# 1 On the causes of geographically heterogeneous parallel

# 2 evolution in sticklebacks

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# 12 Abstract

The three-spined stickleback (Gasterosteus aculeatus) is an important model system for 13 the study of parallel evolution in the wild, having repeatedly colonized and adapted to 14 freshwater from the sea throughout the northern hemisphere. Previous studies identified 15 numerous genomic regions showing consistent genetic differentiation between freshwater 16 and marine ecotypes, but these had typically limited geographic sampling and mostly 17 focused on the Eastern Pacific region. We analysed population genomic data from the 18 three-spined stickleback marine and freshwater ecotypes covering the entire species' 19 range to detect loci involved in parallel evolution at different geographic scales. Most 20 signatures of parallel evolution were unique to the Eastern Pacific and trans-oceanic 21

marine-freshwater differentiation was restricted to a limited number of shared genomic 22 regions, including three chromosomal inversions. Based on simulations and empirical 23 data, we demonstrate that this could result from the stochastic loss of freshwater-adapted 24 alleles during the invasion of the Atlantic basin and selection against freshwater-adapted 25 variants in the sea, both of which can reduce standing genetic variation available for 26 freshwater adaptation outside the Eastern Pacific region. Moreover, the elevated linkage 27 disequilibrium associated with marine-freshwater differentiation in the Eastern Pacific is 28 consistent with secondary contact between marine and freshwater populations that 29 evolved in isolation from each other during past glacial periods. Thus, contrary to what 30 earlier studies from the Eastern Pacific region have led us to believe, parallel marine-31 freshwater differentiation in sticklebacks is far less prevalent and pronounced in all other 32 parts of the species global distribution range. 33

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Keywords: Gasterosteus aculeatus; genetic differentiation; linkage disequilibrium; local
 adaptation; parallel evolution

# 37 Introduction

The extent to which the evolution of similar phenotypes arises by selection acting on 38 shared ancestral polymorphism (i.e. parallel evolution<sup>1</sup>) or via distinct molecular 39 evolutionary pathways (i.e. convergent evolution<sup>2-4</sup>) is a major question in evolutionary 40 biology. A powerful approach to disentangle these processes is to study the genomic 41 architecture underlying the repeated evolution of similar phenotypes in similar 42 environments. After the retreat of Pleistocene glaciers, marine three-spined sticklebacks 43 (Gasterosteus aculeatus) colonized many newly formed freshwater habitats and adopted 44 similar changes in a number of morphological, physiological, life history and behavioural 45 traits<sup>5-9</sup>. Thus, this species has become one of the most widely used model systems to 46 study the molecular basis of adaptive evolution in vertebrates in the wild<sup>10</sup>. 47

Previous studies of the three-spined stickleback model system have quantified the extent 48 of parallel evolution by identifying genomic regions that are consistently differentiated 49 between marine and freshwater ecotypes sampled across different geographic areas<sup>3,11-18</sup>. 50 The focus has historically been on the Eastern Pacific region<sup>11,13,14,19-21</sup>, but several recent 51 studies have focused on Atlantic populations<sup>12,15-18</sup>. However, only two studies have thus 52 far included samples from a larger (global) geographic range<sup>3,11</sup>. Based on whole genome 53 sequence data from a limited number of individuals from Eastern Pacific and Atlantic 54 populations (n = 21), Jones et al.<sup>11</sup> identified ~200 genomic regions that consistently 55 separated marine and freshwater individuals globally, representing roughly 0.5% of the 56 dataset. They also found that 2.83% of the genome showed signatures of parallel selection 57 in Eastern Pacific freshwater locations – approximately six times more than that at the 58 global scale (tree i, Supplementary Fig. 2 and Supplementary Table 7 in Jones et al.<sup>11</sup> – 59 suggesting that more loci contribute to parallel evolution at smaller geographic (regional) 60

scales. However, since this pattern was unique to the Eastern Pacific (and the focus was 61 on global parallelism) its implications for a holistic understanding of marine-freshwater 62 differentiation at both regional and global scales was never discussed. Such global 63 heterogeneous ecotype divergence is consistent with the results of several other studies 64 as well. Focusing on 26 candidate genes in six pairs of marine-freshwater populations 65 across the globe. DeFaveri et al.<sup>3</sup> found that only ~50% of the genes under divergent 66 selection were shared across more than three population pairs, and none were shared 67 among all populations. This suggested a limited re-reuse of ancestral polymorphism at the 68 global scale, implicating either an important role of convergent evolution at larger 69 geographic scales<sup>3</sup>, or geographic heterogeneity in selective pressure among different<sup>3</sup> 70 freshwater ecosystems<sup>3,4</sup>. Furthermore, studies focusing on parallel evolution within 71 oceans, and even smaller geographic regions, show striking differences in the proportion 72 73 of loci involved in parallel freshwater adaptation between Pacific and Atlantic regions<sup>11-</sup> <sup>13,15-18,21</sup>. For instance, Terekhanova et al.<sup>17,18</sup> recovered only 21 highly localized genomic 74 regions involved in parallel freshwater differentiation in the White Sea, in contrast to 75 Hohenlohe et al.<sup>13</sup> and Nelson & Cresko<sup>21</sup> who found large genomic regions involved in 76 parallel freshwater differentiation across almost all chromosomes in the Eastern Pacific 77 populations. Therefore, the potential mechanisms underlying this apparent large-scale 78 geographic heterogeneity in genome-wide patterns of parallel evolution in three-spined 79 sticklebacks remain unexplored. To this end, we analysed population genomic data from a 80 comprehensive sampling of all major geographic areas inhabited by the three-spined 81 stickleback, and employed unsupervised and supervised methods to detect loci involved in 82 parallel marine-freshwater differentiation at different geographical scales. Based on earlier 83 observations<sup>3,12,15-18,22</sup>, we hypothesize that the genetic parallelism in response to 84 freshwater colonization by marine sticklebacks is heterogeneous at the global scale, and 85

that the degree of genetic parallelism is much stronger in the Eastern Pacific region than
 elsewhere.

We further seek to understand and discuss the ultimate causes of the marked regional 88 differences in genome-wide signatures of parallel genetic differentiation among ecotypes. 89 To explain the mechanism behind the repeated use of the same alleles in independent 90 freshwater populations of sticklebacks, Schluter & Conte<sup>1</sup> proposed the "transporter 91 hypothesis". This hypothesis postulates that three-spined sticklebacks have repeatedly 92 colonized and adapted to freshwater environments via selection on standing genetic 93 variation in large marine populations. These freshwater-adapted alleles are in turn 94 maintained in the marine populations by recurrent gene flow with previously colonized 95 freshwater populations. Three-spined stickleback populations have persisted in the 96 Eastern Pacific for approximately 26 Mya<sup>23-26</sup> and from there recolonized the Western 97 Pacific and Atlantic Ocean basins following local extinctions much more recently, during 98 the late Pleistocene (36.9-346.5 kya<sup>27-29</sup>). During bottlenecks and founder events, rare 99 alleles are lost at a higher rate than common alleles<sup>30,31</sup>. Since freshwater-adapted alleles 100 exist in the marine populations only at low frequencies<sup>1</sup>, it is likely that they were lost to a 101 higher degree than neutral variation during geographic range expansions from the Eastern 102 Pacific (via the sea), thereby reducing the amount of standing genetic variation available 103 for freshwater adaptation outside of the Eastern Pacific. To test this hypothesis, we used 104 individual-based forward simulations designed to mimic the transporter hypothesis, and 105 the general global population demographic history of three-spined sticklebacks outlined 106 above. We conclude with a discussion on other potential biological and demographic 107 explanations for the high degree of geographic heterogeneity in patterns of parallel 108 genomic differentiation, and reflect upon the representativeness of the Eastern Pacific 109 three-spined stickleback populations as a general model for the study of parallel evolution. 110

## 112 **Results**

## 113 Marine-freshwater divergence determined by unsupervised and supervised

114 approaches

The Linkage Disequilibrium Network Analysis (LDna) was applied on a dataset including 115 2,511,922 SNPs derived from 166 individuals worldwide to identify and extract clusters of 116 highly correlated loci, i.e. sets of loci affected by the same evolutionary processes (LD-117 clusters). The first step of LDna identified 214,326 loci that were in high LD with at least 118 one other locus within windows of 100 SNPs (Supplementary Information 1). The next step 119 of performing LDna on each chromosome separately (only using one locus from each LD-120 cluster from step one; Supplementary information 1) resulted in 81 distinct LD-clusters. 121 From these, a final 29 LD-clusters were obtained (pooling within chromosome LD-clusters 122 whenever they were grouped by LDna in the final step; Supplementary information 1), 123 containing a total of 71,064 loci (viz. Cluster 1-29). Eight of these LD-clusters associated 124 with geographic structure and genetic parallelism are highlighted in Fig. 2a-h. Details of all 125 29 clusters can be found in Extended Data Fig. 1 and Extended Data Fig. 7. 126

LDna successfully recovered most of the previously identified regions from Jones et al. (2012) that differentiated marine from freshwater ecotypes, and failed to recover only small regions that had low coverage and relatively low levels of marine-freshwater differentiation (Supplementary Information 2 and Extended Data Fig.2).

