

EUROPEAN
HEMATOLOGY
ASSOCIATIONFerrata Storti
Foundation

Low-count monoclonal B-cell lymphocytosis persists after seven years of follow up and is associated with a poorer outcome

Ignacio Criado,¹ Arancha Rodríguez-Caballero,¹ M. Laura Gutiérrez,¹ Carlos E. Pedreira,² Miguel Alcoceba,³ Wendy Nieto,¹ Cristina Teodosio,¹ Paloma Bárcena,¹ Alfonso Romero,⁴ Paulino Fernández-Navarro,⁵ Marcos González,³ Julia Almeida,^{1*} Alberto Orfao^{1*} and The Primary Health Care Group of Salamanca for the Study of MBL

Haematologica 2018
Volume 103(7):1198-1208

¹Cancer Research Centre (IBMCC, USAL-CSIC), Department of Medicine and Cytometry Service (NUCLEUS), University of Salamanca, IBSAL and CIBERONC, Spain; ²Systems and Computing Department (PESC), COPPE, Federal University of Rio de Janeiro (UFRJ), Brazil; ³Hematology Service, University Hospital of Salamanca, IBMCC, IBSAL, CIBERONC and Department of Nursery and Physiotherapy, University of Salamanca, Spain; ⁴Centro de Atención Primaria de Salud Miguel Armijo, Salamanca, Sanidad de Castilla y León (SACYL), Spain and ⁵Centro de Atención Primaria de Salud de Ledesma, Salamanca, Sanidad de Castilla y León (SACYL), Spain

*AO and JA contributed equally to this work.

ABSTRACT

Low-count monoclonal B-cell lymphocytosis is defined by the presence of very low numbers of circulating clonal B cells, usually phenotypically similar to chronic lymphocytic leukemia cells, whose biological and clinical significance remains elusive. Herein, we re-evaluated 65/91 low-count monoclonal B-cell lymphocytosis cases (54 chronic lymphocytic leukemia-like and 11 non-chronic lymphocytic leukemia-like) followed-up for a median of seven years, using high-sensitivity flow cytometry and interphase fluorescence *in situ* hybridization. Overall, the clone size significantly increased in 69% of low-count monoclonal B-cell lymphocytosis cases, but only one subject progressed to high-count monoclonal B-cell lymphocytosis. In parallel, the frequency of cytogenetic alterations increased over time (32% vs. 61% of cases, respectively). The absolute number of the major T-cell and natural killer cell populations also increased, but only among chronic lymphocytic leukemia-like cases with increased clone size vs. age- and sex-matched controls. Although progression to chronic lymphocytic leukemia was not observed, the overall survival of low-count monoclonal B-cell lymphocytosis individuals was significantly reduced vs. non-monoclonal B-cell lymphocytosis controls ($P=0.03$) plus the general population from the same region ($P\leq 0.001$), particularly among females ($P=0.01$); infection and cancer were the main causes of death in low-count monoclonal B-cell lymphocytosis. In summary, despite the fact that mid-term progression from low-count monoclonal B-cell lymphocytosis to high-count monoclonal B-cell lymphocytosis and chronic lymphocytic leukemia appears to be unlikely, these clones persist at increased numbers, usually carrying more genetic alterations, and might thus be a marker of an impaired immune system indirectly associated with a poorer outcome, particularly among females.

Correspondence:

orfao@usal.es

Received: November 3, 2017.

Accepted: March 15, 2018.

Pre-published: March 22, 2018.

doi:10.3324/haematol.2017.183954

Check the online version for the most updated information on this article, online supplements, and information on authorship & disclosures: www.haematologica.org/content/103/7/1198

©2018 Ferrata Storti Foundation

Material published in *Haematologica* is covered by copyright. All rights are reserved to the Ferrata Storti Foundation. Use of published material is allowed under the following terms and conditions:

<https://creativecommons.org/licenses/by-nc/4.0/legalcode>.

Copies of published material are allowed for personal or internal use. Sharing published material for non-commercial purposes is subject to the following conditions:

<https://creativecommons.org/licenses/by-nc/4.0/legalcode>,

sect. 3. Reproducing and sharing published material for commercial purposes is not allowed without permission in writing from the publisher.



Introduction

Chronic lymphocytic leukemia (CLL) is the most common leukemia in adults in the Western world, typically affecting older patients, particularly males, with a median age at diagnosis of 70 years (y) old.¹ It is characterized by the accumulation of mature B cells in peripheral blood (PB), bone marrow (BM) and also secondary lymphoid tissues, with a uniquely aberrant CD19⁺ CD20^{lo} CD5^{+/++} CD23⁺ sIgM^{+/+lo} phenotype and restricted immunoglobulin (Ig) light chain usage.^{2,3} Typically, CLL

shows a heterogeneous clinical outcome; thus, whereas in some patients the disease remains stable and they will never require treatment, in around 70% of cases treatment is required and results in variable outcomes, from complete response and prolonged survival to refractory disease and death.³⁻⁵

Currently, it is well established that virtually every CLL case is preceded by monoclonal B-cell lymphocytosis (MBL) defined by smaller numbers of circulating PB clonal CLL-like B-cells (<5,000 clonal B-cells/ μ L) in the absence of any clinical symptoms or signs of disease.⁶ In 2010, MBL was further subdivided into low-count (MBL^{lo}) and high-count MBL (MBL^{hi}), depending on the number of PB clonal B cells (lower vs. higher than $0.5 \times 10^9/L$, respectively).⁷ While MBL^{hi} has been reported to progress to overt CLL requiring treatment at a rate of 1–2% cases per year,^{8,9} no information is available at present regarding the ≥ 5 -year risk of progression of MBL^{lo} to MBL^{hi} and CLL.¹⁰

The detection of MBL^{lo} has become routinely feasible due to the use of highly sensitive flow cytometry (FCM) approaches for the screening of subjects from the general population who present normal blood cell counts. Of note, the prevalence of MBL^{lo} is significantly higher than that of MBL^{hi} and CLL, with a frequency that ranges between 3% and 14% of the general adult (≥ 40 y) population, depending on the sensitivity of the FCM technique used.¹¹ Independently of the method, it is well-established that the incidence of MBL^{lo} progressively increases with age, with a prevalence >20% among individuals of more than 70 years of age.¹² Whether MBL^{lo} represents the normal counterpart of CLL (e.g., some studies suggest that MBL^{lo} clones are more likely related to immunosenescence)¹³ or a very early stage of development of CLL, remains an open question. This is partially because, in contrast to MBL^{hi}, long-term follow-up studies in large series of MBL^{lo} cases have not been reported thus far, which limits our understanding of the biological and clinical significance of very low numbers of circulating CLL-like clones, as well as those factors and mechanisms involved in potential long-term progression of (conceivably) a minor proportion of all MBL^{lo} cases to MBL^{hi} and CLL; likewise, little information is available about the evolution of non CLL-like MBL. Such information is critical to a better understanding of the ontogenesis of CLL from the very early stages of the disease, and to better identify MBL patients with stable vs. progressive B-cell lymphocytosis who might benefit from a closer clinical follow-up.

Herein, we report on a cohort of 91 MBL^{lo} (CLL-like and non CLL-like) subjects identified in a population-based screening study and followed for a minimum of five years (median >seven years). Our primary goal was to determine the rate of medium-term progression of MBL^{lo} to MBL^{hi} and CLL, and to identify the most relevant clinical and biological characteristics of PB lymphocytes associated with progression.

Methods

Subjects and samples

The baseline study was conducted from December 2007 to October 2009, when PB samples from 639 healthy adult (≥ 40 y) volunteers (54% females/46% males) from the general population of the same geographical area (Salamanca, Northwest of Spain) were screened for the presence of small B-cell clones, using highly

sensitive FCM.^{12,14} At inclusion, all subjects had normal PB cell counts and did not suffer from any hematological/immunological disease, as described elsewhere.^{1,6} In 91/639 subjects studied (14.2%), ≥ 1 PB clonal B-cell population was detected at recruitment; in the vast majority of them (80/91; 88%) clonal B cells were consistent with CLL-like MBL^{lo} (< 0.5×10^9 clonal B cells/L showing a CLL-like phenotype), whereas the remaining 11 individuals (12%) were classified as non CLL-like MBL^{lo}.^{12,14} MBL^{lo} subjects were re-evaluated at a median time of seven years after recruitment (range: 61 to 95 months). All subjects gave their written informed consent at baseline for both the initial and the follow-up studies, and they filled out an epidemiological questionnaire with demographic and (self-reported) medical information, under the supervision of his/her primary care doctor.¹⁵ The study was approved by the Ethics Committee of the University Hospital of Salamanca (Spain).

