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6  
7 RESEARCH ARTICLE

8  
9 **Effects of 2,4-dichlorophenoxyacetic acid on the ventral prostate of rats**  
10 **during the peri-pubertal, pubertal and adult stage**

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15  
16 **Abstract**

17 The herbicide 2,4-dichlorophenoxyacetic acid (2,4-D) is used on a wide variety of terrestrial and  
18 aquatic broadleaf weeds. 2,4-D has been shown to produce a wide range of adverse effects on  
19 animal and human health. The aim of the current study was to evaluate the effects of pre- and  
20 postnatal exposure to 2,4-D on rat ventral prostate (VP). Pregnant rats were exposed daily to  
21 oral doses of 70 mg/kg/day of 2,4-D from 16 days of gestation up to 23 days after delivery.  
22 Then, the treated groups ( $n=8$ ) were fed with a 2,4-D added diet until sacrificed by  
23 decapitation on postnatal day (PND) 45, 60, or 90. Morphometric studies were performed and  
24 androgen receptor (AR) protein levels in the VP were determined. AR, insulin-like growth factor-  
25 I (IGF-1) and insulin-like growth factor-I receptor (IGF-1R) mRNA expression in the VP along with  
26 testosterone (T), dihydroxytestosterone (DHT), growth hormone (GH) and IGF-1 serum levels  
27 were also determined to ascertain whether these parameters were differentially affected.  
28 Results of this study showed that 2,4-D exposure during gestation and until adulthood altered  
29 development of the prostate gland in male rats, delaying it at early ages while increasing its  
30 size in adults, indicate that 2,4-D could behave as endocrine disruptors (EDs).

31  
32  
33 **Introduction**

34 Differentiation of the prostate gland during embryogenesis  
35 and subsequent tissue growth during postnatal life is  
36 controlled by androgenic hormones synthesized in the testes  
37 (George et al., 1991). The two most important androgens are  
38 testosterone (T) and its metabolite, 5 $\alpha$ -dihydrotestosterone  
39 (DHT). Both act through the same receptor and each of these  
40 androgens has its own specific role during male sexual  
41 differentiation (Knobil & Neill, 1994). Some androgenic  
42 effects, such as the promotion of spermatogenesis and the  
43 enhancement of muscle growth, are believed to be mediated  
44 by the testicular androgen testosterone. In other target tissues,  
45 including prostate, testosterone is converted to DHT by the  
46 enzyme steroid 5 $\alpha$ -reductase (George et al., 1991). In addition  
47 to hormonal influences, studies have demonstrated that  
48 several growth factors, such as insulin-like growth factor-I  
49 (IGF-I), display important mitogenic effects on the prostate  
50 and are essential for the development of this gland (Ruan  
51 et al., 1999).

52 Epidemiological studies have indicated the influence of  
53 height, weight, dietary and lifestyle factors on IGF-I serum

33 **Keywords**

34 2,4-Dichlorophenoxyacetic herbicide,  
35 developmental toxicity, hormone,  
36 androgen receptor

37 **History**

38 Received 13 January 2015  
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41 Published online ■ ■ ■

42 levels and several of its binding proteins (Baibas et al., 2003;  
43 Kaklamani et al., 1999; Sandhu et al., 2006; Schneider et al.,  
44 2006). Other factors, such as the environment and dietary  
45 contamination by chemicals should also be taken into  
46 account. The possibility exists that environmental contamin-  
47 ants could influence the IGF system. Thus, studies in animals  
48 and human subjects have demonstrated that environmental  
49 pollutants, such as benzopyrene, dioxins, dibenzofurans and  
50 hexachlorobenzene could alter the normal synthesis and/or  
51 secretion of IGF-I (Randi et al., 2006; Tannheimer et al.,  
52 1998; Wang et al., 2005).

53 Chlorophenoxy herbicides are widely used in agriculture  
54 and forestry for the control of broad-leaved weeds in pastures,  
55 cereal crops, as well as along public rights of way. 2,4-  
56 Dichlorophenoxyacetic acid (2,4-D) is used on a wide variety  
57 of terrestrial and aquatic broadleaf weeds. It has little effect  
58 on grasses (Shaner, 2014). Several studies have shown that  
59 doses of 50, 70 or 100 mg/kg body weight (bw)/day of 2,4-D  
60 produce a wide range of toxic effects on the embryo as well as  
61 on the reproductive and neural systems in animal (mostly rat)  
62 and human models (Barnekow et al., 2001; Charles et al.,  
63 2001; Rosso et al., 2000). Lerda & Rizzi (1991) studied the  
64 reproductive function of 32 male farm sprayers who were  
65 exposed to 2,4-D and found significant levels of asthenos-  
66 permia, necrospermia and teratospermia in exposed workers  
67 compared with unexposed controls. Doses of 50 mg/kg  
68 bw/day of 2,4-D have been reported to increase ventral

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121 prostate (VP) weight in rats (Kim et al., 2002). Treatment  
122 of human prostate cancer cell cultures with 10 nM 2,4-D  
123 enhanced the androgenic activity of dihydroxytestosterone  
124 (DHT) on cell proliferation and transactivation (Kim et al.,  
125 2005).

