frost T, Magnuson JJ. Manipulation of host density affects infestation of a peritrich ciliate (*Epistylis lacustris*) on a calanoid copepod (*Leptodiaptomus minutus*) in Crystal lake, Wisconsin, USA. Journal of Freshwater Ecology. 2001;16(4):557-564

[58] Hanamura Y, Nagasaki K. Occurrence of the sandy beach mysids *Archaeomysis* spp. (Mysidacea) infested by epibiontic peritrich ciliates (Protozoa). Crustacean Research. 1996;25:25-33

[59] Swennes AG, Noe JG, Craig Findly R, Dickerson HW. Differences in virulence between two serotypes of *Ichthyophthirius multifiliis*. Diseases of Aquatic Organisms. 2006;69:227-232

[60] Ito A, Honma H, Gürelli G, Cocmen B, Mishima T, Nakai Y, Imai S. Redescription of *Triplumaria selenica* Latteur et al., 1970 (Ciliophora, Entodiniomorphida) and its phylogenetic position base don the infraciliary bands an 18SSU rRNA gene sequence. European Journal of Protistology. 2010;46:180-188

[61] Ito A, Ishihara M, Imai S. Bozasella gracilis n. sp. (Ciliophora, Entodiniomorphia) from Asian elephant and phylogenetic analysis of entodiniomorphids and vestibuliferids.

European Journal of Protistology. 2014;50:134-152

[62] Lu B, Shen Z, Zhang Q, Hu X, Warren A, Song W. Morphology and molecular analyses of four epibiotic peritrichs on crustacean and polychaete hosts, including descriptions of two new species (Ciliophora, Peritrichia). European Journal of Protistology. 2020;73:125670

[63] Snelling T, Pinloche E, Worgan HJ, Newbold CJ, McEwan NR. Molecular phylogeny of *Spirodinium equi*, *Triadinium caudatum* and *Blepharocorys* sp. from the equine hindgut. Acta Protozoologica. 2011;50:319-326

[64] Tang FH, Zhao YJ, Warren A.Phylogenetic analyses of trichodinids (Ciliophora, Oligohymenophorea) inferred from 18S rRNA gene sequence data. Current Microbiology.2013;66:306-313

[65] Xu K, Lei Y, Song W, Choi JK, Warren A. Diagnoses and probable phylogenetic relationships of the genera in the family Trichodinidae (Ciliophora, Peritrichia). The Yellow Sea. 2000;6:42-49

[66] Gao F, Gao S, Wang P, Katz L, Song W. Phylogenetic analyses of cyclidiis (Protista, Ciliophora, Scuticociliatia) based on multiple genes suggest their close relationship with thigmotrichichids. Molecular Phylogenetics and Evolution. 2014;75:219-226.

[67] Affa'a FM, Hickey DA, Strüder-Kypke M, Lynn DH. Phylogenetic position of species in the genera *Anoplophrya*, *Plagiotoma*, and *Nyctotheroides* (Phylum Ciliophora), endosymbiotic ciliates of annelids and anurans. Journal of Eukaryotic Microbiology. 2004;51(3):301-306

[68] Clamp JC, Bradbury PC, Strüder-Kypke MC, Lynn DH. Phylogenetic position of the apostome ciliates (Phylum Ciliophora, Subclass Apostomatia) tested using small subunit rRNA gene sequences. Denisia. 2008;23:395-402

[69] Fokam Z, Nana PA, Moche K, Bricheux G, Bouchard P, Ngassam P, Sime-Ngando T. Influence of soil physicochemical parameters on the abundance of *Paracoelophrya polymorphus* (Ciliophora; Radiophryidae) commensal of earthworms (Annelida: Glossoscolecidae) collected in Bambui (North-West Cameroon). Journal of Biodiversity and Environmental Sciences. 2015a;6(2): 376-389

[70] Ogut H, Akyol A, Alkan MZ. Seasonality of *Ichthyophthirius multifiliis* in the trout (*Oncorhynchus mykiss*) farms of the Eastern Black Sea region of Turkey. Turkish Journal of Fisheries and Aquatic Sciences. 2005;5:23-27