Flow cytometry immunophenotypic studies

Overall, 1–4 mL of ethylenediamine tetraacetic acid (EDTA)-anticoagulated PB was collected per case and follow-up time-point; subsequently it was processed and analyzed using previously reported highly sensitive FCM approaches.^{12,14,16,17} (*Online Supplementary Methods* and *Online Supplementary Table S1*).

Interphase fluorescence *in situ* hybridization (iFISH) studies

The most common CLL - i.e., del(13q14), trisomy 12, del(11q)(*ATM*) and del(17p)(*TP53*) - along with other B-cell chronic lymphoproliferative disorders (B-CLPD)-associated cytogenetic alterations were investigated by iFISH on fluorescence-activated cell sorting (FACS)-purified (sorted) single clonal B cells ($\geq 95\%$ purity), as previously described¹⁸ (*Online Supplementary Table S2*). A total of 31/91 PB samples studied at baseline and 56/65 at follow-up (year +7) were analyzed by iFISH; in 21 cases (18 CLL-like and three non CLL-like MBL^{lo}) paired samples were analyzed by iFISH at both baseline and year +7. The potential presence of del(13q14) was also tested in non-clonal B-cells from 5/7 MBL^{lo} cases found to have del(13q14)⁺ MBL cells.

Statistical analyses

All conventional statistical analyses (i.e., descriptive statistics, univariate analyses, including overall survival (OS) analysis, as well as multivariate analyses to predict the variables independently associated with a greater/lower risk of death), were performed with SPSS 19.0 software (SPSS-IBM, Armonk, NY, USA), using the tests, databases and statistical significance values detailed in *Online Supplementary Methods*. Appropriate tests were further used to objectively evaluate real changes in the size of the B-cell clones studied during follow-up (resampling bootstrap method)¹⁹ and to build a predictive linear regression model to estimate the time CLL-like MBL^{lo} clones might potentially take to progress to MBL^{hi} and CLL, using MATLAB R2015a (Mathworks, Natick, MA, USA) (*Online Supplementary Methods*).

Results

Follow-up of the MBL^{lo} cohort

From those 91 MBL^{lo} individuals identified in the screening study performed in the general population of Salamanca between 2007 and 2009,^{12,14} 65–71% of MBL^{lo} cases from the original series; 29 males and 36 females; median age at baseline 70 (range: 43–84 years old)-; were re-evaluated after a median follow-up of seven years (range: 61 to 95 months) (Table 1). These 65 individuals

were representative of the original MBL^{lo} cohort for all variables analyzed, except for a significantly lower age ($P=0.02$) vs. those 26 individuals that could not be followed - median age of 75 (range: 48-95 years)-. These later subjects could only be re-evaluated for their death vs. alive status at the end of the study because of: i) 12/26 (46%) died before the fifth year of follow-up; ii) 2 subjects declined continuing their participation in the study; and iii) the remaining 12 cases were lost to follow-up after >5y from recruitment. Eight of 65 cases followed for >5y (12%) died afterward, making a total of 21 (26%) deaths among MBL^{lo} cases included in OS analyses.

In all 65 individuals who were evaluated after five years, ≥ 1 clonal B-cell population was reliably identified in PB at follow-up. In 22/65 (34%) cases ≥ 2 clones were detected (vs. 32% at baseline), resulting in a total of 86 MBL^{lo} clones detected (Table 1 and Table 2). All MBL^{lo} clones showed an identical phenotype at both time-points (Table 2). Thus, 74/86 B-cell clones (86%) showed a classical CLL-like phenotype and 12 (14%) were classified as non CLL-like MBL clones. At year +7, 35/74 CLL-like clones (47%) corresponded to monoclonal cases and the remaining 39 (53%), to 19 subjects with bi(multi)clonal CLL-like MBL^{lo} (Table 2); in two subjects, CLL-like and non CLL-like clones

Table 1. Clinical and biological characteristics of MBL^{lo} subjects at baseline and after follow-up (year +7).

	All subjects (n=65)		CLL-like MBL ^{lo} subjects (n=54)		Non CLL-like MBL subjects (n=11)		P
	Baseline	Follow-up	Baseline	Follow-up	Baseline	Follow-up	
Follow-up time (months)	0	84 (61-95)	0	84 (61-95)	0	83 (63-87)	NA
Male/Female*	29/36 (45%/55%)		22/32 (41%/59%)		7/4 (64%/36%)		NA
Age, years	70 (43-84)	75 (49-91)	68 (43-84)	75 (49-91)	76 (58-81)	83 (65-88)	<0.01 ^{abc}
Leukocytosis (>10x10 ⁹ /L)*	0 (0%)	2 (3%)	0 (0%)	2 (3%)	0 (0%)	0 (0%)	NS
Lymphocytosis (>4x10 ⁹ /L)*	0 (0%)	3 (5%)	0 (0%)	2 (4%)	0 (0%)	1 (9%)	NS
N. total T cells/ μ L	1261 (341-2428)	1448 (276-3753)	1290 (341-2428)	1508 (460-3753)	1111 (796-1965)	1206 (276-2907)	<0.01 ^{ab}
N. CD4 ⁺ T cells/ μ L	687 (253-1572)	840 (184-2045)	684 (253-1572)	898 (227-2045)	732 (351-1395)	629 (184-1995)	0.015 ^{ab}
N. CD8 ⁺ T cells/ μ L	449 (71-1154)	491 (66-1742)	446 (71-1154)	479 (96-1742)	453 (237-750)	617 (66-848)	<0.03 ^{ab}
N. CD4 ⁺ /CD8 ⁺ T cells/ μ L	4.3 (0.19-38)	8.2 (1.1-147)	4.5 (0.55-37)	8.2 (1.3-147)	4.3 (0.19-27)	8.2 (1.1-29)	<0.02 ^{abc}
N. CD4 ⁺ /CD8 ⁻ T cells/ μ L	56 (8.1-254)	62 (1.9-407)	58 (8.0-214)	64 (1.9-338)	36 (11-254)	34 (8.1-406)	<0.05 ^{ab}
N. NK cells/ μ L	304 (76-1138)	373 (87-3415)	297 (76-1138)	373 (89-3415)	394 (150-848)	372 (178-937)	0.001 ^{ab}
N. total B cells/ μ L	133 (26-1173)	155 (22-1218)	132 (41-478)	150 (28-1218)	137 (26-1173)	190 (22-1207)	<0.01 ^{ab}
N. normal B cells/ μ L	119 (23-478)	116 (21-536)	126 (37-478)	140 (26-536)	72 (23-136)	45 (21-190)	0.08 ^b
N. clonal B cells/ μ L	0.99 (0.03-1101)	2.0 (0.05-1149)	0.75 (0.03-66)	1.7 (0.05-808)	56 (0.62-1101)	90 (1.3-1149)	<0.001 ^{ab}
Subjects with ≥ 2 MBL clones*	21 (32%)	22 (34%)	18 (33%)	19 (35%)	3 (27%)	3 (27%)	NS
Progression* (to MBL ^{hi})	NA	1 (2%)	NA	1 (2%)	NA	0 (0%)	NA
Deaths*	NA	8 (12%)	NA	7 (13%)	NA	1 (9%)	NA

CLL-like or non CLL-like with ≥ 1 B-cell clone with different phenotypes were classified depending on the phenotype of the larger clone. Results expressed as median (range) or * as number of cases (percentage). ^aBaseline vs. follow-up (year +7) for all cases. ^bBaseline vs. follow-up (year +7) for CLL-like MBL cases. ^cBaseline vs. follow-up (year +7) for non CLL-like MBL cases. CLL: chronic lymphocytic leukemia; MBL^{hi}: high-count monoclonal B-cell lymphocytosis; MBL^{lo}: low-count monoclonal B-cell lymphocytosis; N: number; NA: not applicable; NK: natural killer; NS: not statistically significantly different ($P>0.05$).

coexisted. Of note, two individuals carrying two CLL-like B-cell clones became “monoclonal” while a second clone emerged in one monoclonal CLL-like MBL^{lo} case at seven years follow-up. In turn, non CLL-like clones (n=12) showed phenotypic profiles identical to those observed at baseline and comparable to those of different B-CLPD, as detailed in *Online Supplementary Table S3*.⁶

Clonal B-cell load in PB at re-evaluation (year +7).

