



***SIMULTANEOUS DETERMINATION OF ENDOCRINE DISRUPTING
COMPOUNDS IN ESTUARINE WATER, SEDIMENT AND MARICULTURE
FISHES THROUGH EXTRACTION COUPLED WITH LIQUID
CHROMATOGRAPHY-MASS SPECTROMETRY AND HUMAN HEALTH
RISK MANAGEMENT***

NUR AFIFAH HANUN ISMAIL

FPAS 2020 7



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By

NUR AFIFAH HANUN ISMAIL

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfillment of the Requirements for the degree of Master of Science**

February 2020

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

SIMULTANEOUS DETERMINATION OF ENDOCRINE DISRUPTING COMPOUNDS IN ESTUARINE WATER, SEDIMENT AND MARICULTURE FISHES THROUGH EXTRACTION COUPLED WITH LIQUID CHROMATOGRAPHY-MASS SPECTROMETRY AND HUMAN HEALTH RISK MANAGEMENT

By

NUR AFIFAH HANUN ISMAIL

February 2020

Chairman: Ahmad Zaharin Aris, PhD
Faculty: Environmental Studies

Endocrine disrupting compounds (EDCs) are emerging pollutants causing global concern because they can disrupt the endocrine system in aquatic organisms, mammals, and humans. Because EDCs have been introduced into aquatic ecosystems, the exposure of humans and animals that depend on aquatic foods, especially fishes, should be seriously considered. These pollutants have been released into the environment through many sources, e.g., wastewater treatment plants, terrestrial run-off (industrial activities, pharmaceuticals, and household waste), and precipitation. The use of hormone, pharmaceuticals, pesticides, and fertilizers for maintaining and increasing fish health and growth also contributes to EDC pollution in the water body. Pulau Kukup, Johor, Malaysia is one of the biggest mariculture areas that is actively involved in marine fish export to other countries. As aquaculture production through mariculture activities in Malaysia support food production, the concentration and distribution of EDCs in aquatic ecosystem need to be monitored to secure the food safety. The aim of this study is to optimize a suitable and reliable method to be applied on environmental samples (estuarine water and sediment) and biota sample (mariculture fish) and for the determination of EDCs pollutant in Pulau Kukup, Johor. Besides, this study also presented the human health risk assessment associated with the consumption of fish from Pulau Kukup, Johor, Malaysia. The method displays a high extraction recovery for estuarine water sample, sediment, and mariculture fish in current study, ranging from 92.02% to 132.32 %, 50.39 to 129.10%, and 52.94- 125.95% respectively. The highest concentration EDCs detected in estuarine water sample is diclofenac (< 0.47-79.72 ng/L) followed by E2 (< 5.28-31.11 ng/L) and EE2 (< 0.30-7.69 ng/L). In sediment, bisphenol A (0.072-0.389 ng/g dry weight) was observed as the highest concentration, followed by diethylstilbestrol (< 0.208-0.331 ng/g dry weight) and propranolol (< 0.250-0.275 ng/g dry weight). Meanwhile in fish, the highest concentration of EDCs were detected in muscle, liver, and reproductive organ is

dexamethasone (15.84 ng/g, dried muscle), dexamethasone (43.56 ng/g), and E2 (44.85 ng/g) respectively. Based on human health risk calculation in this study, five targeted EDCs (progesterone, bisphenol A, primidone, sulfamethoxazole, and diclofenac) shown no potential health risk ($HQ < 1$) with the consumption of fish from this mariculture site. This current study can be a baseline assessment for EDCs pollution profile and distribution in the coastal ecosystem from mariculture site throughout the world especially in Malaysia. The data obtained should be relevance to decision-making legislation and policy ratification for food safety to improve the quality of protein-based food and reduce environmental pollution. Owing to the significant concentration of targeted EDCs detected in estuarine water sample, sediment, and mariculture fish, the need to further monitoring in future are required. Although the concentration of targeted compounds obtained are low but their effects may appear in the long term period and this situation alarms not only the environment health but also cause the potential risk to human.

