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# **6 Fundamental research questions in subterranean biology**

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82	
83	ABSTRACT
84	Five decades ago, a landmark paper in Science titled The Cave Environment heralded caves as
85	ideal natural experimental laboratories in which to develop and address general questions in
86	geology, ecology, biogeography, and evolutionary biology. Although the 'caves as laboratory'
87	paradigm has since been advocated by subterranean biologists, there are few examples of studies
88	that successfully translated their results into general principles. The contemporary era of big
89	data, modelling tools, and revolutionary advances in genetics and (meta)genomics provides an
90	opportunity to revisit unresolved questions and challenges, as well as examine promising new

91 avenues of research in subterranean biology. Accordingly, we have developed a roadmap to guide future research endeavours in subterranean biology by adapting a well-established 92 methodology of 'horizon scanning' to identify the highest priority research questions across six 93 subject areas. Based on the expert opinion of 30 scientists from around the globe with 94 complementary expertise and of different academic ages, we assembled an initial list of 258 95 96 fundamental questions concentrating on macroecology and microbial ecology, adaptation, 97 evolution, and conservation. Subsequently, through online surveys, 130 subterranean biologists 98 with various backgrounds assisted us in reducing our list to 50 top-priority questions. These 99 research questions are broad in scope and ready to be addressed in the next decade. We believe this exercise will stimulate research towards a deeper understanding of subterranean biology and 100 foster hypothesis-driven studies likely to resonate broadly from the traditional boundaries of this 101 field. 102

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104 *Key words*: biospeleology, cave biology, expert opinion, groundwater, horizon scanning,

105 research questions, stygofauna, troglobionts.

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# 123 I. INTRODUCTION

In the era of the Internet, social media, and open-access mega-journals, the amount of accessible 124 125 scientific information is overwhelming (Landhuis, 2016; Wakeling et al., 2016; Fire & Guestrin, 2019; Jarić et al., 2020). It is estimated that more than 50 million peer-reviewed scientific papers 126 exist (Jinha, 2010) and about 1.5 million new articles are published every year (Laurance *et al.*, 127 128 2013). To capitalize on the volume of this information and make the most of it (e.g. Ioannidis, 2005; Jeschke et al., 2019), it is becoming increasingly important for scientists to explore 129 130 effective ways to capture the latest advances in their field or related fields of research. Horizon 131 scanning – i.e. the systematic examination of information to identify emerging issues and opportunities in a given research area - has become a useful tool to summarize and determine 132 research priorities and agendas (Sutherland et al., 2011). The most important questions in 133 134 ecology (Sutherland et al., 2013; McGill et al., 2019), island biogeography (Patiño et al., 2017),

and microbiology (Antwis et al., 2017), the annual identification of emerging issues in global 135 conservation (Sutherland et al., 2020), as well as the 100 articles that every ecologist should read 136 137 (Courchamp & Bradshaw, 2018), are all instructive examples where horizon scanning has successfully synthesized trends or highlighted the most promising future research avenues. 138 Fifty years ago, in a landmark *Science* paper titled *The Cave Environment*, Poulson & 139 140 White (1969) heralded caves as 'natural laboratories', i.e. simplified settings that can be used to understand the principles governing the dynamics of more complex environments. Characterized 141 142 by stringent environmental constraints and simple communities, subterranean habitats have been regarded as ideal systems for investigating many of the unresolved questions in ecology, 143 biogeography, and evolutionary biology (Juan et al., 2010; Sánchez-Fernández et al., 2018; 144 Mammola, 2019). Scientists have also studied subterranean organisms to understand human 145 diseases such as autism (Yoshizawa et al., 2018), diabetes (Riddle et al., 2018), and cancer 146 (Gatenby, Gillies & Brown, 2011), to investigate the engineering potential of biologically 147 148 inspired materials (Lepore *et al.*, 2012), and to discover new drugs and pharmaceutical products (Cheeptham et al., 2013). Others have even looked at caves through the lens of astrobiology, 149 150 showing that the subterranean microbiome might hold clues to life beyond Earth (Northup *et al.*, 151 2011; Popa *et al.*, 2011).

Although the 'caves as laboratory' paradigm is often advocated by subterranean biologists, examples of studies that have successfully translated their results into general principles remain few in number. Five decades after Poulson & White (1969), subterranean biology is entering a new research era dominated by big data (Zagmajster *et al.*, 2019), modelling tools (Flôres *et al.*, 2013; Mammola & Leroy, 2018), and increasingly cheaper molecular approaches (Pérez-Moreno, Iliffe & Bracken-Grissom, 2016; Lefébure *et al.*, 2017).

Concomitantly, we are facing a global crisis that is negatively impacting subterranean 158 159 biodiversity (Mammola et al., 2019b; Boulton, 2020). Therefore, the time is ripe to review the 160 outstanding challenges faced by this broad-in-scope discipline, as well as promising new research avenues where subterranean-based studies may be helpful in answering general and 161 broadly scoped questions. Because gathering multiple views on such an extensive subject is 162 163 difficult, we relied on the well-established methodology of horizon scanning to identify 50 fundamental, but unresolved questions in subterranean biology. With this intellectual exercise, 164 we aimed to develop a roadmap that will guide future research endeavours and stimulate 165 hypothesis-driven studies likely to resonate beyond the boundaries of this discipline. 166

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## 168 II. HORIZON SCANNING PROTOCOL

#### 169 (1) Initial list assembly

We used horizon scanning methodology (Sutherland *et al.*, 2011) and adapted the approach 170 171 developed by Patiño et al. (2017) to identify priority research questions in subterranean biology. Survey coordinators (S.M. and P.C.) identified seven subject areas within the subterranean 172 biology discipline (Table 1), namely: (1) Adaptation, (2) Origin and evolution, (3) Community 173 174 ecology, (4) Macroecology and biogeography, (5) Conservation biology, (6) Microbiology and applied topics, and (7) Other topics. We included the latter subject area to cover additional topics 175 176 or ideas that departed from the six core subject areas and may have been overlooked. For each 177 subject area, survey coordinators invited a senior researcher (highlighted with asterisks in Table 178 1) to act as panel coordinator, with the task of establishing an international panel of experts to identify and formulate a set of fundamental questions. Each panel coordinator selected and 179 180 invited three or four members to join their panel, which included at least one early-career

181 scientist (i.e. a postdoc or researcher with less than 10 years of experience) to obtain a multigenerational perspective on the different topics. Survey coordinators encouraged panel members 182 183 to consult broadly with colleagues and select additional researchers to join their panels if deemed important in providing complementary expertise. In assembling the panels, our goal was to 184 maximize multidisciplinarity, while ensuring that research interests within the seven panels 185 186 covered a broad array of geographic areas, model organisms, and networks of international collaborators. Members of each panel identified at least 20 questions that they viewed as 187 fundamental within their subject area and thus likely to advance the field significantly. 188

In total, we assembled 258 questions, which were screened for duplication or ambiguity 189 by the survey coordinators. In this phase, survey coordinators purged most subterranean-specific 190 jargon from questions and homogenized wording to ensure that all questions were presented in a 191 clear and straightforward manner. Therefore, throughout the survey we operated under the 192 assumption that all questions were characterized by a similar degree of readability (Plavén-193 194 Sigray *et al.*, 2017). After the cleaning procedure and removal of duplicate questions, we assembled a final list of 211 survey questions (hereafter 'List #1'). In assembling List #1, we 195 subsumed questions identified by the panel focusing on 'Other topics' into the six main subject 196 197 areas.

198

# 199 (2) Voting procedure and selection of 50 top-priority questions

We subjected List #1 to an initial round of online voting by all panel members (Survey #1) to select the most voted 20 questions for each of the six subject areas (Fig. 1). Voting was a binary choice, whereby participants scored each question as either of 'major' or 'minor' importance. We randomized question order for each participant. We repeated this voting protocol in all subsequent online surveys. Each panel member voted on all questions irrespective of subject
area, although votes by panelists on their subject area were disregarded in the final ranking of
Survey #1. As a result, survey coordinators culled List #1 to the 120 most-voted questions (20
questions from each of six subject areas), referred to as List #2, thus balancing the number of
questions in subsequent voting rounds.

209 We then subjected List #2 to online voting (Survey #2) by inviting a broad community of subterranean biologists including ca. 170 members of the International Society on Subterranean 210 211 Biology (ISSB), ca. 50 members of the European Cave Organism Network, ca. 100 members of the Anchialine mailing list, as well as other working groups and email listservs related to 212 subterranean biology that we could identify (e.g. national biospeleological groups). Note that 213 members of these different groups often overlapped and some of the emails were no longer 214 active. We estimated that Survey #2 reached an upper boundary of between 200 and 250 unique 215 recipients. Of these, 133 recipients completed the online survey. 216

At the end of Survey #2, we gave participants the opportunity to submit one additional question if they felt this question was missing from List #2. Thus, 25 additional questions were added to the third list of questions (List #3). Questions in List #3 were voted on by all panel members (Survey #3), and then ranked (by percentage of 'major importance' votes per question) together with the 120 questions from List #2. Finally, we selected the highest ranking questions to assemble a list of 50 top-priority questions.

223

## 224 (3) Caveats on interpretation

Some general caveats should be recognized when interpreting the results of any horizon scanning
survey (e.g. Sutherland *et al.*, 2011, 2013; Seddon *et al.*, 2014; Patiño *et al.*, 2017). First, the

227 background knowledge and intellectual passions of the experts involved may introduce subjectivity in the formulation of the initial list of topics and questions. Second, subjectivity 228 229 likely plays a role throughout the voting process, as any voting outcome may be affected by the interests of a particular group of participants. In our case, potential biases in the composition of 230 subterranean biologists sampled may have influenced the final selection of the top-priority 231 232 questions to an extent difficult to quantify precisely. For example, questions related to microbiology received the lowest share of 'major importance' votes (mean  $\pm$  SD: 0.69  $\pm$  0.01). It 233 is understood that microbiology topics are not less important or timely, it is simply that 234 microbiologists are probably underrepresented in the subterranean biology community. Also, an 235 imbalance in the expertise of participants may explain the substantial difference in how the 236 highest priority questions were parsed across the six subject areas – from four in 'Community' 237 ecology' to 12 in 'Conservation biology'. 238

To address these potential shortcomings, we adopted four countermeasures. First, we 239 240 increased the survey audience, by addressing the questionnaire to different groups and associations of subterranean biologists. Second, we diversified the expertise of panel members 241 by including early-stage to mid- and late-career researchers from different disciplines, research 242 243 laboratories, and geographic areas. Third, we included a seventh panel ('Other topics') specifically to fill the gaps in the initial composition of proposed questions. Indeed, it has been 244 245 argued that in horizon scanning, the initial division into subject areas may limit lateral thinking 246 (Sutherland *et al.*, 2013). Finally, by allowing voters to suggest additional questions when voting 247 in the survey, we were able to capture the range of priority topics better.

We are confident these practices minimized some of the biases inherent to this study.Importantly, we believe this 50 top-priority survey served to highlight some of the most timely

and challenging areas of interest in current and future research, rather than providing a

251 comprehensive synthesis of research needs in modern subterranean biology.

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# 253 III. SUMMARY OF THE HORIZON SCAN

In Survey #1, the percentage of 'major importance' votes ranged between 89% (top-voted
question) and 4% (least-voted question). In the extended online voting (Survey #2), 133 voters
participated, of which 71% identified 'subterranean biology' as their primary field of research.

257 Although voters' gender was slightly skewed toward males (76 men versus 57 women),

deviation from the 1:1 male: female ratio was not significant ( $\chi^2 = 2.71$ ; d.f. = 1; P = 0.10),

259 indicating that our sample was not gender-biased. 45% of survey voters were experienced

researchers, with an academic age of more than 10 years since they earned their PhD, while 29% were researchers within 10 years from their PhD. PhD and undergraduate students accounted for 16% of voters. The remaining 10% of participants were other professionals, such as research and field technicians or recreational cavers.

During Survey #2, participants suggested 28 additional questions; three questions were duplicates and were thus excluded. The remaining 25 questions were evaluated during Survey #3, and three made it to the 50 top-priority list. The lower threshold for questions was 67% of 'major importance' votes, whereas the top-voted question garnered 91% votes (Fig. 1).

In the following, we present the 50 top-priority questions in subterranean biology according to the results of Surveys #2 and #3 (the full list of questions is provided as online supporting information in Appendix S1). For clarity, questions were compiled into our six subject areas. We provide information about each question's final rank (#) and percentage of 'major importance' votes received (%), and highlight the three questions suggested by the 273 Survey #2 participants with an asterisk (\*). A glossary of terms is available in Table 2.

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# 275 IV. ADAPTATION

- 276 Q1 What are the drivers of adaptive evolution in caves? [#1; 91%]
- 277 Q2 What are the main constraints to subterranean adaptation? [#4; 83%]
- 278 Q3 What are the degrees of adaptive plasticity of organisms across different subterranean
- 279 environments? [#9; 78%]
- 280 Q4 Which traits of subterranean organisms should be considered as 'adaptive'? [#11; 78%]
- 281 Q5 How have morphological and behavioural traits co-evolved in subterranean organisms?
- 282 [#14; 76%]
- 283 Q6 What is the level and nature of reproductive isolation between cave and surface populations
- and what reproductive barriers are typically involved? [#19; 75%]
- Q7 Do similar traits evolve repeatedly in subterranean organisms due to changes in the same
- genes, genetic pathways, and/or developmental processes? [#23; 73%]
- 287 Q8 Have subterranean species evolved a distinct set of convergent behaviours? [#26; 72%]
- 288 Q9 Are there common developmental pathways that promote or constrain subterranean
- adaptation? [#29; 72%]
- $290 \quad Q10 Do traits that constitute reproductive isolation evolve in the same way across independent$
- closely related subterranean populations or species? [#42; 70%]
- 292
- 293 The morphology of subterranean organisms, which show bizarre convergent adaptations even
- across different animal phyla, has historically attracted the attention of generations of scientists
- 295 (Juan et al., 2010) including Charles Darwin (1859). Therefore, it is no surprise that subterranean

biologists participating in this survey greatly valued the role of subterranean habitats as natural
laboratories for the study of adaptive evolution. Ten questions focusing on adaptation were
included in our top-50 list (Fig. 1).

