

LUND UNIVERSITY

Microvesicle signaling in cardiovascular biology under normal and pathobiological conditions

Bryl-Gorecka, Paulina

2021

Document Version: Publisher's PDF, also known as Version of record

Link to publication

Citation for published version (APA): Bryl-Gorecka, P. (2021). *Microvesicle signaling in cardiovascular biology under normal and pathobiological conditions*. Lund University, Faculty of Medicine.

Total number of authors: 1

General rights

Unless other specific re-use rights are stated the following general rights apply:

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights. • Users may download and print one copy of any publication from the public portal for the purpose of private study

or research.

- You may not further distribute the material or use it for any profit-making activity or commercial gain
 You may freely distribute the URL identifying the publication in the public portal

Read more about Creative commons licenses: https://creativecommons.org/licenses/

Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

LUND UNIVERSITY

PO Box 117 221 00 Lund +46 46-222 00 00

Microvesicle signaling in cardiovascular biology under normal and pathobiological conditions

PAULINA BRYL-GÓRECKA DEPARTMENT OF CARDIOLOGY | FACULTY OF MEDICINE | LUND UNIVERSITY



Microvesicle signaling in cardiovascular biology under normal and pathobiological conditions

Paulina Bryl-Górecka



DOCTORAL DISSERTATION

by due permission of the Faculty of Medicine, Lund University, Sweden. To be defended at Segerfalksalen, BMC A10, Sölvegatan 19, Lund University on 2nd of September 2021 at 09:00.

Faculty opponent

Professor Christina Christersson, MD, PhD, FESC Department of Medical Sciences, Cardiology, Uppsala University, Sweden

Organization LUND UNIVERSITY		Document name Doctoral [Dissertation	
		Date of issue September 2	, 2021	
Author(s) Paulina Bryl-Górecka		Sponsoring organization		
Title and subtitle Microvesicle signaling in cardiovascular biology under normal and pathobiological conditions				
Abstract Cardiovascular disease (CVD) is one directly related to the circulatory s Atherosclerotic plaque build-up can le To prevent or slow down of the CV connected with lifestyle and nutrition. Microvesicles (MVs) belong to a large bodies. These small (<1 µm) vesicle process involving numerous signalir vascular endothelial cells. The release bioactive molecules both on their surd discovery and potential diagnostic a cardiovascular-related pathologies. To analyse plasma MVs, we optimize wave, that isolates vesicles in a no to standard protocols of MV enrichme the results of a clinical study reveale a release of MVs from activated endo vesicular cargo. Bilberries (<i>Vaccinium myrtillus</i>) are of the BEARSMART randomized clinica on circulating MVs in MI patients. A platelet and endothelial WV concentre decreased endothelial vesiculation, w project showed for the first time that r influence of berries on vascular healt In the PROFLOW clinical study, wa a parameter depicting blood flow in t CVD. Proteomic profiling demonstrate of this clinical study pointed at a pote Lastly, we showed that atherosclerof that the vesicles originated from sev represented the pathological process We also demonstrated an advantage The outcome of the studies included Study I and II underlined the importal whereas the Western world is still cha on potential application of MVs as bio and possibly, personalized approach.	of the lead ystem, c ead to a l VD progr r group, c es are re- ng pathw. e is active face and pplication ed an acco- on-contac- ent and c d that ex- sthelium. considere- al study w fiter eight tic plaque yeral type- es within of analys- in this the nce of ex- racterize- markers of cular dise	ding causes of death worldwid levelops due to an interplay reduction in arterial blood flow ession, it is important to prir alled extracellular vesicles that leased from the cell by blebt ays. Circulating MVs originati ted by pro-inflammatory and p inside the lumen that makes the n. The overall aim of this the ustic trapping machine based at manner. We demonstrated ould potentially be used for ve- ercise has a protective effect of The proteomic data showed the d to be a beneficial dietary co- e explored if there was an effec- s weeks of dietary intervention reover, the <i>in vitro</i> part of the s is related to the P2X ₇ purinergi- changes can directly affect MV ed a negative relationship be and levels of endothelial and connection for CFR and seve- ication of MVs, for diagnosis of a released MVs, following ball us of cells and exhibited a pro- the cardiovascular system th sing isolated EVs, compared to essis points at the important rol- ercise and nutrition for preven- d with physical inactivity and in or targets of therapeutics, howe	le. Atherosclerosis, a chronic pathology / between several molecular events. r and finally, myocardial infarction (MI). marily address modifiable risk factors t also includes exosomes and apoptotic oing of plasma membrane, a complex e mainly from blood cells, as well as ro-coagulant conditions. MVs transport hem an interesting target for biomarker sis was to explore the role of MVs in method, employing ultrasonic standing that the new method is comparable scicle-based diagnostics . Furthermore, on the vasculature through decreasing hat exercise changes the pattern of the exponent for patients with CVD and in ect of bilberry powder supplementation n, there was a significant reduction in tudy demonstrated that bilberry extract c receptor pathway. The results of this / release and underlined the protective etween coronary flow reserve (CFR), platet MVs, circulating in patients with tral vesicular biomarkers. The outcome f vascular dysfunction. loon angioplasty. The results revealed bo-atherogenic 'pattern' of proteins that at can lead to the atheroma formation. o crude plasma samples. e of MVs in the cardiovascular system. tion of CVD through decrease in MVs, habalanced diet. Study III and IV focused ever it needs further, thorough research kers, Acoustic trapping	
Supplementary bibliographical information		Language English		
ISSN 1652-8220 Lund University, Faculty of Medic		ledicine Doctoral	ISBN 978-91-8021-081-2	
Recipient's notes	Numbe	er of pages 85	Price	
Security classific		y classification	1	
	•			

I, the undersigned, being the copyright owner of the abstract of the above-mentioned dissertation, hereby grant to all reference sources permission to publish and disseminate the abstract of the above-mentioned dissertation.

Signature Paulino Byr - Govedne Date 2021-07-26

Microvesicle signaling in cardiovascular biology under normal and pathobiological conditions

Paulina Bryl-Górecka



Coverphoto by Paulina Bryl-Górecka, Piotr Górecki & Henryk Górecki

Copyright pp 1-85 (Paulina Bryl-Górecka)

Paper 1 $\ensuremath{\mathbb{C}}$ by the Authors (Published open access, The Royal Society of Chemistry)

Paper 2 © by the Authors (Published open access, Wiley-VCH GmbH)

Paper 3 $\ensuremath{\mathbb{C}}$ by the Authors (Published open access, The American Physiological Society)

Paper 4 © by the Authors (Manuscript unpublished)

Faculty of Medicine Department of Cardiology

ISBN 978-91-8021-081-2 ISSN 1652-8220

Printed in Sweden by Media-Tryck, Lund University Lund 2021



Media-Tryck is a Nordic Swan Ecolabel certified provider of printed material. Read more about our environmental work at www.mediatryck.lu.se MADE IN SWEDEN 'Education is the most powerful weapon, which you can use to change the world' Nelson Mandela

'For a man to conquer himself is the first and noblest of all victories' Plato

To my Family

Table of Contents

List of Papers & Manuscripts	8
Popular Summary	9
Populär Sammanfattning	11
Podsumowanie popularnonaukowe	13
Introduction	15
Cardiovascular disease	15
Endothelial dysfunction and atherosclerosis Atheroma composition	15 17
Effects of nutrition on cardiovascular health Nutrition for prevention and treatment of CVD Bilberries for cardiovascular health	18 18 18
Effects of exercise on cardiovascular health	19
Microvesicles Definition Role in development of atherosclerosis Methods of microvesicle isolation and detection	20 20 21 22
Materials and Methods	25
Acoustic trapping (Papers I-IV)	25
High-speed centrifugation (Paper I)	26
Flow cytometry (Papers I-IV)	26
Proximity Extension Assay (Papers I, III, IV)	27
Transmission Electron Microscopy (Paper I)	28
Nanoparticle Tracking Analysis (Papers I and II)	28
Endothelial vascular cell culture (Paper II)	28
Calcein assay (Paper II)	29
Compound effect on endothelial cells (Paper II)	29
RNA extraction, reverse transcription and quantitative PCR (Paper II)	30
Western Blot (Paper II)	31
Coronary flow reserve measurement (Paper III)	31
Statistics (Papers I-IV)	32

Aims and Hypotheses	
Results and Discussion	
Paper I	
Paper II	
Paper III	45
Paper IV	53
Summary and Perspectives	61
Acknowledgements	63
Podziękowania	65
References	67

List of Papers & Manuscripts

- I. Paulina Bryl-Górecka, Ramasri Sathanoori, Mariam Al-Mashat, Björn Olde, Jonas Jögi, Mikael Evander, Thomas Laurell and David Erlinge. Effect of exercise on the plasma vesicular proteome: A methodological study comparing acoustic trapping and centrifugation. Lab on a Chip 2018, 18(20): 3101-3111; doi: 10.1039/c8lc00686e.
- II. Paulina Bryl-Górecka, Ramasri Sathanoori, Lilith Arevström, Rikard Landberg, Cecilia Bergh, Mikael Evander, Björn Olde, Thomas Laurell, Ole Fröbert and David Erlinge. *Bilberry supplementation after myocardial infarction decreases microvesicles in blood and affects endothelial vesiculation*. Mol Nutr Food Res 2020, 64(20):e2000108; doi: 10.1002/mnfr.202000108.
- III. Paulina Bryl-Górecka, Kreema James, Kristina Torngren, Inger Haraldsson, Li-Ming Gan, Sara Svedlund, Björn Olde, Thomas Laurell, Elmir Omerovic and David Erlinge. *Microvesicles in plasma reflect coronary flow reserve in patients with cardiovascular disease*. Am J Physiol Heart Circ Physiol 2021; doi: 10.1152/ajpheart.00869.2020.
- IV. **Paulina Bryl-Górecka**, Kreema James, Björn Olde, Thomas Laurell, David Erlinge. *Characterization of extracellular vesicles released from atherosclerotic plaque following balloon angioplasty*. Manuscript unpublished.

Popular Summary

Cardiovascular disease (CVD) is one of the leading causes of death worldwide. Major risk factors are both lifestyle-related, as well as non-modifiable. Atherosclerosis is a chronic pathology of the arteries that is demonstrated as a plaque built-up within the vessel. Such deposition can lead to a reduction in blood flow and finally, myocardial infarction. Although many types of anti-atherogenic medications have been already developed, it is important to also address modifiable risk factors connected with lifestyle and nutrition, to prevent or slow down the CVD progression.

Microvesicles (MVs) are a type of so called extracellular vesicles that originate from different types of cells and are released to the environment. MVs found in the circulation are predominantly released from blood cells or endothelial cell layer in the vessels, and are mainly stimulated by pathological, pro-inflammatory conditions. MVs carry many different classes of molecules and thus are a very interesting source of potential biomarkers – indicators of a disease. To analyse circulating MVs, we optimized a new method for MV isolation from plasma, called acoustic trapping. It is a new technology that employs acoustic standing wave that 'traps' vesicles.

In Paper I we showed that acoustic trapping is comparable to standard protocols and could potentially be used for vesicle-based diagnostics. The results of a clinical study revealed that exercise has a beneficial effect on the vasculature through decreasing a release of MVs from endothelium. The analysis also indicated that physical activity changes the protein content of the vesicles.

Bilberries, or Nordic blueberries, are considered beneficial for patients with CVD. Thus, in Paper II we investigated if there was an effect of bilberry powder supplementation on circulating MVs in myocardial infarction patients. Eight weeks of dietary intervention significantly decreased the concentration of endothelial and platelet-derived MVs. *In vitro* culturing of endothelial cells demonstrated that bilberry extract decreased vesicle release that was related to the P2X₇ receptor. This project underlines that nutrional changes can affect MV release and confirm the protective impact of berries on cardiovascular health.

In Paper III, we observed a negative correlation between coronary flow reserve, a parameter depicting blood flow in the heart, and levels of endothelial and platet MVs, circulating in patients with CVD. There was a similar connection for CFR and

several proteins, transported in MVs. The outcome of this clinical study points at a potential application of MVs for diagnosis of vascular dysfunction.

In paper IV we showed that atherosclerotic plaque releases MVs, following balloon angioplasty that originated from several types of cells. The vesicles exhibited a pathological 'pattern' of proteins that represent the processes within the cardiovascular system that lead to the formation of atheroma.

In summary, the outcome of the studies included in this thesis points at the important role of MVs in the cardiovascular system and their potential application as biomarkers.

Populär Sammanfattning

Kardiovaskulär sjukdom (CVD) är en av de vanligaste dödsorsakerna i världen. De största riskfaktorerna är både livsstilsrelaterade och kroniska. Åderförkalkning är en kronisk patologi i artärerna som kännetecknas av plackbildning i blodkärlen. Denna förändring kan i sin tur leda till en minskning av blodflödet och slutligen till hjärtinfarkt. Även om många typer av anti-aterogena läkemedel redan har utvecklats, så är det viktigt att även ta itu med riskfaktorer kopplade till livsstil och näring för att förhindra eller sakta ner progressionen av CVD.

Mikrovesiklar (MV) är extracellulära vesiklar som frigörs från celler till omgivningen genom avknoppning från plasmamembranet. MV som finns i cirkulationen frigörs huvudsakligen från blodkroppar eller från kärlens endotelcellskikt vilket i sin tur kan stimuleras av patologiska, proinflammatoriska tillstånd. MV innehåller många olika typer av molekyler och är därför en mycket intressant källa till potentiella biomarkörer – dvs indikatorer på en sjukdom. För att analysera cirkulerande MV:er optimerade vi en ny metod för MV-isolering från plasma, kallad "acoustic trapping". Det är en ny teknik som använder en akustisk stående våg som "fångar" mikrovesiklar.

I Arbete I visade vi att "acoustic trapping" är jämförbar med standardmetoder och potentiellt kan användas för vesikelbaserad diagnostik. Resultaten av en klinisk studie visade att motion har en gynnsam effekt på kärlsystemet genom att minska frisättningen av MV från endotelet. Analysen visade också att fysisk aktivitet förändrar proteininnehållet i vesiklarna.

Blåbär anses vara fördelaktiga för patienter med CVD. Således undersökte vi i Arbete II om ett diet tillägg av blåbärspulver har effekt på cirkulerande MV hos hjärtinfarktpatienter. Vi fann att åtta veckors dietintervention signifikant minskade koncentrationen av MV från endotel och trombocyter. Blåbärsextrakt minskade frisättningen av vesiklar från odlade endotelceller. Detta projekt visar att näringsförändringar kan påverka MV-frisättning och bekräftar den skyddande effekten av blåbär på hjärt-kärlhälsan.

I Arbete III observerade vi en negativ korrelation mellan koronarflödesreserv, en parameter som visar blodflödet i hjärtat och nivåer av endotel- och trombocyt-MV, hos patienter med CVD. Det fanns liknande kopplingar mellan CFR och flera olika vesikel-proteiner. Resultatet av denna kliniska studie pekar på en potentiell tillämpning av MV för diagnos av vaskulär dysfunktion.

I Arbete IV visade vi att ballongangioplastik orsakar att aterosklerotiska plack frisätter MV, och att dessa härstammar från flera typer av celler. Vesiklarna uppvisade ett patologiskt "mönster" av proteiner som alla representerar processer som leder till bildandet av aterom.

Sammanfattningsvis pekar resultaten, som ingår i denna avhandling, på MV:s viktiga roll i det kardiovaskulära systemet och deras potentiella tillämpning som biomarkörer.

Podsumowanie popularnonaukowe

Choroby układu sercowo-naczyniowego są jedną z głównych przyczyn zgonów na świecie. Czynniki ryzyka tej grupy chorób są zarówno związane ze stylem życia, jak i niemodyfikowalne. Miażdżyca to choroba zwyrodnieniowa tętnic, polegająca na wytworzeniu blaszki miażdżycowej wewnątrz naczynia krwionośnego. Proces ten może doprowadzić do zmniejszenia przepływu krwi, a w skrajnych przypadkach – zawału mięśnia sercowego. Pomimo istnienia wielu typów leków przeciwmiażdżycowych, należy zadbać przede wszystkim o modyfikowalne czynniki ryzyka, związane ze stylem życia i odżywianiem, w celu zapobiegania i spowolnienia rozwoju chorób układu krążenia.

pecherzyków Mikropecherzyki to rodzaj tzw. zewnatrzkomórkowych, wytwarzanych z błony komórkowej, które są uwalniane przez komórki i wysyłane do otoczenia. Proces ten ma miejsce głównie w patologicznych warunkach prozapalnych. Mikropecherzyki krażące we krwi pochodzą przede wszystkim z płytek krwi, leukocytów i erytrocytów, a także śródbłonka, czyli warstwy komórek wyściełających naczynia krwionośne. Mikropęcherzyki transportują wiele różnych rodzajów związków chemicznych i z tego względu są interesującym źródłem potencjalnych biomarkerów - molekularnych wskaźników choroby. Aby zbadać mikropęcherzyki krążące we krwi, opracowaliśmy nową metodę o nazwie "pułapkowanie akustyczne", pozwalającą na ich pozyskanie z osocza. Technologia ta wykorzystuje tzw. stojaca fale akustyczna, która umożliwia "złapanie" i oczyszczenie pęcherzyków.

W publikacji nr 1 pokazaliśmy, że pułapkowanie akustyczne osiąga porównywalne wyniki, jak standardowe protokoły do izolacji mikropęcherzyków i w przyszłości mogłaby być potencjalnie użyta w celach diagnostycznych. Rezultaty testu klinicznego dowiodły, że aktywność fizyczna ma pozytywny wpływ na naczynia krwionośne poprzez zmniejszanie wydzielania mikropęcherzyków przez śródbłonek. Wyniki analizy wskazały również na wpływ ćwiczeń na zmiany w zawartości białek, obecnych w pęcherzykach.

Działanie borówki czarnej, popularnie zwanej jagodą, uznawane jest za korzystne w przypadku chorób sercowo-naczyniowych. Z tego względu, w publikacji nr 2 badaliśmy wpływ suplementacji proszkiem z owoców tej rośliny na mikropęcherzyki we krwi pacjentów po przebytym zawale serca. Ośmiotygodniowa interwencja dietetyczna znacząco zmniejszyła poziom mikropęcherzyków pochodzących ze śródbłonka oraz płytek krwi. Badania *in vitro* na wyizolowanych komórkach śródbłonka ukazały, że ekstrakt z borówki czarnej zmniejszył produkcję pęcherzyków zewnątrzkomórkowych, co było powiązane z receptorem P2X₇. Wyniki tego projektu podkreślają wpływ diety na uwalnianie mikropęcherzyków i potwierdzają pozytywny efekt jagód na układ krążenia.

W publikacji nr 3 zaobserwowaliśmy odwrotną korelację między tzw. rezerwą wieńcową, parametrem obrazującym przepływ w sercu, a poziomami mikropęcherzyków pochodzących z płytek krwi i śródbłonka u pacjentów ze zdiagnozowaną chorobą układu krążenia. Podobny związek miał miejsce również w przypadku rezerwy wieńcowej oraz kilkoma białkami, transportowanymi przez mikropęcherzyki. Rezultaty tego badania wskazują na potencjalne zastosowanie mikropęcherzyków w celu diagnozy dysfunkcji śródbłonka.

W publikacji nr 4 ukazaliśmy, że po zabiegu angioplastyki tętnic, płytka miażdżycowa uwalniała mikropęcherzyki, pochodzące z rożnych rodzajów komórek. Pęcherzyki z blaszki miażdżycowej charakteryzowały się również patologicznym wzorcem białek, co oddaje procesy toczące się w naczyniach krwionośnych, które doprowadzają do rozwoju miażdżycy.

Podsumowując, wyniki naszych badań wskazują na ważną rolę mikropęcherzyków w funkcjonowaniu układu sercowo-naczyniowego i ich potencjalne zastosowanie jako biomarkerów.

