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Chapter

Probiotics as Potential Antimicrobials for the Treatment of Infections: current Reality or Remote Future?

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Abstract

Probiotics are microorganisms that live in symbiosis with the human body. The intake of probiotics in adequate amounts can improve biological functions bringing improvements in the health of the host. Many studies have demonstrated the indisputable antimicrobial activity of probiotics and their potential for an alternative treatment of infections. Nevertheless, the forms of encapsulation, as well as clinical trials on the clinical use of these microorganisms as a recognized and well-established protocol, are still incipient. In this chapter, we provide a general approach to the topic and point to future directions in the probiotics field for this purpose. Moreover, microbial resistance is a current public health problem and the search for new therapeutic alternatives is urgent. Probiotics and other natural therapies have been considered very promising. The approaches of future research should focus mainly on the isolation of new probiotic microorganisms, the definition of inoculum, forms of encapsulation for controlled delivery, and clinical trials for the definition of doses and mechanism of action in the fight against infections.

Keywords: probiotics, pharmacology, antimicrobial activity, microbiota, biomaterials

1. Introduction

The human body is inhabited by numerous microorganisms, including bacteria, fungi, viruses, and protozoa, which represent the human microbiota. Compared to the number of human cells, there is a much larger number of microorganisms [1], which affect the host's physiological functions in different ways [2]. After a long time of science focusing on pathogenic microorganisms that cause human diseases, the interest was also turned to those that provide benefits to the organism, such as probiotics.

The first time that probiotics were mentioned and defined was in 1965 and the concept was restricted to substances produced by bacteria that promote the growth of other bacteria [3]. In 2001, the Food and Agriculture Organization of the United Nations (FAO) updated the concept of probiotics for any living microorganisms that provide health benefits to the host when ingested in adequate quantities [4]. The most widely used and studied probiotics for human health benefits are generally gram-positive bacteria that function primarily as modulators and maintainers of gut health [5]. Examples of widely studied probiotics such as *Lactobacillus*, *Bifidobacterium*, *Escherichia*, *Enterococcus*, *Bacillus* e *Streptococcus* [6].

The commensal intestinal microbiota is related to important functions for maintaining the health of the organism, such as increased resistance against infections, differentiation of the immune system, and synthesis of nutrients [7]. Nevertheless, recent studies have shown that the benefits of probiotics for human health go beyond [8], including anti-inflammatory activities [9], anti-tumor activities [10], antioxidant [11], antimicrobial [12] and modulation of the microbiome [13]. Although, research on the antimicrobial activity of probiotic microorganisms remains incipient and its clinical applicability for the treatment of infections has not been fully explored [14].

Infections have been commonly treated with antibiotics. However, the unrestrained and irrational use of these drugs can range from individual harms, such as specific adverse effects of the drug for the patient, to serious public health problems, such as the selection of drug-resistant microorganisms [15]. Likewise, research on alternative therapies for the treatment of infectious diseases should be encouraged and the field of probiotic microorganisms is very promising. Therefore, in this chapter, we will discuss the current reality of treating infections using probiotic microorganisms and/or their by-products as well as the prospects for this therapy to become a reality in current medicine.

2. Probiotic microorganisms

In 1965, Lilly and Stillwell first used the term probiotic, describing substances that one organism secretes and can stimulate the growth of another [16]. Nonetheless, its use goes back to millennia, as the use of recipes with fermented milk by Greeks and Romans. There are also reports of the use of sour milk in the bible. Thus, it is observed that the benefits of the use of probiotics to human health have been discussed for millennia [17].

These microorganisms, when colonizing the gastrointestinal tract, interact directly with the cells of the immune system, playing an important role in the maintenance and balance of the immune system [18]. The mechanisms of action of probiotics are complex and, in most cases, likely, more than one mechanism occurs simultaneously. The main biological pathways of action include increased epithelial barrier, inhibition of microbial adhesion and competitive exclusion of pathogenic microorganisms in addition to the production of antimicrobial substances, modulation of the immune system, maintenance of normal levels of short-chain fatty acids, and regulation of intestinal absorption of electrolytes [19].

The word “probiotic” comes from Greek and means “for life” [20]. Probiotics are viable live microorganisms, bacteria, and yeasts, which confer benefits to the health of the host when ingested in adequate concentration. Probiotic microorganisms, in general, are part of the intestinal microflora, but can also be found in ecological environments. Many factors need to be considered before isolating a potential probiotic microorganism. Initially, it is necessary that the strain is not pathogenic and shows some type of behavior that reflects in biological activities

Genus	Specie	Main source	Reference
<i>Lactobacillus</i>	<i>L. casei</i> , <i>L. bulgaricus</i> , <i>L. acidophilus</i> , <i>L. rhamnosus</i> , <i>L. reuteri</i> , <i>L. pantarum</i> and <i>L. johnsonii</i>	Dairy and human gastrointestinal tract	[26, 27]
<i>Bifidobacterium</i>	<i>B. animalis</i> , <i>B. bifidum</i> , <i>B. breve</i> , <i>B. infantis</i> , <i>B. lactis</i> , <i>B. longum</i>	Human, Dog, Primate, Pig, Cow and Horse gastrointestinal tract	[28–30]
<i>Streptococcus</i>	<i>Streptococcus thermophilus</i>	Dairy	[31, 32]
<i>Enterococcus</i>	<i>Enterococcus faecalis</i> , <i>Enterococcus faecium</i>	Human, Cow and Pig gastrointestinal tract	[33–35]
<i>Pediococcus</i>	<i>Pediococcus pentosaceus</i> , <i>Pediococcus acidilactici</i>	Dry quark and rice wine	[36]

Table 1.
 Main probiotic microorganisms that are cited in the literature for human health benefits.

for the benefit of the host [21]. Besides, it is important to consider that the probiotic action is not universal for all species and does not work the same in all tissues of the body [22].

Lactic acid bacteria (*Lactococcus*, *Lactobacillus*, *Streptococcus*, and *Enterococcus*) are among the most well-known microorganisms, used and studied by man for probiotic purposes. In addition to these, we can include, *Bifidobacterium* and *Saccharomyces* species, a non-pathogenic yeast [23–25]. **Table 1** summarizes the main probiotic microorganisms mentioned in the literature for the benefit of human health.

Microorganisms can produce lactic acid from different carbon sources, as well as release secondary metabolites, including bacteriocins, exopolysaccharides, and enzyme complexes with antimicrobial properties preventing the installation and growth of other microorganisms [21, 37]. The mechanisms involved in the action of these microbial products are well understood concerning the benefits generated to the human intestine. However, the use of probiotics for alternative antimicrobial therapy against infections, in general, is incipient, although promising. Subsequently, we will discuss how probiotics can affect a human microbiota, ways of encapsulation, and their main uses for treating infections.

3. Probiotics affect the microbiota

In recent years, several findings have revealed benefits in the administration of probiotics, ranging from direct inhibition of pathogenic microorganisms to improvements in host immune system functions [38–43].

Despite a large number of studies with probiotics, most efforts are focused on understanding the benefits for the intestinal health of the host. Probiotics can exert their antimicrobial activity through different mechanisms of action. Generally, it has been reported that these microorganisms control/kill the pathogenic microbiota through the production of inhibitory substances such as bacteriocins and hydrogen peroxide (capable of inhibiting Gram-negative and Gram-positive pathogenic bacteria); interference at adhesion sites; competition for nutrients in the microenvironment, among others [41, 42, 44, 45]. Besides, there is also the modulation of the immune system, which also plays a role in the control of infections, which can occur in several ways: increased non-specific phagocytic activity through the activation of macrophages [9, 45, 46].

Several probiotic species are widely used in research showing its benefits to the host [46, 47]. Among these benefits, antimutagenic properties [48], anticarcinogenic properties [49–51], antidiarrheal drugs [52–54], system stimulation [55], prevention of atopic dermatitis [56–58], reduced blood cholesterol [59, 60].

Therefore, the use of probiotics has been considered a promising strategy for the prevention and control of various infectious diseases [38–40, 42, 43, 48, 61–63].

Some studies have also demonstrated the importance of probiotics relating to multidrug-resistant bacteria [64]. Multidrug-resistant bacteria, such as vancomycin resistant enterococcus (VRE), carbapenemase-producing enterobacteria (CPE), and extended-spectrum beta-lactamase (ESBL)-carrying strains, represent a major public health issue because they are potential pathogens associated with a high mortality rate [64, 65]. Prevention strategies could be based on the use of probiotics to prevent the colonization of the colon microbiota. Transient colonization with multidrug-resistant bacteria could result in the transfer of antibiotic resistance genes in commensals or potential pathogens, resulting in the persistence of the resistance gene in the microbiota, which could be responsible for an increased risk of lethal infection due to the delay in introducing an effective antibiotic [64, 66]. Surprisingly, clinical cases demonstrated that fecal transplantation was able to cause decolonization of microbiota of naturally resistant Extended Spectrum β -lactamase (ESBL) bacterial strains [67–69]. Furthermore, there are reports that the composition of the microbiota of hospitalized patients is related to the susceptibility to colonization with multiresistant bacteria. The use of probiotic microorganisms such as *L. plantarum* or *L. fermentum* reduced the colonization of resistant pathogens such as *Acinetobacter baumannii*, *Pseudomonas aeruginosa* or *Candida albicans* [70, 71]. Nevertheless, an in vitro study showed that the culture supernatants of *Clostridium butyricum*, *C. difficile*, *Clostridium perfringens*, *Enterococcus faecium*, and *L. plantarum* were able to suppress the growth and transmission of gene resistance of bacteria carrying ESBL and Carbapenemase-Producing Enterobacteriaceae (CPE) [64]. It is undeniable that both colonization by probiotics and the use of their by-products have great potential in the treatment and prevention of infections, however these properties are still scarcely explored.