Keywords: Endocrine disrupting compounds, estuarine water sample, sediment, mariculture fish, liquid chromatography mass spectrometer.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**PENENTUAN SERENTAK SEBATIAN PENGENDALA ENDOKRIN (SPE)
PELBAGAI KELAS DALAM AIR MUARA, SEDIMEN, IKAN
MARIKULTUR DENGAN MENGGUNAKAN KAEDAH
PENGEKSTRAKAN BERSAMA SPEKTROMETER JISIM
KROMATOGRAFI CECAIR (SJKC) DAN PENGURUSAN RISIKO
KESIHATAN MANUSIA**

Oleh

NUR AFIFAH HANUN ISMAIL

Februari 2020

Pengerusi: Ahmad Zaharin Aris, PhD

Fakulti: Pengajian Alam Sekitar

Sebatian Pengendala Endokrin (SPE) adalah bahan pencemar yang menyebabkan kebimbangan dunia kerana boleh mengganggu sistem endokrin dalam hidupan akuatik, mamalia, dan manusia. Oleh sebab sebatian SPE telah tersebar dalam ekosistem akuatik, pendedahan manusia dan haiwan lain terhadap makanan laut terutamanya ikan harus diberi pertimbangan yang serius. Bahan pencemar ini telah dibebaskan ke alam sekitar melalui pelbagai sumber, contohnya seperti loji rawatan air kumbahan, sisa daratan (aktiviti perindustrian, sisa ubat-ubatan, dan sisa isi rumah), dan hujan. Penggunaan hormone, ubat-ubatan, racun perosak, dan baja untuk menjaga dan meningkatkan tahap kesihatan dan tumbesaran ikan turut menyumbang kepada pencemaran SPE di dalam ekosistem akuatik. Pulau Kukup, Johor, Malaysia merupakan salah satu kawasan marikultur terbesar yang terlibat aktif dalam mengeksport ikan laut ke negara-negara lain. Sebagai pengeluar makanan akuakultur melalui aktiviti marikultur di Malaysia, kepekatan dan pengagihan SPE dalam ekosistem akuatik perlu dipantau untuk menjamin keselamatan makanan. Tujuan kajian ini adalah untuk mengoptimumkan kaedah yang sesuai dan boleh dipercayai untuk diguna pakai untuk mengekstrak sampel alam sekitar (air muara dan sedimen) dan sampel biota (ikan marikultur), dan dapat memberi input tentang pencemaran SPE di Pulau Kukup, Johor. Selain itu, kajian ini juga membentangkan penilaian risiko kesihatan manusia yang berkaitan dengan pemakanan ikan dari Pulau Kukup, Johor, Malaysia. Kaedah ini memaparkan pemuliharaan ekstraksi tinggi untuk sampel air muara, sedimen, dan ikan marikultur, dari lingkungan 92.02% hingga 132.32%, 50.39 hingga 129.10%, dan 52.94-125.95% bagi sampel masing-masing. Kadar kepekatan SPE yang paling tinggi telah dikesan di dalam air muara adalah diclofenak (<0.47-79.72 ng / L) diikuti oleh E2 (< 5.28-31.11 ng/L) and EE2 (< 0.30-7.69 ng/L). Dalam sedimen, bisphenol A (0.072-0.389