126 There are few studies on developmental toxicology  
127 addressing the effects of 2,4-D on the prostate. Recently we  
128 have found that 2,4-D (70 mg/kg bw/day) induced an increase  
129 in reactive oxygen species (ROS) levels, lipid peroxidation  
130 and protein oxidation, thereby causing oxidative stress in VP.  
131 This, in turn, could provoke important deleterious changes in  
132 the development of the organ at different ages (Pochettino  
133 et al., 2013). The aim of the current study was to evaluate the  
134 effects of pre- and postnatal exposure of 2,4-D on rat VP.  
135 For this purpose, we carried out morphometric studies and  
136 measured AR protein levels in the prostate. AR, IGF-1 and  
137 IGF-1R mRNA expression in the VP along with T, DHT,  
138 growth hormone (GH) and IGF-1 serum levels were also  
139 determined to ascertain whether these parameters were  
140 differentially affected.

## 141 Materials and methods

### 142 Animals and exposure to 2,4-D

143 Nulliparous female rats of Wistar origin, between 90 and 110  
144 days old and weighing approximately 180–210 g were  
145 obtained from the animal breeding colony of the Faculty of  
146 Pharmacy and Biochemistry, Rosario, Argentina. Stages of  
147 the reproductive cycle were monitored via daily cytological  
148 examination of vaginal smears. Females were mated indi-  
149 vidually with fertile males on the night of pro-estrus. This day  
150 was denoted as gestation day 0 (GD 0). At this time, pregnant  
151 females were individually housed in plastic breeding cages in  
152 a temperature-controlled nursery (22–24 °C) and maintained  
153 on a 12-h light/dark cycle. Food (Cargill pellets, Buenos  
154 Aires, Argentina) and water were available ad lib. All  
155 experimental protocols were performed according to the  
156 Regulation for the Care and Use of Laboratory Animals  
157 (File 6109/012 E.C. Document 267/02) approved by the  
158 Institutional Committee for Animal Use of the National  
159 University of Rosario, Argentina. On GD 16, the pregnant  
160 females were randomly divided into two groups, as follows:

161 *2,4-D-treated groups.* Dams treated with a daily oral dose  
162 (by diet) of about 70 mg of 2,4-D per kg body weight (bw) per  
163 day (70 mg/kg/day) from GD 16 until postnatal day (PND) 23.  
164 Selection of the 2,4-D dose was based on previous studies,  
165 which demonstrated behavioral changes (Bortolozzi et al.,  
166 1999), alterations in neurotransmitter levels in adult rats  
167 (Evangelista de Duffard et al., 1990) and in neonate rats (Ferri  
168 et al., 2000, 2003, 2007), when pups were exposed to the  
169 herbicide through mother's milk. The selected dose was lower  
170 than the no-observed-adverse-effect level (NOAEL) for  
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173  
174 Figure 1. Treatment schedule. Pregnant rats  
175 were treated with 2,4-D (70 mg/kg/day) from  
176 gestational day (GD 16) until weaning. Then,  
177 the treated groups were fed with a 2,4-D  
178 added diet until sacrificed by decapitation on  
179 postnatal day (PND) 45, 60, or 90.

181 chronic dietary 2,4-D toxicity in rats, which was established  
182 as 77 mg/kg/day (Munro et al., 1992). An alcoholic solution  
183 of 2,4-D was mixed with the food and allowed to dry before  
184 administration in the diet (Bortolozzi et al., 1999). According  
185 to previous work, the dietary intake of animals was adjusted  
186 to the most recent body weight and food consumption  
187 determinations (Stürtz et al., 2006).

188 *Control groups.* Dams were fed the same food (sprayed  
189 with alcohol and dried), as described for the treated groups  
190 but without the herbicide.

191 After parturition, each litter was reduced to eight male  
192 pups when possible on PND 1 to ensure good nutrition.  
193 Pups were weaned at PND 23. Next, the treated groups were  
194 fed the 2,4-D diet until sacrifice at 45, 60 or 90 days of age  
195 (Figure 1). Animals were weighed, euthanized by decapitation  
196 between 10.00 and 12.00 h, and trunk blood was collected.  
197 Serum was separated by centrifugation at 4 °C for 15 min at  
198 3500 rpm and stored at –80 °C for hormone level determi-  
199 nation. The VP was dissected from the abdominal cavity of  
200 each animal. After weighing, a portion of the VP was fixed in  
201 10% buffered formalin for paraffin embedding. The remaining  
202 tissue was immediately frozen in liquid nitrogen for further  
203 analysis.

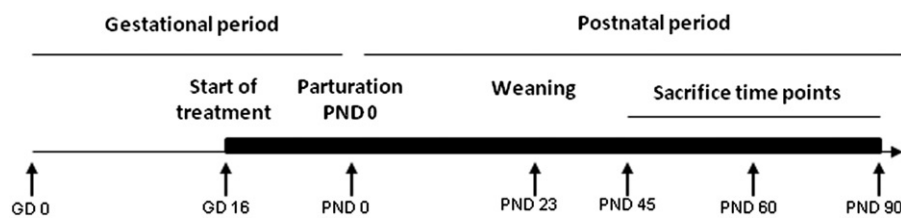
### 204 Histopathological analysis

205 Fixed tissue samples were dehydrated in a graded ethanol  
206 series and embedded in paraffin wax. Sections of 3–5 µm  
207 thickness were cut with a Reichert-Jung Hn 40 microtome and  
208 stained with hematoxylin–eosin (H&E). Slides were exam-  
209 ined under an Olympus Provis microscope (BX40, Olympus  
210 Optical Corp., Toyota, Japan) and images were captured  
211 digitally with the Olympus D-560 camera (Olympus Optical  
212 Corp.).

213 Digital VP images were examined with a digital image  
214 analysis program (ImageJ). Epithelial thickness and alveoli  
215 cell number per unit area were measured and averaged from  
216 four sections per rat (Mandarim-de-Lacerda, 1999; Ma et al.,  
217 2004).

