



LUND UNIVERSITY

Distal cholangiocarcinoma - from novel biomarkers to clinical management and outcome

Byrling, Johannes

2021

Document Version:

Publisher's PDF, also known as Version of record

[Link to publication](#)

Citation for published version (APA):

Byrling, J. (2021). *Distal cholangiocarcinoma - from novel biomarkers to clinical management and outcome*. Lund University, Faculty of Medicine.

Total number of authors:

1

General rights

Unless other specific re-use rights are stated the following general rights apply:

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal

Read more about Creative commons licenses: <https://creativecommons.org/licenses/>

Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

LUND UNIVERSITY

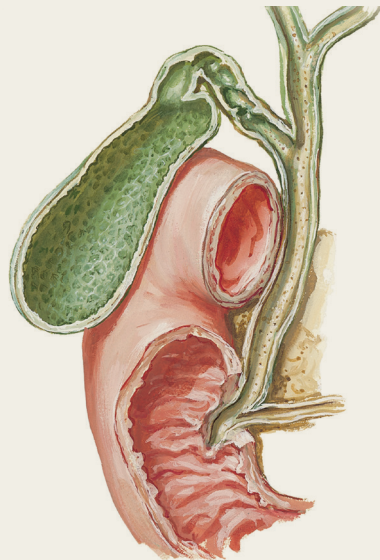
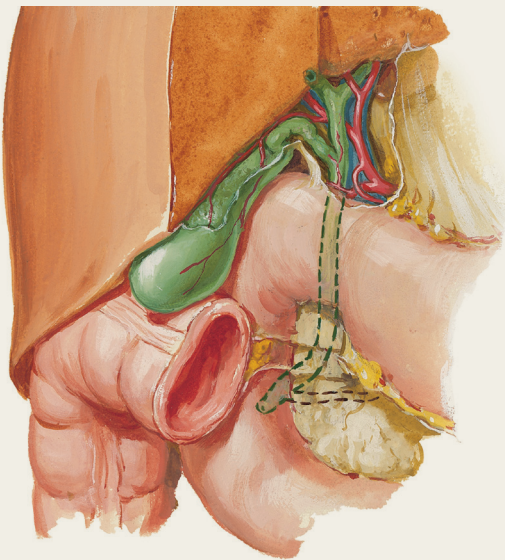
PO Box 117
221 00 Lund
+46 46-222 00 00

Distal cholangiocarcinoma

From novel biomarkers to clinical management and outcome

JOHANNES BYRLING

DEPARTMENT OF SURGERY | CLINICAL SCIENCES, LUND | LUND UNIVERSITY



Distal cholangiocarcinoma

Distal cholangiocarcinoma

From novel biomarkers to clinical management and
outcome

Johannes Byrling



LUND
UNIVERSITY

DOCTORAL DISSERTATION

by due permission of the Faculty of Medicine, Lund University, Sweden.
To be defended at Segerfalk Lecture Hall, BMC A1005, Sölvegatan 17, Lund, on
November 25th, 2021 at 09:00 am.

Faculty opponent

Professor Malin Sund, Department of Surgical and Periooperative
Sciences/Surgery, Umeå University, Umeå, Sweden.

Main supervisor: Associate professor Bodil Andersson.

Co-supervisors: Professor Roland Andersson, Associate professor Daniel Ansari.

Organization LUND UNIVERSITY Faculty of Medicine Department of Clinical Sciences Lund, Division of Surgery, Lund, Sweden		Document name Doctoral dissertation	
		Date of issue November 25 th 2021	
Author: Johannes Byrling		Sponsoring organization	
Title and subtitle Distal cholangiocarcinoma – from novel biomarkers to clinical management and outcome			
Abstract <p>Cholangiocarcinoma is an aggressive malignancy arising from the biliary tree. Anatomical subtypes of cholangiocarcinoma differs in tumor biology and clinical management. Distal cholangiocarcinoma (dCCA) originates from the common bile duct. Radical resection is the only curative treatment, for dCCA it entails a pancreatoduodenectomy (Whipple procedure). Other periampullary cancers treated with pancreatoduodenectomy include pancreatic cancer (PC), ampullary cancer (AC) and duodenal cancer (DC). There is a high rate of recurrence after resection for dCCA. This thesis aimed to evaluate the clinical management of dCCA but also improve understanding of the tumor biology and identify novel biomarkers.</p> <p>In paper I, the outcome and prognostic factors of patients treated with pancreatoduodenectomy for dCCA from 2008 through 2015 at Skane University Hospital were evaluated. We found the median survival to be 22 months which was worse than most previous studies. The presence of lymph node metastasis was confirmed as an important prognostic factor.</p> <p>In paper II, the expression of secreted protein acidic and rich in cysteine (SPARC) in resected dCCA specimens, paired lymph node metastases and normal bile ducts were evaluated using immunohistochemistry (IHC). We found SPARC to be expressed in the stromal compartment of dCCA in 80% of samples. Stromal expression was retained in 68% of lymph node metastases. There was no significant correlation between SPARC expression and survival.</p> <p>In paper III, bottom-up mass spectrometry (MS) followed by verification using parallel reaction monitoring (PRM) was used to identify differentially expressed proteins between dCCA samples and normal bile ducts. Bioinformatic analysis highlighted stromal alterations in dCCA. Forty-six proteins were verified using PRM. Thrombospondin-2 (THBS2) was further validated using IHC. We found THBS2 to be upregulated in dCCA epithelial and stromal compartments. Stromal THBS2 expression was present in 72% of paired lymph node metastases. There was a correlation between stromal THBS2 expression and poor disease-free survival.</p> <p>In paper IV, we studied the utility of serum THBS2 as a diagnostic biomarker for dCCA and PC. THBS2 levels were similar in dCCA and PC. THBS2 + CA 19–9 had an area under the curve of 0.92 in differentiating dCCA + PC from healthy donors. THBS2 did not provide utility in discriminating benign disease however, it was diagnosis dependent.</p> <p>In paper V, we used Swedish National Registry for Pancreatic and Periampullary Cancer to study national trends in frequency of tumor origin, survival, histopathological evaluation and diagnostic accuracy for patients with periampullary cancers. We found PC diagnosis to be more common in unresected patients. Survival was better for AC and DC than dCCA or PC. Median survival was 33 months for dCCA. Regional differences in tumor origin frequency and histopathological outcomes were identified. Clinical rate of misdiagnosis was 15 % for PC and 23% for non-pancreatic periampullary cancers.</p> <p>Key words distal cholangiocarcinoma, biliary tract cancer, periampullary cancer, pancreatoduodenectomy, lymph node metastasis, stroma, SPARC, THBS2, biomarker, mass spectrometry, diagnosis, histopathology</p> <p>Classification system and/or index terms (if any)</p>			
Supplementary bibliographical information		Language English	
ISSN and key title 1652-8220		ISBN 978-91-8021-129-1	
Recipient's notes	Number of pages 76		Price
	Security classification		

I, the undersigned, being the copyright owner of the abstract of the above-mentioned dissertation, hereby grant to all reference sources permission to publish and disseminate the abstract of the above-mentioned dissertation.

Signature

Johannes Byrling

Date 2021-10-20

Distal cholangiocarcinoma

From novel biomarkers to clinical management and
outcome

Johannes Byrling



LUND
UNIVERSITY

Coverphoto by Frank Netter. Netter illustrations used with permission of Elsevier Inc.

Copyright pp 1-76 Johannes Byrling

Paper 1 © Hellenic Society of Gastroenterology

Paper 2 © By the Authors (Open access)

Paper 3 © By the Authors (Open access)

Paper 4 © By the Authors (Open access)

Paper 5 © by the Authors (Manuscript unpublished)

Faculty of Medicine

Department of Surgery, Clinical Sciences, Lund

ISBN 978-91-8021-129-1

ISSN 1652-8220

Printed in Sweden by Media-Tryck, Lund University

Lund 2021



Media-Tryck is a Nordic Swan Ecolabel
certified provider of printed material.
Read more about our environmental
work at www.mediatryck.lu.se

MADE IN SWEDEN 

*“In the realm of ideas, everything depends on enthusiasm. In
the real world, all rests of perseverance.”*

-Johann Wolfgang von Goethe

Table of Contents

List of papers and manuscripts	10
Thesis at a glance	11
Populärvetenskaplig sammanfattning	12
Abbreviations	15
Introduction	17
Anatomy and physiology	17
Nomenclature	17
Epidemiology and risk factors.....	18
Pathobiology	19
Pathological classification	19
Molecular characterization	22
Tumor microenvironment.....	23
Diagnosis and prognosis.....	23
Clinical presentation and diagnostic workup.....	23
Biomarkers	24
Staging and survival.....	25
Clinical prognostic factors.....	26
Impact of histopathological evaluation.....	27
Treatment	27
Surgery	27
Palliative oncological treatment	28
Adjuvant oncological treatment	29
Biomarker investigation	30
Applications of mass spectrometry in cholangiocarcinoma	30
Secreted protein acidic and rich in cysteine	31
Thrombospondin-2	31
Aims of the thesis	32

Methodological considerations	33
Patient cohorts	33
Immunohistochemistry	34
Mass spectrometry	35
Enzyme-linked immunosorbent assay	36
Statistics	37
Ethics	39
Results summary	40
Paper I	40
Paper II	41
Paper III	42
Paper IV	48
Paper V	49
Discussion	51
Aspects of clinical management and outcome	51
Methodological considerations	52
Aspects of tumor biology and biomarkers	52
Methodological considerations	54
Main conclusions	56
Future perspectives	57
Acknowledgements	58
References	60

List of papers and manuscripts

This thesis is based on the following papers. They will be referred to in the text by the corresponding roman numerals.



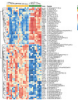


- I. Byrling J, Andersson R, Sasor A, Lindell G, Ansari D, Nilsson J, Andersson B. Outcome and evaluation of prognostic factors after pancreaticoduodenectomy for distal cholangiocarcinoma. *Annals of gastroenterology*. 2017; 30:571-577.
- II. Byrling J, Sasor A, Nilsson J, Said Hilmeresson K, Andersson R, Andersson B. Expression of peritumoral SPARC during distal cholangiocarcinoma progression and correlation with outcome, *Scandinavian Journal of Gastroenterology*. 2020; 55:725-731
- III. Byrling J, Kristl T, Hu D, Pla I, Sanchez A, Sasor A, Andersson R, Marko-Varga G, Andersson B. Mass spectrometry-based analysis of formalin-fixed, paraffin-embedded distal cholangiocarcinoma identifies stromal thrombospondin-2 as a potential prognostic marker. *Journal of translational medicine*. 2020; 18:343.
- IV. Byrling J, Said Hilmeresson K, Ansari D, Andersson R, Andersson B. Thrombospondin-2 as a diagnostic biomarker for distal cholangiocarcinoma and pancreatic ductal adenocarcinoma. *Clin Transl Oncol*. Epub 2021 Jul 29.
- V. Byrling J, Ghazi S, Andersson B. National trends in tumor origin, diagnostic accuracy and histopathological evaluation in patients with periampullary cancer. Manuscript.

Related papers not included in this thesis:

Byrling J, Andersson B, Marko-Varga G, Andersson R. Cholangiocarcinoma – current classification and challenges towards personalized medicine. *Scandinavian journal of gastroenterology*. 2016; 51:641-3.

Byrling J, Sasor A, Nilsson J, Said Hilmeresson K, Andersson R, Andersson B. Expression of fibroblast activation protein and the clinicopathological relevance in distal cholangiocarcinoma. *Scandinavian journal of gastroenterology*. 2020; 55:82-9.

Thesis at a glance

Paper	Objective	Methods	Results	Conclusions
I 	To evaluate survival and prognostic factors of surgically treated dCCA patients.	Clinicopathological data from patients treated with pancreatoduodenectomy for dCCA between 2008 and 2015 at SUS (N=54) were collected.	Median postoperative survival was 22.2 months. Lymph node metastasis was a negative prognostic factor.	Survival after surgery for dCCA was worse than previously presented. The presence of lymph node metastasis was prognostic.
II 	To investigate SPARC expression and prognostic impact in dCCA.	SPARC expression was evaluated by IHC in 59 resected dCCA samples and paired lymph node metastases.	SPARC expression was present in the stromal compartment of dCCA in 80% of patients and in 68% of lymph node metastases. There was no correlation with survival.	Stromal SPARC expression is common in dCCA and frequently retained during metastatic spread to lymph nodes.
III 	To identify dysregulated proteins in dCCA.	Mass spectrometry was used to analyze protein expression in resected dCCA specimens and normal bile ducts. THBS2 was validated using IHC.	Forty-six proteins were found to be dysregulated. Stromal THBS2 expression correlated with poor DFS.	Several dysregulated proteins not previously implicated in dCCA carcinogenesis were identified.
IV 	To analyze serum THBS2 expression and diagnostic biomarker properties in dCCA and PC.	THBS2 and CA 19-9 was measured in preoperative serum samples from patients with dCCA (N=51), PC (N=52), benign diseases (N=27) and healthy donors (N=52) using ELISA.	THBS2 levels were similar in dCCA and PC. THBS2+CA 19-9 had an AUC of 0.92 in differentiating dCCA+PC from healthy donors.	Serum THBS2 together with CA 19-9 has potential as a diagnostic biomarker for dCCA and PC.
V 	To evaluate national trends in tumor origin, survival, histopathological evaluation and diagnostic accuracy for patients with perampullary cancers in Sweden.	Data from patients diagnosed with periampullary cancer from 2010-2019 (N=9143) was retrieved from the Swedish National Registry for Pancreatic and Periampullary Cancer.	PC diagnosis was more common in palliative patients. Survival was better for DC and AC patients than dCCA or PC patients. Regional differences in histopathological outcomes were found. Rate of misdiagnosis was 15 and 23% for PC and non-pancreatic periampullary cancers respectively.	In palliative patients, non-pancreatic periampullary cancers are likely underdiagnosed. There is a need for further harmonization of histopathological evaluation methodology. The clinical rate of misdiagnosis for periampullary cancers needs to be considered.
Abbreviations: AC; ampullary cancer. dCCA; distal cholangiocarcinoma. DC; duodenal cancer. DFS; disease free survival. ELISA; enzyme-linked immunosorbent assay. IHC; immunohistochemistry. PC; pancreatic cancer. SPARC; secreted protein acidic and rich in cysteine. SUS; Skane University Hospital. THBS2; thrombospondin-2.				

Populärvetenskaplig sammanfattning

Gallvägscancer är en ovanlig cancersjukdom, men förödande för dem som drabbas. Radikal kirurgi är det enda botande behandlingsalternativet, dessvärre har en majoritet av patienterna ej botbar sjukdom vid diagnostillfället. Det finns undergrupper av gallvägscancer som baseras på den anatomiska utgångspunkten i gallträdet, dessa undergrupper kräver olika kirurgiskt omhändertagande men uppvisar även skillnader i tumörbiologi. Distal gallvägscancer uppkommer från den gemensamma gallgången. Den gemensamma gallgången sträcker sig från infästningen av gallblåsegången, genom bukspottskörtelns huvud ner till Vaters ampull. Vaters Ampull mynnar därefter i tolvfingertarmen. Det finns få studier på de olika undergrupperna av gallvägscancer, framförallt på distal gallvägscancer. I min doktorsavhandling har jag fokuserat på distal gallvägscancer och forskningen har syftat både till att förbättra det kliniska omhändertagandet men också att bättre förstå tumörbiologin och identifiera nya molekyllära markörer för sjukdomen.

För de patienter som har en resekel tumör utan fjärrspridning rekommenderas kirurgi med så kallad pankreatoduodenektomi, även känt som Whipples operation. Det är en stor operation och även stor risk för återfall efter kirurgi. Tidigare studier har visat stor variation i återfallsrisk och överlevnad efter operation. I min första studie analyserade vi utfallet för patienter som genomgått pankreatoduodenektomi för distal gallvägscancer vid Skånes Universitetssjukhus mellan 2008 och 2015 (54 patienter). Medianöverlevnaden (tiden till hälften av patienterna har avlidit) var 22 månader, detta var lägre än i de flesta tidigare studier. Förekomst av tumörspridning i bortopererade lymfkörtlar bekräftades som en viktig negativ prognostisk faktor.

Utöver cancercellerna består en tumör även av flera andra celltyper, exempelvis bindvävsceller och immunförsvarsceller. I tumören ingår även stödjevävnad. Vid distal gallvägscancer har man funnit att stora delar av tumörmassan utgörs av ärrbildningsliknande stödjevävnad. Denna "mikromiljö" har visats samverka med cancercellerna och bidra till cancers utveckling. Proteinet SPARC, som framförallt återfinns i stödjevävnad, har visats kunna bidra till cancerutveckling och korrelera med dålig prognos i flera cancerformer. I den andra studien i avhandlingen analyserade vi uttrycket av SPARC i bortopererade distala gallvägscancer (59 patienter). Vi fann att SPARC inte uttrycktes i normala gallvägar men uttrycktes i stödjevävnaden i distal gallvägscancer i 80% av tumörerna. SPARC uttrycktes även av stödjevävnad kring cancerspridning i bortopererade lymfkörtlar hos 68%. SPARC uttryck var inte signifikant korrelerat med överlevnad. Resultaten talar för att SPARC har en roll under cancerutvecklingen och spridningsprocessen vid distal gallvägscancer.

Nästan alla funktioner i kroppen styrs av proteiner. Vid cancerutveckling sker skador i arvsmassan. Detta leder till ändrat uttryck och funktion hos proteiner, vilket får cancercellerna att ändra sitt beteende jämfört med en frisk cell. Masspektrometri

är en metod för att studera proteinuttryck i biologiska system och kan användas för att analysera ett stort antal proteiner samtidigt. I studie III använde vi masspektrometri för att analysera proteinuttryck i operationspreparat och jämföra distala gallvägscancer och friska gallvägar. Vi identifierade 46 protein där nivåerna skiljde sig åt, flera av dessa proteiner uttrycks av bindvävsceller eller i stödjevävnad. Vi valde att studera proteinet THBS2 vidare och fann att det uttrycktes av både tumörceller och stödjevävnad i distala kolangiocarcinom men ej i normala gallvägar. Uttryck av THBS2 i tumörstödjevävnad korrelerade med högre risk för återfall. Liksom för SPARC sågs THBS2 uttryck frekvent i stödjevävnad kring cancerspridning i bortopererade lymfkörtlar. Huvudresultatet av studien var identifikationen av flertal protein utan tidigare koppling till distal gallvägscancer som kan studeras vidare. Resultaten talar även för att THBS2 har en roll vid distal gallvägscancers utveckling och korrelerar med sämre överlevnad.

THBS2 har tidigare studerats som en diagnostisk markör för bukspottkörtelcancer. Distal gallvägscancer och bukspottkörtelcancer har flera biologiska likheter. Vi valde därför att studera huruvida THBS2 i blodprover (51 patienter) kunde användas för att diagnosticera distal gallvägscancer. Vi fann att nivåerna av THBS2 var förhöjda i distal gallvägscancer liknade vid bukspottkörtelcancer. THBS2 kunde tillsammans med en redan använd markör (CA 19-9) med god precision användas för att skilja cancerpatienter från friska individer, inklusive patienter med tidig sjukdom. Kombinationen presterade sämre på att skilja godartade sjukdomar från cancer, men det såg ut att vara diagnosberoende. THBS2 har potential att diagnosticera distal gallvägscancer och bukspottkörtelcancer. Ytterligare större studier inkluderande metodoptimering, kartläggning av nivåerna vid olika godartade tillstånd och andra cancerformer krävs innan THBS2 kan användas i kliniken.

En förklaring till skillnaden mellan olika studier avseende förekomst och utfall för patienter med distal gallvägscancer är den metodik som används för att undersöka operationspreparat efter pankreatoduodenektomi. Under det sista årtiondet har en standardiserad undersökningsmetod införts gradvis i Sverige. I den sista studien i avhandlingen användes data från det nationella kvalitetsregistret för tumörer i pankreas och periampullärt. Periampullär cancer är ett samlingsbegrepp för tumörer som uppkommer kring Vaters ampull och inbegriper bukspottkörtelcancer, distal gallvägscancer, ampullär cancer samt tolvfingertarmcancer. De behandlas alla kirurgiskt med pankreatoduodenektomi. I studie V användes registerdata för att jämföra andelen av de olika tumörerna samt skillnader över tid och mellan regioner i Sverige. Vi fann att det var vanligare att patienter som ej kunde opereras diagnosticerades med bukspottkörtelcancer. Standardiseringen korrelerade med att fler tumörer bedömdes som bukspottkörtelcancer samt att andelen med mikroskopiskt icke radikal resektion och förekomst av cancerspridning till lymfkörtlar ökade. Vi fann omotiverat stora skillnader mellan olika regioner avseende bedömt tumörsprung, icke radikal resektion och lymfkörtelspridning. Överlevnaden nationellt var betydligt bättre för tolvfingertarm- och ampullär cancer

än för distal gallvägscancer och bukspottkörtelcancer. Medianöverlevnaden för distal gallvägscancer var 33 månader, vilket var bättre än i första studien. Slutligen fann vi när vi jämförde diagnosen före och efter operation att 15% av patienterna med bukspottkörtelcancer hade annan periampullärcancer diagnos före operation och 23% av patienterna med distal gallvägscancer, ampullär- eller tolvfingertarmscancer hade bukspottkörtelcancerdiagnos före operation. Studien påtalar vikten av fortsatt arbete för att harmonisera undersökningsmetodiken av pankreatoduodenektomipreparat i Sverige (och sannolikt resten av världen) samt att man bör ta hänsyn till osäkerheten i diagnostiken, framförallt om preoperativ cytostatikabehandling övervägs då preparatval skiljer sig mellan de olika cancerformerna.