[71] Xu K, Song W. Two trichodinid ectoparasites from marine molluscs in the Yellow Sea, off China, with the description of *Trichodina caecellae* n sp. (Protozoa: Ciliophora: Peritrichia). Systematic Parasitology. 2008;69:1-11

[72] Heinecke RD, Buchmann K.
Inflammatory response of rainbow trout Oncorhynchus mykiss (Walbaum, 1792)
larvae against Ichthyophthirius multifiliis.
Fish & Shelfish Immunology. 2013;34:
521-528

[73] Njine T, Ngassam P. Morphogenesis of bipartition in two species of the genus *Ptychostomum* (Ciliophora, Hysterocinetidae). European Journal of Protistology. 1993;29:396-406

[74] Landers SC. Morphogenesis in *Hyalophysa chattoni* (Ciliophora: Apostomatida): reduction of oral polykineties in the encysted phoront. Archiv für Protistenkunde. 1997;148: 389-397

[75] Dovgal I, Chatterjee T, Ingole B. An overview of suctorian ciliates (Ciliophora, Suctorea) as epibionts of halacarid mites (Acari, Halacaridae). Zootaxa. 2008a;1810:60-68

[76] Fenchel T. On the ciliate fauna associated with the marine species of the amphipod genus *Gammarus* J. G. Fabricius. Ophelia. 1965;2(2):281-303

[77] Latteur B. *Diplodinium archon* n. sp., ciliate ophryoscolescide du rumen de l'antilope *Tragelaphus scriptus* Pallas. Annales de la Société Belge de Médecine Tropicale. 1966;46(6):727-740

[78] Affa'a FM. Observations morphologiques sur deux nyctothéres (ciliés hétérotriches) commensaux de batraciens anoures du Québec. Canadian Journal of Zoology. 1991;69:2765-2770

[79] Gürelli G. Intestinal ciliate fauna of the Asian elephant from Gaziantep, Turkey and the description of *Brevitentaculum antebum* n. g., n. sp. European Journal of Protistology. 2016;56:26-31

[80] Al-Tayib O, Abdoun KA. *Balantidium coli* infection in hamadryas baboon (*Papio hamadryas*) in Saudi Arabia: a case report. The Journal of Animal and Plant Sciences. 2013;23(3):939-943.

[81] Chatterjee T, Dovgal IV. A checklist of ciliate epibionts (Ciliophora) found on bryozoans. Zootaxa. 2020;4896(4): 547-559

[82] Roy BC, Mondal MMH, Talukder MH, Majumder S. Prevalence of *Balantidium coli* in buffaloes at different areas of Mymensingh. Journal of the Bangladesh Agricultural University. 2011;9(1):67-72

[83] Martinele I, Detoni ML, Rust NM,
D'Agosto M. Protozoários ciliados
(Protista, Ciliophora) no conteúdo do
rúmen e do retículo de bovinos. Revista
Brasileira de Zoociências. 2007;9(1):63-66

[84] Hollande A, Batisse A. Contribution à l'étude des infusoires parasites du coecum

de l'hydrocheire (*Hydrocheirus capybara* L.). I. La famille des Cycloposthiidae. Memórias do Instituto Oswaldo Cruz. 1959;57(1):1-16

[85] Ito A, Imai S. Ciliates from the cecum of capybara (*Hydrochoerus hydrochaeris*) in Bolivia. 2. The family Cycloposthiidae. European Journal of Protistology. 2000;36:169-200

[86] Gürelli G, Akman FTB. Rumen ciliate biota of domestic cattle (*Bos taurus taurus*) in Istanbul, Turkey and infraciliature of *Metadinium medium* (Entodiniomorphida, Ophryoscolecidae). Acta Protozoologica. 2017;56:171-180

[87] Pomajbíková K, Petrzelková KJ,
Petrásová J, Profousová I, Kalousová B,
Jirjú M, Sá RM, Modrý D. Distribution of the entodiniomorphid ciliate *Troglocorys cava* Tokiwa, Modrý, Ito,
Pomajbíková, Petrzelková & Imai, 2010,
(Entodiniomorphida:
Blepharocorythidae) in wild and captive chimpanzees. Journal of Eukaryotic Microbiology. 2012;59(1):97-99