ng / g berat kering) diperhatikan sebagai kepekatan tertinggi, diikuti oleh diethylstilbestrol (<0.208-0.331 ng / g berat kering) dan propranolol (<0.250-0.275 ng / g berat kering). Sementara itu, kepekatan tertinggi EDC yang dikesan dalam isi ikan, hati ikan, dan organ pembiakan ikan adalah dexamethasone (15.84 ng/g, otot kering), dexamethasone (43.56 ng/g) dan E2 (44.85 ng/g) masing-masing. Berdasarkan pengiraan risiko kesihatan manusia dalam kajian ini, lima jenis SPE (progesteron, bisfenol A, primidon, sulfametosazol, dan diklofenak) tidak menunjukkan risiko kesihatan yang berpotensi (HQ <1) jika dikaitkan dengan pengambilan ikan dari lokasi marikultur ini. Kajian semasa ini boleh menjadi penilaian asas untuk memahami profil pencemaran SPE dan penyebaran pencemaran ini dalam ekosistem pantai di tapak marikultur di seluruh dunia terutama di Malaysia. Data yang diperolehi harus relevan untuk merangka dan mengubal dasar bagi keselamatan makanan untuk meningkatkan kualiti makanan berasaskan protein dan mengurangkan pencemaran alam sekitar. Memandangkan kepekatan SPE yang nyata telah dikesan dalam sampel air muara, sedimen, dan ikan marikultur, pemantauan lanjut pada masa akan datang perlu dilakukan. Walaupun kepekatan sebatian pencemaran yang didapati rendah, tetapi kesannya mungkin muncul dalam tempoh jangka panjang dan keadaan ini tidak hanya mengganggu kesihatan persekitaran tetapi juga menyebabkan potensi risiko kepada manusia.

Kata Kunci: Sebatian pemusnah endokrin, air muara, sedimen, ikan marikultur, spektrometer jisim kromatografi cecair.

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Ahmad Zaharin Aris, PhD

Professor
Faculty of Forestry and Environment
Universiti Putra Malaysia
(Chairman)

Nitty Hirawaty Kamarulzaman, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

ZALILLAH MOHD SHARIFF, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

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Signature:

Name of Chairman of
Supervisory
Committee:

Ahmad Zaharin Aris, PhD
Professor
Faculty of Forestry and Environment
Universiti Putra Malaysia

Signature:

Name of Member of
Supervisory
Committee:

Nitty Hirawaty Kamarulzaman, PhD
Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia

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LIST OF ABBREVIATIONS

ADHD	attention deficit hyperactivity disorder
ADI	acceptable daily intake
AP	alkylphenolic
APEOs	alkylphenol ethoxylates
CE	collision energy
dSPE	dispersive solid phase extraction
DLLME	dispersive liquid-liquid microextraction
DO	dissolved oxygen
EDCs	endocrine disrupting compounds
EDI	estimated daily intake
EDTA	ethylenediamine-tetraacetate dehydrate
EP	entrance potential
FAO	Food and Agriculture Organization
FUSLE	focused ultrasound solid-liquid extraction
GH	growth hormone
GnRH	gonadotropin-releasing hormone
HI	health risk
HPA	hypothalamic-pituitary-adrenal
HPLC	high-performance liquid chromatography
HS-SPME	hollow-fiber liquid phase microextraction
LC-MS/MS	liquid chromatography mass spectrometry-mass spectrometry
LIT-MS	linear ion trap
MAE	microwave-assisted extraction

MDL	method detection limit
MQL	method quantification limit
MRM	multiple reaction monitoring
MS	mass-spectrometry
NABC	Needs, Approaches, Benefits, and Challenges
OPP	organophosphorus pesticides
PCA	principle component analysis
PCBs	polychlorinated biphenyls
PTFE	polytetrafluoroethylene
PhACs	Pharmaceutical Active Compounds
PLE	pressurized liquid extraction
POS	polycystic ovarian syndrome
QA	quality assurance
QC	quality control
QUeChERS	Quick, Easy, Cheap, Effective, Rugged, and Safe
rsd	relative standard deviation
Rt	retention time
SPE	solid phase extraction
STP	sewage treatment plants
TDI	tolerable daily intake
TH	thyroid hormone
THQ	target hazard quotient
TOC	total organic carbon
TOF-MS	time-of-flight

TRH	thyrotropin-releasing hormone
TSH	thyroid stimulating hormone
UN	United Nations
UAE	ultrasound-assisted extraction
UHPLC	ultra-high-performance liquid chromatography
USEPA	United States Environmental Protection Agency
WWTPs	waste water treatment plants
WHO	World Health Organization