Min förhoppning med avhandlingen är att bidra med några pusselbitar i det stora pussel som är förståelsen av tumörbiologin vid distal gallvägscancer. Det är även att de kliniska delarna kan användas för att förbättra dagens omhändertagande. Jag hoppas på så sätt att avhandlingen kan bidra till att hjälpa de patienter som drabbas av denna svåra sjukdom.

Abbreviations

AC	Ampullary Cancer
AJCC	American Joint Committee on Cancer
AUC	Area under the curve
BillN	Biliary intraepithelial neoplasm
BTC	Biliary tract cancer
CA 19–9	Carbohydrate antigen 19–9
CCA	Cholangiocarcinoma
CT	Computed tomography
CPH	Cox proportional hazards regression
dCCA	Distal cholangiocarcinoma
DC	Duodenal cancer
DEPs	Differentially expressed proteins
DFS	Disease-free survival
eCCA	Extrahepatic cholangiocarcinoma
ELISA	Enzyme-linked immunosorbent assay
ERCP	Endoscopic retrograde cholangiopancreatography
EUS-FNA	Endoscopic ultrasound-guided fine-needle aspiration
ECM	Extracellular matrix
FDA	Food and drug administration
FGFR	Fibroblast growth factor receptor
FISH	Fluorescence in situ hybridization
FFPE	Formalin-fixed paraffin-embedded
FOLFOX	5-fluorouracil + leucovorin and oxaliplatin
GBC	Gallbladder cancer
GEMCIS	Gemcitabine cisplatin
GEMOX	Gemcitabine oxaliplatin
GO	Gene ontology
HR	Hazard ratio

iCCA	Intrahepatic cholangiocarcinoma
IDH1	Isocitrate dehydrogenase 1
IPNB	Intraductal papillary neoplasm of the bile duct
IHC	Immunohistochemistry
KM	Kaplan-Meier
LC	Liquid chromatography
LC-MS/MS	Liquid phase separation on-line with tandem mass spectrometry
MMR	Mismatch repair
MRCP	Magnetic resonance cholangiopancreatography
MS	Mass spectrometry
NCCN	National comprehensive cancer network
NTRK	Neurotrophic receptor tyrosine kinase
OS	Overall survival
PBD	Preoperative biliary drainage
PC	Pancreatic cancer
pCCA	Perihilar cholangiocarcinoma
PET	Positron emission tomography
PRM	Parallel reaction monitoring
PSC	Primary sclerosing cholangitis
PTC	Percutaneous transhepatic cholangiography
ROC	Receiver operating characteristics
R1 resection	Non-radical resection
SPARC	Secreted protein acidic and rich in cysteine
SUS	Skane University Hospital
THBS2	Thrombospondin-2
5-FU	5-fluorouracil + leucovorin

Introduction

Anatomy and physiology

The primary role of the biliary tract (Figure 1) is the transportation, modulation and storage of bile. Bile flows from the liver to the duodenum. The biliary tract originates at the microscopic interaction surface between hepatocytes and cholangiocytes named the canals of Hering. The small bile ductules in the liver then merge gradually to interlobular, septal and segmental ducts. The segmental ducts, which drain the corresponding liver segments join to form the right and left hepatic ducts. The confluence of the right and left hepatic duct occurs at the liver hilum and the common hepatic duct is formed. The cystic duct originates from the gallbladder, where bile is stored and concentrated, then join the common hepatic duct to form the common bile duct. The common bile duct transverses the pancreatic head and join with the main pancreatic duct leading to the ampulla of Vater which, in turn, opens into the duodenum (1). In the ampullary region, a smooth muscle sphincter called the sphincter of Oddi surround the terminal common bile- and pancreatic ducts.

The main functions of bile are the excretion of waste products and the digestion of fat. Bile components include bile acids, bilirubin, phospholipids, steroids, electrolytes, endogenous and exogenous lipophilic compounds (2). Approximately 600 ml of bile is produced daily in humans (3). The green to yellow colour typical of bile is contributed to the pigment bilirubin which is a by-product from the degradation of red blood cells. Under physiological conditions, the majority of bile is stored and concentrated in the gallbladder and a coordinated release of bile into the duodenum is performed in response to food intake. The coordination is mediated by hormones such as cholecystokinin which induce gallbladder contraction and sphincter of Oddi relaxation (3).

Nomenclature

Cholangiocarcinoma (CCA) is a malignancy arising from the epithelial lining of the biliary tree. Anatomically, CCA is subtyped based on anatomical location as intrahepatic cholangiocarcinoma (iCCA) or extrahepatic cholangiocarcinoma (eCCA). ICCA arises above the second order bile ducts in the liver. ECCA is subclassified as perihilar cholangiocarcinoma (pCCA), sometimes referred to as

Klatskin tumor, which arises between the second order bile ducts and the insertion of the cystic duct or distal cholangiocarcinoma (dCCA) which arises along the common bile duct from the cystic insertion to the ampulla of Vater (Figure 1) (4). Approximately 10-20% of CCAs are iCCA, 50-60% pCCA and 20-30% dCCA (5). Biliary tract cancer (BTC) encompasses all subtypes of cholangiocarcinoma and additionally gallbladder cancer (GBC). Furthermore, cancers originating in the anatomical proximity of the ampulla of Vater are referred to as periampullary cancers and includes pancreatic cancer (PC), dCCA, ampullary cancer (AC) and duodenal cancer (DC) (6).

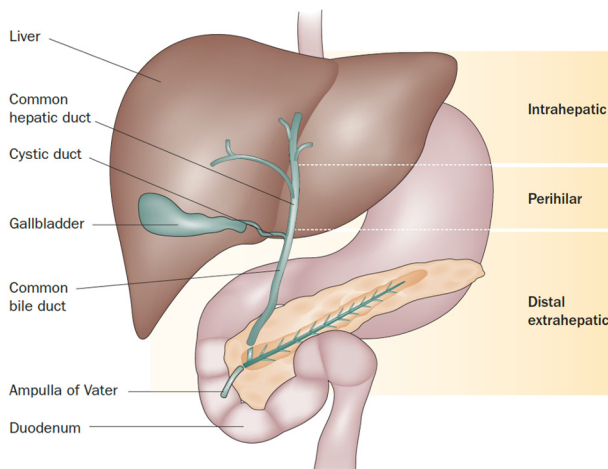


Figure 1. Anatomy of the biliary tract. Anatomical origins of cholangiocarcinoma subtypes are highlighted. Reproduced from Blechacz et al. (7) with permission from Springer Nature.

Epidemiology and risk factors

CCA accounts for approximately 3% of all gastrointestinal cancers (8) and has a slight male predominance. CCA is most commonly diagnosed in the seventh decade of life and is uncommon before the fifth decade of life (9). The incidence trends and risk factors depend on subtype and additionally, a large geographical variation is seen in the incidence of CCA. The incidence is low in western countries (0.35-2 per 100 000 person years) with substantially higher incidences (>14 per 100 000 person years) in some eastern countries (10). The incidence of iCCA has been increasing substantially in western countries during the last decades with the incidence of eCCA unchanged or slightly decreased (10). The epidemiological trends in CCA need to be interpreted with caution given problems with coding and classification of CCA subtypes. PCCA has been classified as intrahepatic in previous versions of the international classification of disease (ICD), with no separate coding for dCCA

prior to the 11th version (not yet implemented) (11). Epidemiological data for dCCA only is limited. In a national cohort from the Netherlands, the incidence of dCCA was 0.55-0.9 per 100 000 person years from 2009 through 2016 (12).

The majority of CCA cases in western countries arise in the absence of identifiable risk factors. However, patients with primary sclerosing cholangitis (PSC) have a substantially elevated lifetime risk of CCA development and the presence of benign biliary strictures which presents a diagnostic challenge (13). The highest incidence rate of CCA in the world (84 per 100 000 person-years) is seen in regions of Thailand where the liver flukes *Opisthorchis viverrini* and *Clonorchis sinensis* are endemic (5). Other risk factors associated with CCA include biliary diseases (bile duct cysts, hepatolithiasis, cholecystolithiasis, choledocholithiasis), liver diseases (cirrhosis, hepatitis, hemochromatosis), digestive diseases (inflammatory bowel disease, chronic pancreatitis, peptic ulcers), metabolic disorders (diabetes mellitus type 2, obesity, non-alcoholic fatty liver disease/steatohepatitis), lifestyle/environmental exposure (smoking, alcohol consumption, thorotrast, asbestos, 1,2-dichloropropane) as well as some genetic polymorphisms (Table 1) (11).

Pathobiology

Pathological classification

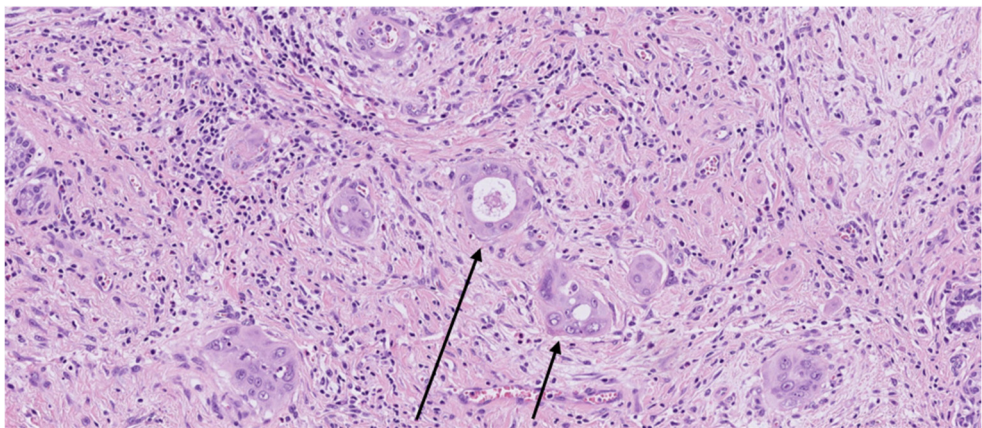
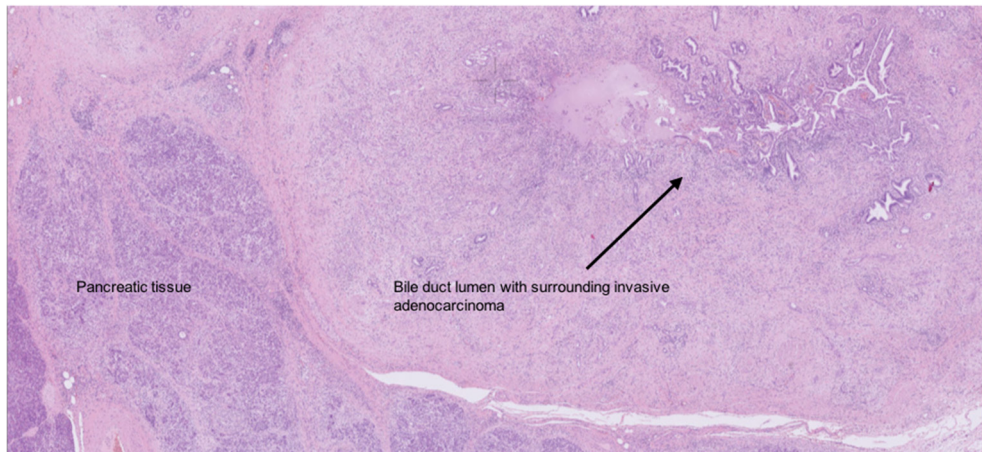
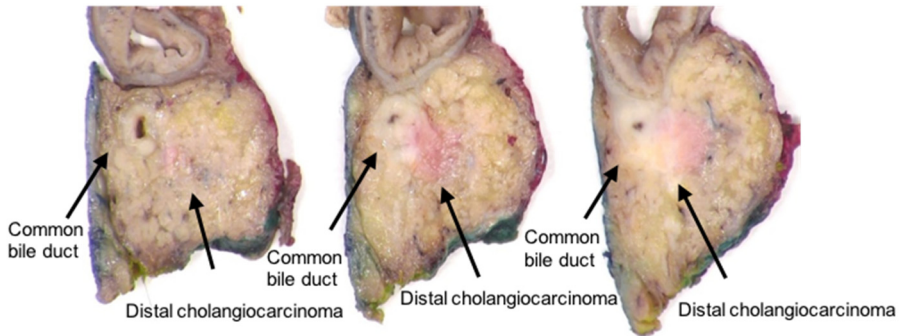
DCCA typically presents as a nodular sclerosing or flat lesion, or less common as an intraductal growing papillary tumor, tubular lesion or superficial spreading tumor (14). The absolute majority are conventional adenocarcinomas (Figure 2). Rare histological variants including squamous or adenosquamous carcinoma, mucinous/signet ring, clear cell, undifferentiated and lymphoepithelial can also occur (14). Periapillary adenocarcinomas can present an intestinal or pancreaticobiliary phenotype, a distinction with prognostic/predictive importance which is done primarily in AC (15, 16). The majority of dCCAs have a pancreatobiliary phenotype. However, approximately 10% have an intestinal phenotype (17). The nodular sclerosing subtype of dCCA is derived from the precursor lesion biliary intraepithelial neoplasia (BilIN), and the rare papillary subtype from intraductal papillary neoplasm of the bile duct (IPNB) (18). BilIN is classified according to the world health organization (19) as BilIN-1 (low-grade dysplasia), BilIN-2 (intermediate grade dysplasia) and BilIN-3 (high-grade dysplasia) (19). The gradual development from low to high grade dysplasia and eventually invasive cancer is believed to represent sequentially acquired genetic alterations in analogy to the well characterized dysplasia-carcinoma sequence in

PC (20). The specific sequence of genetic alterations during the development from BiIN to invasive dCCA is poorly characterized (21).

Table 1. Risk factors for intra- and extrahepatic cholangiocarcinoma. Reproduced from Kahn et al. (11) with permission from John Wiley and Sons.

Risk factor	Strenght of the association in iCCA	Strenght of the association in eCCA
Bile duct cysts	++++	++++
Caroli's disease	++++	++++
PSC/Cholangitis	++++a	++++a
Hepatolithiasis	+++	No association
Cholelithiasis/ choledocholithiasis	++/+++	++++
Cirrhosis	+++ /++++	++/+++
HBV	++/+++	+
HCV	++/+++	+/++
Hemochromatosis ++	++	No association
Wilson's disease	No association	No association
IBD	++	+/++
Chronic pancreatitis	++	+++
Duodenal/gastric ulcer	+	+
Opisthorchis viverrini	+++a	+++a
Clonorchis sinensis	+++a	+++a
Diabetes type II	+	+
Obesity	+a	+a
NAFLD/NASH	+++	++
Alcohol	++	No association
Cigarette smoking	+	+
Thorotrast	++++a	++++a
1,2-dichloropropane	++++a	++++a
Asbestos +++ +/++	+++	+/++

+, weak/modest association (OR: 1-1.7); ++, moderate association (OR:1.7-3); +++, strong association (OR: 3-8); +++++, very strong association (OR > 8). a, Available studies did not distinguish between iCCA and eCCA. Abbreviations: ECCA; extrahepatic cholangiocarcinoma. HBV; hepatitis B virus. HCV; hepatitis C virus. IBD; inflammatory bowel disease. ICCA; intrahepatic cholangiocarcinoma. NASH; non-alcoholic steatohepatitis. NAFLD; non-alcoholic fatty liver disease. PSC; primary sclerosing cholangitis.



Nests of cancer cells surrounded by desmoplastic stroma

Figure 2. Macroscopic picture of a distal cholangiocarcinoma from serial axial sections through the pancreatic head and microscopic pictures from the same patient showing a poorly differentiated adenocarcinoma originating from the common bile duct with an ample desmoplastic reaction. Image courtesy of Dr Agata Sasor.

Molecular characterization

Sequencing studies have elucidated the mutational panorama of CCA. Initial sequencing studies analysed mixed or iCCA cohorts. Clear differences in the mutational spectrum of CCA subtypes were identified (22, 23). The most commonly mutated genes identified in eCCA are KRAS, TP53, ARID1A and SMAD4 (24). Mutations in potentially druggable targets such as EGFR/HER2, PIK3CA, BRCA, ELF3, NTRK, BRAF and mismatch repair (MMR) proteins have been identified in approximately 25% of eCCA patients (24, 25). Few studies have presented genomic data for the dCCA subtype. In 42 dCCA patients subjected to whole exome sequencing the most frequent mutations were TP53, KRAS, SMAD4, and CDKN2A (17). In another study using targeted sequencing of 30 dCCA patients; TP53, KRAS, CDKN21, SMARCA4 and SMAD4 were the most frequently mutated genes (26). Clustering analysis of transcriptomic data has identified 4 molecular subclasses of eCCA. These subclasses serve as a framework for understanding the molecular pathogenesis and also helps identify potential therapeutic targets. The proliferation class is characterized by enrichment of MYC signaling, HER2 overexpression/amplification and mTOR pathway activation. The immune class is characterized by immune infiltrates and overexpression of immune checkpoint molecules. Further, the mesenchymal class is characterized by expression of epithelial-mesenchymal transition markers and TGF- β signaling. Finally, the metabolic class is characterized by overexpression of genes involved in bile- and fatty acid signaling and show hepatocyte features (24). In dCCA, the proliferation class was the most frequent (58%), followed by the immune type (12%), the mesenchymal type (11%) and the metabolic type (4%) (Figure 3).

Proliferation (58%)	Immune (12%)	Mesenchymal (11%)	Metabolic (4%)
HER2 overexpression	Lymphocyte infiltration	Epithelial-mesenchymal transition	Bile acid and fatty acid metabolism
Myc targets, cell cycle signaling and DNA repair	PD1/PDL1 expression	Desmoplastic stroma	Hepatocyte phenotype
Ki-67 high	Interferon signaling	Hedgehog, TNF- α , TGF- β	HDAC6, HNF4A

Figure 3. Molecular features of eCCA subclasses adopted from Montal et al. (24). The frequency of subclasses in dCCA is presented.

Tumor microenvironment

During recent decades, the major importance of the interplay between cancerous cells and components of the tumor microenvironment such as fibroblasts, endothelial cells and immune cell populations have been elucidated (27). CCA is characterized by an extensive stromal reaction, often with extensive fibrosis which has been shown to play an important role during CCA carcinogenesis (28). Stromal components in the tumor microenvironment has been shown to contribute to CCA proliferation, dissemination and development of treatment resistance (29, 30). One study found 58% of dCCA samples to have a tumor-stroma percentage (percentage stroma/tumor cells) above 70% with a higher stroma proportion correlated with worse survival (31).

Diagnosis and prognosis

Clinical presentation and diagnostic workup

The most common presentation of dCCA is obstructive jaundice with or without cholangitis. Other more unspecific symptoms typically indicative of advanced disease are abdominal pain, weight loss, malaise, night sweats, loss of appetite and fatigue (32). Routine chemistry can show elevated liver enzymes with a cholestatic pattern. Initial workup in the case of jaundice include ultrasound and in the absence of a benign explanation for obstruction (gallstone disease) a computed tomography (CT) scan is performed. The CT scan can characterize the tumor and surrounding structures, and additionally identify distant metastases. A dominant biliary stricture without associated tumor is the most common finding. Some patients present with a pancreatic head tumor (7). Further, non-invasive characterization of the biliary tree in the case of unclear diagnosis or surveillance of patients with PSC can be performed using magnetic resonance cholangiopancreatography (MRCP) (33). Positron emission tomography (PET) is not routinely used due to subpar diagnostic performance for the primary tumor but can be considered for improved staging in cases of indeterminate lymph node involvement or suspicion of other metastatic locations (34).

The majority of dCCA patients require alleviation of biliary obstruction with preoperative biliary drainage (PBD) prior to surgery. Notably, PBD compared to upfront surgery is associated with increased risk of complications (35). However, waiting times, very high bilirubin levels and cholangitis (36) are reasons to perform PBD prior to surgery and in clinical routine it's performed in most patients. Endoscopic retrograde cholangiopancreatography (ERCP) is the preferred method with percutaneous transhepatic cholangiography (PTC) as a secondary option (37). During intervention, brush cytology can be obtained to confirm malignancy.

Cytology suffers from limited sensitivity (38), although it has been substantially improved by the use of complementary fluorescence in situ hybridization (FISH). FISH detects specific chromosomal aberrations associated with malignancy (39). In strictures with suspected malignancy and negative cytology/FISH, alternatives such as cholangioscopy with targeted biopsies using the Spyglass system (40) or endoscopic ultrasound-guided fine-needle aspiration (EUS-FNA) (41, 42) are available. Notably, in patients with a clear suspicion of malignancy such as a solid pancreatic head mass which is upfront resectable, biopsy confirmation is not required before proceeding to surgery (43). In patients with unresectable/metastatic disease, percutaneous biopsies from the primary tumor or available metastases are options to histopathologically verify the cancer diagnosis prior to palliative therapy.