[88] Arvy L, Batisse A, Lacombe D. Peritriches épizoïques dans la chambre branchiale des Balanidae (Crustacea: Cirripedia) *Epistylis nigrellii* n. sp. *E. horizontalis* (Chatton, 1930). Annales de Parasitologie Humaine et Comparee. 1969;44(4):351-374

[89] Clamp JC. Revision of the family Lagenophryidae Bütschli, 1889 and description of the family Usconophryidae n. fam. (Ciliophora; Peritricha). Journal of Protozoology. 1991;38(4):355-377

[90] Estes AM, Reynolds BS, Moss AG. *Trichodina ctenophorii* n. sp., a novel symbiont of ctenophores of the Northern coast of the Gulf of Mexico. Journal of Eukaryotic Microbiology.
1997;44(5):420-426.

[91] Ma M, Lu B, Fan X, Shi Y, Chen X. Taxonomic clarification of a well-known pathogenic scuticociliate, *Miamensis* 

*avidus* Thompson & Moewus, 1964 (Ciliophora, Scuticociliatia). Journal Ocean University China (Oceanic and Coastal Sea Research). 2018;17(6):1-12

[92] Gürelli G, Daw AFOE. Endosymbiotic ciliated protozoan biota of dromedary camels and domestic cattle in Tunisia. Zootaxa. 2020;4859(3):409-418

[93] Gürelli G, Ito A. Intestinal ciliated protozoa of the Asian elephant *Elephas maximus* Linnaeus, 1758 with the description of *Triplumaria izmirae* n. sp. European Journal of Protistology. 2014;50:25-32

[94] Corrêa LL, Ceccarelli PS, Tavares-Dias M. An outbreak of *Ichthyophthirius multifiliis* (Ciliophora: Ichthyophthiriidae) in wild endemic fish fauna *Steindachneridium parahybae* (Siluriformes: Pimelodiae) in Brazil. Annals of Parasitology. 2019;65(4): 417-421

[95] El-Tantawy SAM, Reda ESA, Abdel-Aziz A, Abou El-Nour MF, Rady I. *Apiosom*a spp. and *Scopulata epibranchialis* infesting Nile perch fish *Lates niloticus* in Dakahlia Province, Egypt. New York Science Journal. 2013;6(6):111-118

[96] Pritchett KR, Sanders GE. Epistylididae ectoparasites in a colony of African clawed frogs (*Xenopus laevis*). Journal of the American Association for Laboratory Animal Science. 2007;46(2): 86-91

[97] Pomajbíková K, Petržlková KJ, Profousová I, Petrášová J, Modrý D. Discrepancies in the occurrence of *Balantidium coli* between wild and captive African great apes. Journal of Parasitology. 2010;96(6):1139-1144

[98] Gürelli G, Göçmen B. Intestinal ciliate composition found in the feces of the Turk rahvan horse *Equus caballus* Linnaeus, 1758. European Journal of Protistology. 2011;47:245-255 [99] Gürelli G, Canbulat S, Aldayarov N. Fecal ciliate composition of domestic horses (*Equus caballus* Linnaeus, 1758) living in Kyrgyzstan. Zootaxa. 2015; 4039(1):145-156

[100] Bel GS, Couret M. *Balantidium coli* infection in man. The Journal of Infectious Diseases. 1910;7(5):609-624

[101] Bellanger AP, Scherer E, Cazorla A, Grenouillet F. Dysenteric syndrome due to *Balantidium coli*: a case report. New Microbiologica. 2013;36:203-205

[102] Warren A, Robson EA. The identity and occurrence of *Kerona pediculus* (Ciliophora: Hypotrichida), a wellknown epizoite of *Hydra vulgaris* (Cnidaria: Hydrozoa). Zoologische Verhandelingen. 1998;323(31);235-245

[103] Tanahashi M, Meng XY, Fukatsu T. A novel symbiotic ciliate (Ciliophora: Peritrichia) in the hindgut of a stag beetle (Coleoptera: Lucanidae).Zoological Science. 2017;34(3):217-222

[104] Santha Kumari V, Nair NB. Some ciliates from the marine wood-boring isopod *Sphaeroma*. Indian Journal of Fisheries. 1985;32(2):215-223