Overall, a significant ($P \leq 0.001$) increase in the median size of MBL^{lo} clones was found at follow-up, both for CLL-like (≈ 2 -fold median increase) and for non CLL-like MBL^{lo} clones (≈ 3 -fold median increase) (Table 2 and Figure 1A,B). Such increased absolute number of clonal B-cells over time was associated with a significantly increased ($P \leq 0.001$) percentage of clonal B cells from all PB B cells (Table 2). In detail, most MBL^{lo} clones (59/86; 69%) showed significantly increased numbers at re-evaluation vs. baseline, while the remaining 27 B-cell clones persisted at similar (16%) or lower levels (15%); this behavior was very similar for CLL-like and non CLL-like clones (Table 2). Of note, 30/35 (86%) CLL-like clones from (mono)clonal cases increased in size at follow-up vs. only 21/39 (54%) clones from bi(multi)clonal cases ($P=0.004$). Interestingly, among non CLL-like clones, most marginal zone lymphoma-like clones increased (5/6; 83%), while the two mantle cell lymphoma-like B-cell clones decreased significantly in number (*Online Supplementary Table S3*).

Cytogenetic alterations of MBL^{lo} clonal B cells at baseline and follow-up

The overall frequency of CLL-like MBL^{lo} cases carrying CLL-associated cytogenetic alterations, for example del(13q14), trisomy 12, del(11q)(ATM) and del(17p)(TP53), at baseline was of 29% (7/24 cases tested). At recruitment del(13q14)(D13S25) was found in 56%±34% cells from 6/20 cases evaluated (30%), the RB1 gene was additionally involved in 3 of them, and trisomy 12 was present in the remaining case (59% of cells), both as single alterations. After seven years of follow-up, the percentage of cytogenetic altered cases augmented to 62% of MBL^{lo} cases (31/50 cases, including 15 cases studied at baseline). Interestingly, all cytogenetic alterations observed at base-

line also remained at follow-up; in addition, 4/15 (27%) individuals studied at both time-points further acquired del(13q14)(D13S25) (*Online Supplementary Table S4*). Overall, del(13q14)(D13S25) remained the most frequent alteration at follow-up (27/48; 56%), affecting 32±27% of CLL-like cells. Of note, in five cases in which clonal B cells showed del(13q14)(D13S25), non-clonal B cells were also studied for this alteration, and was found to be absent in all of them. RB1 gene involvement was identified in only 1/7 cases tested; furthermore, trisomy 12 was restricted to one patient who had the same abnormality at baseline (Table 3). Clonal B cells from one individual in whom del(17p)(TP53) was not investigated at baseline was found to carry this cytogenetic alteration in 10% of cells at follow-up. Alterations involving 14q32 were investigated only at follow-up in a subset of 20 CLL-like MBL^{lo} cases, being found in five (20%) patients (Table 3).

Regarding non CLL-like clones, t(11;14)(q13-q32) was detected in 100% of clonal B cells from one of the two MCL-like cases studied, while del(7q32) was detected in 2/5 splenic marginal zone lymphoma (SMZL)-like cases (Table 3). None of the cases investigated showed t(14;18) (*data not shown*).

Distribution of normal residual T-, B- and NK-cell populations

The PB counts of total T cells and their CD4⁺CD8⁻, CD8⁺CD4⁻ and CD4⁻CD8^{+/lo} subsets, as well as NK cells and normal residual polyclonal B cells was significantly increased ($P < 0.05$) in CLL-like MBL^{lo} at follow-up vs. baseline (Table 1). In contrast, among non CLL-like MBL cases, CD4⁺CD8⁺ T cells were the only lymphoid subset significantly increased ($P=0.02$) at the seven year follow-up. To rule out a potential age-related bias and further confirm these findings, we compared the number of PB normal lymphocyte subsets at seven years follow-up vs. a large series of non-MBL healthy donors matched per age and sex distribution to the CLL-like MBL^{lo} cases at seven years (*Online Supplementary Table S5*) and the same differences were found, ruling out an impact of sex or more advanced age on the increased PB residual lymphocyte counts. No significant correlation ($P > 0.05$) was revealed between the absolute number of clonal B cells and any of the normal residual PB lymphocyte subsets analyzed (*data not shown*).

Table 2. Biological characteristics of MBL^{lo} clones at baseline and at follow-up (year +7).

	All clones (n=86)		CLL-like MBL ^{lo} clones (n=74)		Non CLL-like MBL clones (n=12)		P
	Baseline	Follow-up	Baseline	Follow-up	Baseline	Follow-up	
N. of clones from monoclonal/Bi(multi)clonal subjects*	44/42 (51%/49%)	42/44 (49%/51%)	36/38 (49%/51%)	35/39 (47%/53%)	8/4 (67%/33%)	7/5 (54%/46%)	NS
N. of clones that increased*	NA	59 (69%)	NA	51 (69%)	NA	8 (67%)	NA
N. clonal B cells/ μ L	0.06 (0.03-1101)	1.3 (0.05-1146)	0.46 (0.03-66)	0.85 (0.05-789)	37 (0.57-1101)	68 (1.3-1146)	<0.001 ^{ab}
% clonal B cells (from total B cells)	0.48% (0.02%-94%)	0.95% (0.02%-97%)	0.35% (0.02%-21%)	0.73% (0.02%-65%)	30% (0.46%-94%)	60% (1.4%-97%)	<0.03 ^{abc}

Results expressed as median (range) or as * number of cases (percentage). ^aBaseline vs. follow-up (year +7) for all cases. ^bBaseline vs. follow-up (year +7) for CLL-like MBL clones. ^cBaseline vs. follow-up (year +7) for non CLL-like MBL clones. CLL: chronic lymphocytic leukemia; MBL^{lo}: low-count monoclonal B-cell lymphocytosis; N: number; NA: not applicable; NS: not statistically significantly different ($P > 0.05$).

Clinical and biological characteristics of CLL-like MBL^{lo} at baseline vs. follow-up, according to the kinetics of the B-cell clone

Upon comparing CLL-like MBL^{lo} cases with increased vs. stable/decreased clonal B-cell numbers at seven years follow-up, the former had a similar male/female distribution, but they were significantly younger (median age: 68y vs. 78y; Table 4).

Strikingly, MBL^{lo} cases who showed larger CLL-like clone sizes over time also showed significantly higher ($P<0.05$) numbers of the distinct normal residual T-, B- and NK-cell subsets at follow-up (vs. baseline) (Table 4). Moreover, in these subjects a direct correlation was observed between the absolute number of clonal B cells and CD4⁺CD8⁻ T cells ($r^2=0.5$; $P=0.001$). In contrast, no significant ($P>0.05$) association was found between higher numbers of clonal CLL-like B cells in PB over time, and an increased frequency of cytogenetic alterations. Interestingly, del(13q14) was the sole genetic alteration detected at the seven year follow-up within cases with stable/decreased CLL-like B-cell clones, while those cases with increased CLL-like B-cell clones at year +7 showed cytogenetic alterations other than del(13q14), e.g., trisomy 12 (1/40), del(17p)(*TP53*) (1/39) and t(14q32) (5/20 cases tested) (Table 4).

Clinical outcome of MBL^{lo} cases

Three subjects developed absolute lymphocytosis after seven years of follow-up (median: 5.3×10^9 lymphocytes/L; range: 4.1×10^9 - 5.9×10^9 /L) in the absence of signs of disease. Two had CLL-like B-cell clones carrying del(13q14), while the remaining case had a non CLL-like clone. In one of the two CLL-like MBL^{lo} cases, the size of the B-cell clone increased over the threshold for MBL^{hi} (>500 clonal B cells/ μ L), while the other two cases remained as MBL^{lo}. Remarkably, these three subjects displayed the highest increase in clone size at re-evaluation: this translated into a significantly lower (estimated) time to progression into CLL (median: 95y; range: 54-128y) according to the predictive mathematical model used. In turn, the estimated time to progression to CLL for the other MBL^{lo} individuals was far beyond a normal life expectancy (median: 54,767y; range: 54->63 million years).