Vancomycin-resistant enterococci (VRE) seem less adapted to survival in the intestinal microbiota. Thus, these pathogens are more susceptible to decolonization when compared to other multiresistant bacteria. The intestinal microbiota in patients suffering from hematologic malignancies is less frequently colonized by VRE in the presence of *Barnesiella* [7]. In vivo evidence demonstrates that supplementing resident microbiota with *Barnesiella* or *Lactobacillus paracasei* CNCM I-3689 reduces VRE colonization in mice [72, 73]. In clinics, a case report showed VRE decolonization after fecal grafting for the treatment of *C. difficile colitis* [64].

The clinical use of probiotics in the treatment of infection is challenging the thinking of encapsulation for delivery. It is necessary to maintain the viability of these microorganisms long enough to compete with pathogenic microorganisms. Next, we'll discuss different potential encapsulation modalities for delivery.

4. Biomaterials for encapsulation of probiotics

The drug delivery systems through liposomes, micelles, carbon nanotubes, and dendrimers allowed the increase of therapeutic efficacy, reduction of toxicity, sustained and controlled release [74, 75]. The biotechnology industry has been aiming at the development of techniques for encapsulating probiotics, since their health benefits are indisputable. However, unlike inert substances, probiotics are live microorganisms, which in a way is a challenge in their manufacture, as they must be

kept in a live/viable state during the processing, storage, and gastrointestinal transit steps to ensure its effectiveness on target sites [75].

The encapsulation technique consists of a set of physical–chemical or mechanical processes in which solid, liquid or gaseous materials are packaged, trapped in another material, usually hydrocolloidal materials, resulting in the formation of particles that vary in shape and size (from nanometer to millimeter) [76–78].

The encapsulated part is named core material, internal phase, active agent, or payload phase, and the encapsulating agent is called the carrier, shell, external phase, or matrix [78]. From these components, the encapsulation forms different structures: reservoir (where the core is surrounded by a shell), matrix (the internal phase is distributed on the surface), or coated matrix, in which matrix is surrounded by an additional coating layer [78].

The use of nanoencapsulation techniques ($<1\ \mu\text{m}$) is not feasible because of the size of the bacteria (1 to 5 μm) [76]. On the other hand, it is possible to obtain microcapsules using other techniques [79, 80]. The first microencapsulation techniques applied were spray drying, freeze-drying or lyophilization, foam drying, and fluidized bed drying [78]. Other techniques used are extrusion, emulsion technologies, gel particles, coacervation, and electrospraying [76, 78, 81].

The encapsulation of probiotics can be made using natural polymers, such as polysaccharides, polypeptides, and polynucleotides, or synthetic polymers. Conventionally, three processes are involved in encapsulation. First, the cells must be incorporated into a matrix, which can be liquid (by dissolution or dispersion) or solid (by agglomeration or adsorption). Then the solution must be dispersed (liquids) or sprayed (solids) on the surface. The last process aims to stabilize the structure, through polymerization, gelling, solidification, evaporation, coacervation, or coalescence [79].

Before choosing the technique, it is necessary to consider some important criteria: the relationship between the composition of the material, type of bacteria, temperature and pH of the medium, as well as the host's immune response. The biocompatibility of the material used in the encapsulation is directly related to the viability of the probiotics, which must remain equal to or greater than 10^7 CFU/ml [82, 83]. Therefore, factors such as solubility, digestibility, and release capacity must also be carefully analyzed [84]. Consequently, it is expected that the biomaterial will be able to form an effective protective barrier to resist pH variations and ensure the survival of bacteria, without causing damage to the host organism. Next, some biomaterials commonly used for the encapsulation of probiotics will be discussed.

4.1 Alginate

It is a natural polysaccharide composed of alginic acid (β -D-manuronic acid and L-gunoronic acid), obtained through some types of seaweed (laminaria). It is considered the most used material for the encapsulation of probiotics. Calcium alginate is preferable because it associates the biocompatibility of the material with a simple and low-cost technique. However, some disadvantages are attributed, such as the high porosity of the particles, which can reduce the protection of cells in the matrix [85] and sensitization in an acid medium [86]. Nonetheless, the association of alginate with other polymeric components or the addition of additives to the surface of the particles can easily overcome these defects [87]. Alginate spheres reach the intestine satisfactorily, without undergoing significant degradation by stomach acids [88]. Besides, the structural configuration of the probiotic encapsulation to alginate is comparable to the beneficial biofilm formation by probiotics bacteria [89].

4.2 Chitosan

It is a biodegradable copolymer obtained from the deacetylation of chitin (polysaccharide) present in the crustacean exoskeleton. It consists of units of D-glucosamine, capable of forming polymeric networks through Cross-link due to the presence of free amino groups. It is commonly found associated with another polymer since studies have shown that its isolated use in the matrix does not contribute to the maintenance of cell viability [86]. When applied in multilayers together with calcium alginate, have shown promising results, where the particles are coated with chitosan forming polyelectrolyte complexes that reinforce the alginate structure [89, 90]. Although its use is relatively common, care should be taken when choosing this biomaterial to encapsulate some types of bacteria, such as those from lactic acid, since chitosan can cause their inhibition [91]. Additionally, its solubility is directly related to the pH of the medium, being insoluble at pH higher than 5.5 [92], which may result in null or insufficient release.

4.3 Carrageenan

These natural polymers are extracted from red algae (Rhodophyceae) and are commonly used as additives in the food industry. Three variables are found: (kappa) κ -carrageenan, (iota) ι -carrageenan and (lambda) λ -carrageenan [93]. The use for encapsulation of probiotics is based on the sol-gel transition characteristics of the types κ -carrageenan and ι -carrageenan [93]. The dissolution of the polymer occurs after heating in a temperature range between 40 and 45°C, at which point the bacteria must be incorporated. Subsequently, the solution is stored at room temperature allowing gelation to occur, forming a three-dimensional gel [87]. Studies have shown that bacteria have been kept viable, demonstrating a promising effect of the use of carrageenan [94–96].

4.4 Gellan gum

This polysaccharide comes from the bacterium *Sphingomonas elodea*. It is composed of glucose (60%), rhamnose (20%), and gluconic acid (20%). These microbial polysaccharides are considered water-soluble polymers and are commonly used as solidifying, gelling, or stabilizing agents [97]. Other microbial polysaccharides, such as arabic gum, jambilam, and xanthan gum, when associated with gelam gum, become very promising for the encapsulation of probiotics [98].

4.5 Cellulose acetate phthalate (CAP)

They are polysaccharides derived from plants that have important characteristics, such as insolubility at pH below 5 and solubility at pH above 6. Thus, it can be used effectively to enable encapsulated probiotics to reach the intestine and be released gradually without being altered by stomach pH [99]. CAP does not form a gel, therefore, it is used as a coating agent for other biomaterials.

4.6 Starches

Another polysaccharide extracted from plants. Resistance to degradation by pancreatic enzymes present in the small intestine is an interesting characteristic that justifies its use as a probiotic delivery agent, guaranteeing the viability of bacteria when reaching the large intestine [86, 100]. It is commonly associated with alginate or carrageenan to form resistant capsules or gels [87, 101].

4.7 Synthetic polymers

The use of synthetic material for encapsulating probiotics requires that it must be a biodegradable material and provide bacterial viability. An example of these polymers is PVA - poly (vinyl alcohol), characterized by being soluble in water, chemically stable, and of low cost. Studies have shown that its use the use of this material alone [102] or associated with other biomaterials [103] is satisfactory while maintaining the viability of probiotic microorganisms. Poly(lactic acid) (PLA), poly(glycolic acid) (PGA), poly(lactic-co-glycolic acid) (PLGA), poly(ethylene glycol) (PEG), poly(ethylene oxide) (PEO), and poly(vinyl pyrrolidone) (PVP) are other synthetic polymers used for encapsulation of probiotics synthetic polymers used for encapsulation of probiotics [84]. The use of these polymers is linked to the technique of producing fibers through electrospinning.

All of these alternatives mentioned aim to encapsulate probiotics for intestinal delivery. Although they can be applied to other tissues of the body, the data in the literature are incipient and need to be better analyzed for application in the treatment of other infections, such as those discussed in the next topic.

5. Prevention and treatment of infection with probiotics

The resistance of pathogenic microorganisms to synthetic antimicrobials and, consequently, the ineffectiveness of conventional therapies and recurrence of infections reflects the need to seek alternative and/or supplementary methods in the treatment protocols [104]. Probiotics are one of the methods and are considered promising, as they provide satisfactory results when facing infections of a bacterial, fungal and viral nature, whether in the intestinal, urinary, respiratory, female genital tracts, and in the oral cavity. In addition, it is safe and does not promote adverse effects on the human body [105, 106]. **Figure 1** schematizes the delivery of microencapsulated probiotic microorganisms in an epithelium colonized by pathogenic microorganisms for the treatment of an infection.