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#### 132 Trans-oceanic marine-freshwater parallelism

LD-clusters 5, 6, 10, 11, 12, 13, 16, 18, 20, 22, 25 and 27 (a total of 2,502 loci, 0.100% of 133 the dataset; see four representatives in Fig. 1e-h, and all in Extended Data Fig.1, 2) 134 grouped multiple freshwater individuals from different geographic regions across the 135 Pacific and Atlantic Oceans (for all P < 0.05, permutation test for ecotype differentiation), 136 reflecting genetic marine-freshwater parallelism on a global (trans-oceanic) scale. Within 137 those, LD-clusters 6, 11, 12, 16 and 27 (a total of 1.639 loci, 0.065% of the dataset) 138 similarly showed high marine-freshwater allelic differentiation ( $F_{ST}$ , Fig 1e-h, Fig. 2c-e) in 139 both the Eastern Pacific and Atlantic, further suggesting global parallelism. Particularly, 140 loci from LD-cluster 11 mapped to four distinct regions on Chr. V of which one (Fig. 2c-e) 141 mark the position of the Ectodysplasin (EDA) gene that known to be responsible for 142 marine-freshwater differences in lateral armour plate development worldwide<sup>20</sup>. In contrast, 143 although the remaining clusters (5, 10, 13, 18, 20, 22, 25, a total of 863 loci, 0.034% of the 144 dataset) also grouped freshwater individuals from both the Pacific and Atlantic Oceans 145 (similar to LD-cluster 29 above; Fig. 2c-e and Extended Data Fig. 1), they showed much 146 less marine-freshwater differentiation in the Atlantic Ocean than in the Eastern Pacific (Fig. 147 2c-e). Among the LD-clusters associated with marine-freshwater differentiation, LD-148 clusters 6 and 22 covered previously known chromosomal inversions on Chr. I and XI, 149 respectively<sup>11</sup> (Fig. 1e, Extended Data Fig. 1, Extended Data Fig. 7). In addition, we also 150 found a putative novel inversion on Chr. V (LD-cluster 19,241 loci) that was not associated 151 with marine-freshwater differentiation (Extended Data Fig. 1, Extended Data Fig. 7). LDna 152 and  $F_{ST}$  analyses did not detect any significant region showing marine-freshwater 153 differentiation in the Western Pacific (Extended Data Fig. 3c) and thus, this region is not 154 considered further here. 155

LD-clusters 2 (53,785 loci, 2.141% of the dataset, Fig. 1b) and 21 (183 loci, 0.007% of the 158 dataset, Fig. 1c) separated Eastern Pacific freshwater individuals exclusively from the 159 remaining samples, a pattern that is not expected by chance alone (permutation test P <160 0.001, Extended Data Fig. 1, Extended Data Fig. 7). Rather, this reflects a shared adaptive 161 response among Eastern Pacific freshwater populations. The exception was two 162 freshwater individuals from the Eastern Pacific (ALA; Alaska) that did not group with the 163 other freshwater individuals from the Eastern Pacific but instead with Atlantic (marine and 164 freshwater), Western Pacific (marine and freshwater) and the marine individuals from 165 Eastern Pacific (Fig. 1c, Extended Data Fig. 4). LD-cluster 29 (2,728 loci, 0.109% of the 166 dataset) covering a known inversion on Chr. XXI (Fig. 1d) grouped the Eastern Pacific 167 freshwater individuals (except the two Alaskan individuals above) together with six Atlantic 168 freshwater individuals and one Eastern Pacific marine individual. Because this LD-cluster 169 maps to an inversion, the groups also represent putative inversion karyotypes<sup>22</sup>. Thus, this 170 inversion shows strong ecotype differentiation not only in the Eastern Pacific, but also in a 171 small proportion of individuals outside of the Eastern Pacific that putatively carry the 172 freshwater-adapted karyotype (i.e. the karyotype with the highest frequency among 173 Eastern Pacific freshwater individuals). Notably, no cluster of similar magnitude to LD-174 cluster 2 – which separates freshwater individuals from one specific region from all 175 remaining samples in the data - could be detected outside of the Eastern Pacific, 176 demonstrating that parallel marine-freshwater differentiation in the Eastern Pacific is much 177 more prevalent than anywhere else in the world. 178

A small proportion of the loci from LD-cluster 2 (28 SNPs) mapped to regions that showed global parallelism in Jones et al. <sup>11</sup> (Extended Data Fig. 2). In addition, <1% of all loci in

LD-cluster 2 (243 SNPs) showed  $F_{ST} > 0.2$  also in the Atlantic (as is evident e.g. from Fig. 2a and Extended Data Fig. 3a). These loci appear to be non-randomly distributed in the genome (Extended Data Fig. 3a), indicating that indeed they are likely to be linked to genomic regions involved in marine-freshwater differentiation in both the Atlantic and the Eastern Pacific. Due to small sample-sizes, the  $F_{ST}$  Manhattan plots display a considerable amount of noise, particularly in the datasets from the Eastern and Western Pacific Oceans (Fig. 2b,e and Extended Data Fig. 3).

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## 189 Geographic structure and regional local adaptation

LD-cluster 1 (10,184 loci, 0.405% of the dataset) separated all Pacific individuals (Eastern 190 and Western) from the Atlantic individuals (Fig. 1a), thus mainly reflecting trans-oceanic 191 geographic structure. LD-clusters 4, 8, 9, 14 and 24 (a total of 526 loci, 0.021% of the 192 dataset, Extended Data Fig. 1) separated freshwater individuals from only one geographic 193 region; this likely reflects geographic clustering, but could also contain some loci involved 194 in non-parallel freshwater adaptation. We therefore could not determine the underlying 195 evolutionary phenomena that produced these clusters (Extended Data Fig. 2). Accordingly, 196 loci from these LD-clusters showed little marine-freshwater differentiation in both the 197 Eastern Pacific and Atlantic (Extended Data Fig. 1), and only 2 loci (from LD-cluster 14) 198 mapped to the marine-freshwater divergent regions identified by Jones et al.<sup>11</sup>) regions 199 (Extended Data Fig. 2). 200

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## 202 **Proof of concept simulations**

Several potential explanations for geographic heterogeneity in parallel patterns of marine-203 freshwater differentiation in three-spined sticklebacks have been suggested<sup>3</sup>. One such 204 explanation that has not received much attention in the context of three-spined 205 sticklebacks is the stochastic loss of freshwater-adapted alleles due to founder events 206 when three-spined sticklebacks colonized the rest of the world from the Eastern Pacific in 207 the late Pleistocene (see Introduction). Thus, as a proof of concept, we used forward-in-208 time simulations to investigate the conditions under which parallel islands of differentiation 209 between marine and freshwater ecotypes can arise under such a scenario. 210

In the simulated data, before Atl (the simulated Atlantic Ocean) was colonized from Pac 211 (the simulated Pacific Ocean) prior to the closing of the Bering Strait (38-40 Kya), all five 212 freshwater populations in Pac were fixed or nearly fixed for the freshwater-adapted alleles 213 of all locally adapted QTL (Fig. 3f). Following the colonization of Atl, the increased 214 frequency of the freshwater allele in the Atlantic freshwater populations depended on both 215 QTL density and the level of gene flow between Pac and Atl (Fig. 3f). The highest increase 216 in freshwater-adapted alleles in Atl was observed when QTL density was low (3 QTL per 217 chromosome) and trans-oceanic gene flow was high (5 migrants/generation, Fig. 3f). 218 During post-glacial colonization of new freshwater habitats from the sea (10 Kya to 219 present), freshwater-adapted alleles (in both *Pac* and *Atl*) gradually increased in the newly 220 formed freshwater populations (Fig. 3f), reflecting local adaptation. This increase was 221 similarly dependent on the QTL density (both in Pac and Atl) and trans-oceanic gene flow 222 (only affecting *Atl*, Fig. 3f). These patterns likely reflect the underlying levels of ancestral 223 variation in the sea available for subsequent freshwater adaptation (Supplementary Fig. 224 1a). The lowest frequencies of freshwater-adapted alleles in the sea were always 225 observed when QTL density was the highest (in both Pac and Atl) and trans-oceanic gene 226 flow was the lowest (only affecting the Atl, Supplementary Fig. 1a). Furthermore, the 227

frequency of freshwater-adapted alleles in both the sea (ancestral variation) and in the 228 post-glacial freshwater populations (local adaptation) depended on whether the QTL were 229 located in low or high recombination regions; the lowest frequencies of freshwater-adapted 230 alleles were always observed in low recombination regions (Supplementary Fig. 1b,c). The 231 freshwater-adapted alleles in both Pac and Atl freshwater populations never reached 232 similar frequencies during the post-glacial colonization (10 Kya; Fig. 3f) as before post-233 glacial colonization (>10 Kya; Fig. 3f), showing that ancestral variation in the sea was not 234 sufficient to allow complete local adaptation (i.e. fixation of all original freshwater-adapted 235 alleles) in our simulations. Note that with the rapid fixation of all the freshwater-adapted 236 alleles (that started at frequency 0.5 in the freshwater populations in Pac) and the low 237 mutation rate used (1e<sup>-8</sup> per site and generation), the contribution of *de novo* mutations (at 238 the QTL) to freshwater adaptation in these simulations are negligible. 239

In the simulations, present-day marine-freshwater differentiation (mean neutral  $F_{ST}$ ) was 240 always low for the two chromosomes without QTL as well as in high recombination regions 241 of chromosomes that contained QTL (Fig. 4; Supplementary Fig. 1d). In contrast,  $F_{ST}$  for 242 low recombination regions of QTL-containing chromosomes was high for Pac (for all 243 parameter settings), indicating strong islands of parallel marine-freshwater differentiation. 244 This was also true for Atl when QTL density was low (3 or 6 QTL per chromosome) and 245 when trans-oceanic gene flow was high, but not when QTL density was high (9 QTL per 246 chromosome; Fig. 4; Supplementary Fig. 1d) and trans-oceanic gene flow was low. 247