Biomarkers

The National Institutes of Health has defined a biomarker as “*a characteristic that is objectively measured and evaluated as an indicator of normal biologic processes, pathogenic processes, or pharmacologic response to a therapeutic intervention*” (44). A biomarker can be diagnostic (used to detect a condition), prognostic (used to indicate the likelihood of an event) or predictive (used to predict response to a treatment). Serum Carbohydrate 19–9 (CA 19–9) is the primary diagnostic biomarker used for CCA and PC. Obstructive jaundice regardless of diagnosis can cause CA 19–9 elevation and 5-10% in a Caucasian population do not genetically express CA 19–9 (45, 46) which limit utility. The diagnostic sensitivity and specificity are approximately 72% and 84% respectively in CCA (47). The low diagnostic performance, in particularly at detecting early-stage disease means CA 19–9 isn’t suitable for screening purposes and the primary use is in monitoring disease progression (48). The colorectal marker carcinoembryonic antigen (CEA) and ovarian cancer marker carbohydrate antigen 125 has lower diagnostic performance than CA 19–9 for CCA diagnosis and monitoring and are infrequently used clinically (49).

Large efforts have been directed towards novel diagnostic biomarkers and prognostic biomarkers for CCA, but none have reached the clinic yet (5). Most evaluated diagnostic candidates have been blood or bile markers. Cell-free DNA, circulating tumor cells, microRNA, extracellular vesicles, proteomic and metabolomic biomarkers/biomarker panels have shown promise (50). Selected potential diagnostic protein biomarkers include MMP-7 (51), osteopontin (52), interleukin-6 (53) and CYFRA (21-1) (54). A meta-analysis of prognostic biomarkers evaluated by immunohistochemistry (IHC) in patients with resected CCA included 4126 patients and found expression of fascin, EGFR, MUC1, MUC4 and p27 to associate with prognosis (55).

Staging and survival

As with most cancers the most important prognostic factor is the stage. The TNM system describes the extent of disease of the primary tumor (T), regional lymph nodes (N) and distant metastasis (M) which are then combined to a prognostic stage grouping which also dictate treatment alternatives. With the introduction of the 7th version of the American Joint Committee on Cancer (AJCC) staging system in 2010, the first separate TNM staging criteria for dCCA was introduced. In 2018 the 8th edition of the AJCC staging manual was introduced which changed the T stages to depth of invasion rather than invasion of specific bile duct structures as well as an expanded N stage category (Table 2). Regional lymph node stations for dCCA include nodes located at the superior/splenic artery (station 11), anterior and posterior pancreaticoduodenal nodes (station 17 and 13), nodes in the hepatoduodenal ligament (station 12), upper mesenteric nodes (station 14), common hepatic artery (station 8) and coeliac nodes (station 9) (56). Common locations for distant metastasis are liver, non-regional lymph nodes, peritoneum and lungs (57).

The lack of early symptoms and aggressive tumor biology translate to a majority of patients with CCA presenting at an advanced disease stage. Approximately one third of patients are considered for surgical treatment which is the only treatment with curative potential (58). Locally advanced disease (T4) or spread beyond regional lymph nodes disqualify surgical treatment of dCCA.

The survival for patients with locally advanced or metastatic dCCA at presentation is dismal. In a national cohort from the Netherlands the median survival for locally advanced unresectable dCCA was 6.7 months with a 5-year survival rate of 3% and for metastatic dCCA 3.6 months with a 5-year survival rate of 0% (12). The rate of recurrence after surgery with curative intent for dCCA is high with an overall median postoperative survival of 33 months and a 5-year survival rate ranging from 13% to 54% according to a meta-analysis of 3258 dCCA patients from 2017 (59). There is a substantial heterogeneity in the reported survival after resection for dCCA in the literature. The majority of studies have historically been from Asian high-volume centers. Postoperative median survival in selected larger western cohorts have ranged from 18 to 40 months (12, 60-65).

Table 2. The TNM staging system for dCCA according to the American Joint Committee on Cancer (AJCC).

	AJCC 7th edition	AJCC 8th edition
Primary tumor (T)		
TX	Primary tumor cannot be assessed	Primary tumor cannot be assessed
T0	No evidence of primary tumor	No evidence of primary tumor
Tis	Carcinoma in situ	Carcinoma in situ
T1	Tumor confined to the bile duct histologically	Tumor invades the bile duct wall with a depth less than 5 mm
T2	Tumor invades beyond the wall of the bile duct	Tumor invades the bile duct wall with a depth of 5-12 mm
T3	Tumor invades the gallbladder, pancreas, duodenum, or other adjacent organs without involvement of the celiac axis, or the superior mesenteric artery	Tumor invades the bile duct wall with a depth greater than 12 mm
T4	Tumor involves the celiac axis, or the superior mesenteric artery	Tumor invades the celiac axis, superior mesenteric artery or common hepatic artery
Regional lymph nodes (N)		
NX	Regional lymph nodes cannot be assessed	Regional lymph nodes cannot be assessed
N0	No regional lymph node metastasis	No regional lymph node metastasis
N1	Regional lymph node metastasis	Metastasis in one to three regional lymph nodes
N2	Not applicable	Metastasis in four or more regional lymph nodes
Distant metastasis (M)		
M0	No distant metastasis	No distant metastasis
M1	Distant metastasis	Distant metastasis
Stage grouping		
	0: TisN0M0	0: TisN0M0
	IA: T1N0M0	I: T1N0M0
	IB: T2N0M0	IIA: T1N1M0 or T2N0M0
	IIA: T3N0M0	IIB: T2N1M0 or T3,N0-1, M0
	IIB: T1-3, N1M0	IIIA: T1-3, N2M0
	III: T4, any N, M0	IIIB: T4, N0-2, M0
	IV: Any T, any N, M1	IV: IV: Any T, any N, M1

Clinical prognostic factors

The overall most influential prognostic factors for patients that undergo resection for dCCA are N stage and presence of non-radical resection (R1 resection) (10). Additional routine histopathological variables associated with prognosis include T stage, histologic grade (well-differentiated, moderately differentiated, poorly differentiated), presence of perineural invasion, lymphatic invasion, venous invasion, pancreatic and peripancreatic fat invasion (59). Preoperative levels of biomarkers CA 19–9 and CEA and postoperative non-normalization has been associated with prognosis (66). A prognostic nomogram based on the preoperative

variable's neutrophil to lymphocyte ratio, peak total bilirubin and major vascular resection has been developed to predict early recurrence (67). Other selected clinical variables that's been associated with prognosis in resected dCCA patients include PBD (68), postoperative complications (69) and blood transfusions (68, 70).

Impact of histopathological evaluation

There is a large discrepancy in the literature regarding the relative frequency of perampullary cancers, R1 resection rate and nodal involvement. The methodology of histopathological evaluation has been shown to influence these assessments (71, 72). Differences in histopathological evaluation could contribute to the heterogenous study results with regard to frequency of dCCA, survival estimates and prognostic factors. The use of standardized histopathological methodology has been shown to increase the rate of non-pancreatic tumor origin, R1 resection and detected lymph node metastases substantially (73). Two main approaches have been developed; axial sectioning as described by Verbeke et al. (74) and bivalving as described by Adsay et al. (75). Both techniques give comparable outcomes (76). In Sweden a standardized protocol based on axial sectioning has been implemented during the 2010s (56). Given that different adjuvant and palliative oncological treatments are used for the individual perampullary cancers, correct identification of tumor origin is of outmost importance.

Treatment

Surgery

For patients with dCCA and other perampullary cancers, the surgical procedure of choice entails a pancreatoduodenectomy, also called the Whipple procedure (77). During the procedure the gallbladder, common bile duct, pancreatic head, duodenum and the distal stomach is removed *en bloc*. The procedure can also be done as a pylorus-preserving resection, which show similar outcome (78). Reconstruction is done with a hepaticojejunostomy, gastroenterostomy and a pancreaticogastrostomy or pancreaticojejunostomy. Some reconstructions also include an entero-entero anastomosis. Venous resection and reconstruction can be performed at high volume centers without increased mortality or morbidity (79). Arterial resection is associated with a high mortality and morbidity but considered in highly selected patients (80). Pancreatoduodenectomy is one of the most extensive surgical procedures used in modern medicine and has historically been associated with a substantial postoperative mortality. Advances in surgical technique and perioperative care have improved the results. The 30-day and 90-day

postoperative mortality after pancreatoduodenectomy in Sweden was 1.5% and 3.5% respectively during the 2010s (81). The postoperative morbidity is still substantial with complications grade 3 or higher according to the Clavien-Dindo classification (82) occurring in approximately 15% of patients (81). In addition to common post-operative complications such as wound infection, thrombosis etc, procedure specific complication include leakage from the pancreatic anastomosis (postoperative pancreatic fistula) (83), delayed gastric emptying (84) and postpancreatectomy hemorrhage (85). DCCA compared to PC has been associated with a higher frequency of pancreatic fistula, which in turn has been associated with worse survival (86). This is presumably due to a higher frequency of soft pancreatic tissue in dCCA compared with PC (87, 88).

Palliative oncological treatment

For the majority of patients with unresectable/metastatic disease palliative chemotherapy is the only treatment option. During the 1990s and 2000s several, mostly smaller, non-randomized trials evaluated the role of chemotherapy in advanced BTC. The primary compounds with evidence of efficacy were the antimetabolites gemcitabine and 5-fluorouracil/leucovorin (5-FU) (leucovorin is a reduced folate which enhances 5-fluorouracil effectiveness) as well as the platinum compounds cisplatin and oxaliplatin (89). In 2010 the randomized ABC-02 trial, which compared first-line gemcitabine and cisplatin (GEMCIS) with gemcitabine singlet established a new standard of care. Treatment with the combination chemotherapy resulted in a significantly improved overall survival (OS) (11.7 vs 8.1 months) (90). The treatment benefit was similar across anatomical subtypes. The toxicity profile of cisplatin (oto/neurotoxicity, high emetogenicity, renal toxicity) has led many centers to substitute cisplatin for the less toxic platinum compound oxaliplatin. No randomized comparisons of GEMCIS and gemcitabine+oxaliplatin (GEMOX) regimens exist. A meta-analysis found a slight survival benefit for GEMCIS (11.7 vs 9.7 months) at the expense of higher toxicity (91). A randomized trial comparing GEMCIS and GEMOX in first line advanced GBC showed similar outcomes, with the numerical survival slightly favoring GEMOX (9 vs 8.3 months) (92). In Sweden the GEMOX regimen is generally preferred and real-world data have shown similar survival to comparable clinical trials (93). A minority of patients receive second line treatment (94), the ABC-06 trial was the first randomized study in the second line setting and compared 5-FU and oxaliplatin (FOLFOX) versus active symptom control after progression on first line GEMCIS treatment. The FOLFOX arm had a modestly improved survival compared to active symptom control (6.2 vs 5.3 months) (95). Slightly lower treatment benefit of FOLFOX was observed in the eCCA patients compared with other locations. Radiotherapy is rarely used for advanced BTC in Sweden but can be considered for symptomatic relief of painful lesions and in highly selected patients for treatment of localized disease.

The poor survival with current treatments options and identification of potentially targetable molecular alterations have led to a multitude of targeted therapies and immunotherapies investigated for the treatment of CCA (96). The most encouraging results have been found with fibroblast growth factor receptor (FGFR) inhibitors in FGFR-2 gene fusion CCA and isocitrate dehydrogenase-1 (IDH1) inhibitors in IDH1 mutant CCA (97). However, these mutations occur almost exclusively in iCCA. No targeted therapy for dCCA is currently used in clinical routine in Sweden. Notable emerging therapies include the immune checkpoint inhibitor pembrolizumab which has been Food and Drug Administration (FDA) approved for solid tumors with palliative treatment intention and MMR deficiency or microsatellite instability. The approval is based on results from basket trials including BTC patients (98, 99). MMR deficiency occurs in approximately 2% of all BTCs. Tumors with neurotrophic receptor tyrosine kinase (NTRK) fusions have shown a high response rate to NTRK inhibitors in basket trials (100-102), getting FDA and European Medicines Agency approval for treatment agnostic of tumor origin in patients without alternative treatment options. NTRK fusions are present in <1% of BTC patients (103). HER2 overexpression/amplification is present in 17% of eCCA, a phase II trial has shown promising results with anti-HER2 antibodies trastuzumab and pertuzumab in pre-treated patients (104) with randomized data eagerly awaited.

Adjuvant oncological treatment

Because of the high rate of recurrence after surgery with curative intent, several adjuvant strategies of chemotherapy or chemoradiotherapy have been evaluated. Until the 2010s, mostly small non-randomized or retrospective studies had been performed which meant adjuvant treatment were given inconsistently and with various regimes in different centres (105). The European Study Group for Pancreatic Cancer (ESPAC)-3 trial enrolled 428 patients with resected periampullary cancers, 96 of which had dCCA. Patients were allocated to 3 arms: observation, 5-FU and gemcitabine. The study did not find differences in the primary outcome of OS, however, a secondary multivariable analysis adjusting for prognostic factors revealed a survival benefit of adjuvant chemotherapy (hazard ratio (HR)= 0.75) with a favourable safety profile of gemcitabine compared to 5-FU (106). The study was not powered for subgroup analysis. Notably, among dCCA patients the survival was not numerically improved with a median survival in the observation, 5-FU and gemcitabine arms of 27.2, 18.3 and 19.5 months respectively. The BCAT trial randomized patients after resection for eCCA to gemcitabine or observation, no significant difference in RFS or OS were seen (107). The PRODIGE-12/ACCORD-18 trial randomized patients after resection to GEMOX or observation, with no significant difference in RFS or OS observed (108). The BILCAP trial was published in 2019 and although somewhat controversial, established a new standard of care for adjuvant treatment in BTC. BTC patients were randomized after

resection to capecitabine (an oral prodrug to 5-FU) for 6 months or observation. The study did not meet the primary endpoint of improved OS in the intention to treat population (numerical survival 52 vs 36 months favouring capecitabine). A pre-specified sensitivity analysis of the intention to treat population with adjustment for prognostic factors revealed a significant benefit from capecitabine treatment (HR=0.71) (109). Based on the results of the BILCAP trial capecitabine is used for adjuvant treatment of BTC in Sweden. Based on the national comprehensive cancer network (NCCN) guidelines, gemcitabine can be offered to patients for whom capecitabine is not an option (110). Chemoradiotherapy in the adjuvant setting is generally not used in Sweden. Based on non-randomized phase II data, American society of clinical oncology and NCCN guidelines suggest it can be considered in patients with non-radical resection or high-risk features (110-112). Neoadjuvant treatment for dCCA is not used outside of clinical trials (113).

Biomarker investigation

Applications of mass spectrometry in cholangiocarcinoma

Proteomics is the large-scale study of proteins involved in a biological condition. Proteins are the functional unit of the cell and often suitable as biomarkers. Mass spectrometry (MS) is the most frequently employed technique for proteomics (114). The basic principle of MS is the ionization of compounds and subsequent separation based on the mass to charge ratio of the generated ions and finally ion detection. Several different MS technologies for analysis of biological compounds have been developed. Commonly used methods such as 2-dimensional gel electrophoresis followed by MS has downsides such as limited reproducibility and difficulty to upscale (115). A rapid technological development during the last decades has enabled the simultaneous identification and quantification of proteomes in complex biological systems such as tissues (116). In particular, the development of bottom-up or “shotgun” proteomics has shown promise for biomarker discovery (117). Several previous MS based proteomics studies of resected CCA specimens have been performed using different analytical approaches (118-131). The majority of these have investigated iCCA or a mixed CCA/BTC cohorts. The first proteomic characterization of an eCCA cohort was done by Maeda et al. (120). They used a bottom-up mass spectrometry workflow to analyse resected formalin-fixed paraffin-embedded (FFPE) specimens and performed sequential targeted verification and IHC validation. The discovery experiment identified 136 overexpressed proteins, 10 of which could be validated as upregulated in eCCA. Takenami et al. used bottom-up proteomics followed by IHC validation to identify novel biomarkers to differentiate dCCA and PC. Eighteen differentially expressed proteins (DEPs) were identified, 5 proteins were validated by IHC as potential novel biomarkers (132).

Secreted protein acidic and rich in cysteine

Secreted protein acidic and rich in cysteine (SPARC) is a 32 kDa matricellular (non-structural) protein component of the extracellular matrix (ECM). SPARC is composed of three structural domains, the acidic N-terminal which has low calcium affinity, the cystine rich follastin-like domain and the C-terminal calcium binding domain (133). SPARC has a diverse functionality and impacts several biological functions such as ECM-remodelling, cell-ECM interaction, cell differentiation, angiogenesis, bone remodelling and wound healing (134-139). SPARC has been studied in various cancer models and a functional role in tumor associated stroma remodelling, tumor growth, migration and apoptosis has been found. Additionally, SPARC can contribute to development of chemotherapy resistance (133). The function of SPARC in cancer is seemingly context dependent and both pro- and antitumoral effects have been identified (140). SPARC has been proposed as a treatment predictive biomarker for treatment with gemcitabine (141) and nab-paclitaxel (142) where contradictory results exist (143). In two cohorts of patients treated with resection for BTC, stromal SPARC expression was associated with poor survival, with the negative prognostic effect enhanced in gemcitabine treated patients suggesting treatment resistance (144, 145).

Thrombospondin-2

Thrombospondin-2 (THBS2) has a mass of 150 kDa and is composed of an N-terminal domain, a disulphide link, a procollagen domain, types I-III repeats and a C terminus. Structurally THBS2 form homotrimers (146). Like SPARC, THBS2 is a matricellular protein with an important role in ECM remodelling. THBS2 is required for normal fibroblast function and can induces dysfunctional activation of matrix metalloproteinases (147). The ECM modulating function of THBS2 has a potent anti-angiogenic role during tissue homeostasis (148). The function of THBS2 during carcinogenesis seems dependent on its cellular context. In a PC model, cancer cell expression of THBS2 inhibited invasiveness through downregulation of matrix metalloproteinase-9 and urokinase-type plasminogen activator (149). In a coculture model, THBS2 expression in stromal pancreatic stellate cells was found to promote invasiveness (150). Kim et al. identified THBS2 as a secreted protein from a PC precursor lesion model using MS. It was further validated in several cohorts and proposed as a novel biomarker for the early diagnosis of PC (151).

Aims of the thesis

The general aim of this thesis was to investigate strategies for improving the management of patients with dCCA. This included the evaluation of clinical outcomes, diagnostic process as well as identification of novel protein biomarkers.

The specific aims of each paper are listed below

- I. To investigate clinical outcome and clinicopathological prognostic factors for patients treated with pancreatoduodenectomy at Skane University Hospital from 2008 through 2015
- II. To examine the expression of SPARC in resected dCCA including primary tumors and paired lymph node metastases and relationship with survival.
- III. To identify novel tissue biomarkers for dCCA using discovery mass spectrometry and targeted verification
- IV. To evaluate the expression of serum THBS2 in dCCA, PC, patients with benign diseases and healthy individuals and evaluate the diagnostic and prognostic biomarker potential.
- V. To investigate national trends in the frequency of periampullary tumor origin, survival, histopathological evaluation and diagnostic accuracy.

Methodological considerations

The detailed methods are presented in the original papers. The methods are briefly presented below.

Patient cohorts

This thesis includes three different patient cohorts. The first cohort was identified from institutional records and included all patients treated with pancreatoduodenectomy for dCCA at Skane University Hospital (SUS) between 2000 and 2015 (N=64), 59 of which had (FFPE) tissues available. A histopathological re-evaluation of available material was performed in order to ensure a standardized assessment, staging was done according to AJCC 7th edition. Demographical, clinical and follow up was collected retrospectively from patient charts. For paper I, the time period was restricted to 2008-2015 (N=54) in order to evaluate a modern time span during which SUS served as a tertiary referral centre (152). Resected tumor specimens from the full cohort (N=59) were used for molecular analysis in papers II and III.

A prospective biobank of patients scheduled for pancreatic resection at SUS was started in 2012. Preoperative serum samples were obtained from patients that underwent pancreatoduodenectomy, in addition healthy controls were obtained from the local blood donation centre. Serum aliquots were stored at -80 °C until analysis. For paper IV, serum samples from all patients with a histopathological dCCA diagnosis treated with surgery from 2012 through 2019 were analysed (N=51). PC patients (N=52) from the cohort were identified and matched to the best extent possible in a 1:1 ratio to the N- and T-stage distribution in the dCCA samples. Available patients from the cohort with benign pancreatic diseases (N=27) were included as well as 51 age- and gender matched healthy donors.

The Swedish National Registry for Pancreatic and Periapillary Cancer was established in 2009. The registry prospectively gathers data from patients with periapillary cancers and patients that undergo pancreatic surgery irrespective of diagnosis. The registry is organized into six parts: registration, preoperative- and intraoperative data, postoperative data, histopathology, oncology and follow up. The registry has been validated with a good coverage against the Swedish Cancer

Registry (81). For paper V, all patients with periampullary tumors between 2010 and 2019 were included (N=9143).