One indication of probiotics refers to the treatment of *Helicobacter pylori* infection, which is one of the most common chronic bacterial infections in humans, with approximately 4.4 billion infected individuals worldwide in 2015 [107]. *H. pylori* infection is associated with the development of gastric cancer, which represents one of the main global causes of cancer-related deaths [108, 109]. The treatment of *H. pylori* infection is based on its eradication, with the use of antibiotics, such as amoxicillin, clarithromycin, and metronidazole. However, antibiotic therapy promotes an imbalance in the intestinal microbiota and increased levels of resistant bacteria [110], as well as species associated with persistent gastric inflammation and gastric carcinogenesis [111]. This situation justifies probiotic supplementation, aiming to reduce undesirable changes in the intestinal microbiota, promote the eradication of *H. pylori* [108, 112], produce significant improvements in gastrointestinal symptoms, and, consequently, in the quality of life of individuals [108, 109]. The combination of probiotics with antibiotic therapy for the eradication of *H. pylori* was suggested in the Thailand Consensus, held in 2015 [110].

Probiotics, in addition to reducing the density of *H. pylori*, promote immune responses with reduced inflammatory status [112–115], significantly reduce adverse events related to antibiotic treatment, and improve patient compliance [109, 116]. Despite this evidence, it was highlighted in the Thailand Consensus, that most studies that evaluated the effects of probiotics on the eradication of *H. pylori* are of poor quality, compromising general recommendations. It has been suggested that

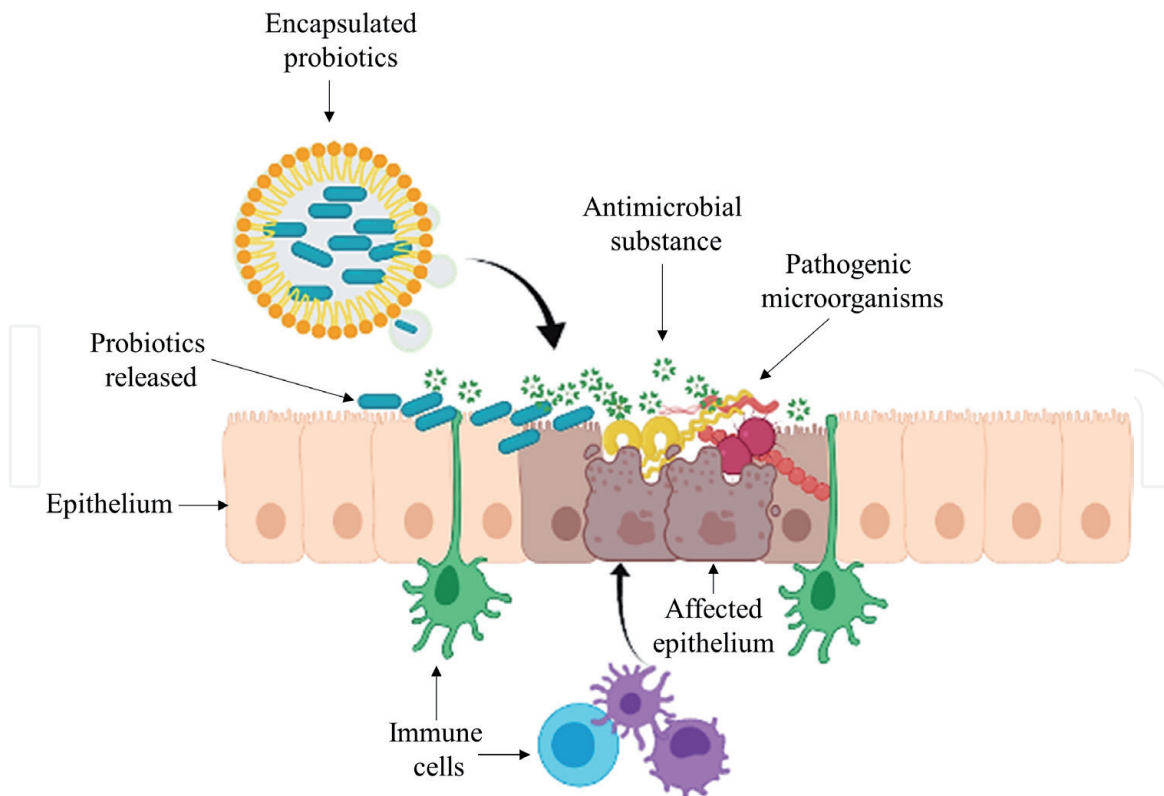


Figure 1.

Microencapsulated probiotics being delivered for the treatment of infection in epithelial tissue. Note that after the exit of the microorganisms from the micelle there is the colonization of the region and release of bacteriocins that in addition to acting as antimicrobials, stimulate the host's immune system.

further studies should be carried out to determine the best strain, the ideal dose, the duration of treatment, effectiveness, contraindications, and cost–benefit [110].

Probiotics are also indicated to reduce or prevent diarrhea associated with antibiotics and infections by *Clostridium difficile*, common in hospitalized patients [117, 118] and the elderly [119]. In children, its effectiveness in preventing antibiotic-associated diarrhea [120] and in treatment for acute gastroenteritis has not been confirmed, despite reducing the duration of hospitalization [121]. The use of probiotics in the treatment of infections by *Enterobacteriaceae* producing extended-spectrum β -lactamase has also been discussed. However, the results are incipient to indicate its use in eradication therapy in patients with prolonged intestinal transport of *Enterobacteriaceae* [122].

Some studies have suggested that supplementation with probiotics can improve the host's innate and acquired immune response, promoting a protective effect against respiratory infections [46, 123, 124]. The increase in the population of T cells, more precisely CD4 and CD8, is one of the most important mechanisms of the anti-infection effect of probiotics [125, 126]. Oral probiotics, when used in children, in addition to improving intestinal microecological balance can reduce the frequency of respiratory tract infections [89], mostly caused by viruses, such as the coronavirus [127], influenza [128], and bacteria, such as *Streptococcus pneumoniae*) [129]. Several studies have found that probiotics reduce episodes of acute respiratory tract infections in children, adults, the elderly, and athletes [89, 125, 126, 130, 131], proving its beneficial effect in these populations, with no reports of adverse effects in children [131].

The high recurrence of urinary tract infections in children [132] and the possibility of developing microbial resistance to drugs used against this disease have justified research with non-antibiotic alternatives, such as the use of probiotics for the prevention of recurrent urinary infections in this population

[133]. Probiotics appear to prevent recurrent urinary infections by contributing to the recovery and maintenance of microbiomes, by reducing the adherence, growth, and colonization of infectious pathogens in the urinary tract, in addition to improving host defenses, and attenuating or eliminating inflammation [105, 134–144]. Unlike the beneficial role of the use of probiotics in preventing urinary infections in children [132], it appears to have no protective effect in adults with severe spinal injuries, who have recurrent urinary infections [145], as well as in healthy young women [146].

Regarding the genital tract, the administration of probiotics, alone or as adjunctive therapy to the use of conventional antimicrobials, demonstrates success in the treatment of infections such as bacterial vaginosis and vulvovaginal candidiasis, common and recurrent infections in women of reproductive age. These infections that produce abnormal vaginal discharge, itching, vulvar odor, are associated with important health complications, such as the increased transmission of sexual infections, risk of premature birth, and pelvic inflammation, with negative impacts on quality of life [138, 147–149].

In infections that affect the mouth, candidiasis is also one of the most prevalent diseases, especially when local factors are predisposing the installation of the infection. Probiotics have been suggested for the treatment of oral candidiasis because they reduce the population of *Candida* spp. [150], the course of treatment with conventional antifungal therapies [151], and the severity of clinical manifestations of the infection associated with prosthetic stomatitis [152, 153], including asymptomatic [62]. Besides, the immunological and antimicrobial potential of probiotics also can be used in the treatment of periodontal disease killing periodontopathogens, as *Porphyromona gingivalis*, and promoting the expression of some favorable immunoregulatory effects [154]. In summary, probiotics favor oral health, increasing fluids in the mucosa, reducing the accumulation of dental biofilm and gingival inflammation, improving the clinical signs characteristic of periodontal infection, such as redness and swelling [63, 155, 156].

Studies show beneficial effects of the combination of probiotics in the treatment regimen for different infections, with improvements in the clinical condition and patient adherence to treatment. Although, researchers warn of the need for further studies to define the best treatment protocol, including the determination of effects, contraindications, and cost–benefit [110].

6. Concluding remarks

Today's society is experiencing a public health problem related to an exponential increase in microbial resistance, compared to the slow evolution of new drug development. The human organism is attacked daily by countless pathogenic microorganisms, many of which cause lethal infections. The use of alternative therapies, alone or as an adjunct to antibiotics, is a reality. Concerning the use of probiotics, its effectiveness in modifying the microbial is unquestionable, either by the production of antimicrobial bacteriocins or by the modulation of the immune system. Nonetheless, there is no consensus or standardization for the clinical use of probiotics for the treatment of infectious diseases, except its use for the recomposition of the intestinal microbiota. Moreover, two important challenges need to be overcome: the standardization of carriers to deliver these microorganisms effectively to the treatment site and the definition of important factors, such as the mechanism of action, standardization of inoculum, and therapeutic protocols, based clinical trials. Thus, although promising, widespread antimicrobial therapy with probiotics is not yet a reality for clinical practice.