CHAPTER 1

INTRODUCTION

1.1 Background of study

The occurrence of endocrine disrupting chemicals (EDCs) in the environment has received broad interest over the past decades due to the hazardous characteristic. Over the past 10 years, the study of EDCs has been progress very fast but unfortunately the full range of early work is not presented. These pollutants are present in the environment and used in daily life as food and consumer product and are grouped to natural and artificial hormone, drugs and pharmaceuticals, industrial and household chemicals, pesticide, alkylphenol, and plasticizer (Olujimi et al., 2010; Kabir et al., 2015; Grześkowiak et al., 2016). EDCs generally are defined as chemicals that may interfere with the function of the endocrine system in living things (Barreiros et al., 2016). EDCs can disrupt various body functions with different pathways and mechanisms. EDCs have the ability to mimic and block the endocrine system in humans, have the potential to elicit negative impacts on the hormonal systems of organisms, and cause severe effect such as cancer, abnormal reproductive growth, and metabolic disorders (diabetes, obesity, and endometriosis), leading to a variety of health problems (Mills & Chichester, 2005; Campbell et al., 2006; Deblonde et al., 2011; Bayen et al., 2013; Aris et al., 2014; Esteban et al., 2014b; Kabir et al., 2015; Legler et al., 2015; Giulivo et al., 2016; Ismail et al., 2018).

EDCs can be grouped into natural or synthetic compounds and were produced and introduced to water bodies through waste water treatment plant (WWTP), industrial waste, household waste, agriculture, aquaculture, pharmaceutical waste, urban activities, and indirect sources such as storm-water runoff (Brion et al., 2004; Falconer et al., 2006; Wang et al., 2013; Esteban et al., 2014a; Camilleri and Vulliet, 2015; Salgueiro-González et al., 2015; Barreiros et al., 2016; Chen and Chou, 2016; Omar et al., 2016; Wee et al., 2016; Tan et al., 2018) (Figure 1.1). Then, these pollutants will be absorbed, accumulated, and biomagnified in air, water, sediment, and biota, respectively (Kabir et al., 2015). EDCs also can transport through the food chain via benthic algae and invertebrates (Omar et al., 2019), which may be consumed by fish or birds. Most of the EDCs waste were directly discharged into the aquatic ecosystem and the process of bioaccumulation and biomagnification were taken place here. EDCs which accumulate in water bodies can be transferred into biota matrixes (fish, clam and plankton). Therefore, this process creates potential routes or pathways for EDC exposure to inland and aquatic organisms. Humans can be exposed toward endocrine chemicals when fish and shellfish are consumed. Therefore, it is important to monitor the environmental concentration of EDCs, including in surface water, sediments, aquatic plants and animals, since human are potentially exposed to EDCs via several pathway (Ismail et al., 2017). In agriculture and aquaculture, these pollutants now are strictly banned in feed production and growth antibiotics usage (Wu et al., 2016). Aquaculture and mariculture practices play an important part in bioaccumulation and biomagnification of EDCs in the biota matrixes (fish and shrimp culture). The use of fertilizer and antibiotics for fish

growth and reproduction also can contribute in EDCs pollution. The rate of bioaccumulation is higher in the fishes and highest consumers of the food web since most of the EDCs are lipophilic and concentrated in the fat of the ingesting organisms (Warring and Harris, 2005). The presence of these pollutants in the aquatic ecosystem (aquaculture and mariculture ecosystem), can cause the production of fish to decrease. EDCs can disturb and penetrate into the endocrine system in human and also other organisms. Thus, some effects can be seen in fish, mammals, and also human when exposed to the EDCs. In fish, for example, can cause irregular vitellogenin induction, disturb sex determination, decrease growth rates, delayed reproduction, and altered fish behaviors (Melvin, 2017; Gilannejad et al., 2016; Aris et al., 2014; Segner et al., 2013). Meanwhile in human, EDCs can alter the reproductive system, obesity and diabetes, induce breast cancer cells, disrupt thyroid function, and affect the nervous system (Ismail et al., 2017; Wee and Aris, 2017).