Colonization of suitable habitat is the initial event leading to subterranean adaptation 299 (details in Section V). Whatever the mode or pathway, colonizers often experience a significant 300 301 change upon entering the subterranean environment (i.e. complete darkness), which results in visual sensory deprivation, challenges in locating mates and food, limited or modified food 302 resources, and physical barriers to dispersal. Adaptive responses to these factors may involve the 303 action of selection on plastic traits already existing in the colonizers (i.e. phenotypic plasticity; 304 Bilandžija et al., 2020), standing genetic variation, or new beneficial mutations. Understanding 305 which of these environmental factors and adaptive responses play a primary role in subterranean 306 adaptation, either acting alone or in various combinations, was the most important question (Q1) 307 in our survey, selected by 91% of participants. Yet, given that some higher taxa are missing or 308 309 understudied in caves (Culver & Pipan, 2019), it remains unclear what are the main constraints to subterranean adaptation (Q2) and whether specific exaptations facilitate successful 310 colonization events (see also Q11 in Section V). Resolving how many phenotypes of 311 312 subterranean dwellers depend on genetic and developmental constraints (Q9), or reflect entrapment at local peaks in adaptive landscapes or recent invasions with insufficient time for 313 314 selection to alter traits, is one of the future challenges for evolutionary biologists.

Additional high-priority questions were focused on subsequent refinements of the initial adaptive responses, such as the repertoire of adaptive plasticity (Q3), the degree to which preexisting genetic variation contributes to subterranean phenotypes, and which traits of subterranean organisms can be considered as adaptive (Q4). Historically, reduction or loss of

traits such as eyes and pigmentation was thought to be driven by random mutations and genetic 319 drift or by natural selection, either directly or indirectly. This controversy has continued to the 320 321 present, with strong adaptationist (Carlini & Fong, 2017) and non-adaptationist (Wilkens & Strecker, 2017) viewpoints. Depending on the species or ecological context, it is possible that all 322 323 of these mechanisms have roles in subterranean adaptation. Resolving this debate will require 324 explanations at the molecular, cellular, and developmental levels in multiple lineages (Jeffery, 2005), and the integration of this information to infer whether convergent traits evolve repeatedly 325 in subterranean animals due to changes in the same or different genes, genetic pathways, and 326 developmental processes (Q7). Answers to all these questions will contribute to our 327 understanding concerning why some species adapt rapidly and evolve when facing new 328 environmental conditions, inside or outside caves, which is a critical question given global 329 climate change (Walther *et al.*, 2002). In turn, this could provide insights about adaptive 330 processes occurring in other ecological settings with a similar set of environmental conditions 331 332 (e.g. permanent darkness, constancy in climatic conditions, food scarcity), such as deep-sea habitats (Trontelj, Borko & Delić, 2019; Mammola, 2020). 333

334 Once survival in a subterranean habitat is ensured, the successful colonizers are subject to 335 adaptive morphological and behavioural (co-)evolution (Q5). Many behavioural changes are probably influenced by the essential requirements of finding food and mates in darkness, and 336 337 may be convergent across different subterranean lineages (Q8). Also, some subterranean animals 338 suddenly attain a new status at the top trophic level and predator release occurs. For example, in 339 the Mexican tetra, Astyanax mexicanus (De Filippi) (Actinopterygii: Characidae), the workhorse of adaptive evolution studies in caves (Jeffery, 2009; Wilkens & Strecker, 2017; Torres-Paz et 340 341 al., 2018), this new ecological status of an apex predator facilitated the evolution of a range of

behaviours that may not be sustainable in a predator-limited surface environment (Yoshizawa *et al.*, 2010; Hyacinthe, Attia & Rétaux, 2019).

344 Most subterranean organisms may also face subsequent invasions of their habitats by new colonizers, of both former surface-dwelling conspecifics (if they are still extant) and other 345 competing species (e.g. Howarth et al., 2007; Wynne et al., 2014). Therefore, to understand 346 347 subterranean adaptations fully, it is crucial to explore the degree and nature of reproductive isolation between the subterranean-adapted lineages and invading surface conspecifics (Q6). The 348 majority of subterranean animals probably arose through the process of ecological speciation in 349 which reproductive isolation evolved as a response to divergent selection between environments 350 351 (Niemiller, Fitzpatrick & Miller, 2008; Mammola et al., 2018). Thus, many subterranean adaptations should at least indirectly favour non-random mating between individuals of the 352 derived subterranean and ancestral surface populations. Understanding this will help to address 353 whether traits that constitute reproductive isolation evolve in the same way in independent 354 355 closely related subterranean populations or species (Q10), and therefore whether and how often parallel speciation occurs in the subterranean realm. Ultimately, this would shed new light 356 357 concerning the intriguing hypothesis on the predictability of evolution (Blount, Lenski & Losos, 358 2018).

359

#### 360 V. ORIGIN AND EVOLUTION

Q11 – Which traits present in surface species (exaptations) facilitate successful subterranean
colonization and adaptation? [#12; 77%]

Q12 – How do, and which, patterns of subterranean species diversification vary across taxa and
habitats? [#13; 77%]

- Q13 What evolutionary processes most commonly triggered radiations of subterranean
  organisms? [#15; 76%]
- 367 Q14 Do subterranean organisms lack genetic variation and thus the ability to adapt to a
- 368 changing environment? [#16; 75%]
- Q15 Does the timeline of subterranean evolution differ among taxa, types of subterranean
- habitats, different biogeographic areas, and different ecological settings? [#22; 74%]
- Q16 What are the impact(s) of biotic and abiotic factors on speciation? [#28; 72%]
- 372 Q17 Why are some lineages successful at colonizing subterranean habitats while others are
- 373 not? [#35; 71%]
- 374 Q18 How old are subterranean species? [#36; 71%]
- 375 Q19 The role of evolutionary processes (convergence/divergence/evolutionary
- 376 stasis/parallelisms) in subterranean organisms: what are the most common evolutionary
- 377 processes? [#40; 70%]
- $Q_{20}$  Are shallow subterranean habitats a gateway to colonize deep zones and is the evolution
- of deep subterranean species conditioned with a colonization of shallow and later deeper zones?[#41; 70%]
- Q21 What is the rate of evolution of different subterranean traits and does the degree of <math>Q21 What is the rate of evolution of different subterranean traits and does the degree of <math>Q21 What is the rate of evolution of different subterranean traits and does the degree of <math>Q21 What is the rate of evolution of different subterranean traits and does the degree of <math>Q21 What is the rate of evolution of different subterranean traits and does the degree of <math>Q21 What is the rate of evolution of different subterranean traits and does the degree of <math>Q21 What is the rate of evolution of different subterranean traits and does the degree of <math>Q21 What is the rate of evolution of different subterranean traits and does the degree of <math>Q21 What is the rate of evolution of different subterranean traits and does the degree of <math>Q21 What is the rate of evolution of different subterranean traits and does the degree of <math>Q21 What is the rate of evolution of different subterranean traits and does the degree of <math>Q21 What is the rate of evolution of different subterranean traits and does the degree of <math>Q21 What is the rate of evolution of different subterranean traits and does the degree of <math>Q21 What is the rate of evolution of different subterranean traits and does the degree of <math>Q21 What is the rate of evolution of different subterranean traits and does the degree of <math>Q21 What is the rate of evolution of different subterranean traits and does the degree of <math>Q21 What is the rate of evolution of different subterranean traits and does the degree of <math>Q21 What is the rate of evolution of different subterranean traits and does the degree of <math>Q21 What is the rate of evolution of evolut
- subterranean adaptation correlate with duration of subterranean inhabitation? [#44; 69%]
- 383
- 384 Subterranean animals have long interested biologists as evolutionary models. Studies of these
- 385 species have endeavoured to improve our understanding of evolution, its repeatability at the
- 386 phenotypic (Friedrich, 2013; Porter & Sumner-Rooney, 2018), physiological (Jones, Cooper &
- 387 Seymour, 2019), and molecular level (Leys *et al.*, 2005; Bilandžija, Četković & Jeffery, 2012;

Niemiller *et al.*, 2013), its reversibility (Copilaş-Ciocianu *et al.*, 2018), and the role of drift in
morphological changes (Martínez *et al.*, 2017; Wilkens, 2020). The eleven questions identified
highlight how, despite advances in the application of genetic tools and techniques in the last 50
years, fundamental questions regarding the origin and evolution of subterranean animals remain
unanswered.

393 Two high-ranked questions (Q11 and Q17) focused on the traits that enable species to successfully colonize and adapt to subterranean habitats. Additional questions focused on the 394 most common evolutionary processes (Q19), and the influence of biotic and abiotic factors (Q16) 395 that lead to different patterns of diversification across subterranean lineages (Q12). Important 396 subterranean radiations are known in all major taxonomic groups (Deharveng & Bedos, 2019), 397 but only a few of them have been well documented. These include Amphipoda (Zakšek et al., 398 2019), Collembola (Lukić et al., 2019), and Coleoptera (Leys et al., 2003; Faille et al., 2010; 399 Njunjić *et al.*, 2018). Which evolutionary processes best explain these radiations remains highly 400 401 debated (Q13) and it would be particularly interesting to compare and contrast radiations of surface-dwelling plants and animals (Gillespie et al., 2020) with subterranean-adapted species to 402 determine if any universal patterns exist. For many animal groups, subterranean species are 403 404 commonly assumed to have evolved from surface species (Barr & Holsinger, 1985; Peck & Finston, 1993), but recent phylogenetic studies suggest that this assumption may not always 405 406 apply (Faille *et al.*, 2010; Juan *et al.*, 2010; Leijs *et al.*, 2012). Speciation and diversification may 407 also occur within the confines of a subterranean habitat, a process referred to as 'endogenous diversification' (Trontelj, 2019). Moreover, some phylogenetic studies suggested that 408 subterranean colonization is not an evolutionary dead end and surface species may actually arise 409 410 from subterranean ancestors (Prendini, Francke & Vignoli, 2010; Niemiller et al., 2013; Copilaş-

Ciocianu *et al.*, 2018). However, cases of endogenous speciation and 'subterranean to surface'
reversals are potentially confounded by extinction of surface lineages (Juan *et al.*, 2010).
Therefore, new approaches are needed that avoid reliance on phylogenetic methods alone to
improve our understanding of these patterns.

Genetic variation enhances the ability of species to adapt and diversify. Additionally, it 415 416 has been shown that some subterranean species may contain high levels of neutral genetic variation (Buhay & Crandall, 2005; Guzik et al., 2009), but it is still unclear whether neutral 417 mutations equates to high levels of adaptive genetic variation. This underpins the question 418 whether subterranean species lack the ability to adapt to changing environments (Q14), including 419 420 increasing temperatures and the introduction of new pathogens (Mammola et al., 2019c). Such hypotheses are obviously not exclusive to the subterranean environment. However, this 421 ecosystem does provide numerous examples of how low genetic variation was hypothesized to 422 be related to low adaptive capacity, a phenomenon more common underground than at the 423 424 surface (Konec et al., 2015; Lefébure et al., 2017; Fumey et al., 2018). Understanding the timeline and direction of subterranean evolution, as well as the age of 425 426 subterranean species, featured prominently in several questions (Q15, Q18, Q20, Q21). 427 Advances in molecular clock calibration (Drummond et al., 2006) and genomic analyses (Pérez-Moreno *et al.*, 2016) are considerably promising and permit the development of robust time trees 428 429 (Pons et al., 2019). However, these analyses are limited by the availability of extant and fossil 430 taxa and the extinction of surface relatives; the latter makes it difficult to pinpoint the initial 431 colonization time of a subterranean habitat by a given species. This is particularly important for ancient lineages of specialized subterranean organisms with marine origin, which often lack 432 433 surface-dwelling relatives and/or show low levels of fossilization (Pérez-Moreno et al., 2016).

This is unfortunate because many of these basally branching lineages are required to reconstruct
trait evolution of major animal lineages (e.g. Johnson *et al.*, 2012; Khodami *et al.*, 2017; LozanoFernandez *et al.*, 2019).

The genetic basis underlying evolution of subterranean traits, and how they are shaped by 437 natural selection and/or neutral processes, are key factors in determining rates of subterranean 438 439 evolution (Q21). Considerable advances have been made through the study of model subterranean species, especially Astyanax mexicanus and the freshwater isopod Asellus aquaticus 440 441 (L.) (Protas & Jeffery, 2012). These species have several independent and recently evolved subterranean populations, as well as extant surface populations, which can be hybridized in the 442 laboratory. Their features allow for the dissection of genes and mutations responsible for traits 443 related to subterranean life and provide information on the processes (e.g. selection or neutral 444 evolution) that shape their evolution. The role of neutral processes in the evolution of 445 subterranean animals has also been explored using alternative model systems (e.g. dytiscid 446 447 beetles and amblyopsid cavefishes). In both cases, species have been evolving underground for millions of years, which is sufficient to enable the fixation of deleterious mutations in genes 448 under relaxed selection (Niemiller et al., 2013; Tierney et al., 2018). These model organisms 449 450 offer great potential to investigate major questions on the origin and evolution of subterranean animals using comparative genomics, and thus may provide insights for similar processes in 451 452 other, non-subterranean, settings.