Conflict of interest

The authors declare no conflict of interest.

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References

- [1] Sender R, Fuchs S, Milo R. Revised Estimates for the Number of Human and Bacteria Cells in the Body. *PLoS biology*. 2016;14(8):e1002533. DOI: 10.1371/journal.pbio.1002533
- [2] Altveş S, Yildiz HK, Vural HC. Interaction of the microbiota with the human body in health and diseases. *Bioscience of microbiota, food and health*. 2020;39(2):23-32. DOI: 10.12938/bmfh.19-023
- [3] Lilly DM, Stillwell RH. Probiotics: growth-promoting factors produced by microorganisms. *Science*. 1965;147:747-748. DOI: 10.1126/science.147.3659.747
- [4] Food and Agriculture Organization and World Health Organization. FAO and WHO to hold first global forum of food safety regulators. [Internet]. 2001.1(101). Available from: https://fao.org/WAICENT/OIS/PRESS_NE/PRESSENG/2001/pren01101.htm [Accessed: 2021-05-22]
- [5] Marco ML, Pavan S, Kleerebezem M. Towards understanding molecular modes of probiotic action. *Curr Opin Biotechnol*. 2006;17(2):204-10. DOI: 10.1016/j.copbio.2006.02.005
- [6] Gupta V, Garg R. Probiotics. *Indian J Med Microbiol*. 2009;27(3):202-9. DOI: 10.4103/0255-0857.53201
- [7] Ubeda C, Pamer EG. Antibiotics, microbiota, and immune defense. *Trends Immunol*. 2012;33(9):459-66. DOI: 10.1016/j.it.2012.05.003
- [8] Tang C, Lu Z. Health promoting activities of probiotics. *J Food Biochem*. 2019;43(8):e12944. DOI: 10.1111/jfbc.12944
- [9] Plaza-Díaz J, Ruiz-Ojeda FJ, Vilchez-Padial LM, Gil A. Evidence of the Anti-Inflammatory Effects of Probiotics and Synbiotics in Intestinal Chronic Diseases. *Nutrients*. 2017;9(6):555. DOI: 10.3390/nu9060555.
- [10] Marinelli L, Tenore GC, Novellino E. Probiotic species in the modulation of the anticancer immune response. *Semin Cancer Biol*. 2017;46:182-190. DOI: 10.1016/j.semcancer.2017.08.007.
- [11] Mishra V, Shah C, Mokashe N, Chavan R, Yadav H, Prajapati J. Probiotics as potential antioxidants: a systematic review. *J Agric Food Chem*. 2015;63(14):3615-26. DOI: 10.1021/jf506326t
- [12] Baumgardner RM, Berreta A, Kopper JJ. Evaluation of commercial probiotics for antimicrobial resistance genes. *Can Vet J*. 2021;62(4):379-383.
- [13] Quigley EMM. Prebiotics and Probiotics in Digestive Health. *Clin Gastroenterol Hepatol*. 2019;17(2):333-344. DOI: 10.1016/j.cgh.2018.09.028
- [14] Silva DR, Sardi JCO, Pitanguí NS, Roque SM, Silva ACB, Rosalen PL. Probiotics as an alternative antimicrobial therapy: Current reality and future directions. 2020:104080. DOI: 10.1016/j.jff.2020.104080
- [15] Yang H, Sun Y, Cai R, Chen Y, Gu B. The impact of dietary fiber and probiotics in infectious diseases. *Microbial Pathogenesis*. 2019:103931.
- [16] Lilly DM, Stillwell RH. Probiotics: Growth-promoting factors produced by microorganisms. *Science*. 1965;147(3659):747-8. DOI: 10.1126/science.147.3659.747.
- [17] Hosono A. Fermented milk in the orient. In: Nakazawa Y, Hosono A. *Functions of Fermented Milk: Challengers for the Health Sciences*. Barking (UK): Elsevier Science Publishers Ltd; 1992. p. 61-78.

- [18] Bezirtzoglou E, Stavropoulou E. Immunology and probiotic impact of the newborn and young children intestinal microflora. *Anaerobe*. 2011;17(6):369-74. DOI: 10.1016/j.anaerobe.2011.03.010.
- [19] Bermudez-Brito M, Plaza-Díaz J, Muñoz-Quezada S, Gómez-Llorente C, Gil A. Probiotic mechanisms of action. *Ann Nutr Metab*. 2012;61(2):160-74. DOI: 10.1159/000342079.
- [20] Reid G, Jass J, Sebulsky MT, McCormick JK. Potential uses of probiotics in clinical practice. *Clin Microbiol Rev*. 2003;16(4):658-72. DOI: 10.1128/cmr.16.4.658-672.2003.
- [21] Melo Pereira GV, de Oliveira Coelho B, Magalhães Júnior AI, Thomaz-Soccol V, Soccol CR. How to select a probiotic? A review and update of methods and criteria. *Biotechnol Adv*. 2018;36:2060-2076.
- [22] Stavropoulou E, Bezirtzoglou E. Probiotics in Medicine: A Long Debate. *Frontiers in immunology*. 2020;11:2192. DOI: 10.3389/fimmu.2020.02192
- [23] Doron S, Snyderman DR. Risk and safety of probiotics. *Clin Infect Dis*. 2015;60:S129–S134.
- [24] Prado FC, Lindner JD, Inaba J, Thomaz-Soccol V, Brar SK, Soccol CR. Development and evaluation of a fermented coconut water beverage with potential health benefits. *J Funct Foods*. 2015;12:489-497.
- [25] Soccol CR, Prado MRM, Garcia LMB, Rodrigues C, Medeiros ABP, Thomaz-Soccol V. Current developments in probiotics. *J Microb Biochem Technol*. 2015;7:11-20.
- [26] Karami S, Roayaei M, Hamzavi H, Bahmani M, Hassanzad-Azar H, Leila M, Rafieian-Kopaei M. Isolation and identification of probiotic *Lactobacillus* from local dairy and evaluating their antagonistic effect on pathogens. *International journal of pharmaceutical investigation*. 2017;7(3):137-141. DOI: 10.4103/jphi.JPHI_8_17
- [27] Bazireh H, Shariati P, Azimzadeh Jamalkandi S, Ahmadi A, Boroumand MA. Isolation of Novel Probiotic *Lactobacillus* and *Enterococcus* Strains From Human Salivary and Fecal Sources. *Front Microbiol*. 2020;11:597946. DOI: 10.3389/fmicb.2020.597946.
- [28] Jungersen M, Wind A, Johansen E, Christensen JE, Stuer-Lauridsen B, Eskesen D. The Science behind the Probiotic Strain *Bifidobacterium animalis* subsp. *lactis* BB-12(®). *Microorganisms*. 2014;2(2):92-110. doi: 10.3390/microorganisms2020092
- [29] Reuter G. The *Lactobacillus* and *Bifidobacterium* microflora of the human intestine: composition and succession. *Curr Issues Intest Microbiol*. 2001;2(2):43-53.
- [30] Milani C, Mangifesta M, Mancabelli L, Lugli GA, James K, Duranti S, Turrone F, Ferrario C, Ossiprandi MC, van Sinderen D, Ventura M. Unveiling bifidobacterial biogeography across the mammalian branch of the tree of life. *ISME J*. 2017;11(12):2834-2847. DOI: 10.1038/ismej.2017.138
- [31] Yamamoto E, Watanabe R, Koizumi A, Ishida T, Kimura K. Isolation and characterization of *Streptococcus thermophilus* possessing *prtS* gene from raw milk in Japan. *Biosci Microbiota Food Health*. 2020;39(3):169-174. DOI: 10.12938/bmfh.2019-052
- [32] Xiong ZQ, Kong LH, Lai PF, Xia YJ, Liu JC, Li QY, Ai LZ. Genomic and phenotypic analyses of exopolysaccharide biosynthesis in