Immunohistochemistry

IHC is widely used in biology and clinical pathology to visualize antigen expression and localization in tissues (Figure 4). Most commonly, the antigen is a protein. When applied to FFPE-tissues the first steps include mounting on a glass slide, deparaffinization and rehydration through incubation with xylene and graded ethanol solutions. Next, in order to break the methylene bridges that cross link proteins and “mask” antigens an “antigen retrieval” is performed, in this thesis a heat-induced antigen retrieval was used throughout. An antibody targeting the antigen of interest (primary antibody) is then incubated on the tissue specimen. The primary antibody can be either monoclonal, which means it targets one specific epitope of the antigen, such as the SPARC antibody used in paper II, or polyclonal, such as the THBS2 antibody used in paper III, where different epitopes of the same antigen are targeted. Benefits of monoclonal antibodies include lower background, and low lot-to-lot variation, whilst polyclonal antibodies are less sensitive to fixation and processing (153). After washing a secondary antibody targeting the primary antibody is applied. The secondary antibody is labelled, typically with a polymer-linked enzyme (paper II) or biotin, which forms an avidin-biotin-enzyme complex in an additional step (paper III). Finally, the enzyme catalyses a reaction after the addition of a chromogenic substrate producing a visible colour, the intensity of which correlate to the presence of antigen in the tissue (154).

The evaluation of IHC consists of quality assessment, including the evaluation of adequate external positive and negative controls, and any available internal controls (cell types such as immune cells or endothelial cells predicted to express or not express the antigen of interest in the stained section). A valuation of the expression includes assessment of which cell types express the antigen such as cancer cells, stromal fibroblasts, endothelial cells or immune populations. A valuation of the cellular compartment (membranous, cytoplasmic or nuclear) and whether consistent with expected antigen expression compartment should be performed. The staining is by nature semi quantitative and both the intensity and percentage can be quantified. Different scoring systems have been developed and validated. Commonly used scoring systems include the Allred, H-score and immunoreactive score, additionally various statistical methods can be used to identify the optimal cutpoint for prognostic differentiation (155). For the evaluation of SPARC, a previously used scoring system was employed for dichotomization into high and low expression (156). There was no previously described scoring system for THBS2, and in paper III the prognostic correlation was done between samples with

and without THBS2 expression rather than performing data-driven cut point optimization in the relatively small cohort.

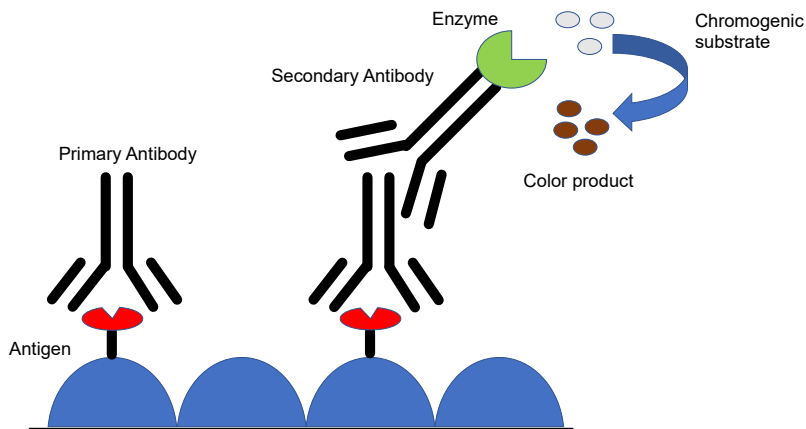


Figure 4. Schematic illustration of immunohistochemistry. A primary antibody binds to an antigen in a tissue. A secondary antibody with a conjugated enzyme then binds the primary antibody. The enzyme catalyzes a chemical reaction in which a visible precipitate is generated, the intensity of which correlates to the presence of the antigen in the sample.

Mass spectrometry

In a bottom-up proteomic workflow proteins are extracted and digested into peptides enzymatically. Peptides are then separated using liquid chromatography (LC) and injected into the MS instrument where they are ionized and separated based on their mass/charge ratio. The most intense precursors are fragmented and analyzed in a second MS run. The resulting fragmentation ions are used to perform peptide sequencing and subsequent in-silico protein prediction. Quantification can be performed either based on label free or labelled methods. Previously, MS has mostly been used to analyze fresh frozen tissues. However, protocols for the analysis of FFPE tissues have been developed (157, 158). In paper III, a “bottom-up” proteomics workflow using LC on-line with tandem mass spectrometry (LC-MS/MS) was used to compare the proteomes of resected dCCA FFPE specimens (N=20) and FFPE normal bile ducts (N=6). Specimens were macrodissected. Protein extraction, deparaffinization and protein cross-link breakage was done using heat induced antigen retrieval similar to IHC. Proteins were digested to peptides using trypsin. The peptides were separated using an LC on-line to the MS instrument

(Q Exactive Plus, Thermofisher Scientific). Protein identification and quantification was done using the software Proteome discoverer and alternative software programs (MaxQuant and OpenMS). Label free quantification was performed using a precursor ion area detector.

Limitations of bottom-up proteomics include quantitative imprecision, limited sensitivity and difficulty in validating a large number of identified DEPs (159, 160). In order to overcome this, a verification of DEPs was done using targeted MS. Most commonly used targeted proteomics methods are multiple-reaction monitoring and parallel reaction monitoring (PRM). PRM is performed using a quadrupole Orbitrap MS such as the Q Exactive Plus used for the discovery bottom-up proteomics. Specific peptides are selected for identification and the instrument is set to identify the specific precursor ions and a full MS/MS scan of precursor and fragment ions are analyzed. PRM is able to detect peptides at a lower concentration and with improved accuracy compared to discovery bottom-up proteomics (161, 162). In paper III, new material from 16 dCCA specimens and 9 controls were analyzed using PRM (the same sample preparation and MS as during the discovery study was used) in order to verify selected DEPs from the discovery study. Quantification based on precursor ion intensity was done using the Skyline software (163).

Enzyme-linked immunosorbent assay

Enzyme-linked immunosorbent assay (ELISA) is frequently used in biological and clinical research for quantitative measurement of antigens in fluids such as serum/plasma, bile, urine, cellular extracts and more. Like IHC, ELISA uses specific antibodies to bind the antigen of interest. The most commonly used version is the “sandwich” ELISA. Two antibodies targeting different epitopes of an antigen are used (Figure 5). In paper IV, a commercial sandwich ELISA (DTSP20, R&D Systems, Minneapolis, MN, USA) was used to measure THBS2 concentration in serum samples. The ELISA was selected as it was used in the majority previous studies evaluating THBS2 as a diagnostic biomarker (151, 164-166). Principally, the samples are incubated in a well plate with the first antibody (capture antibody) coated. After washing, the second antibody (detection antibody) with a conjugated enzyme is added to the wells. After additional washing a chromogen is added. The enzyme catalyses the chromogenic reaction and a change in colour takes place proportional to the presence of the antigen. A microplate reader is used to measure the density of each well at a specified wavelength. Standards of a known concentration are included, and a standard curve is calculated. For THBS2 concentration calculation a four-parameter logistic regression standard curve was calculated using MyAssays online data analysis tool.

Serum CA 19–9 concentrations were measured in a clinical laboratory using an accredited Chemiluminescent immunoassay (Ref.11776193 122 Cobas/Roche).

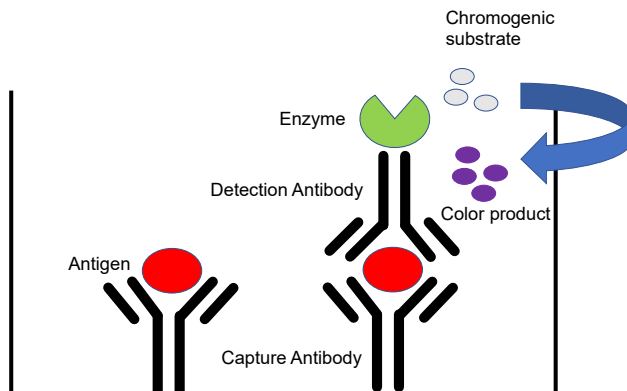


Figure 5. Schematic illustration of a sandwich ELISA. The antigen binds to the coated capture antibody. A detection antibody with a conjugated enzyme then binds a different epitope of the antigen. The enzyme catalyzes a chemical reaction in which a colored product is generated, the intensity of which correlates to the presence of the antigen in the sample.

Statistics

Survival analysis was used in all papers included in the thesis. Survival outcomes included disease-free survival (DFS; time to recurrence or death) and OS. While OS is considered the golden standard for cancer research, DFS is a commonly used surrogate endpoint that can identify statistically relevant differences at a shorter follow up time (167). The Kaplan-Meier (KM) method was used to estimate the survival of patients. The KM method uses the exact time to event or censoring (loss to follow up) and incorporates the number of patients at risk in the calculation of survival. For an adequate estimation, it is important that censoring is not related to the groups investigated. The KM survival estimate is combined with the log-rank test to compare differences between groups. The log-rank test compares a weighted average of recorded events and expected events at all time points. When interpreting KM survival curves the precision in the survival estimate is reduced when few patients at risk remain and a presentation of number at risk and confidence intervals are recommended (168). Although survival calculation is performed using all

available data, the graphical presentation of survival in Paper I could have been shorter since few patients remained past 3 years.

A regression model is a statistical way of estimating the relationship between an outcome variable and one or more predictor variables. In this thesis, the relationship between predictor variables and survival was performed using Cox proportional hazard regression (CPH) (paper I, III) and predictor variables and diagnosis using logistic regression (paper IV). The CPH model calculates the HR of predictor variables. The HR is the difference in the likelihood of the outcome event occurring when the predictor variable is present compared to not present (categorical variables) or when increased by one unit (continuous variables). With inclusion of multiple covariables (multivariable regression) the HR estimates are adjusted for one another which is used to adjust for confounding and identify variables independently associated with survival. CPH-models are built on the assumption that the hazard associated with a variable does not vary over time. Other important aspects are to avoid overfitting and collinearity (169). The selection of variables for a multivariable CPH can be performed using different methods (170). In paper I, the goal was to identify the strongest predictor for survival. Backward elimination was used which step- by step remove variables that do not contribute to model prediction. In paper III the goal was to adjust for confounders of THBS2 association with survival and variable selection of clinically relevant confounders were done manually.

The statistical analysis of MS data in paper III was done using the dedicated software Perseus (171). Important analytical steps included data filtering (only proteins quantified in 50% of samples in each group was used), normalization, imputation and group comparison with adjustment for multiple testing. Normalization was done to account for variations of cellularity in the samples. The normalization in the discovery experiment was based on median protein intensity in each sample. In the PRM verification, only pre specified proteins were quantified and normalization was done using common housekeeping proteins glyceraldehyde 3-phosphate dehydrogenase and tubulin beta chain. DEPs were identified using t-tests with a permutation based false discovery rate adjustment. In order to further describe the biological properties of identified DEPs bioinformatic analysis was performed. The Gene ontology (GO) project has standardized the description of genes and gene products. Specifically, the cellular component, molecular function and biological process are annotated (172). PANTHER database (173) was used to perform GO classification of identified DEPS and identify enrichment. Different ways of annotating signaling pathways have been developed and enrichment analyses of pathways were done using additional reference databases KEGG (174) and REACTOME (175). Protein-protein interaction patterns were analysed using the STRING database (176).

Diagnostic accuracy of biomarkers THBS2 and CA 19–9 was evaluated in paper IV. Central properties include the sensitivity (proportion of patients with a condition

correctly classified by the biomarker) and specificity (proportion of patients without the condition correctly classified by the biomarker). In clinical practice, the actual performance of a biomarker is also highly dependent on the prevalence of the condition investigated (177). Receiver operating characteristics (ROC) analysis is performed by plotting the sensitivity on the y axis and (1-specificity) on the x-axis of a two-dimensional plot. It provides a visual representation of the trade-off between sensitivity and specificity at different cutpoints which can aid in cutpoint selection. Additionally, the area under the curve (AUC) provides a summary statistic of diagnostic performance. Logistic regression models were used to generate AUCs of THBS2 and CA 19-9 individually and combined.

Ethics

The studies in this thesis were approved by the regional ethics committee at Lund University. Dnr 2015-392 (papers I-III). Dnr 2010/684, 2012/661, 2015/266 (Paper IV). Dnr 2015/392, 2021/00622 (Paper V). None of the studies included in this thesis had an impact on medical treatment of the patients. All data was anonymized and analysed on group level.

Results summary

Paper I

The first paper of this thesis evaluated outcome and prognostic factors in patients treated with pancreatoduodectomy for dCCA at SUS between 2008-2015 ((N=54). The mean age was 68 ± 7.7 years and 38% were women. The most common presenting symptoms were jaundice (72%), abdominal pain (35%) and weight loss (19%). PBD was performed in 98% of patients. Postoperative complications ≥ 3 according to the Clavien-Dindo classification occurred in 17% of patients. After histopathological re-evaluation, 72% of patients had lymph node metastasis. Sixty-three percent of patients had R1 resections. Chemotherapy was administered in 52% of patients with the most common regimen being gemcitabine monotherapy. The median time to recurrence was 13.2 months and 52% and 82% of patients had recurrence at year 1- and 3 respectively. The most common location of recurrence was the liver (31%), followed by local recurrence (24%). The median survival was 22.2 months. The OS at year 1 was 80%, year 3 (21%) and year 5 (9.2%).

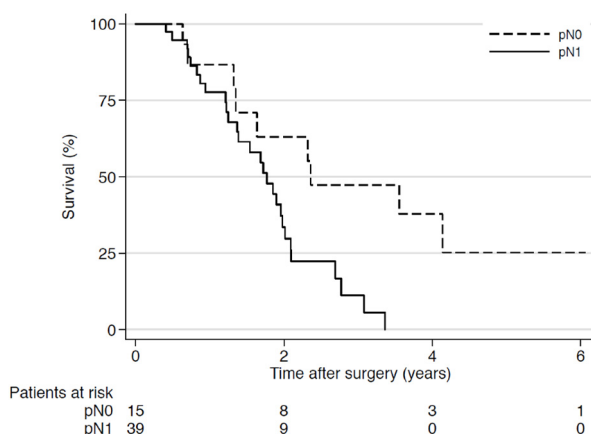


Figure 6. Overall survival after resection for distal cholangiocarcinoma stratified by lymph node status. Survival is estimated using the Kaplan-Meier method.

Using univariable CPH, the presence of lymph node metastasis (categorical) HR 2.9 ($P=0.016$) and lymph node metastases (continuous) HR 1.08 ($P=0.017$) were significantly associated with survival. A CPH model with stepwise backward selection found no other variable to be significantly associated with survival when adjusted for lymph node metastasis. Analysing number of lymph node metastases or the lymph node ratio did not improve the prediction of the model compared to presence of lymph node metastasis alone. Survival of dCCA patients stratified by lymph node status is presented in Figure 6.

Paper II

The second paper evaluated the expression pattern and prognostic significance of SPARC in patients with dCCA treated with pancreatoduodenectomy between 2000-2015 ($N=59$) using IHC. Both primary tumors and paired lymph node metastases were evaluated. No SPARC immunoreactivity was detected in normal bile ducts. Tumor cell immunoreactivity was weak and focal and thus was not quantified further. Stromal immunoreactivity intensity was absent in 20%, weak in 10%, intermediate in 39% and strong in 31% of patients. The proportion of SPARC positive stroma was absent in 20%, 11-25% in 1.7% of patients, 26-50% in 15% of patients, 51-75% in 25% of patients and $\geq 75\%$ in 37% of patients (Figure 7). An intermediate or strong staining in $>25\%$ of stromal cells was classified as high expression which entailed 68% of patients. Stromal (non-lymphoid) SPARC expression was present in 68% of lymph node metastases. When comparing paired primary tumors and paired lymph node metastases 96% and 68% ($P=0.016$) respectively had stromal SPARC expression present. High stromal SPARC expression was associated with presence of lymph node metastasis (78 vs 47%; $P=0.013$). No significant association between SPARC expression and survival was found.

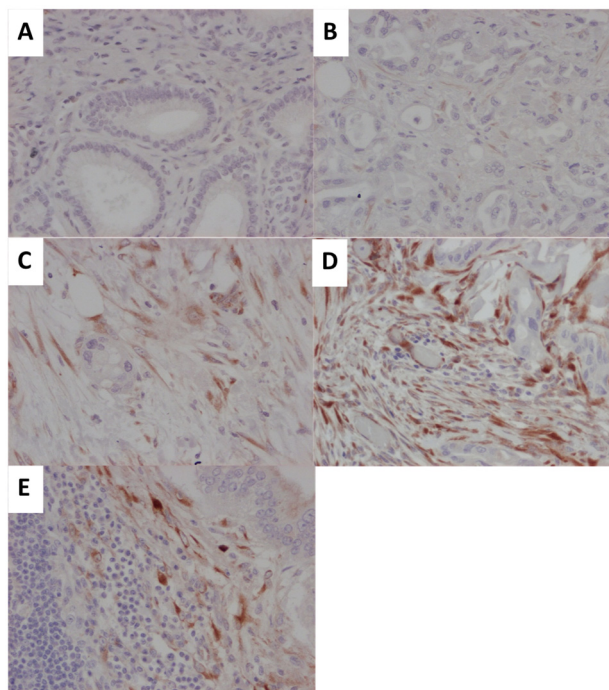


Figure 7. Representative immunohistochemical images at 200 x magnification of SPARC expression in the stromal compartment of distal cholangiocarcinoma (A), negative expression, (B) weak expression, (C) moderate expression, (D) strong expression, (E) Strong peritumoral stromal SPARC expression in a paired lymph node metastasis.

Paper III

Paper III encompasses a proteomic profiling of resected FFPE dCCA specimens and normal bile ducts using discovery bottom-up MS, followed by verification using PRM and further validation of THBS2 expression using IHC. A total of 3037 proteins were identified. Eighty-seven DEPs were identified, 31 proteins were upregulated and 56 downregulated in dCCA compared to normal bile ducts. Bioinformatic analysis of DEPs from the discovery study revealed enrichment of several extracellular components and signaling pathways such as integrin signaling, ECM-receptor interaction and collagen degradation.

In the PRM verification, 122 peptides mapping to 79 proteins could be quantified. In total, 65 peptides were found to be differentially expressed corresponding to 46 proteins. Twenty-eight proteins were upregulated and 18 downregulated. Several previously described tentative CCA biomarkers (S100-P, CECAM-6, cytokeratin 17, thymidine phosphorylase and heat-shock protein 90) were correctly identified as upregulated and several novel proteins without previous association to CCA

biology were identified. The identified peptides/ proteins and their previous association with CCA/BTC are presented in Table 3.

IHC was used to further investigate the expression of THBS2 in dCCA, paired lymph node metastases and normal bile ducts. THBS2 was predominantly negative in normal biliary epithelium but weak focal expression was detected. THBS2 immunoreactivity was seen in dCCA cancer cells with a membranous/cytoplasmic staining pattern and in the stroma. Epithelial immunoreactivity was absent in 10%, 1+ in 51%, 2+ in 32% and 3+ in 7%. The stromal intensity was absent in 8%, 1+ in 31%, 2+ in 42% and 3+ in 11% (Figure 8). When comparing primary tumors and paired lymph node metastases, epithelial THBS2 expression was present in 96% and 54% ($P=0.001$) respectively. Stromal THBS2 expression was present in 96% and 72% ($P=0.031$) respectively. Epithelial THBS2 expression was not associated with survival. Stromal THBS2 was not significantly associated with DFS ($P=0.105$) or OS ($P=0.079$) using KM analysis. After adjustment for confounders in a multivariable CPH, stromal THBS2 was significantly associated with poor DFS (HR 3.95, $P = 0.037$) and a trend was seen for OS (HR 3.34, $P = 0.062$).

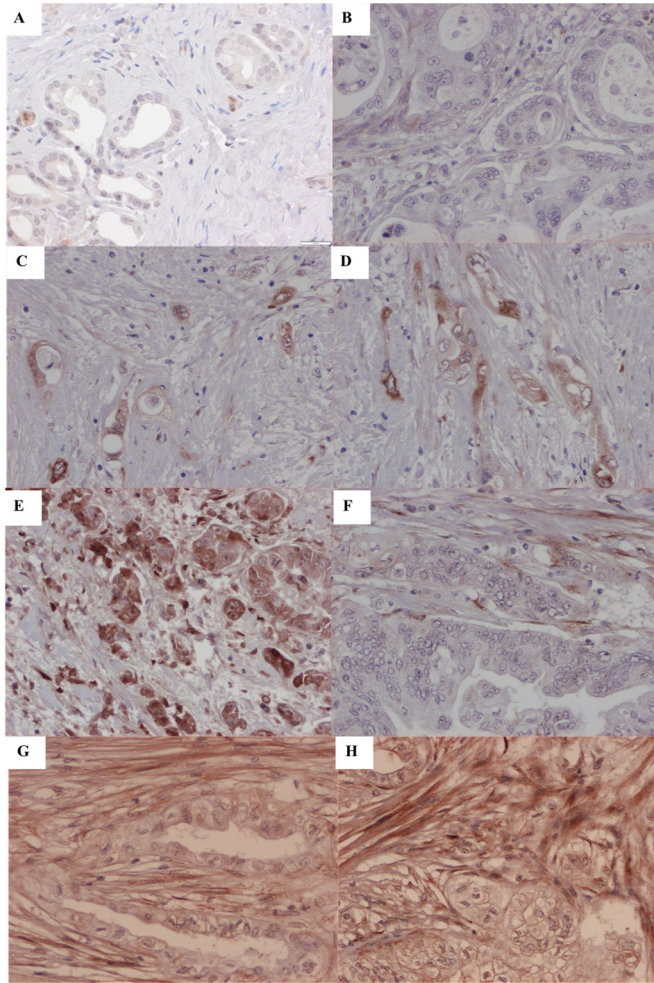


Figure 8. Representative immunohistochemical images of THBS2 expression at at 200 x magnification. Normal bile duct; (A) no expression. Distal cholangiocarcinoma; (B) no expression, (C) weak epithelial expression, (D) moderate epithelial expression, (E) strong epithelial expression, (F) weak stromal expression, (G) moderate stromal expression and (H) strong stromal expression.

