# UNIVERSIDADE FEDERAL DO RIO GRANDE DO SUL

Faculdade de Farmácia

Disciplina de Trabalho de Conclusão de Curso de Farmácia

Synergistic effect of ibuprofen with itraconazole and fluconazole against

Cryptococcus neoformans

Letícia Fernandes da Rocha

Porto Alegre, dezembro de 2017

# UNIVERSIDADE FEDERAL DO RIO GRANDE DO SUL

Faculdade de Farmácia

Disciplina de Trabalho de Conclusão de Curso de Farmácia

Synergistic effect of ibuprofen with itraconazole and fluconazole against Cryptococcus neoformans

> Trabalho de Conclusão apresentado ao Curso de Farmácia da Universidade Federal do Rio Grande do Sul como parte dos requisitos para a obtenção do título de Farmacêutica

Letícia Fernandes da Rocha Prof<sup>a</sup> Adelina Mezzari Orientadora Bruna Pippi Coorientadora

Porto Alegre, dezembro de 2017

### Agradecimentos

À Faculdade de Farmácia por ter contribuído no despertar do meu fascínio pela Ciência, paixão pela área da Saúde e pela prática farmacêutica.

Às professoras e professores que tive ao longo da graduação que lecionaram com sabedoria e empatia, meus sinceros sentimentos de gratidão por todos os ensinamentos.

À Professora orientadora Adelina que embarcou na minha ideia e deu o suporte necessário para a realização desta última etapa.

À co-orientadora e amiga Bruna, que com toda a paciência, compreensão e inteligência me fez chegar nos resultados finais, minha gratidão e admiração.

Às mestrandas, doutorandas e professor do Laboratório de Micologia Aplicada, meu muito obrigada por toda a ajuda.

Aos meus amigos da vida e daqueles que me foram presenteados pela graduação e que hoje são essenciais no meu dia-a-dia, agradeço imensamente por trazerem leveza nesta dura caminhada.

À minha família, em especial minha mãe e meu irmão que conviveram diariamente com as minhas angústias dessa longa trajetória, meu muito obrigada pelo amor e suporte. Ao meu namorado que, sempre compreensível, me apoiou, confortou e ajudou desde o início da graduação, minha profunda admiração e gratidão. Este artigo foi elaborado segundo as normas da revista *Letters in Applied Microbiology* apresentadas em anexo, na qualidade de "Artigo Original". Adequações serão elaboradas após as correções e sugestões da banca revisora.

## Synergistic effect of ibuprofen with itraconazole and fluconazole against

## Cryptococcus neoformans

# Letícia Fernandes da Rocha<sup>1</sup>, Bruna Pippi<sup>2</sup>, Adelina Mezzari<sup>3</sup>

<sup>1</sup>Faculade de Farmácia, Universidade Federal do Rio Grande do Sul, Porto Alegre, Brazil.
<sup>2</sup>Programa de Pós-Graduação em Microbiologia Agrícola e do Ambiente, Universidade
Federal do Rio Grande do Sul, Porto Alegre, Brazil.

<sup>3</sup>Departamento de Análises, Faculdade de Farmácia da Universidade Federal do Rio Grande do Sul, Porto Alegre, Brazil.

Running headline: Synergism of ibuprofen and azoles

## Address correspondence:

Adelina Mezzari

Departamento de Análises, Faculdade de Farmácia, Universidade Federal do Rio Grande do Sul. Avenida Ipiranga, 2752, Porto Alegre, RS, Brasil. CEP 90.610-000

E-mail address: mezzari@ufrgs.br

## Significance and Impact of the Study

The high incidence of resistance development in *Cryptococcus neoformans* isolates drives the search for new strategies of treatment such as the association of drugs acting synergistically. Non-antifungal drugs such as ibuprofen have been the subject of research because of the low cost and good knowledge of its adverse and therapeutic effects. Ibuprofen has been explored for its potential ability to increase susceptibility of strains to antifungal and to help combat the inflammatory symptoms that fungal diseases may cause.