Overall survival of MBL vs. non-MBL individuals

At the end of the study (January 2017), the clinical records and epidemiological questionnaires from all individuals recruited at baseline were reviewed. During follow-up, 21/89 (24%) MBL^{lo} cases and 41/290 (14%) age- and sex-matched non-MBL^{lo} subjects from the original cohort had died ($P=0.03$). Though the median OS for the two groups

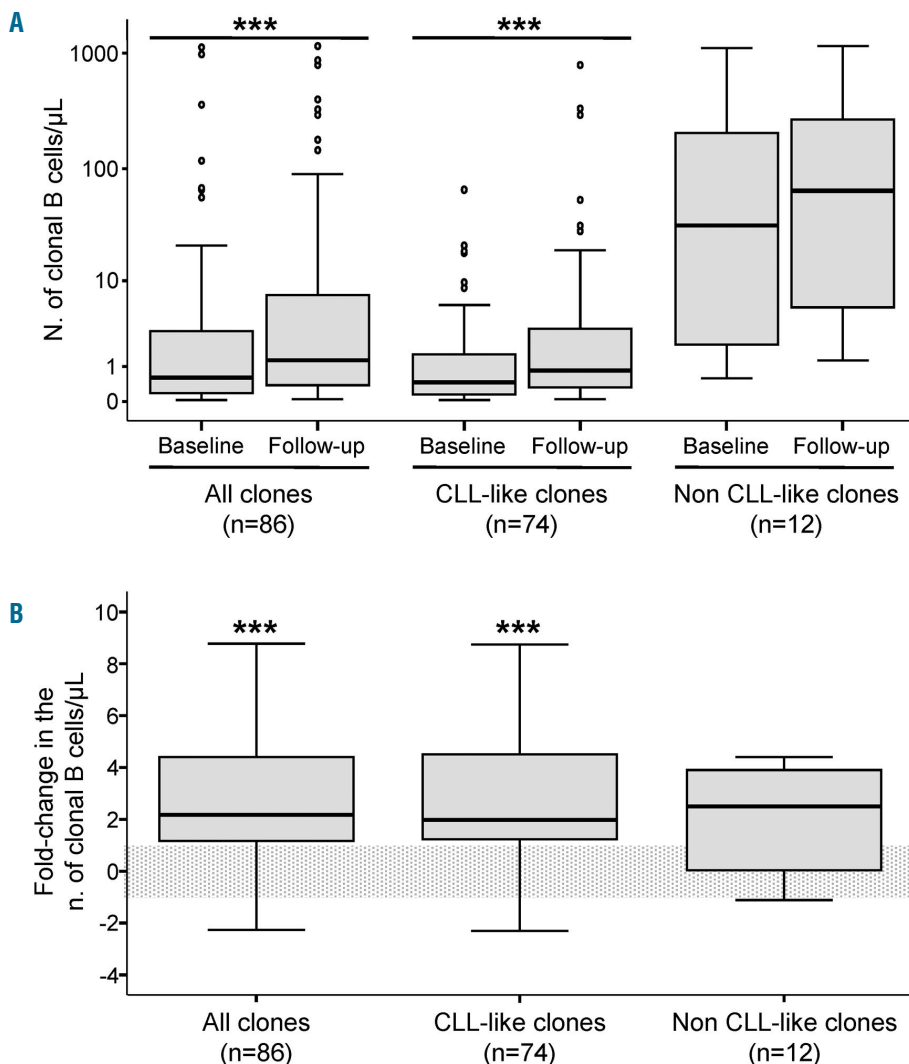


Figure 1. Changes in the number of clonal B cells during follow-up. Panel A shows the absolute number of PB clonal B cells/ μ L detected in MBL^{lo} individuals at baseline and at follow-up, according to the phenotype of the clonal population. Panel B represents the fold-change in the number of clonal B cells/ μ L from baseline, which is represented by the horizontal light gray box. Notched boxes represent 25th and 75th percentile values; the lines in the middle correspond to median values (50th percentile) and vertical lines represent the highest and lowest values that are neither outliers nor extreme values, which are represented as single dots. *** P -value <0.001. N: number; CLL: chronic lymphocytic leukemia.

had not been reached yet, a significantly shorter OS was observed for MBL^{lo} individuals vs. age- and sex-matched non-MBL controls from the same cohort and geographical area (10y OS rates of 76% vs. 86%, respectively; $P=0.03$) (Figure 2A,B). Moreover, MBL^{lo} subjects also showed a significantly shortened survival vs. age-matched individuals of the general population from the same geographical region (8.0% vs. 1.8% in the period 2015-2016, respectively; $P<0.001$) (Online Supplementary Figure S1). Interestingly, such differences in OS were at the expense of a lower OS of CLL-like MBL^{lo} females, who showed a significantly ($P=0.01$) higher risk of death (hazard ratio (HR) of 2.5; 95% confidence interval (CI) of 1.2-5.4) than non-MBL females of the same age (Figure 2C,F). Infections (21%; mostly respiratory infections and sepsis), cancer (36%; all solid tumors except for an essential thrombocythemia) and cardiovascular diseases (29%; i.e., myocardial infarction and acute ischemic stroke) were the main causes of death among MBL^{lo} subjects. Overall, infections were overrepresented among the MBL^{lo} cohort vs. age- and sex-matched subjects from the general population of the same geographical area (21% vs. 1.4%, respectively; $P\leq 0.001$). In contrast, the proportion of deaths caused by tumors (36% vs. 26%, respectively; $P>0.05$) and by cardiovascular diseases (29% vs. 33%, respectively; $P>0.05$) were similar in both groups. In

turn, no MBL^{lo} subjects died as a cause of non-infectious respiratory tract diseases or genitourinary diseases, diabetes, dementia or other nervous system disorders, which accounted for $\approx 30\%$ of deaths in the age- and sex-matched general population cohort living in the same geographical area. In order to identify those variables independently associated with OS, a multivariate Cox regression analysis, including laboratory, epidemiological and medical information, was carried out. Advanced age- HR of 5.1; 95% CI: 1.5-17.5; $P=0.01$ -, co-existing cardiovascular diseases (HR: 2.7; 95% CI: 1.3-5.4; $P=0.01$), solid tumors (HR: 2.9; 95% CI: 1.3-6.5; $P=0.007$) and, to a lesser extent, the presence of MBL^{lo} clones (HR: 2.1, 95% CI: 0.97-4.7; $P=0.06$), were independently associated with a shorter OS in the whole cohort (Table 5 and Online Supplementary Table S6).

Discussion

Several preceding studies have shown that virtually all CLL cases are preceded by MBL^{hi},^{8,20,21} in contrast, such a relationship has not been demonstrated for MBL^{lo} cases, its role as a preleukemic condition still remaining to be confirmed.^{9,21} In fact, there exist very few studies with short-term follow-up (i.e., $\leq 3y$) which have investigated

Table 3. Frequency of cases with CLL-associated cytogenetic alterations and percentage of cells affected by each genetic abnormality.

	All MBL ^{lo} cases		CLL-like MBL ^{lo} cases		Non CLL-like MBL ^{lo} cases		P
	Baseline (n=31)	Follow-up (n=56)	Baseline (n=24)	Follow-up # (n=50)	Baseline (n=7)	Follow-up (n=6)	
N. of cases with cytogenetic alterations (%)	10/31 (32%)	34/56 (61%)	7/24 (29%)	31/50 (62%)	3/7 (43%)	3/6 (50%)	0.01 ^{ab}
Chromosomal region							
del(13q14) (<i>DI3S25</i>)	7/22 (32%)	28/54 (52%)	6/20 (30%)	27/48 (56%)	1/2 (50%)	1/6 (17%)	0.06 ^c
% altered cells	49 \pm 36%	31 \pm 27%	56 \pm 34%	32 \pm 27%	8%	7%	NA
del(13q14) (<i>RBI1</i>)	3/15 (20%)	1/7 (14%)	3/15 (20%)	1/7 (14%)	NA	NA	NS
% altered cells	14 \pm 3%	47%	14 \pm 3%	47%			
Trisomy 12	2/21 (10%)	2/55 (3.6%)	1/19 (5.3%) [¥]	1/49 (2%) [¥]	1/2 (50%)	1/6 (17%)	NS
% altered cells	34 \pm 35%	45 \pm 35%	59%	70%	9%	20%	
del(11q) (<i>ATM</i>)	2/12 (17%)	1/54 (1.9%)	0/10 (0%)	0/48 (0%)	2/2 (100%)	1/6 (17%)	NS
% altered cells	39 \pm 44%	50%	NA	NA	39 \pm 44%	50%	
del(17p) (<i>TP53</i>)	1/10 (10%)	1/54 (1.9%)	0/8 (0%)	1/48 (2.1%)	1/2 (50%)	0/6 (0%)	NS
% altered cells	13%	10%	NA	10%	13%	NA	
t(14q32)*	0/5 (0%)	7/27 (26%)	NA	5/23 (22%)	0/4 (0%)	2/4 (50%)	NS
% altered cells	NA	33 \pm 30%		31 \pm 33%	NA	38 \pm 30%	
t(11;14)(q13-q32)	1/2 (50%)	NA	NA	NA	1/2 (50%)	NA	NA
% altered cells	100%				100%		
del(7q32)	0/1 (0%)	2/5 (40%)	NA	NA	0/1 (0%)	2/5 (40%)	NS
% altered cells	NA	20 \pm 2.1%			NA	20 \pm 2.1%	
3q27 (<i>BCL6</i>)	0/1 (0%)	0/5 (0%)	NA	NA	0/1 (0%)	0/5 (0%)	NS
% altered cells	NA	NA			NA	NA	
18q21 (<i>MALT1</i>)	0/2 (0%)	0/4 (0%)	NA	NA	0/2 (0%)	0/4 (0%)	NA
% altered cells	NA	NA			NA	NA	