### 218 Western blotting analysis for androgen receptor (AR)

219 Prostate samples were mechanically homogenized in buffer  
220 containing 50 mM Tris–HCl (pH 7.4), 150 mM NaCl, 1 mM  
221 EDTA, 1% Triton X-100, 1% sodium deoxycholate, 0.1%  
222 sodium dodecylsulphate (SDS) and 1 mM phenylmethylsul-  
223 fonyl fluoride (PMSF) by means of a Polytron for 30 s at 4 °C.  
224 Following centrifugation of the homogenate, proteins were  
225 extracted from the supernatant and quantified by the Lowry  
226 method (Lowry et al., 1951). Aliquots of each sample  
227 containing equal amounts of protein were loaded onto 8%  
228 SDS-polyacrylamide gels for SDS-polyacrylamide gel elec-  
229 trophoresis under reducing conditions. After electrophoresis,  
230



241 proteins were transferred onto a nitrocellulose membrane  
 242 (Sigma). The blot was blocked with 10% nonfat dry milk in  
 243 phosphate buffered saline/3% milk powder/0.1% triton X100  
 244 (PBSX) for 1 h, incubated overnight at 4 °C with 3% BSA in  
 245 PBSX containing a 1:1000 dilution of the anti-AR (Sigma) or  
 246 anti-βactin (Sigma) primary antibodies and washed twice for  
 247 20 min in PBSX. Blots were subsequently incubated with  
 248 appropriate alkaline phosphatase-labeled secondary antibody  
 249 for 1 h, and then visualized by incubating the membrane  
 250 for 15 min in a solution containing nitroblue tetrazolium and  
 251 5-bromo-4-chloro-3-indolyl-phosphate. AR and β-actin pro-  
 252 tein expression were quantified by densitometric analysis of  
 253 the bands as integrated optical density (IOD). AR expression  
 254 was normalized to βactin values.

## 255 Hormone assays

256 GH was measured by double antibody radioimmunoassay  
 257 (RIA) using materials generously provided by A. F. Parlow  
 258 and the NHPP (National Hormone and Pituitary Program,  
 259 Harbor-UCLA Medical Center, Torrance, CA). Hormones  
 260 were radio-iodinated using the Chloramine T method and  
 261 purified by passage through Sephadex G75 (Rosato et al.,  
 262 1992). Results were expressed in terms of the rat GH RP-2  
 263 standard preparations. Assay sensitivity was 0.5 g l<sup>-1</sup> serum  
 264 and the inter- and intra-assay variation coefficients  
 265 were <10%.

266 Rat IGF-I, testosterone and DHT concentrations in sera  
 267 were measured by radioimmunoassay using commercial kits  
 268 for total hormones (DSL-2900, DSL-4100, DSL-9600 double  
 269 antibody radioimmunoassay, respectively; all from Diagnostic  
 270 Products Corporation, Los Angeles, CA).

## 271 RNA isolation, cDNA synthesis and qPCR

272 Total RNA was isolated from VP using TRIzol (Invitrogen,  
 273 Carlsbad, CA) according to manufacturer's recommendations.  
 274 RNA pellets were dissolved in RNase-free water and stored  
 275 at -80 °C until analysis. Total RNA was quantified by OD  
 276 260 nm spectrophotometry (Beckman DU 640  
 277 Spectrophotometer). Integrity of purified RNA was deter-  
 278 mined by 2% agarose gel electrophoresis. cDNA was  
 279 synthesized from 2 µg of total RNA using oligo (dt) primer  
 280 (Biodynamics S.R.L, Argentina) and 200 U M-MLV reverse  
 281 transcriptase (Promega, WI). Briefly, 5 × M-MLV Reaction  
 282 Buffer, 0.4 mM dNTPs (Promega, WI); 21.5 U RNase  
 283 Inhibitor (Promega, WI), 0.4 µM oligo (dt) primers; 2 mM  
 284 MgCl<sub>2</sub> (Invitrogen) and RNase-free water for 50 µl of final  
 285 volume. Retrotranscription cycling programs consisted of  
 286 5 min at 65 °C, 1 h at 40 °C followed by enzyme inactivation  
 287 at 95 °C for 3 min. cDNA was stored at -80 °C until use.  
 288 qPCR was performed with the ABI PRISM 7500 Real Time  
 289 PCR System (Applied Biosystems, Foster City, CA) using  
 290 10 µl of a 1/200 dilution of cDNA, 0.4 µM of each primer  
 291 (Invitrogen, Argentina) (Table 1) and 25 µl of FastStart  
 292 Universal SYBR Green Master (ROX) (Roche Applied  
 293 Science) in a final volume of 50 µl. The reaction mixture  
 294 was run online at 50 °C for 2 min and 95 °C for 10 min,  
 295 followed by 35 cycles at 95 °C for 1 min, 55 °C for 1 min and  
 296 72 °C for 1 min, with an extension phase of 1 cycle at 95 °C  
 297 for 1 min, 60 °C for 1 min and 95 °C for 1 min.

Table 1. Description of primers used in this study.