- Streptococcus thermophilus S-3. *J Dairy Sci.* 2019;102(6):4925-4934. DOI: 10.3168/jds.2018-15572
- [33] Fisher K, Phillips C. The ecology, epidemiology and virulence of *Enterococcus*. *Microbiology (Reading)*. 2009;155(Pt 6):1749-1757. DOI: 10.1099/mic.0.026385-0
- [34] Reuben RC, Roy PC, Sarkar SL, Alam RU, Jahid IK. Isolation, characterization, and assessment of lactic acid bacteria toward their selection as poultry probiotics. *BMC Microbiol.* 2019;19(1):253. DOI: 10.1186/s12866-019-1626-0
- [35] Madoshi BP, Mtambo MMA, Muhairwa AP, Lupindu AM, Olsen JE. Isolation of vancomycin-resistant *Enterococcus* from apparently healthy human animal attendants, cattle and cattle wastes in Tanzania. *J Appl Microbiol.* 2018;124(5):1303-1310. DOI: 10.1111/jam.13722
- [36] Bhagat D, Raina N, Kumar A, Katoch M, Khajuria Y, Slathia PS, Sharma P. Probiotic properties of a phytase producing *Pediococcus acidilactici* strain SMVDUDB2 isolated from traditional fermented cheese product, Kalarei. *Sci Rep.* 2020;10(1):1926. DOI: 10.1038/s41598-020-58676-2
- [37] Leroy F, De Vuyst L. Simulation of the effect of sausage ingredients and technology on the functionality of the bacteriocin-producing *Lactobacillus sakei* CTC 494 strain. *Int J Food Microbiol.* 2005;100(1-3):141-52. DOI: 10.1016/j.ijfoodmicro.2004.10.011
- [38] Sajedinejad N, Paknejad M, Houshmand B, Sharafi H, Jelodar R, Shahbani Zahir H, Noghabi KA. *Lactobacillus salivarius* NK02: a Potent Probiotic for Clinical Application in Mouthwash. *Probiotics Antimicrob Proteins.* 2018;10(3):485-495. DOI: 10.1007/s12602-017-9296-4
- [39] Lopes EG, Moreira DA, Gullón P, Gullón B, Cardelle-Cobas A, Tavaría FK. Topical application of probiotics in skin: adhesion, antimicrobial and antibiofilm in vitro assays. *J Appl Microbiol.* 2017;122(2):450-461. DOI: 10.1111/jam.13349
- [40] Rossoni RD, Fuchs BB, de Barros PP, Velloso MD, Jorge AO, Junqueira JC, Mylonakis E. *Lactobacillus paracasei* modulates the immune system of *Galleria mellonella* and protects against *Candida albicans* infection. *PLoS One.* 2017;12(3):e0173332. DOI: 10.1371/journal.pone.0173332
- [41] Markowiak P, Ślizewska K. Effects of Probiotics, Prebiotics, and Synbiotics on Human Health. *Nutrients.* 2017;9(9):1021. DOI: 10.3390/nu9091021
- [42] de Moraes GMD, de Abreu LR, do Egito AS, Salles HO, da Silva LMF, Nero LA, Todorov SD, Dos Santos KMO. Functional Properties of *Lactobacillus mucosae* Strains Isolated from Brazilian Goat Milk. *Probiotics Antimicrob Proteins.* 2017;9(3):235-245. DOI: 10.1007/s12602-016-9244-8
- [43] Goderska K, Agudo Pena S, Alarcon T. *Helicobacter pylori* treatment: antibiotics or probiotics. *Appl Microbiol Biotechnol.* 2018;102(1):1-7. DOI: 10.1007/s00253-017-8535-7
- [44] Neal-McKinney JM, Lu X, Duong T, Larson CL, Call DR, Shah DH, Konkel ME. Production of organic acids by probiotic lactobacilli can be used to reduce pathogen load in poultry. *PLoS One.* 2012;7(9):e43928. DOI: 10.1371/journal.pone.0043928
- [45] Sikorska H, Smoragiewicz W. Role of probiotics in the prevention and treatment of meticillin-resistant *Staphylococcus aureus* infections. *Int J Antimicrob Agents.* 2013;42(6):475-81. DOI: 10.1016/j.ijantimicag.2013.08.003

- [46] Dong H, Rowland I, Thomas LV, Yaqoob P. Immunomodulatory effects of a probiotic drink containing *Lactobacillus casei* Shirota in healthy older volunteers. *Eur J Nutr*. 2013;52(8):1853-1863.
- [47] Ganguli K, Meng D, Rautava S, Lu L, Walker WA, Nanthakumar N. Probiotics prevent necrotizing enterocolitis by modulating enterocyte genes that regulate innate immune-mediated inflammation. *Am J Physiol Gastrointest Liver Physiol*. 2013;304(2):G132-41. DOI: 10.1152/ajpgi.00142.2012
- [48] Matsubara VH, Wang Y, Bandara HMHN, Mayer MPA, Samaranayake LP. Probiotic lactobacilli inhibit early stages of *Candida albicans* biofilm development by reducing their growth, cell adhesion, and filamentation. *Appl Microbiol Biotechnol*. 2016;100(14):6415-6426. DOI: 10.1007/s00253-016-7527-3
- [49] Yang HL, Xia HQ, Ye YD, Zou WC, Sun YZ. Probiotic *Bacillus pumilus* SE5 shapes the intestinal microbiota and mucosal immunity in grouper *Epinephelus coioides*. *Dis Aquat Organ*. 2014;111(2):119-27. DOI: 10.3354/dao02772
- [50] Yu AQ, Li L. The Potential Role of Probiotics in Cancer Prevention and Treatment. *Nutr Cancer*. 2016;68(4):535-44. DOI: 10.1080/01635581.2016.1158300
- [51] Wollowski I, Rechkemmer G, Pool-Zobel BL. Protective role of probiotics and prebiotics in colon cancer. *Am J Clin Nutr*. 2001;73(2):451S-455S. DOI: 10.1093/ajcn/73.2.451s
- [52] Devaraj NK, Suppiah S, Veettil SK, Ching SM, Lee KW, Menon RK, Soo MJ, Deuraseh I, Hoo FK, Sivaratnam D. The Effects of Probiotic Supplementation on the Incidence of Diarrhea in Cancer Patients Receiving Radiation Therapy: A Systematic Review with Meta-Analysis and Trial Sequential Analysis of Randomized Controlled Trials. *Nutrients*. 2019;11(12):2886. DOI: 10.3390/nu11122886
- [53] Clancy R. Immunobiotics and the probiotic evolution. *FEMS Immunol Med Microbiol*. 2003;38(1):9-12. DOI: 10.1016/S0928-8244(03)00147-0
- [54] Liu MM, Li ST, Shu Y, Zhan HQ. Probiotics for prevention of radiation-induced diarrhea: A meta-analysis of randomized controlled trials. *PLoS One*. 2017;12(6):e0178870. DOI: 10.1371/journal.pone.0178870
- [55] Casas-Solís J, Huizar-López M, Irecta-Nájera C, Pita-López M, Santerre A. Immunomodulatory Effect of *Lactobacillus casei* in a Murine Model of Colon Carcinogenesis. *Probiotics and Antimicrobial Proteins*. 2020;12. DOI: 10.1007/s12602-019-09611-z
- [56] Rather IA, Bajpai VK, Kumar S, Lim J, Paek WK, Park YH. Probiotics and Atopic Dermatitis: An Overview. *Front Microbiol*. 2016;7:507. DOI: 10.3389/fmicb.2016.00507
- [57] Huang R, Ning H, Shen M, Li J, Zhang J, Chen X. Probiotics for the Treatment of Atopic Dermatitis in Children: A Systematic Review and Meta-Analysis of Randomized Controlled Trials. *Front Cell Infect Microbiol*. 2017;7:392. DOI: 10.3389/fcimb.2017.00392
- [58] Lise M, Mayer I, Silveira M. Use of probiotics in atopic dermatitis. *Rev Assoc Med Bras (1992)*. 2018;64(11):997-1001. DOI: 10.1590/1806-9282.64.11.997
- [59] Shimizu M, Hashiguchi M, Shiga T, Tamura HO, Mochizuki M. Meta-Analysis: Effects of Probiotic Supplementation on Lipid Profiles in Normal to Mildly Hypercholesterolemic

Individuals. PLoS One.

2015;10(10):e0139795. DOI: 10.1371/journal.pone.0139795

[60] Nath A, Molnár MA, Csighy A, Kószegi K, Galambos I, Huszár KP, Koris A, Vatai G. Biological Activities of Lactose-Based Prebiotics and Symbiosis with Probiotics on Controlling Osteoporosis, Blood-Lipid and Glucose Levels. *Medicina (Kaunas)*. 2018;54(6):98. DOI: 10.3390/medicina54060098

[61] Tahmourespour A, Kermanshahi RK. The effect of a probiotic strain (*Lactobacillus acidophilus*) on the plaque formation of oral Streptococci. *Bosn J Basic Med Sci*. 2011;11(1):37-40. DOI: 10.17305/bjbms.2011.2621

[62] Ishikawa KH, Mayer MP, Miyazima TY, Matsubara VH, Silva EG, Paula CR, Campos TT, Nakamae AE. A multispecies probiotic reduces oral *Candida* colonization in denture wearers. *J Prosthodont*. 2015;24(3):194-9. DOI: 10.1111/jopr.12198.

[63] Kuru BE, Laleman I, Yalınizoğlu T, Kuru L, Teughels W. The Influence of a *Bifidobacterium animalis* Probiotic on Gingival Health: A Randomized Controlled Clinical Trial. *J Periodontol*. 2017;88(11):1115-1123. DOI: 10.1902/jop.2017.170213.

[64] Wieërs G, Belkhir L, Enaud R, Leclercq S, Philippart de Foy JM, Dequenne I, de Timary P, Cani PD. How Probiotics Affect the Microbiota. *Front Cell Infect Microbiol*. 2020;9:454. DOI: 10.3389/fcimb.2019.00454

[65] Caballero S, Carter R, Ke X, Sušac B, Leiner IM, Kim GJ, Miller L, Ling L, Manova K, Pamer EG. Distinct but Spatially Overlapping Intestinal Niches for Vancomycin-Resistant *Enterococcus faecium* and Carbapenem-Resistant *Klebsiella pneumoniae*. *PLoS Pathog*. 2015;11(9):e1005132. doi: 10.1371/journal.ppat.1005132.