Table 3. Proteins found to be differentially expressed using parallel reaction monitoring ($q < 0.05$, $FC \geq 2$ or ≤ 0.5) between distal cholangiocarcinoma samples and normal bile ducts. Proteins are ordered by descending FC. Abbreviations: CA; cancer. CCA; cholangiocarcinoma. CO; control. FC; fold change.

#	Accession	Gene	Protein Name	Peptide sequence	Valid Cancer	Valid Control	q-value	FC Ca/Co	Literature CCA
1	P35442	TSP2	Thrombospondin-2	LVFNPDQ EDLDG GR	16	9	< 0.0001	29.3	
2	P35442	TSP2	Thrombospondin-2	FDYIPPV NADDLSK	16	9	< 0.0001	8.3	
3	Q6UX06	OLFM4	Olfactomedin-4	LLEYR	16	8	< 0.0001	7.8	(120, 178)
4	P25815	S100P	Protein S100-P	YSGSEG STQTLTK	16	9	< 0.0001	7.3	(179, 180)
5	P40199	CEAM6	Carcinoembryonic antigen-related cell adhesion molecule 6	IGYSWYK	16	9	0.0007	7.2	181
6	P25815	S100P	Protein S100-P	ELPGFLQ SGK	16	8	< 0.0001	6.4	(179, 180)
7	Q04695	K1C17	Keratin, type I cytoskeletal 17	ASLEGNL AETENR	16	9	0.0044	5.5	(182, 183)
8	Q99439	CNN2	Calponin-2	GLQSGV DIGVK	16	8	< 0.0001	4.8	
9	Q96CG8	CTHR1	Collagen triple helix repeat-containing protein 1	VLFSGSL R	16	9	< 0.0001	4.5	
10	P08238	HS90B	Heat shock protein HSP 90-beta	NPDDITQ EEYGEFY K	16	9	< 0.0001	4.2	(184)
11	P19971	TYPH	Thymidine phosphorylase	MLAAQG VDPGLAR	16	9	< 0.0001	4.1	(185, 186)
12	P31949	S10AB	Protein S100-A11	DGYNYTL SK	16	9	< 0.0001	3.8	(187)
13	P19971	TYPH	Thymidine phosphorylase	VAAALDD GSALGR	16	9	< 0.0001	3.6	(185, 186)
14	P19827	ITIH1	Inter-alpha-trypsin inhibitor heavy chain H1	AAISGEN AGLVR	16	9	0.0030	3.5	
15	Q96HE7	ERO1A	ERO1-like protein alpha	LGAVDES LSEETQK	16	9	< 0.0001	3.0	
16	Q01518	CAP1	Adenylyl cyclase-associated protein 1	VENQEN VSNLVIE DTELK	16	9	< 0.0001	2.9	
17	Q9UBR2	CATZ	Cathepsin Z	NVDGVN YASITR	16	9	0.0004	2.7	
18	P50454	SERP H	Serpin H1	AVLSAEQ LR	16	9	< 0.0001	2.7	(188)

#	Accession	Gene	Protein Name	Peptide sequence	Valid Cancer	Valid Control	q-value	FC Ca/Co	Literature CCA
19	P21291	CSRP1	Cysteine and glycine-rich protein 1	GYGYGQ GAGTLSTDK	16	9	0.0018	2.7	
20	Q9UBR2	CATZ	Cathepsin Z	NSWGEP WGER	16	9	0.0007	2.6	
21	P02792	FRIL	Ferritin light chain	ALFQDIK	16	9	0.0002	2.6	
22	Q01518	CAP1	Adenylyl cyclase-associated protein 1	LSDLLAPI SEQIK	16	9	< 0.0001	2.5	
23	P42224	STAT1	Signal transducer and activator of transcription 1-alpha/beta	TELISVSE VHPSR	14	4	0.0033	2.5	(189)
24	Q9NZM1	MYOF	Myoferlin	ANVTVLD TQIR	16	6	0.0020	2.4	
25	Q96CG8	CTHR1	Collagen triple helix repeat-containing protein 1	IIIEELPK	16	6	0.0020	2.4	
26	P00338	LDHA	L-lactate dehydrogenase A chain	SADTLW GIQK	16	9	< 0.0001	2.4	(190, 191)
27	P00338	LDHA	L-lactate dehydrogenase A chain	VTLTSEE EAR	16	9	< 0.0001	2.3	
28	O75369	FLNB	Filamin-B	FNDEHIP ESPYLVP VIAPSDDAR	16	9	0.0005	2.3	
29	P43490	NAMPT	Nicotinamide phosphoribosyltransferase	STQAPLII RPDSGN PLDTVLK	15	8	< 0.0001	2.2	
30	Q9NZM1	MYOF	Myoferlin	GPVGTVS EAQLAR	16	9	< 0.0001	2.2	
31	P40121	CAPG	Macrophage-capping Protein	ANAQAAA LYK	16	6	0.0006	2.2	(130)
32	P38606	VATA	V-type proton ATPase catalytic subunit A	TVISQSL SK	16	9	< 0.0001	2.2	
33	P14618	KPYM	Pyruvate kinase PKM	LDIDSPPI TAR	16	9	< 0.0001	2.1	(192)
34	P08238	HS90B	Heat shock protein HSP 90-beta	ALLFIPR	14	5	0.0021	2.1	(184)

#	Accession	Gene	Protein Name	Peptide sequence	Valid Cancer	Valid Control	q-value	FC Ca/Co	Literature CCA
35	P27348	1433 T	14–3-3 protein theta	YLIANATNPESK	16	9	0.0078	2.1	(118, 193)
36	P07900	HS90A	Heat shock protein HSP 90-alpha	NPDDITN EEYGEFYK	16	9	0.0092	2.1	(184)
37	P43490	NAMPT	Nicotinamide phosphoribosyltransferase	AVPEGFVIPR	16	7	< 0.0001	2.1	
38	P14618	KPYM	Pyruvate kinase PKM	GDLGIEIPAEK	16	9	< 0.0001	2.1	(192)
39	P10809	CH60	60 kDa heat shock protein, Mitochondrial	LVQDVAN NTNEEAG DGTTTAT VLAR	16	9	0.0408	2.0	(194)
40	P00352	AL1A1	Retinal dehydrogenase 1	TIPIDGNFTYTR	16	9	0.0457	0.46	(195)
41	P04040	CATA	Catalase	ADVLTTGAGNPVGDK	16	9	< 0.0001	0.34	(196)
42	P04040	CATA	Catalase	FNTANDDNVTQVR	16	9	< 0.0001	0.32	(196)
43	P09525	ANXA4	Annexin A4	GLGTDDNTLIR	16	9	0.0005	0.32	(126)
44	Q16853	AOC3	Membrane primary amine Oxidase	YQLAVTQ R	16	9	< 0.0001	0.29	
45	O95994	AGR2	Anterior gradient protein 2 Homolog	LPQTLSR	16	9	< 0.0001	0.29	(197)
46	O60218	AK1BA	Aldo–keto reductase family 1-member B10	SGDDLFPK	16	9	0.0152	0.27	
47	Q13228	SBP1	Methanethiol oxidase	IYVVDVGSEPR	16	9	< 0.0001	0.26	(126)
48	P00167	CYB5	Cytochrome b5	FLEEHPG GEEVLR	16	9	< 0.0001	0.25	(126)
49	Q13228	SBP1	Methanethiol oxidase	LVLPSLISR	14	5	0.0004	0.24	(126)
50	P51884	LUM	Lumican	ISNIPDEYFK	16	9	< 0.0001	0.18	
51	Q9UBX5	FBLN5	Fibulin-5	DQPFTILYR	16	9	< 0.0001	0.18	(198)
52	Q07507	DERM	Dermatopontin	YFESVLD R	16	9	< 0.0001	0.18	(199)
53	P51884	LUM	Lumican	ILGPLSYSK	16	9	< 0.0001	0.16	
54	P55083	MFAP4	Microfibril-associated glycoprotein 4	GFYYSLK	16	9	< 0.0001	0.16	(200)
55	P07585	PGS2	Decorin	VSPGAFTPLVK	16	9	< 0.0001	0.16	(201)

#	Accession	Gene	Protein Name	Peptide sequence	Valid Cancer	Valid Control	q-value	FC Ca/Co	Literature CCA
56	P51888	PRELP	Prolargin	NQLEEVPSALPR	16	9	< 0.0001	0.15	
57	Q07507	DERM	Dermatopontin	GATTTFS AVER	16	9	< 0.0001	0.14	(1992)
58	P55083	MFAP4	Microfibril-associated glycoprotein 4	WTVFQK	14	5	< 0.0001	0.12	(200)
59	P00325	ADH1B	Alcohol dehydrogenase 1B	AAVLWEVK	14	5	< 0.0001	0.12	
60	P07585	PGS2	Decorin	ASYSGVSLFSNPVQ YWEIQPS TFR	16	9	< 0.0001	0.11	(201)
61	P08294	SODE	Extracellular superoxide dismutase	VTGVVLF R	14	5	< 0.0001	0.11	
62	P23141	EST1	Liver carboxylesterase 1	FTPPQPA EPWSFVK	16	9	< 0.0001	0.08	(126, 198)
63	P20774	MIME	Mimecan	LTLFNAK	16	9	< 0.0001	0.07	(198)
64	P23141	EST1	Liver carboxylesterase 1	TVIGDHG DELFSVF GAPFLK	16	9	< 0.0001	0.06	(126, 198)
65	P20774	MIME	Mimecan	DFADIPN LR	16	9	< 0.0001	0.05	(198)

Paper IV

The utility of serum THBS2 as a biomarker for the diagnosis of dCCA (N=51) and PC (N=52) was further explored in paper IV. There was no significant difference in THBS2 levels between dCCA and PC. Unlike CA 19–9, THBS2 levels did not correlate with stage. The combination of THBS2 and CA 19–9 (≥ 35) had a discriminatory power (AUC) of 0.94 in separating dCCA and healthy controls (Figure 9). THBS2 + CA 19–9 (≥ 35) had an AUC of 0.92 for the discrimination of dCCA+ PC from healthy controls. When THBS2 was dichotomized at 51 ng/ml THBS2 (≥ 51) + CA 19–9 (≥ 35) had a sensitivity of 79% with a specificity of 96% for the discrimination of dCCA and PC from healthy controls. THBS2 provided benefit compared to CA 19–9 alone in early- stage disease. In N0 dCCA patients the AUC increased from 0.69 to 0.87 with the addition of THBS2. For N0 PC patients, the AUC increased from 0.70 to 0.81. THBS2 did not provide value to CA 19–9 in discriminating dCCA and PC from the benign pancreatic disease's cohort (N=27). Notably, large heterogeneity within the benign cohort was noted 3/5

patients with autoimmune pancreatitis had substantially elevated THBS2 levels. No correlation between THBS2 levels and prognosis was found.

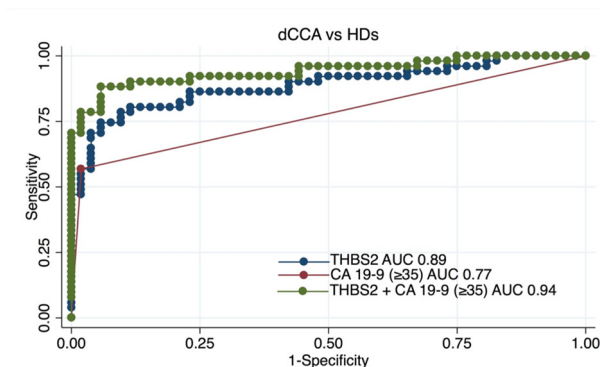


Figure 9. ROC curves for THBS2, CA 19–9 and THBS2 + CA 19–9 combined for dCCA vs healthy donors.

Paper V

The inspiration for the final paper of this thesis came from the findings in paper I and the large discrepancies in the literature regarding dCCA frequency and outcome after surgery. The Swedish National Registry for Pancreatic and Periapillary Cancer was used to evaluate national trends in assessment of periapillary tumor origin, diagnostic accuracy, histopathological evaluation and outcome in Sweden. The frequency of individual periapillary cancers in patients with unresectable/metastatic disease at diagnosis (N=4428) was PC in 84%, dCCA in 4%, AC in 5% and DC 7%. Among patients treated with pancreatoduodenectomy (N=2760) the histopathological diagnosis was PC in 63%, dCCA in 13%, AC in 17% and DC in 7%. Comparing before and after histopathological standardization 57% vs 64% of patients was diagnosed with PC, 15% vs 12% dCCA, 20% vs 16 % AC and 9% vs 7% DC. When stratified by region, the frequency range of PC was 47-71%, dCCA 9-25%, AC 14-21% and DC 3-9%. The rate of R1 resection increased after standardization (32% vs 45%) and large regional differences were seen (12-65%). A similar pattern was found for nodal stage. Standardization increased the frequency of N1 stage (62% vs 77%) and regional differences (52-82%) were found. Median and 5-year OS was 24 months and 21% for PC, 33 months and 31% for dCCA. Median survival was not reached in AC or DC and 5-year survival rates were 54% and 56% respectively (Figure 10). Among patients with PC on final histology 15% had a preoperative diagnosis of non-pancreatic

periampullary tumor. Among patients with non-pancreatic periampullary cancer 23% had a preoperative diagnosis of pancreatic tumor.

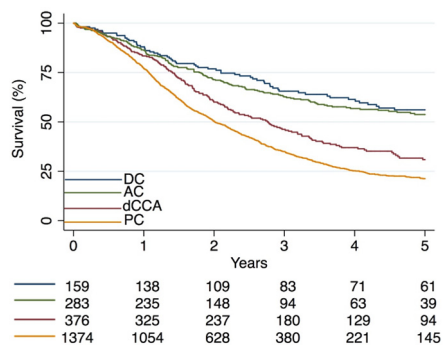


Figure 10. Postoperative survival for patients treated with pancreatoduodenectomy for periampullary cancer stratified by tumor origin.

Discussion

Aspects of clinical management and outcome

In paper I, we found the survival following pancreatoduodenectomy for dCCA (median survival 22.2 months) to be worse than most previous studies (59). A high frequency of negative prognostic factors, such as lymph node metastasis and R1 resection were present, which could have contributed. In paper V, which presented outcome in a national cohort during a comparable time period, the median survival was 33.3 months, which is a substantially better. There were large regional differences with respect to frequency of identified tumor origin in paper V, (9%-25%) for dCCA. It is possible that samples which would have been classified as PC in other centers/studies were included in the paper I cohort, contributing to worse survival. The large discrepancy in frequency of individual periampullary cancers in the literature (72) have most likely influenced survival estimates and contributed to the heterogeneity seen. Further research in dCCA and other periampullary cancers would benefit from reporting the methodology of histopathological evaluation and frequency of individual periampullary cancers in the underlying cohort. The fact that differences in histopathological evaluation of pancreatoduodenectomy specimens exists in Sweden, where a national protocol for evaluation exists, suggest additional quality efforts are warranted to ensure consistency. We believe a similar problem exists globally.

In paper V, we found a diagnosis of PC to be more common in patients with non-resectable periampullary cancer. Non-pancreatic origin was found in 37% of resected patients and 16% of non-resected patients. Although a lower rate of resectable tumors for patients with PC could contribute, this result is likely due to the difficulty in diagnosing individual periampullary cancers clinically. When comparing the pre- and postoperative diagnosis of patients, 15% of PC patients and 23% of non-pancreatic periampullary cancers had an incorrect preoperative diagnosis. The clinical rate of misdiagnosis was similar to a recent study from the Netherlands (202). Distal bile duct tumors had the worst diagnostic accuracy, the concordance between pre- and postoperative diagnosis was 37%. The use of neoadjuvant therapy is emerging primarily for PC (203) but also in CCA (113). In today's oncological treatment algorithm, tumor origin dictates chemotherapy regimens used. Admittedly, there is an overlap in the regimens used for periampullary cancers, which makes the impact of misclassification difficult to

estimate. Several studies have suggested the morphology (intestinal or pancreatobiliary differentiation) to better reflect prognosis and treatment prediction than anatomical tumor origin in periampullary cancers (15-17). Further research is required to define a treatment predictive, biologically relevant tumor stratification and improve diagnostic precision in periampullary cancers. The rate of misdiagnosis should be considered when neoadjuvant therapy is an option and in the design of clinical trials.

In paper I, the presence of lymph node metastasis was confirmed as an important prognostic factor, in concordance with the literature (204-208). Number of lymph node metastases or lymph node ratio did not improve the prediction. Since the publication of Paper I, the AJCC 8th edition was published which redefined the N stage in dCCA (Table 2). The optimal predictive lymph node variable for prognosis is still under debate with heterogeneous study results, probably due to differences in patient selection, surgical technique and histopathological evaluation influencing results (209, 210).

Methodological considerations

Paper I had a retrospective design and a small sample size. Consequently, there was a limiting precision of survival estimates and low statistical power. Paper V was based on a large, validated, prospectively collected national cohort. Limitations of paper V are inherent to registry data, such as quality of registration and availability of variables. We did not have the possibility of comparing diagnostic precision in patients based on preoperative histopathological verification or certainty of preoperative diagnosis. We did also not have data on intestinal or pancreatobiliary differentiation and limited oncological treatment data.

Aspects of tumor biology and biomarkers

In paper III, which represents the first quantitative comparison of proteomes in dCCA and normal bile ducts, bioinformatic analysis of DEPs from the discovery experiment revealed enrichment of the extracellular compartment and extracellular signalling pathways. Studies on the functional properties of the tumor stroma in CCA have been done mostly in iCCA cohorts (211). Given the drastically different microenvironments of the liver and extrahepatic biliary tree, finding from iCCA cannot be directly extrapolated to dCCA. Tumor microenvironment scores based on the amount of desmoplasia and infiltration of immune cell subsets have shown to correlate with prognosis in dCCA. However, few studies on the unique microenvironment in dCCA has been performed (31). One study characterized the tumor-stroma fraction in 16 solid malignancies found pancreatobiliary type

periampullary cancer to have the highest stroma proportion. A high stromal fraction was correlated with better survival (212). Studies on stromal biology in PC have highlighted the importance of tumor-stroma interactions for cancer development, but also the impact of stromal heterogeneity. Tumor suppressive functions of stromal components have been described. Stromal therapeutic targeting has been notoriously difficult, but promising candidates are in development (213). A more thorough description of individual proteins identified in paper III is available in the discussion of the manuscript. Further studies on the unique functional properties of the stroma in dCCA is merited.

Both SPARC and THBS2 are matricellular ECM proteins. In paper II, we found SPARC expression to be frequent in dCCA stroma and also frequently maintained in paired lymph node metastases. Previous studies in BTC cohorts have identified a negative prognostic impact of stromal SPARC expression (144, 145), which was not found in the present study. In paper III, we found THBS2 to be expressed in dCCA stromal, but also epithelial compartments. THBS2 had not previously been associated with CCA. A negative prognostic correlation between stromal THBS2 and DFS was seen. Previously, THBS2 secreted from stromal stellate cells has been shown to promote PC cell invasion in vitro (150). One hypothesis could be that a similar mechanism is in play in dCCA and that THBS2 secreted in dCCA microenvironment contributes to cancer invasiveness and poor prognosis. Like SPARC, stromal THBS2 expression was frequently maintained in lymph node metastases. Previous studies have found primary tumors to prime distant microenvironments and stromal reestablishment as a mechanism driving metastasis outgrowth (214). Pre-metastatic lymph node alterations with increased immunosuppressive signaling and following metastasis, lymph node remodelling are important parts of the metastatic process (215). The microenvironment of metastatic CCA is largely unexplored. The functional roles of SPARC and THBS2 in dCCA and role during metastatic development merits further investigation.

The main finding in paper IV was that serum THBS2 was elevated in dCCA, similarly to PC. THBS2 could aid in identifying early-stage disease compared with CA 19-9 alone in the preoperative setting. There was no benefit of THBS2 in separating dCCA and PC from benign controls. However, heterogeneity was seen in the benign cohort. Three out of five patients with autoimmune pancreatitis had substantially elevated levels of THBS2. After the initiation of this study, Large et al. evaluated THBS2 as a diagnostic biomarker for dCCA and found comparable diagnostic performance (216). Similarly, they found no additive value of THBS2 to CA 19-9 in discriminating benign diseases as a group, however, it did improve performance in patients with high bilirubin. This was a hypothesis we could not test in our study since we had few jaundiced patients in the cohort. Another study evaluated THBS2 expression in prospectively collected pre-diagnostic samples. Unfortunately, THBS2 failed to identify presymptomatic PC development and as

such won't be suitable for screening of asymptomatic individuals (217). Whether this is also the case for dCCA remains to be elucidated, although plausible.

Given the rarity of CCA, and even more dCCA, screening of asymptomatic individuals is not realistic. Any clinical application of a new biomarker would be in the screening of high- risk individuals such as PSC patients or as differential diagnostic tools against benign conditions after a suspicious finding. Evaluations of THBS2 utility in larger cohorts (preferentially including other CCA subtypes) representing these clinical scenarios, either alone or as part of a panel, should be the next step. If utility is found, further method optimization and sequential validation is required prior to clinical use (218).