In the LD-clusters with the strongest Pacific-freshwater (*PF*) versus non-*PF* genetic
differentiation in the simulated data, the clusters separation score (CSSs; the scaled
centroid distance based on the two first principal components [PCs], with range [0,1]) were
always high (>0.75) between *PF* and the Atlantic populations (Atlantic-marine and

freshwater, AF and AM, respectively), similar to LD-cluster 2 (Fig. 3g-h, Supplementary 252 Fig. 1e). However, the CSS between PF and Pacific-marine (PM) for LD-cluster 2 was also 253 high (0.62), in contrast to the simulated data where this score was low (starting from < 0.2) 254 but increased with QTL density, and more so when migration rate during colonization of Atl 255 from *Pac* was high (5 migrants/generation; Fig. 3g-h, Supplementary Fig. 1e). This is likely 256 due to QTL density increasing the distance (in the Principal Component Analyses [PCA]) 257 between *PF* and *PM* and migration rate decreasing the distance between *Pac* and *Atl* 258 individuals, as CSS here is scaled by the maximum Euclidean distance between any two 259 points in the data. Furthermore, when migration rate was low, no LD-cluster showed any 260 significant CSS between AF and AM. However, when migration rate was high and with 261 increasing QTL density, LD-clusters similar to LD-cluster 2 were readily observed also in 262 Atl. This shows that in simulations, low migration rates and high QTL densities are 263 required to produce patterns similar the observed data. 264

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## 266 **Discussion**

Using genome-wide SNP data from a comprehensive global sampling of marine and 267 freshwater stickleback ecotypes, we demonstrate that a much smaller proportion of the 268 genome (0.208% of the dataset) is involved in global parallel marine-freshwater 269 differentiation than exclusively in the Eastern Pacific (2.149% of the dataset). This shows 270 that parallel evolution in the three-spined stickleback is much more pervasive in the 271 Eastern Pacific than anywhere else in the world. Indeed, the LD signal from marine-272 freshwater differentiation in the Eastern Pacific is even stronger than that from geographic 273 structuring between the Pacific and Atlantic Oceans – LD-clusters separating freshwater 274 individuals from the Eastern Pacific comprised five times as many loci than the LD-cluster 275

reflecting geographic structuring between Pacific and Atlantic Oceans. With simulations, 276 we demonstrate that this pattern could partly be explained by the stochastic loss of low 277 frequency freshwater-adapted alleles in the sea during range expansion from the Eastern 278 Pacific. As predicted, the discrepancy between the simulated Pacific and Atlantic 279 populations in both  $F_{ST}$  and CSS analyses was the highest when trans-oceanic gene flow 280 was low (stronger founder event), but this also required the QTL density of locally adapted 281 loci to be high, as this reduced the levels of standing variation of freshwater-adapted 282 alleles in the Atlantic. However, the loss of ancestral variation due to founder effects and 283 the transporter hypothesis is likely not the only explanation for the large discrepancy in 284 patterns of marine-freshwater differentiation between the Eastern Pacific and Atlantic 285 Oceans. In the following, we discuss other alternative biological processes that could 286 potentially contribute to this discrepancy. 287

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## 289 Geographic heterogeneity in standing genetic variation

The "transporter hypothesis"<sup>1</sup> postulates that a low frequency of freshwater-adapted alleles 290 is maintained in the sea via recurrent gene flow between ancestral marine and previously 291 colonized freshwater populations. This standing genetic variation is what selection acts on 292 during the subsequent colonization of freshwater habitats. This implicitly assumes that a 293 large pool of locally adapted alleles has accumulated over a long period of time, as gene 294 flow is expected to spread potentially beneficial mutations across demographically 295 independent populations<sup>32,33</sup>. In support of this hypothesis, it has been shown that 296 haplotypes repeatedly used in freshwater adaptation are identical by descent<sup>20,34</sup> and old – 297 on average, six million years (My), but some are reported to be as old as 15 My<sup>21</sup>. Notably, 298 these studies analysed populations from the Eastern Pacific region, which represents the 299

oldest and most ancestral marine population<sup>27,28</sup> where three-spined sticklebacks are 300 thought to have persisted since the split from their close relative, the nine-spined 301 stickleback (*Pungitius pungitius*), approximately 26 Mya<sup>23-26,35</sup>. However, populations in the 302 Western Pacific and the Atlantic are much younger, as they were colonized from the 303 Eastern Pacific during the late Pleistocene (36.9-346.5 kya<sup>27,28</sup>). Furthermore, there is no 304 evidence for trans-oceanic admixture<sup>27,28</sup> following the split of Pacific and Atlantic clades, 305 and there are no extant populations of three-spined sticklebacks in arctic Russia between 306 the Kara Sea and the Eastern Siberian Sea. Thus, the spread of freshwater-adapted 307 alleles from the Eastern Pacific to elsewhere via migration through the Bering Strait is 308 unlikely, and has probably not occurred in recent times. Our simulations show that 309 following colonization of freshwater populations from the sea, the accessibility of 310 freshwater-adapted alleles - which is a function of colonization history, QTL-density and 311 recombination rate – largely determines the number of loci that show high marine-312 freshwater differentiation. Thus, consistent with previous simulations<sup>34,36</sup>, genomic islands 313 of differentiation in linked neutral loci require several QTL to cluster in low recombination 314 regions (Fig. 4 and Supplementary Fig. 1d). Furthermore, when trans-Atlantic gene flow 315 was low and QTL density was high, we readily observed LD-clusters that showed high 316 marine-freshwater differentiation only in the Eastern Pacific, not in the Atlantic. 317

Our simulations and empirical data suggest that both stochastic loss of genetic diversity and selection against freshwater-adapter variants played a role in reducing the pool of standing genetic variation of freshwater-adapted alleles in the Atlantic region. During range expansions, genetic diversity is expected to decrease with distance from the source population from which the expansion started<sup>37</sup>. This pattern was clear in our simulations as well as in the empirical data, both of which show a statistically significant reduction in heterozygosity in the Atlantic region as compared to the Eastern Pacific region (Fig. 3i,

Supplementary Information 3). These results are consistent with a moderate founder effect 325 following colonisation of the Atlantic basin from the Eastern Pacific (Supplementary 326 Information 3), which could account for the random loss of standing genetic variation of 327 freshwater adapted alleles. Furthermore, since freshwater-adapted alleles are selected 328 against in the sea and thus occur at low frequencies in marine environments, they are 329 even less likely to spread to new geographic regions than neutral alleles<sup>30,31</sup>. Consistent 330 with this, the mean individual heterozygosity for LD-cluster 2 loci was 29 times higher in 331 the Pacific compared to the Atlantic Ocean; a very pronounced difference compared to 332 that in the rest of the genome (Extended Data Fig. 5b). However, the between-ocean 333 differences in marine-freshwater differentiation in the simulated data (Fig. 4) were much 334 less pronounced compared to the empirical data (Fig. 2d). Thus, founder effects and 335 selection are likely not the only factors affecting patterns of marine-freshwater 336 differentiation in the three-spined sticklebacks at the global level. 337

Alternative explanations for the observed discrepancy in patterns of marine-freshwater 338 differentiation between the Eastern Pacific and Atlantic include *i*) stronger spatial genetic 339 structure in marine populations outside of the Eastern Pacific causing heterogeneity in 340 standing genetic variation available for freshwater adaptation, and *ii*) heterogeneity in 341 selective regimes among freshwater habitats, both between Atlantic and Eastern Pacific 342 Oceans and between different geographic areas in the Atlantic. We have further tested 343 these hypotheses but found little or inconclusive support in our data and in other studies 344 (Supplementary Information 3). 345

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#### 347 Secondary contact in the Eastern Pacific

All of the above hypotheses assume that the original source of the ancestral variation in 348 the Eastern Pacific and elsewhere is the same. That is, ancient Eastern Pacific marine 349 populations carried most ancestral variation of freshwater adapted alleles at low 350 frequencies, sourcing the Atlantic region with freshwater adapted alleles which were 351 partially lost either stochastically or due to selection. However, an alternative hypothesis is 352 that modern Eastern Pacific freshwater variants were not present in the marine ancestors 353 but rather in land-locked, ice-lake freshwater populations<sup>38</sup>. As glaciers melted, those 354 populations could have followed meltwater downstream, establishing freshwater 355 populations with a different stock of alleles. Secondary contact with marine sticklebacks 356 during this time might have eroded genetic differentiation across most of the genome, with 357 the exception of those regions involved in freshwater adaptation. In this scenario, the 358 standing genetic variation responsible for Eastern Pacific freshwater adaptation may not 359 have entered the Eastern Pacific marine population until after the end of the last glaciation. 360 Since there is no evidence of gene flow between Atlantic and Pacific populations after the 361 closing of the Bering Strait (60-30 Kya), this ancestral variation could have remained 362 unique to the Pacific Ocean. 363

There is ample evidence for large ice-lakes during the last glacial period (LGP) in North 364 America, with little (if any) connection to the sea <sup>39-42</sup>. Thus, a large part of the genetic 365 variation underlying marine and freshwater adaptation in the Eastern Pacific could in 366 principle have evolved in allopatry i.e. separately among the freshwater ice-lake 367 populations and in the sea (and any other potential freshwater water bodies the sea is in 368 contact with). Consistent with this hypothesis is the strong pattern of long-range LD 369 observed among Eastern Pacific marine individuals<sup>43</sup>, as well as our LDna results which 370 revealed one large cluster separating the Pacific and Atlantic Ocean individuals, and one 371 that specifically separates all Eastern Pacific freshwater individuals from all other 372

individuals. The secondary contact hypothesis is also consistent with close to zero 373 heterozygosity in Atlantic marine individuals observed for LD-cluster 2 loci (Extended Data 374 Fig. 5a), as well as the mismatches marine-freshwater differentiation in the simulated 375 compared to the empirical data. Curiously, we found significant isolation by distance in the 376 Atlantic but not in the Eastern Pacific where overall population structuring was 377 nevertheless higher than in the Atlantic (Extended Data Fig. 5d). This could be consistent 378 with the secondary contact hypothesis, if introgression was stronger in some regions of the 379 Eastern Pacific compared to others. However, further empirical and simulation studies are 380 needed to test the extent to which this secondary contact hypothesis provides a better 381 explanation for the observed data than the transporter hypothesis alone. 382