### Abstract

The present study investigated the association of the non-steroidal anti-inflammatory drug ibuprofen with itraconazole, fluconazole and amphotericin B against *Cryptococcus neformans* isolates. The minimal inhibitory concentration (MIC) was found according to M27-A3 protocol and the *in vitro* interactions were evaluated using a checkerboard microdilution method. Synergism was demonstrated between azoles and ibuprofen for most isolates. However, no synergistic effects were seen when amphotericin B was combined with ibuprofen. Therefore, our results suggest that ibuprofen presents clinical potential when combined with azole drugs in the treatment of cryptococcosis.

Keywords: Cryptococcus neoformans, ibuprofen, itraconazole, fluconazole, synergism

#### Introduction

Opportunistic pathogenic fungi such as *Candida, Aspergillus* and *Cryptococcus* species are responsible for systemic infections affecting mainly immunodeficient patients such as neonates, transplanted and patients with acquired immunodeficiency syndrome (AIDS) (Grimaldi *et al.* 2010). Cryptococcosis is an infection mainly caused by encapsulated yeast fungus such as *Cryptococcus neoformans*. This microorganism is found in bird droppings and contaminated soil with higher prevalence in tropical and subtropical regions (Ramos-e-Silva *et al.* 2012). It has the airway as portal of entry causing pulmonary infection and can be disseminated to the brain resulting in severe meningoencephalitis (Prates *et al.* 2013; Chen *et al.* 2015). It is estimated that cryptococcal meningitis result in 120.000 to 240.000 deaths per year worldwide (Rajasingham,*et al.* 2017).

The gold standard treatment for cryptococcosis is the combination of amphotericin B and 5-flucytosine (Reichert-Lima *et al.* 2016). However, due to its high cost, 5-flucytosine is not present in therapeutic protocols in several countries giving place to azole drugs (Smith *et al.* 2015). Nevertheless, azoles have shown ineffectiveness in several studies due to the development of strains resistant (Gullo *et al.* 2013). In addition, although the polyene resistance is considered to be rare and has a good spectrum of activity, this drug has restricted use due to nephrotoxicity problems. (Kagan *et al.* 2012; Xie *et al.* 2014).

Currently few antifungals are commercially available and the development of new drugs does not accompany the high incidence of the development of resistant strains (Liu *et al.* 2013). Combination therapy with two or more antifungals has the potential to reduce antifungal resistance and decrease toxicity of each drug, but its side effects should be

evaluated with caution (Hatipoglu and Hatipoglu 2013). Thus, *in vitro* association studies with non-antifungal agents in order to at the potentiation of antifungal drugs have been performed and are still required to delineate *in vivo* assays and consequent clinical trials (Venturini *et al.* 2011; Hatipoglu and Hatipoglu 2013). Ibuprofen is a non-steroidal anti-inflammatory drug easily accessible because it is inexpensive and has shown a synergistic effect when combined with fluconazole in *Candida* strains (Hatipoglu and Hatipoglu 2013; Liu *et al.* 2013). So, the present study aims to test the association of ibuprofen with itraconazole, fluconazole and amphotericin B against *C. neoformans* isolates.

### **Results and Discussion**

The MIC values of each antifungal agents against twenty-five *C. neoformans* isolates were determined. MIC range, Geometric means (GM), MIC<sub>50</sub> (MIC value which inhibits 50% of the isolates) and MIC<sub>90</sub> (MIC value that inhibits 90% of the isolates) for itraconazole (ITC), fluconazole (FLC), amphotericin B (AMB) and ibuprofen (IBP) are presented in Table 1. It can be observed that isolates with low sensitivity to the antifungal agents were found. The use of IBP alone resulted in 50% growth reduction. Since IBP is not an antifungal and there is no standardization in relation to the evaluation of its inhibitory effect, we consider as MIC the concentration that reduces 50% fungal growth. Based on the high MICs, the non-antifungal agent showed a weak antifungal activity against *C. neoformans*.

Table 2 prsent the effects of antifungal agent combination, which demonstrated synergistic or indifference. The combination of azoles (ITC and FLC) with IBP resulted predominantly in synergism, which was detected in 75% of the isolates for combination with FLC and in 62% of the isolates for combination with ITC. On the other hand, AMB

associated with IBP resulted in 100 % of indifference against *C. neoformans*. Antagonism was not detected against both groups.