Introduction

Cardiovascular disease

Cardiovascular disease (CVD) is a term grouping diseases related to circulatory system, mainly blood vessels and the heart. CVD is a serious global burden, as it accounts for approximately one third of deaths worldwide.¹ Major risk factors comprise smoking, obesity, genetics, hypertension, diabetes, high cholesterol levels, physical inactivity, alcohol consumption and air pollution. In the past years, mainly due to an increase in CVD treatment options, a reduction in morbidity and mortality has been observed. Examples of applied medications include statins, beta-blockers, angiotensin converting enzyme inhibitors, angiotensin receptor blockers, warfarin, platelet aggregation-regulating compounds such as aspirin, clopidogrel or newer: prasugrel and ticagrelor.¹ However, modifiable risk factors should be primarily addressed by modification of lifestyle (e.g. regular exercise) and diet.

One of the diseases belonging to the CVD group is coronary artery disease (CAD), also known as ischemic heart disease.² It is connected with accumulation of atherosclerotic plaque in the heart arteries that leads to a decrease in the blood flow. CAD includes stable and unstable angina, cardiac arrest and finally, myocardial infarction or heart attack (Fig. 1), characterized by heart tissue damage due to very low oxygen levels and release of troponin into the circulation. Other types of CVD include e.g. heart failure, stroke, cardiomyopathy, hypertensive heart disease and aortic aneurysm. While not all CVD types are connected with atherosclerotic plaques, CAD is directly related to this vascular pathology.

Endothelial dysfunction and atherosclerosis

Endothelial dysfunction is the first step towards development and progression of atherosclerosis and further CAD. It involves several molecular events, such as decrease in nitric oxide (NO) production, release of pro-inflammatory and pro-thrombotic molecules, increased platelet aggregation that lead to a decline in vascular relaxation and subsequent reduction in blood flow in coronary arteries.^{3,4}



Fig. 1 Types of cardiovascular diseases, based on Olvera Lopez et al.² Abbreviations: STEMI – ST elevation myocardial infarction, NSTEMI – non-ST elevation myocardial infarction.

Transition from endothelial dysfunction to a chronic disease called atherosclerosis is caused by interplay between several molecules that switch the vasculature to increasingly pathological status. Interestingly, atherosclerosis is sometimes referred to as 'inflammatory disease', reflecting the complexity of this process.⁵ The occurring molecular events include e.g. high levels of LDL in blood, accumulation of oxidized lipids in the vascular wall, leukocyte adhesion to the endothelial layer, proliferation of vascular smooth muscle cells (SMCs), extracellular matrix deposition, as well as calcium accumulation that finally lead to atherosclerotic vessel, decrease formation in the of the vessel plaque lumen and deterioration of blood flow (Fig. 2).⁶ Substantial blood flow limitation in the arteries is manifested as arterial stenosis that is an advanced stadium of the disease.^{4,7}



Fig. 2 Schematic representation on how the presence of atherosclerotic plaque decreases arterial blood flow.

Atheroma composition

Atherosclerotic plaque, or atheroma, has a diverse composition that is generally divided into the following sections: a necrotic core, containing macrophages and cholesterol crystals, as well as a fibrous cap, consisting of extracellular matrix, SMCs and leukocytes (Fig. 3). The lesions are considered as vulnerable, when the cap is thin or stable, when the cap is thick due to abundant presence of SMCs.⁸

In severe cases, such as arterial occlusion and MI, the plaque is ruptured that results in an exposure to the bloodstream of prothrombotic material, derived from the atheroma and further, formation of a thrombus (Fig. 3).⁸



Fig. 3 Schematic representation of atheroma composition and progression.

A standard method to increase the lumen of the narrowed vessel and improve the blood flow is balloon angioplasty.⁹ The procedure applies a catheter with a deflated balloon that is placed in the artery and moved to the respective site of the plaque. Once the balloon reaches the atheroma, it becomes inflated and compresses the plaque, thus increasing blood flow (Fig. 4).



Fig. 4 Schematic representation of balloon angioplasty procedure.

Effects of nutrition on cardiovascular health

Nutrition for prevention and treatment of CVD

Diet is one of the essential lifestyle-related factors, profoundly affecting human health. Proper nutrition is a weapon combating not only excessive body weight, but also preventing from several pathologies, especially diseases of affluence, including CVDs. Current recommendations from the American Heart Association for a healthy, heart-friendly diet include limiting intake of sweets, sodium, alcohol and animal-derived products rich in saturated fats, e.g. red meat and high-fat dairy; incorporation of substantial amounts of vegetables, fruits, whole grains, nuts, low fat dairy, fish and plant oils rich in polyunsaturated fatty acids (PUFAs).¹⁰ Some of the officially recommended approaches that fulfil these criteria are Dietary Approach to stop Hypertension (DASH),^{11,12} plant-based ^{13,14} and Mediterranean diets.^{15,16} There is also a growing interest in nutritional supplementation for CVD prevention and additional treatment. For example, intake of standardized anti-inflammatory Omega-3 PUFAs, found naturally in e.g. fatty fish, flax seeds and canola oil, has recently gained attention as potential beneficial anti-CVD compound, although the results of meta-analyses have not been consistent so far.¹⁷⁻²¹

Bilberries for cardiovascular health

It has been reported that diet rich in berries exhibits protective effects on the cardiovascular system.^{22,23} Majority of these beneficial properties are attributed to a high content of polyphenolic pigments called anthocyanins. The mechanism of action on CVD-related pathologies is based on e.g. decreasing local oxidative

stress,²⁴⁻²⁶ improving NO production²⁷ or inflammation inhibition.²⁸ Summary of favourable influence of bilberry consumption is presented on Fig. 5. As bilberry (*Vaccinium myrtillus*) is a widely consumed berry species, its incorporation in the diet as a component or a supplement, appears to be a promising way of improving the status of CVD patients.



Fig. 5 Proposed benefits of bilberry dietary supplementation (upper panel), based on the current clinical and *in vivo* studies and structure of anthocyanins (lower panel), plant pigments connected with beneficial effects evoked by the bilberry intake. Based on Cutler et al.²⁹

Effects of exercise on cardiovascular health

Physical activity is another factor, connected to lifestyle that influences health status. It is a tool to fight excess kilograms, however it is also crucial for maintaining healthy cardiovascular system, as decreased muscle mass and strength are known CVD risk factors. World Health Organisation recommends at least 150 min of moderate-to-vigorous physical activity each week or 20 min daily for adults.³⁰ Interestingly, a positive dose-response effect of the amount of exercise and vascular status has been observed.³¹ It is important to note that in the Western world a sedentary lifestyle prevails and a so-called sarcopenic obesity, a state of simultaneous obesity with low muscle strength due to lack of training, is very common. Thus, an active lifestyle is a cornerstone of CVD prevention and treatment, especially in the elderly population.^{30,32}

Regular training stimulates anti-atherosclerotic changes in endothelium, decreases blood pressure, improves insulin sensitivity, vasodilation and coronary flow reserve; is cardioprotective, anti-arrhythmic and anti-thrombotic.^{30,32,33} Furthermore, it is beneficial for cardiac regeneration via stimulation

of angiogenesis. An important player in promoting vascular health is muscle tissue that releases myokines and other factors into the circulation, thus influencing the whole circulatory system.³⁰ These muscle-derived 'messengers' have gained more attention in the past decade and the knowledge in this field is still expanding.

Microvesicles

Definition

Microvesicles (MVs), sometimes referred to as microparticles or ectosomes, are a subtype of a larger group, called extracellular vesicles (EVs) that also comprises exosomes and apoptotic bodies.³⁴ This division is based on size or biogenesis process (Fig. 6). MV size ranges between 30 - 1000 nm and this type of EVs is produced by shedding of plasma membrane. The membrane blebbing involves numerous intracellular and membrane-bound proteins, as well as activation of different signalling pathways that depend on the cell type. Similar to MVs, apoptotic bodies are membrane-derived vesicles, however their size varies between 1 to 5 μ m and the release occurs during apoptosis. Exosomes are the smallest vesicles, less than 100 nm in diameter that are released from the cell in a complex and controlled process of exocytosis, involving several transport-associated proteins and cytoskeletal remodelling.



Fig. 6 Subtypes of extracellular vesicles, based on the size and biogenesis process.

Cellular origin of plasma microvesicles

The first plasma MVs were discovered by British physician Peter Wolf that specialised in blood coagulation.³⁵ In 1967 he published an article 'The nature and significance of platelet products in human plasma' that described the presence of so called 'platelet dust' and its procoagulant activity. From then on, the research about plasma MVs has developed and the knowledge about these small, but very important cellular particles has been substantially broadened.

The majority of plasma MVs stem from cellular components circulating in blood, such as platelets, erythrocytes and leukocytes.^{34,36} Furthermore, a very important source of plasma MVs are vascular endothelial cells (ECs), especially in the context of endothelial dysfunction and atherosclerosis progression. Pro-inflammatory and pro-coagulant conditions trigger EC activation and a release of MVs, presenting E-selectin (CD62E) on their surface.³⁷ Similarly, proatherosclerotic conditions also lead to shedding of platelet MVs.³⁸

In vivo experiments in rodents demonstrated physiological removal of circulating MVs by organs such as liver and spleen. The exact mechanism of MV clearance is yet unknown, however the available data proposes lipid- and protein-directed phagocytosis that depends on the recipient cell, as well as the donor cell of MVs.³⁹

As MVs are membrane-derived vesicles, they carry biologically active molecules on their surface,^{40,41} such as receptors and phospholipids, originating from the mother cell. MVs also transport a molecular cargo in their lumen: proteins,⁴²⁻⁴⁵ miRNAs⁴⁶ and long non-coding RNAs (Fig. 6).⁴⁷ This enrichment of different classes of particles, present both inside and on the membrane, underlines the value of MVs in mechanistic studies, cardiovascular biomarker discovery and potential diagnostic application in the future.^{48,49}

Role in development of atherosclerosis

Low levels of MVs are also detected in blood of healthy people that suggests an existence of vesicle release in physiological conditions. Nevertheless, there are strong indications that elevated levels of circulating MVs are the hallmarks of vascular dysfunction and CVD.⁵⁰ Endothelial and platelet MVs have been associated with a range of cardiovascular system pathologies, such as hypertension, CAD and MI.^{34,36,51-54} Currently available data shows that MVs can affect cardiovascular system via many molecular mechanisms. For example, activated ECs release MVs, augmenting platelet aggregation *ex vivo* that occurs through a von Willebrand factor (vWF)-dependent crosslinking.⁵⁵ MVs can further facilitate pro-atherogenic processes by e.g. activating clotting cascade, promoting monocyte-endothelium interactions and regulating endothelial and vascular SMC proliferation.^{56,57} As MVs are a rich source of potential biomarkers and are involved in vascular pathologies, the projects included in this thesis have mainly been focused on vesicles of the MV subtype.

Methods of microvesicle isolation and detection

Characterization of circulating MVs in blood and so-called 'liquid biopsies' provide a novel approach in both pre-clinical research, as well as clinical diagnostics. However, MV isolation from patient plasma remains technically challenging. Most of the currently used isolation techniques (e.g. ultracentrifugation, iodixanol/sucrose gradient centrifugation, immunoaffinity beads) were first developed to isolate EVs, especially exosomes from culture medium used in the *in vitro* experiments. It is, however, possible to apply them for MV isolation from human plasma.⁵⁸ Nevertheless, these methods exhibit several disadvantages, especially with regards to limited-volume biobank samples, as they require substantial sample volumes^{59,60} and can lead to loss of vesicles and EV integrity.⁶¹⁻⁶³ Thus, the traditional methods are challenging to use in the cardiovascular-related diagnostic context.

Traditionally, high-speed centrifugation has been used for plasma MV isolation. Compared to ultracentrifugation that is usually used for culture medium, this technique uses much lower than 100.000 x g rotor speed, ranging between 13.000 and 20.000 x g for flow cytometric or e.g. proteomic/miRNA analyses (Fig. 7). One of the aims of this thesis was to optimize a novel method for MV enrichment from plasma, called acoustic trapping that is a non-contact method, based on a microscale technology. The method applies acoustic standing wave instead of g force, thus preventing vesicle disruption and plasma aggregation. Furthermore, the volume of plasma needed for a successful MV isolation is very low, reaching 50 for proteomic analysis. Another u1 per sample e.g. advantage of acoustic trapping is no need for staining before the isolation, so that the whole range of different MVs is isolated. A detailed technical description of the technique is given in the 'Materials and Methods' section and Article I. Both high-speed centrifugation and acoustic trapping are used for MV isolation from frozen patient samples, thus it is vital to ensure a proper handling of the samples and removal of blood cells prior freezing.⁶¹ As numerous novel microscale approaches for EV isolation are currently appearing and different lab-on-a-chip devices are being developed for liquid biopsies,⁶⁴⁻⁶⁷ several interesting discoveries in the field should be expected in the future.

Flow cytometry is a widely-applied method for measuring concentration and analysis of cellular origin of plasma MVs.⁶⁸ It is based on a staining of MVs with fluorescently-labelled antibodies, directed against proteins, present on the outer membrane of the vesicles.⁶⁹ Most of the currently available research on MVs has been done on conventional flow cytometers that usually enable measuring vesicles not smaller than 500-200 nm.⁷⁰ However, new branches, called high-sensitive (sometimes also referred to as micro or nano) flow cytometry⁷¹⁻⁷³ and imaging flow

cytometry^{74,75} have recently appeared and are developing rapidly. This thesis presents data obtained both on a conventional (Study I. II and III), as well as a micro (Study III and IV) flow cytometer.

Other methods for MV detection and enumeration that allow to measure other physical parameters of the vesicles, involve more or less-known techniques such as electron and atomic force microscopy, nanoparticle tracking analysis, zeta potential analysis, dynamic light scattering and tunable resistive pulse sensing (Fig. 8). It is important to note that all above-mentioned techniques have strengths and weaknesses, especially with regards to potential application in clinical diagnostics and the best approaches for MV isolation and analyses are yet to be determined.



Fig. 7 Different methods of microvesicle isolation from plasma, presented in this thesis.



Fig. 8 Examples of methods used for extracellular vesicle analysis, with given measured physicochemical properties.^{68,75-82}

Materials and Methods

Acoustic trapping (Papers I-IV)

As mentioned in the Introduction, acoustic trapping is a novel method for MV enrichment from plasma samples. The platform (AcouTrap, AcouSort AB) consists of a robotic unit, suitable for use of 96-well plates, a glass capillary, a transducer and pumps used for sample aspiration and washing steps (Fig. 9). Briefly, the procedure involves 'seed particle' aspiration and washing, plasma sample aspiration and MV trapping within the seed particle cluster, washing and elution of the MV suspension.



Fig. 9 Acoustic trapping platform at the Molecular Cardiology laboratory.

A rectangular glass capillary was used and a 4 MHz PZT transducer was attached through a thin layer of glycerol (Paper I and II) or glue (Paper III and IV) to the outside of the capillary. Both glycerol and glue ensure uniform acoustic coupling over time, without evaporation. The transducer generates a local acoustic standing wave in the capillary that enables particles to be retained or 'trapped' in a noncontact way. Because primary acoustic radiation force is dependent on the particle volume, smaller particles (below 2 μ m), e.g. MVs, cannot be directly retained in the acoustic trap, as the force is not sufficient for particle focusing. Larger particles, however, can be retained against the flow, thus 12 μ m polystyrene beads were used as 'seed particles'. After aspiration of the diluted plasma sample, the scattered sound field between the seed particles and MVs present in the sample induces enrichment of MVs within the seed particle cluster. The cluster is further washed and subsequently released in a defined volume for further analyses, e.g. proteomic or miRNA profiling. For detailed graphical explanation of acoustic trapping working principles, please refer to the experimental section in Paper I.

High-speed centrifugation (Paper I)

To compare acoustic trapping with standard isolation protocol for flow cytometric analysis of MVs, plasma samples were centrifuged at 1600 x g for 15 min at RT, followed by 13 000 x g for 2 min. 80% of the supernatant was aspirated and diluted with DPBS for subsequent staining with fluorochrome-conjugated antibodies (Abs). For proteomic profiling, patient samples were centrifuged at 1600 x g for 15 min at and further RT 100 μl of plasma was centrifuged at 20 000 x g for 60 min at RT, resulting in two fractions: MV-rich pellet and MVfree supernatant. Detailed explanation of the workflow and method comparison is included in Paper I.

Flow cytometry (Papers I-IV)

Flow cytometry (FCM) is a method that uses optical characteristics of the cells or vesicles for real-time visualisation and enumeration. MVs stained with fluorescently labelled antibodies are injected into the cytometer, positioned in a central fluid stream (hydrodynamic focusing), illuminated with a laser beam and further detected.

In Papers I-III Accuri C6 flow cytometer (BD Accuri) was used for enumerating platelet and endothelial MVs isolated with acoustic trapping or high-speed centrifugation. Polystyrene size standard beads $(0.1-3 \mu m)$ were used to establish a flow cytometric 'MV gate', corresponding to the size of MVs. 100 µl of MV suspension was incubated with PE-conjugated anti-CD62E (E-selectin) or CD42a (glycoprotein IX) Abs for 30 min at RT. 20 000 events were collected in the MV gate and the obtained data is shown as the number of fluorescently labelled MVs/µl. Fluorescence-based gates were established using respective Ab isotype controls. ApoGee Micro flow cytometer (Apogee Flow Systems) was used in Papers III and Size-calibrated IV. silica beads (ApogeeMix) were used to calibrate the MV gate. For Papers III (BAFF analysis section) and IV, patient plasma was diluted 1:200 with DPBS and 200 μ l was stained with 2 μ l of Abs for 30 min in RT. 110 μ l was aspirated and analysed by the cytometer and the obtained data is represented as the number of fluorescently labelled MVs/ μ l. For additional gating strategies or detailed antibody description, please refer to the experimental sections or supplementary information of the respective publications.

Proximity Extension Assay (Papers I, III, IV)

To perform proteomic profiling of MVs, isolated and lysed MVs were analysed with Cardiovascular II and III Proseek Multiplex 196 x 96 panels (Olink Proteomics). This proximity extension assay (PEA) allows to simultaneously analyze 92 known potential cardiovascular biomarkers in 96 samples, using or 0.5-1 μ g/ μ l protein. Each of the panels was evaluated by the manufacturer for specificity, precision, sensitivity, dynamic range, matrix effects and interference. The method involves three steps: incubation, extension and detection (Fig. 10). In the incubation phase, two specific antibody probes with oligonucleotide tags bind in close proximity to the target protein. In the extension phase, a PCR target sequence is formed that allows DNA polymerization during quantitative real-time PCR (qRT-PCR). The last step is a detection of the amplified sequence. The generated qRT-PCR data is further transformed to relative quantification in log2scale as Normalized Protein eXpression (NPX). Detailed description controls used for the analysis, as well as equations used for NPX calculation are provided on the manufacturer's website (www.olink.com). The analysis was performed at Clinical Biomarkers Facility at the Science for Life Laboratory, Uppsala University.



Fig. 10 Principle of Olink proximity extension assay.

Transmission Electron Microscopy (Paper I)

To visualize EVs in our sample set, transmission electron microscopy (TEM) was applied. This technique creates an image by transmitting a beam of electrons through an ultrathin specimen. Briefly, to obtain a TEM image of EVs, plasma sample was diluted with DPBS, centrifuged and washed. EV-rich pellet was fixed with paraformaldehyde in Sorensen's phosphate buffer solution. The fixed pellet was resuspended in DPBS and stained with mouse anti-human CD62E Ab. A glow discharge carbon-coated 400 mesh grid was blocked with BSA. The sample was then placed on the grid and the staining was performed with goat anti-mouse Ab conjugated with 10 nm of gold. After series of washing with PBS, the samples were fixed with glutaraldehyde, stained with uranyl acetate and dried. The imaging was done on FEI Tecnai Biotwin TEM.