Methodological considerations

The cohorts used for IHC analysis in papers II and III are retrospectively acquired and relatively small, especially with regards to paired lymph nodes. Thus, the statistical power for detection of prognostic markers is low. The difference in prognostic implication of SPARC expression between the current study and previous studies could in addition to patient selection and the inclusion of all subtypes of BTC in the other studies be due to low power, antibody specificity or cutpoint selection. An additional weakness is that the IHC scoring was done by a single person.

Limitations in the MS analysis in paper III include low sample size. The control group was normal bile ducts from separate patients rather than the more commonly used normal adjacent to tumor cells. Although this can introduce bias, it can also help identify additional proteins since normal appearing cells adjacent to tumors can be influenced by signaling from the tumor "field cancerization" (219). Samples were not similarly orientated to the bile duct. Discovery and verification were done using mostly overlapping samples. Due to practical considerations no additional pre-fractionation of samples was performed, which could have been used to further improve analytical performance. A housekeeping gene normalization was used for the PRM verification. Although housekeeping protein are presumably stably expressed this is not always the case and introduces a source of bias (220).

In paper IV there are some imbalances in the sample matching with cancers patients being slightly older than benign and normal cohorts. Additionally, PC samples had higher N stages than the dCCA cohort. A benefit of the benign group was that it represented "realistic" benign samples with a high enough suspicion of malignancy to motivate surgery, however this also meant it was small and heterogenous with very few individual benign diagnoses. Additionally, no relevant benign biliary controls were included. Preoperative bilirubin levels were obtained from patient charts at varying time points from surgery (and PBD) as well as THBS2/CA 19-9 measurements limiting utility. The THBS2 level in the healthy controls were higher

than previous studies due to unknown technical variation. However, the diagnostic performance was quite similar.

Main conclusions

- Lymph node metastasis is an important prognostic factor after resection for dCCA.
- SPARC is upregulated in dCCA stroma and frequently maintained in lymph node metastases
- Several previously unknown differentially expressed proteins were identified in dCCA. Bioinformatic analysis highlighted stromal alterations in dCCA. THBS2 was validated as upregulated in dCCA cancer cells and stroma. Stromal THBS2 expression is a potential negative prognostic marker. Stromal THBS2 expression was also frequently retained in lymph node metastases
- Serum THBS2 has potential as a diagnostic biomarker for dCCA together with CA 19–9.
- PC diagnosis is more frequent in patients that do not undergo surgical resection for periampullary cancer. Standardization has improved histopathological evaluation of pancreatoduodenectomy specimens, but unmotivated regional differences exist. AC and DC has a significantly better survival following pancreatoduodenectomy than dCCA or PC. The rate of clinical misdiagnosis should be considered when preoperative therapy is an option.
- Further knowledge about the tumor biology in dCCA and optimization of clinical management is necessary to improve survival for patients with this dismal disease.

Future perspectives

The research in CCA has taken large steps forward during the last decades, with an increased understanding of the genetic and molecular alterations involved in cancer development, and improvements in surgical technique and oncological treatment options. Despite this, the prognosis of patients remains poor. This thesis highlights areas of improvement in the clinical management of dCCA, where further research hopefully can contribute to better diagnostic precision, both in the pre- and postoperative setting. The studies of protein expression have highlighted the importance of the stromal compartment in dCCA. Further knowledge regarding the function of stroma during dCCA development and metastatic spread is merited. There is an unmet need for novel diagnostic and prognostic/predictive biomarkers for CCA/dCCA but prior to implementation of a biomarker, extensive method development and validation are required. In the short term, no new biomarkers are likely to be approved. Hopefully, some of the proteins in this thesis can be further studied to elucidate their function during and dCCA development and their biomarker potential, either alone or as part of a panel.

Acknowledgements

I would like to express my sincerest gratitude to all the people involved in the creation of this thesis and supporting me throughout. In particular I would like to thank:

Bodil Andersson, main supervisor. Thank you for enthusiastically and compassionately sharing your knowledge with me and introducing me to the world of bile duct cancer. You are a role model, both as a doctor and researcher, and I will always be grateful for your support and belief in me, even when I did not do so myself.

Roland Andersson, co-supervisor. For sharing your vast experience and never-ending enthusiasm and letting me join your research group what feels like an eternity ago.

Daniel Ansari, co-supervisor. For your vast devotion to research which is an influence on everyone around you.

Katarzyna Said Hilmersson. For invaluable assistance and knowledge in the maze that is laboratory work.

Agata Sator. For devoting your time and energy to make all the histopathological evaluations in this thesis possible.

Current and former members of the research team: **Linus Aronsson**, for enduring living with me for years and being a great friend. **Monika Bauden**, for your many kind words, **Carlos Urey**, for inspirational endurance in research and **Emelie Karnevi**, for teaching me laboratory work.

György Mark-Varga. For gently introducing me to the complicated world of mass spectrometry and creating a warm, welcoming and supportive research environment.

Theresa Kristl, **Dingyuan Hu**, **Indira Pla Parada**, **Aniel Sanchez**, **Melinda Rezeli**, and all other members of the Center of Excellence in Biological and Medical Mass Spectrometry. For your kind help and assistance in making paper III go from a thought to reality.

Co-authors not previously mentioned, **Gert Lindell**, **Johan Nilsson** and **Sam Ghazi**. For providing valuable knowledge and input to make this thesis better.

Monica Keidser, for excellent administrative support.

Richard Fristedt and **Emma Niméus** for excellent opposition and feedback during my halftime seminar.

All current and former colleagues at the departments of Surgery and Oncology at Skane University Hospital for your dedication to patients and for continuously helping me become a better doctor. In particular, I would like to thank my clinical supervisor **Eva Forsland** for guiding me through the world of oncology and making sure I stay on track.

Former schoolteachers **Tenzing, Andreas** and **Elisabeth**, for your dedication to all students and for inspiring me to pursue knowledge.

My friends **Edvard, Gabriel, Sebastian, Isak, Linus, Björn** and **Marcus**, for making life enjoyable through everything from board game evenings to obscure concerts.

My fantastic family with parents **Sonnie** and **Britta**, sisters **Lovisa** and **Sofie** and our pälsklung **Winston**. For your unconditional love and support through all of life.

Charlotte, for endlessly supporting and carrying me through the hardships of science and the creation of this thesis. You are my best friend and every day with you is a good day. I am looking forward to many more, love you lots!

References

1. Roskams TA, Theise ND, Balabaud C, Bhagat G, Bhathal PS, Bioulac-Sage P, et al. Nomenclature of the finer branches of the biliary tree: canals, ductules, and ductular reactions in human livers. *Hepatology* (Baltimore, Md). 2004;39(6):1739-45.
2. Boyer JL. Bile formation and secretion. *Compr Physiol*. 2013;3(3):1035-78.
3. Hundt M, Basit H, John S. Physiology, Bile Secretion. StatPearls. Treasure Island (FL): StatPearls Publishing. Copyright © 2021, StatPearls Publishing LLC.; 2021.
4. Razumilava N, Gores GJ. Classification, diagnosis, and management of cholangiocarcinoma. *Clinical gastroenterology and hepatology : the official clinical practice journal of the American Gastroenterological Association*. 2013;11(1):13-21.e1; quiz e3-4.
5. Banales JM, Marin JJG, Lamarca A, Rodrigues PM, Khan SA, Roberts LR, et al. Cholangiocarcinoma 2020: the next horizon in mechanisms and management. *Nature reviews Gastroenterology & hepatology*. 2020;17(9):557-88.
6. Sarmiento JM, Nagomey DM, Sarr MG, Farnell MB. Periapillary cancers: are there differences? *The Surgical clinics of North America*. 2001;81(3):543-55.
7. Blehacz B, Komuta M, Roskams T, Gores GJ. Clinical diagnosis and staging of cholangiocarcinoma. *Nature reviews Gastroenterology & hepatology*. 2011;8(9):512-22.
8. Rizvi S, Gores GJ. Pathogenesis, diagnosis, and management of cholangiocarcinoma. *Gastroenterology*. 2013;145(6):1215-29.
9. Tyson GL, El-Serag HB. Risk factors for cholangiocarcinoma. *Hepatology* (Baltimore, Md). 2011;54(1):173-84.
10. Valle JW, Kelley RK, Nervi B, Oh DY, Zhu AX. Biliary tract cancer. *Lancet* (London, England). 2021;397(10272):428-44.
11. Khan SA, Tavolari S, Brandi G. Cholangiocarcinoma: Epidemiology and risk factors. *Liver international : official journal of the International Association for the Study of the Liver*. 2019;39 Suppl 1:19-31.
12. Strijker M, Belkous A, van der Geest LG, van Gulik TM, van Hooft JE, de Meijer VE, et al. Treatment and survival of resected and unresected distal cholangiocarcinoma: a nationwide study. *Acta oncologica* (Stockholm, Sweden). 2019;58(7):1048-55.
13. Bergquist A, von Seth E. Epidemiology of cholangiocarcinoma. *Best practice & research Clinical gastroenterology*. 2015;29(2):221-32.
14. Nakanuma Y, Kakuda Y. Pathologic classification of cholangiocarcinoma: New concepts. *Best Practice & Research Clinical Gastroenterology*. 2015;29(2):277-93.

15. Bronsert P, Kohler I, Werner M, Makowiec F, Kuesters S, Hoepfner J, et al. Intestinal-type of differentiation predicts favourable overall survival: confirmatory clinicopathological analysis of 198 periampullary adenocarcinomas of pancreatic, biliary, ampullary and duodenal origin. *BMC cancer*. 2013;13:428.
16. Westgaard A, Pomianowska E, Clausen OP, Gladhaug IP. Intestinal-type and pancreatobiliary-type adenocarcinomas: how does ampullary carcinoma differ from other periampullary malignancies? *Annals of surgical oncology*. 2013;20(2):430-9.
17. Gingras MC, Covington KR, Chang DK, Donehower LA, Gill AJ, Ittmann MM, et al. Ampullary Cancers Harbor ELF3 Tumor Suppressor Gene Mutations and Exhibit Frequent WNT Dysregulation. *Cell reports*. 2016;14(4):907-19.
18. Nakanuma Y, Jang KT, Fukushima N, Furukawa T, Hong SM, Kim H, et al. A statement by the Japan-Korea expert pathologists for future clinicopathological and molecular analyses toward consensus building of intraductal papillary neoplasm of the bile duct through several opinions at the present stage. *Journal of hepato-biliary-pancreatic sciences*. 2018;25(3):181-7.
19. WHO classification of tumours of the digestive system, fourth edition. Bosman, F T; World Health Organization; International Agency for Research on Cancer.
20. Feldmann G, Beatty R, Hruban RH, Maitra A. Molecular genetics of pancreatic intraepithelial neoplasia. *Journal of hepato-biliary-pancreatic surgery*. 2007;14(3):224-32.
21. Aishima S, Kubo Y, Tanaka Y, Oda Y. Histological features of precancerous and early cancerous lesions of biliary tract carcinoma. *Journal of hepato-biliary-pancreatic sciences*. 2014;21(7):448-52.
22. Rizvi S, Gores GJ. Emerging molecular therapeutic targets for cholangiocarcinoma. *Journal of hepatology*. 2017;67(3):632-44.
23. Weinberg BA, Xiu J, Lindberg MR, Shields AF, Hwang JJ, Poorman K, et al. Molecular profiling of biliary cancers reveals distinct molecular alterations and potential therapeutic targets. *J Gastrointest Oncol*. 2019;10(4):652-62.
24. Montal R, Sia D, Montironi C, Leow WQ, Esteban-Fabrá R, Pinyol R, et al. Molecular classification and therapeutic targets in extrahepatic cholangiocarcinoma. *Journal of hepatology*. 2020.
25. Rizzo A, Tavolari S, Ricci AD, Frega G, Palloni A, Relli V, et al. Molecular Features and Targeted Therapies in Extrahepatic Cholangiocarcinoma: Promises and Failures. *Cancers (Basel)*. 2020;12(11):3256.
26. Lundgren S, Hau SO, Elebro J, Heby M, Karnevi E, Nodin B, et al. Mutational Landscape in Resected Periampullary Adenocarcinoma: Relationship With Morphology and Clinical Outcome. *JCO Precision Oncology*. 2019(3):1-8.
27. Hanahan D, Coussens Lisa M. Accessories to the Crime: Functions of Cells Recruited to the Tumor Microenvironment. *Cancer cell*. 2012;21(3):309-22.
28. Sirica AE, Gores GJ. Desmoplastic Stroma and Cholangiocarcinoma: Clinical Implications and Therapeutic Targeting. *Hepatology (Baltimore, Md)*. 2014;59(6):2397-402.

29. Brivio S, Cadamuro M, Strazzabosco M, Fabris L. Tumor reactive stroma in cholangiocarcinoma: The fuel behind cancer aggressiveness. *World Journal of Hepatology*. 2017;9(9):455-68.
30. Cadamuro M, Stecca T, Brivio S, Mariotti V, Fiorotto R, Spirli C, et al. The deleterious interplay between tumor epithelia and stroma in cholangiocarcinoma. *Biochimica et biophysica acta Molecular basis of disease*. 2018;1864(4 Pt B):1435-43.
31. Hwang HW, Kim JY, Lee SE, Choi YS, Sook-Hee H, Lee TJ, et al. Prognostic Effects of Histology-Based Tumour Microenvironment Scores in Resected Distal Bile Duct Cancer. *Histopathology*. 2020.
32. Forner A, Vidili G, Rengo M, Bujanda L, Ponz-Sarvisé M, Lamarca A. Clinical presentation, diagnosis and staging of cholangiocarcinoma. *Liver international : official journal of the International Association for the Study of the Liver*. 2019;39 Suppl 1:98-107.
33. Jhaveri KS, Hosseini-Nik H. MRI of cholangiocarcinoma. *Journal of magnetic resonance imaging : JMRI*. 2015;42(5):1165-79.
34. Lamarca A, Barriuso J, Chander A, McNamara MG, Hubner RA, ÓReilly D, et al. (18)F-fluorodeoxyglucose positron emission tomography ((18)FDG-PET) for patients with biliary tract cancer: Systematic review and meta-analysis. *Journal of hepatology*. 2019;71(1):115-29.
35. van der Gaag NA, Rauws EA, van Eijck CH, Bruno MJ, van der Harst E, Kubben FJ, et al. Preoperative biliary drainage for cancer of the head of the pancreas. *The New England journal of medicine*. 2010;362(2):129-37.
36. Latenstein AEJ, Mackay TM, van Huijgevoort NCM, Bonsing BA, Bosscha K, Hol L, et al. Nationwide practice and outcomes of endoscopic biliary drainage in resectable pancreatic head and periampullary cancer. *HPB*. 2021;23(2):270-8.
37. Lorenz JM. Management of Malignant Biliary Obstruction. *Semin Intervent Radiol*. 2016;33(4):259-67.
38. Trikudanathan G, Navaneethan U, Njei B, Vargo JJ, Parsi MA. Diagnostic yield of bile duct brushings for cholangiocarcinoma in primary sclerosing cholangitis: a systematic review and meta-analysis. *Gastrointestinal endoscopy*. 2014;79(5):783-9.
39. Barr Fritcher EG, Voss JS, Brankley SM, Campion MB, Jenkins SM, Keeney ME, et al. An Optimized Set of Fluorescence In Situ Hybridization Probes for Detection of Pancreatobiliary Tract Cancer in Cytology Brush Samples. *Gastroenterology*. 2015;149(7):1813-24.e1.
40. Navaneethan U, Hasan MK, Lourdasamy V, Njei B, Varadarajulu S, Hawes RH. Single-operator cholangioscopy and targeted biopsies in the diagnosis of indeterminate biliary strictures: a systematic review. *Gastrointestinal endoscopy*. 2015;82(4):608-14.e2.
41. Fritscher-Ravens A, Broering DC, Knoefel WT, Rogiers X, Swain P, Thonke F, et al. EUS-guided fine-needle aspiration of suspected hilar cholangiocarcinoma in potentially operable patients with negative brush cytology. *The American journal of gastroenterology*. 2004;99(1):45-51.

42. Mohamadnejad M, DeWitt JM, Sherman S, LeBlanc JK, Pitt HA, House MG, et al. Role of EUS for preoperative evaluation of cholangiocarcinoma: a large single-center experience. *Gastrointestinal endoscopy*. 2011;73(1):71-8.
43. Asbun HJ, Conlon K, Fernandez-Cruz L, Friess H, Shrikhande SV, Adham M, et al. When to perform a pancreatoduodenectomy in the absence of positive histology? A consensus statement by the International Study Group of Pancreatic Surgery. *Surgery*. 2014;155(5):887-92.
44. Biomarkers and surrogate endpoints: preferred definitions and conceptual framework. *Clinical pharmacology and therapeutics*. 2001;69(3):89-95.
45. Marrelli D, Caruso S, Pedrazzani C, Neri A, Fernandes E, Marini M, et al. CA19-9 serum levels in obstructive jaundice: clinical value in benign and malignant conditions. *American journal of surgery*. 2009;198(3):333-9.
46. Duffy MJ, Sturgeon C, Lamerz R, Haglund C, Holubec VL, Klapdor R, et al. Tumor markers in pancreatic cancer: a European Group on Tumor Markers (EGTM) status report. *Annals of oncology : official journal of the European Society for Medical Oncology / ESMO*. 2010;21(3):441-7.
47. Liang B, Zhong L, He Q, Wang S, Pan Z, Wang T, et al. Diagnostic Accuracy of Serum CA19-9 in Patients with Cholangiocarcinoma: A Systematic Review and Meta-Analysis. *Medical science monitor : international medical journal of experimental and clinical research*. 2015;21:3555-63.
48. Ducreux M, Cuhna AS, Caramella C, Hollebecque A, Burtin P, Goéré D, et al. Cancer of the pancreas: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Annals of oncology : official journal of the European Society for Medical Oncology / ESMO*. 2015;26 Suppl 5:v56-68.
49. Malaguarnera G, Paladina I, Giordano M, Malaguarnera M, Bertino G, Berretta M. Serum markers of intrahepatic cholangiocarcinoma. *Dis Markers*. 2013;34(4):219-28.
50. Macias RIR, Banales JM, Sangro B, Muntané J, Avila MA, Lozano E, et al. The search for novel diagnostic and prognostic biomarkers in cholangiocarcinoma. *Biochimica et biophysica acta Molecular basis of disease*. 2018;1864(4 Pt B):1468-77.
51. Leelawat K, Sakchinabut S, Narong S, Wannaprasert J. Detection of serum MMP-7 and MMP-9 in cholangiocarcinoma patients: evaluation of diagnostic accuracy. *BMC gastroenterology*. 2009;9:30.
52. Loosen SH, Roderburg C, Kauertz KL, Pombeiro I, Leyh C, Benz F, et al. Elevated levels of circulating osteopontin are associated with a poor survival after resection of cholangiocarcinoma. *Journal of hepatology*. 2017;67(4):749-57.
53. Cheon YK, Cho YD, Moon JH, Jang JY, Kim YS, Kim YS, et al. Diagnostic utility of interleukin-6 (IL-6) for primary bile duct cancer and changes in serum IL-6 levels following photodynamic therapy. *The American journal of gastroenterology*. 2007;102(10):2164-70.
54. Huang L, Chen W, Liang P, Hu W, Zhang K, Shen S, et al. Serum CYFRA 21-1 in Biliary Tract Cancers: A Reliable Biomarker for Gallbladder Carcinoma and Intrahepatic Cholangiocarcinoma. *Digestive diseases and sciences*. 2015;60(5):1273-83.

55. Ruys AT, Groot Koerkamp B, Wiggers JK, Klumpen HJ, ten Kate FJ, van Gulik TM. Prognostic biomarkers in patients with resected cholangiocarcinoma: a systematic review and meta-analysis. *Annals of surgical oncology*. 2014;21(2):487-500.
56. Björnstedt M, Danielsson O, Fernandez Moro C, Ghazi S, Glaumann H, Nedkova A, et al. Gastrointestinal pathology-pancreas and periampullary region. Recommendations from the KVASt study group of the Swedish society for pathology.
<https://www.svfp.se/foreningar/uploads/L15178/kvast/lever/KVASt%20Pankreas%202021/KVASt%20Pankreas%202021%20dok%20uppdaterad%2010813.pdf>. Accessed September 2021
57. Wang X, Yu GY, Chen M, Wei R, Chen J, Wang Z. Pattern of distant metastases in primary extrahepatic bile-duct cancer: A SEER-based study. *Cancer medicine*. 2018;7(10):5006-14.
58. Khan SA, Davidson BR, Goldin RD, Heaton N, Karani J, Pereira SP, et al. Guidelines for the diagnosis and treatment of cholangiocarcinoma: an update. *Gut*. 2012;61(12):1657-69.
59. Zhou Y, Liu S, Wu L, Wan T. Survival after surgical resection of distal cholangiocarcinoma: A systematic review and meta-analysis of prognostic factors. *Asian journal of surgery / Asian Surgical Association*. 2015.
60. DeOliveira ML, Cunningham SC, Cameron JL, Kamangar F, Winter JM, Lillemoe KD, et al. Cholangiocarcinoma: thirty-one-year experience with 564 patients at a single institution. *Annals of surgery*. 2007;245(5):755-62.
61. Ethun CG, Lopez-Aguilar AG, Pawlik TM, Poultsides G, Idrees K, Fields RC, et al. Distal Cholangiocarcinoma and Pancreas Adenocarcinoma: Are They Really the Same Disease? A 13-Institution Study from the US Extrahepatic Biliary Malignancy Consortium and the Central Pancreas Consortium. *Journal of the American College of Surgeons*. 2017;224(4):406-13.
62. Ecker BL, Vining CC, Roses RE, Maggino L, Lee MK, Drebin JA, et al. Identification of Patients for Adjuvant Therapy After Resection of Carcinoma of the Extrahepatic Bile Ducts: A Propensity Score-Matched Analysis. *Annals of surgical oncology*. 2017;24(13):3926-33.
63. Petrova E, Ruckert F, Zach S, Shen Y, Weitz J, Grutzmann R, et al. Survival outcome and prognostic factors after pancreatoduodenectomy for distal bile duct carcinoma: a retrospective multicenter study. *Langenbeck's archives of surgery / Deutsche Gesellschaft für Chirurgie*. 2017;402(5):831-40.
64. Tol JA, Brosens LA, van Dieren S, van Gulik TM, Busch OR, Besselink MG, et al. Impact of lymph node ratio on survival in patients with pancreatic and periampullary cancer. *The British journal of surgery*. 2015;102(3):237-45.
65. Courtin-Tanguy L, Turrini O, Bergeat D, Truant S, Darnis B, Delpero JR, et al. Multicentre study of the impact of factors that may affect long-term survival following pancreaticoduodenectomy for distal cholangiocarcinoma. *HPB : the official journal of the International Hepato Pancreato Biliary Association*. 2018;20(5):405-10.