MIC for AMB combined with IBP was chosen when fungal growth was reduced in 100%.

The limited efficacy and the difficulty to introduce new antifungal drugs into the market make the drugs association an important therapeutic strategy to treat potentially life-threatening invasive fungal infections (Fuentefria et al. 2017). Previous studies have detected synergism between IBP and azole agains *Candida albicans* and *Cryptococcus neoformans* increasing the susceptibility of the isolates to these antifungal agents (Ricardo *et al* 2009; Ogundeji *et al* 2016), corroborating with our research. Several mechanisms may be involved in the selection of azole resistant strains, such as mutations causing structural changes in enzyme affinity, overproduction of enzymes and overexpression of genes that encode proteins that cause drug efflux (Gullo *et al.* 2013). The efflux bombs are plasma membrane transport proteins. AFR1, MDR1 and AFR2 genes plays an important role in encoding of these proteins in *C. neoformans* and *C. gatti.* When these genes are overexpressed occurs to expulsion of the azoles out of the cell contributing to the decrease of the concentration of drug at action site and explains part of the resistance to azole drugs (Basso Jr *et al.* 2015).

Understanding the resistance mechanisms of azoles and the action of IBP helps to explain our findings of *in vitro* synergy. IBP is an efflux pump blocker and can prevent the output of azole from the fungal cell. Thus, the high susceptibility of cells to IBP+azoles association may be attributed to the increase in intracellular concentration of the antifungal (Pina-Vaz *et al* 2005). On the other hand, AMB do not require internalization into fungal cells for exert their antifungal activity and so they escape from efflux systems (Vandeputte *et al.* 2012). This may justify the indifferent effect of the IBP+AMB association found in the present study. In addition, previous studies showed that IBP causes fungal membrane damage and can be considered, depending on the dose, fungicide or fungistatic (Argenta *et al.* 2012; Arai *et al.* 2005). Our results are in agreement since IBP alone was able to inhibit the cell growth of *C. neoformans* isolates.

An additional benefit in using a nonsteroidal anti-inflammatory drugs such as ibuprofen should be considered since prostaglandins may be involved in fungal colonizations and its anti-inflammatory mechanism works mostly by inhibiting cyclooxygenase isoenzymes (Rusu *et al.* 2014). Thus, in addition to synergism, IBP has advantages because of the clinical manifestations of the disease, which, besides classic pulmonary and central nervous system manifestations, also causes infections of the skin, prostate, eyes and other parts of the body (Maziarz and Perfect 2016).

The results of this present study suggest that the combination between IBP and azole drugs may be suitable for cryptococcosis therapy since synergism was demonstrated. Further *in vivo* studies in clinical situations are still required to prove the effects of the combination of ibuprofen and azoles antifungals.

## Material and methods

#### **Fungal Strains**

A total of twenty five clinical isolates of *C. neoformans* were included in this study. All isolates had PCR-confirmed molecular identification through primers CNa-70S (5'-ATTGCGTCCACCAAGGAGCTC-3') and CNa-70A (5'-ATTGCGTCCATGTTACGTGGC-3'). The isolates were provided by the Clinical Analysis Departmentof the Federal University of Rio Grande do Sul, Porto Alegre, RS. All isolates were grown on Sabouraud dextrose agar at 35 ° C for 48 h prior to the experiments.

### Drugs

According to CLSI recommendations, FLC stock solution (Metrochem Api Private Limited, India) was prepared in distilled water. IBP (Sigma-Aldrich, USA), ITC (Metrochem Api Private Limited), and AMB (Metrochem Api Private Limited) stock solution were prepared in DMSO (Nuclear, Brazil). For the experiments, the compounds were diluted in RPMI 1640 medium (Sigma-Aldrich) to obtain a maximum concentration of 2% DMSO.