[105] Jankowski AW. *Eufolliculina brunea* sp. nov. (Ciliophora, Heterotrichida), a symbiont of wood-boring *Limnoria* isopods. Zoosystematica Rossica. 2009b;18(2):179-183

[106] Dovgal I, Chatterjee T, Ingole B, Nanajkar. First report of *Limnoricus ponticus* Dovgal & Lozowski (Ciliophora: Suctorea) as epibionts on *Pycnophyes* (Kinorhynch) from the Indian Ocean with keys to species of the genus *Limnoricus*. Cahiers de Biologie Marine. 2008b;49:381-385

[107] Cerón-Cuchi M, Marcoppido G, Dekker A, Fondevila M, De la Fuente G, Morici G, Cravero S. Ciliate protozoa of the forestomach of llamas (*Lama glama*) from locations at different altitude in Argentina. Zootaxa. 2016;4067(1):49-56 [108] Hinde K. Milk composition varies in relation to the presence and abundance of *Balantidium coli* in the mother in cautive *Rhesus macaques* (*Macaca mulatta*). American Journal of Primatology. 2007;69:625-634

[109] Nakauchi K. The prevalence of *Balantidium coli* infection in fifty-six mammalian species. Journal of Veterinary Medical Science. 1999;61(1):63-65

[110] Chatterjee T, Dovgal I, Fernandez-Leborans G. A checklist of suctorian epibiont ciliates (Ciliophora) found on meiobenthic marine nematodes. Journal of Natural History. 2019b;53(33-34):2133-2143

[111] Fokam Z, Ngassam P, Boutin CL, Zebazé-Togouet SH. Trois espèces nouvelles de *Coelophrya*, ciliés astomes endocommensaux d'*Alma nilotica* (oligochète terricole) du Cameroun. Bulletin de la Société d'histoire Naturelle de Toulouse. 2008;144:27-33

[112] Ngassam P, Grain J. Description of the new genus *Hysterocinetoides*(Ciliophora, Hysterocinetidae) and revision of the systematics of the subclass Hysterocinetia. Protistology.
2002;38:11-17

[113] Jankowski AW. *Tokophrya sibirica* sp. nov. (Ciliophora: Acinetida), a symbiont of freshwater ostracods inIrkutsk Province. Zoosystematica Rossica. 2014;23(2):174-177

[114] Lom J, Kozloff EN. The ultrastructure of *Phalacrocleptes verruciformis*, an unciliated ciliate parasitizing the polychaete *Schizobranchia insignis*. The Journal of Cell Biology. 1967;33:355-364

[115] Jankowski AW. *Aspidisca beringiana* sp. nov. and *Simbiodisca* subgen. nov. (Ciliophora: Euplotida), a symbiont of terebellid polychaetes in the Bering Sea. Zoosystematica Rossica. 2009a;18(2): 184-190 [116] Obanda V, Gakuya F, Lekolool I, Chege S, Okita B, Manyibe T. Ciliated intestinal protozoa of Black (*Diceros bicornis michaeli*) and white rhinoceroses (*Ceratotherium simum simum*) in Kenya. African Journal of Ecology. 2007;46: 144-148

[117] Lynn DH, Strüder-Kypke M. Phylogenetic position of *Licnophora*, *Lechriopyla*, and *Schizocaryum*, three unusual ciliates (Phylum Ciliophora) endosymbiotic in echinoderms (Phylum Echinodermata). Journal of Eukaryotic Microbiology. 2002;49(6):460-468

[118] Gürelli G, Lyons ET, Kesbiç FI. Hindgut ciliate composition of throughbred mares in Kentucky, USA, and binary fission in *Polymorphella ampulla*. Zootaxa. 2019;4646(2):369-384

[119] Rataj M, Vďačný P. Living morphology and molecular phylogeny of oligohymenophorean ciliates associated with freshwater turbellarians. Diseases of Aquatic Organisms. 2019;134:147-166

[120] Lynn DH, Wright ADG. Biodiversity and molecular phylogeny of Australian *Clevelandella* species (Class Armophorea, Order Clevelandellida, Family Clevelandellidae), intestinal endosymbiotic ciliates in the woodfeeding roach *Panesthia cribrata* Saussure, 1864. Journal of Eukaryotic Microbiology. 2013;60:335-341