# 453 VI. COMMUNITY ECOLOGY

- 454 Q22 What are the main ecological and ecosystem services provided by subterranean
- 455 populations and communities? [#20; 75%]
- 456 Q23 What are the key food-web processes influencing subterranean community dynamics?
- 457 [#24; 73%]
- 458 Q24 How do stochastic events interact with long-term trends in subterranean ecosystems?
  459 [#30; 72%]
- 460 Q25 How do basic life-history characteristics differ among subterranean communities and
- 461 between subterranean and surface communities? [#33; 71%]

462

Subterranean habitats are well-suited systems to address general problems in community ecology 463 (Mammola, 2019). Foremost, caves are often semi-closed environments extensively replicated 464 across the Earth (Culver, 1970; Culver & Pipan, 2019; Itescu, 2019; Mammola, 2019). Second, 465 466 subterranean communities generally exhibit lower diversity and abundance of organisms than surface ones and are characterized by a bottom-truncated functional diversity (Gibert & 467 468 Deharveng, 2002), allowing us to disentangle the effect of abiotic conditions and biotic 469 interactions in filtering species possessing specific traits within the community (Cardoso, 2012). Third, caves have some conspicuous environmental gradients from the surface towards the 470 471 subsurface (Howarth, 1982; Tobin, Hutchins & Schwartz, 2013; Mammola et al., 2019d), 472 offering a mosaic structure of subterranean microhabitats defined by distinct habitat-filtering 473 properties (Trontelj, Blejec & Fišer, 2012; Mammola et al., 2020). Four questions in community ecology made it to the top-50 list. This result reflects a 474 475 general trend in subterranean biology, where researchers have primarily focused on caves as

model systems for evolutionary studies (Juan *et al.*, 2010), and secondarily used caves as
convenient settings to address fundamental ecological questions (Mammola, 2019). Yet, these
four questions fell within general and timely areas of current ecological research (see Sutherland *et al.*, 2013).

480 The top-ranked question underscored the importance of services provided to humans by 481 subterranean species and ecosystems (Q22), rather than on theoretical aspects of community ecology. Examples of ecosystem services provided by subterranean ecosystems include 482 pollination, seed dispersal, and agricultural pest control by bats (Kunz et al., 2011; Medellin, 483 Wiederholt & Lopez-Hoffman, 2017), provision of clean water (Griebler & Avramov, 2015), 484 serving as a source for new pharmaceutical products (Cheeptham et al., 2013), and even cheese 485 production (Ozturkoglu-Budak et al., 2016). While services with direct benefit to humans have 486 received some attention, values provided by subterranean ecosystems extend far beyond direct 487 human needs. In light of emerging conservation issues associated with subterranean ecosystems 488 489 (Mammola et al., 2019b), investigating ecological services and links between above- and belowground diversity in ecosystem functioning is crucial. 490

491 Two questions called for more research into life-history characteristics (e.g. growth rates, 492 age and size at sexual maturity, longevity, and survival rates; Q25) and food-web specificities of subterranean communities (Q23). Interactions among life-history traits determine the fitness of 493 494 each population, while interactions between populations and the environment dictate the 495 distribution of species (Steranrs, 1992). Only a few studies have described life histories of 496 subterranean species, and this is partially explained by the challenges of captive breeding and the technical problems and effort necessary to conduct in situ comprehensive studies (Vonk & 497 498 Nijman, 2006; Voituron et al., 2011; Venarsky, Huryn & Benstead, 2012; Riesch et al., 2016;

Simon *et al.*, 2017). Consequently, the lack of knowledge on cave species traits limits our
understanding of evolutionary and ecological processes occurring in subterranean ecosystems.

501 Energy limitation is considered a primary mechanism influencing both evolutionary and ecological processes in subterranean environments (Venarsky & Huntsman, 2018). However, a 502 503 more nuanced understanding of subterranean food-web dynamics (Q23) will require other 504 research actions, including to (i) understand the spatial and temporal dynamics of energy resources; (ii) compare resource quality with consumers' physiological requirements; and (iii) 505 compare consumption rates with resource availability in subterranean habitats with different 506 environmental conditions (e.g. terrestrial versus aquatic, fresh versus salt water, and detrital 507 508 versus chemolithoautotrophic food webs).

Finally, understanding the role of stochastic events in caves was highlighted as a deficient 509 area in community ecology (Q24). Given that these events are increasing in frequency amid the 510 environmental crisis of the new millennium (Rahmstorf & Coumou, 2011), the study of 511 512 stochastic phenomena has emerged as a central topic in ecology (Scheffer *et al.*, 2001). Recent papers used groundwater crustaceans to elucidate some of the mechanisms by which earthquakes 513 514 affect the composition and structure of biological communities (Galassi et al., 2014; Fattorini et 515 al., 2017; Fattorini, Di Lorenzo & Galassi, 2018; Morimura et al., 2020). Additional studies have focused on the effect of other events, such as heavy precipitation (Calderón-Gutiérrez, Sánchez-516 517 Ortiz & Huato-Soberanis, 2018) and flooding (Pacioglu et al., 2019). Although it may seem 518 counterintuitive to study stochastic environmental shifts in caves, as they have been traditionally 519 perceived as stable ecosystems, these examples show how caves may represent promising model systems for quantifying the impacts of abrupt environmental shifts driving ecosystem evolution 520 521 (Mammola, 2019).

# 522 VII. MACROECOLOGY AND BIOGEOGRAPHY

Q26 – What drives subterranean patterns of phylogenetic and functional diversity? [#21; 75%]
Q27 – Would the use of novel molecular methods (e.g. metabarcoding, environmental DNA)
provide new insights on subterranean biodiversity patterns and affect known patterns? [#27;
72%]
Q28 – What is the species richness pattern of subterranean organisms globally? [#31; 72%]
Q29 – What factors drive the relative importance of speciation, extinction, and dispersal in

shaping subterranean diversity patterns across regions? [#34; 71%]

530 Q30 – Are current subterranean biodiversity patterns best explained by history of colonization of

surface ancestors or by *in situ* speciation and dispersal in subterranean habitats? [#39; 70%]

532 Q31 – How can sampling effort be standardized so that comparisons of species richness are

533 unbiased? [#43; 69%]

534

535 Over the last 20 years, research in subterranean ecology is shifting from local to landscape 536 studies aiming to document and understand biodiversity patterns at regional to global scales (Zagmajster et al., 2019). This transition is not without difficulties, as it requires linking 537 538 biodiversity patterns to eco-evolutionary processes with little to no possibility for manipulative experiments. Six questions in 'Macroecology and biogeography' were identified in the top-50 539 540 list (Fig. 1). These questions mirror the main challenges faced when documenting and 541 understanding broad-scale biodiversity patterns at the surface. The first challenge is assembling 542 the data required to bring out the characteristic features of biodiversity patterns at such broad scales, while ensuring these patterns are not biased by sampling effort (Q28, Q31). Secondly, to 543 544 combine multiple sampling techniques, species identification methods (e.g. morphological and

545 DNA-based identification), and biodiversity metrics (e.g. alpha, beta, and gamma diversity) in a 546 meaningful way to elucidate the many facets of biodiversity patterns (e.g. taxonomic,

phylogenetic, and/or functional diversity; Jarzyna & Jetz, 2016) (Q27, Q26). Lastly, the relative
contributions of different evolutionary processes (Q29) and diversification hypotheses (Q30) in
shaping biodiversity patterns should be fully examined.

550 The publication of global subterranean diversity maps and databases is a recent phenomenon (Culver & Pipan, 2019; Zagmajster et al., 2019). While diversity maps are 551 informative as they portray differences in species richness among regions or countries, we still 552 lack global maps showing species richness for spatial units of equal area [but see Zagmajster, 553 Culver & Sket (2008), Niemiller & Zigler (2013), and Eme et al. (2015) for examples of 554 regional- and continental-scale diversity maps]. Several approaches have been developed to 555 minimize differences in species richness due to sampling bias (Q31). This issue is particularly 556 germane to difficulties in sampling subterranean habitats. For example, sampling protocols were 557 558 typically standardized among sites and completeness of species inventories were assessed using accumulation and rarefaction curves (Zagmajster et al., 2008; Dole-Olivier et al., 2009; Wynne 559 et al., 2018). Also, observed species richness patterns were tested for robustness using species 560 561 richness estimators (Zagmajster et al., 2014), or complemented with species richness predictions modelled from environmental data (Mokany et al., 2019). 562

Beyond accounting for sampling biases, molecular methods are increasingly useful in understanding subterranean biodiversity patterns (Q27). For example, a recent study comparing latitudinal patterns of crustacean species range size obtained from morphology- and DNA-based species delimitation showed that the pattern of increasing median range size at higher latitudes was more evident when delimiting species with DNA (Eme *et al.*, 2018) (Fig. 2). As sequencing 568 becomes increasingly applied to subterranean taxa, environmental DNA sampling and monitoring may be also used to detect these species in areas difficult to access (Gorički et al., 569 570 2017; Niemiller *et al.*, 2018), thus resulting in more accurate maps of their distributions. To our knowledge, patterns of phylogenetic and functional diversity at continental to global scales have 571 not been documented for any subterranean taxon (Q26), despite the growing knowledge of 572 573 phylogenetic relationships and species traits (Morvan et al., 2013; Fernandes, Batalha & Bichuette, 2016; Fišer et al., 2019; Mammola et al., 2020). Documenting these patterns will 574 575 further underscore the relative importance of dispersal, extinction, and different speciation modes in shaping geographic variation of species richness. Given the differences in global 576 diversity patterns between subterranean and surface habitats, comparing the two systems might 577 help further to elucidate the key drivers of diversity. 578

Recent macroecological studies have shown that historical climatic variability, spatial 579 heterogeneity, and energy contribute to species richness patterns of subterranean taxa in Europe. 580 581 However, the contributions of these factors vary regionally and across taxa (Eme *et al.*, 2015; Bregović & Zagmajster, 2016; Bregović, Fišer & Zagmajster, 2019; Mammola et al., 2019a). At 582 a landscape scale, linking environmental factors with speciation, extinction, and dispersal 583 584 dynamics (Q29), as well as diversification processes (Q30), remains challenging and requires the use of phylogenetic methods and a large number of specimens for DNA analysis (Stern et al., 585 586 2017). Yet phylogenetic methods encompass uncertainties that are highly sensitive to sampling 587 bias and the confounding effect of extinction, both obscuring the inference of transitions to 588 subterranean life. To ameliorate this, genes that lose their function soon after the transition should be used (Lefébure et al., 2017) (see also Section V). 589

590

## 591 VIII. CONSERVATION

- 592 Q32 How does climate change affect subterranean-adapted organisms? [#2; 84%]
- 593 Q33 What are the effects of pollution on subterranean-restricted microorganisms, arthropods,
- 594 and vertebrates? [#3; 84%]
- Q34 What is the impact of above-ground disturbance on subterranean environments and their
  fauna? [#5; 82%]
- 597 Q35 How can we evaluate the ecological status of subterranean ecosystems? [#6; 80%]
- 598 Q36 How can we protect subterranean-adapted species from invasive species? [#7; 80%]
- 599 Q37 How can we combine policy, education, research, and management to safeguard
- 600 subterranean biodiversity effectively? [#8; 80%]
- Q38\* What factors determine the size and location of effective protected areas in subterranean
  environments? [#10; 78%]
- $Q39^*$  How can we effectively involve governments and key stakeholders in the conservation of
- 604 caves and other subterranean systems? [#17; 75%]
- 605 Q40 What would be the best monitoring protocols to quantify long-term changes in the
- distribution and abundance of subterranean invertebrates? [#18; 75%]
- 607 Q41 How do we address the lack of knowledge (biodiversity shortfalls) about the biology of
- subterranean species to enhance proper conservation measures? [#25; 73%]
- Q42 Can subterranean-adapted organisms be used as bioindicators of the health of
- 610 subterranean ecosystems? [#45; 69%]
- 611 Q43 How does the use of caves by humans (e.g. tourism, religious, therapeutic, and
- recreational activities) affect subterranean ecosystems? [#48; 68%]
- 613

Ecosystems are experiencing biodiversity loss at an unprecedented rate worldwide (Barnosky et 614 al., 2011; Dirzo et al., 2014; IPBES, 2018; Cardoso et al., 2020). Thus, conservation and 615 616 management of cave biological diversity is of the utmost concern among subterranean biologists (Mammola et al., 2019b). Conservation questions comprised most of the questions (24%) in our 617 top-50 list (Fig. 1). Of these, 10 questions were part of the initial List #1, while two additional 618 619 questions were suggested by survey participants. Three questions (Q32, Q33, and Q36) highlighted three of the greatest threats to biodiversity worldwide – climate change (Ripple *et al.*, 620 2019), pollution (Ripple et al., 2017), and invasive alien species (Pyšek et al., 2020) – whose 621 effects are pervasive also underground (Mammola *et al.*, 2019b). Additional questions were 622 centred on the impacts of above-ground disturbance (Q34) and human activities (Q43) on 623 subterranean habitats. All these threats can be combined and described as 'habitat loss and 624 degradation', which is one of the most important drivers of biodiversity loss globally (IPBES, 625 2018). Subterranean habitat loss and degradation is primarily due to surface activities, such as 626 627 agricultural expansion and intensification, urbanization, and mining activities (Reboleira et al., 2013; Mammola et al., 2019b; Castaño-Sánchez, Hose & Reboleira, 2020). Human activities 628 inside caves may also constitute localized threats, with recreational use and tourism activities 629 630 being of particular concern (Fernandez-Cortes et al., 2011; Faille, Bourdeau & Deharveng, 2015). In certain areas, people are even poaching rare invertebrate species for private collections 631 632 (Simičević, 2017), as in the discussed case of Anophthalmus hitleri Scheibel (Coleoptera: 633 Carabidae) (Berenbaum, 2010). 634

Evaluating, understanding, and mitigating these threats are primarily hampered by our scarce knowledge of subterranean organisms' biology (Q41), especially life-history traits (see Q25 in Section VI). Understanding changes in species' abundance and distribution will be crucial to halting biodiversity loss in subterranean habitats. Studies aimed at identifying
bioindicator species (Q42) to help bolster long-term monitoring programs (Q40) are needed.
Additionally, improved sampling procedures and characterizing cave communities in previously
undocumented areas would both enhance our knowledge of subterranean biodiversity (Mammola *et al.*, 2019*b*) and improve the effectiveness of conservation measures (Q41).