383

#### 384 Conditions that allow global parallelism

Genomic islands of parallel ecotype divergence were more likely to arise in the simulations 385 when several QTL clustered in the same low recombination region. Surprisingly these 386 were also the QTL where the frequency of the freshwater-adapted allele showed the 387 lowest frequencies in the sea and thus, were least likely to spread to Atlantic during 388 colonisation from Pacific. Since QTL in low recombination regions are less likely to be 389 separated by recombination when freshwater-adapted individuals migrate to the sea, it is 390 reasonable to assume that the selection pressure against these "haplotypes" in the sea is 391 stronger<sup>43</sup>. However, this is not consistent with the empirical data showing that the 392 genomic regions most likely to show global parallel ecotype divergence are inversions, 393 where recombination in heterokaryotypes is particularly restricted. Our simulations assume 394 that freshwater-adapted alleles are selected against in the sea (and the strength of this 395 selection is equal for all QTL) while in reality, selection against some of the "freshwater 396

haplotypes/karyotypes" in the sea may be weak or even absent, allowing them to easily 397 spread during range expansions. Consistent with this reasoning, in PCAs based on loci 398 from LD-clusters corresponding to inversions (LD-clusters 6, 22 and 29) several marine 399 individuals also cluster with the freshwater individuals (Fig. 1d,e, Extended Data Fig. 1), 400 indicating frequent occurrence of the "freshwater karyotypes" in the sea. Indeed, 401 Terekhanova et al.<sup>17</sup> found that the genomic regions most commonly involved in local 402 adaptation in multiple independent freshwater populations were also those with the highest 403 frequencies in the sea. In other words, the most geographically widespread genomic 404 regions involved in freshwater adaptation (sensu the transporter hypothesis) are likely to 405 experience the weakest selection against them in the sea, allowing them to remain at 406 higher frequencies in the sea as standing genetic variation <sup>17</sup>. 407

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#### Are three-spined sticklebacks a representative model to study parallel evolution?

Since the pattern of parallel genetic differentiation between marine and freshwater 410 stickleback ecotypes in the Eastern Pacific is in stark contrast to what is seen across other 411 parts of the species distribution range, it is reasonable to question the generality of the 412 findings from the Eastern Pacific stickleback studies with respect to parallel evolution on 413 broader geographic scales. A recent review of parallel evolution suggests that even 414 dramatic phenotypic parallelism can be generated by a continuum of parallelism at the 415 genetic level <sup>44</sup>. For instance, the coastal ecotypes of *Senecio lautus* exhibit only partial 416 reuse of particular QTL among replicate populations<sup>45</sup>, and genetic redundancy frequently 417 underlies polygenic adaptation in *Drosophila*<sup>46</sup>. Similarly, using  $F_{ST}$  outliers to detect 418 putative genomic targets of selection, Kautt et al.<sup>47</sup> (cichlid fishes), Le Moan et al.<sup>48</sup> 419

420 (anchovy) and Westram et al.<sup>49</sup> (periwinkles) showed that phenotypically very similar 421 populations often share only a small proportion of their  $F_{ST}$  outliers.

One exception that seems more general across taxa is the repeated involvement of 422 chromosomal inversions in parallel evolution. Chromosomal inversions could store 423 standing variation as a balanced polymorphism and distribute it to fuel parallel 424 adaptation<sup>50</sup>. For instance, the same Chr. I inversion involved in global marine-freshwater 425 differentiation in three-spined sticklebacks <sup>11,15,17,18, this study</sup> also differentiates stream and 426 lake ecotypes in the Lake Constance basin in Central Europe<sup>51</sup>. Two other clear examples 427 where most genetic differentiation between ecotypes at larger geographic scales is 428 partitioned into inversions come from monkey flowers (*Mimulus guttatus*<sup>52</sup>) and marine 429 periwinkles (*Littorina saxatilis*<sup>53,54</sup>). 430

While our study focuses exclusively on marine-freshwater ecotype pairs of three-spined 431 sticklebacks, other ecotype pairs within freshwater habitats, such as stream vs. lake and 432 benthic vs. limnetic, also exist. A recent study focusing on stream-lake populations found 433 that putative selected loci showed greater parallelism in the Eastern Pacific (Vancouver 434 Island) than the global scale (North America and Europe<sup>55</sup>), i.e. a similar pattern as 435 reported by our study. Furthermore, Conte et al.<sup>56</sup> studied the extent of QTL reuse in 436 parallel phenotypic divergence of limnetic and benthic three-spined sticklebacks within 437 Paxton and Priest Lakes (British Columbia), and found that although 76% of 42 phenotypic 438 traits diverged in the same direction, only 49% of the underlying QTL evolved in parallel in 439 both lakes. For highly parallel traits in two other pairs of benthic-limnetic sticklebacks, only 440 32% of the underlying QTL were reported to be shared<sup>57</sup>. Thus, these studies are also in 441 stark contrast to the original conclusions of widespread genetic parallelism in three-spined 442 sticklebacks. Notably, the two freshwater individuals from the Eastern Pacific that did not 443

cluster with the remaining freshwater individuals from the Eastern Pacific (and were 444 subsequently removed from the datasets used for  $F_{ST}$  genome scans) were from Alaska. 445 These two individuals are also phylogenetically distinct from other freshwater individuals 446 from the Eastern Pacific <sup>28</sup>. One explanation for this could be that the highly divergent 447 freshwater populations in the Eastern Pacific have a different colonization history than the 448 Alaskan lakes. More specifically, the former could have been colonized from some 449 divergent ice-lake refugia (see above), whereas the latter could have independently been 450 colonized from the sea. 451

452

## 453 **Conclusions**

Our results demonstrate that genetic parallelism in the marine-freshwater three-spined 454 stickleback model system is in fact not as pervasive as some earlier studies focusing on 455 Eastern Pacific populations have led us to believe. Our analysis of geographically more 456 comprehensive data, with similar and less assumption-burdened methods as used in 457 earlier studies, shows that the extraordinary genetic parallelism observed in the Eastern 458 Pacific Ocean is not detectable elsewhere in the world (e.g. Atlantic Ocean, Western 459 Pacific Ocean). Hence, the focus on the Eastern Pacific has generated a perception bias -460 the patterns detected there do not actually apply to the rest of the world. Furthermore, our 461 simulations show that the spread of freshwater-adapted alleles can be hampered if 462 colonization of the Atlantic from the Pacific was limited, particularly for QTL clustered in 463 low recombination regions (i.e. those most likely to result in parallel islands of ecotype 464 differentiation). Therefore, geographic differences in the incidence and pervasiveness of 465 parallel evolution in three-spined sticklebacks likely stem from geographic heterogeneity in 466 access to, and amount of, standing genetic variation, which in turn has been influenced by 467

selection as well as historical population demography. Such historical demographic factors
include founder events as well as the potential accumulation of genetic ecotypic
differences in allopatry during the last glacial maximum, followed by a secondary contact
only after the Atlantic Ocean was colonized via the sea from the Eastern Pacific. Hence,
while striking genome-wide patterns of genetic parallelism exist (e.g. in Eastern Pacific
sticklebacks), the conditions under which such patterns can occur may be far from
common, perhaps even exceptional.

475

## 476 Material and Methods

### 477 Sample collection

We obtained population genomic data from 166 individuals representing both marine and 478 freshwater ecotypes from the Eastern and Western Pacific, as well as from the Eastern 479 and Western Atlantic Oceans (Fig. 1i, Extended Data Fig. 6, Supplementary Table 1). 480 Additional data from previously published studies were retrieved from GenBank. Fish 481 collected for this study were sampled with seine nets, minnow traps and electrofishing. 482 Specimens were preserved in ethanol after being euthanized with an overdose of Tricaine 483 mesylate (MS222). The samples were collected under appropriate national fishing or 484 ethical licenses granted to collectors of the respective countries listed in acknowledgement 485 in Fang et al. (2018). In Finland, the fishing is permitted by land owner according to the 486 Finnish Fishing Law (5§ 27.5.2011/600). The research does not involve animal 487 experiments according to Act of Animal Experimentation (FINLEX 497/2013). 488

To study the extent of genetic parallelism among freshwater sticklebacks with different
 phylogeographic histories, we classified global samples into seven biogeographic regions

<sup>491</sup> based on their phylogenetic affinities: (i) Eastern Pacific, (ii) Western Pacific, (iii) Western
<sup>492</sup> Atlantic, (iv) White and Barents Seas, (v) North Sea and British Isles, (vi) Baltic Sea and
<sup>493</sup> (vii) Norwegian Sea <sup>28</sup> (Fig. 1i). A summary of coordinates, ecotype and population
<sup>494</sup> information on the sampled individuals and re-acquired samples is given in the
<sup>495</sup> Supplementary Table 1.