[121] Scheltema RS. Dispersal of the protozoan *Folliculina simplex* Dons (Ciliophora, Heterotricha) throughout the North Atlantic Ocean on the shells of gastropod veliger larvae. Journal of Marine Research. 1973;31(1):11-20

[122] Curds CR. A guide to the species of the genus *Euplotes* (Hypotrichida, Ciliatea), Bulletin of the British Museum (Natural History) Zoology. 1975;28(1): 1-61

[123] Obert T, Vďačný P. Evolutionary origin and host range of *Plagiotoma* 

*lumbrici* (Ciliophora, Hypotrichia), an obligate gut symbiont of lumbricid earthworms. Journal of Eukaryotic Microbiology. 2020;67:176-189

[124] Li M, Ponce-Gordo F, Grim JN, Wang C, Nilsen F. New insights into the molecular phylogeny of *Balantidium* (Ciliophora, Vestibuliferida) based on the analysis of new sequences of species from fish hosts. Parasitology Research. 2014;113:4327-4333

[125] Kittelmann S, Devente SR, Kirk MR, Seedorf H, Dehority BA, Janssen PH. Phylogeny of intestinal ciliates, including *Charonina ventriculi*, and comparison of microscopy and 18S rRNA gene pyrosequencing for rumen ciliate community structure analysis. Applied and Environmental Microbiology. 2015;81:2433-2444

[126] Cameron SL, Wright ADG, O'Donoghue PJ. An expanded phylogeny of the Entodiniomorphida (Ciliophora: Litostomatea). Acta Protozoologica. 2003;42:1-6

[127] Foissner W. Ontogenesis in ciliated protozoa, with emphasis on stomatogenesis. In: Hausmann K, Bradbury PC, editors. Ciliates cells as organisms. Stuttgart: Gustav Fischer Verlag; 1996. p. 95-158

[128] Fahrni JF. Morphologie et ultrastucture de *Spirochona gemmipara* Stein, 1852 (Ciliophora, Chonotrichida). III. Appareil de fixation chez la larve et chez l'adulte. Journal of Protozoology. 1984;31(2):221-232

[129] Bardele CF. Budding and metamorphosis in *Acineta tuberosa*. An electron microscopic study on morphogenesis in Suctoria. Journal of Protozoology. 1970;17(1):51-70

[130] Yi YL, Lu C, Hu XG, Ling F,
Wang GX. Antiprotozoal activity of medicinal plants against *lchthyophthirius multifiliis* in goldfish (*Carassius auratus*).
Parasitology Research. 2012;111:1771-1778 [131] Raabe Z. Ordo Thigmotricha (Ciliata-Holotricha). II. Familia Hemispeiridae. Acta Protozoologica. 1970a;7:117-180

[132] Raabe Z. Ordo Thigmotricha (Ciliata-Holotricha). III. Familiae Ancistrocomidae et Sphenopryidae. Acta Protozoologica. 1970b;7:385-463

[133] Raabe Z. Ordo Thigmotricha (Ciliata-Holotricha). IV. Familia Thigmophryidae. Acta Protozoologica. 1971;9:121-170

[134] Raabe Z. Ordo Thigmotricha (Ciliata-Holotricha). V. Familiae Hysterocinetidae et Protoanoplophryidae. Acta Protozoologica. 1972;10:115-184

[135] Lynn DH, Strüder-Kypke, MC. The sanguicolous apostome *Metacollinia luciensis* Jankowski 1980 (Colliniidae, Apostomatia, Ciliophora) is not closely related to other sanguicolous apostomes. Journal of Eukaryotic Microbiology. 2018;66:140-146

[136] Landers SC, Gómez-Gutiérrez J, Peterson WT. *Gymnodinioides pacifica*, n. sp., an exuviotrophic ciliated protozoan (Ciliophora, Apostomatida) from euphausiids of the Northeastern Pacific. European Journal of Protistology. 2006;42:97-106

[137] Lee RF, Walker AN, Landers SC, Walters TL, Powell SA, Frischer ME. Black spot gill syndrome in the Northern shrimp, *Pandalus borealis*, caused by the parasitic ciliate *Synophrya* sp. Journal of Invertebrate Pathology. 2019;161:40-46