### Antifungal susceptibility testing:

Minimum inhibitory concentrations (MICs) of IBP and antifungal agents were determined in duplicate by broth microdilution method according to M27-A3 protocol (CLSI, 2008). Serial two-fold dilutions were made in RPMI 1640 medium (Sigma-Aldrich) buffered with MOPS (Sigma-Aldrich) and the concentrations ranges tested were:  $16 - 0.0312 \mu g/ml$  of ITC,  $0.125 - 64 \mu g/ml$  of FLC,  $0.0312 - 16 \mu g/ml$  of AMB and  $1 - 512 \mu g/ml$  of IBP. The experiments were carried out in duplicate. MICs values were defined as the lowest concentration of compounds at which the microorganisms tested did not show visible growth (AMB) or reduced 50% of growth (FLC, IBP and ITC) in 72 h.

## **Checkerboard assay**

The interaction between IBP and each antifungal was evaluated for eight *C. neoformans* isolates using the checkerboard method (Johnson et al. 2004). The assay lead to forty nine different concentration combinations between IBP and antifungal agents in concentrations of the MIC/8, MIC/4, MIC/2, MIC, MICx2, MICx4 and MICx8. The experiments were conducted in duplicate and incubated at 35°C for 72 h. The effect of

the combinations was classified by determining the fractional inhibitory concentration index (FICI) expressed as the sum of the fractional inhibitory concentrations (FIC), as defined by the following equation:

 $FICI = FIC_A + FIC_B = \frac{MIC_A \text{ in combination}}{MIC_A \text{ tested alone}} + \frac{MIC_B \text{ in combination}}{MIC_B \text{tested alone}}$ 

where MIC<sub>A</sub> and MIC<sub>B</sub> are the MICs of ibuprofen and antifungal agent, respectively (Mukherjee *et al*, 2005). Sinergism was defined when FICI  $\leq 0.5$ , indifference when 0.5 <FICI  $\leq 4$  and antagonism when FICI > 4 (Odds 2003).

# Acknowledgements

To the Department of Analysis of the Faculty of Pharmacy of the Federal University of Rio Grande do Sul, Porto Alegre, RS, Brazil, for the possibility of carrying out the present study as a Conclusion of the Pharmacy Course

## **Conflict of Interest**

The authors declare no conflict of interest.

### References

Arai, R., Sugita, T. and Nishikawa, A. (2015) Reassessment of the in vitro synergistic effect of fluconazole with the non-steroidal anti-inflammatory agent ibuprofen against Candida albicans. *Mycoses* 48, 38–41.

Argenta, J.S., Alves, S.H., Silveira, F., Maboni, G., Zanette, R.A., Cavalheiro, A.S., Pereira, P.L., Pereira, D.I.B., Sallis, E.S.V., Pötter, L., Santurio, J.M. and Ferreiro, L. (2012) In vitro and in vivo susceptibility of two-drug and three-drug combinations of terbinafine, itraconazole, caspofungin, ibuprofen and fluvastatin against Pythium insidiosum. *Veterinary Microbiology* 157, 137–142.

Basso Jr, L.R., Gast, C.E., Bruzual, I. and Wong, B. (2015) Identification and properties of plasma membrane azole efflux pumps from the pathogenic fungi Cryptococcus gattii and Cryptococcus neoformans. *J Antimicrob Chemother* 70, 1396–1407.

Chen, S., Yan, H., Zhang, L., Kong, W., Sun, Y., Zhang, W., Chen, Y. and Deng, A. (2015) Cryptococcus Neoformans Infection and Immune Cell Regulation in Human Monocytes. *Cell Physiol Biochem* 37, 537-547.

Clinical And Laboratory Standards Institute (CLSI). Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts; Approved Standard – Third Edition. CLSI Document M27-A3. Clinical Laboratory Standards Institute, Wayne, PA, USA, 2008.

Fuentefria, A.M., Pippi, B., Dalla Lana, D.F., Doanto K.K. and De Andrade, S.F. (2017) Antifungals discovery: an insight into new strategies to combat antifungal resistance. *Letters in Applied Microbiology*, doi:10.1111/lam.12820 Grimaldi, M., De Rosa, M., Di Marino, S., Scrima, M., Posteraro, B., Sanguinetti, M., Fadda, G., Soriente, A. and D'Ursi, A.M. (2010) Synthesis of new antifungal peptides selective against Cryptococcus neoformans. *Bioorganic & Medicinal Chemistry* 18, 7985–7990.