Primers	Type	Primer sequence	Length (bp)
AR	Forward	5'-TAGCAGGGCAGATCCTGTCT-3'	197
AR	Reverse	5'-CCACCGAATTCCTTATCCT-3'	197
IGF-1	Forward	5'-TCCGCTGAAGCCTACAAAGT-3'	200
IGF-1	Reverse	5'-GGGAGGCTCCTCCTACATTC-3'	100
IGF-1R	Forward	5'-GACAGTGAATGAGGCTGCAA-3'	197
IGF-1R	Reverse	5'-TCTCCACCTCTGGCCTTAGA-3'	100
GADPH	Forward	5'-TGCCAAGGCTGTGGGCAAGG-3'	197
GADPH	Reverse	5'-GCTTACCACCTTCTTGATG-3'	197

Primer sequences were designed according to cDNA  
 sequence from Genbank® (Table 1).

Samples were deemed positive at any given cycle when the  
 value of the emitted fluorescence was greater than the  
 threshold value calculated by the instrument's software  
 (Sequence Detector Ver. 1.9.1). The threshold cycle (Ct),  
 which is defined as the cycle at which PCR amplification  
 reaches a significant value (i.e. usually 15 times greater than  
 the standard deviation of the baseline), is given as the mean  
 value. Relative expression of each mRNA was calculated by  
 the ΔCt method (where ΔCt is the value obtained by  
 subtracting the Ct value of GADPH mRNA from the Ct  
 value of the target mRNA), specifically, the amount of target  
 mRNA relative to GADPH mRNA is expressed as 2<sup>-ΔΔCt</sup>.  
 Data are expressed as the ratio of the target mRNA to  
 GADPH mRNA. Each PCR run included a no-template  
 control and a sample without reverse transcriptase.

## Statistical analyses

Data are presented as mean ± standard error (SE) of each  
 group. All statistical comparisons were performed between  
 the control and treated groups for each period of study: 45, 60  
 and 90 PND. Comparisons were analyzed by Student's *t* test.  
 Differences of *p* < 0.05 were considered significant. Litters  
 with *n* = 8 per each treated or control group were evaluated  
 in every case.

## Results

No differences were observed in food and water consumption  
 between control and treated groups. Maternal exposure to  
 70 mg/kg/day 2,4-D had no effect on body weight gain during  
 gestation or lactation, on the number of pups born or on post-  
 natal mortality. In agreement with our previous study, 2,4-D  
 reduced slightly the pup weight gain (8–12%) (Bortolozzi  
 et al., 1999).

Absolute and relative VP weight decreased significantly  
 in the treated groups when compared with controls (47.2%  
 and 39.7% at PND 45 and 54.9% and 33.6% at 60  
 PND, respectively). However, absolute and relative VP  
 weight increased at PND 90 (19.1% and 26.1%, respectively)  
 (Table 2).

## Effects of 2,4-D on the histology of rat prostate

As shown in Figure 2(A, C and E), the alveoli of control  
 prostates were lined with a layer of tall columnar epithelial

361 cells with a high cytoplasm/nuclear ratio. The luminal  
 362 epithelial cells showed a significant reduction in cytoplasmic  
 363 area after 2,4-D treatment in the VP at PND 45 and 90  
 364 (Figure 2B and F). The increase in luminal volume was  
 365 accounted for by a significant ( $p < 0.01$ ) decrease in the  
 366 average cell number per unit area, reaching 29.5% of control  
 367 rats (Table 3).

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371 Table 2. Body, absolute and relative prostate weight of male rats,  
 372 controls and 2,4-D treated with 70 mg/kg/day.

374 Age	Body weight (g)	Absolute prostate weight (g)	Relative prostate weight (g/g bw)
376 45 PND			
377 Control	139.83 ± 2.61	0.091 ± 0.005	0.066 ± 0.003
378 2,4-D	121.36 ± 3.72**	0.048 ± 0.006***	0.039 ± 0.003**
379 60 PND			
380 Control	198.73 ± 5.62	0.253 ± 0.012	0.122 ± 0.004
381 2,4-D	176.51 ± 6.63*	0.114 ± 0.004***	0.081 ± 0.008***
382 90 PND			
383 Control	310.53 ± 4.77	0.301 ± 0.013	0.096 ± 0.003
384 2,4-D	284.30 ± 5.72**	0.343 ± 0.016	0.121 ± 0.005*

384 2,4-D treated versus controls: each value is the mean ± SEM ( $n=8$ ).

385 \* $p < 0.05$ ;

386 \*\* $p < 0.01$ ;

387 \*\*\* $p < 0.001$ .

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390 Figure 2. Prostate for males at 3 ages studied.  
 391 Sections stained with H-E. The bar represents  
 392 50  $\mu$ m (A) and (B) rats of 45 days old, control  
 393 and treated respectively. (C) and (D) rat 60-  
 394 day-old control and treated. (E) and (F) rat  
 395 90-day-old control and treated. Lumen (L) of  
 396 prostatic alveoli and prostatic epithelium  
 397 (arrow).

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## Effects of 2,4-D on AR expression

In the 2,4-D-treated rats, VP AR protein abundance decreased significantly at PND 45 (22%). However, we observed an increase in AR (37.7%) with respect to controls (Figure 3) at PND 90.