642 Furthermore, it is crucial to adopt innovative approaches to safeguard subterranean biodiversity (Q37), as well as to determine the size and location of effective protected areas 643 (Q38). Standardized systematic sampling techniques have been applied to terrestrial (Wynne et 644 al., 2018, 2019) and aquatic subterranean invertebrate species (Dole-Olivier et al., 2009); to be 645 optimally beneficial to conservation and monitoring, these techniques will need to be further 646 scrutinized across a large breadth of taxa and systems. Recently, a cave vulnerability assessment 647 protocol has been developed for bat cave roosts (Tanalgo, Tabora & Hughes, 2018) and, if 648 refined, would hold promise for use with other subterranean animals. 649

650 Protected areas are the most crucial measure to safeguard specific subterranean habitats and the sensitive animal populations they often support (Q38). Indices have been developed for 651 site selection and conservation prioritization (e.g. Borges et al., 2012; Rabelo, Souza-Silva & 652 653 Ferreira, 2018; Strona et al., 2019; Fattorini et al., 2020) which are often based on complementarity, flexibility, and irreplaceability principles (Michel et al., 2009). Yet, rigorous 654 655 geospatial analysis is still rarely applied when the extents of protected areas are being 656 determined. Further considerations should include managing lands upslope from caves or entire 657 watersheds supporting sensitive subterranean habitats. If a species-level approach is taken for establishing a protected area, it would be reasonable to protect the land at the hydrogeologic unit 658 659 (i.e. watershed or karst/volcanic unit) level – as animals are expected to use mesocaverns or

unconsolidated sediments for dispersal (Howarth, 1983; Malard *et al.*, 2017; Trontelj, 2019).
Importantly, such an approach should be based on the most accurate estimation of the relevant
animal's distributional range.

While effective legislation and/or management plans exist for some subterranean species 663 and some regions of the world, overall management policies for most regions of speleological 664 665 importance are lacking (Q39). Only a few countries have national cave protection laws. For example, the United States Federal Cave Protection Act of 1988 has been used as a tool to 666 manage caves on federally owned lands, while Brazil requires geological and biological 667 assessments of caves and stipulates mitigation of any human activities that may negatively 668 impact cave natural resources. In any case, to be fully operational, such legislative and 669 management tools need to be based on the best available science including a comprehensive 670 knowledge of fauna distribution (Brooks, Da Fonseca & Rodrigues, 2004; Samways et al., 2020) 671 and traits of the species of concern (Chichorro, Juslén & Cardoso, 2019; Fattorini et al., 2020). 672 673 Importantly, management plans will require both financial, governmental, and local community support for their implementation. Unfortunately, most countries lack the capacity or legislation to 674 675 protect and conserve sensitive subterranean resources.

676

#### 677 IX. MICROBIOLOGY AND APPLIED TOPICS

- 678 Q44 What is the role of Bacteria, Archaea, fungi, and viruses in nutrient cycling in
- 679 subterranean systems? [#32; 71%]
- 680 Q45 How adaptable are cave microorganisms to changing environmental conditions (e.g.
- 681 climate change)? [#37; 70%]
- 682 Q46 How do other organisms (humans and other animals), and their activities (e.g. visiting

humans and global climate change) influence cave microbiome diversity patterns? [#38; 70%]

Q47 – How does the range of energy sources and quantity influence the diversity of subterranean
microbiota? [#46; 68%]

686 Q48 – What are the limiting nutrients for subterranean microbiota and how do they affect overall
687 subterranean microbial diversity? [#47; 68%]

Q49 – How do subterranean microorganisms cycle key elements – nitrogen, iron, carbon, sulfur,
and phosphorus? [#49; 67%]

690  $Q50^*$  – What is the role of microorganisms in cave-formation processes (speleogenesis) in

691 subterranean environments? [#50; 67%]

692

Without a doubt, topics such as adaptation, origin and evolution, community dynamics, and 693 biogeographic distribution patterns are similarly important and actively targeted in microbial 694 ecology (Antwis et al., 2017). However, research in macroecology and microbial ecology is 695 696 often conducted separately rather than hand-in-hand. For nearly 200 years, subterranean ecosystems have been studied from a macroscopic perspective. Subterranean microbiological 697 research is a relatively new discipline with most research having been conducted since the 698 699 middle of the last century (Griebler & Lueders, 2009). A modern ecosystem approach to subterranean biota requires consideration across all trophic levels and scales (Hershey & Barton, 700 701 2019), especially since the 1980s, when the first cave ecosystems fully sustained by *in situ* 702 chemosynthetic primary production were discovered (Sarbu, Kane & Kinkle, 1996; Kumaresan 703 *et al.*, 2014).

The seven questions on the top-50 list address general problems that have been frequently
examined for various subterranean ecosystems, such as alluvial aquifers, however, less

706	systematically for cave environments. Three questions focused on the active role of
707	microorganisms in nutrient cycling (Q44, Q49) and how nutrient limitations influence microbial
708	diversity (Q48). Although we know that microbes rule the subsurface in terms of element cycles
709	(Ortiz et al., 2014; Kimble et al., 2018) and constitute the basis of the food web, we still lack
710	detailed information on conversion rates and growth kinetics. In addition, subterranean
711	organisms often persist with limited energy resources. Thus, understanding their specific
712	adaptations would help advance our understanding of adaptive strategies for microorganisms in
713	other ecosystems (e.g. mountain-summit and deep-sea habitats). Additionally, the role of viruses,
714	which only recently has been recognized as 'tremendous' for groundwater ecosystems (Griebler,
715	Malard & Lefébure, 2014), has not been investigated for terrestrial subterranean systems (Q44).
716	Two questions further addressed the resistance and resilience of cave microbial
717	communities to disturbance from changes in environmental conditions (Q45) (Cavicchioli et al.,
718	2019), and the impacts of other organisms (in particular, humans; Moldovan et al., 2020;
719	Martínez et al., 2020) on microbial diversity (Q46). These questions also were related to
720	conservation issues from a microbiological perspective. The adverse impacts of the fungus
721	Pseudogymnoascus destructans that causes white-nose syndrome in North American bats is a
722	prominent example. To date, P. destructans occurs in 38 U.S. states and seven Canadian
723	provinces (see http://www.whitenosesyndrome.org), which raises serious concerns for the
724	conservation of hibernating bat species and the ecosystem services they provide (Kunz et al.,
725	2011; Boyles et al., 2011; Medellin et al., 2017; Mammola et al., 2019b). The fungus is an
726	opportunistic environmental pathogen, which can remain in the subterranean environment and
727	contribute to the cave microbiome even in the absence of its host (Lorch et al., 2013).
728	It has been hypothesized that microbial communities with high diversity and functional

729 redundancy do not select for ecosystems poor in energy and stable in environmental conditions (Griebler & Lueders, 2009). Thus, the introduction of novel species may have a destabilizing 730 effect on a cave's biological equilibrium (Q46). The same is true for the introduction of 731 contaminants, such as organic compounds and nutrients that provide additional energy. We are 732 only beginning to understand whether and how energy-diversity relationships known from 733 734 macroecology apply to complex natural bacterial communities (Q47). In fact, there is a growing body of evidence that diversity-productivity relationships also drive microbial communities 735 (Smith, 2007), but this question has not been examined systematically in subterranean 736 ecosystems yet. 737

Finally, Q50 points to the potential contribution of microorganisms in speleogenetic processes, such as weathering and rock formation *via* inducing precipitation. Specifically, in terms of (inorganic) carbon cycling in face of climate change, the role of microbes in the formation of caves may be of great relevance, and has yet to be fully examined.

742

#### 743 X. CONCLUSIONS

(1) The 50th anniversary of Poulson & White's (1969) article was the perfect time to reflect on 744 745 milestone scientific achievements obtained in the natural laboratories offered by caves, while also delineating the most important research priorities for years to come. We have shown how 746 747 subterranean biology has contributed strongly to general scientific questions via the study of 748 evolutionary and ecological processes along the vertical dimension (i.e. the evolutionary 749 transition from the surface to the subsurface). These accomplishments resonate with the 750 sentiments of Poulson & White (1969) and we anticipate that biologists will continue to unravel 751 the mysteries of subterranean ecosystems and contribute to scientific knowledge more broadly,

insofar as revolutionary advances in approaches and technologies continue to foster and nurturenovel paradigms.

754 (2) There is a significant lack of knowledge concerning eco-evolutionary processes underlying biodiversity patterns along the horizontal gradient (i.e. within subterranean habitats). This is 755 756 largely driven by a paucity of functional ecology studies, the weakness of trait-based approaches 757 (Cardoso, 2012; Fernandes et al., 2016; Fišer et al., 2019; Mammola et al., 2020), and the lack of robust systematic sampling techniques for most taxonomic groups (Wynne et al., 2019). 758 759 Bridging these gaps will significantly influence how we address and prioritize future research on the conservation and ecosystem services of subterranean habitats (e.g. Fattorini *et al.*, 2020), as 760 emphasized by the large number of unresolved questions in conservation biology (representing 761 nearly 25% of the top-50 list). 762

(3) We also invite scientists to redouble their efforts to understand the diversity of subterranean 763 life across all its components, with a special focus on linking macroscopic and microbial ecology 764 765 (Foulquier et al., 2011; Mermillod-Blondin, 2011). This will enable us to achieve a mechanistic understanding of subterranean eco-evolutionary processes and ecosystem function. This 766 767 information will be critical in guiding future policy decisions as human activities and global 768 environmental change increasingly impact and strain the subterranean realm. (4) There is a concern that simple voting exercises such as this one may favour general over 769 770 specific questions. Perhaps as a result of this bias, some of the top-voted questions appear to be 771 broad in scope (e.g. Q1, Q2, and Q32). While these questions were able to capture important 772 general lines of inquiry, specific questions may be more useful for setting applied agendas.

773 Therefore, we invite interested readers to consult Appendix S1, which contains our complete list

of 120 questions.

775 (5) While the 'caves as laboratory' paradigm is an effective way to frame broadly scoped studies, we recognize the top-50 list of questions primarily pertains to unresolved issues within the 776 777 borders of subterranean biology. Yet subterranean habitats offer much more. Deep subterranean habitats are one of the few natural systems defined by highly stable and homogenous climatic 778 conditions tantamount to those maintained in a laboratory (Sánchez-Fernández et al., 2018). 779 780 These systems have an island-like nature (Itescu, 2019), and often support communities characterized by highly specialized organisms interacting in simplified ecological networks 781 (Mammola, 2019). By extension, a robust understanding of these rather simplified settings may 782 enable researchers to disentangle the complexities of more diverse systems (e.g. deep-sea 783 habitats). 784

(6) Ultimately, all these features point at subterranean ecosystems as ideal settings in which to
tackle general questions. We strived to provide examples of how some of our survey questions
may aid in addressing non-cave specific agendas. Our hope is that this horizon scan exercise both
underscores the importance of caves for addressing a range of eco-evolutionary questions, as
well as stimulates researchers to redouble their efforts to address some of these lingering
questions in subterranean biology.

791

# 792 XI. ACKNOWLEDGEMENTS

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## 822 XII. REFERENCES

- 823 ANTWIS, R.E., GRIFFITHS, S.M., HARRISON, X.A., ARANEGA-BOU, P., ARCE, A., BETTRIDGE, A.S.,
- 824 BRAILSFORD, F.L., DE MENEZES, A., DEVAYNES, A., FORBES, K.M., FRY, E.L., GOODHEAD,
- 825 I., HASKELL, E., HEYS, C., JAMES, C., *ET AL*. (2017). Fifty important research questions in
- microbial ecology. *FEMS Microbiology Ecology* **93**, 10.1093/femsec/fix044.
- AUDRA, P. & PALMER, A.N. (2011). The pattern of caves: controls of the epigenic speleogensis. *Géomorphologie* 17, 359–378.
- 829 BARNOSKY, A.D., MATZKE, N., TOMIYA, S., WOGAN, G.O.U., SWARTZ, B., QUENTAL, T.B.,
- 830 MARSHALL, C., MCGUIRE, J.L., LINDSEY, E.L., MAGUIRE, K.C., MERSEY, B. & FERRER,
- E.A. (2011). Has the Earth's sixth mass extinction already arrived? *Nature* **471**, 51–57.
- BARR, T.C. & HOLSINGER, J.R. (1985). Speciation in cave faunas. *Annual Review of Ecology and Systematics* 16, 313–337.
- BERENBAUM, M. (2010). ICE Breakers. American Entomologist 56, 132–185.
- 835 BILANDŽIJA, H., ĆETKOVIĆ, H. & JEFFERY, W.R. (2012). Evolution of albinism in cave
- planthoppers by a convergent defect in the first step of melanin biosynthesis. *Evolution* &
- 837 *Development* **14**, 196–203.
- 838 BILANDŽIJA, H., HOLLIFIELD, B., STECK, M., MENG, G., NG, M., KOCH, A.D., GRAČAN, R.,
- 839 ĆETKOVIĆ, H., PORTER, M.L., RENNER, K.J. & JEFFERY, W. (2020). Phenotypic plasticity as
  840 a mechanism of cave colonization and adaptation. *eLife* 9, e51830.
- 841 BLOUNT, Z.D., LENSKI, R.E. & LOSOS, J.B. (2018). Contingency and determinism in evolution:
- replaying life's tape. *Science* **362**, eaam5979.
- 843 BORGES, P.A.V., CARDOSO, P., AMORIM, I.R., PEREIRA, F., CONSTÂNCIA, J.P., NUNES, J.C.,