Nanoparticle Tracking Analysis (Papers I and II)

Nanoparticle Tracking Analysis (NTA) is a method that allows to visualise, enumerate and assess the size distribution of EVs. It calculates the diameter of particles suspended in liquids based on Brownian motion. The analysis of isolated EVs was performed using NanoSight LM10 (Malvern) equipped with a 488 nm blue laser. EVs were stained with PE-conjugated CD62E or CD42a Abs and diluted for fluorescence-based NTA. Ab-only and PBS controls were used to ensure measurement of the vesicles.

Endothelial vascular cell culture (Paper II)

Human umbilical vein endothelial cells (HUVECs), pooled from multiple donors, were purchased from Life Technologies. Three different HUVEC lots below passage 4 were used in the *in vitro* experiments. The cells were seeded in dishes covered with attachment factor and cultured in Medium 200 with gentamycin/amphotericin and low serum growth supplement (LSGS, final serum concentration 2%), containing hydrocortisone, epidermal growth factor, basic fibroblast growth factor and heparin (Thermo Fisher Scientific).

Calcein assay (Paper II)

In study II we examined the effect of bilberry extract (BE) on endothelial vesicle release, hypothetically linked to the $P2X_7$ purinergic receptor. To study the vesiculation, ECs were stained with calcein AM, a hydrophobic dye that emits fluorescence inside the intact cells or EVs due to removal of acetomethoxy groups by endogenous intracellular esterase activity (Fig. 11). The green fluorescent signal can be further measured with e.g. fluorometer. Calcein is a reliable dye to measure vesicle release, as it does not emit fluorescence outside the cells or EVs due to its degradation, thus does not give false positive or unspecific signal like some lipophilic dyes do.⁸³



Fig. 11 Principle of calcein staining.

Compound effect on endothelial cells (Paper II)

To assess the effect of different concentrations of BE on viability and proliferation of HUVECs, trypan blue was used. The highest tested concentration of 1000 μ g/ml of BE was chosen for further experiments due to the lack of toxic effects on the cells. HUVECs were seeded in 12-well dishes and cultured for 24 h. 1 mg/ml BE-containing medium was prepared from 100 mg/ml stock. The mixture or control medium were incubated with confluent cells for 24 h. 300 μ m of P2X₇ agonist bzATP (Sigma-Aldrich) and 100 nm of P2X₇ antagonist AZ11645373 (Sigma-Aldrich) were prepared from stock solutions in the culture medium. Calcein AM (Thermo Fisher Scientific) stock was prepared in DMSO and diluted in HBSS to obtain 2 μ m working solution. The cells were washed with HBSS, 1 ml of calcein solution was added per well and the cells were incubated at 37°C for 15 min. After washing, the diluted compounds or fresh culture medium were added to the calcein-stained cells and incubated for the next 24 h. The medium was further collected, centrifuged to remove cellular debris and analysed using CLARIOstar plate reader (BMG Labtech). The workflow of the vesiculation measurement is presented on Fig. 12. All values were normalized to that of the control cells.



Fig. 12 Workflow of endothelial vesiculation analysis.

RNA extraction, reverse transcription and quantitative PCR (Paper II)

To measure gene expression, a real-time or quantitative polymerase chain reaction (qPCR) was performed on isolated RNA. The first step applies reverse transcriptase (RT) to create complementary DNA from RNA. (RT-PCR). The second step is based on a real-time measurement of a target DNA sequence amplification by DNA polymerase (qPCR) that reflects the expression of the gene of interest.

The cells were harvested with 700 μ L QIAzol (Qiagen) and frozen at -80°C. RNA isolation was performed with miRNeasy kit with DNase digestion (Qiagen) as per manufacturer's instructions. RNA concentration and quality was assessed with NANODROP 2000C spectrophotometer. High Capacity RNA to cDNA kit (Thermo Fisher Scientific) was used for reverse transcription. The mRNA expression was assessed using TaqMan Fast Universal PCR Master Mixes (Thermo Fisher Scientific) with 20 ng template as per manufacturer's instructions. The TaqMan employs annealing of the template with dye-labelled probes. assay In case of no DNA polymerization, the dye is quenched and there is no fluorescence. However, when the DNA polymerase is active, it releases the dye from the quencher that results in the fluorescent signal that is proportional to the amount of the synthetized DNA. qPCR reaction was performed on StepOnePlus Real-Time PCR System (Thermo Fisher Scientific). GAPDH was used as a housekeeping gene for the reference (Δ Ct) and the data was normalized to the

mean of the controls ($\Delta\Delta$ Ct), according to the formula $2^{-\Delta\Delta Ct}$, giving the relative quantification (RQ) parameter.

Western Blot (Paper II)

Western Blot is a technique that allows to quantify protein expression based on the antibody binding. HUVECs were plated in 6-well plates and cultured for 24 h. 1 mg/ml BE was added and the cells were harvested and lysed with 200 µL RIPA buffer (Cell Signaling) containing phosphatase inhibitors. Protein concentration was measured using BCA Protein Assay Kit (Thermo Fisher Scientific). 10 µg of protein was loaded on pre-cast polyacrylamide midi gels (Thermo Fisher Scientific) and separated with electrophoresis based on the molecular weight. Further, the proteins were transferred via iBlot 2 transferring system (Thermo Fisher Scientific) on a nitrocellulose membrane. To prevent non-specific binding of Abs, the membrane was blocked with protein-free blocking buffer (Thermo Fisher Scientific). Primary Abs against the proteins of interest were incubated with the membrane overnight and washed. Then, a secondary Ab, conjugated with horseradish peroxidase (HRP) and binding to the primary Ab was added. Visualisation was performed with Pico Chemiluminescent Assay (Thermo Fisher Scientific), containing HRP substrate, on Odyssey Fc (LI-COR) based on the densitometric properties of the membrane. β -actin was used as a reference protein. The phosphorylation levels of phospho-Akt (pAkt) and phospho-p38 (pp38) were normalised to the total levels of these proteins. The data is represented as normalised to the control.

Coronary flow reserve measurement (Paper III)

Coronary Flow Reserve (CFR) is a clinical parameter used for measuring vascular blood flow. It assesses both atherosclerotic lesions in the coronary arteries and microvascular dysfunction. As CFR represents blood delivery to the heart during higher demand,^{84,85} high CFR values usually mean better coronary circulation (Fig. 13). Low CFR is a strong and independent predictor of cardiovascular events.⁸⁵⁻⁸⁸ In Paper III, CFR was measured in LAD with transthoracic echocardiography by experienced operators at Skåne University Hospital and Sahlgrenska University Hospital. The above-mentioned method is an alternative to invasive methods and carries strong prognostic information.⁸⁵ In order to obtain maximal hyperaemia in coronary arteries, the procedure involved adenosine administration to patients. This purine nucleoside acts via A_{2A} G_s-protein coupled receptors, present in the vasculature that stimulate adenylyl cyclase.^{89,90} cAMP further leads to SMC

hyperpolarization through potassium channels, subsequent endothelial-independent SMC relaxation and finally vasodilation of the coronary artery.⁸⁹



Fig. 13 Relationship between coronary flow reserve values and the level of stenosis. Based on https://thoracickey.com/24-fractional-flow-reserve/.

Statistics (Papers I-IV)

For all studies p-values <0.05 were considered as statistically significant. The normality of the data was determined using Shapiro test. Statistical analyses were performed using GraphPad Prism or STATA 14 software. In Papers I, III and IV, the analysis of proteomic profiling with Olink PEA was done on NPX (Normalized Protein Expression) values. Proteins with $50\% \leq$ values lower than the limit of detection were removed from the analysis and analysed with paired t-test or Wilcoxon signed-rank test. In study I, Wilcoxon signed rank test for paired data was used for FCM comparison between the time points (before and 1 h after exercise). In Paper II, Mann-Whitney U test was applied to compare differences between BE supplementation and no dietary intervention after 8 weeks. One-way ANOVA with false discovery rate (FDR) correction of Benjamini and Hochberg was used to analyse the in vitro effects of BE and compounds on ECs. In Paper III Spearman's coefficients were calculated to determine correlations. Multiple linear regression was used to measure the relationship between CFR and certain factors. In Paper IV, paired t-test or Wilcoxon-signed rank tests were used to analyse the differences in FCM and proteomic data between proximal and distal samples to the plaque. For detailed description of the applied statistical methods, please refer to the respective publications.

Aims and Hypotheses

The overall objective of this thesis was to study the role of MVs in cardiovascularrelated pathologies, as well as to optimize the novel method for MV enrichment – acoustic trapping.

The specific objectives of each study are given below:

The primary aim of **Paper I** was to optimize the acoustic trapping machine for MV isolation from plasma and compare it to standard high-speed centrifugation protocols for flow cytometric and proteomic profiling. A clinical study was performed to also study the effect of exercise on the vasculature in the MV context.

Paper II investigated if there is an influence of bilberry extract dietary intervention in MI patients on MV levels in plasma. The *in vitro* part of the study aimed to explore the effect of bilberry extract on endothelial vesiculation, related to the $P2X_7$ receptor.

Paper III studied the relationship between coronary flow reserve, a parameter depicting blood flow in the heart and MV levels or vesicle-related biomarkers, circulating in patients with CVD.

Paper IV focused on exploring the composition and cellular origin of plaquereleased EVs, following balloon angioplasty. It also examined if there was an advantage of analysing isolated EVs, compared to crude plasma samples.
Results and Discussion

Paper I

Plasma-derived MVs offer a promising approach in diagnostics of several clinical pathologies. As MV isolation by currently available methods is both technically challenging and requires large sample volumes,⁶¹ we used an alternative method, based on an acoustic standing wave technology.

Optimization of acoustic trapping

The optimization of the acoustic trapping was established by monitoring the efficiency of the machine that was calculated based on the levels of CD42a+ platelet MVs (Fig. 14A), as the majority of circulating MVs is of platelet origin.⁴⁰ For this purpose, pooled frozen plasma from volunteers was used as an internal control to monitor the device stability. Our results showed that acoustic trapping exhibited high and stable MV recovery throughout the experiment. We further compared platelet MV isolation efficiency with a commonly used high-speed centrifugation protocol. As presented on Fig. 14B, the loss of MVs during isolation is substantially lower for acoustic trapping that can he related to non-contact principle of isolation and thus decreased aggregation/disruption of the vesicles. Improved design of the acoustic trapping machine increased MV recovery compared to earlier reports⁹¹ and enhanced the long-term stability.



Fig. 14 Results of acoustic trapping optimization for plasma microvescicle isolation. A: Efficiency of acoustic trapping. B: Comparison of acoustic trapping and high-speed centrifugation recovery. Mean ± SEM. One-way ANOVA with FDR correction of Benjamini and Hochberg. *Statistically significant, p<0.05.

Exercise study worklflow

To add a clinical context to the comparison, a pilot study consisting of 20 subjects who underwent physical exercise was carried out. Plasma samples were obtained before and 1 h after the training. The complete study workflow is presented on Fig. 15. MVs were isolated both with acoustic trapping and standard high-speed centrifugation protocol.



Fig. 15 Acoustic trapping and high-speed centrifugation comparison workflow.

Flow cytometric analysis of MVs

The methods were further compared by FCM. The analysis revealed a rapid decrease in CD62E+ (E-selectin) endothelial MVs 1 h after exercise on stationary bicycle that was, importantly, consistent for both methods (Fig. 16).

It has been described that physical exercise has an impact on MV concentration in blood. However, exercise protocols, MV isolation methods and analysed markers vary substantially between the studies, making the outcome comparisons challenging.⁹²⁻¹⁰⁰ A study by Babbitt et al. also evaluated the levels of CD62E+ MVs, but after a long-term exercise intervention in African Americans.⁹⁵ The results showed that six months of aerobic exercise training significantly decreased circulating MVs derived from activated endothelium. One of the possible explanations of our and Babbitt's observations is a protective effect of exercise on human body, including the vasculature and inflammatory response¹⁰¹⁻¹⁰⁵ that is reflected in the levels of CD62E+ MVs in blood.



Fig. 16 Comparison of acoustic trapping (A) and high-speed centrifugation (B) for flow cytometric analysis of plasma microvesicles. Mean ± SD. Wilcoxon signed-rank test. *Statistically significant, p<0.05.

Nanoparticle tracking analysis of vesicles

NTA was another method, used for the comparison of the methods (Fig. 17). It revealed that both methods of isolation obtained similar results regarding EV size distribution, with CD62E+ vesicles primarily in the MV size range. Additionally, acoustic trapping yielded higher levels of CD62E+ EVs (Fig. 17B), compared to the centrifugation protocol (Fig. 17D), which is consistent with the results from the FCM analysis.



Fig. 17 Comparison of acoustic trapping (upper panel) and high-speed centrifugation (lower panel) with nanoparticle tracking analysis of plasma microvesicles. A, C – evaluation of total concentration of vesicles (unstained). B, D – evaluation of fluorescently labeled CD62E+ vesicles.

Proteomic profiling of MVs

We also compared the protein content of vesicles circulating before and 1 h after physical exercise, isolated with acoustic trapping and high-speed centrifugation. The proteomic analysis with Olink PEA cardiovascular panels showed 54 significantly changed proteins in the MV fraction (acoustic trapping and pellet fractions) in response to physical exercise, whereas the MV-free plasma proteome (supernatant) displayed only four differentially regulated proteins (Fig. 18). As the vast majority of differentially regulated proteins were present in the vesicles, this finding underlines the role of MVs in cellular communication and their potential application as plasma-derived biomarkers. STRING database search revealed that most of the changed proteins were associated with inflammatory response (e.g. CCL17, CCL24, CXCL1, CXCL16, IL-1RA, IL-18), angiogenesis (angiopoietin 1, TIE-2) and coagulation (PAR1, P-selectin, Src, vWF) that stands in line with the current knowledge about the effects of exercise on biochemical parameters.^{102,106-118} Furthermore, most possible sources of the vesicular exercise-regulated proteins were ECs, leukocytes, muscle tissue and platelets.



Fig. 18 Venn diagram showing comparison of acoustic trapping and high-speed centrifugation (microvesiclerich pellet and microvesicle-free supernatant), based on the proteomic profiling of exercise-regulated proteins. Numbers in the circles represent significantly changed proteins after the exercise. Bolded proteins have been previously described as of vesicular origin, based on Vesiclepedia.¹¹⁹ Abbreviations of all exercise-regulated proteins are explained in Paper I and its supplementary material.

Since the manuscript publication in 2018, several new original, as well as review reports regarding exercise and EVs have appeared.^{100,120-125} Some researchers point at the role of muscle tissue as an endocrine organ and exercise-released exosomes as important reservoirs of different chemokines and growth factors, sometimes referred to as 'exerkines'.^{120,122,123} This hypothesis is consistent with our findings from the proteomic study that identified exercise-regulated proteins of muscle tissue origin, such as myoglobin and follistatin. As described above, the NTA revealed a very small amount of EVs isolated with acoustic trapping with diameter below 100 nm that could be of exosome subtype of vesicles, transporting exerkines in the circulation during/after the physical exercise.

Transmission electron microscopy of vesicles

We also performed additional experiments to validate the presence of CD62E marker on the vesicles in our material. For this purpose, immunogold staining against CD62E was performed and analysed with TEM. It confirmed the presence of both the intact vesicles and the marker on the vesicle surface (Fig. 19).



Fig. 19 CD62E immunogold staining for transmission electron microscopy. Red arrows indicate the presence of CD62E antigen.

In summary, results presented in Paper I showed that acoustic trapping is a fast and efficient method that is comparable with the standard centrifugation protocols for MV isolation. It has the advantage of using lower sample volumes and contact-free separation with higher yield. Furthermore, this study underlines the potential application of acoustic trapping in the clinical vesicle-based diagnostics. The results of the clinical study point at a protective effect of exercise on the vasculature.

Paper II

BEARSMART clinical study

In the BEARSMART study, MI patients received an 8-week bilberry extract (BE) dietary supplementation,¹²⁶ whereas the control group did not take any dietary supplementation (Fig. 20). MVs were isolated from the patient plasma with acoustic trapping. CD42a+ platelet MVs and CD62E+ endothelial MVs were measured with FCM.



Fig. 20 Study II workflow – investigation of the effect of bilberry extract on circulating platelet and endothelial microvesicles.

Flow cytometric analysis of microvesicles

FCM analysis revealed that 8-week BE dietary intervention significantly decreased levels of both circulating platelet (Fig. 21A) and endothelial MVs (Fig. 21B), compared to the control group.

Anthocyanins are the major group of compounds present in bilberries that could be linked to the reduced vesiculation. These polyphenols reduce both platelet^{23,127,128} and endothelial activation,^{129,130} thus possibly the vesicle release. As MVs are related to the cardiovascular pathologies, our results demonstrate an important effect of dietary intervention with BE supplementation on MI patients.



Fig. 21 Eight-week supplementation of bilberry extract (BE) significantly decreases circulating platelet (A) and endothelial (B) microvesicles. Median \pm 95% CI. Mann-Whitney U test. *Statistically significant, p<0.05.

Nanoparticle tracking analysis of patient samples

We also performed NTA to determine vesicle size distribution in the samples (Fig. 22A). The majority of isolated vesicles were of 200-400 nm diameter, thus was in the range of MVs. The analysis also confirmed the presence of CD62E+ (Fig. 22B) and CD42a+ (Fig. 22C) vesicles in patient plasma.



Fig. 22 Evaluation of size distribution of vesicles present in patient samples, isolated with acoustic trapping. A- size distribution of all vesicles (unstained). B- size distribution of vesicles originating from activated endothelium. Csize distribution of platelet-derived vesicles.

Effect of bilberry extract on endothelial cells

As BE supplementation decreased circulating endothelial MVs, we further investigated the effect of BE solution on endothelial vesicle release, using HUVECs and calcein staining. Concentration of 1 mg/ml of BE was chosen due to the lack of toxic effects on the cells. Based on the literature, $P2X_7$ receptor was selected as a target for induction of vesiculation.¹³¹ Western blot and RT-qPCR techniques were applied to explore the mechanisms of BE influence on ECs (Fig. 23).



Fig. 23 Study II workflow – *in vitro* experimental part investigating the influence of bilberry extract on endothelial vesiculation, as well as the mechanisms underlying this effect.

Influence of bilberry extract on endothelial vesiculation

Our results show that *in vitro* treatment of ECs with BE decreased endothelial vesicle release (Fig. 24). $P2X_7$ receptor agonist, bzATP, increased the vesiculation compared to the control cells that was abrogated in the presence of $P2X_7$ receptor antagonist AZ11645373. Pre-incubation with BE further prevented bz-ATP evoked vesiculation. Thus, our data shows that BE reduces both unstimulated and $P2X_7$ -stimulated vesicle release from ECs.



Fig. 24 Effect of bilberry extract (BE) on endothelial vesiculation. BE incubation significantly decreased vesicle release and bzATP increased the vesiculation compared to the control cells. Pre-incubation with BE prevented bz-ATP evoked vesiculation. Mean ± SD. One-way ANOVA with FDR correction of Benjamini and Hochberg. *Statistically significant, p<0.05.

Modulation of protein phosphorylation by bilberry extract

Based on the vesiculation results, Akt and p38 were selected as candidate proteins for further investigation, as both take part in P2X₇ signaling pathways, as well as vesicle release.¹³²⁻¹³⁵ Furthermore, it has been previously reported that bilberryderived compounds affect both Akt and p38 phosphorylation *in vitro* and *in vivo*.¹³⁶⁻¹³⁹ Our results show that incubation of ECs with BE decreased Akt phosphorylation after 3 h (Fig. 25A), but not after 6 h (Fig. 25B). p38 phosphorylation, however, did not change at both time points (Fig. 25C,D). This suggests that p38 is not directly involved in the observed BE-related vesiculation decrease.



Fig. 25 Effect of bilberry extract (BE) on protein phosphorylation. BE incubation for 3 h significantly decreased Akt phosphorylation compared to the control cells. No effect was observed on p38 phosphorylation. Mean ± SD. One-way ANOVA with FDR correction of Benjamini and Hochberg. *Statistically significant, p<0.05.