[66] Kaushik SB, Lebwohl MG. Psoriasis: Which therapy for which patient: Focus on special populations and chronic infections. *J Am Acad Dermatol*. 2019;80(1):43-53. DOI: 10.1016/j.jaad.2018.06.056

[67] Singh V, Kumar A, Raheja G, Anbazhagan AN, Priyamvada S, Saksena S, Jhandier MN, Gill RK, Alrefai WA, Borthakur A, Dudeja PK. *Lactobacillus acidophilus* attenuates downregulation of DRA function and expression in inflammatory models. *Am J Physiol Gastrointest Liver Physiol*. 2014;307(6):G623-31. DOI: 10.1152/ajpgi.00104.2014

[68] Crum-Cianflone NF, Sullivan E, Ballon-Landa G. Fecal microbiota transplantation and successful resolution of multidrug-resistant-organism colonization. *J Clin Microbiol*. 2015;53(6):1986-9. DOI: 10.1128/JCM.00820-15

[69] Millan B, Park H, Hotte N, Mathieu O, Burguiere P, Tompkins TA, Kao D, Madsen KL. Fecal Microbial Transplants Reduce Antibiotic-resistant Genes in Patients With Recurrent *Clostridium difficile* Infection. *Clin Infect Dis*. 2016;62(12):1479-1486. DOI: 10.1093/cid/ciw185

[70] Singhi SC, Kumar S. Probiotics in critically ill children. *F1000Res*. 2016;5:F1000 Faculty Rev-407. doi: 10.12688/f1000research.7630.1

[71] Soltan Dallal MM, Zamaniahari S, Davoodabadi A, Hosseini M, Rajabi Z. Identification and characterization of probiotic lactic acid bacteria isolated from traditional persian pickled vegetables. *GMS Hyg Infect Control*. 2017;12:Doc15. DOI: 10.3205/dgkh000300

[72] Tannock GW, Munro K, Harmsen HJ, Welling GW, Smart J, Gopal PK. Analysis of the fecal microflora of human subjects

consuming a probiotic product containing *Lactobacillus rhamnosus* DR20. *Appl Environ Microbiol.* 2000;66(6):2578-88. DOI: 10.1128/aem.66.6.2578-2588.2000

[73] Crouzet L, Derrien M, Cherbuy C, Plancade S, Foulon M, Chalin B, van Hylckama Vlieg JET, Grompone G, Rigottier-Gois L, Serron P. *Lactobacillus paracasei* CNCM I-3689 reduces vancomycin-resistant *Enterococcus* persistence and promotes *Bacteroidetes* resilience in the gut following antibiotic challenge. *Sci Rep.* 2018;8(1):5098. DOI: 10.1038/s41598-018-23437-9

[74] Singh MN, Hemant KS, Ram M, Shivakumar HG. Microencapsulation: A promising technique for controlled drug delivery. *Res Pharm Sci.* 2010;5(2):65-77.

[75] Sarao LK, Arora M. Probiotics, prebiotics, and microencapsulation: A review. *Crit Rev Food Sci Nutr.* 2017;57(2):344-371. DOI: 10.1080/10408398.2014.887055

[76] Champagne CP, Fustier P. Microencapsulation for the improved delivery of bioactive compounds into foods. *Curr Opin Biotechnol.* 2007;18(2):184-90.

[77] Chen MJ, Chen KN. Applications of Probiotic Encapsulation. In: *Dairy Products. Encapsulation Control Release Technol Food Syst.* 2007. p. 83-112. DOI: 10.1002/9780470277881.ch4

[78] Asgari S, Pourjavadi A, Licht TR, Boisen A, Ajallouei F. Polymeric carriers for enhanced delivery of probiotics. *Adv Drug Deliv Rev.* 2020;161-162:1-21. DOI: 10.1016/j.addr.2020.07.014

[79] Burgain J, Gaiani C, Linder M, Scher J. Encapsulation of probiotic living cells: From laboratory scale to industrial applications. *J Food Eng. Elsevier Ltd.* 2011;104:467-83. DOI: 10.1016/j.jfoodeng.2010.12.031

[80] Sundus KSY, Sayantani N, Moses DJA. Targeted Delivery of Probiotics : Perspectives on Research and Commercialization. *Probiotics Antimicrob. Proteins.* Springer US. 2021. DOI: 10.1007/s12602-021-09791-7

[81] Rodrigues FJ, Cedran MF, Bicas JL, Sato HH. Encapsulated probiotic cells: Relevant techniques, natural sources as encapsulating materials and food applications – A narrative review. *Food Res Int.* 2020;137:109682. DOI: 10.1016/j.foodres.2020.109682

[82] Rossier-Miranda FJ, Schroën K, Boom R. Mechanical characterization and pH response of fibril-reinforced microcapsules prepared by layer-by-layer adsorption. *Langmuir.* 2010;26(24):19106-13. DOI: 10.1021/la1033542

[83] Serna L, Vallejo-Castillo, V. Probiotic encapsulation. *African journal of microbiology research.* 2013;7:4743. DOI: 10.5897/AJMR2013.5718.

[84] Yoha KS, Nida S, Dutta S, Moses JA, Anandharamakrishnan C. Targeted Delivery of Probiotics: Perspectives on Research and Commercialization. *Probiotics Antimicrob Proteins.* 2021;27:1-34. DOI: 10.1007/s12602-021-09791-7

[85] Gouin S. Microencapsulation: Industrial appraisal of existing technologies and trends. *Trends Food Sci Technol.* 2004;15:330-47.

[86] Mortazavian AM, Azizi A, Ehsani MR, Razavi SH, Mousavi SM, Sohrabvandi S, Reinheimer JA. Survival of encapsulated probiotic bacteria in Iranian yogurt drink (Doogh) after the product exposure to simulated gastrointestinal conditions. *Milchwissenschaft.* 2008;63(4):427-429.

[87] Krasaekoopt W, Bhandari B, Deeth H. Evaluation of encapsulation techniques of probiotics for yoghurt. *Int Dairy J.* 2003;13:3-13.

- [88] Rayment P, Wright P, Hoad C, Ciampi E, Haydock D, Gowland P, et al. Investigation of alginate beads for gastro-intestinal functionality, Part 1: In vitro characterisation. *Food Hydrocoll.* Elsevier Ltd. 2009;23:816-22. DOI: 10.1016/j.foodhyd.2008.04.011
- [89] Li KL, Wang BZ, Li ZP, Li YL, Liang JJ. Alterations of intestinal flora and the effects of probiotics in children with recurrent respiratory tract infection. *World J Pediatr.* 2019;15(3):255-261. DOI: 10.1007/s12519-019-00248-0.
- [90] Lee JS, Cha DS, Park HJ. Survival of freeze-dried *Lactobacillus bulgaricus* KFRI 673 in chitosan-coated calcium alginate microparticles. *J Agric Food Chem.* 2004;52:7300-5.
- [91] Groboillot AF, Champagne CP, Darling GD, Poncelet D, Neufeld RJ. Membrane formation by interfacial cross-linking of chitosan for microencapsulation of *Lactococcus lactis*. *Biotechnol Bioeng.* 1993;42(10):1157-63. DOI: 10.1002/bit.260421005
- [92] Huguet ML, Neufeld RJ, Dellacherie E. Calcium-alginate beads coated with polycationic polymers: Comparison of chitosan and DEAE-dextran. *Process Biochem.* 1996;31:347-53.
- [93] Yuguchi Y, Thu Thuy TT, Urakawa H, Kajiwara K. Structural characteristics of carrageenan gels: Temperature and concentration dependence. *Food Hydrocoll.* 2002;16:515-22.
- [94] Dinakar P, Mistry VV. Growth and viability of *Bifidobacterium bifidum* in cheddar cheese. *J Dairy Sci.* 1994; 77(10):2854-64. DOI: 10.3168/jds.S0022-030(94)77225-8
- [95] Dafe A, Etemadi H, Zarredar H, Mahdavinia GR. Development of novel carboxymethyl cellulose/k-carrageenan blends as an enteric delivery vehicle for probiotic bacteria. *Int J Biol Macromol.* Elsevier B.V. 2017;97:299-307. DOI: 10.1016/j.ijbiomac.2017.01.016
- [96] Gutiérrez-Zamorano C, González-Ávila M, Díaz-Blas G, Smith CT, González-Correa C, García-Cancino A. Increased anti-*Helicobacter pylori* effect of the probiotic *Lactobacillus fermentum* UCO-979C strain encapsulated in carrageenan evaluated in gastric simulations under fasting conditions. *Food Res Int.* Elsevier. 2019;121:812-6. DOI: 10.1016/j.foodres.2018.12.064
- [97] Bajaj IB, Survase SA, Saudagar PS, Singhal RS. Gellan gum: Fermentative production, downstream processing and applications. *Food Technol Biotechnol.* 2007;45:341-54.
- [98] Jiménez-Pranteda ML, Poncelet D, Nader-Macías ME, Arcos A, Aguilera M, Monteoliva-Sánchez M, et al. Stability of lactobacilli encapsulated in various microbial polymers. *J Biosci Bioeng* [Internet]. The Society for Biotechnology, Japan. 2012;113:179-84. DOI: 10.1016/j.jbiosc.2011.10.010
- [99] Fávoro-Trindade CS, Grosso CRF. Microencapsulation of *L. acidophilus* (La-05) and *B. Lactis* (Bb-12) and evaluation of their survival at the pH values of the stomach and in bile. *Journal of Microencapsulation.* 2002;19(4):485-494.
- [100] Crittenden R, Laitila A, Forsell P, Matto J, Saarela M, Mattila-Sandholm T, Myllarinen P. Adhesion of bifidobacteria to granular starch and its implications in probiotic technologies. *Applied and Environmental Microbiology.* 2001;67(8):3469-3475.
- [101] Martin MJ, Lara-Villoslada F, Ruiz MA, Morales ME. Effect of unmodified starch on viability of alginate-encapsulated *Lactobacillus*

fermentum CECT5716. LWT - Food Sci Technol. Elsevier Ltd. 2013;53:480-6. DOI: 10.1016/j.lwt.2013.03.019

[102] López-Rubio A, Sanchez E, Sanz Y, Lagaron JM. Encapsulation of living bifidobacteria in ultrathin PVOH electrospun fibers. Biomacromolecules. 2009;10:2823-9.