Gullo, F.P., Rossi, S. A., Sardi, J., Teodoro, V. L. I., Mendes-Giannini, M. J. S. & Fusco-Almeida A. M. (2013) Cryptococcosis: epidemiology, fungal resistance, and new alternatives for treatment. *Eur J Clin Microbiol Infect Dis* 32, 1377–1391

Hatipoglu, N. and Hatipoglu, H. (2013) Combination antifungal therapy for invasive fungal infections in children and adults. *Expert Rev. Anti Infect. Ther.* 11, 523-535.

Johnson, M., Macdougall, C., Ostrosky-Zeichner, L., Perfect, J. and Rex, J. (2004) Combination antifungal therapy. *Antimicrob Agents Chemother* 48, 693–715.

Kagan, S,. Ickowicz, D., Shmuel, M., Altschuler, Y., Sionov, E., Pitusi, M., Weiss, A., Farber, S., Domb, A. & Polachecka I. (2012) Toxicity mechanisms of amphotericin B and its neutralization by conjugation with arabinogalactan. *Antimicrobial Agents and Chemotherapy* 56, 5303-5611.

Liua, S., Houb, Y., Chenc, X., Gaoa, Y., Li, H., & Sund, S. (2014) Combination of fluconazole with non-antifungal agents: A promising approach to cope with resistant Candida albicans infections and insight into new antifungal agent discovery. *International Journal of Antimicrobial Agents* 43, 395-402.

Maziarz, E.L., and Perfect, J.R. (2016) Cryptococcosis. *Infect Dis Clin N Am* 30, 179-206.

Mukherjee P.K., Sheehan D.J., Hitchcock CA, Ghannoum MA. (2005) Combination Treatment of Invasive Fungal Infections. *Clin Microbiol Rev* 18, 163–194.

14

Odds, F.C. (2003) Synergy, antagonism, and what the chequerboard puts between them. *J Antimicrob Chemother* 52, 1.

Ogundeji, A.O, Pohl, C.O. & Sebolai, O.M. (2016) Repurposing of Aspirin and Ibuprofen as Candidate Anti-Cryptococcus Drugs. *Antimicrobial Agents and Chemotherapy* 60, 4799-4808.

Rajasingham, R., Smith, R.M., Park, B.J., Jarvis, J.N., Govender, N.P., Chiller, T.M., Denning, D.W., Loyse, A., Boulware, D.R. (2017) Global burden of disease of HIV-associated cryptococcal meningitis: an updated analysis. *Lancet Infect Dis* 17, 873-881.

Renato, A., Prates, R.A., Fuchs, B.B., Mizuno, K., Naqvi, Q., Kato, I.K., Ribeiro, M.R., Mylonakis, E., Tegos, G.P. & Hamblin, M.R. (2013) Effect of Virulence Factors on the Photodynamic Inactivation of Cryptococcus neoformans. *PLoS ONE* 8, e54387-.

Ramos-e-Silva, M., Lima, C.M.O., Schechtman, R.C., Trope, B.M., & Carneiro, S. (2012) Systemic mycoses in immunodepressed patients (AIDS). *Clinics in Dermatology* 30, 616-627.

Pina-Vaz, C., Rodrigues, A.G., Costa-De-Oliveira, S., Ricardo, E. and Mardh, P.A. (2005) Potent synergic effect between ibuprofen and azoles on Candida resulting from blockade of efflux pumps as determined by FUN-1 staining and flow cytometry. *J Antimicrob Chemother*, 56, 678-685.

Reichert-Lima, F., Busso-Lopes, A.F., Lyra, L., Haddad Peron, I., Taguchi, H., Mikami, Y., Kamei, K., Moretti, M.L. and Schreiber, A.Z. (2016) Evaluation of antifungal combination against Cryptococcus spp. *Mycoses* 59, 585–593.

Ricardo, E., Costa-de-Oliveira, S., Silva Dias, A., Guerra, J., Gonc<sub>alves</sub> Rodrigues, A. And Pina-Vaz, C. (2009) Ibuprofen reverts antifungal resistance onCandida albicans showing overexpression of CDRgenes. *FEMS Yeast Res* 9, 618–625.