Effect of bilberry extract on gene expression

Since it has been shown that Akt signaling pathway regulates $P2X_7$ expression in the neural cell line,¹⁴⁰ we also investigated if the observed decrease in the vesiculation is associated with BE effect on $P2X_7$ transcription. As seen on Fig. 26, BE treatment decreased $P2X_7$ gene expression that could explain a decrease in the bzATP-stimulated vesiculation after pre-treatment with BE. A similar pattern was observed after bzATP incubation that can be explained by a negative-feedback loop and inhibition of $P2X_7$ transcription after the receptor stimulation.¹⁴¹ However, BE decreased the receptor mRNA levels more profoundly than the agonist. Furthermore, treatment of HUVECs with BE and subsequent stimulation with bzATP also decreased $P2X_7$ expression.



Fig. 26 Effect of bilberry extract (BE) on P2X₇ gene expression. BE incubation significantly decreased P2X₇ mRNA levels compared to the control cells. BzATP also decreased P2X₇ transcription. Mean ± SD. One-way ANOVA with FDR correction of Benjamini and Hochberg. *Statistically significant, p<0.05. RQ – relative quantification.

We further selected four vesiculation-related genes that could potentially be affected by BE. Rab27b and Rab27a control different stages of vesicle release,¹⁴² SMPD1 regulates P2X₇-dependent vesiculation in glial cells¹⁴³ and ARF6 is involved in MV release from tumour cell line,¹⁴⁴ as well as from macrophages via P2X₇ activation.¹⁴⁵ Our results show that BE treatment decreased the expression of Rab27a and Rab27b genes (Fig. 27A,B). Pre-incubation with BE diminished the bzATP-induced increase in Rab27b, Rab27a and SMPD1 gene expression (Fig. 27A-C). The results were not conclusive for ARF6 expression, as its transcription decreased after all combinations of treatments (Fig. 27D).

Based on our findings, we propose a model, where BE and its major active constituents, anthocyanins, reduce Akt phosphorylation and further $P2X_7$ expression that subsequently decreases endothelial vesiculation. Importantly, we also believe that the *in vitro* observed effect of BE reflects the decrease in plasma endothelial MV concentration in BE-supplemented MI patients.

Furthermore, as BE is a complex mixture and the BE concentration used in the *in vitro* study was relatively high, other potential mechanisms could be involved in the reduction of circulating endothelial MVs. Anthocyanins or anthocyanin-

derived metabolites could potentially act locally as free radical scavengers^{22,146} and thus prevent endothelial activation and vesiculation.³⁷



Fig. 27 Effect of bilberry extract (BE) on vesiculation-related gene expression. BE incubation significantly decreased mRNA levels of Rab27b and Rab27a (A, B). BzATP increased the transcription compared to the control cells (A, B, C). Pre-incubation with BE prevented bz-ATP evoked vesiculation (A, B, C). No effect was observed on ARF6 gene expression. Mean ± SD. One-way ANOVA with FDR correction of Benjamini and Hochberg. *Statistically significant, p<0.05. RQ – relative quantification

In summary, the results of Paper II demonstrated that BE dietary intervention improves the profile of circulating MVs in blood of CVD patients. Additionally, we show that BE evokes effects on endothelial vesiculation through several molecular mechanisms, connected with $P2X_7$ signaling pathway, such as Akt phosphorylation, $P2X_7$ and vesiculation-related gene expression. We reported for the first time that

dietary compounds have impact on vesiculation, both *in vitro* and *in vivo*. This supports the importance of our findings for potential dietary recommendations for cardiovascular patients, as well as a possible part of nutritional prevention of CVD.

Paper III

PROFLOW clinical study

The prospective, exploratory and open PROspective Evaluation of Coronary FLOW Reserve and Molecular Biomarkers in Patients with Established Coronary Artery Disease (PROFLOW) study has previously been described by Haraldsson et al.¹⁴⁷ It high-risk patients with CAD, recruited by Department involved 619 of Cardiology at Skåne University Hospital in Lund and Sahlgrenska University Hospital in Göteborg. Swedish Coronary Angiography and Angioplasty registry (SCAAR) was used to identify patients with high risk for future major adverse cardiovascular events. This study presents the data from 220 patients recruited in Lund. Patients were clustered into high and low CFR groups based on the median value. CFR measured in LAD with CFR was ultrasound system as described previously¹⁴⁷ and adenosine-assisted transthoracic echocardiography protocol was applied. MVs were isolated from plasma with acoustic trapping and analysed with FCM and Olink proteomic panels.

Coronary flow reserve and microvesicles

We investigated the relationship between platelet and endothelial MVs and CFR, as the currently available data points at their contribution to endothelial dysfunction and development of atherosclerosis. Our results show a negative correlation between CFR and platelet and endothelial MV levels in plasma (Fig. 28 B, D). MV levels were also compared in high and low CFR groups. According to our results, the low CFR group had significantly higher concentration of circulating platelet (Fig. 26A) and endothelial MVs (Fig. 26C), compared to the high CFR group. Furthermore, we found a positive correlation between platelet and endothelial MV levels (Fig. 29). We did not observe any relationship between MVs and risk factors, as well as disease or inflammatory burden.

We also applied a statistical approach, involving explanatory multiple linear regression, to investigate the relationship of CFR with patient characteristics such as BMI, smoking, age, diabetes, hypertension, dyslipidaemia, low density lipoprotein (LDL) levels and circulating platelet and endothelial MV concentrations. The results showed a statistical significance for platelet MV levels parameter estimate, which means that for each increase by 100 platelet MVs/µl in plasma, the CFR value decreases on average by 0.06.

The above results could be explained with the notion that circulating MVs could contribute to a decrease in CFR. It is known that MVs can affect vascular physiology and contribute to formation of atherosclerotic plaques.^{34,39,46,148} The lower CFR values can also result from the presence of MVs released by already dysfunctional endothelium that further decrease NO production and vascular relaxation in healthy ECs.¹⁴⁹ Moreover, MVs generated by ageing ECs leadto progression of the plaque through increased calcification in the vessel.³⁹



Fig. 28 Coronary flow reserve negatively correlates with circulating platelet and endothelial microvesicles. A, C: Patients with high CFR have lower concentration of circulating platelet (A) and endothelial (C) MVs, compared to the low CFR group. Median \pm 95% CI. Mann-Whitney U test. *Statistically significant, p<0.05. B, D: Platelet (B) and endothelial (D) MVs in plasma negatively correlate with CFR, Rho - Spearman's coefficient, p <0.05.



Fig. 29 Circulating endothelial microvesicle levels positively correlate with platelet microvesicles. Rho - Spearman's coefficient, p<0.05.

Proteomic analysis of microvesicles

As our data demonstrated that there is a relationship between CFR and MVs in plasma, the notion if this is reflected in the MV content was further explored. Olink CVD panels were used to perform a proteomic analysis on MVs enriched from patient plasma. The statistical analysis of CFR and NPX levels showed seven negatively correlating proteins (Fig. 30): NF κ B essential modulator (NEMO), resistin, Tyrosine-protein kinase receptor UFO (AXL), perlecan, B-cell activating factor (BAFF), Insulin-like growth factor-binding protein 7 (IGFBP7) and CD163. No positively correlating proteins with CFR were found.

The comparison of high and low CFR groups revealed that the low CFR group exhibited higher concentration in the vesicles of nearly all above-mentioned proteins, with CD163 as an exception that did not reach statistical significance, despite a visible trend toward higher values in the low CFR group (Fig. 31).



Fig. 30 Correlation plots showing results of proteomic analysis with Olink cardiovascular panels. Seven proteins negatively correlate with patient coronary flow reserve, based on the normalized protein levels (NPX), Rho – Spearman's coefficient, p<0.05.



Fig. 31 Patients from the low CFR group are characterized with significantly higher levels of cardiovascular biomarkers levels, based on the normalized protein expression (NPX) values. Median \pm range, Mann-Whitney U test for unpaired data. *Statistically significant, p<0.05.

According to the current knowledge, all of the significantly correlating proteins are involved in pathological and pro-atherosclerotic processes.^{38,150-163} It is noteworthy that as most of the proteins have been measured as soluble forms in plasma, there is a possibility that they could have been partially measured as MV components. Thus, this study confirms for the first time that these proteins are present in the vesicular fraction of plasma.

Analysis of circulating BAFF-expressing microvesicles

One of the significant proteins, BAFF, is a protein that is directly involved in atherogenesis and its role is implicated in CAD.¹⁴⁸ It is also a pharmacological target for developing anti-inflammatory therapeutics.¹⁶⁴ Furthermore, its role has been implied in the progression of atherosclerosis in mice¹⁵⁰ and application of anti-BAFF receptor Ab reduces the lesions.¹⁶⁵ BAFF-related B-cell depletion in mice decreases cardiac inflammation and infarct size, as well as circulating levels of BAFF correlate with adverse effects in MI patients.¹⁶⁶

Thus, we performed FCM analysis to identify cellular origin of circulating BAFF-expressing MVs. Platelet, leukocyte, endothelial and erythrocyte origin of BAFF+ MVs was explored. As seen on Fig. 32, our results showed that BAFF+ MVs in plasma co-stain with CD42b and CD31 markers and therefore have platelet origin. Our data revealed that leukocytes (CD45 and CD16 markers), activated endothelial cells (CD62E marker) or erythrocytes (CD235a marker) were not the major sources of circulating BAFF+ MVs (Fig. 32).



Fig. 32 Investigation of cellular origin of BAFF+ microvesicles in plasma. Patient plasma was stained both with BAFF and platelet (upper panel), leukocyte (middle paned), endothelial (lower panel, left) and erythrocyte (lower panel, right) markers. Dot plots show that circulating BAFF+ MVs represent a fraction of platelet MVs.

As small fraction of CD31 is also expressed by ECs, we co-stained CD31 with platelet marker CD42b and endothelial marker CD144 (VE-cadherin) (Fig. 33). The staining confirmed platelet (Fig. 33A) and not endothelial origin (Fig. 33B) of BAFF+ CD31+ MVs.



Fig. 33 Co-staining of CD31 marker with platelet (A) and endothelial (B) markers to determine major CD31 cellular origin in plasma. Dot plots show that CD31+ MVs are mainly of platelet origin, thus confirm that BAFF-expressing MVs are a fraction of platelet MVs.

To further explore circulating BAFF+ MV, we investigated the expression on their surface of CD36 that is an oxidized LDL receptor, involved in pro-atherogenic processes¹⁶⁷ (Fig. 34). The staining demonstrated that a fraction of BAFF+ MVs expressed CD36 and the marker was of platelet (Fig. 34B) and not endothelial (Fig. 34C), leukocyte (Fig. 32D) or erythrocyte (Fig. 34E) origin.

Previous reports considered leukocytes as the main cellular source of BAFF in plasma.¹⁶⁴ However, our data demonstrated that the vast majority of circulating BAFF+ MVs is a subpopulation of platelet MVs. Our findings are supported by the results of other groups who discovered BAFF expression on platelets¹⁶⁸ and megakaryocytes.¹⁶⁹ Furthermore, as BAFF+ MVs also carry pro-atherosclerotic CD36, they represent a fraction of MVs involved in pro-inflammatory and pro-atherogenic processes.

Atherosclerosis is sometimes described as an inflammatory disease,⁵ thus to further explore if pro-inflammatory conditions are connected with BAFF+ MV release, we stimulated a megakaryocyte cell line with TNF α , a cytokine related to endothelial dysfunction.¹⁷⁰ Addition of TNF α to the culture medium increased the production of BAFF-expressing MVs by MEG01 cells (Fig. 35B), compared to the lack of stimulation (Fig. 35A). The above results could partially explain the higher levels BAFF in MVs from the low CFR group. BAFF appears to be a promising drug target for treatment of vascular dysfunction, as therapeutics against it already exist.



Fig. 34 Analysis of CD36 marker expression on BAFF+ microvesicles (A) and co-staining with platelet (B), endothelial (C), leukocyte (D) and erythrocyte (E) markers to determine its cellular origin in plasma. Dot plots demonstrate that CD36+ MVs are mainly of platelet origin.



Fig. 35 Pro-inflammatory conditions stimulate a release of BAFF+ microvesicles from megakaryocyte cell line. MEG01 cells were stimulated with TNF α that substantially increased BAFF+ MV release (B), compared to the unstimulated cells (A).

In summary, in Paper III we investigated if there was a connection between CFR values and circulating MVs. The low CFR group was characterized with significantly increased concentration of platelet and endothelial MV levels in plasma. Proteomic analysis of biomarker cargo in MVs showed that AXL, CD163, IGFBP-7, NEMO, resistin, BAFF and perlecan levels negatively correlated with CFR. All of the above-mentioned proteins are involved in inflammatory and pro-atherogenic processes, thus they reflect the pathological vascular status of the patients from the low CFR group and have potential clinical value as biomarkers of deterioration of vascular status. Moreover, anti-BAFF therapeutics could be potentially explored for treatment against CVD.

Paper IV

'Plaque-released MVs' clinical study

The study consisted of 20 participants with a mean age of 66 years that were recruited at Department of Cardiology, Skåne University Hospital in Lund. All patients had a diagnosed coronary arterial stenosis and had undergone balloon angioplasty. Blood samples were derived from aortic root (proximal to the plaque), reflecting circulating MVs or the coronary artery part distal to the plaque, containing plaque-released MVs, following balloon angioplasty.

Flow cytometric analysis of plaque-released MVs

We applied FCM to explore if atherosclerotic plaque contains MVs that are released after balloon angioplasty. Our analysis showed that total concentration of MVs derived from part distal to the plaque was significantly higher, compared to the MV levels present in the circulation (Fig. 36). Although a substantial increase in total MV concentration after balloon angioplasty was observed, no subtype of MVs increased significantly more than the other.

We further investigated the cellular origin of plaque-derived MVs with FCM. Several cellular origins of MVs have been detected (Fig. 37): platelet (CD42b+, CD62P+, CD36+, BAFF+), endothelial (CD62E+, CD201+), leukocyte (CD45+, CD16+, CD16+, CD14+, CD204+), SMC (CD140b+) and erythrocyte (CD235a+).

It has been implicated that the above-mentioned MVs are hallmarks of CVD and atherosclerosis^{34,52,149,171} and their markers play several roles in pro-inflammatory and pro-atherosclerotic processes.^{34,172-175} Generally, high concentration of MVs in blood, expressing proteins involved in progression of atherosclerosis, are related to a degree of CVD and points at a pathological environment within the vascular system.



Fig. 36 Flow cytometric analysis of a total MV release from atherosclerotic plaque, following balloon angioplasty. Median ± 95% CI. Wilcoxon signed rank test for paired data. * Statistically significant, p<0.05

Proteomic analysis of plaque-released EVs

To explore a protein cargo of plaque-released MVs after balloon angioplasty, we performed an analysis with Olink CVD panels. Additional aim of this study was to examine if there was an advantage of analysing enriched EVs. Thus, we compared the proteomic results from EVs isolated with acoustic trapping with the data obtained from crude plasma samples that also contain soluble, non-vesicular proteins.

In case of enriched vesicles, 29 proteins had significantly higher concentration in plaque released-MVs, compared to circulating MVs (Table 1). Analysis of crude plasma samples revealed 49 proteins with significantly higher levels in samples derived from distal part to the plaque (Table 1). There was an overlap of 12 significant proteins between enriched MVs and plasma samples (Fig. 38, Table 1).



Fig. 37 Analysis of cellular origin of plaque-released MVs with flow cytometry. Plasma samples were stained with Abs against platelet markers: CD42b, CD36, CD62P, BAFF (A), endothelial markers: CD62E, CD201 (B), leukocyte markers: CD45, CD16, CD14, CD204 (C), erythrocyte marker CD235a and SMC marker CD140b (D).

Abbreviation	Protein name	Enriched in plaque- released EVs	Enriched in distal plasma samples
ACE2	Angiotensin-converting enzyme 2		+
ADM	Pro-adrenomedullin		+
AGRP	Agouti-related protein		+
AMBP	Alpha-1-microglobulin/bikunin precursor		+
AZU1	Azurocidin	+	
BOC	Brother of CDO		+
CCL3	c-c motif chemokine 3 (MIP1)		+
CD163	Scavenger receptor cysteine-rich type 1 protein M130	+	
CD4	T-cell surface glycoprotein cluster of differentiation 4		+
CD93	Complement component C1q receptor	+	
CEACAM8	Carcinoembryonic antigen-related cell adhesion molecule 8 (CD66b)		+
CHI3L1	Chitinase-3-like protein 1	+	
CPB1	Carboxypeptidase B	+	
CTRC	Chymotrypsin-C		+
CTSL	Procathepsin L		+
DCN	Decorin		+
FABP2	Fatty acid-binding protein, intestinal		+
FGF21	Fibroblast growth factor 21		+
FS	Follistatin	+	
Gal-9	Galectin-9	+	+
GDF-2	Growth/differentiation factor 2/BMP-9		+
GIF	Cobalamin binding intrinsic factor		+
GRN	Progranulin	+	
HAOX1	Hydroxyacid oxidase 1		+
HO-1	Heme oxygenase-1	+	+
IGFBP-7	Insulin-like growth factor-binding protein 7	+	
IL-6RA	Interleukin-6 receptor subunit alpha	+	
IL16	Pro-interleukin-16		+
IL1RA	Interleukin-1 receptor antagonist		+
IL1RL2	Interleukin-1 receptor-like 2		+
IL4RA	Interleukin-4 receptor subunit alpha		+
IL6	Interleukin-6		+
KIM1	Hepatitis A virus cellular receptor 1 (HAVCR1, TIM1)		+
LEP	Leptin		+
LOX-1	Lectin-like oxidized LDL receptor 1	+	+
LPL	Lipoprotein lipase	+	
MARCO	Macrophage receptor MARCO	+	+
MB	Myoglobin	+	
MERTK	Tyrosine-protein kinase Mer		+
MMP12	Matrix metalloproteinase-12	+	+
OPN	Osteopontin	+	
OSCAR	Osteoclast-associated immunoglobulin-like receptor	+	+
PARP-1	Poly (ADP-ribose) polymerase 1		+
PD-L2	Programmed cell death 1 ligand 2		+
PGLYRP1	Peptidoglycan recognition protein 1	+	
PLC	Perlecan	+	
PlgR	Polymeric immunoglobulin receptor		+

 Table 1. List of proteins with significantly higher levels in plaque-released Evs and distal plasma samples, idetified with Olink proteomic panels.

PRELP	Prolargin	+	
PRSS8	Prostasin	+	+
RAGE	Receptor for advanced glycosylation end products	+	+
REN	Renin		+
SCF	Kit ligand/Stem cell factor		+
SELE	E-selectin	+	
SERPINA12	Serpin A12/Vaspin		+
SPON2	Spondin-2	+	+
STK4	Serine/threonine-protein kinase 4/Mst1		+
TF	Tissue Factor		+
TGM2	Protein-glutamine gamma-glutamyltransferase 2	+	+
THBS2	Thrombospondin-2	+	+
TIE2	Angiopoietin-1 receptor		+
ТМ	Thrombomodulin		+
TNFRSF13B	Tumor necrosis factor receptor superfamily member 13B (TACI)	+	+
TNFSF13B	B-cell activating factor (BAFF)	+	
TRAIL-R2	Tumor necrosis factor receptor superfamily member 10B		+
VEGFD	Vascular endothelial growth factor D		+
XCL1	Lymphotactin		+

This study was of an exploratory kind, thus the proteomic analysis aimed to study the biomarker patterns in plaque-released MVs, rather than concentrating on particular proteins. We compared the results obtained from isolated MVs (vesicular fraction) with plasma samples (vesicular + soluble fractions) to investigate if there was any advantage of MV purification.