[103] Çanga EM, Dudak FC. Improved digestive stability of probiotics encapsulated within poly(vinyl alcohol)/cellulose acetate hybrid fibers. Carbohydr Polym. Elsevier Ltd. 2021;264.

[104] Klarin B, Adolfsson A, Torstensson A, Larsson A. Can probiotics be an alternative to chlorhexidine for oral care in the mechanically ventilated patient? A multicentre, prospective, randomised controlled open trial. Crit Care. 2018;22(1):272. DOI: 10.1186/s13054-018-2209-4.

[105] Rostok M, Hütt P, Rööp T, Smidt I, Štšepetova J, Salumets A, Mändar R. Potential vaginal probiotics: safety, tolerability and preliminary effectiveness. Benef Microbes. 2019;10(4):385-393. DOI: 10.3920/BM2016.0123.

[106] De Gregorio PR, Maldonado NC, Pingitore EV, Terraf MCL, Tomás MSJ, de Ruiz CS, Santos V, Wiese B, Bru E, Paiz MC, Reina MF, Schujman DE, Nader-Macías MEF. Intravaginal administration of gelatine capsules containing freeze-dried autochthonous lactobacilli: a double-blind, randomised clinical trial of safety. Benef Microbes. 2020;11(1):5-17. DOI: 10.3920/BM2019.0081.

[107] Hooi JKY, Lai WY, Ng WK, Suen MMY, Underwood FE, Tanyingoh D, Malfertheiner P, Graham DY, Wong VWS, Wu JCY, Chan FKL, Sung JY, Kaplan GG, Ng SC. Global Prevalence of Helicobacter pylori

Infection: Systematic Review and Meta-Analysis. Gastroenterology. 2017;153(2):420-429. DOI: 10.1053/j.gastro.2017.04.022

[108] Oh B, Kim BS, Kim JW, Kim JS, Koh SJ, Kim BG, Lee KL, Chun J. The Effect of Probiotics on Gut Microbiota during the Helicobacter pylori Eradication: Randomized Controlled Trial. Helicobacter. 2016;21(3):165-74. DOI: 10.1111/hel.12270.

[109] Poonyam P, Chotivitayatarakorn P, Vilaichone RK. High Effective of 14-Day High-Dose PPI-Bismuth-Containing Quadruple Therapy with Probiotics Supplement for Helicobacter Pylori Eradication: A Double Blinded-Randomized Placebo-Controlled Study. Asian Pac J Cancer Prev. 2019;20(9):2859-2864. DOI: 10.31557/APJCP.2019.20.9.2859

[110] Mahachai V, Vilaichone RK, Pittayanon R, Rojborwonwitaya J, Leelakusolvong S, Kositchaiwat C, Mairiang P, Praisontarangkul OA, Ovarlarnporn B, Sottisuporn J, Pisespongsa P, Maneerattanaporn M, Sony R, Sirinthornpunya S, Chaiyamahapurk O, Wiwattanachang O, Sansak I, Harnsomboon P, Chitapanarux T, Chuenrattanukul S. Thailand Consensus on Helicobacter pylori Treatment 2015. Asian Pac J Cancer Prev. 2016;17(5):2351-2360. DOI: 10.7314/APJCP.2016.17.5.2351

[111] Sung JY, Coker OO, Chu E, Szeto CH, Luk STY, Lau HCH, Yu J. Gastric microbes associated with gastric inflammation, atrophy and intestinal metaplasia 1 year after Helicobacter pylori eradication. Gut. 2020;69:1572-1581. DOI: 10.1136/gutjnl-2019-319826

[112] Yoon JY, Cha JM, Hong SS, Kim HK, Kwak MS, Jeon JW, Shin HP. Fermented milk containing Lactobacillus paracasei and *Glycyrrhiza glabra* has a beneficial effect in patients with Helicobacter pylori infection. A

- randomized, double-blind, placebo-controlled study. *Medicine*. 2019;98:35. DOI: 10.1097/MD.00000000000016601
- [113] Çekin AH, Şahintürk Y, Harmandar FA, Uyar S, Yolcular BO, Çekin Y. Use of probiotics as an adjuvant to sequential *H. pylori* eradication therapy: impact on eradication rates, treatment resistance, treatment-related side effects, and patient compliance. *Turk J Gastroenterol*. 2017;28(1):3-11. DOI: 10.5152/tjg.2016.0278.
- [114] Chen L, Xu W, Lee A, He J, Huang B, Zheng W, Su T, Lai S, Long Y, Chu H, Chen Y, Wang L, Wang K, Si J, Chen S. The impact of *Helicobacter pylori* infection, eradication therapy and probiotic supplementation on gut microenvironment homeostasis: An open-label, randomized clinical trial. *EBioMedicine*. 2018;35: 87-96. DOI: 10.1016/j.ebiom.2018.08.028
- [115] Underwood MA. Probiotics and the prevention of necrotizing enterocolitis. *Journal of Pediatric Surgery*. 2019;54(3):405-412. DOI: 10.1016/j.jpedsurg.2018.08.055
- [116] Seddik H, Boutallaka H, Elkoti I, Nejjari F, Berraida R, Berrag S, Loubaris K, Sentissi S, Benkirane A. *Saccharomyces boulardii* CNCM I-745 plus sequential therapy for *Helicobacter pylori* infections: a randomized, open-label trial. *Eur J Clin Pharmacol*. 2019;75(5):639-645. DOI: 10.1007/s00228-019-02625-0
- [117] Barker AK, Duster M, Valentine S, Hess T, Archbald-Pannone L, Guerrant R, Safdar N. A randomized controlled trial of probiotics for *Clostridium difficile* infection in adults (PICO). *J Antimicrob Chemother*. 2017;72:3177-3180. DOI: 10.1093/jac/dkx254
- [118] Alberda C, Marcushamer S, Hewan T, Journault N, Kutsogiannis D. Feasibility of a *Lactobacillus casei* Drink in the Intensive Care Unit for Prevention of Antibiotic Associated Diarrhea and *Clostridium difficile*. *Nutrients*. 2018;10:539. DOI: 10.3390/nu10050539
- [119] van Wietmarschen HA, Busch M, van Oostveen A, Pot G, Jong MC. Probiotics use for antibiotic-associated diarrhea: a pragmatic participatory evaluation in nursing homes. *BMC Gastroenterology*. 2020;20:151. DOI: 10.1186/s12876-020-01297-w
- [120] Kołodziej M, Szajewska H. *Lactobacillus reuteri* DSM 17938 in the prevention of antibiotic-associated diarrhoea in children: a randomized clinical trial. *Clinical Microbiology and Infection*. 2019;25:699-704. DOI: 10.1016/j.cmi.2018.08.017 1198-743
- [121] Szymański H, Szajewska H. Lack of Efficacy of *Lactobacillus reuteri* DSM 17938 for the Treatment of Acute Gastroenteritis: A Randomized Controlled Trial. *Pediatr Infect Dis J*. 2019;38(10):e237-e242. DOI: 10.1097/INF.0000000000002355
- [122] Ljungquist O, Kampmann C, Resman F, Riesbeck K, Tham J. Probiotics for intestinal decolonization of ESBL-producing Enterobacteriaceae: a randomized, placebo-controlled clinical trial. *Clinical Microbiology and Infection*. 2020;26:456-462. DOI: 10.1016/j.cmi.2019.08.019 1198-743X
- [123] Nova E, Wörnberg J, Gómez-Martínez S, Díaz LE, Romeo J, Marcos A. Immunomodulatory effects of probiotics in different stages of life. *Br J Nutr*. 2007;98(1):90-95.
- [124] Kawase M, He F, Kubota A, Yoda K, Miyazawa K, Hiramatsu M. Heatkilled *Lactobacillus gasseri* TMC0356 protects mice against influenza virus infection by stimulating gut and respiratory immune responses. *FEMS Immunol Med Microbiol*. 2012;64(2):280-288.
- [125] Pu F, Guo Y, Li M, Zhu H, Wang S, Shen X, He M, Huang C, He F. Yogurt

supplemented with probiotics can protect the healthy elderly from respiratory infections: A randomized controlled open-label trial. *Clin Interv Aging*. 2017;12:1223-1231. DOI: 10.2147/CIA.S141518.

[126] Michalickova D, Minic R, Dikic N, Andjelkovic M, Kostic-Vucicevic M, Stojmenovic T, Nikolic I, Djordjevic B. Lactobacillus helveticus Lafti L10 supplementation reduces respiratory infection duration in a cohort of elite athletes: a randomized, double-blind, placebo-controlled trial. *Appl Physiol Nutr Metab*. 2016;41(7):782-9. DOI: 10.1139/apnm-2015-0541.

[127] Peiris JS. Severe acute respiratory syndrome (SARS). *J Clin Virol*. 2003;28:245-7.