496

## 497 Sequencing and genotype likelihood estimation

Restriction site associated DNA sequencing (RADseq) using the enzyme *Pstl* was 498 performed for the 62 individuals sampled in this study, using the same protocol as in Fang 499 et al. (2018), where DNA library preparation and sequencing method are described in 500 detail. The raw RAD sequencing data has been uploaded to GenBank. Previously 501 published RADseg and whole genome sequencing (WGS) data for an additional 104 502 individuals from 62 populations were retrieved from GenBank (Supplementary Table 1). All 503 RADseg and WGS datasets were mapped to the three-spined stickleback reference 504 genome (release-92, retrieved from Ensembl<sup>58</sup> using BWA mem v0.7.17<sup>59</sup>. PCR duplicates 505 were removed using the program Stacks v2.5<sup>60</sup> for pair-end RAD data, and SAMtools v1.9 506 (function "markdup"<sup>61</sup>) for whole genome data. Given the heterogeneity in sequencing 507 depth among different datasets, and particularly the very low coverage of the data 508 retrieved from Jones et al.<sup>11</sup>, most of the analyses were performed directly using genotype 509 likelihoods, avoiding variant calling whenever possible. Genotype likelihoods where 510 estimated from the mapped reads using the model of SAMtools<sup>61</sup> as implemented in the 511 program suite ANGSD v0.929<sup>62</sup>. Full scripts for the genotype likelihood estimation and 512 filtering parameters are publically available through DRYAD. Bases with a q-score below 513 20 (-minQ 20) and reads with mapping quality below 25 (-minMapQ 25) were removed, 514

and variants were only retained if they had a p-value smaller than 1e<sup>-6</sup> (-SNP\_pval 1e<sup>-6</sup> flag
in ANGSD). We retained sites with a minimum read depth of two (-minIndDepth 2) in at
least 80% of the sampled individuals (-minInd 133). The sex chromosome (Chr. XIX<sup>63,64</sup>)
was excluded from downstream analyses due to sex-specific genomic heterogeneity<sup>65,66</sup>.
The raw output of genotype likelihoods from all 166 individuals comprised 2,511,922
genome-wide loci.

521

## 522 Unsupervised approach to determine marine-freshwater differentiation

LDna uses a pairwise matrix of LD values, estimated by  $r^2$ , to produce a single linkage 523 clustering tree. The hierarchical clustering algorithm uses the LD matrix to combine two 524 clusters connected to each other by at least one edge. In the resulting tree, the nodes 525 represent clusters of loci connected by LD values above thresholds, where the threshold 526 value is proportional to the distance from the root<sup>22</sup>. As the LD threshold is sequentially 527 lowered, an increasing number of loci will be connected to each other in a fashion that 528 reflects their similarity in phylogenetic signals. For each cluster merger (with decreasing 529 LD threshold), the change in median LD between all pairwise loci in a cluster before and 530 after the merger is estimated as  $\lambda$ . When two highly interconnected clusters merge,  $\lambda$  will 531 be large (unlike when only a single locus is added to an existing cluster), signifying that 532 these two clusters bear distinct phylogenetic signals. LDna is currently limited to ~20,000 533 SNPs at a time due to its dependence on LD estimates for all pairwise comparisons 534 between loci in the dataset. To analyse the whole dataset, we applied a novel three-step 535 LDna approach to reduce the complexity of the data in a nested fashion. First, we started 536 with non-overlapping windows within each chromosome<sup>67</sup>, then performed the analysis on 537 each chromosome individually<sup>22</sup>, and finally on the whole dataset (Supplementary 538

Information 1). In all steps of LDna, we estimated LD between loci from genotype
likelihoods using the program ngsLD<sup>68</sup>, setting the minimum SNP minor allele frequency at
0.05. Full scripts for the LDna analyses are provided in DRYAD. In the first step, we only
kept loci that were in high LD with at least one locus (*r*<sup>2</sup>>0.8) within a window of 100 SNPs,
as most SNPs in the data were not correlated with any other adjacent loci (so-called
singleton clusters<sup>67</sup>), and thus, are unlikely to be informative in the LDna analyses
(Supplementary Information 1).

The main evolutionary phenomena that cause elevated LD between large sets of loci in 546 population genomic datasets are polymorphic inversions, population structure and local 547 adaptation, all of which are expected to be present in our dataset<sup>22</sup>. There are specific and 548 distinct predictions about the population genetic signal and the distribution of loci in the 549 genome that arise from these evolutionary phenomena<sup>22</sup>. First, clusters with LD signals 550 caused by polymorphic inversions are expected to predominantly map to the specific 551 genomic region where the inversions are situated. In addition, PCAs of these loci are 552 expected to separate individuals based on karyotypes. In general, the heterokaryotype is 553 expected to be intermediate to the two alternative homokaryotypes (provided that all 554 karyotypes exist in the dataset), and the heterokaryotypic individuals are expected to show 555 higher observed heterozygosity than the homokaryotypes. However, this is not always so 556 clear, for instance when the inversion is new and mutational differences have not yet 557 accumulated. Second, a PCA based on loci whose frequencies are shaped by genetic drift 558 is expected to separate individuals on the basis of geographic location, with no (or very 559 little) separation between marine and freshwater ecotypes. Third, an LD signal caused by 560 local adaptation (globally) is expected to cluster individuals based on ecotype, regardless 561 of geographic location, with both the locus distribution and LD patterns to some extent 562 being negatively correlated with local recombination rate<sup>34,69</sup>. The reason for this 563

correlation is that gene flow between ecotypes erodes genetic differentiation in sites linked 564 to locally adapted loci with the exception of regions where recombination is restricted (for 565 instance in inversions, or close to centromeres or telomeres). No such pattern is expected 566 for LD caused by population structuring, as the main source of this LD is the random 567 genetic drift that, in the absence of gene flow, generates LD in a fashion that is 568 independent of genome position (Kemppainen et al. 2015) and background selection is 569 also not expected to result in strong patterns of within-species genetic differentiation, 570 particularly when there is at least some level of gene-flow<sup>70,71</sup>. If a set of loci contributes to 571 local adaptation exclusively in a particular geographic area, a PCA based on these loci will 572 only separate individuals based on ecotype in that region. We considered loci to be 573 involved in parallel evolution only if they grouped individuals of the same ecotype from 574 more than one independent location. Otherwise, it is not possible to discern drift from local 575 adaptation, particularly if N<sub>e</sub> is small (i.e. genetic drift is strong). To determine if an LD-576 cluster was likely associated with parallel freshwater differentiation, we first used 577 expectation maximisation and hierarchical clustering methods to identify clusters of 578 individuals in PCAs that contained a minimum of seven individuals, of which at least 85% 579 are freshwater ecotypes (the "in-group"; dotted line; Fig. 1a-h, Extended Data Fig. 1). With 580 less than seven in-group individuals, there was no power to detect significant associations, 581 even if all individuals were freshwater ecotypes. Second, if such in-groups were detected, 582 we used permutations to further test whether this cluster contained more freshwater 583 individuals than expected by chance (Supplementary Information 4). We benchmarked 584 LDna by quantifying the proportion of regions previously identified by Jones et al.<sup>11</sup> as 585 involved in marine-freshwater ecotype differentiation (globally and within the Eastern 586 Pacific) that were correctly recovered by LD-cluster loci (Supplementary Information 2). 587 Note that freshwater individuals from locations without marine individuals were also 588

important for the analyses, as they can inform us about the geographic scale of parallel
 marine-freshwater differentiation (for the LD-clusters where marine-freshwater
 differentiation also involved geographic regions where marine samples were available).

592

#### 593 Supervised approaches to determine marine-freshwater differentiation

Genome-wide allelic differentiation ( $F_{ST}$  estimated from genotype likelihoods in ANGSD) 594 between marine and freshwater ecotypes was estimated separately for the three major 595 oceans in our study: Eastern Pacific, Western Pacific and Atlantic Oceans. All available 596 samples were always used, but due to the small number of available Pacific marine 597 individuals, marine individuals from the Eastern (n=4) and Western Pacific (n=13) were 598 pooled and treated as a combined Pacific marine group in Eastern and Western Pacific 599 ecotype comparisons (Extended Data Fig. 8) in order to reduce the noise of non-window 600 based (single SNP) analyses. To determine whether pooling of Eastern and Western 601 Pacific marine individuals could bias F<sub>ST</sub> estimates, we first estimated F<sub>ST</sub> in 100 kb 602 windows for Eastern Pacific marine vs. Eastern Pacific freshwater and Western Pacific 603 marine vs. Eastern Pacific freshwater individuals, as using large windows allowed us to 604 obtain precise estimates even when the marine group comprised of only four marine 605 individuals from the Eastern Pacific. The two sets of window-based pairwise  $F_{ST}$  estimates 606 were highly correlated (r = 0.904; P<0.001; Extended Data Fig. 3d-g), suggesting that 607 pooling marine individuals from Eastern and Western Pacific should not strongly affect 608 SNP based estimates. Note that from the results of the unsupervised LDna, two Eastern 609 Pacific freshwater individuals from Kodiak Island, Alaska (ALA population) never grouped 610 with the other Eastern Pacific freshwater individuals. Therefore, in agreement with earlier 611 phylogenetic analyses<sup>28</sup>, these two individuals were excluded from the supervised 612

analyses. The squared correlation coefficient of  $F_{ST}$  before and after this exclusion was 0.88, indicating that this did not affect the results.