66. Lang SA, Bednarsch J, Joechle K, Amygdalos I, Czigany Z, Heij L, et al. Prognostic biomarkers for cholangiocarcinoma (CCA): state of the art. Expert review of gastroenterology & hepatology. 2021;15(5):497-510.
67. Sahara K, Tsilimigras DI, Toyoda J, Miyake K, Ethun CG, Maithele SK, et al. Defining the Risk of Early Recurrence Following Curative-Intent Resection for Distal Cholangiocarcinoma. Annals of surgical oncology. 2021.
68. Beetz O, Klein M, Schrem H, Gwiasda J, Vondran FWR, Oldhafer F, et al. Relevant prognostic factors influencing outcome of patients after surgical resection of distal cholangiocarcinoma. BMC surgery. 2018;18(1):56.
69. Andrianello S, Marchegiani G, Malleo G, Rusev BC, Scarpa A, Bonamini D, et al. Over 700 Whipples for Pancreaticobiliary Malignancies: Postoperative Morbidity Is an Additional Negative Prognostic Factor for Distal Bile Duct Cancer. Journal of gastrointestinal surgery : official journal of the Society for Surgery of the Alimentary Tract. 2017;21(3):527-33.
70. Lopez-Aguilar AG, Ethun CG, Pawlik TM, Tran T, Poultsides GA, Isom CA, et al. Association of Perioperative Transfusion with Recurrence and Survival After Resection of Distal Cholangiocarcinoma: A 10-Institution Study from the US Extrahepatic Biliary Malignancy Consortium. Annals of surgical oncology. 2019;26(6):1814-23.
71. Soer E, Brosens L, van de Vijver M, Dijk F, van Velthuisen ML, Farina-Sarasqueta A, et al. Dilemmas for the pathologist in the oncologic assessment of pancreatoduodenectomy specimens : An overview of different grossing approaches and the relevance of the histopathological characteristics in the oncologic assessment of pancreatoduodenectomy specimens. Virchows Archiv : an international journal of pathology. 2018;472(4):533-43.
72. Verbeke CS, Gladhaug IP. Resection margin involvement and tumour origin in pancreatic head cancer. The British journal of surgery. 2012;99(8):1036-49.
73. Menon KV, Gomez D, Smith AM, Anthoney A, Verbeke CS. Impact of margin status on survival following pancreatoduodenectomy for cancer: the Leeds Pathology Protocol (LEEPP). HPB : the official journal of the International Hepato Pancreato Biliary Association. 2009;11(1):18-24.
74. Verbeke CS, Leitch D, Menon KV, McMahon MJ, Guillou PJ, Anthoney A. Redefining the R1 resection in pancreatic cancer. The British journal of surgery. 2006;93(10):1232-7.
75. Adsay NV, Basturk O, Saka B, Bagci P, Ozdemir D, Balci S, et al. Whipple made simple for surgical pathologists: orientation, dissection, and sampling of pancreaticoduodenectomy specimens for a more practical and accurate evaluation of pancreatic, distal common bile duct, and ampullary tumors. The American journal of surgical pathology. 2014;38(4):480-93.
76. van Roessel S, Soer EC, van Dieren S, Koens L, van Velthuisen MLF, Doukas M, et al. Axial slicing versus bivalving in the pathological examination of pancreatoduodenectomy specimens (APOLLO): a multicentre randomized controlled trial. HPB : the official journal of the International Hepato Pancreato Biliary Association. 2021.

77. Whipple AO, Parsons WB, Mullins CR. TREATMENT OF CARCINOMA OF THE AMPULLA OF VATER. *Annals of surgery*. 1935;102(4):763-79.
78. Hackert T, Probst P, Knebel P, Doerr-Harim C, Bruckner T, Klaiber U, et al. Pylorus Resection Does Not Reduce Delayed Gastric Emptying After Partial Pancreatoduodenectomy: A Blinded Randomized Controlled Trial (PROPP Study, DRKS00004191). *Annals of surgery*. 2018;267(6):1021-7.
79. Liles JS, Katz MH. Pancreaticoduodenectomy with vascular resection for pancreatic head adenocarcinoma. Expert review of anticancer therapy. 2014;14(8):919-29.
80. Tee MC, Krajewski AC, Groeschl RT, Farnell MB, Nagorney DM, Kendrick ML, et al. Indications and Perioperative Outcomes for Pancreatectomy with Arterial Resection. *Journal of the American College of Surgeons*. 2018;227(2):255-69.
81. Tingstedt B, Andersson B, Jönsson C, Formichov V, Bratlie S-O, Öhman M, et al. First results from the Swedish National Pancreatic and Periampullary Cancer Registry. *HPB*. 2019;21(1):34-42.
82. Dindo D, Demartines N, Clavien PA. Classification of surgical complications: a new proposal with evaluation in a cohort of 6336 patients and results of a survey. *Annals of surgery*. 2004;240(2):205-13.
83. Bassi C, Marchegiani G, Dervenis C, Sarr M, Abu Hilal M, Adham M, et al. The 2016 update of the International Study Group (ISGPS) definition and grading of postoperative pancreatic fistula: 11 Years After. *Surgery*. 2017;161(3):584-91.
84. Wente MN, Bassi C, Dervenis C, Fingerhut A, Gouma DJ, Izbicki JR, et al. Delayed gastric emptying (DGE) after pancreatic surgery: a suggested definition by the International Study Group of Pancreatic Surgery (ISGPS). *Surgery*. 2007;142(5):761-8.
85. Wente MN, Veit JA, Bassi C, Dervenis C, Fingerhut A, Gouma DJ, et al. Postpancreatectomy hemorrhage (PPH): an International Study Group of Pancreatic Surgery (ISGPS) definition. *Surgery*. 2007;142(1):20-5.
86. Andrianello S, Paiella S, Allegrini V, Ramera M, Pulvirenti A, Malleo G, et al. Pancreaticoduodenectomy for distal cholangiocarcinoma: surgical results, prognostic factors, and long-term follow-up. *Langenbeck's archives of surgery / Deutsche Gesellschaft für Chirurgie*. 2015;400(5):623-8.
87. Pratt WB, Callery MP, Vollmer CM, Jr. Risk prediction for development of pancreatic fistula using the ISGPF classification scheme. *World journal of surgery*. 2008;32(3):419-28.
88. Williamsson C, Stenvall K, Wennerblom J, Andersson R, Andersson B, Tingstedt B. Predictive Factors for Postoperative Pancreatic Fistula—A Swedish Nationwide Register-Based Study. *World journal of surgery*. 2020;44(12):4207-13.
89. Eckel F, Schmid RM. Chemotherapy in advanced biliary tract carcinoma: a pooled analysis of clinical trials. *British journal of cancer*. 2007;96(6):896-902.
90. Valle J, Wasan H, Palmer DH, Cunningham D, Anthoney A, Maraveyas A, et al. Cisplatin plus gemcitabine versus gemcitabine for biliary tract cancer. *The New England journal of medicine*. 2010;362(14):1273-81.

91. Fiteni F, Nguyen T, Vernerey D, Paillard MJ, Kim S, Demarchi M, et al. Cisplatin/gemcitabine or oxaliplatin/gemcitabine in the treatment of advanced biliary tract cancer: a systematic review. *Cancer medicine*. 2014;3(6):1502-11.
92. Sharma A, Kalyan Mohanti B, Pal Chaudhary S, Sreenivas V, Kumar Sahoo R, Kumar Shukla N, et al. Modified gemcitabine and oxaliplatin or gemcitabine + cisplatin in unresectable gallbladder cancer: Results of a phase III randomised controlled trial. *European journal of cancer (Oxford, England : 1990)*. 2019;123:162-70.
93. Lagenfelt H, Blomstrand H, Elander NO. Real-World Evidence on Palliative Gemcitabine and Oxaliplatin (GemOx) Combination Chemotherapy in Advanced Biliary Tract Cancer. *Cancers (Basel)*. 2021;13(14):3507.
94. Adeva J, Sangro B, Salati M, Edeline J, La Casta A, Bittoni A, et al. Medical treatment for cholangiocarcinoma. *Liver international : official journal of the International Association for the Study of the Liver*. 2019;39 Suppl 1:123-42.
95. Lamarca A, Palmer DH, Wasan HS, Ross PJ, Ma YT, Arora A, et al. Second-line FOLFOX chemotherapy versus active symptom control for advanced biliary tract cancer (ABC-06): a phase 3, open-label, randomised, controlled trial. *The Lancet Oncology*. 2021;22(5):690-701.
96. Valle JW, Lamarca A, Goyal L, Barriuso J, Zhu AX. New Horizons for Precision Medicine in Biliary Tract Cancers. *Cancer Discov*. 2017;7(9):943-62.
97. Valle JW. Targeted therapy for cholangiocarcinoma. *The lancet Gastroenterology & hepatology*. 2019;4(9):661-2.
98. Le DT, Durham JN, Smith KN, Wang H, Bartlett BR, Aulakh LK, et al. Mismatch repair deficiency predicts response of solid tumors to PD-1 blockade. *Science (New York, NY)*. 2017;357(6349):409-13.
99. Marabelle A, Le DT, Ascierto PA, Di Giacomo AM, De Jesus-Acosta A, Delord JP, et al. Efficacy of Pembrolizumab in Patients With Noncolorectal High Microsatellite Instability/Mismatch Repair-Deficient Cancer: Results From the Phase II KEYNOTE-158 Study. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology*. 2020;38(1):1-10.
100. Drilon A, Laetsch TW, Kummar S, DuBois SG, Lassen UN, Demetri GD, et al. Efficacy of Larotrectinib in TRK Fusion-Positive Cancers in Adults and Children. *The New England journal of medicine*. 2018;378(8):731-9.
101. Drilon A, Siena S, Ou SI, Patel M, Ahn MJ, Lee J, et al. Safety and Antitumor Activity of the Multitargeted Pan-TRK, ROS1, and ALK Inhibitor Entrectinib: Combined Results from Two Phase I Trials (ALKA-372-001 and STARTRK-1). *Cancer Discov*. 2017;7(4):400-9.
102. Doebele RC, Drilon A, Paz-Ares L, Siena S, Shaw AT, Farago AF, et al. Entrectinib in patients with advanced or metastatic NTRK fusion-positive solid tumours: integrated analysis of three phase 1-2 trials. *The Lancet Oncology*. 2020;21(2):271-82.
103. Demols A, Perez-Casanova L, Rocq L, Charry M, Nève ND, Verrellen A, et al. NTRK gene fusions in bilio-pancreatic cancers. *Journal of Clinical Oncology*. 2020;38(15_suppl):e16664-e.

104. Javle M, Borad MJ, Azad NS, Kurzrock R, Abou-Alfa GK, George B, et al. Pertuzumab and trastuzumab for HER2-positive, metastatic biliary tract cancer (MyPathway): a multicentre, open-label, phase 2a, multiple basket study. *The Lancet Oncology*. 2021;22(9):1290-300.
105. Lamarca A, Edeline J, McNamara MG, Hubner RA, Nagino M, Bridgewater J, et al. Current standards and future perspectives in adjuvant treatment for biliary tract cancers. *Cancer treatment reviews*. 2020;84:101936.
106. Neoptolemos JP, Moore MJ, Cox TF, Valle JW, Palmer DH, McDonald AC, et al. Effect of adjuvant chemotherapy with fluorouracil plus folinic acid or gemcitabine vs observation on survival in patients with resected perampullary adenocarcinoma: the ESPAC-3 perampullary cancer randomized trial. *Jama*. 2012;308(2):147-56.
107. Ebata T, Hirano S, Konishi M, Uesaka K, Tsuchiya Y, Ohtsuka M, et al. Randomized clinical trial of adjuvant gemcitabine chemotherapy versus observation in resected bile duct cancer. *The British journal of surgery*. 2018;105(3):192-202.
108. Edeline J, Benabdelghani M, Bertaut A, Watelet J, Hammel P, Joly JP, et al. Gemcitabine and Oxaliplatin Chemotherapy or Surveillance in Resected Biliary Tract Cancer (PRODIGE 12-ACCORD 18-UNICANCER GI): A Randomized Phase III Study. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology*. 2019;37(8):658-67.
109. Primrose JN, Fox RP, Palmer DH, Malik HZ, Prasad R, Mirza D, et al. Capecitabine compared with observation in resected biliary tract cancer (BILCAP): a randomised, controlled, multicentre, phase 3 study. *The Lancet Oncology*. 2019;20(5):663-73.
110. National Comprehensive Cancer Network. Biliary Tract Cancer: Extrahepatic cholangiocarcinoma. NCCN guidelines, Version 5, 2021, .
111. Ben-Josef E, Guthrie KA, El-Khoueiry AB, Corless CL, Zalupski MM, Lowy AM, et al. SWOG S0809: A Phase II Intergroup Trial of Adjuvant Capecitabine and Gemcitabine Followed by Radiotherapy and Concurrent Capecitabine in Extrahepatic Cholangiocarcinoma and Gallbladder Carcinoma. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology*. 2015;33(24):2617-22.
112. Shroff RT, Kennedy EB, Bachini M, Bekaii-Saab T, Crane C, Edeline J, et al. Adjuvant Therapy for Resected Biliary Tract Cancer: ASCO Clinical Practice Guideline. *Journal of Clinical Oncology*. 2019;37(12):1015-27.
113. Rizzo A, Brandi G. Neoadjuvant therapy for cholangiocarcinoma: A comprehensive literature review. *Cancer Treatment and Research Communications*. 2021;27:100354.
114. Crutchfield CA, Thomas SN, Sokoll LJ, Chan DW. Advances in mass spectrometry-based clinical biomarker discovery. *Clinical Proteomics*. 2016;13:1.
115. Magdeldin S, Enany S, Yoshida Y, Xu B, Zhang Y, Zureena Z, et al. Basics and recent advances of two dimensional- polyacrylamide gel electrophoresis. *Clinical Proteomics*. 2014;11(1):16.
116. Mallick P, Kuster B. Proteomics: a pragmatic perspective. *Nature biotechnology*. 2010;28(7):695-709.
117. Zhang Y, Fonslow BR, Shan B, Baek MC, Yates JR. Protein Analysis by Shotgun/Bottom-up Proteomics. *Chemical reviews*. 2013;113(4):2343-94.

118. Darby IA, Vuillier-Devillers K, Pinault E, Sarrazy V, Lepreux S, Balabaud C, et al. Proteomic analysis of differentially expressed proteins in peripheral cholangiocarcinoma. *Cancer Microenvironment*. 2011;4(1):73-91.
119. Kawase H, Fujii K, Miyamoto M, Kubota KC, Hirano S, Kondo S, et al. Differential LC-MS-based proteomics of surgical human cholangiocarcinoma tissues. *Journal of proteome research*. 2009;8(8):4092-103.
120. Maeda S, Morikawa T, Takadate T, Suzuki T, Minowa T, Hanagata N, et al. Mass spectrometry-based proteomic analysis of formalin-fixed paraffin-embedded extrahepatic cholangiocarcinoma. *Journal of hepato-biliary-pancreatic sciences*. 2015.
121. Padden J, Megger DA, Bracht T, Reis H, Ahrens M, Kohl M, et al. Identification of novel biomarker candidates for the immunohistochemical diagnosis of cholangiocellular carcinoma. *Molecular & cellular proteomics : MCP*. 2014;13(10):2661-72.
122. Le Faouder J, Laouirem S, Alexandrov T, Ben-Harzallah S, Leger T, Albuquerque M, et al. Tumoral heterogeneity of hepatic cholangiocarcinomas revealed by MALDI imaging mass spectrometry. *Proteomics*. 2014;14(7-8):965-72.
123. Shi Y, Deng X, Zhan Q, Shen B, Jin X, Zhu Z, et al. A prospective proteomic-based study for identifying potential biomarkers for the diagnosis of cholangiocarcinoma. *Journal of gastrointestinal surgery : official journal of the Society for Surgery of the Alimentary Tract*. 2013;17(9):1584-91.
124. Matsuda A, Kuno A, Matsuzaki H, Kawamoto T, Shikanai T, Nakanuma Y, et al. Glycoproteomics-based cancer marker discovery adopting dual enrichment with *Wisteria floribunda* agglutinin for high specific glyco-diagnosis of cholangiocarcinoma. *Journal of proteomics*. 2013;85:1-11.
125. Maimonis PJ, Zen Y, Britton DJ, Brand A, Ward M, Pike I, et al. Identification of new liver tumor biomarkers using proteomics. *Journal of Clinical Oncology*. 2012;30(15).
126. Dos Santos A, Court M, Thiers V, Sar S, Guettier C, Samuel D, et al. Identification of cellular targets in human intrahepatic cholangiocarcinoma using laser microdissection and accurate mass and time tag proteomics. *Molecular and Cellular Proteomics*. 2010;9(9):1991-2004.
127. Kristiansen TZ, Harsha HC, Gronborg M, Maitra A, Pandey A. Differential membrane proteomics using 18O-labeling to identify biomarkers for cholangiocarcinoma. *Journal of proteome research*. 2008;7(11):4670-7.
128. Thanan R, Oikawa S, Yongvanit P, Hiraku Y, Ma N, Pinlaor S, et al. Inflammation-induced protein carbonylation contributes to poor prognosis for cholangiocarcinoma. *Free radical biology & medicine*. 2012;52(8):1465-72.
129. Padden J, Ahrens M, Kalsch J, Bertram S, Megger DA, Bracht T, et al. Immunohistochemical Markers Distinguishing Cholangiocellular Carcinoma (CCC) from Pancreatic Ductal Adenocarcinoma (PDAC) Discovered by Proteomic Analysis of Microdissected Cells. *Molecular & cellular proteomics : MCP*. 2016;15(3):1072-82.

130. Morofuji N, Ojima H, Onaya H, Okusaka T, Shimada K, Sakamoto Y, et al. Macrophage-capping protein as a tissue biomarker for prediction of response to gemcitabine treatment and prognosis in cholangiocarcinoma. *Journal of proteomics*. 2012;75(5):1577-89.
131. Onsurathum S, Haonon O, Pinlaor P, Pairojkul C, Khuntikeo N, Thanan R, et al. Proteomics detection of S100A6 in tumor tissue interstitial fluid and evaluation of its potential as a biomarker of cholangiocarcinoma. *Tumour biology : the journal of the International Society for Oncodevelopmental Biology and Medicine*. 2018;40(4):1010428318767195.
132. Takenami T, Maeda S, Karasawa H, Suzuki T, Furukawa T, Morikawa T, et al. Novel biomarkers distinguishing pancreatic head Cancer from distal cholangiocarcinoma based on proteomic analysis. *BMC cancer*. 2019;19(1):318-.
133. Nagaraju GP, Dontula R, El-Rayes BF, Lakka SS. Molecular mechanisms underlying the divergent roles of SPARC in human carcinogenesis. *Carcinogenesis*. 2014;35(5):967-73.
134. Bradshaw AD, Sage EH. SPARC, a matricellular protein that functions in cellular differentiation and tissue response to injury. *The Journal of clinical investigation*. 2001;107(9):1049-54.
135. Brekken RA, Sage EH. SPARC, a matricellular protein: at the crossroads of cell-matrix communication. *Matrix biology : journal of the International Society for Matrix Biology*. 2001;19(8):816-27.
136. Rivera LB, Bradshaw AD, Brekken RA. The regulatory function of SPARC in vascular biology. *Cellular and molecular life sciences : CMLS*. 2011;68(19):3165-73.
137. Delany AM, Hankenson KD. Thrombospondin-2 and SPARC/osteonectin are critical regulators of bone remodeling. *Journal of cell communication and signaling*. 2009;3(3-4):227-38.
138. Bradshaw AD. The role of SPARC in extracellular matrix assembly. *Journal of cell communication and signaling*. 2009;3(3-4):239-46.
139. Bradshaw AD. Diverse biological functions of the SPARC family of proteins. *The international journal of biochemistry & cell biology*. 2012;44(3):480-8.
140. Arnold SA, Brekken RA. SPARC: a matricellular regulator of tumorigenesis. *Journal of Cell Communication and Signaling*. 2009;3(3-4):255-73.
141. Liang C, Shi S, Meng Q, Liang D, Ji S, Zhang B, et al. Do anti-stroma therapies improve extrinsic resistance to increase the efficacy of gemcitabine in pancreatic cancer? *Cellular and molecular life sciences : CMLS*. 2018;75(6):1001-12.
142. Giordano G, Pancione M, Olivieri N, Parcesepe P, Velocci M, Di Raimo T, et al. Nano albumin bound-paclitaxel in pancreatic cancer: Current evidences and future directions. *World journal of gastroenterology*. 2017;23(32):5875-86.
143. Hidalgo M, Plaza C, Musteanu M, Illei P, Brachmann CB, Heise C, et al. SPARC Expression Did Not Predict Efficacy of nab-Paclitaxel plus Gemcitabine or Gemcitabine Alone for Metastatic Pancreatic Cancer in an Exploratory Analysis of the Phase III MPACT Trial. *Clinical cancer research : an official journal of the American Association for Cancer Research*. 2015;21(21):4811-8.