Fish are a primary source of protein for humans, and aquaculture is one of the fastest-growing activities globally. The decline in global fish populations has been exacerbated by an increase in the pollution released into the environment. Currently, a global concern is the occurrence of EDCs in the aquatic ecosystem. Asia accounts for 88% of global aquaculture production and is by far the most intense growth in aquaculture (Fiedler et al., 2016). In the 11th Malaysia Plan (2016–2020), aquaculture had become a very crucial activity where it was appointed as the next (third) engine of national growth (Zainol et al., 2016). Pulau Kukup, Johor, Malaysia is among the largest mariculture sites and involved in active mariculture activity, industrial and commercial activities (Ismail et al., 2018). Fish and other marine products here are actively exported to the neighboring countries like Indonesia and Singapore. Near the mariculture cage, there is a national park (Pulau Kukup National Park) protected by the state of Johor. Thus, this current study was performed to examine the level of pollution of multi-class EDCs in this active mariculture cages. About 20 multi-residues of EDCs consist of pesticides (quinalphos, diazinon, and chlorpyrifos), pharmaceuticals and medical drugs (caffeine, chloramphenicol, amoxicillin, propranolol, atorvastatin, diethylstilbestrol, nitrofurazone, dexamethasone, primidone, sulfamethoxazole, and diclofenac), steroid hormone (testosterone, progesterone, 17 β -estradiol, estrone, and 17 α -ethynylestradiol), and phenolic compound (bisphenol A) were examined in the seawater, sediment, and mariculture fish using analytical procedure. Due to the importance of coastal ecosystem to the human and mainly to aquatic life, some monitoring activities need to be done regularly to provide a monitoring data on the occurrence and distribution profile of these emerging pollutants mainly EDCs in the mariculture area. The data obtained should be of relevance to decision-making legislation and policy ratification for food safety to improve the quality of protein-based food and reduce environmental pollution.

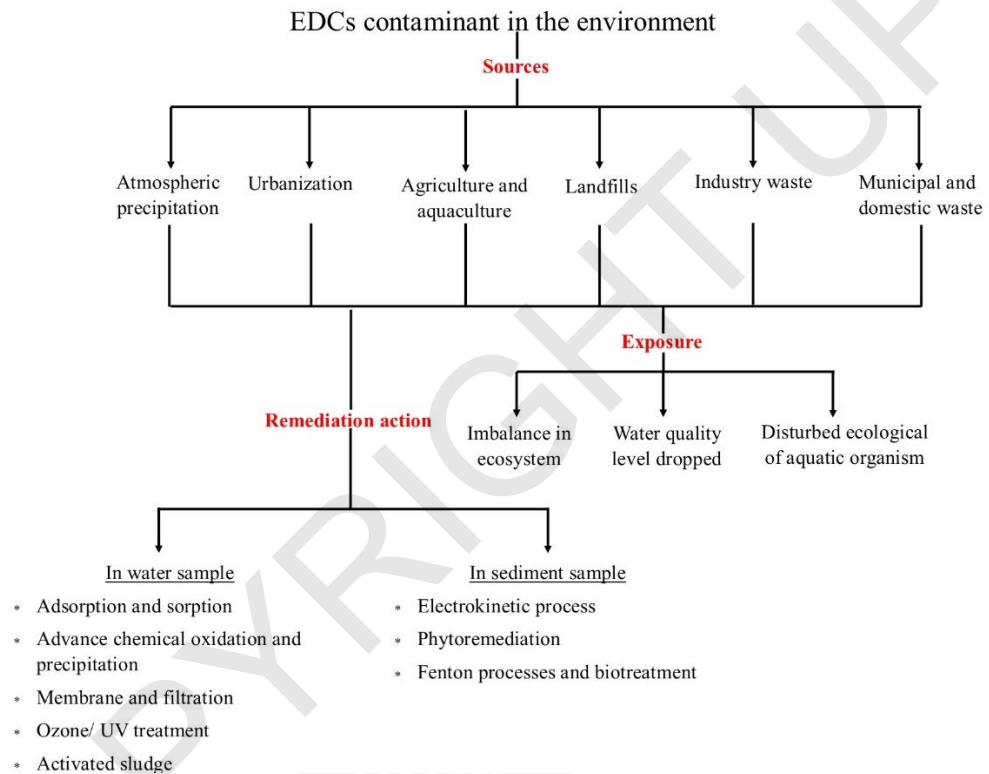


Figure 1.1: Conceptual diagram of occurrence and remediation action of EDCs in the environment.

