

2019

## Data on foliar nutrient concentration of invasive plants in the recipient habitat and their native habitat

Pushpa Soti

*The University of Texas Rio Grande Valley*, [pushpa.soti@utrgv.edu](mailto:pushpa.soti@utrgv.edu)

Matthew Purcell

Krish Jayachandran

Follow this and additional works at: [https://scholarworks.utrgv.edu/bio\\_fac](https://scholarworks.utrgv.edu/bio_fac)



Part of the [Biology Commons](#), and the [Plant Sciences Commons](#)

---

### Recommended Citation

Soti, P., Purcell, M. F., & Jayachandran, K. (2019). Data on foliar nutrient concentration of invasive plants in the recipient habitat and their native habitat. *Data in brief*, 25. <https://doi.org/10.1016/j.dib.2019.104201>

This Article is brought to you for free and open access by the College of Sciences at ScholarWorks @ UTRGV. It has been accepted for inclusion in Biology Faculty Publications and Presentations by an authorized administrator of ScholarWorks @ UTRGV. For more information, please contact [justin.white@utrgv.edu](mailto:justin.white@utrgv.edu), [william.flores01@utrgv.edu](mailto:william.flores01@utrgv.edu).



ELSEVIER

Contents lists available at ScienceDirect

## Data in brief

journal homepage: [www.elsevier.com/locate/dib](http://www.elsevier.com/locate/dib)

## Data Article

## Data on foliar nutrient concentration of invasive plants in the recipient habitat and their native habitat

Pushpa Soti <sup>a,\*</sup>, Matthew F. Purcell <sup>b</sup>, Krish Jayachandran <sup>c</sup><sup>a</sup> University of Texas Rio Grande Valley, USA<sup>b</sup> USDA ARS Australian Biological Control Laboratory, Australia<sup>c</sup> Florida International University, USA

## ARTICLE INFO

## Article history:

Received 12 April 2019

Received in revised form 13 June 2019

Accepted 20 June 2019

Available online 27 June 2019

## Keywords:

Exotic invasive species

Plant growth

## ABSTRACT

Higher foliar nitrogen concentration in plants is often attributed to higher biomass assimilation and subsequently higher plant growth rate. To understand the underlying mechanism of extensive growth rate of an invasive plant, Old World climbing fern (*Lygodium microphyllum*), we analyzed the leaf tissue samples from the native and invaded habitats. In each habitat we selected 3 different locations with varying habitat characteristics (soil type, land use history and coexisting vegetation). Plant aboveground tissue collected from each site were analyzed for macro and micro nutrients. Total C and N were measured with a Truspec CN Analyzer. Total Ca, Fe, Mg, K, Mn, and P in plant tissue samples were measured using inductively coupled plasma mass spectrometry (ICP–MS). Here we present the difference in foliar nutrient concentration of invasive plant species in their native habitats and invaded habitats.

© 2019 The Authors. Published by Elsevier Inc. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

## 1. Data

We present the data collected from an extensive survey of a highly invasive plant, *Lygodium microphyllum*, in its native habitat in Queensland, Australia and recipient habitat in Florida,

\* Corresponding author.

E-mail address: [Pushpa.soti@utrgv.edu](mailto:Pushpa.soti@utrgv.edu) (P. Soti).

Specifications Table

| Subject area               | Ecology  |
|----------------------------|--|
| More specific subject area | Invasion ecology, plant sciences   |
| Type of data               | Map, table and pictures  |
| How data was acquired      | Field survey and lab analysis using ICP –MS, Truspec CN analyzer   |
| Data format                | Raw and analyzed   |
| Experimental factors       | Fully grown young plant tissue were collected for nutrient analysis from all the sampling sites.   |
| Experimental features      | Data were subjected to analysis of variance (ANOVA) using SAS Version 9.2 software, and means were separated using Fisher LSD ( $P$ -values $\leq 0.05$ ).   |
| Data source location       | Australia (Native)<br>Site 1.16° 15' 25.57" S, 145° 24' 3.94" E<br>Site 2.27° 40' 4.16" S, 153° 16' 0.44" E<br>Site 3.27° 22' 31.12" S, 153° 5' 39.42" E<br>Florida (Recipient)<br>Site 1.26° 4' 0.04" N, 80° 16' 5.88" W<br>Site 2.28° 23' 4.03" N, 81° 44' 41.30" W<br>Site 3.27° 0' 37.33" N, 80° 7' 20.28" W |
| Data accessibility         | Data are available within this article.  |
| Related research article   | Soti, P. G., Jayachandran, K., Purcell, M., Volin, J. C., & Kitajima, K. (2014). Mycorrhizal symbiosis and <i>Lygodium microphyllum</i> invasion in south Florida—a biogeographic comparison. <i>Symbiosis</i> , 62 (2), 81–90 [1].  |