Results expressed as number of cases (percentage of cases) and mean \pm SD of percentage of cells affected by each specific genetic alteration. ^aBaseline vs. follow-up (year +7) for all cases. ^bBaseline vs. follow-up (year +7) for CLL-like MBL cases. ^cBaseline vs. follow-up (year +7) for non CLL-like MBL cases. [¥]2/50 individuals carried a clonal MBL^{lo} CLL-like population along with at least one MBL^{lo} non CLL-like clone. ^{*}The same case at baseline and follow-up. ^{*}Other than t(11;14). CLL: chronic lymphocytic leukemia; MBL^{lo}: low-count monoclonal B-cell lymphocytosis; N: number; NA: not applicable; NS: not statistically significantly different ($P>0.05$).

the progression rate from MBL^{lo} to MBL^{hi} and CLL thus far.^{10,22,25} Hence, Fazi *et al.* showed persistent MBL^{lo} clones over time in 90% of CLL-like MBL^{lo} and only 67% of non CLL-like clones, after a median follow-up of ≈ 3 y.¹⁰ Herein, we demonstrate the systematic persistence of both CLL-like and non CLL-like MBL^{lo} B-cell clones with an identical phenotype to baseline after seven years follow-up in 65/65 MBL^{lo} cases, confirming that MBL^{lo} is not a transient condition. Similarly, Matos *et al.* also found the persistence of B-cell clones in their limited series of CLL-like MBL^{lo} cases (n=5) after a median follow-up of ≈ 7 y.²³ Interestingly, in 3/56 CLL-like MBL^{lo} cases, the number of clones identi-

fied at seven years follow-up changed, which might suggest the emergence of MBL^{lo} from an oligoclonal background that mirrors competition and natural selection among multiple coexisting clones.²⁴ Changes observed in the VDJ sequences of the expanded B cells from most of these cases (*data not shown*), together with the progressively decreasing rate of oligoclonality from MBL^{lo} (12-19%) to MBL^{hi} (2.9-13%) and CLL (0.7-3.4%), would further support this hypothesis.^{9,12,25-27} The significance of such oligoclonal B-cell expansions in MBL^{lo} remains unknown, but might be the consequence of the early stages of altered oligoclonal immune responses against multiple antigens,

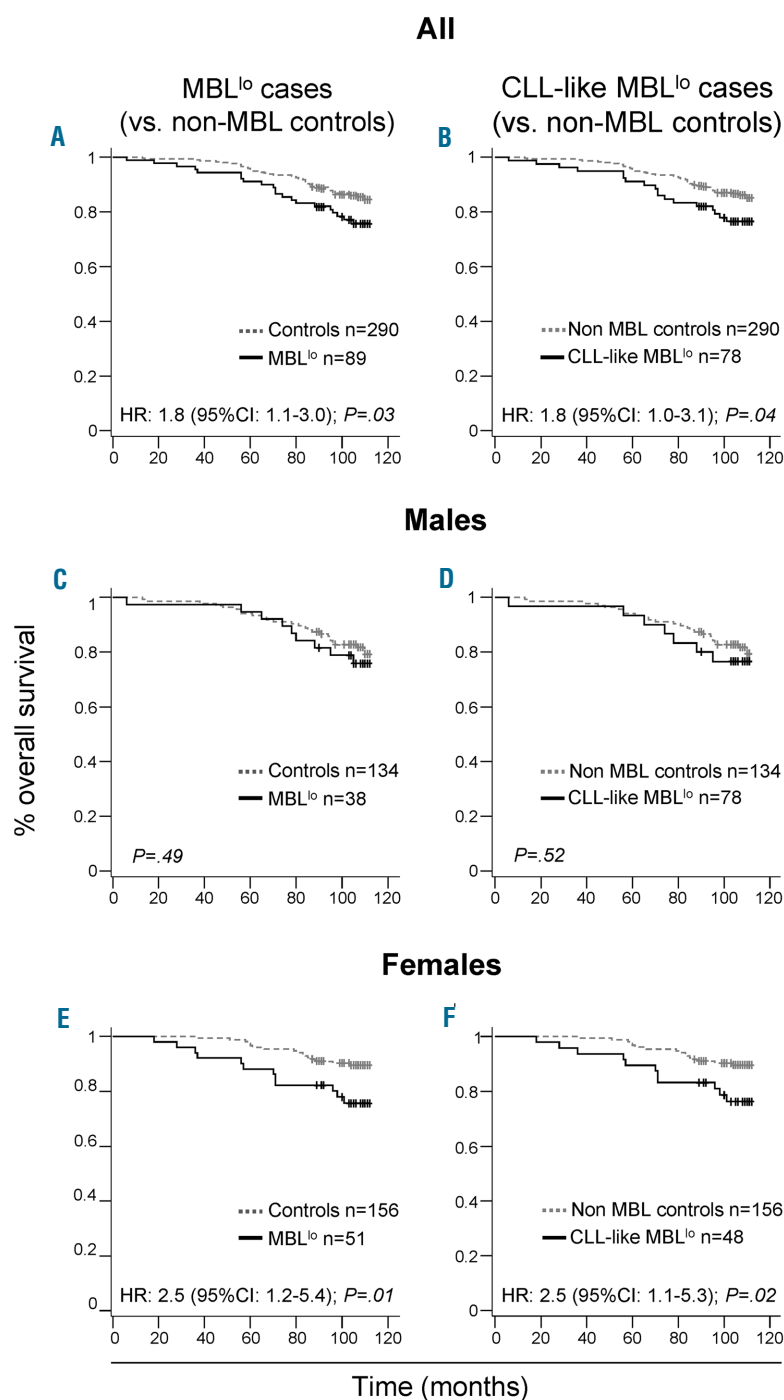


Figure 2. Overall survival from baseline (mortality rates) of MBL^{lo} individuals vs. age- and sex-matched non-MBL controls. Left column panels represent comparisons of overall survival curves from MBL^{lo} subjects (black) and age- and sex-matched non-MBL controls (dotted gray). The same comparison is depicted for all individuals (Panel A) and separately for males (Panel C) and females (Panel E). In the right column, overall survival curves comparing all MBL^{lo} subjects with a CLL-like phenotype vs. all age- and sex-matched non-MBL controls (Panel B). The same subjects distributed according to sex are shown in Panel D (males) and in Panel F (females). MBL^{lo}: low-count monoclonal B-cell lymphocytosis.

Table 4. Clinical and biological characteristics of CLL-like MBL^b subjects at baseline and at follow-up (+7 years) according to the kinetics of the MBL clone in PB (decreased/stable vs. increased size).