## Effects of 2,4-D on T, DHT, GH and IGF-I circulating levels

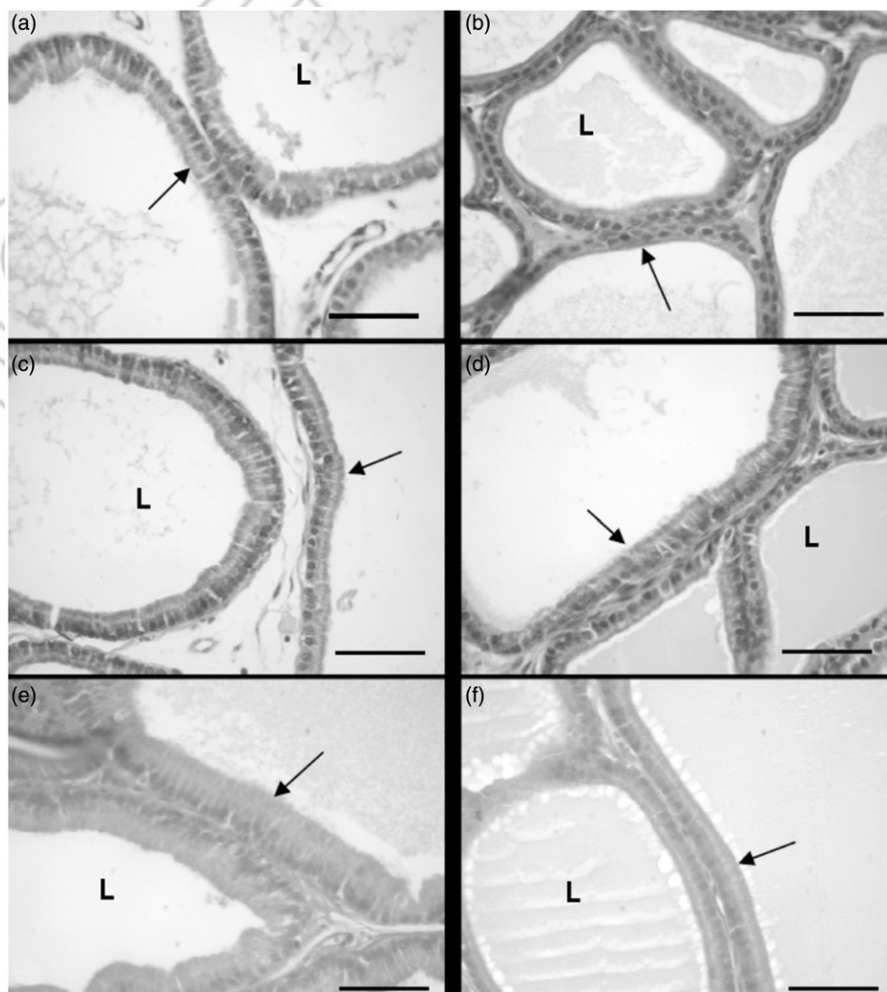
T and DHT serum concentrations at PND 45 and 60 were significantly lower than controls (97% and 96%; 88% and

Table 3. Effects of 2,4-D (70 mg/kg/day) on the epithelial thickness and cell numbers per selected field.

Age	Epithelial thickness ( $\mu$ m)	Cell number
45 PND		
Control	12.9 ± 0.8	52.6 ± 1.2
2,4-D	8.2 ± 0.4*	48.5 ± 1.5
60 PND		
Control	12.2 ± 1.2	51.7 ± 2.5
2,4-D	10.3 ± 1.5	58.1 ± 2.4
90 PND		
Control	16.8 ± 0.9	55.4 ± 2.1
2,4-D	9.8 ± 1.1*	39.1 ± 1.5*

Data are expressed as mean ± SE for at least eight rats for each experimental group.

\* $p < 0.01$  with reference to control values.



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481 76%, respectively) (Figure 4A and B). However in adult  
482 treated animals (PND 90), the levels of both androgens were  
483 similar to control values.

484 IGF-1 serum levels were significantly reduced by 2,4-D  
485 at all ages compared with the respective controls (70%,  
486 55% and 71% at PND 45, 60 and 90, respectively)  
487 (Figure 4C).

488 Conversely, 2,4-D did not affect serum GH levels at any  
489 age studied (Figure 4D).

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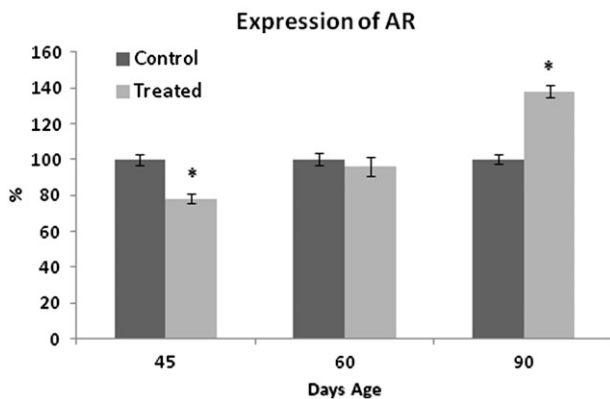


Figure 3. Effects of 2,4-D on the expression of AR in the ventral prostate gland. Rats were treated as described in Figure 1. Densitometric scanning of the AR bands after being normalized to the levels of  $\beta$ -actin. Data are expressed as the mean of 8 samples  $\pm$  SEM. Control, its value was considered as 100% of the intensity of the band.