144. Toyota K, Murakami Y, Kondo N, Uemura K, Nakagawa N, Takahashi S, et al. Impact of Secreted Protein Acidic and Rich in Cysteine (SPARC) Expression on Prognosis After Surgical Resection for Biliary Carcinoma. *Journal of Gastrointestinal Surgery*. 2017;21(6):990-9.
145. Nakashima S, Kobayashi S, Sakai D, Tomokuni A, Tomimaru Y, Hama N, et al. Prognostic impact of tumoral and/or peri-tumoral stromal SPARC expressions after surgery in patients with biliary tract cancer. *Journal of surgical oncology*. 2014;110(8):1016-22.
146. Nakamura T, Bornstein P. Matricellular Proteins*. In: Janes SM, editor. *Encyclopedia of Respiratory Medicine (Second Edition)*. Oxford: Academic Press; 2022. p. 137-51.
147. Calabro NE, Kristofik NJ, Kyriakides TR. Thrombospondin-2 and extracellular matrix assembly. *Biochimica et biophysica acta*. 2014;1840(8):2396-402.
148. de Fraipont F, Nicholson AC, Feige JJ, Van Meir EG. Thrombospondins and tumor angiogenesis. *Trends in molecular medicine*. 2001;7(9):401-7.
149. Nakamura M, Oida Y, Abe Y, Yamazaki H, Mukai M, Matsuyama M, et al. Thrombospondin-2 inhibits tumor cell invasion through the modulation of MMP-9 and uPA in pancreatic cancer cells. *Molecular medicine reports*. 2008;1(3):423-7.
150. Farrow B, Berger DH, Rowley D. Tumor-Derived Pancreatic Stellate Cells Promote Pancreatic Cancer Cell Invasion Through Release of Thrombospondin-2. *Journal of Surgical Research*. 2009;156(1):155-60.
151. Kim J, Bamlet WR, Oberg AL, Chaffee KG, Donahue G, Cao X-J, et al. Detection of early pancreatic ductal adenocarcinoma with thrombospondin-2 and CA19-9 blood markers. *Science translational medicine*. 2017;9(398):5583.
152. Ansari D, Williamsson C, Tingstedt B, Andersson B, Lindell G, Andersson R. Pancreaticoduodenectomy--the transition from a low- to a high-volume center. *Scandinavian journal of gastroenterology*. 2014;49(4):481-4.
153. Lipman NS, Jackson LR, Trudel LJ, Weis-Garcia F. Monoclonal versus polyclonal antibodies: distinguishing characteristics, applications, and information resources. *ILAR journal*. 2005;46(3):258-68.
154. Taylor CR, Rudbeck L. *Immunohistochemical staining methods*. 6th ed. Glostrup, Denmark. Dako Corporation. 2013.
155. Fedchenko N, Reifenrath J. Different approaches for interpretation and reporting of immunohistochemistry analysis results in the bone tissue - a review. *Diagnostic pathology*. 2014;9:221.
156. Ormanns S, Haas M, Baechmann S, Altendorf-Hofmann A, Remold A, Quietzsch D, et al. Impact of SPARC expression on outcome in patients with advanced pancreatic cancer not receiving nab-paclitaxel: a pooled analysis from prospective clinical and translational trials. *British journal of cancer*. 2016;115(12):1520-9.
157. Gustafsson OJ, Arentz G, Hoffmann P. Proteomic developments in the analysis of formalin-fixed tissue. *Biochimica et biophysica acta*. 2015;1854(6):559-80.
158. Steiner C, Ducret A, Tille J-C, Thomas M, McKee TA, Rubbia-Brandt LA, et al. Applications of mass spectrometry for quantitative protein analysis in formalin-fixed paraffin-embedded tissues. *Proteomics*. 2014;14(4-5):441-51.

159. Polanski M, Anderson NL. A list of candidate cancer biomarkers for targeted proteomics. *Biomarker insights*. 2007;1:1-48.
160. Peterson AC, Russell JD, Bailey DJ, Westphall MS, Coon JJ. Parallel Reaction Monitoring for High Resolution and High Mass Accuracy Quantitative, Targeted Proteomics. *Molecular & cellular proteomics : MCP*. 2012;11(11):1475-88.
161. Bourmaud A, Gallien S, Domon B. Parallel reaction monitoring using quadrupole-Orbitrap mass spectrometer: Principle and applications. *Proteomics*. 2016;16(15-16):2146-59.
162. Rauniyar N. Parallel Reaction Monitoring: A Targeted Experiment Performed Using High Resolution and High Mass Accuracy Mass Spectrometry. *Int J Mol Sci*. 2015;16(12):28566-81.
163. Schilling B, Rardin MJ, MacLean BX, Zawadzka AM, Frewen BE, Cusack MP, et al. Platform-independent and label-free quantitation of proteomic data using MS1 extracted ion chromatograms in skyline: application to protein acetylation and phosphorylation. *Molecular & cellular proteomics : MCP*. 2012;11(5):202-14.
164. Simpson RE, Yip-Schneider MT, Wu H, Fan H, Liu Z, Korc M, et al. Circulating Thrombospondin-2 enhances prediction of malignant intraductal papillary mucinous neoplasm. *American journal of surgery*. 2019;217(3):425-8.
165. Peng HY, Chang MC, Hu CM, Yang HL, Lee WH, Chang YT. Thrombospondin-2 is a Highly Specific Diagnostic Marker and is Associated with Prognosis in Pancreatic Cancer. *Annals of surgical oncology*. 2019;26(3):807-14.
166. Berger AW, Schwerdel D, Reinacher-Schick A, Uhl W, Algul H, Friess H, et al. A Blood-Based Multi Marker Assay Supports the Differential Diagnosis of Early-Stage Pancreatic Cancer. *Theranostics*. 2019;9(5):1280-7.
167. Nie RC, Zou XB, Yuan SQ, Chen YB, Chen S, Chen YM, et al. Disease-free survival as a surrogate endpoint for overall survival in adjuvant trials of pancreatic cancer: a meta-analysis of 20 randomized controlled trials. *BMC cancer*. 2020;20(1):421.
168. Pocock SJ, Clayton TC, Altman DG. Survival plots of time-to-event outcomes in clinical trials: good practice and pitfalls. *Lancet (London, England)*. 2002;359(9318):1686-9.
169. Bradburn MJ, Clark TG, Love SB, Altman DG. Survival analysis part II: multivariate data analysis--an introduction to concepts and methods. *British journal of cancer*. 2003;89(3):431-6.
170. Bursac Z, Gauss CH, Williams DK, Hosmer DW. Purposeful selection of variables in logistic regression. *Source code for biology and medicine*. 2008;3:17-.
171. Tyanova S, Temu T, Sinitcyn P, Carlson A, Hein MY, Geiger T, et al. The Perseus computational platform for comprehensive analysis of (prote)omics data. *Nature methods*. 2016;13(9):731-40.
172. Gene Ontology C. Gene Ontology Consortium: going forward. *Nucleic acids research*. 2015;43(Database issue):D1049-D56.
173. Mi H, Huang X, Muruganujan A, Tang H, Mills C, Kang D, et al. PANTHER version 11: expanded annotation data from Gene Ontology and Reactome pathways, and data analysis tool enhancements. *Nucleic acids research*. 2017;45(D1):D183-d9.

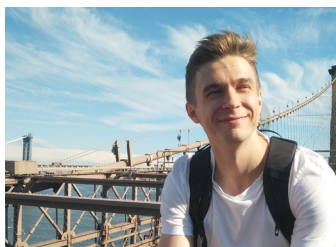
174. Kanehisa M, Sato Y, Kawashima M, Furumichi M, Tanabe M. KEGG as a reference resource for gene and protein annotation. *Nucleic acids research*. 2016;44(D1):D457-62.
175. Fabregat A, Jupe S, Matthews L, Sidiropoulos K, Gillespie M, Garapati P, et al. The Reactome Pathway Knowledgebase. *Nucleic acids research*. 2018;46(D1):D649-d55.
176. Szklarczyk D, Morris JH, Cook H, Kuhn M, Wyder S, Simonovic M, et al. The STRING database in 2017: quality-controlled protein-protein association networks, made broadly accessible. *Nucleic acids research*. 2017;45(D1):D362-D8.
177. Leeftang MMG, Rutjes AWS, Reitsma JB, Hooft L, Bossuyt PMM. Variation of a test's sensitivity and specificity with disease prevalence. *CMAJ*. 2013;185(11):E537-E44.
178. Lukic N, Visentin R, Delhay M, Frossard JL, Lescuyer P, Dumonceau JM, et al. An integrated approach for comparative proteomic analysis of human bile reveals overexpressed cancer-associated proteins in malignant biliary stenosis. *Biochimica et biophysica acta*. 2014;1844(5):1026-33.
179. Sato Y, Harada K, Sasaki M, Nakanuma Y. Clinicopathological significance of S100 protein expression in cholangiocarcinoma. *Journal of gastroenterology and hepatology*. 2013;28(8):1422-9.
180. Nakanuma Y, Uchida T, Sato Y, Uesaka K. An S100P-positive biliary epithelial field is a preinvasive intraepithelial neoplasm in nodular-sclerosing cholangiocarcinoma. *Human pathology*. 2017;60:46-57.
181. Rose JB, Correa-Gallego C, Li Y, Nelson J, Alseidi A, Helton WS, et al. The Role of Biliary Carcinoembryonic Antigen-Related Cellular Adhesion Molecule 6 (CEACAM6) as a Biomarker in Cholangiocarcinoma. *PloS one*. 2016;11(3):e0150195-e.
182. Fernández Moro C, Fernandez-Woodbridge A, Alistair D'souza M, Zhang Q, Bozoky B, Kandaswamy SV, et al. Immunohistochemical Typing of Adenocarcinomas of the Pancreatobiliary System Improves Diagnosis and Prognostic Stratification. *PloS one*. 2016;11(11):e0166067-e.
183. Stroescu C, Herlea V, Dragnea A, Popescu I. The diagnostic value of cytokeratins and carcinoembryonic antigen immunostaining in differentiating hepatocellular carcinomas from intrahepatic cholangiocarcinomas. *Journal of gastrointestinal and liver diseases : JGLD*. 2006;15(1):9-14.
184. Shiota T, Ojima H, Hiraoka N, Shimada K, Rokutan H, Arai Y, et al. Heat shock protein 90 is a potential therapeutic target in cholangiocarcinoma. *Molecular cancer therapeutics*. 2015.
185. Won HS, Lee MA, Chung ES, Kim DG, You YK, Hong TH, et al. Comparison of thymidine phosphorylase expression and prognostic factors in gallbladder and bile duct cancer. *BMC cancer*. 2010;10:564.
186. Thanasai J, Limpiboon T, Jearanaikoon P, Sripan B, Pairojkul C, Tantimavanich S, et al. Effects of thymidine phosphorylase on tumor aggressiveness and 5-fluorouracil sensitivity in cholangiocarcinoma. *World journal of gastroenterology : WJG*. 2010;16(13):1631-8.

187. Zhang MX, Gan W, Jing CY, Zheng SS, Yi Y, Zhang J, et al. S100A11 promotes cell proliferation via P38/MAPK signaling pathway in intrahepatic cholangiocarcinoma. *Molecular carcinogenesis*. 2019;58(1):19-30.
188. Bertram S, Padden J, Kalsch J, Ahrens M, Pott L, Canbay A, et al. Novel immunohistochemical markers differentiate intrahepatic cholangiocarcinoma from benign bile duct lesions. *J Clin Pathol*. 2016;69(7):619-26.
189. Meissl K, Macho-Maschler S, Müller M, Strobl B. The good and the bad faces of STAT1 in solid tumours. *Cytokine*. 2017;89:12-20.
190. Thonsri U, Seubwai W, Warasawapati S, Sawanyawisuth K, Vaeteewoottacharn K, Boonmars T, et al. Overexpression of lactate dehydrogenase A in cholangiocarcinoma is correlated with poor prognosis. *Histology and histopathology*. 2017;32(5):503-10.
191. Yu Y, Liao M, Liu R, Chen J, Feng H, Fu Z. Overexpression of lactate dehydrogenase-A in human intrahepatic cholangiocarcinoma: its implication for treatment. *World journal of surgical oncology*. 2014;12:78.
192. Huang QX, Cui JY, Ma H, Jia XM, Huang FL, Jiang LX. Screening of potential biomarkers for cholangiocarcinoma by integrated analysis of microarray data sets. *Cancer Gene Therapy*. 2015;23:48.
193. Singrang N, Kittisenachai S, Roytrakul S, Svasti J, Kangsamaksin T. NOTCH1 regulates the viability of cholangiocarcinoma cells via 14-3-3 theta. *J Cell Commun Signal*. 2019;13(2):245-54.
194. Shen J, Wang W, Wu J, Feng B, Chen W, Wang M, et al. Comparative Proteomic Profiling of Human Bile Reveals SSP411 as a Novel Biomarker of Cholangiocarcinoma. *PloS one*. 2012;7(10):e47476.
195. Shuang ZY, Wu WC, Xu J, Lin G, Liu YC, Lao XM, et al. Transforming growth factor-beta1-induced epithelial-mesenchymal transition generates ALDH-positive cells with stem cell properties in cholangiocarcinoma. *Cancer letters*. 2014;354(2):320-8.
196. Loilome W, Kadsanit S, Muisook K, Yongvanit P, Namwat N, Techasen A, et al. Imbalanced adaptive responses associated with microsatellite instability in cholangiocarcinoma. *Oncology Letters*. 2017;13(2):639-46.
197. Chevet E, Fessart D, Delom F, Mulot A, Vojtesek B, Hrstka R, et al. Emerging roles for the pro-oncogenic anterior gradient-2 in cancer development. *Oncogene*. 2013;32(20):2499-509.
198. Miller G, Socci ND, Dhall D, D'Angelica M, DeMatteo RP, Allen PJ, et al. Genome wide analysis and clinical correlation of chromosomal and transcriptional mutations in cancers of the biliary tract. *Journal of experimental & clinical cancer research : CR*. 2009;28(1):62-.
199. Gu X, Li B, Jiang M, Fang M, Ji J, Wang A, et al. RNA sequencing reveals differentially expressed genes as potential diagnostic and prognostic indicators of gallbladder carcinoma. *Oncotarget*. 2015;6(24):20661-71.
200. Thuwajit C, Utispan K, Abiko Y, Jarngkaew K, Puapairoj A, Chau-In S, et al. Stromal fibroblast-derived periostin promotes cancer progression and serves as

- diagnostic and poor prognostic factors in cholangiocarcinoma. *Cancer Microenvironment*. 2009;2:S158.
201. Yu X, Zou Y, Li Q, Mao Y, Zhu H, Huang G, et al. Decorin-mediated inhibition of cholangiocarcinoma cell growth and migration and promotion of apoptosis are associated with E-cadherin in vitro. *Tumour biology : the journal of the International Society for Oncodevelopmental Biology and Medicine*. 2014;35(4):3103-12.
 202. van Roessel S, Soer EC, Daamen LA, van Dalen D, Fariña Sarasqueta A, Stommel MWJ, et al. Preoperative misdiagnosis of pancreatic and periampullary cancer in patients undergoing pancreatoduodenectomy: A multicentre retrospective cohort study. *European journal of surgical oncology : the journal of the European Society of Surgical Oncology and the British Association of Surgical Oncology*. 2021.
 203. Müller PC, Frey MC, Ruzza CM, Nickel F, Jost C, Gwerder C, et al. Neoadjuvant Chemotherapy in Pancreatic Cancer: An Appraisal of the Current High-Level Evidence. *Pharmacology*. 2021;106(3-4):143-53.
 204. Wiltberger G, Krenzien F, Benzing C, Atanasov G, Klein F, Hau HM, et al. Prognostic Accuracy of the Seventh Edition of the TNM Classification Compared with the Fifth and Sixth Edition for Distal Cholangiocarcinoma. *Ann Surg Oncol*. 2015;23(4):1320-6.
 205. Kiriya M, Ebata T, Aoba T, Kaneoka Y, Arai T, Shimizu Y, et al. Prognostic impact of lymph node metastasis in distal cholangiocarcinoma. *The British journal of surgery*. 2015;102(4):399-406.
 206. Chung YJ, Choi DW, Choi SH, Heo JS, Kim DH. Prognostic factors following surgical resection of distal bile duct cancer. *Journal of the Korean Surgical Society*. 2013;85(5):212-8.
 207. Pomianowska E, Westgaard A, Mathisen Ø, Clausen OPF, Gladhaug IP. Prognostic Relevance of Number and Ratio of Metastatic Lymph Nodes in Resected Pancreatic, Ampullary, and Distal Bile Duct Carcinomas. *Annals of surgical oncology*. 2012;20(1):233-41.
 208. Murakami Y, Uemura K, Hayashidani Y, Sudo T, Hashimoto Y, Ohge H, et al. Prognostic significance of lymph node metastasis and surgical margin status for distal cholangiocarcinoma. *Journal of surgical oncology*. 2007;95(3):207-12.
 209. You Y, Shin YC, Choi DW, Heo JS, Shin SH, Kim N, et al. Proposed Modification of Staging for Distal Cholangiocarcinoma Based on the Lymph Node Ratio Using Korean Multicenter Database. *Cancers (Basel)*. 2020;12(3):762.
 210. Jun S-Y, Sung Y-N, Lee JH, Park K-M, Lee Y-J, Hong S-M. Validation of the Eighth American Joint Committee on Cancer Staging System for Distal Bile Duct Carcinoma. *Cancer research and treatment*. 2019;51(1):98-111.
 211. Gentilini A, Pastore M, Marra F, Raggi C. The Role of Stroma in Cholangiocarcinoma: The Intriguing Interplay between Fibroblastic Component, Immune Cell Subsets and Tumor Epithelium. *Int J Mol Sci*. 2018;19(10).
 212. Micke P, Strell C, Mattsson J, Martín-Bernabé A, Brunnström H, Huvila J, et al. The prognostic impact of the tumour stroma fraction: A machine learning-based analysis in 16 human solid tumour types. *EBioMedicine*. 2021;65:103269.

213. Hosein AN, Brekken RA, Maitra A. Pancreatic cancer stroma: an update on therapeutic targeting strategies. *Nature reviews Gastroenterology & hepatology*. 2020;17(8):487-505.
214. Sleeman JP. The metastatic niche and stromal progression. *Cancer metastasis reviews*. 2012;31(3-4):429-40.
215. Pereira ER, Jones D, Jung K, Padera TP. The lymph node microenvironment and its role in the progression of metastatic cancer. *Semin Cell Dev Biol*. 2015;38:98-105.
216. Le Large TYS, Meijer LL, Paleckyte R, Boyd LNC, Kok B, Wurdinger T, et al. Combined Expression of Plasma Thrombospondin-2 and CA19-9 for Diagnosis of Pancreatic Cancer and Distal Cholangiocarcinoma: A Proteome Approach. *The oncologist*. 2020;25(4):e634-e43.
217. Udgata S, Takenaka N, Bamlet WR, Oberg AL, Yee SS, Carpenter EL, et al. THBS2/CA19-9 Detecting Pancreatic Ductal Adenocarcinoma at Diagnosis Underperforms in Prediagnostic Detection: Implications for Biomarker Advancement. *Cancer prevention research (Philadelphia, Pa)*. 2020.
218. Rifai N, Gillette MA, Carr SA. Protein biomarker discovery and validation: the long and uncertain path to clinical utility. *Nature biotechnology*. 2006;24(8):971-83.
219. Aran D, Camarda R, Odegaard J, Paik H, Oskotsky B, Krings G, et al. Comprehensive analysis of normal adjacent to tumor transcriptomes. *Nature communications*. 2017;8(1):1077-.
220. Hu X, Du S, Yu J, Yang X, Yang C, Zhou D, et al. Common housekeeping proteins are upregulated in colorectal adenocarcinoma and hepatocellular carcinoma, making the total protein a better "housekeeper". *Oncotarget*. 2016;7(41):66679-88.

Distal cholangiocarcinoma



Johannes Byrling, MD, has during his doctoral studies investigated aspects of the rare but devastating cancer distal cholangiocarcinoma - from novel bio-markers to clinical management and outcome.

Department of Surgery
Clinical Sciences, Lund

Lund University, Faculty of Medicine
Doctoral Dissertation Series 2021:122
ISBN 978-91-8021-129-1
ISSN 1652-8220