- 844 BARCELOS, P., COSTA, P., GABRIEL, R. & DAPKEVICIUS, M. DE L. (2012). Volcanic caves:
- priorities for conserving the Azorean endemic troglobiont species. *International Journal of Speleology* 41, 101–112.
- 847 BOULTON, A.J. (2020). Conservation of groundwaters and their dependent ecosystems:
- 848 integrating molecular taxonomy, systematic reserve planning and cultural values. *Aquatic*
- 849 *Conservation: Marine and Freshwater Ecosystems* **30**, 1–7.
- BOYLES, J.G., CRYAN, P.M., MCCRACKEN, G.F. & KUNZ, T.H. (2011). Economic importance of
  bats in agriculture. *Science* 332, 41–42.
- 852 BREGOVIĆ, P., FIŠER, C. & ZAGMAJSTER, M. (2019). Contribution of rare and common species to
- subterranean species richness patterns. *Ecology and Evolution* **9**, 11606–11618.
- 854 BREGOVIĆ, P. & ZAGMAJSTER, M. (2016). Understanding hotspots within a global hotspot –
- identifying the drivers of regional species richness patterns in terrestrial subterranean
- habitats. *Insect Conservation and Diversity* **9**, 268–281.
- 857 BROOKS, T., DA FONSECA, G.A.B. & RODRIGUES, A. (2004). Species, data, and conservation
- planning. *Conservation Biology* **18**, 1682–1688.
- 859 BUHAY, J.E. & CRANDALL, K.A. (2005). Subterranean phylogeography of freshwater crayfishes
- shows extensive gene flow and surprisingly large population sizes. *Molecular Ecology* 14,
  4259–4273.
- 862 CALDERÓN-GUTIÉRREZ, F., SÁNCHEZ-ORTIZ, C.A. & HUATO-SOBERANIS, L. (2018). Ecological
- patterns in anchialine caves. *PLoS ONE* **13**, e0202909.
- 864 CARDOSO, P. (2012). Diversity and community assembly patterns of epigean vs. troglobiont
- spiders in the Iberian Peninsula. *International Journal of Speleology* **41**, 83–94.
- 866 CARDOSO, P., BARTON, P.S., BIRKHOFER, K., CHICHORRO, F., DEACON, C., FARTMANN, T.,

867	FUKUSHIMA, C.S., GAIGHER, R., HABEL, J.C., HALLMANN, C.A., HILL, M.J., HOCHKIRCH, A.,			
868	KWAK, M.L., MAMMOLA, S., ARI NORIEGA, J., ET AL. (2020). Scientists' warning to			
869	humanity on insect extinctions. Biological Conservation 242, 108426.			
870	CARLINI, D.B. & FONG, D.W. (2017). The transcriptomes of cave and surface populations of			
871	Gammarus minus (Crustacea: Amphipoda) provide evidence for positive selection on cave			
872	downregulated transcripts. PLoS ONE 12, e0186173.			
873	CASTAÑO-SÁNCHEZ, A., HOSE, G.C. & REBOLEIRA, A.S.P.S. (2020). Ecotoxicological effects of			
874	anthropogenic stressors in subterranean organisms: a review. Chemosphere 244, 125422.			
875	CAVICCHIOLI, R., RIPPLE, W.J., TIMMIS, K.N., AZAM, F., BAKKEN, L.R., BAYLIS, M.,			
876	BEHRENFELD, M.J., BOETIUS, A., BOYD, P.W., CLASSEN, A.T., CROWTHER, T.W.,			
877	DANOVARO, R., FOREMAN, C.M., HUISMAN, J., HUTCHINS, D.A., ET AL. (2019). Scientists'			
878	warning to humanity: microorganisms and climate change. Nature Reviews Microbiology			
879	<b>17</b> , 569–586.			
880	CHEEPTHAM, N., SADOWAY, T., RULE, D., WATSON, K., MOOTE, P., SOLIMAN, L.C., AZAD, N.,			
881	DONKOR, K.K. & HORNE, D. (2013). Cure from the cave: volcanic cave actinomycetes and			
882	their potential in drug discovery. International Journal of Speleology 42, 35-47.			
883	CHICHORRO, F., JUSLÉN, A. & CARDOSO, P. (2019). A review of the relation between species			
884	traits and extinction risk. Biological Conservation 237, 220-229.			
885	COPILAȘ-CIOCIANU, D., FIŠER, C., BORZA, P. & PETRUSEK, A. (2018). Is subterranean lifestyle			
886	reversible? Independent and recent large-scale dispersal into surface waters by two species			
887	of the groundwater amphipod genus Niphargus. Molecular Phylogenetics and Evolution			
888	<b>119</b> , 37–43.			
889	COURCHAMP, F. & BRADSHAW, C.J.A. (2018). 100 articles every ecologist should read. Nature			

- 890 *Ecology & Evolution* **2**, 395–401.
- CULVER, D.C. (1970). Analysis of simple cave communities I. Caves as islands. *Evolution* 24,
  463–474.
- 893 CULVER, D.C. & PIPAN, T. (2019). The biology of caves and other subterranean habitats. Oxford
- 894 University Press, USA.
- 895 CURL, L.R. (1964). On the definition of a cave. *National Speleological Society Bullettin* 26, 1–6.
- B96 DARWIN, C. (1859). On the origin of species by means of natural selection, or the preservation of

*favoured races in the struggle of life*. John Murray, London.

- 898 DEHARVENG, L. & BEDOS, A. (2019). Diversity of terrestrial invertebrates in subterranean
- habitats. In *Cave Ecology* (eds T. MOLDOVAN, L. KOVÁČ & S. HALSE), pp. 107–172.
- 900 Springer, Cham, Switzerland.
- 901 DIRZO, R., YOUNG, H.S., GALETTI, M., CEBALLOS, G., ISAAC, N.J.B. & COLLEN, B. (2014).

902 Defaunation in the Anthropocene. *Science* **345**, 401–406.

- 903 DOLE-OLIVIER, M.J., CASTELLARINI, F., COINEAU, N., GALASSI, D.M.P., MARTIN, P., MORI, N.,
- 904 VALDECASAS, A. & GIBERT, J. (2009). Towards an optimal sampling strategy to assess
- groundwater biodiversity: comparison across six European regions. *Freshwater Biology* 54,
  777–796.
- DRUMMOND, A.J., HO, S.Y.W., PHILLIPS, M.J. & RAMBAUT, A. (2006). Relaxed phylogenetics
  and dating with confidence. *PLoS Biology* 4, e88.
- 909 EME, D., ZAGMAJSTER, M., DELIĆ, T., FIŠER, C., FLOT, J.-F., KONECNY-DUPRÉ, L., PÁLSSON, S.,
- 910 STOCH, F., ZAKŠEK, V., DOUADY, C.J. & MALARD, F. (2018). Do cryptic species matter in
- 911 macroecology? Sequencing European groundwater crustaceans yields smaller ranges but
- does not challenge biodiversity determinants. *Ecography* **41**, 424–436.

913	EME, D., ZAGMAJSTER	. M.	. Fišer. C	2	GALASSI. D.	. MARMONIER.	P.,	STOCH. F	F C	CORNU. J.F.

- 914 OBERDORFF, T. & MALARD, F. (2015). Multi-causality and spatial non-stationarity in the
- determinants of groundwater crustacean diversity in Europe. *Ecography* **38**, 531–540.
- 916 FAILLE, A., BOURDEAU, C. & DEHARVENG, L. (2015). Weak impact of tourism activities on
- 917 biodiversity in a subterranean hotspot of endemism and its implications for the conservation
- 918 of cave fauna. *Insect Conservation and Diversity* **8**, 205–215.
- 919 FAILLE, A., RIBERA, I., DEHARVENG, L., BOURDEAU, C., GARNERY, L., QUÉINNEC, E. & DEUVE,
- 920 T. (2010). A molecular phylogeny shows the single origin of the Pyrenean subterranean
- 921 *Trechini* ground beetles (Coleoptera: Carabidae). *Molecular Phylogenetics and Evolution*
- **54**, 97–106.
- 923 FATTORINI, S., LOMBARDO, P., FIASCA, B., DI CIOCCIO, A., DI LORENZO, T. & GALASSI, D.M.P.
- 924 (2017). Earthquake-related changes in species spatial niche overlaps in spring communities.
  925 *Scientific Reports* 7, 443.
- 926 FATTORINI, S., DI LORENZO, T. & GALASSI, D.M.P. (2018). Earthquake impacts on
- 927 microcrustacean communities inhabiting groundwater-fed springs alter species-abundance
- 928 distribution patterns. *Scientific Reports* **8**, 1501.
- 929 FATTORINI, S., FIASCA, B., DI LORENZO, T., DI CICCO, M. & GALASSI, D.M.P. (2020). A new
- protocol for assessing the conservation priority of groundwater-dependent ecosystems.
- 931 *Aquatic Conservation: Marine and Freshwater Ecosystems*, doi:10.1002/aqc.3411
- 932 FERNANDES, C.S., BATALHA, M.A. & BICHUETTE, M.E. (2016). Does the cave environment
- reduce functional diversity? *PLoS ONE* **11**, e0151958.
- 934 FERNANDEZ-CORTES, A., CUEZVA, S., SANCHEZ-MORAL, S., CAÑAVERAS, J.C., PORCA, E.,
- 935 JURADO, V., MARTIN-SANCHEZ, P.M. & SAIZ-JIMENEZ, C. (2011). Detection of human-

- 936 induced environmental disturbances in a show cave. *Environmental Science and Pollution*
- 937 *Research* **18**, 1037–1045.
- FIRE, M. & GUESTRIN, C. (2019). Over-optimization of academic publishing metrics: observing
  Goodhart's Law in action. *GigaScience* 8, giz053.
- 940 FIŠER, C., DELIĆ, T., LUŠTRIK, R., ZAGMAJSTER, M. & ALTERMATT, F. (2019). Niches within a
- 941 niche: ecological differentiation of subterranean amphipods across Europe's interstitial
  942 waters. *Ecography* 42, 1212–1223.
- 943 FLÔRES, D.E.F.L., TOMOTANI, B.M., TACHINARDI, P., ODA, G.A. & VALENTINUZZI, V.S. (2013).
- 944 Modeling natural photic entrainment in a subterranean rodent (*Ctenomys* aff. *knighti*), the
- 945 Tuco-Tuco. *PLoS ONE* **8**, e68243.
- 946 FOULQUIER, A., MALARD, F., MERMILLOD-BLONDIN, F., MONTUELLE, B., DOLÉDEC, S., VOLAT,
- 947 B. & GIBERT, J. (2011). Surface water linkages regulate trophic interactions in a
- groundwater food web. *Ecosystems* **14**, 1339–1353.
- 949 FRIEDRICH, M. (2013). Biological clocks and visual systems in cave-adapted animals at the dawn
- 950 of Speleogenomics. *Integrative and Comparative Biology* **53**, 50–67.
- 951 FUMEY, J., HINAUX, H., NOIROT, C., THERMES, C., RÉTAUX, S. & CASANE, D. (2018). Evidence
- 952 for late Pleistocene origin of *Astyanax mexicanus* cavefish. *BMC Evolutionary Biology* 18,
  953 43.
- 954 GALASSI, D.M.P., LOMBARDO, P., FIASCA, B., DI CIOCCIO, A., DI LORENZO, T., PETITTA, M. &
- DI CARLO, P. (2014). Earthquakes trigger the loss of groundwater biodiversity. *Scientific Reports* 4, 6273.
- GATENBY, R.A., GILLIES, R.J. & BROWN, J.S. (2011). Of cancer and cave fish. *Nature Reviews Cancer* 11, 237.

- GIBERT, J. & DEHARVENG, L. (2002). Subterranean ecosystems: a truncated functional
  biodiversity. *BioScience* 52, 473–481.
- 961 GILLESPIE, R.G., BENNETT, G.M., DE MEESTER, L., FEDER, J.L., FLEISCHER, R.C., HARMON, L.J.,
- 962 HENDRY, A.P., KNOPE, M.L., MALLET, J., MARTIN, C., PARENT, C.E., PATTON, A.H.,
- 963 PFENNIG, K.S., RUBINOFF, D., SCHLUTER, D., *ET AL*. (2020). Comparing adaptive radiations
  964 across space, time, and taxa. *Journal of Heredity* 111, 1–20.
- 965 GORIČKI, Š., STANKOVIC, D., SNOJ, A., KUNTNER, M., JEFFERY, W.R., TRONTELJ, P., PAVIC, M.,
- 966 GRIZELJ, Z., NAPARUS-ALJANCIC, M. & ALJANCIC, G. (2017). Environmental DNA in
- 967 subterranean biology: Range extension and taxonomic implications for *Proteus*. *Scientific*
- 968 *Reports* **7**, 91–93.
- 969 GOULD, S.J. & VRBA, E.S. (1982). Exaptation a missing term in the science of form.
- 970 *Paleobiology* **8**, 4–15.
- GRIEBLER, C. & AVRAMOV, M. (2015). Groundwater ecosystem services: a review. *Freshwater Science* 34, 355–367.
- 973 GRIEBLER, C. & LUEDERS, T. (2009). Microbial biodiversity in groundwater ecosystems.
- 974 *Freshwater Biology* **54**, 649–677.
- 975 GRIEBLER, C., MALARD, F. & LEFÉBURE, T. (2014). Current developments in groundwater
- 976 ecology-from biodiversity to ecosystem function and services. *Current Opinion in*
- 977 *Biotechnology* **27**, 159–167.
- 978 GUZIK, M.T., COOPER, S.J.B., HUMPHREYS, W.F. & AUSTIN, A.D. (2009). Fine-scale
- comparative phylogeography of a sympatric sister species triplet of subterranean diving
- 980 beetles from a single calcrete aquifer in Western Australia. *Molecular Ecology* **18**, 3683–
- 981 3698.