	CLL-like MBL ^b subjects (n=56)		P
	Decrease/stable B-cell clones (n=9)	Increased B-cell clones (n=47)#	
Male/Female*	4/5 (44%/56%)	18/29 (38%/62%)	NS
Age at baseline, years	78 (55-84)	68 (43-81)	0.03
Monoclonal at follow-up*	5/9 (56%)	30/47 (64%)	NS
Leukocytosis (>10x10 ⁹ /L) at follow-up *	0 (0%)	2 (4%)	NS
Lymphocytosis (>4x10 ⁹ /L) at follow-up *	0 (0%)	2 (4%)	NS
N. total T cells/ μ L			
Baseline	1471 (1105-2035)	1285 (341-2428)	NS
follow-up	1406 (711-2313)	1520 (460-3753)	NS
P	NS	<0.01	
N. CD4 ⁺ T cells/ μ L			
Baseline	821 (461-1186)	448 (253-1572)	NS
follow-up	792 (295-1327)	908 (227-2045)	NS
P	NS	<0.01	
N. CD8 ⁺ T cells/ μ L			
Baseline	452 (374-900)	448 (72-1154)	NS
follow-up	491 (245-1469)	467 (96-1742)	NS
P	NS	0.02	
N. CD4 ⁺ /CD8 ⁺ T cells/ μ L			
Baseline	4.3 (0.97-17)	4.6 (0.55-37)	NS
follow-up	4.7 (2.3-13)	8.6 (1.3-147)	0.03
P	NS	<0.001	
N. CD4 ⁻ /CD8 ⁻ T cells/ μ L			
Baseline	70 (8.2-214)	58 (8.0-190)	NS
follow-up	60 (7.2-272)	65 (1.9-338)	NS
P	NS	0.02	
N. total B cells/ μ L			
Baseline	110 (41-263)	139 (50-1066)	NS
follow-up	80 (29-390)	175 (28-1218)	0.02
P	NS	<0.01	
N. normal B cells/ μ L			
Baseline	94 (37-256)	122 (50-478)	NS
follow-up	79 (26-389)	140 (27-536)	NS
P	NS	0.03	
N. clonal B cells/ μ L			
Baseline	0.80 (0.13-23)	0.71 (0.03-66)	NS
follow-up	0.60 (0.05-3.2)	2.0 (0.10-808)	0.03
P	0.02	<0.001	
% clonal B cells			
Baseline	0.92 (0.10-20)	0.53 (0.02-21)	NS
follow-up	0.44 (0.04-10)	1.0 (0.06-66)	NS
P	0.05	<0.001	
N. NK cells/ μ L			
Baseline	304 (167-874)	292 (76-1138)	NS
follow-up	492 (310-1066)	361 (87-3415)	NS
P	NS	<0.01	
Cytogenetic alterations			
Baseline	0/4 (0%)	6/15 (38%)	NS
follow-up	6/9 (67%)	26/41 (63%)	NS
P	0.03	0.14	
del(13q14)(D13S25)			
Baseline	0/2 (0%)	5/13 (39%); 57 \pm 38%	NS
follow-up	6/9 (67%); 17 \pm 9%	21/39 (54%); 37 \pm 29%	NS

continued in the next page

del(13q14) (<i>RBI</i>)			
Baseline	0/2 (0%)	3/13 (23%); 14±3.2%	NS
follow-up	NA	1/7 (14%); 47%	NA
Trisomy 12			
Baseline	0/3 (0%)	1/11 (9%); 59%	NS
follow-up	0/9 (0%)	1/40 (3%); 70%	NS
del(11q) (<i>ATM</i>)			
Baseline	0/2 (0%)	0/6 (0%)	NS
follow-up	0/9 (0%)	0/39 (0%)	NS
del(17p) (<i>TP53</i>)			
Baseline	0/2 (0%)	0/5 (0%)	NS
follow-up	0/9 (0%)	1/39 (3%); 10%	NS
t(14q32)			
Baseline	NA	NA	NA
follow-up	0/3 (0%)	5/20 (25%); 31±33%	NS

²/56 individuals carried a clonal MBL^{lo} CLL-like population along with at least one MBL^{lo} non CLL-like clone. Results expressed as median (range) or as *number of cases (percentage). Cytogenetic alterations are expressed as percentage of cases and mean percentage of cells affected ± SD. *P*-values shown in the right column refer to comparisons between MBL^{lo} subjects who showed decreased/stable vs. increase clone sizes, while *P*-values shown in rows represent differences among subjects within each group at baseline and after seven years of follow-up. CLL: chronic lymphocytic leukemia; MBL^{lo}: low-count monoclonal B-cell lymphocytosis; N.: number; NA: not applicable; NK: natural killer; NS: not statistically significantly different (*P*>0.05).

in which a single clone had not yet emerged as dominant vs. the others, as might occur at the latter, e.g., CLL stage.

Most importantly, over two thirds of all CLL-like MBL^{lo} clones showed a significantly increased size in PB after seven years, while for non CLL-like clones more variable kinetics were observed, depending on the specific phenotype of clonal B-cells. Interestingly, we also observed a significant increase in the frequency of cytogenetic alterations over time, evidencing that B-cell clones are not only dynamic in terms of clone size, but also regarding their capacity to acquire new cytogenetic alterations. Of note, del(13q14), which has been found to be a common mosaicism in the general population,^{28,29} was absent in non-clonal B cells from 5/5 cases investigated in which CLL-like clonal cells did carry this alteration, indicating that the emergence of this alteration in MBL^{lo} is specific for the clonal population. Altogether, these findings suggest that cytogenetic alterations are a relatively early, but not primary, event in the natural history of MBL/CLL, and might have a potential role in the progression of MBL^{lo} to MBL^{hi} and CLL.

The presence and type of cytogenetic lesions, the *IGHV* mutational status, or the presence of stereotyped receptors are some of the most important prognostic factors in CLL, which also define the outcome of MBL^{hi} individuals; furthermore, it might identify a subset of cases in whom the presence of the B-cell clonal population influences OS.³⁰⁻³³ Unfortunately, in the present study, the mutational status and *VDJ* rearrangements were only assessed (both baseline and follow-up) in 8/65 MBL^{lo} individuals (*data not shown*), making it impossible to validate solid conclusions regarding the potential association with the risk for progression into MBL^{hi} and CLL. To the best of our knowledge, the frequency and impact on disease progression of recurrent mutations (i.e., *NOTCH1*, *SF3B1*, *MYD88*, etc.) found in CLL, and also in MBL^{hi}, to a lesser extent, has not been elucidated for MBL^{lo}.³⁴⁻³⁶ Therefore, analysis of these CLL-related mutations in MBL^{lo} cases might further contribute to an improvement in better delineating intrinsic tumor cell factors associated to disease progression.

In addition, the environment in which CLL-like MBL^{lo}

Table 5. Variables studied in the Cox regression multivariate analysis showing an independent impact (*P*<0.1) on OS for the whole MBL^{lo} plus non-MBL cohort.

Variables	HR (95%CI)	<i>P</i>
Whole cohort		
Cardiovascular disease	2.65 (1.30 - 5.41)	0.007
Age (<65y vs. ≥65y)	5.08 (1.48 - 17.49)	0.01
Solid tumor	2.86 (1.26 - 6.46)	0.01
MBL ^{lo} clones	2.14 (0.97 - 4.72)	0.06

CI: confidence interval; HR: hazard ratio; MBL^{lo}: low-count monoclonal B-cell lymphocytosis; N: number; OS: overall survival; PB: peripheral blood. The complete list of variables analyzed in the Cox regression model is provided in *Online Supplementary Table S6*.

clones develop might be influenced by chronic immune responses against e.g., host viruses, that might play a critical role in the expansion of clonal B cells, as recently suggested.³⁷ In line with this hypothesis, herein we also show that the expansion of CLL-like MBL^{lo} clones after seven years of follow-up (vs. baseline) is accompanied by a significant increase of all T-cell (but CD4⁺CD8⁺ cytotoxic T-cells) and NK-cell populations in PB.