In each comparison, the sites were firstly filtered from raw mapped reads, retaining sites 615 with less than 25% missing data with quality control (-minIndDepth 1, -uniqueOnly 1, -616 remove\_bads 1, -minMapQ 20, -minQ 20). We retained only variable sites (-SNP\_pval 1e<sup>-</sup> 617 <sup>6</sup>) in each region, resulting in 1,218,858 SNPs in the Eastern Pacific, 1,072,257 SNPs in 618 the Western Pacific, and 1,681,923 SNPs in the Atlantic Ocean. We then obtained 619 genotype likelihoods and site allele frequency likelihoods of the variants (-GL 1, -doSaf 1). 620 Based on these likelihoods, we estimated the two-dimensional site-frequency spectrum 621 (SFS) for each pair of ecotypes (realSFS) and calculated the pairwise weighted  $F_{ST}$ 622 (realSFS fst). 623

624

## 625 **Proof of concept using simulated data**

The simulations were performed with quantiNemo<sup>72</sup>. and aimed to recreate the transporter 626 hypothesis model in the Eastern Pacific (referred to as "Pac" in the context of simulations), 627 to simulate the colonization of the Atlantic (referred to as "Atl" in the context of simulations) 628 from Pac 60-30 Kya during the last known opening of the Bering Strait<sup>27,73,74</sup> and the 629 subsequent post-glacial (10 Kya) colonization of newly formed freshwater habitats in both 630 oceans (simulation details can be found in Supplementary Information 4). In short, 631 simulations begin with one marine population in Pac connected to five independent 632 freshwater populations by symmetrical gene flow (i.e. no gene flow exists between any of 633 the freshwater populations; Fig. 3a) for 10k generations (40-50 kya). This is followed by 634 colonisation of Atl from Pac (with Atl having identical population structure to Pac) by 635

allowing one or five migrants per generation between the oceans for 2 Ky (38-40 Kya), 636 after which no further gene flow is possible. The retreat of the Pleistocene continental ice 637 sheets (at 10 Kya; Fig. 3d) and the colonization of newly formed freshwater habitats is 638 simulated by removing four of the freshwater populations, immediately followed by the 639 emergence of four new (post-glacial) freshwater populations (in both Pac and Atl; Fig. 3e). 640 The fifth freshwater population remains as a "glacial refugia" that continues to feed 641 freshwater-adapted alleles to the sea as standing genetic variation. Post-glacial local 642 adaptation is thus only possible due to the spread of freshwater-adapted alleles from the 643 sea in accordance with the transporter hypothesis<sup>1</sup> (Fig. 3a-e). 644

Marine-freshwater differentiation was based on bi-allelic QTL with allelic effects of either 645 zero or ten, with the selection optima in the marine habitat being zero and the selection 646 optima in all freshwater populations being 20. Thus, a freshwater individual homozygous 647 for allele 2 for a given QTL meant that the individual was at its optimal phenotype, and vice 648 versa for marine individuals. Selection intensities were such that a sufficient amount of 649 standing genetic variation was allowed in the sea and rapid local adaptation in freshwater 650 was possible (see Supporting Information 4 for details). In simulations, all allele 651 frequencies started from 0.5 in all populations (including the QTL in the freshwater 652 habitats). The simulated genome was comprised of ten equally sized chromosomes, with a 653 total genome size of 1000 bps. Regions of both low (centromeric regions) and high 654 (chromosome arms) recombination were represented (Supplementary Information 4). 655 Either 3, 6 or 9 QTL per chromosome were randomly placed in eight of the chromosomes, 656 after which the positions were fixed, leaving the last two chromosomes without any QTL. 657 Twenty replicate simulations were run for each of the six different parameter settings (two 658 levels of trans-oceanic gene flow rates and three different QTL densities). The frequency 659 of freshwater-adapted alleles was recorded at 50-generation intervals throughout the 660

simulations. Population genomic data were saved at the end of the simulations(representing present-day sampling).

663

#### 664 Linking empirical data to simulated data

LDna identified one major cluster (LD-cluster 2; see Results) that separated all Eastern 665 Pacific freshwater individuals from the remaining individuals (Atlantic, Western Pacific and 666 marine samples from the Eastern Pacific, pooled). From the simulated data, we first sub-667 sampled individuals from Pac and Atl to match the samples size of the empirical data 668 (excluding the Western Pacific samples, as this ocean was not included in the 669 simulations), and used LDna to detect clusters similar to LD-cluster 2 using Cluster 670 Separation Scores (CSS; custom R-scripts available from DRYAD). Cluster Separation 671 Scores were calculated as the Euclidean centroid distance in a PCA (based on 672 coordinates from the two first principal components scaled by their eigenvalues) between 673 two groups of individuals, standardized by the longest distance between any two 674 individuals in the PCA (CSS thus ranges between [0,1]). PCA of the simulated datasets 675 were performed by the function snpgdsPCA from the R-package SNPRelate<sup>75</sup>. CSS 676 scores are known to correlate with  $F_{ST}$ , but give higher resolution when genetic 677 differentiation is high, and are less sensitive to small sample sizes<sup>11</sup>. In LDna, we are only 678 interested in clusters with high  $\lambda$ -values (see above and Supplementary Information 1). 679 Therefore, from the ten LD-clusters with the highest  $\lambda$ -values (from each simulated 680 dataset), we considered the cluster with the highest CSS between PF (Eastern Pacific 681 freshwater) and non-PF individuals to be the strongest candidates for showing high 682 ecotype differentiation specifically in Pac, and thus, the most similar to LD-cluster 2 in the 683 empirical data. The non-PF individuals were comprised of PM (Eastern Pacific marine), AF 684

(Atlantic freshwater) and AM (Atlantic marine) individuals pooled. To further assess how 685 similar the patterns of population differentiation (in PCAs) were in the above LD-clusters 686 (simulated data) and empirically obtained LD-cluster 2, we compared the CSS's for all 687 pairwise comparisons between PF and the other three groups of individuals (i.e. "PF vs. 688 *PM*', "*PF* vs. *AF*' and "*PF* vs. *AM*') in the simulated and empirical datasets. To further 689 assess the extent to which clusters similar to LD-cluster 2 could be produced in the Atl, we 690 used the same procedure as above to look for the LD-clusters with the highest CSS scores 691 between AF and non-AF individuals (AM, PF and PM), and calculated CSS scores 692 between AF individuals and the three non-AF groups. 693

694

# 695 Data availability

The new RAD sequencing data have been uploaded to the GenBank under accession numbers SAMN14078677-SAMN14078738. Previously published sequencing data are retrieved from studies specified in Supplementary Table 1.

699

# 700 Code availability

- 701 The scripts used for analysing empirical data (genotype likelihood estimation, filtering,
- LDna) and simulated data are available in DRYAD repository:
- 703 https://doi.org/10.5061/dryad.b2rbnzsb1.

# 705 Author contributions

PK and JM conceive the concept of the study, with contributions from PM and BF. BF and
 PK carried out analyses with significant contributions from PM. PK and BF lead the writing,
 with significant contributions from PM and JM. XF contributed to lift-over analysis. BF

visualised the data. All authors accepted the final version.

710

# 711 Competing interests

The authors declare no competing interests.

713

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729		References
730 731 732	1	Schluter, D. & Conte, G. L. Genetics and ecological speciation. <i>Proc. Natl. Acad. Sci. USA</i> <b>106</b> , 9955-9962, doi:10.1073/pnas.0901264106 (2009).
733 734	2	Arendt, J. & Reznick, D. Convergence and parallelism reconsidered: what have we learned about the genetics of adaptation? <i>Trends Ecol. Evol.</i> <b>23</b> , 26-32 (2008).
735 736 737	3	DeFaveri, J., Shikano, T., Shimada, Y., Goto, A. & Merila, J. Global analysis of genes involved in freshwater adaptation in threespine sticklebacks (Gasterosteus aculeatus). <i>Evolution</i> <b>65</b> , 1800-1807, doi:10.1111/j.1558-5646.2011.01247.x (2011).
738 739	4	Stern, D. L. The genetic causes of convergent evolution. <i>Nat. Rev. Genet.</i> <b>14</b> , 751-764, doi:10.1038/nrg3483 (2013).
740 741	5	Bell, M. A. & Foster, S. A. <i>The evolutionary biology of the threespine stickleback</i> . (Oxford University Press, 1994).
742 743	6	Gibson, G. The synthesis and evolution of a supermodel. <i>Science</i> <b>307</b> , 1890-1891 (2005).
744 745	7	Hendry, A. P., Peichel, C. L., Matthews, B., Boughman, J. W. & Nosil, P. Stickleback research: the now and the next. <i>Evol. Ecol. Res.</i> <b>15</b> , 111-141 (2013).
746 747	8	Lescak, E. A. <i>et al.</i> Evolution of stickleback in 50 years on earthquake-uplifted islands. <i>Proc. Natl. Acad. Sci. USA</i> <b>112</b> , E7204-7212, doi:10.1073/pnas.1512020112 (2015).
748 749	9	Östlund-Nilsson, S., Mayer, I. & Huntingford, F. A. <i>Biology of the three-spined stickleback</i> . (CRC Press, 2006).
750 751 752	10	McKinnon, J. S. & Rundle, H. D. Speciation in nature: the threespine stickleback model systems. <i>Trends Ecol. Evol.</i> <b>17</b> , 480-488, doi:doi 10.1016/S0169-5347(02)02579-X (2002).
753 754	11	Jones, F. C. <i>et al.</i> The genomic basis of adaptive evolution in threespine sticklebacks. <i>Nature</i> <b>484</b> , 55-61, doi:10.1038/nature10944 (2012).
755 756 757	12	Ferchaud, A. L. & Hansen, M. M. The impact of selection, gene flow and demographic history on heterogeneous genomic divergence: three-spine sticklebacks in divergent environments. <i>Mol. Ecol.</i> <b>25</b> , 238-259, doi:10.1111/mec.13399 (2016).