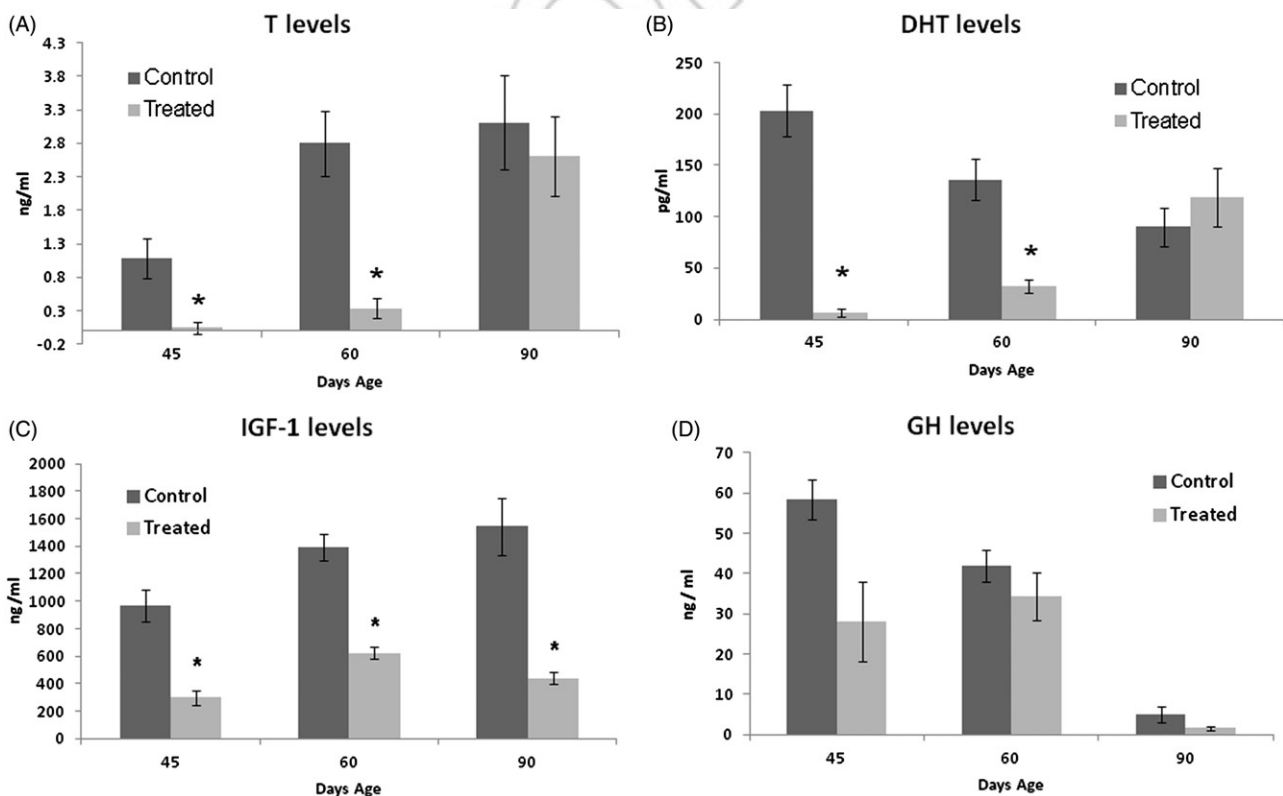


Figure 4. Serum T (A), DHT (B), IGF-I (C) and GH levels (D) expressed as ng/ml in male rats treated with 2,4-D at all ages studied. Control (black bar) and treated group (70 mg/kg/day, gray bar). Data are expressed as mean  $\pm$  SE for at least eight rats for each experimental group. (\*) Significance levels observed are  $p < 0.05$  in comparison with control group.

## Effect of 2,4-D on AR, IGF-1 and IGF-1R mRNA from VP 541

To assess the effect of 2,4-D on VP at 45, 60, and 90 days of age, AR, IGF-1 and IGF-1R mRNA levels of in VP were determined by real time quantitative PCR. As indicated in Figure 5(A), IGF-1 mRNA levels decreased significantly in VP from treated groups with compared controls at PND 45 and 60 (31% and 32%, respectively). On the other hand, IGF-1R mRNA levels increased significantly 20% and 42% with respect to controls at PND 45 and PND 60, respectively (Figure 5B). However, 2,4-D treatments did not affect AR mRNA levels at any age studied (data not shown).

## Discussion 553

Many studies have focused on chemicals that modify the function of the endocrine system. Depending on the beginning and length of exposure, such chemicals alter growth and development of hormone-sensitive organs such as the prostate gland.

The present study shows that rats exposed through the mother during pregnancy and postnatal life until weaning and treated later during development with 70 mg/kg/day of 2,4-D through diet, did not show external signs of toxicity, such as changes in body weight of pups at birth or fetal toxicity. However, a slight decrease in body weight (between 8 and 12%) at sacrifice (45, 60 and 90 days of age) was detected. In no case such decrease surpassed 15% and therefore, according to previous data from our laboratory, it was not considered to be toxicologically relevant since it did not critically affect the overall development of the animal (Bortolozzi et al., 1999).

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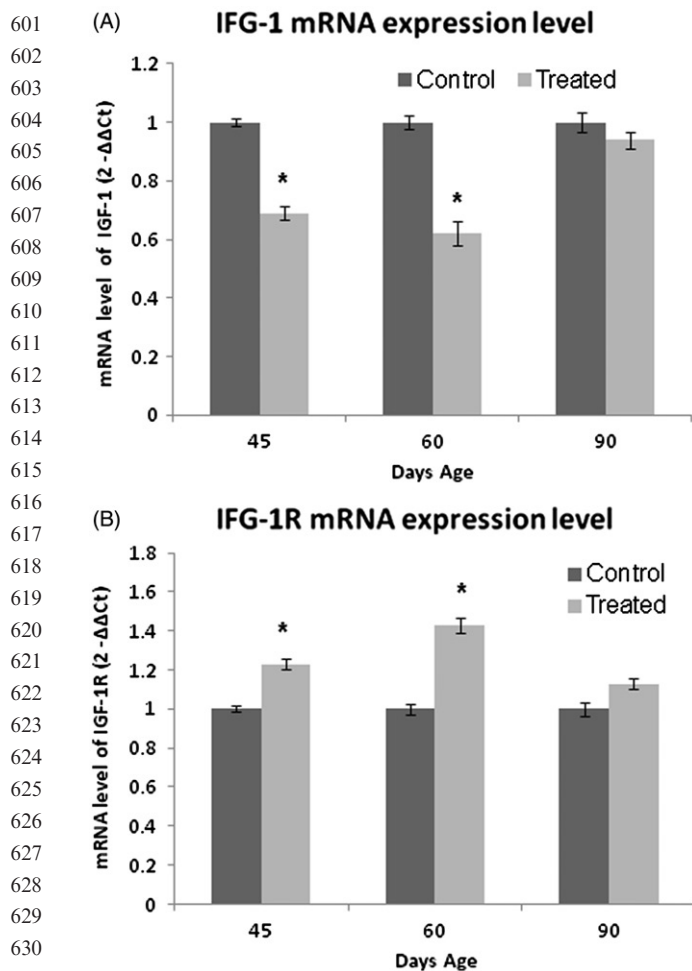