Controversial results have been reported regarding PB T-cell numbers in MBL^{lo}. Hence, while te Raa *et al.* found normal CD4⁺ and CD8⁺ T-cell counts in PB of MBL^{hi},³⁸ other studies have demonstrated that around half of the MBL^{lo} individuals show ≥1 clonal/oligoclonal CD4⁺CD8⁺ T-cell population, with an overall increased frequency of clonal T-cell populations vs. age-matched individuals from the general population.^{10,39} However, the presence of clonal (CD4⁺CD8⁺ and other) T-cell expansions has also been described as a common event in older individuals, and has been associated with the ageing of the immune system.³⁹ In this respect, we demonstrate herein that changes in the number of circulating PB T-cell and NK-cell populations among our CLL-like MBL^{lo} subjects were not age-related, *via* a parallel analysis of a large group of 250 age- and sex-matched non-MBL controls (*Online Supplementary Table S5*). From a pathophysiological point of view, the increase in most PB T- and NK-cell populations could be associated

with either a potentially protective or activating effect of these cellular components of the immune system (microenvironment) on the expanded clonal B-cells.^{40,41} Therefore, on one hand, increased numbers of (functionally impaired) T cells have been described in CLL^{38,42,43} while on the other hand, we have recently shown increased titers of plasma antibodies against CMV and EBV in MBL^{hi} and CLL patients vs. MBL^{lo} and non-MBL controls, despite their antibody (immune)deficient state.³⁷ Taken together, these latter findings might further support the existence of additional signals coming from immune cells other than clonal B cells, that could already contribute to the expansion of (cyto)genetically altered CLL-like clones at the earliest stages of disease, by promoting activation, proliferation and/or survival of specific B-cell clones.

A major goal of our study was to investigate the medium-term rate of progression of MBL^{lo} to MBL^{hi} and (potentially also) CLL. Overall, only one subject evolved from MBL^{lo} to MBL^{hi}, and none transformed to CLL, which would translate into a progression rate from MBL^{lo} to MBL^{hi} of 1.8% after seven years of follow-up. Despite the fact that the rate of progression of MBL^{lo} to MBL^{hi} and CLL appears to be extremely low, one of the most astonishing findings of our follow-up study was the significantly higher frequency of deaths among MBL^{lo} subjects, associated with a significant adverse impact on OS vs. both non-MBL controls, particularly among females, and the general population (of similar age and sex distribution) living in the same region in Spain. However, comparisons with the general population must be considered with care, since the conditions of this population might differ from that of non-MBL individuals recruited at the Primary Health Services. Multivariate analysis showed a borderline significant association between the presence of MBL^{lo} clones and a shorter survival. Despite this, the specific mechanisms responsible for the higher frequency of infections and deaths observed, particularly among women, are unknown, and further studies are required to validate and clarify these results. In this regard, controversial results have been reported on MBL^{hi} subjects in the literature. Thus, while Shanafelt *et al.* showed no differences in OS of MBL^{hi} vs. the general population,³³ Shim *et al.* pointed out a higher frequency of deaths in their MBL^{lo} cohort (4/11; 36%), albeit no statistically significant differences were found vs. non-MBL controls in the latter study, probably due to the small sample size.²² In addition, Fazi *et al.* also reported that 16/137 (12%) CLL-like MBL^{lo} subjects died before re-evaluation after a median time of three years, which is a high proportion of their whole cohort.¹⁰ However, in the aforementioned report no information about the age of the deceased subjects is provided, and therefore, if it is the case they were older (than those subjects remaining alive) such high mortality rates might have been expected. Even more strikingly is the overrepresenta-

tion of infections as causes of death in MBL^{lo} compared to that of our non-MBL cohort. Impaired immune responses and higher frequencies of infection have been recurrently reported in both MBL^{hi} and CLL,^{44–47} but so far very little information exists in MBL^{lo}, and such an association deserves further investigations. Several groups pointed out that the frequency of clonal hematopoiesis dramatically increases with age in the general population, especially among the elderly, in a similar way to the increased frequency of MBL^{lo}, reflecting a clear relationship between clonal hematopoiesis and a higher risk of death.^{48,49} Whether or not both phenomena are related to MBL^{lo} deserves future investigations.

In summary, we show herein that although MBL^{lo} is a persistent and dynamic condition with a progressive acquisition of cytogenetic alterations, usually associated with an increased clone size and higher T- and NK-cell numbers in PB over time, progression of MBL^{lo} to MBL^{hi} and CLL is extremely rare in the medium-term. Despite this, the MBL^{lo} subjects analyzed herein, particularly women, showed a shortened OS associated with an increased risk of death, particularly due to infections, further supporting the notion that MBL^{lo} could be a marker of an impaired immune system, indirectly associated with a poorer outcome. Additional studies are necessary to confirm these findings and shed light onto the specific immune defects and microenvironmental factors involved in MBL^{lo}.

Funding

This work was supported by the RD06/0020/0035 and RD12/0036/0048 grants from Red Temática de Investigación Cooperativa en Cáncer (RTICC), Instituto de Salud Carlos III, Ministerio de Economía y Competitividad, (Madrid, Spain and FONDOS FEDER); CB16/12/00400 grant (CIBERONC, Instituto de Salud Carlos III, Ministerio de Economía y Competitividad, (Madrid, Spain and FONDOS FEDER); the FIS PI06/0824-FEDER, PS09/02430-FEDER, PI12/00905-FEDER, DTS15/00119-FEDER, PI16/00787-FEDER and PI17/00399-FEDER grants, from the Fondo de Investigación Sanitaria of Instituto de Salud Carlos III; the GRS206/A/08 grant, (Ayuda al Grupo GR37 de Excelencia, SAN/1778/2009) from the Gerencia Regional de Salud (Consejería de Educación and Consejería de Sanidad of Castilla y León, Valladolid, Spain) and the SA079U14 grant (Consejería de Educación and Consejería de Sanidad of Castilla y León, Valladolid, Spain). ML Gutiérrez is supported by grant PTA2014-09963-I from the Instituto de Salud Carlos III.

Acknowledgments

The authors would like to thank “The Primary Health Care Group of Salamanca for the Study of MBL” for their contribution to the study; a complete list of members is included in the Online Supplementary Information.

References

- Rai KR, Jain P. Chronic lymphocytic leukemia (CLL)-Then and now. *Am J Hematol.* 2016;91(3):330–340.
- Hallek M, Cheson BD, Catovsky D, et al. Guidelines for the diagnosis and treatment of chronic lymphocytic leukemia : a report from the International Workshop on Chronic Lymphocytic Leukemia updating the National Cancer Institute – Working Group 1996 guidelines. *Blood.* 2008;111(12):5446–5456.
- Scarfo L, Ferreri AJM, Ghia P. Chronic lymphocytic leukaemia. *Crit Rev Oncol Hematol.* 2016;104:169–182.
- Bachow SH, Lamanna N. Evolving strategies for the treatment of chronic lymphocytic leukemia in the upfront setting. *Curr Hematol Malig Rep.* 2016;11(1):61–70.
- Chiorazzi N. Implications of new prognostic markers in chronic lymphocytic leukemia. *Hematol Am Soc Hematol Educ Progr.* 2012;2012:76–87.
- Swerdlow SH, Campo E, Harris NL, et al. WHO Classification of Tumours of