[138] Bradbury PC. *Conidophrys pitelkae*, a new species of pilisuctorian from cuticular hairs of *Crangon crangon* (Linneus). Acta Protozoologica.
1975;14(2):161-170

[139] de Padúa SB, Ishikawa MM, Ventura AS, Jeronimo GT, Martins ML, Tavares LER. Brazilian catfish parasitized by *Epistylis* sp. (Ciliophora, Epistylididae), with description of parasite intensity score. Parasitology Research. 2013;112:443-446

[140] Valladao GMR, Leyva-Pereira N, Viadanna PHO, Gallani SU, Farias THV, Pilarski F. Haematology and histopathology of Nile tilapia parasitised by *Epistylis* sp., an emerging pathogen in South America. Bulletin of the European Association of Fish Pathologists. 2014;35(1):14-20

[141] Saha M, Bandyopadhyay PK, Göcmen B. First record of ectoparasitic ciliates, of genus *Trichodina* (Ciliophora: Trichodinidae) parasiting cultured Oranda Gold fish (*Carassius auratus auratus*) in India. Zootaxa. 2017; 4319(1):128-142

[142] Fariya N, Abidi R, Chauhan UK. Morphological and molecular description of a new species, *Trichodina cirhinii* sp. nov. (Ciliophora: Trichodinidae), infesting native freshwater fish *Cirrhinus mrigala*. Journal of Biological Sciences and Medicine. 2017;3(1):10-17

[143] Akşit D, Falakali-Mutaf B, Göçmen B, Gürelli G. A preliminary observations on *Trichodina* sp. (Ciliophora: Peritricha) on the gills of limpets (*Patella* spp.) in Antalya (Turkey). North-Western Journal of Zoology. 2008;4(2):295-299

[144] van As LL, Basson L, van As JG. Mobiline peritrichs (Ciliophora) collected from the gills of African limpets. Acta Protozoologica. 2017;56:245-254

[145] Wahl M. Marine epibiosis. I.Fouling and antifouling: some basic aspects. Marine Ecology Progress Series.1989;58:175-189

[146] Wahl M, Mark O. The predominantly facultative nature of epibiosis: experimental and observational evidence. Marine Ecology Progress Series. 1999;187:59-66 [147] Xu Z. The abundance of epizoic ciliate *Epistylis daphniae* related to their host *Moina macrocopa* in an urban stream. Journal of Invertebrate Pathology. 1992;60:197-200

[148] Schuwerack PM, Lewis JW, Jones PW. Pathological and physiological changes in the South African freshwater crab *Potamonautes warreni* Calman induced by microbial gill infections. Journal of Invertebrate Pathology. 2001;77:269-279

[149] Puckett GL, Carman KR. Ciliate epibiont effects on feeding, energy reserves, and sensitivity to hydrocarbon contaminants in an estuarine harpacticoid copepod. Estuaries. 2002;25(3):372-381

[150] Gregori M, Fernandez-Leborans G, Roura Á, González ÁF, Pascual S. Description of a new epibiotic relationship (Suctorian–Copepoda) in NE Atlantic waters: from morphological to phylogenetic analyses. Acta Zoologica. 2016;97(2):165-176

[151] Burris ZP, Dam HG. Deleterious effects of the ciliate epibiont *Zoothamnium* sp. on fitness of the copepod *Acartia tonsa*. Journal of Plankton Research. 2014;36(3):788-799

[152] Fernandez-Leborans G, Gabilondo R. Taxonomy and distribution of the hydrozoan and protozoan epibionts on *Pagurus bernhardus* (Linnaeus 1758) (Crustacea Decapoda) from Scotland. Acta Zoologica. 2006;87(1):33-48

[153] Hanamura Y. Seasonality and infestation pattern of epibiosis in the beach mysid *Archaemysis articulata*. Hydrobiologia. 2000;427:121-127

[154] Utz LRP. Growth of the peritrich
epibiont *Zoothamnium intermedium*Precht, 1935 (Ciliophora, Peritrichia)
estimated from laboratory experiments.
Brazilian Journal of Biology.
2008;68:441-446