#### Value of the Data

- To our knowledge this is the first comparative data on foliar nutrients concentration of an invasive plants growing in their native habitats and in the invaded or recipient habitats.
- This dataset can potentially provide some insight on the extensive aboveground growth and nutrient turnover rate of an invasive species in the recipient habitats.
- This data can be useful to researchers studying the ecology of exotic invasive plants.

United States (Fig. 1). The difference in above ground growth of *L. microphyllum* in both the habitats is presented in Fig. 2. Data on the variation in the plant tissue nutrient content is presented in Table 1.

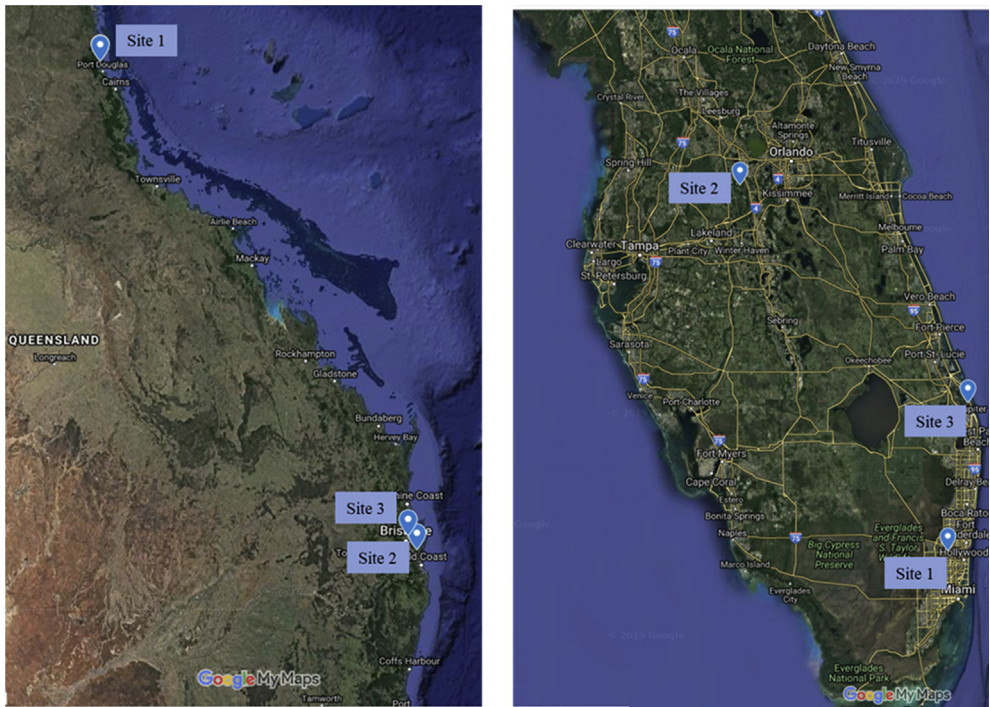
## 2. Experimental design, materials, and methods

### 2.1. Sampling sites

Leaf tissue samples of wild *L. microphyllum* were collected from 3 different locations each in south Florida and Queensland, Australia (Fig. 1). In each location young, fully-grown above-ground plant tissue samples were collected from 6 different plants selected randomly resulting a total of 36 samples.

### 2.2. Sample processing and analysis

The plant tissue samples were dried in an oven at 60 °C for one week and finely ground using a mortar and pestle. Total C and N were measured with a Truspec CN analyzer. Total Ca, Fe, Mg, K, Mn, and P in plant tissue samples were measured with an ICP –MS at USDA, ARS Laboratory, Miami, Florida.