- Hohenlohe, P. A. *et al.* Population genomics of parallel adaptation in threespine
   stickleback using sequenced RAD tags. *PLoS Genet.* 6, e1000862,
   doi:10.1371/journal.pgen.1000862 (2010).
- 14 Hohenlohe, P. A. & Magalhaes, I. S. in *Population Genomics* (Springer. Cham, 2019).
- Liu, S., Ferchaud, A. L., Gronkjaer, P., Nygaard, R. & Hansen, M. M. Genomic
   parallelism and lack thereof in contrasting systems of three-spined sticklebacks. *Mol. Ecol.* 27, 4725-4743, doi:10.1111/mec.14782 (2018).
- Pujolar, J. M., Ferchaud, A. L., Bekkevold, D. & Hansen, M. M. Non-parallel
   divergence across freshwater and marine three-spined stickleback Gasterosteus
   aculeatus populations. *J. Fish Biol.* **91**, 175-194, doi:10.1111/jfb.13336 (2017).
- Terekhanova, N. V., Barmintseva, A. E., Kondrashov, A. S., Bazykin, G. A. & Mugue,
   N. S. Architecture of Parallel Adaptation in Ten Lacustrine Threespine Stickleback
   Populations from the White Sea Area. *Genome Biol. Evol.* **11**, 2605-2618,
   doi:10.1093/gbe/evz175 (2019).
- Terekhanova, N. V. *et al.* Fast evolution from precast bricks: genomics of young
   freshwater populations of threespine stickleback Gasterosteus aculeatus. *PLoS Genet.* 10, e1004696, doi:10.1371/journal.pgen.1004696 (2014).
- 19 Chan, Y. F. *et al.* Adaptive evolution of pelvic reduction in sticklebacks by recurrent
   deletion of a Pitx1 enhancer. *Science* 327, 302-305, doi:10.1126/science.1182213
   (2010).
- Colosimo, P. F. *et al.* Widespread parallel evolution in sticklebacks by repeated
   fixation of Ectodysplasin alleles. *Science* **307**, 1928-1933,
   doi:10.1126/science.1107239 (2005).
- Nelson, T. C. & Cresko, W. A. Ancient genomic variation underlies repeated ecological adaptation in young stickleback populations. *Evol. Lett.* 2, 9-21, doi:10.1002/evl3.37
   (2018).
- Kemppainen, P. *et al.* Linkage disequilibrium network analysis (LDna) gives a global
   view of chromosomal inversions, local adaptation and geographic structure. *Mol. Ecol. Resour.* 15, 1031-1045, doi:10.1111/1755-0998.12369 (2015).
- Betancur, R. R., Orti, G. & Pyron, R. A. Fossil-based comparative analyses reveal
   ancient marine ancestry erased by extinction in ray-finned fishes. *Ecol. Lett.* 18, 441 450, doi:10.1111/ele.12423 (2015).

- Matschiner, M., Hanel, R. & Salzburger, W. On the origin and trigger of the
   notothenioid adaptive radiation. *PLoS ONE* 6, e18911,
   doi:10.1371/journal.pone.0018911 (2011).
- Meynard, C. N., Mouillot, D., Mouquet, N. & Douzery, E. J. A phylogenetic perspective
   on the evolution of Mediterranean teleost fishes. *PLoS ONE* 7, e36443,
   doi:10.1371/journal.pone.0036443 (2012).
- 26 Sanciangco, M. D., Carpenter, K. E. & Betancur, R. R. Phylogenetic placement of
   enigmatic percomorph families (Teleostei: Percomorphaceae). *Mol. Phylogenet. Evol.* 94, 565-576, doi:10.1016/j.ympev.2015.10.006 (2016).
- Fang, B., Merila, J., Matschiner, M. & Momigliano, P. Estimating uncertainty in
   divergence times among three-spined stickleback clades using the multispecies
   coalescent. *Mol. Phylogenet. Evol.* 142, 106646, doi:10.1016/j.ympev.2019.106646
   (2020).
- Fang, B., Merila, J., Ribeiro, F., Alexandre, C. M. & Momigliano, P. Worldwide
   phylogeny of three-spined sticklebacks. *Mol. Phylogenet. Evol.* 127, 613-625,
   doi:10.1016/j.ympev.2018.06.008 (2018).
- Orti, G., Bell, M. A., Reimchen, T. E. & Meyer, A. Global survey of mitochondrial DNA
   sequences in the threespine stickleback: evidence for recent migrations. *Evolution* 48,
   608-622 (1994).
- 30 Halliburton, R. & Halliburton, R. Introduction to population genetics. (Pearson/Prentice
   Hall Upper Saddle River, NJ, 2004).
- 812
   31
   Hyten, D. L. *et al.* Impacts of genetic bottlenecks on soybean genome diversity. *Proc.* 

   813
   Natl. Acad. Sci. USA 103, 16666-16671, doi:10.1073/pnas.0604379103 (2006).
- 32 Johannesson, K. *et al.* Repeated evolution of reproductive isolation in a marine snail:
   unveiling mechanisms of speciation. *Philos Trans R Soc Lond B Biol Sci* 365, 1735 1747, doi:10.1098/rstb.2009.0256 (2010).
- Kemppainen, P., Lindskog, T., Butlin, R. & Johannesson, K. Intron sequences of
   arginine kinase in an intertidal snail suggest an ecotype-specific selective sweep and a
   gene duplication. *Heredity* **106**, 808-816, doi:10.1038/hdy.2010.123 (2011).
- Roesti, M., Gavrilets, S., Hendry, A. P., Salzburger, W. & Berner, D. The genomic
   signature of parallel adaptation from shared genetic variation. *Mol. Ecol.* 23, 3944 3956, doi:10.1111/mec.12720 (2014).

- 35 Varadharajan, S. *et al.* A high-quality assembly of the nine-spined stickleback
   (Pungitius pungitius) genome. *Genome Biol. Evol.*, doi:10.1093/gbe/evz240 (2019).
- 36 Feder, J. L. & Nosil, P. The efficacy of divergence hitchhiking in generating genomic
   islands during ecological speciation. *Evolution* 64, 1729-1747, doi:10.1111/j.1558 5646.2010.00943.x (2010).
- Ramachandran, S. *et al.* Support from the relationship of genetic and geographic
   distance in human populations for a serial founder effect originating in Africa. *Proc. Natl. Acad. Sci. USA* 102, 15942-15947, doi:10.1073/pnas.0507611102 (2005).
- Bierne, N., Gagnaire, P. A. & David, P. The geography of introgression in a patchy
  environment and the thorn in the side of ecological speciation. *Curr. Zool.* 59, 72-86,
  doi:DOI 10.1093/czoolo/59.1.72 (2013).
- 39 Baker, V. R. & Bunker, R. C. Cataclysmic Late Pleistocene Flooding from Glacial Lake
   Missoula a Review. *Quat. Sci. Rev.* 4, 1-41, doi:Doi 10.1016/0277-3791(85)90027-7
   (1985).
- Bretz, J. H. The Lake Missoula floods and the channeled scabland. *J Geol.* 77, 505 543 (1969).
- 41 Oviatt, C. G. Chronology of Lake Bonneville, 30,000 to 10,000 yr BP. *Quat. Sci. Rev.* 110, 166-171, doi:10.1016/j.quascirev.2014.12.016 (2015).
- 42 Upham, W. *The glacial lake agassiz*. Vol. 25 (US Government Printing Office, 1896).
- 43 Hohenlohe, P. A., Bassham, S., Currey, M. & Cresko, W. A. Extensive linkage
  disequilibrium and parallel adaptive divergence across threespine stickleback
  genomes. *Philos Trans R Soc Lond B Biol Sci* 367, 395-408,
  doi:10.1098/rstb.2011.0245 (2012).
- 44 Bolnick, D. I., Barrett, R. D. H., Oke, K. B., Rennison, D. J. & Stuart, Y. E.
   (Non)Parallel Evolution. *Annu. Rev. Ecol. Evol. Syst.* 49, 303-330,
   doi:10.1146/annurev-ecolsys-110617-062240 (2018).
- 45 Roda, F., Walter, G. M., Nipper, R. & Ortiz-Barrientos, D. Genomic clustering of
  adaptive loci during parallel evolution of an Australian wildflower. *Mol. Ecol.* 26, 36873699, doi:10.1111/mec.14150 (2017).
- 46 Barghi, N. *et al.* Genetic redundancy fuels polygenic adaptation in Drosophila. *PLoS Biol.* **17**, e3000128, doi:10.1371/journal.pbio.3000128 (2019).

- 47 Kautt, A. F., Elmer, K. R. & Meyer, A. Genomic signatures of divergent selection and
   speciation patterns in a 'natural experiment', the young parallel radiations of N
   icaraguan crater lake cichlid fishes. *Mol. Ecol.* 21, 4770-4786 (2012).
- 48 Le Moan, A., Gagnaire, P. A. & Bonhomme, F. Parallel genetic divergence among
   coastal-marine ecotype pairs of European anchovy explained by differential
   introgression after secondary contact. *Mol. Ecol.* 25, 3187-3202 (2016).
- 49 Westram, A. *et al.* Do the same genes underlie parallel phenotypic divergence in different L ittorina saxatilis populations? *Mol. Ecol.* **23**, 4603-4616 (2014).
- 50 Morales, H. E. *et al.* Genomic architecture of parallel ecological divergence: beyond a single environmental contrast. *Sci. Adv.* **5**, eaav9963 (2019).
- Roesti, M., Kueng, B., Moser, D. & Berner, D. The genomics of ecological vicariance
   in threespine stickleback fish. *Nat. Commun.* 6, 8767, doi:10.1038/ncomms9767
   (2015).
- Twyford, A. D. & Friedman, J. Adaptive divergence in the monkey flower Mimulus
   guttatus is maintained by a chromosomal inversion. *Evolution* 69, 1476-1486,
   doi:10.1111/evo.12663 (2015).
- 53 Faria, R. *et al.* Multiple chromosomal rearrangements in a hybrid zone between
   Littorina saxatilis ecotypes. *Mol. Ecol.* 28, 1375-1393, doi:10.1111/mec.14972 (2018).
- 54 Westram, A. M. *et al.* Clines on the seashore: The genomic architecture underlying rapid divergence in the face of gene flow. *Evol. Lett.* **2**, 297-309 (2018).
- Paccard, A. *et al.* Repeatability of Adaptive Radiation Depends on Spatial Scale:
   Regional Versus Global Replicates of Stickleback in Lake Versus Stream Habitats. *J. Hered.* 111, 43-56, doi:10.1093/jhered/esz056 (2020).
- Source of Conte, G. L. *et al.* Extent of QTL Reuse During Repeated Phenotypic Divergence of Sympatric Threespine Stickleback. *Genetics* 201, 1189-1200, doi:10.1534/genetics.115.182550 (2015).
- S7 Conte, G. L., Arnegard, M. E., Peichel, C. L. & Schluter, D. The probability of genetic
   parallelism and convergence in natural populations. *Proc. Biol. Sci.* 279, 5039-5047,
   doi:10.1098/rspb.2012.2146 (2012).
- 58 Hubbard, T. et al. Ensembl 2005. Nucleic Acids Res. 33, D447-D453 (2005).