Figure 5. Effect of 2,4-D on IGF-1 (A) and IGF-1R (B) mRNA expression level in rat ventral prostate at the ages of 45, 60 and 90 days by real time quantitative PCR. The housekeeping gene GAPDH was used as an internal positive control standard for quantitative analysis. The relative expression of target genes was calculated using  $2^{-\Delta\Delta C_t}$ . At each age controls were assigned the reference value 1 and all data were expressed as mean  $\pm$  SEM of at least 4 independent determinations. (\*) Significance levels observed are  $p < 0.05$  in comparison with control group.

The neonatal development period is considered the most vulnerable to the action of xenobiotics (Dencker & Eriksson, 1998). Pharmacological doses of diethylstilbestrol (DES) (Singh & Handelsman, 1999) and vinclozolin (Yu et al., 2004) in rats exposed during development produced a reduction in the growth and size of the prostate gland. Moreover, *in utero* exposure to these chemicals induced a higher incidence of prostate lesions in old age, including atrophy and prostatitis (Cowin et al., 2008).

In this work, we show that 2,4-D diminished VP weight as well as height of epithelial the cell layer of the alveoli in animals of 45 and 60 days of age. These data correlate with previous results from our laboratory showing delayed puberty in male rats treated with the herbicide, evidenced as a decrease in the number of sperm cells with normal morphology (Madariaga, 2007). This observation is supported by the markedly low T levels observed in peri and pubertal (45 and 60 days) animals reported in this study. At 60 days of age, the low T level was accompanied by similarly low levels of DHT, the androgen responsible for stimulating growth and

function of the prostate gland, thus explaining the delayed organ development.

Since androgens exert their action on the prostate gland through AR, AR levels were also determined. It has been reported that certain chemicals, including bisphenol A, nonylphenol and fenthionare capable of interacting directly with the AR, activating transcription of AR-dependent genes in mammalian cells (Kitamura et al., 2003; Lee et al., 2003). It has also been shown that estrogenization during development reduces AR protein levels, which also decreases the response capacity to DHT and T, without modification of AR mRNA expression (Prins, 1997). The decrease in receptor protein levels was due to increased proteolytic degradation (Woodham et al., 2003). In the present work we found that AR levels, were decreased in treated animals compared to controls at 45 days of age, without changes in mRNA expression, indicating that the herbicide, may be increasing the degradation or decreasing AR protein synthesis in peripubertal rats. On the other hand, since AR expression in epithelial prostatic tissue increases with age (Prins & Birch, 1995), decreased AR protein expression may be a consequence of delayed puberty and maturation of reproductive organs.

Even though the prostate is sensitive to T and DHT during development, ductal branching morphogenesis occurs before puberty, when androgen levels are low (Donjacour & Cunha, 1988). At this early stage, circulating IGF-1 and GH are elevated and therefore play a critical role in prostate development (Sandhu et al., 2006). For this reason, serum levels of GH were evaluated showing the typical pattern observed during growth, with high levels that decline with age in the young control animals, and no significant effect of 2,4-D treatment. Since GH levels are typically pulsatile, and measurement at only one point in time may not reflect possible effects of the herbicide upon GH pulsatility (amplitude or frequency), we decided to determine circulating IGF-1 that, as previously mentioned, displays more stable circulating levels. This approach, reflects more accurately the state of GH secretion, since systemic and local IGF-I synthesis depends largely on GH (Rajaram et al., 1997). IGF-1 levels decreased in 45 and 60 day old animals treated with 2,4-D. IGF-1  $-/-$  mice show overall reduction in prostate size and specific structural deficit, including the number of ductal terminals. Administration of IGF-1 reverses these effects (Ruan et al., 1999), demonstrating the role of circulating IGF-1 in prostate development. Therefore we can conclude that at least some of the effects of 2,4-D in the development of the gland in prepubertal and pubertal animals may be mediated through the decrease in circulating IGF-1 produced by the herbicide.

In addition to hormonal influences and circulating IGF-1, local factors may play a critical role in prostate normal growth. IGF-1 is produced by stromal cells. acting as a paracrine factor on epithelial cells through IGF-1R (Lipschutz et al., 1999). We found diminished IGF-1 mRNA level at 45 and 60 days of age in treated animals, accompanied by increased expression levels of its receptor. IGF-1 mRNA expression at the tissue level is also regulated by circulating GH and thus the herbicide could affect IGF-1 transcription on tissues directly, or through undetected effects on GH

721 pulsatility, as mentioned above. On the other hand, the  
722 increase in IGF-1R expression levels at the local level could  
723 be compensating for the decreased local or circulating IGF-1.