**Fig. 1.** Sampling sites in native habitat, Queensland Australia (left) and recipient habitat, Florida (right).

Samples for ICP-MS analysis were prepared following the slightly modified acid digestion method [2]. 0.5 g of finely ground plant tissue samples were transferred to large glass tubes and mixed with 10 ml of 30%  $\text{HNO}_3$ . The tubes were covered with a vapor recovery system and heated to  $95 \pm 5$  °C and refluxed for 10 minutes without boiling under the hood in a heating block maintained with a Partlow Mic 6000 Profile Process Controller. After cooling to 40 °C, 2 ml of DI water and 3 ml of 30%  $\text{H}_2\text{O}_2$  was added and heated until the effervescence subsided. The samples were cooled and diluted to 50 ml with DI water, centrifuged at 2000 rpm for 10 minutes and filtered with a Whatman No. 41 filter paper.

### 2.3. Data analysis

Data on the foliar nutrient concentration were subjected to analysis of variance (ANOVA) using SAS Version 9.2 software. Means were separated using Fisher LSD ( $P$ -values  $\leq 0.05$ ).



**Fig. 2.** Above ground growth of *L. microphyllum* in native habitat (left) and recipient habitat (right). In the recipient habitats in Florida, *L. microphyllum* grows over trees up to 30 m in height and creates thick fern mats, smothering trees and shrubs, however in their native habitats these plants are much smaller in height and do not create a thick fern mat.

**Table 1**

Nutrient concentration in the leaf tissue from the two habitats (recipient and native) of *L. microphyllum*. Different letters within the column indicate significantly different means at the 0.05 level.

|           | Ca (mg g <sup>-1</sup> ) | Fe (mg g <sup>-1</sup> ) | Mg (mg g <sup>-1</sup> ) | K (mg g <sup>-1</sup> ) | Mn (mg g <sup>-1</sup> ) | Zn (mg g <sup>-1</sup> ) | C:N     |
|-----------|--------------------------|--------------------------|--------------------------|-------------------------|--------------------------|--------------------------|---------|
| Recipient |                          |                          |                          |                         |                          |                          |         |
| Site 1    | 5.73b                    | 0.22a                    | 2.28ab                   | 23.54ab                 | 0.12a                    | 0.10a                    | 12.70c  |
| Site 2    | 7.37a                    | 0.14b                    | 1.99bc                   | 20.49bc                 | 0.10b                    | 0.09ab                   | 16.11b  |
| Site 3    | 5.11b                    | 0.15b                    | 2.55a                    | 18.97c                  | 0.09b                    | 0.06c                    | 12.33c  |
| Native    |                          |                          |                          |                         |                          |                          |         |
| Site 1    | 3.24cd                   | 0.12b                    | 1.68c                    | 18.10c                  | 0.08c                    | 0.07bc                   | 25.00a  |
| Site 2    | 2.82d                    | 0.12b                    | 1.83c                    | 24.94a                  | 0.06d                    | 0.10a                    | 23.738a |
| Site 3    | 3.97c                    | 0.14b                    | 1.73c                    | 19.63c                  | 0.08c                    | 0.08abc                  | 12.33a  |

## Acknowledgments

The authors acknowledge Cheryl Millett, Central Florida Nature Conservancy; Patricia L. Howell, Natural Areas Specialist, Broward County; and Jeffrey Bach, Environmental Specialist, Jonathan Dickinson State Park, for their help in site selection and sample collection. We also thank staff of the USDA ARS Australian Biological Control Laboratory for assistance with sample collecting in Australia. We thank Dr. Stewart Reed, USDA-ARS, Subtropical Horticulture Research: Miami, FL, for his help in sample analysis.

### **Conflict of interest**

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

### **Appendix A. Supplementary data**

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.dib.2019.104201>.

### **References**

- [1] P.G. Soti, K. Jayachandran, M. Purcell, J.C. Volin, K. Kitajima, Mycorrhizal symbiosis and *Lygodium microphyllum* invasion in south Florida—a biogeographic comparison, *Symbiosis* 62 (2) (2014) 81–90.
- [2] J. Retka, A. Maksymowicz, D. Karmasz, Determination of Cu, Ni, Zn, Pb, Cd by ICP-MS and Hg by AAS in Plant Samples, in: *Proceedings of 15th International Conference on Heavy Metals in the Environment*, 2010, p. 1071.