982	HERSHEY, O.S. & BARTON, H.A. (2019). The microbial diversity of caves. In Cave Ecology (eds
983	T. MOLDOVAN, L. KOVÁČ & S. HALSE), pp. 69–90. Springer, Cham, Switzerland.
984	HOWARTH, F.G. (1982). Bioclimatic and geologic factors governing the evolution and
985	distribution of Hawaiian cave insects. Entomologia Generalis 8, 17–26.
986	HOWARTH, F.G. (1983). Ecology of cave arthropods. Annual Review of Entomology 28, 365–389.
987	HOWARTH, F.G., JAMES, S.A., MCDOWELL, W., PRESTON, D.J. & IMADA, C.T. (2007).
988	Identification of roots in lava tube caves using molecular techniques: implications for
989	conservation of cave arthropod faunas. Journal of Insect Conservation 11, 251–261.
990	HYACINTHE, C., ATTIA, J. & RÉTAUX, S. (2019). Evolution of acoustic communication in blind
991	cavefish. Nature Communications 10, 4231.
992	IOANNIDIS, J.P.A. (2005). Why most published research findings are false. PLOS Medicine 2,
993	e124.

994 IPBES (2018). Summary for policymakers of the thematic assessment report on land degradation

and restoration of the Intergovernmental Science-Policy Platform on Biodiversity and

996 Ecosystem Services. *Preliminary guide regarding diverse conceptualization of multiple* 

997 values of nature and its benefits, including biodiversity and ecosystem functions and

998 services (deliverable 3 (d)).

- 999 ITESCU, Y. (2019). Are island-like systems biologically similar to islands? A review of the
  1000 evidence. *Ecography* 42, 1298–1314.
- 1001 JARIĆ, I., CORREIA, R.A., BROOK, B.W., BUETTEL, J.C., COURCHAMP, F., DI MININ, E., FIRTH,
- 1002 J.A., GASTON, K.J., JEPSON, P., KALINKAT, G., LADLE, R., SORIANO-REDONDO, A., SOUZA,
- 1003 A.T. & ROLL, U. (2020). iEcology: harnessing large online resources to generate ecological
- 1004 insights. *Trends in Ecology & Evolution* **35**, 630–639.

- JARZYNA, M.A. & JETZ, W. (2016). Detecting the multiple facets of biodiversity. *Trends in Ecology & Evolution* 31, 527–538.
- JEFFERY, W.R. (2005). Adaptive evolution of eye degeneration in the Mexican blind cavefish. *Journal of Heredity* 96, 185–196.
- JEFFERY, W.R. (2009). Regressive evolution in *Astyanax* cavefish. *Annual Review of Genetics*43, 25–47.
- JESCHKE, J.M., LOKATIS, S., BARTRAM, I. & TOCKNER, K. (2019). Knowledge in the dark:
  scientific challenges and ways forward. *FACETS* 4, 423–441.
- 1013 JINHA, A.E. (2010). Article 50 million: an estimate of the number of scholarly articles in
- 1014 existence. *Learned Publishing* **23**, 258–263.
- 1015 JOHNSON, G.D., IDA, H., SAKAUE, J., SADO, T., ASAHIDA, T. & MIYA, M. (2012). A 'living fossil'
- 1016 eel (Anguilliformes: Protanguillidae, fam. nov.) from an undersea cave in Palau.
- 1017 *Proceedings of the Royal Society B: Biological Sciences* **279**, 934–943.
- 1018 JONES, K.K., COOPER, S.J.B. & SEYMOUR, R.S. (2019). Cutaneous respiration by diving beetles
- 1019 from underground aquifers of Western Australia (Coleoptera: Dytiscidae). Journal of
- 1020 *Experimental Biology* **222**, jeb196659.
- 1021 JUAN, C., GUZIK, M.T., JAUME, D. & COOPER, S.J.B. (2010). Evolution in caves: Darwin's
- 1022 'wrecks of ancient life' in the molecular era. *Molecular Ecology* **19**, 3865–3880.
- 1023 KHODAMI, S., MCARTHUR, J.V., BLANCO-BERCIAL, L. & MARTINEZ ARBIZU, P. (2017).
- Molecular phylogeny and revision of copepod Orders (Crustacea: Copepoda). *Scientific Reports* 7, 9164.
- 1026 KIMBLE, J.C., WINTER, A.S., SPILDE, M.N., SINSABAUGH, R.L. & NORTHUP, D.E. (2018). A
- 1027 potential central role of Thaumarchaeota in N-Cycling in a semi-arid environment, Fort

- 1028 Stanton Cave, Snowy River passage, New Mexico, USA. *FEMS microbiology ecology* 94.
- 1029 KONEC, M., PREVORČNIK, S., SARBU, S.M., VEROVNIK, R. & TRONTELJ, P. (2015). Parallels
- between two geographically and ecologically disparate cave invasions by the same species,
- 1031 *Asellus aquaticus* (Isopoda, Crustacea). *Journal of Evolutionary Biology* **28**, 864–875.
- 1032 KUMARESAN, D., WISCHER, D., STEPHENSON, J., HILLEBRAND-VOICULESCU, A. & MURRELL, J.C.
- 1033 (2014). Microbiology of movile cave-A chemolithoautotrophic ecosystem.
- 1034 *Geomicrobiology Journal* **31**, 186–193.
- 1035 KUNZ, T.H., BRAUN DE TORREZ, E., BAUER, D., LOBOVA, T. & FLEMING, T.H. (2011). Ecosystem
- services provided by bats. *Annals of the New York Academy of Sciences* **1223**, 1–38.
- 1037 LANDHUIS, E. (2016). Scientific literature: information overload. *Nature* 535, 457–458.
- LAURANCE, W.F., USECHE, D.C., LAURANCE, S.G. & BRADSHAW, C.J.A. (2013). Predicting
  publication success for biologists. *BioScience* 63, 817–823.
- 1040 LEFÉBURE, T., MORVAN, C., MALARD, F., FRANÇOIS, C., KONECNY-DUPRÉ, L., GUÉGUEN, L.,
- 1041 WEISS-GAYET, M., SEGUIN-ORLANDO, A., ERMINI, L., SARKISSIAN, C. DER, PIERRE
- 1042 CHARRIER, N., EME, D., MERMILLOD-BLONDIN, F., DURET, L., VIEIRA, C., *ET AL*. (2017).
- 1043 Less effective selection leads to larger genomes. *Genome Research* 27, 1016–1028.
- 1044 LEIJS, R., VAN NES, E.H., WATTS, C.H., COOPER, S.J.B., HUMPHREYS, W.F. & HOGENDOORN, K.
- 1045 (2012). Evolution of blind beetles in isolated aquifers: a test of alternative modes of
- 1046 speciation. *PLoS ONE* **7**, e34260.
- 1047 LEPORE, E., MARCHIORO, A., ISAIA, M., BUEHLER, M.J. & PUGNO, N.M. (2012). Evidence of the
- most stretchable egg sac silk stalk, of the European Spider of the Year *Meta menardi*. *PLoS ONE* 7, e30500.
- 1050 LEYS, R., COOPER, S.J.B., STRECKER, U. & WILKENS, H. (2005). Regressive evolution of an eye

- pigment gene in independently evolved eyeless subterranean diving beetles. *Biology Letters*1, 496–499.
- 1053 LEYS, R., WATTS, C.H.S., COOPER, S.J.B. & HUMPHREYS, W.F. (2003). Evolution of
- 1054 subterranean diving beetles (Coleoptera: Dytiscidae: Hydroporini, Bidessini) in the arid
- 1055 zone of Australia. *Evolution* **57**, 2819–2834.
- 1056 LORCH, J.M., MULLER, L.K., RUSSELL, R.E., O' CONNOR, M., LINDNER, D.L. & BLEHERT,
- 1057 D.S. (2013). Distribution and environmental persistence of the causative agent of White-
- 1058 Nose Syndrome, *Geomyces destructans*, in bat hibernacula of the Eastern United. *Applied*
- and Environmental Microbiology **79**, 1293–1301.
- 1060 LOZANO-FERNANDEZ, J., GIACOMELLI, M., FLEMING, J.F., CHEN, A., VINTHER, J., THOMSEN,
- 1061 P.F., GLENNER, H., PALERO, F., LEGG, D.A., ILIFFE, T.M., PISANI, D. & OLESEN, J. (2019).
- 1062 Pancrustacean evolution illuminated by taxon-rich genomic-scale sata sets with an
- 1063 expanded Remipede sampling. *Genome Biology and Evolution* **11**, 2055–2070.
- 1064 LUKIĆ, M., DELIĆ, T., PAVLEK, M., DEHARVENG, L. & ZAGMAJSTER, M. (2019). Distribution
- 1065 pattern and radiation of the European subterranean genus Verhoeffiella (Collembola,
- 1066 Entomobryidae). *Zoologica Scripta* **49**, 86–100.
- 1067 MALARD, F., CAPDERREY, C., CHURCHEWARD, B., EME, D., KAUFMANN, B., KONECNY-DUPRÉ,
- 1068 L., LÉNA, J.-P., LIÉBAULT, F. & DOUADY, C.J. (2017). Geomorphic influence on
- 1069 intraspecific genetic differentiation and diversity along hyporheic corridors. *Freshwater*
- 1070 *Biology* **62**, 1955–1970.
- MAMMOLA, S. (2019). Finding answers in the dark: caves as models in ecology fifty years after
  Poulson and White. *Ecography* 42, 1331–1351.
- 1073 MAMMOLA, S. (2020). On deepest caves, extreme habitats, and ecological superlatives. Trends in

1074 *Ecology & Evolution* **35**, 469–472.

- 1075 MAMMOLA, S., ARNEDO, M.A., FIŠER, C., CARDOSO, P., DEJANAZ, A.J. & ISAIA, M. (2020).
- 1076 Environmental filtering and convergent evolution determine the ecological specialization of
- subterranean spiders. *Functional Ecology* **34**, 1064–1077.
- 1078 MAMMOLA, S., ARNEDO, M.A., PANTINI, P., PIANO, E., CHIAPPETTA, N. & ISAIA, M. (2018).
- 1079 Ecological speciation in darkness? Spatial niche partitioning in sibling subterranean spiders
- 1080 (Araneae : Linyphiidae : *Troglohyphantes*). *Invertebrate Systematics* **32**, 1069–1082.
- 1081 MAMMOLA, S., CARDOSO, P., ANGYAL, D., BALÁZS, G., BLICK, T., BRUSTEL, H., CARTER, J.,
- 1082 ĆURČIĆ, S., DANFLOUS, S., DÁNYI, L., DÉJEAN, S., DELTSHEV, C., ELVERICI, M.,
- 1083 FERNÁNDEZ, J., GASPARO, F., ET AL. (2019a). Local- versus broad-scale environmental
- 1084 drivers of continental  $\beta$ -diversity patterns in subterranean spider communities across
- 1085 Europe. *Proceedings of the Royal Society B: Biological Sciences* **286**, 20191579.
- 1086 MAMMOLA, S., CARDOSO, P., CULVER, D.C., DEHARVENG, L., FERREIRA, R.L., FIŠER, C.,
- 1087 GALASSI, D.M.P., GRIEBLER, C., HALSE, S., HUMPHREYS, W.F., ISAIA, M., MALARD, F.,
- 1088 MARTINEZ, A., MOLDOVAN, O.T., NIEMILLER, M.L., *ET AL*. (2019b). Scientists' warning on
- the conservation of subterranean ecosystems. *BioScience* **69**, 641–650.
- MAMMOLA, S. & LEROY, B. (2018). Applying species distribution models to caves and other
  subterranean habitats. *Ecography* 41, 1194–1208.
- 1092 MAMMOLA, S., PIANO, E., CARDOSO, P., VERNON, P., DOMÍNGUEZ-VILLAR, D., CULVER, D.C.,
- 1093 PIPAN, T. & ISAIA, M. (2019*c*). Climate change going deep: the effects of global climatic
- alterations on cave ecosystems. *The Anthropocene Review* **6**, 2053019619851594.
- 1095 MAMMOLA, S., PIANO, E., MALARD, F., VERNON, P. & ISAIA, M. (2019d). Extending Janzen's
- 1096 hypothesis to temperate regions: a test using subterranean ecosystems. *Functional Ecology*

**33**, 1638–1650.

- 1098 MARTÍNEZ, A., DI CESARE, A., MARI-MENA, N., GARCÍA-GÓMEZ, G., GARCIA-HERRERO, A.,
- 1099 CORNO, G., FONTANETO, D. & ECKERT, E.M. (2020). Tossed 'good luck' coins as vectors
- 1100 for anthropogenic pollution into aquatic environment. *Environmental Pollution* **259**,
- 1101 113800.

1103

- 1102 MARTÍNEZ, A., KVINDEBJERG, K., ILIFFE, T.M. & WORSAAE, K. (2017). Evolution of cave
- 1104 MCGILL, B.J., CHASE, J.M., HORTAL, J., OVERCAST, I., ROMINGER, A.J., ROSINDELL, J., BORGES,

suspension feeding in Protodrilidae (Annelida). Zoologica Scripta 46, 214–226.