Isolated plaque-released MVs, compared to circulating MVs, were characterised with significantly higher levels of proteins involved in progression of atherosclerosis, e.g. inflammation (RAGE, CHI3L1, E-selectin, PGLYRP1, galectin-9, progranulin, azurocidin, IL-6 receptor, BAFF and BAFF receptor – TNFRSF13B, OSCAR),¹⁷⁶⁻¹⁸⁶ macrophage activation (CD163, CD93, MARCO),^{163,187,188} oxidized lipid uptake (LOX-1),¹⁸⁹ foam cell formation (follistatin),¹⁹⁰ extracellular matrix remodelling (MMP12, TGM2, spondin-2, thrombospondin-2, prolargin, perlecan, osteopontin, CHI3L1, IGFBP7),^{160,191-199} proteolysis (prostasin, CPB1)^{200,201} and muscle tissue biology (myoglobin).²⁰²

Distal plasma samples, containing both soluble and vesicle-derived proteins, were characterized with significantly higher levels of proteins related to e.g. inflammation (IL-6, IL-16, IL-1RA, IL-4 receptor, IL-1RL2, RAGE, CCL3, CD4, TNFRSF13B, leptin, CEACAM8, OSCAR, PD-L2, PlgR, XCL1, galectin-9, STK4),^{176,178,182,183,186,203-215} activation of macrophages (MARCO, KIM1, MERTK),^{188,216,217} endothelial dysfunction (leptin, GDF2),^{215,218} calcification (GDF2),²¹⁹ coagulation (TF, thrombomodulin),^{220,221} biology of fatty acids (FABP2, LOX1, HAOX1, TIE2),^{189,222-224} extracellular matrix (decorin, spondin-2, thrombospondin-2, TGM2),^{191,192,194,225} apoptosis (TRAIL-R2, STK4),^{226,227}

proteolysis (MMP12, chymotrypsin-C, procathepsin L, prostasin),^{200,228,229} angiogenesis (TIE2, VEGFD),^{224,230} antioxidant activity (HO1, AMBP),^{231,232} muscle tissue (BOC),²³³ DNA repair (PARP1),²³⁴ proliferation (SCF)²³⁵ and feeding behaviour (AGRP, FGF21).²³⁶⁻²³⁸ Interestingly, we observed an increase in distal plasma samples of vasoconstrictors ACE2²³⁹ and proadrenomedullin,²⁴⁰⁻²⁴³ as well as a potent vasoconstrictor, renin.²⁴⁴

Bioinformatic analysis of isolated EVs and plasma

We further explored the cargo of plaque-released MVs with STRING database search (Fig. 41). The biological process analysis showed that vesicle preparations, containing atheroma-derived MVs, were generally related to pro-atherogenic processes, such as stress and inflammatory responses (macrophage activation, chemotaxis, cytokine production, cell adhesion, leukocyte migration), as well as regulation of SMC biology and lipid signaling (Fig. 39A). A high number of proteins enriched in plaque-released MVs was involved in vesicle-mediated transport, thus pointing at their vesicular origin (Fig. 39A). This finding could also be connected with an overall higher level of MVs, as shown by FCM. Similar results were obtained after analysing significantly higher proteins in crude, distal plasma samples (Fig. 39D).

The majority of proteins higher in plaque-released MVs were mainly derived from plasma membrane, cytoplasmic vesicles and secretory granules that further corroborates their MV origin (Fig. 39B). Importantly, the results of cellular component analysis for crude plasma did not reveal vesicle-related terms for significant proteins in distal samples (Fig. 39E). This discrepancy underlines an advantage of vesicle isolation, instead analysing crude plasma samples, for studying MV-derived proteins, as the sensitivity of detecting the differences in protein concentration may be too low in crude plasma samples.

The cellular origin analysis revealed that proteins with significantly higher levels in plaque-released MVs were predominantly derived from macrophages, ECs, SMCs, neutrophils, platelets and other sources (Fig. 39C), similarly to the FCM results. Augmented concentration of macrophage-derived molecules in plaque-released MVs implies a significant role of these cells in progression of atherosclerosis.^{7,245} Analogous picture was presented by results from analysis of distal plasma samples (Fig. 39F).

In conclusion, we believe that the above results confirm that atherosclerotic plaque releases EVs that present a proatherogenic proteomic 'pattern' that represents the pathological molecular events within the cardiovascular system, as well as the health status of the patients.



Fig. 38 proteomic content comparison of isolated MVs and crude plasma samples. The numbers reflect significantly higher proteins (p<0.05), which are listed in the boxes. Protein abbreviations are explained in Table 1.



Fig. 39 Results of biological process, cellular compartment and tissue/cellular origin analyses for isolated MVs (left panel) and plasma (right panel). The graphs were created based on significantly higher proteins in distal samples, identified with proteomic panels. Biological process and cellular compartments analyses were performed with STRING. Tissue origin was assessed with The Human Protein Atlas and The Platelet Web.

Summary and Perspectives

The overall aim of this thesis was to investigate the role of MVs in the physiology and pathology of the cardiovascular system. To answer our questions, both clinical studies and *in vitro* experiments were performed.

In Paper I we showed that a novel method for vesicle isolation, acoustic trapping, is a valid method for MV isolation that could potentially be applied in clinical diagnostics. Furthermore, the results of the clinical study also demonstrated a protective effect of exercise on the vasculature, including decrease of MV release from the endothelium.

Paper II revealed that dietary supplementation with bilberry extract exhibit favourable effect on MI patient health status, by decreasing the concentration of platelet and endothelial MVs in blood. Additionally, we discovered that bilberry extract positively affects endothelial cells by reducing the vesiculation via several molecular mechanisms, connected with P2X₇ receptor. This study reports for the first time that dietary compounds have an effect on vesicle release, both *in vitro* and *in vivo*. These findings are important for potential future dietary recommendations for patients with CVD, as well as disease prevention with nutrition.

Paper III showed that there is a relationship between coronary flow reserve and MVs in blood. The low CFR group, thus the group with a decreased blood flow, exhibited significantly higher levels of platelet and endothelial MVs. The group was also characterized with increased concentration of several pro-atherosclerotic proteins in the vesicles. These results reflect the pathological status of cardiovascular system of the patients and have a potential clinical value as biomarkers of deterioration of endothelial function. It also raises the possibility that anti-BAFF therapeutics could be further explored as treatment against CVD.

In Study IV we demonstrated that atherosclerotic plaque releases MVs, following balloon angioplasty. The vesicles originate from several types of cells and present a pro-atherogenic 'pattern' of proteins. These results represent the pathological processes within the cardiovascular system that lead to the formation of atherosclerotic plaque.

The outcome of the studies included in this thesis points at the important role of MVs in the cardiovascular system. Study I and II underlined the importance of exercise and nutrition for prevention of CVD through decrease in MVs, whereas the

Western world is still characterized with physical inactivity and diet with low nutritional value. Study III and IV focused on potential application of MVs as biomarkers or targets of therapeutics, however it needs further, thorough research and possibly, personalized approach.

Acknowledgements

Gratitude is the beautiful form of happiness: this is its cause and its result.

Walter Dirks

Exactly six years ago I got a chance to take part in a long, exciting, sometimes exhausting, but very meaningful educational journey - I became a PhD student at Lund University.

It would not be possible without an approval of my supervisor, David Erlinge. It has been an honor and pleasure to be a part of the Molecular Cardiology lab and my PhD studies will always have a special place in my heart. I am enormously grateful for inviting me to Sweden and giving me this wonderful opportunity to develop as a researcher, but also as a person. This experience will always stay with me and I strongly believe it will bear fruit in many aspects of my life in the future, irrespective of which path I will choose. David, I would also like to thank you for your guidance throughout my studies, for your kindness, understanding and for the atmosphere that you create in the group. I consider myself lucky to get such interesting and fruitful projects and to collaborate with several creative researchers. I remember when I decided, a bit unexpectedly, to study biology. I had so many questions and there were so many things to explore. Thank you so much for allowing me to add my own, small piece of cobblestone to the road of knowledge, it is really meaningful to me.

This journey would not happen either without my second advisor, Björn Olde. It is really challenging to list everything that I would like to thank you for, Björn, so this time I will summarize it in a bit different way. A red-haired singer sang some time ago that *Girls just want to have fun* and I must say that I really had fun during my PhD studies, thanks to you.

I would like to cordially thank Siv Svensson who has been my guide at BMC during this whole time. Thank you, Siv, for a very warm welcome in the lab, for your help and advice. It was a pleasure to be your roommate and I am grateful for all for our conversations, including those about Swedish culture and places to visit.

I would also like to acknowledge Ramasri Sathanoori. Rama, it was an honor and pleasure to be your colleague. Thanks to your efforts I developed a lot as a researcher, both in terms of critical thinking and performing experiments. I am

enormously grateful for your involvement during my PhD studies and I will always remember that when you give someone your time, you are giving them a portion of your life that you will never get back. (...) That is why the greatest gift you can give someone is your time.

I would like to express my gratitude to all other members of the Molecular Cardiology lab, the Department of Cardiology, BMC D12 and Vesicle Lab Group. Kreema James, Mario Grossi, Olof Gidlöf, Selvi Celik, Neha Pimpalwar, Seher Alcevska, Maya Karbalaei Sadegh and Ihdina Sukma Dewi – it was a pleasure to meet and work with you. Thank you for your kindness and the atmosphere in the lab. Monica Magnusson, thank you so much for your help with everything connected with the defence organization and administrative issues. Rebecca Rylance, I am very grateful for your help with statistical problems and importantly, understanding statistics. Hooi Ching Lim and Anson Ku, I appreciate our nice conversations and good time in Toronto. I would also like to thank Anna, Emma and Elsa, students who visited the lab during my studies that I had a pleasure to introduce to the field of our research.

I am very grateful to all my collaborators: Acousort AB, especially Thomas Laurell and Mikael Evander, Ole Fröbert, Rikard Landberg, Cecilia Bergh, Lilith Arevström, Elmir Omerovic, Inger Haraldsson, Kristina Torngren, Sara Svedlund, Li-Ming Gan, Jonas Jögi, Mariam Al-Mashat, Lina Gefors, Lotta Cinthio and all Skåne University Hospital employees who were engaged in the projects included in this thesis, for their work and support with article reviews.

Last, but not least, I would like to thank the Polish Club: Joanna Daszkiewicz-Nilsson, Kasia Said-Hilmersson (Kasia 1), Kasia Zielińska (Kasia 2) and Kasia Krawczyk (Kasia 3), for the lovely atmosphere in BMC D12, our conversations about everything and, despite being so different, for getting along with so well. It was a pleasure to meet and share with you a part of my life. I wish you all the best and I hope our club will survive also outside the walls of BMC.

Podziękowania

Kiedy czasem zdam sobie sprawę z olbrzymich konsekwencji całkiem małych rzeczy... Nie mogę odeprzeć myśli... że małych rzeczy nie ma.

Bruce Barton

Myślę, że gdy wiele lat temu usłyszałam słowa: *Paulinka, pamiętaj, uczysz się dla siebie*, nikt nie pomyślałby, że sprawy zajdą tak daleko, że w wieku trzydziestu lat będę bronić doktoratu na Uniwersytecie w Lund w Szwecji. Niestety nie wszystkie moi bliscy doczekali tej chwili. Jest wiele bliskich mi osób, którym chciałabym podziękować za to, że jestem tu, gdzie jestem i muszę przyznać, że pisanie tych słów wiele dla mnie znaczy.

Jedząc owoc, pamiętaj o tym, kto zasadził drzewo.

przysłowie wietnamskie

Przede wszystkim dziękuję moim Rodzicom, którzy zawsze we mnie wierzyli, wspierali mnie i nigdy nie bali się moich, czasem dziwacznych, pomysłów na przyszły zawód. Od piosenkarki i weterynarza (era przedszkolna), przez filologa i teatrologa (gimnazjum), wreszcie psychologa (liceum). Na tym oczywiście nie poprzestałam przyprawiać ich o kołatanie serca, no bo po maturach postanowiłam jednak studiować biologię, no a potem biochemię, a jeszcze później w ogóle mi się zachciało jakiś kardiologicznych doktoratów ;) Nigdy nie usłyszałam od Was, że coś jest dla mnie za trudne, że nie warto próbować, a jednocześnie zawsze wykazywaliście zrozumienie i nie stawialiście poprzeczki zbyt wysoko. Dzięki temu jestem teraz szczęśliwym człowiekiem, założyłam wspaniałą rodzinę i cieszę się udanym życiem osobistym i zawodowym. Wasza rola w moim sukcesie jest nie do przecenienia.

Chciałabym również podziękować reszcie Rodziny, tej która jest i tej, której niestety już z nami nie ma. Dziękuję za to, że dzięki Wam zostałam niejako "zamarynowana" w akceptacji i poczuciu własnej wartości, a także tematach związanych z rozwojem, edukacją, a to wszystko podane w sosie z bliskich relacji i wsparcia. Moim Ciociom i Wujkom dziękuję przede wszystkim za wszystkie

rozmowy i wspólnie spędzony czas. Te pozornie drobne i nieistotne wydarzenia odegrały ważna rolę w tym, kim teraz jestem i jak patrzę na życie. Moim Kuzynom dziękuję za ciekawe rozmowy, motywację i świetny przykład. Nie szkodzi, że jednak nie udało Wam się wysłać mnie w kosmos, myślę, że i tak jest nieźle ;)

Bądźmy wdzięczni wobec ludzi, którzy czynią nas szczęśliwymi. Oni są uroczymi ogrodnikami, dzięki którym nasze dusze rozkwitają.

Marcel Proust

Drogi Piotrze, ciężko wyrazić mi słowami moją wdzięczność za towarzyszenie mi w tej sześcioletniej drodze. To dla mnie zaszczyt, że już od dziesięciu lat jesteś ze mną i wspierasz mnie we wszystkich aspektach życia. Dziękuję za Twój rozsądek, wrażliwość, zrozumienie oraz ogromne oddanie i miłość, których codziennie doświadczam. Dziękuję za to, że mnie inspirujesz i motywujesz, a także dbasz o równowagę zawodowo-osobistą w naszym życiu. Dziękuję Ci za to, że dzięki Twojemu zaangażowaniu i wsparciu mogłam realizować się zarówno rodzinnie, jak i naukowo.

Drogi Henryku, tak naprawdę byłeś przy mnie przez większość mojego doktoratu. Chciałabym Ci bardzo podziękować za Twój entuzjazm, dociekliwość, cudowną wyobraźnię, no i oczywiście miłość. Tak jak Twój Tata, jesteś niesamowicie inspirującą osobą i mam nadzieję, że Twoja radość życia dostarczy Ci wielu interesujących wrażeń i doświadczeń w życiu. I pamiętaj: *Don't grow up – it's a trap!*

References

- Joseph P, Leong D, McKee M, Anand SS, Schwalm J-D, Teo K, Mente A and Yusuf S, 2017. *Reducing the Global Burden of Cardiovascular Disease, Part 1*. Circulation Research 121:677-694.
- E. Olvera Lopez, B.D. Ballard and Jan A, 2020. *Cardiovascular Disease*, Treasure Island (FL): StatPearls Publishing, https://www.ncbi.nlm.nih.gov/books/NBK535419/.
- 3. Teixeira BC, Lopes AL, Macedo RCO, Correa CS, Ramis TR, Ribeiro JL and Reischak-Oliveira A, 2014. *Inflammatory markers, endothelial function and cardiovascular risk*. Jornal Vascular Brasileiro 13:108-115.
- 4. Libby P, Ridker PM and Hansson GK, 2011. *Progress and challenges in translating the biology of atherosclerosis*. Nature 473:317-325.
- 5. Ross R, 1999. *Atherosclerosis--an inflammatory disease*. The New England journal of medicine 340:115-126.
- 6. Schiro A, Wilkinson FL, Weston R, Smyth JV, Serracino-Inglott F and Alexander MY, 2014. *Endothelial microparticles as conveyors of information in atherosclerotic disease*. Atherosclerosis 234:295-302.
- 7. Epstein FH, 1999. *Atherosclerosis an inflammatory disease*. The New Egland Journal of Medicine 340:115-126.
- 8. Magnoni M, Ammirati E and Camici PG, 2015. *Non-invasive molecular imaging of vulnerable atherosclerotic plaques*. Journal of Cardiology 65:261-269.
- 9. Chhabra L, Zain MA and J. SW, 2020. *Angioplasty*, Treasure Island (FL): StatPearls Publishing, StatPearls [Internet].
- Van Horn L, Carson JAS, Appel LJ, Burke LE, Economos C, Karmally W, Lancaster K, Lichtenstein AH, Johnson RK, Thomas RJ, et al., 2016. Recommended Dietary Pattern to Achieve Adherence to the American Heart Association/American College of Cardiology (AHA/ACC) Guidelines: A Scientific Statement From the American Heart Association. Circulation 134.
- 11. Siervo M, Lara J, Chowdhury S, Ashor A, Oggioni C and Mathers JC, 2014. *Effects* of the Dietary Approach to Stop Hypertension (DASH) diet on cardiovascular risk factors: a systematic review and meta-analysis. British Journal of Nutrition 113:1-15.
- 12. Soltani S, Arablou T, Jayedi A and Salehi-Abargouei A, 2020. *Adherence to the dietary approaches to stop hypertension (DASH) diet in relation to all-cause and cause-specific mortality: a systematic review and dose-response meta-analysis of prospective cohort studies*. Nutrition Journal 19.

- Kim H, Caulfield LE, Garcia-Larsen V, Steffen LM, Coresh J and Rebholz CM, 2019. Plant-Based Diets Are Associated With a Lower Risk of Incident Cardiovascular Disease, Cardiovascular Disease Mortality, and All-Cause Mortality in a General Population of Middle-Aged Adults. Journal of the American Heart Association 8.
- 14. Satija A and Hu FB, 2018. *Plant-based diets and cardiovascular health*. Trends in Cardiovascular Medicine 28:437-441.
- 15. Widmer RJ, Flammer AJ, Lerman LO and Lerman A, 2015. *The Mediterranean Diet, its Components, and Cardiovascular Disease*. The American Journal of Medicine 128:229-238.
- 16. Martínez-González MA, Gea A and Ruiz-Canela M, 2019. *The Mediterranean Diet and Cardiovascular Health*. Circulation Research 124:779-798.
- Abdelhamid AS, Brown TJ, Brainard JS, Biswas P, Thorpe GC, Moore HJ, Deane KHO, AlAbdulghafoor FK, Summerbell CD, Worthington HV, et al., 2018. Omega-3 fatty acids for the primary and secondary prevention of cardiovascular disease. Cochrane Database of Systematic Reviews, DOI: 10.1002/14651858.CD003177.pub3.
- 18. Mozaffarian D and Wu JHY, 2011. *Omega-3 Fatty Acids and Cardiovascular Disease*. Journal of the American College of Cardiology 58:2047-2067.
- 19. Kromhout D, Yasuda S, Geleijnse JM and Shimokawa H, 2011. *Fish oil and omega-3 fatty acids in cardiovascular disease: do they really work?* European Heart Journal 33:436-443.
- 20. Colussi G, Catena C, Novello M, Bertin N and Sechi LA, 2017. *Impact of omega-3 polyunsaturated fatty acids on vascular function and blood pressure: Relevance for cardiovascular outcomes*. Nutrition, Metabolism and Cardiovascular Diseases 27:191-200.
- Bernasconi AA, Wiest MM, Lavie CJ, Milani RV and Laukkanen JA, 2021. Effect of Omega-3 Dosage on Cardiovascular Outcomes. Mayo Clinic Proceedings 96:304-313.
- 22. Rodriguez-Mateos A, Heiss C, Borges G and Crozier A, 2014. *Berry (poly)phenols and cardiovascular health.* J Agric Food Chem 62:3842-3851.
- 23. Erlund I, Koli R, Alfthan G, Marniemi J, Puukka P, Mustonen P, Mattila P and Jula A, 2008. *Favorable effects of berry consumption on platelet function, blood pressure and HDL cholesterol.* Am J Clin Nutr 87:323-331.
- 24. Rodriguez-Mateos A, Rendeiro C, Bergillos-Meca T, Tabatabaee S, George TW, Heiss C and Spencer JP, 2013. *Intake and time dependence of blueberry flavonoidinduced improvements in vascular function: a randomized, controlled, double-blind, crossover intervention study with mechanistic insights into biological activity.* Am J Clin Nutr 98:1179-1191.
- 25. Bornsek SM, Ziberna L, Polak T, Vanzo A, Ulrih NP, Abram V, Tramer F and Passamonti S, 2012. *Bilberry and blueberry anthocyanins act as powerful intracellular antioxidants in mammalian cells*. Food Chem 134:1878-1884.