Rusu, E., Radu-Popescu, M., Pelinescu, D. and Vassu, T. (2014) Treatment with some anti-inflammatory drugs reduces germ tube formation in Candida albicans strains. *Brazilian Journal of Microbiology* 45, 1379-1383.

Smith, K.D., Achan, B., Huppler Hullsiek, K., McDonald, T.R., Okagaki, L.H., Alhadab, A.A., Akampurira, A., Rhein, J.R., Meya, D.B., Boulware, D.R. and Nielsen, K. (2015) Increased Antifungal Drug Resistance in Clinical Isolates of Cryptococcus neoformans in Uganda. *Antimicrobial Agents and Chemotherapy* 59, 7197–7204.

Vandeputte P., Ferrari S. and Coste AT. (2012) Antifungal Resistance and New Strategies to Control Fungal Infections. *Int J Microbiol* 3, 1-26.

Venturini, T.P., Rossato, L., Spader, T.B., Tronco-Alves, G.R., Azevedo, M.I., Weiler, C.B., Santurio, J.M. and Hartz Alves, S. (2011) In vitro synergisms obtained by amphotericin B and voriconazole associated with non-antifungal agents against Fusarium spp. *Diagnostic Microbiology and Infectious Disease* 71, 126–130.

Xie, J. L., Polvi1, E., Shekhar-Guturja, T. & Cowen, L. (2014) Elucidating drug resistance in human fungal pathogens. *Future Microbiol.* 9, 523-542.

# Tables

**Table 1.** Susceptibility profile ( $\mu$ g/ml) of twenty five isolates of *Cryptococcus neoformans* to antifungal agents expressed in ranges of variation of minimum and maximum MIC values (MIC ranges), geometric mean (GM), MIC<sub>50</sub> (MIC value that inhibits 50% of the isolates) and MIC<sub>90</sub> (MIC value that inhibits 90% of the isolates).

Agents	MIC range (µg/ml)	GM (µg/ml)	MIC <sub>50</sub> (µg/ml)	MIC <sub>90</sub> (µg/ml)
ITC	0.03125 - 1	0.66	1	1
FLC	0.25 - 8	1.74	2	4
AMB	0.5 - 16	6.17	4	16
IBP	128 - 512	319.57	256	512

Isolata		MIC (ug/mL)			IBP + FLC		IBP + ITC			IBP + AMB						
Isolate		MIC (µg/IIIL)			M	MIC combination (µg/mL)		MIC combination (µg/mL)			MIC combination (µg/mL)					
	IBP	FLC	ITC	AMB	IBP	FLC	FICI	Interaction	IBP	ITC	FICI	Interaction	IBP	AMB	FICI	Interaction
CN06	512	4	0.25	2	8	1	0.26	Syn	256	0.0625	0.5	Syn	8	2	1.01	Ind
CN07	512	8	0.5	2	128	0.25	0.25	Syn	8	0.25	0.52	Ind	16	2	1.03	Ind
CN10	512	2	0.25	2	64	0.5	0.375	Syn	64	0.03125	0.25	Syn	512	1	1.5	Ind
CN11	512	4	0.5	4	8	2	0.52	Ind	16	0.125	0.28	Syn	256	2	1	Ind
CN17	512	4	0.25	0.5	8	1	0.27	Syn	8	0.125	0.52	Ind	256	0.25	1	Ind
CN19	512	4	0.5	2	64	1	0.38	Syn	8	0.125	0.27	Syn	512	1	1.5	Ind
CN24	256	4	0.25	4	4	1	0.26	Syn	64	0.125	0.75	Ind	512	2	2.5	Ind
CN25	512	2	0.25	2	256	0.5	0.75	Ind	128	0.0625	0.5	Syn	256	0.25	0.63	Ind

**Table 2.** *In vitro* susceptibility of *Cryptococcus neoformans* to ibuprofen (IBR) combined with itraconazole (ITC), fluconazole (FLC) and amphotericin B (AMB).

Syn = Synergism Ind= indifferent