- Haematopoietic and Lymphoid Tissues. 4th ed. Lyon (France): IARC; 2017.
7. Rawstron AC, Shanafelt T, Lanasa MC, et al. Different biology and clinical outcome according to the absolute numbers of clonal B-cells in monoclonal B-cell lymphocytosis (MBL). *Cytometry B Clin Cytom.* 2010;78(Suppl. 1):19–23.
 8. Rawstron AC, Bennett FL, O'Connor SJM, et al. Monoclonal B-cell lymphocytosis and chronic lymphocytic leukemia. *N Engl J Med.* 2008;359(6):575–583.
 9. Vardi A, Dagklis A, Scarfo L, et al. Immunogenetics shows that not all MBL are equal: the larger the clone, the more similar to CLL. *Blood.* 2013;121(22):4521–4528.
 10. Fazi C, Scarfo L, Pecciarini L, et al. General population low-count CLL-like MBL persists over time without clinical progression, although carrying the same cytogenetic abnormalities of CLL. *Blood.* 2011;118(25):6618–6625.
 11. Shim YK, Middleton DC, Caporaso NE, et al. Prevalence of monoclonal B-cell lymphocytosis: a systematic review. *Cytometry B Clin Cytom.* 2010;78(Suppl. 1):10–18.
 12. Nieto WG, Almeida J, Romero A, et al. Increased frequency (12%) of circulating chronic lymphocytic leukemia-like B-cell clones in healthy subjects using a highly sensitive multicolor flow cytometry approach. *Blood.* 2009;114(1):33–37.
 13. Scarfo L, Fazi C, Ghia P. MBL versus CLL: how important is the distinction? *Hematol Oncol Clin North Am.* 2013;27(2):251–265.
 14. Nieto WG, Teodosio C, López A, et al. Non-CLL-like monoclonal B-Cell lymphocytosis in the general population: Prevalence and phenotypic/genetic characteristics. *Cytometry B Clin Cytom.* 2010;78(Suppl. 1):24–34.
 15. Casabonne D, Almeida J, Nieto WG, et al. Common infectious agents and monoclonal B-cell lymphocytosis: a cross-sectional epidemiological study among healthy adults. *PLoS One.* 2012;7(12):e52808.
 16. Kalina T, Flores-Montero J, van der Velden VHJ, et al. EuroFlow standardization of flow cytometer instrument settings and immunophenotyping protocols. *Leukemia.* 2012;26(9):1986–2010.
 17. Flores-Montero J, Sanoja-Flores L, Paiva B, et al. Next Generation Flow for highly sensitive and standardized detection of minimal residual disease in multiple myeloma. *Leukemia.* 2017;31(10):2094–2103.
 18. Quijano S, López A, Rasillo A, et al. Impact of trisomy 12, del(13q), del(17p), and del(11q) on the immunophenotype, DNA ploidy status, and proliferative rate of leukemic B-cells in chronic lymphocytic leukemia. *Cytometry B Clin Cytom.* 2008;74(3):139–149.
 19. Efron B, Tibshirani R. Bootstrap methods for standard errors, confidence intervals, and other measures of statistical accuracy. *Stat Sci.* 1986;1(1):54–77.
 20. Landgren O, Albitar M, Ma W, et al. B-cell clones as early markers for chronic lymphocytic leukemia. *N Engl J Med.* 2009;360(7):659–667.
 21. Shanafelt TD, Ghia P, Lanasa MC, Landgren O, Rawstron AC. Monoclonal B-cell lymphocytosis (MBL): biology, natural history and clinical management. *Leukemia.* 2010;24(3):512–520.
 22. Shim YK, Vogt RF, Middleton D, et al. Prevalence and natural history of monoclonal and polyclonal B-cell lymphocytosis in a residential adult population. *Cytometry B Clin Cytom.* 2007;72(5):344–353.
 23. Matos DM, Furtado FM, Falcao RP. Monoclonal B-cell lymphocytosis in individuals from sporadic (non-familial) chronic lymphocytic leukemia families persists over time, but does not progress to chronic B-cell lymphoproliferative diseases. *Rev Bras Hematol Hemoter.* 2015;37(5):292–295.
 24. Greaves M, Maley CC. Clonal evolution in cancer. *Nature.* 2012;481(7381):306–313.
 25. Henriques A, Rodriguez-Caballero A, Nieto WG, et al. Combined patterns of IGHV repertoire and cytogenetic/molecular alterations in monoclonal B lymphocytosis versus chronic lymphocytic leukemia. *PLoS One.* 2013;8(7):e67751.
 26. Sanchez M-L, Almeida J, Gonzalez D, et al. Incidence and clinicobiologic characteristics of leukemic B-cell chronic lymphoproliferative disorders with more than one B-cell clone. *Blood.* 2003;102(8):2994–3002.
 27. Kern W, Bacher U, Schmittger S, et al. Flow cytometric identification of 76 patients with biconal disease among 5523 patients with chronic lymphocytic leukaemia (B-CLL) and its genetic characterization. *Br J Haematol.* 2014;164(4):565–569.
 28. Jacobs KB, Yeager M, Zhou W, et al. Detectable clonal mosaicism and its relationship to aging and cancer. *Nat Genet.* 2012;44(6):651–658.
 29. Machiela MJ, Zhou W, Caporaso N, et al. Mosaic 13q14 deletions in peripheral leukocytes of non-hematologic cancer cases and healthy controls. *J Hum Genet.* 2016;61(5):411–418.
 30. Shanafelt TD, Kay NE, Rabe KG, et al. Brief report: natural history of individuals with clinically recognized monoclonal B-cell lymphocytosis compared with patients with Rai 0 chronic lymphocytic leukemia. *J Clin Oncol.* 2009;27(24):3959–3963.
 31. Kern W, Bacher U, Haferlach C, et al. Monoclonal B-cell lymphocytosis is closely related to chronic lymphocytic leukaemia and may be better classified as early-stage CLL. *Br J Haematol.* 2012;157(1):86–96.
 32. Strati P, Shanafelt TD. Monoclonal B-cell lymphocytosis and early-stage chronic lymphocytic leukemia: diagnosis, natural history, and risk stratification. *Blood.* 2015;126(4):454–462.
 33. Shanafelt TD, Kay NE, Rabe KG, et al. Survival of patients with clinically identified monoclonal B-cell lymphocytosis (MBL) relative to the age- and sex-matched general population. *Leukemia.* 2012;26(2):373–376.
 34. Landau DA, Tausch E, Taylor-Weiner AN, et al. Mutations driving CLL and their evolution in progression and relapse. *Nature.* 2015;526(7574):525–530.
 35. Barrio S, Shanafelt TD, Ojha J, et al. Genomic characterization of high-count MBL cases indicates that early detection of driver mutations and subclonal expansion are predictors of adverse clinical outcome. *Leukemia.* 2017;31(1):170–176.
 36. Agathangelidis A, Ljungström V, Scarfò L, et al. Highly similar genomic landscapes in monoclonal B-cell lymphocytosis and ultra-stable chronic lymphocytic leukemia with low frequency of driver mutations. *Haematologica.* 2018 Feb 15. [Epub ahead of print].
 37. Criado I, Muñoz-Criado S, Rodríguez-Caballero A, et al. Host virus and pneumococcus-specific immune responses in high-count monoclonal B-cell lymphocytosis and chronic lymphocytic leukemia: implications for disease progression. *Haematologica.* 2017;102(7):1238–1246.
 38. te Raa GD, Tonino SH, Remmerswaal EBM, et al. Chronic lymphocytic leukemia specific T-cell subset alterations are clone-size dependent and not present in monoclonal B lymphocytosis. *Leuk Lymphoma.* 2012;53(11):2321–2325.
 39. Ghia P, Prato G, Stella S, Scielzo C, Geuna M, Caligaris-Cappio F. Age-dependent accumulation of monoclonal CD4+CD8+ double positive T lymphocytes in the peripheral blood of the elderly. *Br J Haematol.* 2007;139(5):780–790.
 40. Purroy N, Wu CJ. Coevolution of leukemia and host immune cells in chronic lymphocytic leukemia. *Cold Spring Harb Perspect Med.* 2017;7(4):a026740.
 41. Os A, Bürgler S, Ribes AP, et al. Chronic lymphocytic leukemia cells are activated and proliferate in response to specific T helper cells. *Cell Rep.* 2013;4(3):566–77.
 42. Ramsay AG, Clear AJ, Fatah R, Gribben JG. Multiple inhibitory ligands induce impaired T-cell immunologic synapse function in chronic lymphocytic leukemia that can be blocked with lenalidomide: establishing a reversible immune evasion mechanism in human cancer. *Blood.* 2012;120(7):1412–1421.
 43. Riches JC, Davies JK, McClanahan F, et al. T cells from CLL patients exhibit features of T-cell exhaustion but retain capacity for cytokine production. *Blood.* 2013;121(9):1612–1621.
 44. Morrison VA. Infectious complications of chronic lymphocytic leukaemia: pathogenesis, spectrum of infection, preventive approaches. *Best Pract Res Clin Haematol.* 2010;23(1):145–153.
 45. Moreira J, Rabe KG, Cerhan JR, et al. Infectious complications among individuals with clinical monoclonal B-cell lymphocytosis (MBL): a cohort study of newly diagnosed cases compared to controls. *Leukemia.* 2013;27(1):136–141.
 46. Forconi F, Moss P. Perturbation of the normal immune system in patients with CLL. *Blood.* 2015;126(5):573–81.
 47. Solomon BM, Chaffee KG, Moreira J, et al. Risk of non-hematologic cancer in individuals with high-count monoclonal B-cell lymphocytosis. *Leukemia.* 2016;30(2):331–336.
 48. Jaiswal S, Fontanillas P, Flannick J, et al. Age-related clonal hematopoiesis associated with adverse outcomes. *N Engl J Med.* 2014;371(26):2488–2498.
 49. Genovese G, Kahler AK, Handsaker RE, et al. Clonal hematopoiesis and blood-cancer risk inferred from blood DNA sequence. *N Engl J Med.* 2014;371(26):2477–2487.