[128] Peiris JS, Tu WW, Yen HL. A novel H1N1 virus causes the first pandemic of the 21st century. *Eur J Immunol*. 2009;39:2946-54.

[129] Marsland BJ, Gollwitzer ES. Host-microorganism interactions in lung diseases. *Nat Rev Immunol*. 2014;14:827-835. DOI: 10.1038/nri3769

[130] Wang B, Hylwka T, Smieja M, Surette M, Bowdish DME, Loeb M. Probiotics to Prevent Respiratory Infections in Nursing Homes: A Pilot Randomized Controlled Trial. *J Am Geriatr Soc*. 2018;66(7):1346-1352. DOI: 10.1111/jgs.15396

[131] Campanella V, Syed J, Santacroce L, Saini R, Ballini A, Inchingolo F. Oral probiotics influence oral and respiratory tract infections in pediatric population: a randomized double-blinded placebo-controlled pilot study. *Eur Rev Med Pharmacol Sci*. 2018;22(22):8034-8041. DOI: 10.26355/eurrev_201811_16433.

[132] Sadeghi-Bojd S, Naghshizadian R, Mazaheri M, Ghane Sharbaf F, Assadi F. Efficacy of Probiotic Prophylaxis After The First Febrile Urinary Tract Infection

in Children With Normal Urinary Tracts. *J Pediatric Infect Dis Soc*. 2020;9(3):305-310. DOI: 10.1093/jpids/piz025.

[133] Nelson CP, Hoberman A, Shaikh N, et al. Antibiotic resistance and urinary tract infection recurrence. *Pediatrics*. 2016;137:pii:e20152490. DOI: 10.1542/peds.2015-2490.

[134] Libertucci J, Young VB. The role of the microbiota in infectious diseases. *Nat Microbiol*. 2019;4:35-45.

[135] Matsuzaki T, Chin J. Modulating immune responses with probiotic bacteria. *Immunol Cell Biol*. 2000;78:67-73.

[136] Cadieux PA, Burton J, Devillard E, Reid G. Lactobacillus by-products inhibit the growth and virulence of uropathogenic *Escherichia coli*. *J Physiol Pharmacol*. 2009;60(6):13-8.

[137] Kovachev SM, Vatcheva-Dobrevska RS. Local Probiotic Therapy for Vaginal *Candida albicans* Infections. *Probiotics Antimicrob Proteins*. 2015;7(1):38-44. DOI: 10.1007/s12602-014-9176-0.

[138] Pendharkar S, Brandsborg E, Hammarström L, Marcotte H, Larsson PG. Vaginal colonisation by probiotic lactobacilli and clinical outcome in women conventionally treated for bacterial vaginosis and yeast infection. *BMC Infect Dis*. 2015;15:255. DOI: 10.1186/s12879-015-0971-3.

[139] Barthow C, Wickens K, Stanley T, Mitchell EA, Maude R, Abels P, Purdie G, Murphy R, Stone P, Kang J, Hood F, Rowden J, Barnes P, Fitzharris P, Craig J, Slykerman RF, Crane J. The Probiotics in Pregnancy Study (PiP Study): rationale and design of a double-blind randomised controlled trial to improve maternal health during pregnancy and prevent infant eczema and allergy. *BMC Pregnancy Childbirth*.

2016;16(1):133. DOI: 10.1186/s12884-016-0923-y.

[140] Davar R, Nokhostin F, Eftekhar M, Sekhavat L, Bashiri Zadeh M, Shamsi F. Comparing the Recurrence of Vulvovaginal Candidiasis in Patients Undergoing Prophylactic Treatment with Probiotic and Placebo During the 6 Months. *Probiotics Antimicrob Proteins*. 2016;8(3):130-3. DOI: 10.1007/s12602-016-9218-x.

[141] Russo R, Superti F, Karadja E, De Seta F. Randomised clinical trial in women with Recurrent Vulvovaginal Candidiasis: Efficacy of probiotics and lactoferrin as maintenance treatment. *Mycoses*. 2019;62(4):328-335. DOI: 10.1111/myc.12883.

[142] Vladareanu R, Mihu D, Mitran M, Mehedintu C, Boianu A, Manolache M, Vladareanu S. New evidence on oral *L. plantarum* P17630 product in women with history of recurrent vulvovaginal candidiasis (RVVC): a randomized double-blind placebo-controlled study. *Eur Rev Med Pharmacol Sci*. 2018;22(1):262-267. DOI: 10.26355/eurrev_201801_14128.

[143] Reznichenko H, Henyk N, Maliuk V, Khyzhnyak T, Tynna Y, Filipiuk I, Veresniuk N, Zubrytska L, Quintens J, Richir K, Gerasymov S. Oral Intake of Lactobacilli Can Be Helpful in Symptomatic Bacterial Vaginosis: A Randomized Clinical Study. *J Low Genit Tract Dis*. 2020;24(3):284-289. DOI: 10.1097/LGT.0000000000000518.

[144] Sgibnev A, Kremleva E. Probiotics in addition to metronidazole for treatment *Trichomonas vaginalis* in the presence of BV: a randomized, placebo-controlled, double-blind study. *Eur J Clin Microbiol Infect Dis*. 2020;39(2):345-351. DOI: 10.1007/s10096-019-03731-8.

[145] Toh SL, Lee BB, Ryan S, Simpson JM, Clezy K, Bossa L, Rice SA,

Marial O, Weber GH, Kaur J, Boswell-Ruys CL, Goodall S, Middleton JW, Tuderhope M, Kotsiou G. Probiotics [LGG-BB12 or RC14-GR1] versus placebo as prophylaxis for urinary tract infection in persons with spinal cord injury [ProSCIUTTU]: a randomised controlled trial. *Spinal Cord*. 2019;57(7):550-561. DOI: 10.1038/s41393-019-0251-y.

[146] de Wolff MG, Johansen M, Ersbøll AS, Rosthøj S, Brunsgaard A, Midtgaard J, Tabor A, Hegaard HK. Efficacy of a midwife-coordinated, individualized, and specialized maternity care intervention (ChroPreg) in addition to standard care in pregnant women with chronic disease: protocol for a parallel randomized controlled trial. *Trials*. 2019;20(1):291. DOI: 10.1186/s13063-019-3405-5

[147] Gallo MF, Macaluso M, Warner L, Fleenor ME, Hook EW 3rd, Brill I, Weaver MA. Bacterial vaginosis, gonorrhea, and chlamydial infection among women attending a sexually transmitted disease clinic: a longitudinal analysis of possible causal links. *Ann Epidemiol*. 2012;22(3):213-20. DOI: 10.1016/j.annepidem.2011.11.005.

[148] Macklaim JM, Clemente JC, Knight R, Gloor GB, Reid G. Changes in vaginal microbiota following antimicrobial and probiotic therapy. *Microb Ecol Health Dis*. 2015;26:27799. DOI: 10.3402/mehd.v26.27799.

[149] Russo R, Karadja E, De Seta F. Evidence-based mixture containing *Lactobacillus* strains and lactoferrin to prevent recurrent bacterial vaginosis: a double blind, placebo controlled, randomised clinical trial. *Benef Microbes*. 2019;10(1):19-26. DOI: 10.3920/BM2018.0075.

[150] Doppalapudi R, Vundavalli S, Prabhat MP. Effect of probiotic bacteria on oral *Candida* in head- and neck-radiotherapy patients: A randomized

clinical trial. *J Cancer Res Ther.* 2020;16(3):470-477. DOI: 10.4103/jcrt.JCRT_334_18.

[151] Hu L, Mao Q, Zhou P, Lv X, Hua H, Yan Z. Effects of *Streptococcus salivarius* K12 with nystatin on oral candidiasis-RCT. *Oral Dis.* 2019;25(6):1573-1580. DOI: 10.1111/odi.13142.

[152] Lee X, Vergara C, Lozano CP. Severity of *Candida*-associated denture stomatitis is improved in institutionalized elders who consume *Lactobacillus rhamnosus* SP1. *Aust Dent J.* 2019;64(3):229-236. DOI: 10.1111/adj.12692.

[153] Miyazima TY, Ishikawa KH, Mayer M, Saad S, Nakamae A. Cheese supplemented with probiotics reduced the *Candida* levels in denture wearers-RCT. *Oral Dis.* 2017;23(7):919-925. DOI: 10.1111/odi.12669.

[154] Invernici MM, Furlaneto FAC, Salvador SL, Ouwehand AC, Salminen S, Mantziari A, Vinderola G, Ervolino E, Santana SI, Silva PHF, Messoria MR. *Bifidobacterium animalis* subsp *lactis* HN019 presents antimicrobial potential against periodontopathogens and modulates the immunological response of oral mucosa in periodontitis patients. *PLoS One.* 2020;15(9):e0238425. DOI: 10.1371/journal.pone.0238425.

[155] Sabatini S, Lauritano D, Candotto V, Silvestre FJ, Nardi GM. Oral probiotics in the management of gingivitis in diabetic patients: a double blinded randomized controlled study. *J Biol Regul Homeost Agents.* 2017;31(2 Suppl 1):197-202.

[156] Tobita K, Watanabe I, Tomokiyo M, Saito M. Effects of heat-treated *Lactobacillus crispatus* KT-11 strain consumption on improvement of oral cavity environment: a randomised double-blind clinical trial. *Benef Microbes.* 2018;9(4):585-592. DOI: 10.3920/BM2017.0137.