724 Kim et al. (2005) reported that 2,4-D (50 mg/kg/day)  
725 caused an increase in the weight of androgen-dependent  
726 tissues. When administered simultaneously with T to  
727 castrated adult animals, an increase in the VP weight was  
728 observed. In this work similar results were found in 90 day old  
729 animals, in the presence of normal T and DHT levels. Thus, it  
730 can be concluded that the increase in gland weight at this age  
731 may be produced by 2,4-D acting synergistically with the  
732 normal circulating androgens.

733 It is worth mentioning that, some pesticides such as  
734 atrazine (nonsteroidal compound) produce different changes  
735 in the prostate. Also, a brief exposure to this compound  
736 through breastfeeding on postnatal days 1–9, suppressed  
737 suckling induced PRL release in the mothers and resulted in  
738 an adverse effect such as inflammation of the prostate in adult  
739 male offspring (Stoker et al., 1999). Also in a previous work  
740 from our laboratory, exposure to 2,4-D during lactation  
741 (postnatal days 1–16) resulted in a partial blockade in  
742 suckling induced oxytocin and PRL release in treated mothers  
743 (Stürtz et al., 2010). Therefore, the observed increase in  
744 prostate gland weight may not only be associated with  
745 hormonal induction, but also with the effects that the  
746 herbicide may have caused during lactation. This is due to  
747 the fact that, in our experimental scheme, animals are exposed  
748 from gestation to adulthood.

749 It has been reported that some phytochemicals, depending  
750 on their concentration, may exhibit agonist/antagonist activity  
751 on the steroid hormone system. For example, *in vitro* studies  
752 have shown that quercetin at low concentrations acts as  
753 agonist (Taepongsorat et al., 2008). The *in vivo* effect of this  
754 compound is not well known, but it has been observed that  
755 treatment with quercetin caused a dramatic expansion of the  
756 prostatic lumen, which was filled with secretion, indicating  
757 that quercetin may have increased the secretory activity of the  
758 epithelial cells. Likewise, the increase in luminal volume  
759 produces a decrease in the number of cells per unit area  
760 compared to the control (Ma et al., 2004). In the histological  
761 and morphometric observation of prostates of 90 day old male  
762 rats treated with 2,4-D, we found changes similar to those  
763 found in the previously mentioned paper, suggesting that the  
764 herbicide may act similarly. Additionally, although androgens  
765 and IGF-1 levels were restored in the adult glands of treated  
766 animals, epithelial tissue morphology remained altered, as it  
767 is expressed in the decrease in height of its cells.

768 Administration of low doses of estrogen to the mother  
769 during gestation increases the size of the prostate as well as  
770 AR expression in the adult offspring (Nagel et al., 1997).  
771 Studies performed on cell line cultures of human prostate  
772 cancer indicate that 2,4-D and 2,4-dichlorophenol (DCP) in  
773 combination with DHT have androgenic activity in cell  
774 proliferation and induce transactivation by androgen, possibly  
775 through increased translocation to the nucleus without  
776 alteration in AR expression levels (Kim et al., 2005). On  
777 the contrary, in this work we found a 40% increase in AR  
778 protein levels for the treated group at 90 days of age.  
779 However, the aforementioned results were observed in  
780 isolated tumoral epithelial cells, and thus cannot always be

781 interpreted as a reflection of what would take place in normal  
782 tissue *in vivo*, as the prostatic epithelial tissue depends on  
783 stromal factors for its correct development and maintenance.  
784 Since AR mRNA levels in adult animals were not modified,  
785 the action of 2,4-D could be exerted on AR protein synthesis  
786 or degradation rates, increasing its abundance. It is interesting  
787 to point out that AR synthesis is induced by DHT, thus the  
788 herbicide would enhance this effect in animals of 90 days of  
789 age, since in the absence of androgen, 2,4-D alone did not  
790 increase AR expression in mature animals.

## 791 Conclusion

792 2,4-D belongs to the group of non-steroidal environmental  
793 substances with the ability to act as endocrine disruptors  
794 (EDs) (Diamanti-Kandarakis et al., 2009). The National  
795 Institutes of Health (USA) considers 2,4-D as a potential ED  
796 (Anon, 2004). Numerous papers indicate that IGF-1 and  
797 RIGF-1 expression are influenced by steroid and peptide  
798 hormones (Yu & Rohan, 2000). Therefore, variations detected  
799 in the members of the IGF family could be partially due to a  
800 modification of gonadal steroid concentrations and their  
801 receptors since the herbicide produces:

- 802 (1) Decrease of serum androgen and AR levels in the  
803 prostate of male pups at their youngest age. This effect is  
804 similar to the one observed for environmental substances  
805 with estrogenic effect (Singh & Handelsman, 1999). The  
806 opposite effect was observed in adult age, where T levels  
807 were normal and AR protein expression was induced in  
808 the prostate was induced.
- 809 (2) Serum IGF-1 levels were diminished in pups at the three  
810 ages studied.
- 811 (3) VP from prepub and puber males showed a decrease in  
812 IGF-1 mRNA levels along with an increase in its receptor  
813 expression.

814 These results indicate that 2,4-D could behave as an ED,  
815 affecting prostate development. Future research should focus  
816 on the nature of the major deleterious effects produced by the  
817 herbicide on VP, establishing whether if such changes are  
818 permanent or reversible and able to if they affect male fertility  
819 and/or prostate function.

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## 824 Declaration of interest

825 The authors report no declarations of interest.

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