- 1105 P.A. V, Emerson, B.C., Etienne, R.S., Hickerson, M.J., Mahler, D.L., Massol, F.,
- 1106 MCGAUGHRAN, A., NEVES, P., PARENT, C., *ET AL*. (2019). Unifying macroecology and
- 1107 macroevolution to answer fundamental questions about biodiversity. *Global Ecology and*
- 1108 *Biogeography* 28, 1925–1936.
- 1109 MEDELLIN, R.A., WIEDERHOLT, R. & LOPEZ-HOFFMAN, L. (2017). Conservation relevance of bat
- 1110 caves for biodiversity and ecosystem services. *Biological Conservation* **211**, 45–50.
- 1111 MERMILLOD-BLONDIN, F. (2011). The functional significance of bioturbation and biodeposition
- 1112 on biogeochemical processes at the water–sediment interface in freshwater and marine
- 1113 ecosystems. *Journal of the North American Benthological Society* **30**, 770–778.
- 1114 MICHEL, G., MALARD, F., DEHARVENG, L., DI LORENZO, T., SKET, B. & DE BROYER, C. (2009).
- 1115 Reserve selection for conserving groundwater biodiversity. *Freshwater Biology* 54, 861–
  1116 876.
- 1117 MOKANY, K., HARWOOD, T.D., HALSE, S.A. & FERRIER, S. (2019). Riddles in the dark:
- 1118 Assessing diversity patterns for cryptic subterranean fauna of the Pilbara. *Diversity and*
- 1119 *Distributions* **25**, 240–254.

1120	MOLDOVAN, O.T., BERCEA, S., NĂSTASE-BUCUR, R., CONSTANTIN, S., KENESZ, M., MIREA, I.C.,				
1121	PETCULESCU, A., ROBU, M. & ARGHIR, R.A. (2020). Management of water bodies in show				
1122	caves – A microbial approach. Tourism Management 78, 104037.				
1123 1124 1125	MORIMURA, S., ZENG, X., NOBORU, N. & HOSONO, T. (2020). Changes to the microbial				
1126	communities within groundwater in response to a large crustal earthquake in Kumamoto,				
1127	southern Japan. Journal of Hydrology 581, 124341.				
1128	Morvan, C., Malard, F., Paradis, E., Lefébure, T., Konecny-Dupré, L. & Douady, C.J.				
1129	(2013). Timetree of aselloidea reveals species diversification dynamics in groundwater.				
1130	Systematic Biology 62, 512–522.				
1131	NIEMILLER, M.L., FITZPATRICK, B.M. & MILLER, B.T. (2008). Recent divergence with gene flow				
1132	in Tennessee cave salamanders (Plethodontidae: Gyrinophilus) inferred from gene				
1133	genealogies. Molecular Ecology 17, 2258–2275.				
1134	NIEMILLER, M.L., FITZPATRICK, B.M., SHAH, P., SCHMITZ, L. & NEAR, T.J. (2013). Evidence for				
1135	repeated loss of selective constraint in rhodopsin of amblyopsid cavefishes (Teleostei:				
1136	Amblyopsidae). Evolution 67, 732–748.				
1137	NIEMILLER, M.L., PORTER, M.L., KEANY, J., GILBERT, H., FONG, D.W., CULVER, D.C., HOBSON,				
1138	C.S., KENDALL, K.D., DAVIS, M.A. & TAYLOR, S.J. (2018). Evaluation of eDNA for				
1139	groundwater invertebrate detection and monitoring: a case study with endangered				
1140	Stygobromus (Amphipoda: Crangonyctidae). Conservation Genetics Resources 10, 247-				
1141	257.				
1142	NIEMILLER, M.L. & ZIGLER, K.S. (2013). Patterns of cave biodiversity and endemism in the				

- 1143 Appalachians and Interior Plateau of Tennessee, USA. *PLoS ONE* **8**, e64177.
- 1144 NJUNJIĆ, I., PERRARD, A., HENDRIKS, K., SCHILTHUIZEN, M., PERREAU, M., MERCKX, V.,
- 1145 BAYLAC, M. & DEHARVENG, L. (2018). Comprehensive evolutionary analysis of the
- 1146 *Anthroherpon* radiation (Coleoptera, Leiodidae, Leptodirini). *PLoS ONE* **13**, e0198367.
- 1147 NORTHUP, D.E., MELIM, L.A., SPILDE, M.N., HATHAWAY, J.J.M., GARCIA, M.G., MOYA, M.,
- 1148 STONE, F.D., BOSTON, P.J., DAPKEVICIUS, M.L.N.E. & RIQUELME, C. (2011). Lava cave
- microbial communities within mats and secondary mineral deposits: implications for life
  detection on other planets. *Astrobiology* 11, 601–618.
- 1151 ORTIZ, M., LEGATZKI, A., NEILSON, J.W., FRYSLIE, B., NELSON, W.M., WING, R.A., SODERLUND,
- 1152 C.A., PRYOR, B.M. & MAIER, R.M. (2014). Making a living while starving in the dark:
- metagenomic insights into the energy dynamics of a carbonate cave. *ISME Journal* 8, 478–
  491.
- 1155 OZTURKOGLU-BUDAK, S., GURSOY, A., AYKAS, D.P., KOÇAK, C., DÖNMEZ, S., DE VRIES, R.P. &
- 1156 BRON, P.A. (2016). Volatile compound profiling of Turkish Divle Cave cheese during
- production and ripening. *Journal of Dairy Science* **99**, 5120–5131.
- 1158 PACIOGLU, O., IANOVICI, N., FILIMON, M., SINITEAN, A., IACOB, G., BARABAS, H., PAHOMI, A.,
- 1159 ACS, A., MUNTEAN, H. & PÂRVULESCU, L. (2019). The multifaceted effects induced by
- 1160 floods on the macroinvertebrate communities inhabiting a sinking cave stream.
- 1161 International Journal of Speleology 48, 167–177.
- 1162 PATIÑO, J., WHITTAKER, R.J., BORGES, P.A. V, FERNÁNDEZ-PALACIOS, J.M., AH-PENG, C.,
- 1163 ARAÚJO, M.B., ÁVILA, S.P., CARDOSO, P., CORNUAULT, J., DE BOER, E.J., DE NASCIMENTO,
- 1164 L., GIL, A., GONZÁLEZ-CASTRO, A., GRUNER, D.S., HELENO, R., *ET AL*. (2017). A roadmap
- 1165 for island biology: 50 fundamental questions after 50 years of the Theory of Island

- Biogeography. *Journal of Biogeography* **44**, 963–983.
- 1167 PECK, S.B. & FINSTON, T.L. (1993). Galapagos islands troglobites: the questions of tropical
- troglobites, parapatric distributions with eyed-sister-species, and their origin by parapatric
- speciation. *Memoires de Biospeologie* **20**, 19–37.
- 1170 PÉREZ-MORENO, J.L., ILIFFE, T.M. & BRACKEN-GRISSOM, H.D. (2016). Life in the Underworld:
- Anchialine cave biology in the era of speleogenomics. *International Journal of Speleology*49, 149–170.
- 1173 PLAVÉN-SIGRAY, P., MATHESON, G.J., SCHIFFLER, B.C. & THOMPSON, W.H. (2017). The
- 1174 readability of scientific texts is decreasing over time. *eLife* **6**, e27725.
- 1175 PONS, J., JURADO-RIVERA, J.A., JAUME, D., VONK, R., BAUZÀ-RIBOT, M.M. & JUAN, C. (2019).
- The age and diversification of metacrangonyctid subterranean amphipod crustaceans
  revisited. *Molecular Phylogenetics and Evolution* 140, 10659.
- 1178 POPA, R., SMITH, A.R., POPA, R., BOONE, J. & FISK, M. (2011). Olivine-respiring bacteria
- isolated from the rock-ice interface in a lava-tube cave, a mars analog environment.
- 1180 *Astrobiology* **12**, 9–18.
- PORTER, M.L. & SUMNER-ROONEY, L. (2018). Evolution in the dark: unifying our understanding
  of eye loss. *Integrative and Comparative Biology* 58, 367–371.
- 1183 POULSON, T.L. & WHITE, W.B. (1969). The cave environment. *Science* **165**, 971–981.
- 1184 PRENDINI, L., FRANCKE, O.F. & VIGNOLI, V. (2010). Troglomorphism, trichobothriotaxy and
- 1185 typhlochactid phylogeny (Scorpiones, Chactoidea): more evidence that troglobitism is not
- an evolutionary dead-end. *Cladistics* **26**, 117–142.
- 1187 PROTAS, M. & JEFFERY, W.R. (2012). Evolution and development in cave animals: from fish to
- 1188 crustaceans. *Wiley interdisciplinary reviews. Developmental biology* **1**, 823–845.

- 1189 PYŠEK, P., HULME, P.E., SIMBERLOFF, D., BACHER, S., BLACKBURN, T.M., CARLTON, J.T.,
- 1190 DAWSON, W., ESSL, F., FOXCROFT, L.C., GENOVESI, P., JESCHKE, J.M., KÜHN, I., LIEBHOLD,
- 1191 A.M., MANDRAK, N.E., MEYERSON, L.A., *ET AL*. (2020). Scientists' warning on invasive
- alien species. *Biological Reviews*, 10.1111/brv.12627
- 1193 RABELO, L.M., SOUZA-SILVA, M. & FERREIRA, R.L. (2018). Priority caves for biodiversity
- conservation in a key karst area of Brazil: comparing the applicability of cave conservation
  indices. *Biodiversity and Conservation* 27, 2097–2129.
- 1196 RAHMSTORF, S. & COUMOU, D. (2011). Increase of extreme events in a warming world.

1197 *Proceedings of the National Academy of Sciences* **108**, 17905–17909.

- 1198 REBOLEIRA, A.S.P.S., ABRANTES, N., OROMÍ, P. & GONÇALVES, F. (2013). Acute toxicity of
- 1199 copper sulfate and potassium dichromate on stygobiont *Proasellus*: general aspects of
- 1200 groundwater ecotoxicology and future perspectives. *Water, Air, & Soil Pollution* 224, 1550.
- 1201 RIDDLE, M.R., ASPIRAS, A.C., GAUDENZ, K., PEUß, R., SUNG, J.Y., MARTINEAU, B., PEAVEY, M.,
- 1202 BOX, A.C., TABIN, J.A., MCGAUGH, S., BOROWSKY, R., TABIN, C.J. & ROHNER, N. (2018).
- 1203 Insulin resistance in cavefish as an adaptation to a nutrient-limited environment. *Nature*
- **555**, 647.
- 1205 RIESCH, R., REZNICK, D.N., PLATH, M. & SCHLUPP, I. (2016). Sex-specific local life-history

adaptation in surface- and cave-dwelling Atlantic mollies (*Poecilia mexicana*). Scientific

- 1207 *Reports* **6**, 22968.
- 1208 RIPPLE, W.J., WOLF, C., NEWSOME, T.M., BARNARD, P. & MOOMAW, W.R. (2019). World
- scientists' warning of a climate emergency. *BioScience* **70**, 8–12.
- 1210 RIPPLE, W.J., WOLF, C., NEWSOME, T.M., GALETTI, M., ALAMGIR, M., CRIST, E., MAHMOUD,
- 1211 M.I., LAURANCE, W.F. & 15364 SCIENTIST SIGNATORIES FROM 184 COUNTRIES (2017).

- 1212 World scientists' warning to humanity: a second notice. *BioScience* **67**, 1026–1028.
- 1213 SAMWAYS, M.J., BARTON, P.S., BIRKHOFER, K., CHICHORRO, F., DEACON, C., FARTMANN, T.,
- 1214 FUKUSHIMA, C.S., GAIGHER, R., HABEL, J.C., HALLMANN, C.A., HILL, M.J., HOCHKIRCH, A.,
- 1215 KAILA, L., KWAK, M.L., MAES, D., ET AL. (2020). Solutions for humanity on how to
- 1216 conserve insects. *Biological Conservation* **242**, 108427.
- 1217 SÁNCHEZ-FERNÁNDEZ, D., RIZZO, V., BOURDEAU, C., CIESLAK, A., COMAS, J., FAILLE, A.,
- 1218 FRESNEDA, J., LLEOPART, E., MILLÁN, A., MONTES, A., PALLARES, S. & RIBERA, I. (2018).
- 1219 The deep subterranean environment as a potential model system in ecological,
- biogeographical and evolutionary research. *Subterranean Biology* **25**, 1–7.
- 1221 SARBU, S.M., KANE, T.C. & KINKLE, B.K. (1996). A chemoautotrophically based cave
- 1222 ecosystem. *Science* **272**, 1953–1955.
- 1223 SCHEFFER, M., CARPENTER, S., FOLEY, J.A., FOLKE, C. & WALKER, B. (2001). Catastrophic
- 1224 shifts in ecosystems. *Nature* **413**, 591–596.
- 1225 SEDDON, A.W.R., MACKAY, A.W., BAKER, A.G., BIRKS, H.J.B., BREMAN, E., BUCK, C.E., ELLIS,
- 1226 E.C., FROYD, C.A., GILL, J.L., GILLSON, L., JOHNSON, E.A., JONES, V.J., JUGGINS, S.,
- 1227 MACIAS-FAURIA, M., MILLS, K., *ET AL*. (2014). Looking forward through the past:
- identification of 50 priority research questions in palaeoecology. *Journal of Ecology* **102**,
- 1229 256–267.
- SIMIČEVIĆ, V. (2017). Poachers threaten Balkans underground biodiversity. *Science* 358, 1116
   LP 1117.
- 1232 SIMON, V., ELLEBOODE, R., MAHÉ, K., LEGENDRE, L., ORNELAS-GARCIA, P., ESPINASA, L. &
- 1233 RÉTAUX, S. (2017). Comparing growth in surface and cave morphs of the species Astyanax
- *mexicanus*: insights from scales. *EvoDevo* **8**, 23.