- 26. Del Bo C, Riso P, Campolo J, Moller P, Loft S, Klimis-Zacas D, Brambilla A, Rizzolo A and Porrini M, 2013. *A single portion of blueberry (Vaccinium corymbosum L) improves protection against DNA damage but not vascular function in healthy male volunteers*. Nutr Res 33:220-227.
- 27. Park SH, Jeong SO, Chung HT and Pae HO, 2015. *Pterostilbene, an Active Constituent of Blueberries, Stimulates Nitric Oxide Production via Activation of Endothelial Nitric Oxide Synthase in Human Umbilical Vein Endothelial Cells.* Plant Foods Hum Nutr 70:263-268.
- 28. Vendrame S, Daugherty A, Kristo AS, Riso P and Klimis-Zacas D, 2013. *Wild blueberry (Vaccinium angustifolium) consumption improves inflammatory status in the obese Zucker rat model of the metabolic syndrome*. J Nutr Biochem 24:1508-1512.
- 29. Cutler BR, Petersen C and Anandh Babu PV, 2017. *Mechanistic insights into the vascular effects of blueberries: Evidence from recent studies*. Mol Nutr Food Res 61.
- Fiuza-Luces C, Santos-Lozano A, Joyner M, Carrera-Bastos P, Picazo O, Zugaza JL, Izquierdo M, Ruilope LM and Lucia A, 2018. *Exercise benefits in cardiovascular* disease: beyond attenuation of traditional risk factors. Nature Reviews Cardiology 15:731-743.
- 31. Ekelund U, Brown WJ, Steene-Johannessen J, Fagerland MW, Owen N, Powell KE, Bauman AE and Lee IM, 2019. *Do the associations of sedentary behaviour with cardiovascular disease mortality and cancer mortality differ by physical activity level? A systematic review and harmonised meta-analysis of data from 850 060 participants*. Br J Sports Med 53:886-894.
- 32. Jakovljevic DG, 2018. *Physical activity and cardiovascular aging: Physiological and molecular insights*. Experimental Gerontology 109:67-74.
- 33. Sharman JE, La Gerche A and Coombes JS, 2014. *Exercise and Cardiovascular Risk in Patients With Hypertension*. American Journal of Hypertension 28:147-158.
- 34. Hafiane A and Daskalopoulou SS, 2018. *Extracellular vesicles characteristics and emerging roles in atherosclerotic cardiovascular disease*. Metabolism 85:213-222.
- 35. Hargett LA and Bauer NN, 2013. On the Origin of Microparticles: From "Platelet Dust" to Mediators of Intercellular Communication. Pulmonary Circulation 3:329-340.
- 36. Jansen F, Nickenig G and Werner N, 2017. *Extracellular vesicles in cardiovascular disease*. Circulation Research 120:1649-1657.
- Deng F, Wang S, Xu R, Yu W, Wang X and Zhang L, 2018. Endothelial microvesicles in hypoxic hypoxia diseases. J Cell Mol Med, DOI: 10.1111/jcmm.13671.
- 38. Badimon L, Suades R, Fuentes E, Palomo I and Padro T, 2016. *Role of Platelet-Derived Microvesicles As Crosstalk Mediators in Atherothrombosis and Future Pharmacology Targets: A Link between Inflammation, Atherosclerosis, and Thrombosis*. Front Pharmacol 7:293.
- 39. Boulanger CM, Loyer X, Rautou P-E and Amabile N, 2017. *Extracellular vesicles in coronary artery disease*. Nature Reviews Cardiology 14:259-272.
- 40. Mause SF and Weber C, 2010. *Microparticles: protagonists of a novel communication network for intercellular information exchange*. Circ Res 107:1047-1057.
- 41. El Andaloussi S, Mager I, Breakefield XO and Wood MJ, 2013. *Extracellular vesicles: biology and emerging therapeutic opportunities*. Nat Rev Drug Discov 12:347-357.
- 42. Milioli M, Ibanez-Vea M, Sidoli S, Palmisano G, Careri M and Larsen MR, 2015. *Quantitative proteomics analysis of platelet-derived microparticles reveals distinct protein signatures when stimulated by different physiological agonists.* J Proteomics 121:56-66.
- 43. Haas B, Serchi T, Wagner DR, Gilson G, Planchon S, Renaut J, Hoffmann L, Bohn T and Devaux Y, 2011. *Proteomic analysis of plasma samples from patients with acute myocardial infarction identifies haptoglobin as a potential prognostic biomarker*. J Proteomics 75:229-236.
- 44. Bosman GJ, Lasonder E, Groenen-Dopp YA, Willekens FL and Werre JM, 2012. *The proteome of erythrocyte-derived microparticles from plasma: new clues for erythrocyte aging and vesiculation.* J Proteomics 76 Spec No.:203-210.
- 45. Chaichompoo P, Kumya P, Khowawisetsut L, Chiangjong W, Chaiyarit S, Pongsakul N, Sirithanaratanakul N, Fucharoen S, Thongboonkerd V and Pattanapanyasat K, 2012. Characterizations and proteome analysis of platelet-free plasma-derived microparticles in beta-thalassemia/hemoglobin E patients. J Proteomics 76 Spec No.:239-250.
- 46. Gidlöf O, van der Brug M, Öhman J, Gilje P, Olde B, Wahlestedt C and Erlinge D, 2013. *Platelets activated during myocardial infarction release functional miRNA, which can be taken up by endothelial cells and regulate ICAM1 expression*. Blood 121:3908-3917.
- 47. Zhang Y, Zhang L, Wang Y, Ding H, Xue S, Qi H and Li P, 2019. *MicroRNAs or long noncoding RNAs in diagnosis and prognosis of coronary artery disease*. Aging Dis 10:353-366.
- 48. Gidlof O, Evander M, Rezeli M, Marko-Varga G, Laurell T and Erlinge D, 2019. Proteomic profiling of extracellular vesicles reveals additional diagnostic biomarkers for myocardial infarction compared to plasma alone. Sci Rep 9:8991.
- 49. Bryl-Gorecka P, Sathanoori R, Al-Mashat M, Olde B, Jogi J, Evander M, Laurell T and Erlinge D, 2018. *Effect of exercise on the plasma vesicular proteome: a methodological study comparing acoustic trapping and centrifugation*. Lab Chip 18:3101-3111.
- 50. Panagiotou N, Neytchev O, Selman C and Shiels PG, 2018. *Extracellular vesicles, ageing, and therapeutic interventions*. Cells 7.
- 51. Eichner NZM, Erdbrugger U and Malin SK, 2018. *Extracellular vesicles: a novel target for exercise-mediated reductions in type 2 diabetes and cardiovascular disease risk.* J Diabetes Res 2018:7807245.
- 52. Hu SS, Zhang HG, Zhang QJ and Xiu RJ, 2014. *Small-size circulating endothelial microparticles in coronary artery disease*. PLoS One 9:e104528.

- 53. Mallat Z, Hugel B, Ohan J, Leseche G, Freyssinet JM and Tedgui A, 1999. *Shed membrane microparticles with procoagulant potential in human atherosclerotic plaques*. Circulation 99:348-353.
- 54. Helbing T, Olivier C, Bode C, Moser M and Diehl P, 2014. *Role of microparticles in endothelial dysfunction and arterial hypertension*. World J Cardiol 6:1135-1139.
- 55. Jy W, Jimenez JJ, Mauro LM, Horstman LL, Cheng P, Ahn CR, Bidot CJ and Ahn YS, 2004. *Endothelial microparticles induce formation of platelet aggregates via a von Willebrand factor/ristocetin dependent pathway, rendering them resistant to dissociation*. J Thromb Haemost 3:1301-1308.
- 56. Curtis AM, Edelberg J, Jonas R, Rogers WT, Moore JS, Syed W and Mohler ER, 3rd, 2013. *Endothelial microparticles: sophisticated vesicles modulating vascular function*. Vasc Med 18:204-214.
- 57. Buendia P, Montes de Oca A, Madueno JA, Merino A, Martin-Malo A, Aljama P, Ramirez R, Rodriguez M and Carracedo J, 2015. *Endothelial microparticles mediate inflammation-induced vascular calcification*. FASEB J 29:173-181.
- 58. Momen-Heravi F, Balaj L, Alian S, Mantel P-Y, Halleck AE, Trachtenberg AJ, Soria CE, Oquin S, Bonebreak CM, Saracoglu E, *et al.*, 2013. *Current methods for the isolation of extracellular vesicles*. Biological Chemistry 394:1253-1262.
- 59. Rito-Palomares M, Baranyai T, Herczeg K, Onódi Z, Voszka I, Módos K, Marton N, Nagy G, Mäger I, Wood MJ, et al., 2015. Isolation of Exosomes from Blood Plasma: Qualitative and Quantitative Comparison of Ultracentrifugation and Size Exclusion Chromatography Methods. Plos One 10:e0145686.
- 60. Lobb RJ, Becker M, Wen Wen S, Wong CSF, Wiegmans AP, Leimgruber A and Möller A, 2015. *Optimized exosome isolation protocol for cell culture supernatant and human plasma*. Journal of Extracellular Vesicles 4:27031.
- 61. Chandler WL, 2013. *Microparticle counts in platelet-rich and platelet-free plasma, effect of centrifugation and sample-processing protocols*. Blood Coagul Fibrinolysis 24:125-132.
- 62. Linares R, Tan S, Gounou C, Arraud N and Brisson AR, 2015. *High-speed centrifugation induces aggregation of extracellular vesicles*. Journal of Extracellular Vesicles 4:29509.
- 63. Yuana Y, Levels J, Grootemaat A, Sturk A and Nieuwland R, 2014. *Co-isolation of extracellular vesicles and high-density lipoproteins using density gradient ultracentrifugation*. Journal of Extracellular Vesicles 3:23262.
- 64. Contreras-Naranjo JC, Wu H-J and Ugaz VM, 2017. *Microfluidics for exosome isolation and analysis: enabling liquid biopsy for personalized medicine*. Lab on a Chip 17:3558-3577.
- 65. Lee K, Shao H, Weissleder R and Lee H, 2015. *Acoustic Purification of Extracellular Microvesicles*. ACS Nano 9:2321-2327.
- 66. Liu F, Vermesh O, Mani V, Ge TJ, Madsen SJ, Sabour A, Hsu E-C, Gowrishankar G, Kanada M, Jokerst JV, *et al.*, 2017. *The Exosome Total Isolation Chip.* ACS Nano 11:10712-10723.
- 67. Jeong S, Park J, Pathania D, Castro CM, Weissleder R and Lee H, 2016. *Integrated Magneto–Electrochemical Sensor for Exosome Analysis*. ACS Nano 10:1802-1809.

- 68. Muller G, 2015. *Novel Tools for the Study of Cell Type-Specific Exosomes and Microvesicles*. Journal of Bioanalysis & Biomedicine 04.
- 69. Chandler WL, 2016. *Measurement of microvesicle levels in human blood using flow cytometry*. Cytometry Part B: Clinical Cytometry 90:326-336.
- 70. Mobarrez F, Antovic J, Egberg N, Hansson M, Jörneskog G, Hultenby K and Wallén H, 2010. *A multicolor flow cytometric assay for measurement of platelet-derived microparticles*. Thrombosis Research 125:e110-e116.
- 71. Morales-Kastresana A, Telford B, Musich TA, McKinnon K, Clayborne C, Braig Z, Rosner A, Demberg T, Watson DC, Karpova TS, *et al.*, 2017. *Labeling Extracellular Vesicles for Nanoscale Flow Cytometry*. Scientific Reports 7.
- 72. Gasecka A, Nieuwland R, Budnik M, Dignat-George F, Eyileten C, Harrison P, Lacroix R, Leroyer A, Opolski G, Pluta K, *et al.*, 2020. *Ticagrelor attenuates the increase of extracellular vesicle concentrations in plasma after acute myocardial infarction compared to clopidogrel.* Journal of Thrombosis and Haemostasis 18:609-623.
- 73. Rosińska J, Ambrosius W, Maciejewska J, Narożny R, Kozubski W and Łukasik M, 2019. Association of platelet-derived microvesicles and their phenotypes with carotid atherosclerosis and recurrent vascular events in patients after ischemic stroke. Thrombosis Research 176:18-26.
- 74. Headland SE, Jones HR, D'Sa ASV, Perretti M and Norling LV, 2014. *Cutting-Edge Analysis of Extracellular Microparticles using ImageStreamX Imaging Flow Cytometry*. Scientific Reports 4.
- 75. Erdbrügger U, Rudy CK, E. Etter M, Dryden KA, Yeager M, Klibanov AL and Lannigan J, 2014. *Imaging flow cytometry elucidates limitations of microparticle analysis by conventional flow cytometry*. Cytometry Part A 85:756-770.
- 76. Midekessa G, Godakumara K, Ord J, Viil J, Lättekivi F, Dissanayake K, Kopanchuk S, Rinken A, Andronowska A, Bhattacharjee S, et al., 2020. Zeta Potential of Extracellular Vesicles: Toward Understanding the Attributes that Determine Colloidal Stability. ACS Omega 5:16701-16710.
- 77. Arraud N, Linares R, Tan S, Gounou C, Pasquet JM, Mornet S and Brisson AR, 2014. *Extracellular vesicles from blood plasma: determination of their morphology, size, phenotype and concentration*. Journal of Thrombosis and Haemostasis 12:614-627.
- Ashcroft AA, de Sonneville J, Yuana Y, Ostanto S, Bertina RM, Kuil ME and Oosterkamp TH, 2012. Determination of the size distribution of blood microparticles directly in plasma using atomic force microscopy and microfluidics. Biomed Microdevices 14:641-649.
- 79. Vogel R, Coumans FAW, Maltesen RG, Böing AN, Bonnington KE, Broekman ML, Broom MF, Buzás EI, Christiansen G, Hajji N, et al., 2016. A standardized method to determine the concentration of extracellular vesicles using tunable resistive pulse sensing. Journal of Extracellular Vesicles 5:31242.
- 80. Dragovic RA, Gardiner C, Brooks AS, Tannetta DS, Ferguson DJ, Hole P, Carr B, Redman CW, Harris AL, Dobson PJ, *et al.*, 2011. *Sizing and phenotyping of cellular vesicles using Nanoparticle Tracking Analysis*. Nanomedicine 7:780-788.

- 81. Gardiner C, Ferreira YJ, Dragovic RA, Redman CWG and Sargent IL, 2013. *Extracellular vesicle sizing and enumeration by nanoparticle tracking analysis.* Journal of Extracellular Vesicles 2:19671.
- 82. van der Pol E, Coumans F, Varga Z, Krumrey M and Nieuwland R, 2013. *Innovation in detection of microparticles and exosomes*. Journal of Thrombosis and Haemostasis 11:36-45.
- 83. Marfin YS, Aleksakhina EL, Merkushev DA, Rumyantsev EV and Tomilova IK, 2015. *Interaction of BODIPY Dyes with the Blood Plasma Proteins*. Journal of Fluorescence 26:255-261.
- 84. Corcoran D, Berry C and Oldroyd K, 2015. *Current frontiers in the clinical research of coronary physiology*. Interv. Cardiol. 7:1-11.
- 85. Gan LM, Svedlund S, Wittfeldt A, Eklund C, Gao S, Matejka G, Jeppsson A, Albertsson P, Omerovic E and Lerman A, 2017. *Incremental value of transthoracic doppler echocardiography-assessed coronary flow reserve in patients with suspected myocardial ischemia undergoing myocardial perfusion scintigraphy*. J Am Heart Assoc 6.
- Cortigiani L, Rigo F, Gherardi S, Bovenzi F, Molinaro S, Picano E and Sicari R, 2012. Coronary flow reserve during dipyridamole stress echocardiography predicts mortality. JACC Cardiovasc Imaging 5:1079-1085.
- 87. Murthy VL, Naya M, Foster CR, Hainer J, Gaber M, Di Carli G, Blankstein R, Dorbala S, Sitek A, Pencina MJ, et al., 2011. *Improved cardiac risk assessment with noninvasive measures of coronary flow reserve*. Circulation 124:2215-2224.
- 88. Murthy VL, Naya M, Foster CR, Gaber M, Hainer J, Klein J, Dorbala S, Blankstein R and Di Carli MF, 2012. *Association between coronary vascular dysfunction and cardiac mortality in patients with and without diabetes mellitus*. Circulation 126:1858-1868.
- Sato A, Terata K, Miura H, Toyama K, Loberiza FR, Hatoum OA, Saito T, Sakuma I and Gutterman DD, 2005. *Mechanism of vasodilation to adenosine in coronary arterioles from patients with heart disease*. American Journal of Physiology-Heart and Circulatory Physiology 288:H1633-H1640.
- 90. Kaur H, Carvalho J, Looso M, Singh P, Chennupati R, Preussner J, Günther S, Albarrán-Juárez J, Tischner D, Classen S, *et al.*, 2017. *Single-cell profiling reveals heterogeneity and functional patterning of GPCR expression in the vascular system*. Nature Communications 8.
- 91. Evander M, Gidlof O, Olde B, Erlinge D and Laurell T, 2015. *Non-contact acoustic capture of microparticles from small plasma volumes*. Lab Chip 15:2588-2596.
- 92. Chen YW, Chen JK and Wang JS, 2010. *Strenuous exercise promotes shear-induced thrombin generation by increasing the shedding of procoagulant microparticles from platelets*. Thromb Haemost 104:293-301.
- 93. Chen YW, Chen YC and Wang JS, 2013. *Absolute hypoxic exercise training* enhances in vitro thrombin generation by increasing procoagulant platelet-derived microparticles under high shear stress in sedentary men. Clin Sci (Lond) 124:639-649.