[155] Bickel SL, Tang KW, Grossart HP.
Ciliate epibionts associated with crustacean zooplankton in German lakes: distribution, motility, and bacterivory. Frontiers in Microbiology.
2012;3(243):1-11

[156] Jones S, Carrasco NK, Perissinotto R, Vosloo A. Association of the epibiont *Epistylis* sp. with a calanoid copepod in the St Lucia Estuary South Africa. Journal of Plankton Research. 2016; 38(6):1404-1411

[157] Utz LRP, Coats DW. Spatial and temporal patterns in the occurrence of peritrich ciliates as epibionts on calanoid copepods in the Chesapeake Bay, USA. Journal of Eukaryotic Microbiology, 2005;52(3):236-244

[158] Souissi A, Souissi S, Hwang JS. The effect of epibiont ciliates on the behavior and mating success of the copepod *Eurytemora affinis*. Journal of Experimental Marine Biology and Ecology. 2013;445:38-43

[159] Mackie RI. Mutualistic fermentative digestion in the gastrointestinal tract: diversity and evolution. Integrative and Comparative Biology. 2002;42:319-326

[160] Schmidt GD, Roberts LS. Foundations of parasitology. 9 ed. New York: The McGraw-Hill Company; 2013. 670p

[161] Vďačný P. Evolutionary associations of endosymbiotic ciliates shed light on the timing of the marsupial-placental split. Molecular Biology and Evolution. 2018;35(7):1757-1769

[162] Corliss JO. The ciliated protozoa; characterization, classification and guide to the literature. 2nd ed. Oxford: Pergamon Press; 1979. 455p

[163] Williams AG, Coleman GS. The rumen protozoa. New York: Springer; 1992. 442p [164] Kumar S, Pitta DW. Revolution in Rumen Microbiology. In: Puniya AK, Singh R, Kamra, DN, editors. Rumen Microbiology, From Evolution to Revolution. New Delhi: Springer; 2015. p. 357-379

[165] Mizrahi I. Rumen symbioses. In: Rosenberg E, DeLong F, Lory S,
Stackebrandt E, Thompson F, editors.
The Prokaryotes. Berlin Heidelberg:
Springer Science & Business Media;
2013. p. 533-544

[166] Nagaraja TG. Microbiology of the rumen. In: Millen DD, Arrigoni MB, Pacheco RDL, editors. Rumenology. Switzerland: Springer; 2016. p. 39-62

[167] Imai S. Phylogenetic taxonomy of rumen ciliate protozoa based on their morphology and distribution. Journal of Applied Animal Research. 1998;13:17-36

[168] Paul SS, Dey A. Domesticated rare animals (yak, mithun and camel): rumen microbial diversity. In: Puniya AK, Singh R, Kamra DN, editors. Rumen Microbiology: From Evolution to Revolution. New Delhi: Springer; 2015. p. 31-36

[169] Wright ADG, Lynn DH. Maximum ages of ciliate lineages estimated using a small Subunit rRNA molecular clock: crown eukaryotes date back to the Paleoproterozoic. Archiv für Protistenkunde. 1997;148:329-341

[170] Pianka ER. Evolutionary ecology. Sixth edition. San Francisco: Addison Wesley Longman; 2000. 512p

[171] Fenchel T. Ecology of protozoa the biology on free-living phagotrophic protists. New York: Springer; 1987. 197p

[172] Fernandes NM, Schrago CG. A multigene timescale and diversification dynamics of Ciliophora evolution.Molecular Phylogenetics and Evolution.2019;139:106521 [173] Gentekaki E, Kolisko M, Gong Y, Lynn DH. Phylogenomics solves a long-standing evolutionary puzzle in the ciliate world: the subclass Peritrichia is monophyletic. Molecular Phylogenetics and Evolution. 2017;106:1-5

[174] Vďačný P, Érseková E, Šoltys K, Budiš J, Pecina L, Rurik, I. Co-existence of multiple bacterivorous clevelandellid ciliate species in hindgut of woodfeeding cockroaches in light of their prokaryotic consortium. Scientific Reports. 2018;8(1):1-13

[175] Jackson A. The evolution of parasite genomes and the origins of parasitism. Parasitology. 2015;142:S1-S5