- SMITH, V.H. (2007). Microbial diversity-productivity relationships in aquatic ecosystems. *FEMS Microbiology Ecology* 62, 181–186.
- 1237 STERANRS, S. (1992). The Evolution of Life Histories. Oxford University Press, New York.
- 1238 STERN, D.B., BREINHOLT, J., PEDRAZA-LARA, C., LÓPEZ-MEJÍA, M., OWEN, C.L., BRACKEN-
- 1239 GRISSOM, H., FETZNER JR., J.W. & CRANDALL, K.A. (2017). Phylogenetic evidence from
- 1240 freshwater crayfishes that cave adaptation is not an evolutionary dead-end. *Evolution* 71,
  1241 2522–2532.
- 1242 STRONA, G., FATTORINI, S., FIASCA, B., DI LORENZO, T., DI CICCO, M., LORENZETTI, W.,
- 1243 BOCCACCI, F. & GALASSI, D.M.P. (2019). Aqualife software: a new tool for a standardized
- 1244 ecological assessment of groundwater dependent ecosystems. *Water* **11**, 2574.
- 1245 SUTHERLAND, W.J., DIAS, M.P., DICKS, L. V, DORAN, H., ENTWISTLE, A.C., FLEISHMAN, E.,
- 1246 GIBBONS, D.W., HAILS, R., HUGHES, A.C., HUGHES, J., KELMAN, R., LE ROUX, X.,
- 1247 LEANSTEY, B., LICKORISH, F.A., MAGGS, L., *ET AL*. (2020). A horizon scan of emerging
- 1248 global biological conservation issues for 2020. *Trends in Ecology & Evolution* **35**, 81–90.
- 1249 SUTHERLAND, W.J., FLEISHMAN, E., MASCIA, M.B., PRETTY, J. & RUDD, M.A. (2011). Methods
- 1250 for collaboratively identifying research priorities and emerging issues in science and policy.
- 1251 *Methods in Ecology and Evolution* **2**, 238–247.
- 1252 SUTHERLAND, W.J., FRECKLETON, R.P., GODFRAY, H.C.J., BEISSINGER, S.R., BENTON, T.,
- 1253 CAMERON, D.D., CARMEL, Y., COOMES, D.A., COULSON, T., EMMERSON, M.C., HAILS, R.S.,
- 1254 HAYS, G.C., HODGSON, D.J., HUTCHINGS, M.J., JOHNSON, D., ET AL. (2013). Identification of
- 1255 100 fundamental ecological questions. *Journal of Ecology* **101**, 58–67.
- 1256 TANALGO, K.C., TABORA, J.A.G. & HUGHES, A.C. (2018). Bat cave vulnerability index (BCVI):
- 1257 a holistic rapid assessment tool to identify priorities for effective cave conservation in the

- 1258 tropics. *Ecological Indicators* **89**, 852–860.
- 1259 TIERNEY, S.M., LANGILLE, B., HUMPHREYS, W.F., AUSTIN, A.D. & COOPER, S.J.B. (2018).
- 1260 Massive parallel regression: a précis of genetic mechanisms for vision loss in diving beetles.
- 1261 *Integrative and Comparative Biology* **58**, 465–479.
- 1262 TOBIN, B.W., HUTCHINS, B.T. & SCHWARTZ, B.F. (2013). Spatial and temporal changes in
- invertebrate assemblage structure from the entrance to deep-cave zone of a temperate
- 1264 marble cave. *International Journal of Speleology* **42**, 203–214.
- 1265 TORRES-PAZ, J., HYACINTHE, C., PIERRE, C. & RÉTAUX, S. (2018). Towards an integrated
- approach to understand Mexican cavefish evolution. *Biology Letters* **14**, 20180101.
- 1267 TRONTELJ, P. (2019). Structure and genetics of cave populations. In *Cave Ecology* (eds T.
- 1268 MOLDOVAN, L. KOVÁČ & S. HALSE), pp. 269–296. Springer, Cham, Switzerland.
- 1269 TRONTELJ, P., BLEJEC, A. & FIŠER, C. (2012). Ecomorphological convergence of cave
- 1270 communities. *Evolution* **66**, 3852–3865.
- 1271 TRONTELJ, P., BORKO, Š. & DELIĆ, T. (2019). Testing the uniqueness of deep terrestrial life.
  1272 Scientific Reports 9, 15188.
- 1273 VENARSKY, M.P. & HUNTSMAN, B.M. (2018). Food webs in caves. In *Cave Ecology* (eds O.
- 1274 MOLDOVAN, L. KOVÁČ & S. HALSE), pp. 309–328. Springer, Cham, Switzerland.
- 1275 VENARSKY, M.P., HURYN, A.D. & BENSTEAD, J.P. (2012). Re-examining extreme longevity of
- 1276 the cave crayfish *Orconectes australis* using new mark–recapture data: a lesson on the
- 1277 limitations of iterative size-at-age models. *Freshwater Biology* **57**, 1471–1481.
- 1278 VOITURON, Y., DE FRAIPONT, M., ISSARTEL, J., GUILLAUME, O. & CLOBERT, J. (2011). Extreme
- 1279 lifespan of the human fish (*Proteus anguinus*): A challenge for ageing mechanisms. *Biology*
- 1280 *Letters*, 7.

- 1281 VONK, R. & NIJMAN, V. (2006). Sex ratio and sexual selection in wormshrimps (Crustacea,
- 1282 Amphipoda, Ingolfiellidea). *Contributions to Zoology* **75**, 189–194.
- 1283 WAKELING, S., WILLETT, P., CREASER, C., FRY, J., PINFIELD, S. & SPEZI, V. (2016). Open-access
- 1284 mega-journals: a bibliometric profile. *PLoS ONE* **11**, e0165359.
- 1285 WALTHER, G.-R., POST, E., CONVEY, P., MENZEL, A., PARMESAN, C., BEEBEE, T.J.C.,
- FROMENTIN, J.-M., HOEGH-GULDBERG, O. & BAIRLEIN, F. (2002). Ecological responses to
  recent climate change. *Nature* 416, 389–395.
- WILKENS, H. (2020). The role of selection in the evolution of blindness in cave fish. *Biological Journal of the Linnean Society* 130, 421–432.
- WILKENS, H. & STRECKER, U. (2017). *Evolution in the dark: Darwin's loss without selection*.
  Springer, Cham, Switzerland.
- 1292 WYNNE, J.J., BERNARD, E.C., HOWARTH, F.G., SOMMER, S., SOTO-ADAMES, F.N., TAITI, S.,
- 1293 MOCKFORD, E.L., HORROCKS, M., PAKARATI, L. & PAKARATI-HOTUS, V. (2014).
- 1294 Disturbance relicts in a rapidly changing world: the Rapa Nui (Easter Island) factor.
- 1295 *BioScience* **64**, 711–718.
- 1296 WYNNE, J.J., HOWARTH, F.G., SOMMER, S. & DICKSON, B.G. (2019). Fifty years of cave
- arthropod sampling: techniques and best practices. *International Journal of Speleology* 48,
  33–48.
- 1299 WYNNE, J.J., SOMMER, S., HOWARTH, F.G., DICKSON, B.G. & VOYLES, K.D. (2018). Capturing
- arthropod diversity in complex cave systems. *Diversity and Distributions* **24**, 1478–1491.
- 1301 YOSHIZAWA, M., GORIČKI, Š., SOARES, D. & JEFFERY, W.R. (2010). Evolution of a behavioral
- shift mediated by superficial neuromasts helps cavefish find food in darkness. *Current*
- 1303 *Biology* **20**, 1631–1636.

- 1304 YOSHIZAWA, M., SETTLE, A., HERMOSURA, M.C., TUTTLE, L.J., CETRARO, N., PASSOW, C.N. &
- MCGAUGH, S.E. (2018). The evolution of a series of behavioral traits is associated with
  autism-risk genes in cavefish. *BMC Evolutionary Biology* 18, 89.
- 1307 ZAGMAJSTER, M., CULVER, D.C. & SKET, B. (2008). Species richness patterns of obligate
- subterranean beetles (Insecta: Coleoptera) in a global biodiversity hotspot effect of scale
  and sampling intensity. *Diversity and Distributions* 14, 95–105.
- 1310 ZAGMAJSTER, M., EME, D., FIŠER, C., GALASSI, D., MARMONIER, P., STOCH, F., CORNU, J.F. &
- 1311 MALARD, F. (2014). Geographic variation in range size and beta diversity of groundwater
- 1312 crustaceans: Insights from habitats with low thermal seasonality. *Global Ecology and*
- 1313 *Biogeography* **23**, 1135–1145.
- 1314 ZAGMAJSTER, M., MALARD, F., EME, D. & CULVER, D.C. (2019). Subterranean biodiversity
- 1315 patterns from global to regional scales. In *Cave Ecology* (eds T. MOLDOVAN, L. KOVÁČ &
- 1316 S. HALSE) pp. 195–227. Springer, Cham, Switzerland.
- 1317 ZAKŠEK, V., DELIĆ, T., FIŠER, C., JALŽIĆ, B. & TRONTELJ, P. (2019). Emergence of sympatry in a
- radiation of subterranean amphipods. *Journal of Biogeography* **46**, 657–669.
- 1319 ZHANG, J., KAPLI, P., PAVLIDIS, P. & STAMATAKIS, A. (2013). A general species delimitation
- method with applications to phylogenetic placements. *Bioinformatics* **29**, 2869–2876.
- 1321

## 1322 XIII. SUPPORTING INFORMATION

- Additional supporting information may be found online in the Supporting Information section atthe end of the article.
- **Appendix S1.** Questions from List #2 (i.e. 120 questions selected from List #1 during Survey#1)
- and List #3 (i.e. 25 additional questions suggested by Survey #2 participants) ranked based on

1327 the percentage of 'major importance' votes.

- 1329 Table 1. Subject areas, general topics addressed, panel member composition (\*= panel
- 1330 coordinator;  $^{\circ}$ = postdoc or early career researcher), and number of questions included in the top-
- 1331 50 list out of the total retained in List #1. Panel members are listed alphabetically by surname.
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Subject area	General topics	Panel members	Number of questions
Adaptation	Morphological, physiological and behavioural adaptations to the subterranean environment	Žiga Fišer°, Daniel W. Fong, Tanja Pipan*, William R. Jeffery, Jure Jugovic	10 out of 43
Origin and evolution	Cave ontology and past climate change, migration–speciation– extinction dynamics, and speciation and diversification	Steven J.B. Cooper*, Matthew Niemiller, Alejandro Martínez°, Meredith Protas	11 out of 36
Community ecology	Population dynamics, community assembly, biotic interaction, trophic webs, and energy flows	Rodrigo L. Ferreira*, Cene Fišer, Thais G. Pellegrini°, Michael Venarsky°	4 out of 32
Macroecology and biogeography	Global diversity patterns (taxonomic, phylogenetic, functional), biogeography theory, and diversity drivers	Maria E. Bichuette, David Eme°, Florian Malard*, Maja Zagmajster°	6 out of 32
Conservation biology	Climate change, habitat loss, invasive species, conservation and management policies, and show-cave-related issues	Isabel R. Amorim <sup>°</sup> , Paulo A. V. Borges <sup>*</sup> , Louis Deharveng, J. Judson Wynne, Ana Sofia P. S. Reboleira	12 out of 37
Microbiology and applied topics	Microbial communities, industrial and pharmaceutical potential, epidemics, and exobiology	Naowarat Cheeptham, Thomas M. Lilley*, Melissa B. Meierhofer°, Diana E. Northup	7 out of 31
Other topics	Any topic falling outside the scope of the six core subject areas	David C. Culver*, Christian Griebler, Johanna Kowalko, Raoul Manenti°	n/a (merged within the other subject areas)

# 1334 Table 2. Glossary of terms.

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Term	General definition
Cave	A human-accessible subterranean space, either a single chamber or series of chambers, formed within different substrata (Curl, 1964). Note that a cave is just one among the wide variety of subterranean habitats (see definition below).
Exaptation	A trait shaped by selection or neutral evolution co-opted for a new function (Gould & Vrba, 1982).
Speleogenetic process	The process of water dissolving surrounding rock, gradually forming passages that evolve into cave systems (Audra & Palmer, 2011).
Subterranean habitat(s) / ecosystem(s)	The breadth of underground voids of different sizes, either dry or filled with water, sharing two main ecological features: the absence of sunlight and buffered climatic conditions. Examples of subterranean habitats include caves, groundwater, anchialine systems, artificially excavated underground voids, shallow subterranean habitats, as well as deep maze of fissures and pore spaces with size prohibiting human entry (Culver & Pipan, 2019).

#### 1337 FIGURE LEGENDS

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Fig. 1. Survey workflow, summary statistics of survey participants, and the breakdown bysubject area of the 50 highest priority research questions.

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1342 Fig. 2. The relationship between median range size (maximum linear extent) per latitudinal band and latitude for 147 European groundwater species of Niphargidae (Amphipoda) and Aselloidea 1343 (Isopoda) delimited using morphology (A) and a molecular species delimitation method (B). 1344 1345 Molecular delimitation was performed by a Bayesian implementation of the Poisson tree processes (Zhang et al., 2013) approach based on molecular phylogenies inferred from 2883 1346 cytochrome c oxidase subunit I sequences. Black horizontal bars, dots, and boxes show the 1347 median, average, and interquartile range, respectively, for 0.9° latitudinal bands. The maximum 1348 length of each whisker is up to 1.5 times the interquartile range. Trend lines (with 95% 1349 1350 confidence intervals) represent the fit of a gamma generalized linear model to the averages of latitudinal bands and its quadratic (A) and cubic (B) term. Data re-analysed from Eme et al. 1351 (2018). 1352

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