- 94. Guiraud T, Gayda M, Juneau M, Bosquet L, Meyer P, Theberge-Julien G, Galinier M, Nozza A, Lambert J, Rheaume E, et al., 2013. A single bout of high-intensity interval exercise does not increase endothelial or platelet microparticles in stable, physically fit men with coronary heart disease. Can J Cardiol 29:1285-1291.
- 95. Babbitt DM, Diaz KM, Feairheller DL, Sturgeon KM, Perkins AM, Veerabhadrappa P, Williamson ST, Kretzschmar J, Ling C, Lee H, et al., 2013. Endothelial activation microparticles and inflammation status improve with exercise training in african americans. Int J Hypertens 2013:538017.
- 96. Kim JS, Kim B, Lee H, Thakkar S, Babbitt DM, Eguchi S, Brown MD and Park JY, 2015. *Shear stress-induced mitochondrial biogenesis decreases the release of microparticles from endothelial cells*. Am J Physiol Heart Circ Physiol 309:H425-433.
- 97. Fruhbeis C, Helmig S, Tug S, Simon P and Kramer-Albers EM, 2015. *Physical exercise induces rapid release of small extracellular vesicles into the circulation*. J Extracell Vesicles 4:28239.
- 98. Wahl P, Jansen F, Achtzehn S, Schmitz T, Bloch W, Mester J and Werner N, 2014. *Effects of high intensity training and high volume training on endothelial microparticles and angiogenic growth factors*. PLoS One 9:e96024.
- 99. Whitham M, Parker BL, Friedrichsen M, Hingst JR, Hjorth M, Hughes WE, Egan CL, Cron L, Watt KI, Kuchel RP, et al., 2018. Extracellular Vesicles Provide a Means for Tissue Crosstalk during Exercise. Cell Metabolism 27:237-251.e234.
- 100. Wilhelm EN, Mourot L and Rakobowchuk M, 2018. *Exercise-Derived Microvesicles: A Review of the Literature*. Sports Medicine 48:2025-2039.
- 101. Warburton DE, Nicol CW and Bredin SS, 2006. *Health benefits of physical activity: the evidence*. CMAJ 174:801-809.
- 102. Sun MW, Zhong MF, Gu J, Qian FL, Gu JZ and Chen H, 2008. *Effects of different levels of exercise volume on endothelium-dependent vasodilation: roles of nitric oxide synthase and heme oxygenase*. Hypertens Res 31:805-816.
- 103. Terra R, Goncalves da Silva SA, Salerno Pinto V and Lourenco Dutra PM, 2012. *Effect of exercise on the immune system: response, adaptation and cell signaling.* Rev Bras Med Esporte 18:208-214.
- 104. Walsh NP, Gleeson M, Pyne DB, Nieman DC, Dhabhar FS, Shephard RJ, Oliver SJ, Bermon S and Kajaniene A, 2011. *Position Statement Part two: Maintaining immune health.* Exerc Immunol Rev:64-103.
- 105. Walsh NP, Gleeson M, Pyne DB, Nieman DC, Dhabhar FS, Shephard RJ, Oliver SJ, Bermon S and Kajaniene A, 2011. *Position statement Part one: Immune function end exercise*. Exerc Immunol Rev:6-63.
- 106. Pedersen L, Pilegaard H, Hansen J, Brandt C, Adser H, Hidalgo J, Olesen J, Pedersen BK and Hojman P, 2011. *Exercise-induced liver chemokine CXCL-1 expression is linked to muscle-derived interleukin-6 expression*. J Physiol 589:1409-1420.
- 107. Ulven SM, Foss SS, Skjolsvik AM, Stadheim HK, Myhrstad MC, Raael E, Sandvik M, Narverud I, Andersen LF, Jensen J, et al., 2015. An acute bout of exercise modulate the inflammatory response in peripheral blood mononuclear cells in healthy young men. Arch Physiol Biochem 121:41-49.

- 108. Timmons JA, Jansson E, Fischer H, Gustafsson T, Greenhaff PL, Ridden J, Rachman J and Sundberg CJ, 2005. Modulation of extracellular matrix genes reflects the magnitude of physiological adaptation to aerobic exercise training in humans. BMC Biol 3:19.
- 109. Peake J, Wilson G, Mackinnon L and Coombes JS, 2005. *Carbohydrate* supplementation and alterations in neutrophils, and plasma cortisol and myoglobin concentration after intense exercise. Eur J Appl Physiol 93:672-678.
- 110. Hansen J, Brandt C, Nielsen AR, Hojman P, Whitham M, Febbraio MA, Pedersen BK and Plomgaard P, 2011. Exercise induces a marked increase in plasma follistatin: evidence that follistatin is a contraction-induced hepatokine. Endocrinology 152:164-171.
- 111. Thompson HS, Scordilis SP, Clarkson PM and Lohrer WA, 2001. A single bout of eccentric exercise increases HSP27 and HSC/HSP70 in human skleletal muscle. Acta Physiol Scand 171:187-193.
- 112. van Loon JE, Sonneveld MA, Praet SF, de Maat MP and Leebeek FW, 2014. Performance related factors are the main determinants of the von Willebrand factor response to exhaustive physical exercise. PLoS One 9:e91687.
- 113. Claus RA, Bockmeyer CL, Sossdorf M, Lösche W and Hilberg T, 2006. *Physical stress as a model to study variations in ADAMTS-13 activity, von Willebrand factor level and platelet activation.* J Thromb Haemost 4:902-905.
- 114. Wang JS, 2004. Intense exercise increases shear-induced platelet aggregation in men through enhancement of von Willbrand factor binding, glycoprotein IIb/IIIa activation, and P-selectin expression on platelets. Eur J Appl Physiol 91:741-747.
- 115. Landers-Ramos RQ, Jenkins NT, Spangenburg EE, Hagberg JM and Prior SJ, 2014. *Circulating angiogenic and inflammatory cytokine responses to acute aerobic exercise in trained and sedentary young men.* Eur J Appl Physiol 114:1377-1384.
- 116. Roupas ND, Mamali I, Maragkos S, Leonidou L, Armeni AK, Markantes GK, Tsekouras A, Sakellaropoulos GC, Markou KB and Georgopoulos NA, 2013. The effect of prolonged aerobic exercise on serum adipokine levels during an ultramarathon endurance race. Hormones 12:275-282.
- 117. Sharafi H and Rahimi R, 2012. *The effect of resistance exercise on p53, caspase-9, and caspase-3 in trained and untrained men.* Journal of strength and conditioning research 26:1142-1148.
- Robson-Ansley P, Cockburn E, Walshe I, Stevenson E and Nimmo M, in *Exercise Immunology Review*, Association for the Advancement of Sports Medicine, 2010, vol. 16, pp. 56-76.
- 119. Kalra H, Simpson RJ, Hong J and Aikawa E, 2012. Vesiclepedia: a compendium for extracellular vesicles with continuous community annotation. PLOS Biology 10:1-5.
- 120. Safdar A and Tarnopolsky MA, 2018. *Exosomes as Mediators of the Systemic Adaptations to Endurance Exercise*. Cold Spring Harbor Perspectives in Medicine 8:a029827.
- 121. Highton PJ, Martin N, Smith AC, Burton JO and Bishop NC, 2018. *Microparticles and exercise in clinical populations*. Exerc Immunol Rev 24:46-58.

- 122. Trovato E, Di Felice V and Barone R, 2019. *Extracellular Vesicles: Delivery Vehicles of Myokines*. Frontiers in Physiology 10.
- 123. Vechetti IJ, Valentino T, Mobley CB and McCarthy JJ, 2020. *The role of extracellular vesicles in skeletal muscle and systematic adaptation to exercise*. The Journal of Physiology, DOI: 10.1113/jp278929.
- 124. Brahmer A, Neuberger E, Esch-Heisser L, Haller N, Jorgensen MM, Baek R, Möbius W, Simon P and Krämer-Albers E-M, 2019. *Platelets, endothelial cells and leukocytes contribute to the exercise-triggered release of extracellular vesicles into the circulation*. Journal of Extracellular Vesicles 8:1615820.
- 125. Rigamonti AE, Bollati V, Pergoli L, Iodice S, De Col A, Tamini S, Cicolini S, Tringali G, De Micheli R, Cella SG, et al., 2019. Effects of an acute bout of exercise on circulating extracellular vesicles: tissue-, sex-, and BMI-related differences. International Journal of Obesity 44:1108-1118.
- 126. Arevstrom L, Bergh C, Landberg R, Wu H, Rodriguez-Mateos A, Waldenborg M, Magnuson A, Blanc S and Frobert O, 2019. *Freeze-dried bilberry (Vaccinium myrtillus) dietary supplement improves walking distance and lipids after myocardial infarction: an open-label randomized clinical trial.* Nutr Res 62:13-22.
- 127. Thompson K, Hosking H, Pederick W, Singh I and Santhakumar AB, 2017. *The effect of anthocyanin supplementation in modulating platelet function in sedentary population: a randomised, double-blind, placebo-controlled, cross-over trial.* Br J Nutr 118:368-374.
- 128. Song F, Zhu Y, Shi Z, Tian J, Deng X, Ren J, Andrews MC, Ni H, Ling W and Yang Y, 2014. *Plant food anthocyanins inhibit platelet granule secretion in hypercholesterolaemia: Involving the signalling pathway of PI3K-Akt.* Thromb Haemost 112:981-991.
- 129. Maestro A, Terdoslavich M, Vanzo A, Kuku A, Tramer F, Nicolin V, Micali F, Decorti G and Passamonti S, 2010. *Expression of bilitranslocase in the vascular endothelium and its function as a flavonoid transporter*. Cardiovasc Res 85:175-183.
- 130. Speciale A, Cimino F, Saija A, Canali R and Virgili F, 2014. *Bioavailability and molecular activities of anthocyanins as modulators of endothelial function*. Genes Nutr 9:404.
- 131. Morelli A, Chiozzi P, Chiesa A, Ferrari D, Sanz JM, Falzoni S, Pinton P, Rizutto R, Olson MF and Di Virgilio F, 2003. *Extracellular ATP causes ROCK-I-dependent bleb formation in P2X7-transfected HEK293 cells*. Molecular Biology of the Cell 14:2655-2664.
- 132. Li J, Li X, Jiang X, Yang M, Yang R, Burnstock G, Xiang Z and Yuan H, 2017. Microvesicles shed from microglia activated by the P2X7-p38 pathway are involved in neuropathic pain induced by spinal nerve ligation in rats. Purinergic Signal 13:13-26.
- 133. Sathanoori R, Sward K, Olde B and Erlinge D, 2015. *The ATP Receptors P2X7 and P2X4 Modulate High Glucose and Palmitate-Induced Inflammatory Responses in Endothelial Cells.* PLoS One 10:e0125111.

- 134. Jacques-Silva MC, Rodnight R, Lenz G, Liao Z, Kong Q, Tran M, Kang Y, Gonzalez FA, Weisman GA and Neary JT, 2004. *P2X7 receptors stimulate AKT phosphorylation in astrocytes*. Br J Pharmacol 141:1106-1117.
- 135. Kostova EB, Beuger BM, Klei TR, Halonen P, Lieftink C, Beijersbergen R, van den Berg TK and van Bruggen R, 2015. *Identification of signalling cascades involved in red blood cell shrinkage and vesiculation*. Biosci Rep 35.
- 136. Ahmet I, Spangler E, Shukitt-Hale B, Joseph JA, Ingram DK and Talan M, 2009. Survival and cardioprotective benefits of long-term blueberry enriched diet in dilated cardiomyopathy following myocardial infarction in rats. PLoS One 4:e7975.
- 137. Lim W, Jeong W and Song G, 2016. *Delphinidin suppresses proliferation and migration of human ovarian clear cell carcinoma cells through blocking AKT and ERK1/2 MAPK signaling pathways*. Mol Cell Endocrinol 422:172-181.
- 138. Hao J, Du H, Li W, Liu F, Lu J, Yang X and Cui W, 2016. *Anthocyanins protected hearts against ischemic injury by reducing MMP-2 activity via Akt/P38 pathways*. Am J Transl Res 8:1100-1107.
- Ogawa K, Tsuruma K, Tanaka J, Kakino M, Kobayashi S, Shimazawa M and Hara H, 2013. The protective effects of bilberry and lingonberry extracts against UV lightinduced retinal photoreceptor cell damage in vitro. J Agric Food Chem 61:10345-10353.
- 140. Gomez-Villafuertes R, Garcia-Huerta P, Diaz-Hernandez JI and Miras-Portugal MT, 2015. PI3K/Akt signaling pathway triggers P2X7 receptor expression as a prosurvival factor of neuroblastoma cells under limiting growth conditions. Sci Rep 5:18417.
- 141. Salvestrini V, Zini R, Rossi L, Gulinelli S, Manfredini R, Bianchi E, Piacibello W, Caione L, Migliardi G, Ricciardi MR, et al., 2012. Purinergic signaling inhibits human acute myeloblastic leukemia cell proliferation, migration, and engraftment in immunodeficient mice. Blood 119:217-226.
- 142. Ostrowski M, Carmo NB, Krumeich S, Fanget I, Raposo G, Savina A, Moita CF, Schauer K, Hume AN, Freitas RP, *et al.*, 2010. *Rab27a and Rab27b control different steps of the exosome secretion pathway*. Nat Cell Biol 12:19-30; sup pp 11-13.
- 143. Bianco F, Perrotta C, Novellino L, Francolini M, Riganti L, Menna E, Saglietti L, Schuchman EH, Furlan R, Clementi E, *et al.*, 2009. *Acid sphingomyelinase activity triggers microparticle release from glial cells*. EMBO J 28:1043-1054.
- 144. Muralidharan-Chari V, Clancy J, Plou C, Romao M, Chavrier P, Raposo G and D'Souza-Schorey C, 2009. *ARF6-regulated shedding of tumor cell-derived plasma membrane microvesicles*. Curr Biol 19:1875-1885.
- 145. Rothmeier AS, Marchese P, Langer F, Kamikubo Y, Schaffner F, Cantor J, Ginsberg MH, Ruggeri ZM and Ruf W, 2017. *Tissue Factor Prothrombotic Activity Is Regulated by Integrin-arf6 Trafficking*. Arterioscler Thromb Vasc Biol 37:1323-1331.
- 146. Edwards M, Czank C, Woodward GM, Cassidy A and Kay CD, 2015. *Phenolic metabolites of anthocyanins modulate mechanisms of endothelial function*. J Agric Food Chem 63:2423-2431.

- 147. Haraldsson I, Gan L-M, Svedlund S, Torngren K, Westergren HU, Redfors B, Lagerström-Fermer M, Angerås O, Råmunddal T, Petursson P, et al., 2019. PROspective evaluation of coronary FLOW reserve and molecular biomarkers in patients with established coronary artery disease the PROFLOW-trial: crosssectional evaluation of coronary flow reserve. Vascular Health and Risk Management 15:375-384.
- 148. Natorska J, Marek G, Sadowski J and Undas A, 2016. *Presence of B cells within aortic valves in patients with aortic stenosis: Relation to severity of the disease*. J Cardiol 67:80-85.
- 149. Jansen F, Li Q, Pfeifer A and Werner N, 2017. Endothelial- and immune cell-derived extracellular vesicles in the regulation of cardiovascular health and disease. JACC: Basic to Translational Science 2:790-807.
- 150. Ait-Oufella H, Herbin O, Bouaziz JD, Binder CJ, Uyttenhove C, Laurans L, Taleb S, Van Vre E, Esposito B, Vilar J, et al., 2010. B cell depletion reduces the development of atherosclerosis in mice. J Exp Med 207:1579-1587.
- 151. Brand K, Page S, Rogler G, Bartsch A, Brandl R, Knuechel R, Page M, Kaltschmidt C, Baeuerle PA and Neumeier D, 1996. *Activated transcription factor nuclear factor-kappa B is present in the atherosclerotic lesion*. J Clin Invest 97:1715-1722.
- 152. Brand K, Page S, Walli A, Neumeier D and Baeuerle PA, 1997. *Role of Nuclear Factor-KB in Atherogenesis*. Experimental Physiology 82:297-304.
- 153. Parreno E, Palomares C, Martinez M, Ballester R, Martinez L and Fornovi A, 2018. *Resistin and Cardiovascular Disease.* Journal of Cardiovascular Diseases & Diagnosis 06.
- 154. Gencer B, Auer R, de Rekeneire N, Butler J, Kalogeropoulos A, Bauer DC, Kritchevsky SB, Miljkovic I, Vittinghoff E, Harris T, et al., 2016. Association between resistin levels and cardiovascular disease events in older adults: The health, aging and body composition study. Atherosclerosis 245:181-186.
- 155. Fontana A, Spadaro S, Copetti M, Spoto B, Salvemini L, Pizzini P, Frittitta L, Mallamaci F, Pellegrini F, Trischitta V, et al., 2015. Association between resistin levels and all-cause and cardiovascular mortality: a new study and a systematic review and meta-analysis. PLoS One 10:e0120419.
- 156. Muse ED, Feldman DI, Blaha MJ, Dardari ZA, Blumenthal RS, Budoff MJ, Nasir K, Criqui MH, Cushman M, McClelland RL, et al., 2015. The association of resistin with cardiovascular disease in the Multi-Ethnic Study of Atherosclerosis. Atherosclerosis 239:101-108.
- 157. Park HK, Kwak MK, Kim HJ and Ahima RS, 2017. *Linking resistin, inflammation, and cardiometabolic diseases.* Korean J Intern Med 32:239-247.
- 158. Cosemans JM, Van Kruchten R, Olieslagers S, Schurgers LJ, Verheyen FK, Munnix IC, Waltenberger J, Angelillo-Scherrer A, Hoylaerts MF, Carmeliet P, et al., 2010. Potentiating role of Gas6 and Tyro3, Axl and Mer (TAM) receptors in human and murine platelet activation and thrombus stabilization. J Thromb Haemost 8:1797-1808.
- 159. Caldentey G, De Frutos P, Cristóbal H, Garabito M, Berruezo A, Bosch X, San Antonio R, Flores-Umanzor E, Perea RJ, De Caralt TM, *et al.*, 2017. *Serum levels of*

GrowthAarrest-Specific 6 protein and soluble AXL in patients with ST-segment elevation myocardial infarction. European Heart Journal: Acute Cardiovascular Care:1-9.

- 160. Segev A, Nili N and Strauss B, 2004. *The role of perlecan in arterial injury and angiogenesis*. Cardiovascular Research 63:603–610.
- 161. Guo XH, Liu LX, Zhang HY, Zhang QQ, Li Y, Tian XX and Qiu ZH, 2014. *Insulinlike growth factor binding protein-related protein 1 contributes to hepatic fibrogenesis.* J Dig Dis 15:202-210.
- 162. Moreno JA, Munoz-Garcia B, Martin-Ventura JL, Madrigal-Matute J, Orbe J, Paramo JA, Ortega L, Egido J and Blanco-Colio LM, 2009. *The CD163-expressing macrophages recognize and internalize TWEAK: potential consequences in atherosclerosis*. Atherosclerosis 207:103-110.
- 163. Aristoteli LP, Moller HJ, Bailey B, Moestrup SK and Kritharides L, 2006. *The* monocytic lineage specific soluble CD163 is a plasma marker of coronary atherosclerosis. Atherosclerosis 184:342-347.
- 164. Samy E, Wax S, Huard B, Hess H and Schneider P, 2017. *Targeting BAFF and APRIL in systemic lupus erythematosus and other antibody-associated diseases*. Int Rev Immunol 36:3-19.
- 165. Kyaw T, Cui P, Tay C, Kanellakis P, Hosseini H, Liu E, Rolink AG, Tipping P, Bobik A and Toh BH, 2013. BAFF receptor mAb treatment ameliorates development and progression of atherosclerosis in hyperlipidemic ApoE(-/-) mice. PLoS One 8:e60430.
- 166. Zouggari Y, Ait-Oufella H, Bonnin P, Simon T, Sage AP, Guerin C, Vilar J, Caligiuri G, Tsiantoulas D, Laurans L, et al., 2013. B lymphocytes trigger monocyte mobilization and impair heart function after acute myocardial infarction. Nat Med 19:1273-1280.
- 167. Podrez EA, Byzova TV, Febbraio M, Salomon RG, Ma Y, Valiyaveettil M, Poliakov E, Sun M, Finton PJ, Curtis BR, et al., 2007. Platelet CD36 links hyperlipidemia, oxidant stress and a prothrombotic phenotype. Nat Med 13:1086-1095.
- 168. Momoi A, Fuse I, Tsukada N and Aizawa Y, 2004. *BAFF is expressed on human platelets and upregulated by activation*. Acta Medica et Biologica 52:117-126.
- 169. Finkielsztein A, Schlinker AC, Zhang L, Miller WM and Datta SK, 2015. *Human megakaryocyte progenitors derived from hematopoietic stem cells of normal individuals are MHC class II-expressing professional APC that enhance Th17 and Th1/Th17 responses*. Immunology Letters 163:84-95.
- 170. Cicha I and Urschel K, 2015. *TNF-α in the cardiovascular system: from physiology to therapy*. International Journal of Interferon, Cytokine and Mediator Research, DOI: 10.2147/ijicmr.s64894:9.
- Li K-Y, Zheng L, Wang Q and Hu Y-W, 2016. Characteristics of erythrocytederived microvesicles and its relation with atherosclerosis. Atherosclerosis 255:140-144.
- Dong ZM, Chapman SM, Brown AA, Frenette PS, Hynes RO and Wagner DD, 1998. *The combined role of P- and E-selectins in atherosclerosis*. Journal of Clinical Investigation 102:145-152.