[176] Luong L, Mathot K. Facultative parasites as evolutionary steppingstones towards parasitic lifestyles. Biology Letters. 2019;15

[177] Poulin R, Randhawa H. Evolution of parasitism along convergent lines: from ecology to genomics. Parasitology. 2013;142:S6-S15

[178] Viney M. How can we understand the genomic basis of nematode parasitism?. Trends in Parasitology.2017;33(6):444-452

[179] Baker J. The origins of parasitism in the protists. International Journal of Parasitology. 1994;24:1131-1137

[180] Kreier J, Baker JR. Parasitic Protozoa. Winchester, Massachusetts, USA: Allen & Unwin; 1987. 256p

[181] Leung TLF. Fossils of parasites: what can the fossil record tell us about the evolution of parasitism?. Biological Reviews. 2017;92(1):410-430

[182] Rataj M, Vďačný P. Multi-gene phylogeny of *Tetrahymena* refreshed with three new histophagous species invading freshwater planarians. Parasitology Research. 2020;119:1523-1545 [183] Strüder-Kypke M.C, Wright ADG, Jerome CA, Lynn DH. Parallel evolution of histophagy in ciliates of the genus *Tetrahymena*. BMC Evolutionary Biology. 2001;(1)5

[184] Clamp JC, Lynn DH. Investigation the biodiversity of ciliates in the 'Age of Integration'. European Journal of Protistology. 2017;61:314-322

[185] Jamy M, Foster R, Barbera P, Czech L, Kozlov A, Stamatakis A, Bending G, Hilton S, Bass D, Bur F. Long read metabarcoding of the eukaryotic rDNA operon to phylogenetically and taxonomically resolve environmental diversity. Molecular Ecology Resources. 2020;20(2):429-443

[186] Pylro V, Roesch L. The Brazilian microbiome. Current status and perspectives. Switzerland: Springer International Publishing; 2017. 128p

[187] Santoferrara L, Burki F, Filker S, Logares R, Dunthorn M, McManus GB. Perspectives from ten years of protist studies by high-throughput metabarcoding. Journal of Eukaryotic Microbiology. 2020;67:612-622

[188] Mayr E. Systematics and the origin of species from the viewpoint of a zoologist. New York: Columbia University Press; 1942. 334p

[189] Rissler LJ, Apodaca JJ. Adding more ecology into species delimitation: ecological niche models and phylogeography help define cryptic species in the black salamander (*Aneides flavipunctatus*). Systematic Biology. 2007;56:924-942

[190] Knowles LL, Carstens B. Delimiting species without monophyletic gene trees. Systematic Biology. 2007;56:887-895

[191] Pons J, Barraclough TG, Gomez-Zurita J, Cardoso A, Duran DP, Hazell S, Kamoun S, Sumlin, WD,

Vogler AP. Sequence- based species delimitation for the DNA taxonomy of undescribed insects. Systematic Biology. 2006;55:595-609

[192] Yang Z, Rannala B. Bayesian species delimitation using multilocus sequence data. PNAS. 2010;107:9264-9269

[193] Zhang J, Kapli P, Pavlidis P, Stamatakis A. A general species delimitation method with applications to phylogenetic placements. Bioinformatics. 2013;29:2869-2876

[194] Zhao Y, Yi Z, Gentekaki E, Zhan A, Al-Farraj SA, Song W. Utility of combining morphological characters, nuclear and mitochondrial genes: an attempt to resolve the conflicts of species identification for ciliated protists. Molecular Phylogenetics and Evolution. 2016;94:718-729

[195] Shazib SUA, Vd'ac ny'P, Kim JH, Jang SW, Shin MK. Molecular phylogeny and species delimitation within the ciliate genus *Spirostomum* (Ciliophora, Postciliodesmatophora, Heterotrichea), using the internal transcribed spacer region. Molecular Phylogenetics and Evolution. 2016;102:128-144

[196] Mehlhorn H. Animal Parasites. Diagnosis, treatment, prevention. Switzerland; Springer International Publishing ; 2016. 730p

[197] Ohtsuka S, Suzaki T, Horiguchi T, Suzuki N, Not F. Marine Protists. Diversity and dynamics. New York: Springer; 2015. 637p