- Li, H. & Durbin, R. Fast and accurate short read alignment with Burrows–Wheeler
   transform. *Bioinformatics* 25, 1754-1760 (2009).
- 60 Catchen, J., Hohenlohe, P. A., Bassham, S., Amores, A. & Cresko, W. A. Stacks: an
  analysis tool set for population genomics. *Mol. Ecol.* 22, 3124-3140,
  doi:10.1111/mec.12354 (2013).
- Li, H. A statistical framework for SNP calling, mutation discovery, association mapping
   and population genetical parameter estimation from sequencing data. *Bioinformatics* 27, 2987-2993, doi:10.1093/bioinformatics/btr509 (2011).
- Korneliussen, T. S., Albrechtsen, A. & Nielsen, R. ANGSD: Analysis of Next
   Generation Sequencing Data. *BMC Bioinform.* 15, 356, doi:10.1186/s12859-014-0356 4 (2014).
- Kitano, J. *et al.* A role for a neo-sex chromosome in stickleback speciation. *Nature* 461, 1079 (2009).
- Natri, H. M., Shikano, T. & Merilä, J. Progressive recombination suppression and
   differentiation in recently evolved neo-sex chromosomes. *Mol. Biol. Evol.* **30**, 1131 1144 (2013).
- 65 Hedrick, P. W. Sex: differences in mutation, recombination, selection, gene flow, and genetic drift. *Evolution* **61**, 2750-2771 (2007).
- 66 Schaffner, S. F. The X chromosome in population genetics. *Nat. Rev. Genet.* 5, 43-51,
   doi:10.1038/nrg1247 (2004).
- Li, Z., Kemppainen, P., Rastas, P. & Merila, J. Linkage disequilibrium clustering-based approach for association mapping with tightly linked genomewide data. *Mol. Ecol. Resour.* 18, 809-824, doi:10.1111/1755-0998.12893 (2018).
- Fox, E. A., Wright, A. E., Fumagalli, M. & Vieira, F. G. ngsLD: evaluating linkage
   disequilibrium using genotype likelihoods. *Bioinformatics* 35, 3855-3856,
   doi:10.1093/bioinformatics/btz200 (2019).
- 69 Roesti, M., Moser, D. & Berner, D. Recombination in the threespine stickleback
  genome--patterns and consequences. *Mol. Ecol.* 22, 3014-3027,
  doi:10.1111/mec.12322 (2013).
- Matthey-Doret, R. & Whitlock, M. C. Background selection and FST: consequences for
   detecting local adaptation. *Mol. Ecol.* 28, 3902-3914 (2019).

- Stankowski, S. *et al.* Widespread selection and gene flow shape the genomic
   landscape during a radiation of monkeyflowers. *PLoS Biol.* **17**, e3000391 (2019).
- Neuenschwander, S., Hospital, F., Guillaume, F. & Goudet, J. quantiNemo: an
   individual-based program to simulate quantitative traits with explicit genetic
   architecture in a dynamic metapopulation. *Bioinformatics* 24, 1552-1553,
   doi:10.1093/bioinformatics/btn219 (2008).
- 73 Hu, A. *et al.* Influence of Bering Strait flow and North Atlantic circulation on glacial sea level changes. *Nat. Geosci.* 3, 118-121, doi:10.1038/ngeo729 (2010).
- Meiri, M. *et al.* Faunal record identifies Bering isthmus conditions as constraint to end Pleistocene migration to the New World. *Proc. Biol. Sci.* 281, 20132167,
   doi:10.1098/rspb.2013.2167 (2014).
- 75 Zheng, X. *et al.* A high-performance computing toolset for relatedness and principal component analysis of SNP data. *Bioinformatics* 28, 3326-3328,
- 928 doi:10.1093/bioinformatics/bts606 (2012).

#### 930 FIGURES

Figure 1 | Linkage Disequilibrium network analysis (LDna). (a-h) Eight main clusters of 931 loci identified by LDna (LD-clusters). In each panel (LD-cluster), the top and middle plots 932 present the marine-freshwater differentiation ( $F_{ST}$ ) between Atlantic and Eastern Pacific 933 samples, respectively. The bottom plot shows the principal component analysis (PCA) 934 based on the LD-cluster loci. The seven different colours represent the geographic origin 935 of populations. Solid and open circles refer to freshwater and marine ecotypes, 936 respectively. All identified LD-clusters (29 in total) and corresponding information are 937 presented in Extended Data Fig. 1 and Extended Data Fig. 7. (j) Map of the sampled 938 populations; colours match those in the PCA results. A Mercator projection of the sampling 939 map is shown in Extended Data Fig. 6. 940

941

### Figure 2 | Genetic parallelism identified by the unsupervised and supervised

**methods.** (a) Comparison of marine-freshwater differentiation ( $F_{ST}$ ) in the Atlantic (x-axis) 943 and Eastern Pacific (y-axis) datasets for the three LD-clusters (LD-clusters 2, 21 and 29) 944 associated with strong marine-freshwater parallelism in the Eastern Pacific. (b) Genome-945 wide  $F_{ST}$  of the Eastern Pacific samples for loci of the LD-clusters coloured as in (a). (c) 946 The same as (a) but for the twelve LD-clusters (5, 6, 10, 11, 12, 13, 16, 18, 20, 22, 25 and 947 27) that are involved in global marine-freshwater genetic parallelism. (d) and (e) Genome-948 wide F<sub>ST</sub> of the Atlantic and Eastern Pacific samples, respectively, with colours 949 corresponding to LD-cluster loci in (c). The position of the Ectodysplasin (EDA) locus is 950 indicated in (d) and (e). 951

Figure 3 | Ecological genetics in simulated data. (a-e) A schematic of the demographic 953 scenario used for the simulations that is consistent with the "transporter hypothesis". (a) 954 Initial local adaption of the freshwater populations in the Pacific. (b) The colonization of 955 stickleback populations from the Pacific to the Atlantic. (c) Geographic isolation between 956 the two oceans. (d) Extinction of lakes during the last glacial period (LGP) with the survival 957 of refuge populations. (e) The post-glacial colonization of the new freshwater populations. 958 (f) Frequency of selected (freshwater-adapted) alleles in the newly established freshwater 959 populations through generations at high and low levels of trans-oceanic gene flow and 960 different QTL-densities. (g) PCA of the empirical data (LD-cluster 2; left) and the simulated 961 data (right), with ecotypes and geographical regions as shown in the figure legend. (h) 962 Cluster separation score (CSS) of the empirical and simulated data in the Pacific and 963 Atlantic oceans, respectively. (i) Boxplots of observed heterozygosity in different 964 geographical regions in the empirical and simulated data (empirical data, GLM, 965 F<sub>2,64</sub>=43.05, P<0.001; simulated data: GLM, F<sub>1,238</sub>=509.7, P<0.001; Supplementary 966 Information 3). Only trends, rather than absolute values, of heterozygosity should be 967 compared between empirical and simulated data (refer to Extended Data Fig. 5 for further 968 information) 969

970

Figure 4 | Genomic differentiation in simulated data. (a, b) Genome-wide marinefreshwater differentiation (*F*<sub>ST</sub>) from simulated data (data from the last generation
representing present day sampling). For each parameter combination, loci from all 20
replicates were pooled. Red dots indicate QTL, and blue dots indicate loci from LDclusters that were the most similar to LD-cluster 2 (empirical data) showing the strongest
marine-freshwater differentiation in the Eastern Pacific (grey represent non-LD cluster

- 977 loci). (c) *F*<sub>ST</sub> distribution of QTL in the simulations (all replicates pooled), indicating the
- proportion of loci that were either fixed ( $F_{ST}$ ~1), lost ( $F_{ST}$ ~0), or were fixed to different
- degrees in only 1, 2 or 3 of the four freshwater populations (0.1  $\leq F_{ST} \leq 0.9$ ) since post
- glacial colonisation. A small amount of noise (along the x-axis) has been added to the QTL
- 981 positions to improve their visibility.

982









Chr.1 

MUTATION DRIFT BALANCE (40–50kya) PACIFIC  $\bigcirc$  $\bigcirc$ 0- $\bigcirc$ COLONIZATION OF ATLANTIC (38–40 kya) PACIFIC  $\bigcirc$ ATLANTIC -0  $\bigcirc$  $\bigcirc$ \_\_\_\_  $\bigcirc$ GEOGRAPHIC ISOLATION (10–38 kya)  $\bigcirc$  $\bigcirc$ 0 0--0  $\bigcirc$ -V **EXTINCTION OF LGP LAKES** (10 kya) Refugee  $\bigcirc$ Population Refugee Population POST-GLACIAL COLONISATION (10 kya–Present)  $\bigcirc$ 0-\_\_\_\_ 0 0--0 

PLEISTOCENE • LAST GLACIAL PERIOD (LGP)

HOLOCENI

9 PCA

PC 2

PCA of Empirical Data (LD-Cluster 2)

PC 1

## **I** FREQUENCY OF SELECTED ALLELES IN FREASHWATER POPULATIONS



PCA of Simulated Data

Pacific Freshwater (PF)

Atlantic Freshwater (AF)

Pacific Marine (PM)

Atlantic Marine (AM)

PC 1

## **D** CLUSTER SEPARATION SCORE IN THE PACIFIC OCEAN





Region (Empirical Data)



Marine-freshwater differentiation ( $F_{s\tau}$ )