- 173. Gimbrone MA and García-Cardeña G, 2016. *Endothelial Cell Dysfunction and the Pathobiology of Atherosclerosis*. Circulation Research 118:620-636.
- 174. Stojkovic S, Thulin Å, Hell L, Thaler B, Rauscher S, Baumgartner J, Gröger M, Ay C, Demyanets S, Neumayer C, et al., 2017. IL-33 stimulates the release of procoagulant microvesicles from human monocytes and differentially increases tissue factor in human monocyte subsets. Thrombosis and Haemostasis 117:1379-1390.
- 175. Unutmaz D, Sarkar A, Mitra S, Mehta S, Raices R and Wewers MD, 2009. Monocyte Derived Microvesicles Deliver a Cell Death Message via Encapsulated Caspase-1. PLoS ONE 4:e7140.
- 176. Egaña-Gorroño L, López-Díez R, Yepuri G, Ramirez LS, Reverdatto S, Gugger PF, Shekhtman A, Ramasamy R and Schmidt AM, 2020. *Receptor for Advanced Glycation End Products (RAGE) and Mechanisms and Therapeutic Opportunities in Diabetes and Cardiovascular Disease: Insights From Human Subjects and Animal Models*. Frontiers in Cardiovascular Medicine 7.
- 177. Rohatgi A, Ayers CR, Khera A, McGuire DK, Das SR, Matulevicius S, Timaran CH, Rosero EB and de Lemos JA, 2009. *The association between peptidoglycan recognition protein-1 and coronary and peripheral atherosclerosis: Observations from the Dallas Heart Study.* Atherosclerosis 203:569-575.
- 178. Moar P and Tandon R, 2021. *Galectin-9 as a biomarker of disease severity*. Cellular Immunology 361:104287.
- 179. Federici M, Yoo HJ, Hwang SY, Hong HC, Choi HY, Yang SJ, Choi DS, Baik SH, Blüher M, Youn B-S, et al., 2013. Implication of Progranulin and C1q/TNF-Related Protein-3 (CTRP3) on Inflammation and Atherosclerosis in Subjects with or without Metabolic Syndrome. PLoS ONE 8:e55744.
- 180. Vincent FB, Saulep-Easton D, Figgett WA, Fairfax KA and Mackay F, 2013. *The BAFF/APRIL system: emerging functions beyond B cell biology and autoimmunity*. Cytokine Growth Factor Rev 24:203-215.
- 181. Ipek E, Yolcu M, Yildirim E, Altinkaynak K, Ozbek Sebin S, Kalkan K, Gulcu O, Ermis E and Ozturk M, 2018. A Novel Marker of Inflammation: Azurocidin in Patients with ST Segment Elevation Myocardial Infarction. International Journal of Molecular Sciences 19:3797.
- 182. Jordan SC, Choi J, Kim I, Wu G, Toyoda M, Shin B and Vo A, 2017. Interleukin-6, A Cytokine Critical to Mediation of Inflammation, Autoimmunity and Allograft Rejection. Transplantation 101:32-44.
- 183. Yu G, Boone T, Delaney J, Hawkins N, Kelley M, Ramakrishnan M, McCabe S, Qiu W, Kornuc M, Xia XZ, et al., 2000. APRIL and TALL-1 and receptors BCMA and TACI: system for regulating humoral immunity. Nat Immunol 1:252-256.
- 184. Tvaroška I, Selvaraj C and Koča J, 2020. *Selectins—The Two Dr. Jekyll and Mr. Hyde Faces of Adhesion Molecules—A Review*. Molecules 25:2835.
- 185. Sinningen K, Rauner M, Goettsch C, Al-Fakhri N, Schoppet M and Hofbauer LC, 2013. Monocytic expression of osteoclast-associated receptor (OSCAR) is induced in atherosclerotic mice and regulated by oxidized low-density lipoprotein in vitro. Biochemical and Biophysical Research Communications 437:314-318.

- 186. Hofbauer LC, Schoppet M, Bornstein SR, Aikawa E, Kopprasch S, Hamann C, Nemeth K, Al-Fakhri N, Helas S, Sinningen K, et al., 2011. The Osteoclast-Associated Receptor (OSCAR) Is a Novel Receptor Regulated by Oxidized Low-Density Lipoprotein in Human Endothelial Cells. Endocrinology 152:4915-4926.
- 187. Jeon J-W, Jung J-G, Shin E-C, Choi HI, Kim HY, Cho M-L, Kim S-W, Jang Y-S, Sohn M-H, Moon J-H, et al., 2010. Soluble CD93 Induces Differentiation of Monocytes and Enhances TLR Responses. The Journal of Immunology 185:4921-4927.
- 188. Alberts A, Klingberg A, Hoffmeister L, Wessig AK, Brand K, Pich A and Neumann K, 2020. *Binding of Macrophage Receptor MARCO, LDL, and LDLR to Disease-Associated Crystalline Structures.* Frontiers in Immunology 11.
- 189. Kattoor AJ, Goel A and Mehta JL, 2019. LOX-1: Regulation, Signaling and Its Role in Atherosclerosis. Antioxidants 8:218.
- 190. Kozaki K, Akishita M, Eto M, Yoshizumi M, Toba K, Inoue S, Ishikawa M, Hashimoto M, Kodama T, Yamada N, *et al.*, 1997. *Role of Activin-A and Follistatin in Foam Cell Formation of THP-1 Macrophages*. Arteriosclerosis, Thrombosis, and Vascular Biology 17:2389-2394.
- 191. Pohjolainen V, Mustonen E, Taskinen P, Näpänkangas J, Leskinen H, Ohukainen P, Peltonen T, Aro J, Juvonen T, Satta J, *et al.*, 2012. *Increased thrombospondin-2 in human fibrosclerotic and stenotic aortic valves*. Atherosclerosis 220:66-71.
- 192. Liao C-H, Yeh S-C, Huang Y-H, Chen R-N, Tsai M-M, Chen W-J, Chi H-C, Tai P-J, Liao C-J, Wu S-M, et al., 2010. Positive regulation of spondin 2 by thyroid hormone is associated with cell migration and invasion. Endocrine-Related Cancer 17:99-111.
- 193. Gandhi PU, Gaggin HK, Redfield MM, Chen HH, Stevens SR, Anstrom KJ, Semigran MJ, Liu P and Januzzi JL, Jr., 2016. Insulin-Like Growth Factor-Binding Protein-7 as a Biomarker of Diastolic Dysfunction and Functional Capacity in Heart Failure With Preserved Ejection Fraction: Results From the RELAX Trial. JACC Heart Fail 4:860-869.
- 194. Johnson KA, Polewski M and Terkeltaub RA, 2008. *Transglutaminase 2 Is Central* to Induction of the Arterial Calcification Program by Smooth Muscle Cells. Circulation Research 102:529-537.
- 195. Talusan P, Bedri S, Yang S, Kattapuram T, Silva N, Roughley PJ and Stone JR, 2005. Analysis of intimal proteoglycans in atherosclerosis-prone and atherosclerosis-resistant human arteries by mass spectrometry. Mol Cell Proteomics. 4:1350-1357.
- 196. Ding Y, Chen J, Cui G, Wei Y, Lu C, Wang L and Diao H, 2016. Pathophysiological role of osteopontin and angiotensin II in atherosclerosis. Biochemical and Biophysical Research Communications 471:5-9.
- 197. Rathcke CN and Vestergaard H, 2009. YKL-40 an emerging biomarker in cardiovascular disease and diabetes. Cardiovascular Diabetology 8:61.
- 198. Chen Q, Jin M, Yang F, Zhu J, Xiao Q and Zhang L, 2013. *Matrix Metalloproteinases: Inflammatory Regulators of Cell Behaviors in Vascular Formation and Remodeling*. Mediators of Inflammation 2013:1-14.

- 199. Dwivedi A, Slater SC and George SJ, 2009. *MMP-9 and -12 cause N-cadherin* shedding and thereby β -catenin signalling and vascular smooth muscle cell proliferation. Cardiovascular Research 81:178-186.
- 200. Chen L-M, Skinner ML, Kauffman SW, Chao J, Chao L, Thaler CD and Chai KX, 2001. Prostasin Is a Glycosylphosphatidylinositol-anchored Active Serine Protease. Journal of Biological Chemistry 276:21434-21442.
- 201. Moore GJ and Benoiton NL, 1975. *Effect of modifiers on the hydrolysis of basic and neutral peptides by carboxypeptidase B1*. Can J Biochem 53:747-757.
- 202. Qiu Y, Sutton L and Riggs AF, 1998. *Identification of Myoglobin in Human Smooth Muscle*. Journal of Biological Chemistry 273:23426-23432.
- 203. Grönberg C, Nilsson J and Wigren M, 2017. *Recent advances on CD4 + T cells in atherosclerosis and its implications for therapy*. European Journal of Pharmacology 816:58-66.
- 204. Alipour A, van Oostrom AJHHM, Izraeljan A, Verseyden C, Collins JM, Frayn KN, Plokker TWM, Elte JWF and Castro Cabezas M, 2008. *Leukocyte Activation by Triglyceride-Rich Lipoproteins*. Arteriosclerosis, Thrombosis, and Vascular Biology 28:792-797.
- 205. Wilson KC, Center DM and Cruikshank WW, 2009. *Mini ReviewThe Effect of Interleukin-16 and its Precursor on T Lymphocyte Activation and Growth*. Growth Factors 22:97-104.
- 206. Perrier S, Darakhshan F and Hajduch E, 2006. *IL-1 receptor antagonist in metabolic diseases: Dr Jekyll or Mr Hyde?* FEBS Letters 580:6289-6294.
- 207. Nelms K, Keegan AD, Zamorano J, Ryan JJ and Paul WE, 1999. *The IL-4 receptor:* signaling mechanisms and biologic functions. Annu Rev Immunol 17:701-738.
- 208. Blumberg H, Dinh H, Dean C, Trueblood ES, Bailey K, Shows D, Bhagavathula N, Aslam MN, Varani J, Towne JE, *et al.*, 2010. *IL-1RL2 and Its Ligands Contribute to the Cytokine Network in Psoriasis.* The Journal of Immunology 185:4354-4362.
- 209. Singh AK, Stock P and Akbari O, 2011. *Role of PD-L1 and PD-L2 in allergic diseases and asthma*. Allergy 66:155-162.
- 210. Turula H and Wobus C, 2018. *The Role of the Polymeric Immunoglobulin Receptor and Secretory Immunoglobulins during Mucosal Infection and Immunity*. Viruses 10:237.
- 211. Lei Y and Takahama Y, 2012. *XCL1 and XCR1 in the immune system*. Microbes and Infection 14:262-267.
- 212. Maurer M and von Stebut E, 2004. *Macrophage inflammatory protein-1*. The International Journal of Biochemistry & Cell Biology 36:1882-1886.
- 213. Geng J, Sun X, Wang P, Zhang S, Wang X, Wu H, Hong L, Xie C, Li X, Zhao H, et al., 2015. Kinases Mst1 and Mst2 positively regulate phagocytic induction of reactive oxygen species and bactericidal activity. Nature Immunology 16:1142-1152.
- 214. Abella V, Scotece M, Conde J, Pino J, Gonzalez-Gay MA, Gómez-Reino JJ, Mera A, Lago F, Gómez R and Gualillo O, 2017. *Leptin in the interplay of inflammation, metabolism and immune system disorders*. Nature Reviews Rheumatology 13:100-109.

- 215. Hou N and Luo J-D, 2011. *Leptin and cardiovascular diseases*. Clinical and Experimental Pharmacology and Physiology 38:905-913.
- 216. Hein RM and Woods ML, 2007. *TIM-1 regulates macrophage cytokine production and B7 family member expression*. Immunology Letters 108:103-108.
- 217. Lemke G, 2019. *How macrophages deal with death*. Nature Reviews Immunology 19:539-549.
- 218. Mitrofan C-G, Appleby SL, Nash GB, Mallat Z, Chilvers ER, Upton PD and Morrell NW, 2017. Bone morphogenetic protein 9 (BMP9) and BMP10 enhance tumor necrosis factor-α-induced monocyte recruitment to the vascular endothelium mainly via activin receptor-like kinase 2. Journal of Biological Chemistry 292:13714-13726.
- 219. Zhu D, Mackenzie NCW, Shanahan CM, Shroff RC, Farquharson C and MacRae VE, 2015. BMP-9 regulates the osteoblastic differentiation and calcification of vascular smooth muscle cells through an ALK1 mediated pathway. Journal of Cellular and Molecular Medicine 19:165-174.
- 220. Grover SP and Mackman N, 2018. *Tissue Factor*. Arteriosclerosis, Thrombosis, and Vascular Biology 38:709-725.
- 221. Loghmani H and Conway EM, 2018. *Exploring traditional and nontraditional roles* for thrombomodulin. Blood 132:148-158.
- 222. Chiu KC, Chuang LM and Yoon C, 2001. *The A54T polymorphism at the intestinal fatty acid binding protein 2 is associated with insulin resistance in glucose tolerant Caucasians*. BMC Genet 2.
- 223. Jones JM, Morrell JC and Gould SJ, 2000. *Identification and Characterization of HAOX1, HAOX2, and HAOX3, Three Human Peroxisomal 2-Hydroxy Acid Oxidases.* Journal of Biological Chemistry 275:12590-12597.
- 224. Ou X, Gao J-H, He L-H, Yu X-H, Wang G, Zou J, Zhao Z-W, Zhang D-W, Zhou Z-j and Tang C-K, 2020. *Angiopoietin-1 aggravates atherosclerosis by inhibiting cholesterol efflux and promoting inflammatory response*. Biochimica et Biophysica Acta (BBA) Molecular and Cell Biology of Lipids 1865:158535.
- 225. Vu TT, Marquez J, Le LT, Nguyen ATT, Kim HK and Han J, 2018. *The role of decorin in cardiovascular diseases: more than just a decoration*. Free Radical Research 52:1210-1219.
- 226. Gonçalves I, Singh P, Tengryd C, Cavalera M, Yao Mattisson I, Nitulescu M, Flor Persson A, Volkov P, Engström G, Orho-Melander M, et al., 2019. sTRAIL-R2 (Soluble TNF [Tumor Necrosis Factor]-Related Apoptosis-Inducing Ligand Receptor 2) a Marker of Plaque Cell Apoptosis and Cardiovascular Events. Stroke 50:1989-1996.
- 227. Ura S, Masuyama N, Graves JD and Gotoh Y, 2001. *Caspase cleavage of MST1* promotes nuclear translocation and chromatin condensation. Proceedings of the National Academy of Sciences 98:10148-10153.
- 228. Rosendahl J, Witt H, Szmola R, Bhatia E, Ózsvári B, Landt O, Schulz H-U, Gress TM, Pfützer R, Löhr M, et al., 2007. Chymotrypsin C (CTRC) variants that diminish activity or secretion are associated with chronic pancreatitis. Nature Genetics 40:78-82.

- 229. Ishidoh K and Kominami E, 1995. *Procathepsin L degrades extracellular matrix proteins in the presence of glycosaminoglycans in vitro*. Biochem Biophys Res Commun 217:624-631.
- 230. Stacker S and Achen M, 2018. *Emerging Roles for VEGF-D in Human Disease*. Biomolecules 8:1.
- Åkerström B, Maghzal GJ, Winterbourn CC and Kettle AJ, 2007. *The Lipocalin α1-Microglobulin Has Radical Scavenging Activity*. Journal of Biological Chemistry 282:31493-31503.
- 232. Simsek MA, Korkmaz B, Ezici A, Turer Cabbar A, Aslanger E, Ozkalayci F, Karabay CY and Degertekin M, 2021. *The Association between Serum Heme* Oxygenase-1 Levels and Coronary SYNTAX Score. Cardiology, DOI: 10.1159/000513144:1-7.
- 233. Kang JS, Mulieri PJ, Hu Y, Taliana L and Krauss RS, 2002. BOC, an Ig superfamily member, associates with CDO to positively regulate myogenic differentiation. EMBO J 21:114-124.
- 234. Henning RJ, Bourgeois M and Harbison RD, 2018. *Poly(ADP-ribose) Polymerase* (*PARP*) and *PARP Inhibitors: Mechanisms of Action and Role in Cardiovascular Disorders*. Cardiovascular Toxicology 18:493-506.
- 235. Björkbacka H, Yao Mattisson I, Wigren M, Melander O, Fredrikson GN, Bengtsson E, Gonçalves I, Almgren P, Lagerstedt JO, Orho-Melander M, et al., 2017. Plasma stem cell factor levels are associated with risk of cardiovascular disease and death. Journal of Internal Medicine 282:508-521.
- 236. Gropp E, Shanabrough M, Borok E, Xu AW, Janoschek R, Buch T, Plum L, Balthasar N, Hampel B, Waisman A, et al., 2005. Agouti-related peptide–expressing neurons are mandatory for feeding. Nature Neuroscience 8:1289-1291.
- 237. Holstein-Rathlou S and Gillum MP, 2019. *Fibroblast growth factor 21: an endocrine inhibitor of sugar and alcohol appetite*. The Journal of Physiology 597:3539-3548.
- 238. von Holstein-Rathlou S, BonDurant Lucas D, Peltekian L, Naber Meghan C, Yin Terry C, Claflin Kristin E, Urizar Adriana I, Madsen Andreas N, Ratner C, Holst B, et al., 2016. FGF21 Mediates Endocrine Control of Simple Sugar Intake and Sweet Taste Preference by the Liver. Cell Metabolism 23:335-343.
- 239. Narula S, Yusuf S, Chong M, Ramasundarahettige C, Rangarajan S, Bangdiwala SI, van Eikels M, Leineweber K, Wu A, Pigeyre M, et al., 2020. Plasma ACE2 and risk of death or cardiometabolic diseases: a case-cohort analysis. The Lancet 396:968-976.
- 240. Silva GM, França-Falcão MS, Calzerra NTM, Luz MS, Gadelha DDA, Balarini CM and Queiroz TM, 2020. *Role of Renin-Angiotensin System Components in Atherosclerosis: Focus on Ang-II, ACE2, and Ang-I–7.* Frontiers in Physiology 11.
- 241. Beygui F, Wild PS, Zeller T, Germain M, Castagné R, Lackner KJ, Münzel T, Montalescot G, Mitchell GF, Verwoert GC, *et al.*, 2014. *Adrenomedullin and Arterial Stiffness*. Circulation: Cardiovascular Genetics 7:634-641.
- 242. Nakayama M, Takahashi K, Murakami O, Murakami H, Sasano H, Shirato K and Shibahara S, 1999. *Adrenomedullin in monocytes and macrophages: possible*

involvement of macrophage-derived adrenomedullin in atherogenesis. . Clin Sci (Lond) 97:247-251.

- 243. Hojo Y, Ikeda U, Ohya K, Ichida M, Kario K, Takahashi M, Ikeda M, Minota S, Isumi Y, Minamino N, et al., 2001. Interaction between monocytes and vascular endothelial cells induces adrenomedullin production. Atherosclerosis 155:381-387.
- 244. Gonçalves I, Edsfeldt A, Colhoun HM, Shore AC, Palombo C, Natali A, Fredrikson GN, Björkbacka H, Wigren M, Bengtsson E, *et al.*, 2016. *Association between renin and atherosclerotic burden in subjects with and without type 2 diabetes*. BMC Cardiovascular Disorders 16.
- 245. Chistiakov DA, Orekhov AN and Bobryshev YV, 2015. *Extracellular vesicles and atherosclerotic disease*. Cell Mol Life Sci 72:2697-2708.

Microvesicle signaling in cardiovascular biology under normal and pathobiological conditions



This PhD thesis is a compilation of my work at the Molecular Cardiology group, Lund University, Sweden. The main goal of the publications included in this thesis was to measure and characterize microvesicles present in blood of patients with cardiovascular disease, as well as optimize a novel, non-contact method for vesicle isolation from plasma, called acoustic trapping.



FACULTY OF MEDICINE

Department of Cardiology

Lund University, Faculty of Medicine Doctoral Dissertation Series 2021:75 ISBN 978-91-8021-081-2 ISSN 1652-8220