1.2 Problem statement

EDCs have been identified as an emerging pollutant that arise the awareness around the researchers because of their hazardous and toxicant properties. EDCs have the ability to disrupt and cause the malfunction of endocrine disrupting system in human, terrestrial animals, aquatic organisms, and plants. A very low concentration (pg/L to ng/L) still can give a serious impact to the environment. Although there are lots of studies on EDCs pollutants in environmental matrices, but the suitable method for quantifying multi-residues of EDCs is still inadequate (Kim and Carlson, 2005). In environment, both natural and artificial EDCs are commonly present in the range of pg/L to ng/L (Beck et al., 2005; Hibberd et al., 2009). However, due to the lack of research done on estuarine ecosystem mainly mariculture area, these pollutants remain very much unknown due to the variety and complex of matrixes (Bayen et al., 2013).

Owing to the low concentration of EDCs found in the environment, highly sensitive analytical procedures are needed to identify and quantify these low environmental levels (Álvarez-Muñoz et al., 2015; Yu and Wu, 2015; Ros et al., 2016). In the current study, analytical method for the analysis of a wide range of multi-class EDCs in water sample was optimized using solid phase extraction (SPE). SPE is the simplest and less complex extraction and cleanup steps. Meanwhile, for sediment, the extraction was performed using Soxhlet extraction coupled with SPE and biota sample (mariculture fish) were extracted by sonicator extraction altogether with SPE clean-up method. Furthermore, without considering the selectivity of residues, it is challenging to optimize chromatographic condition and recovery for each of residue when determining multi-residues in a single analytical run (Wee et al., 2016). Due to the others limitation like different solubility for each multi-residue, it is very difficult to determine the high recovery percentage (%) for each of the compounds.

Although the existence of these organic pollutants in water body is still poorly regulated, it still can be a good marker of the human impact on the environment plus be an excellent indicator for water quality (Esteban et al., 2014b). Furthermore, due to the higher dilution in estuarine water, the concentrations of EDCs are expected to be low and these have led to aquatic ecosystems receiving only a little attention in recent years (Beck et al., 2005). Several studies have been conducted to examine heavy metal contaminants in the water, sediment and in other aquatic matrices of marine and estuarine ecosystems in Malaysia (Looi et al., 2013; Achary et al., 2017; Zhang et al., 2017; Wang et al., 2018). However, there is limited information on the organic chemical pollutants in the Malaysia coastal ecosystem particularly for EDCs. In a previous report, organophosphorus pesticides were detected in the Langat River, Selangor, Malaysia (Wee et al., 2016) and multi-residues of pharmaceutically active compounds were detected in the Klang River estuary, Malaysia (Omar et al., 2018). Other studies by Osman et al. (2012), Santhi and Mustafa (2013), Veerasingam and Mustafa, (2013), and Fang et al. (2019) have reported organic pollutants in surface river water in Malaysia. To date, there is scarce literature review has shown on the presence of EDCs in the mariculture site. Due to the importance of coastal ecosystem to the human and mainly to aquatic life, some monitoring activities on organic pollutants, in Pulau Kukup, Johor will be provided a baseline data on the occurrence and profile of these emerging pollutants in the mariculture area.

In addition, due to the sedimentation, resuspension of the bed sediment, high salinity, high matrix effect, and contain lots of various organic matter, analysis of sediment matrices from coastal ecosystem are difficult to be achieved compared to the other matrices. Numerous analytical procedures have been developed and optimized to measure EDCs presence in environment, but the previous studies only focused on the water samples and leaving sediment analyses largely less concerned. According to Zhou and Broodbank (2014), to understand the long-term occurrence of EDCs pollutant, sediment analysis is highly required rather than only water analyses.

Sediment is one of the crucial environmental matrices in the surrounding ecosystem and numerous types of chemicals and pollutants precipitate sank in the sediment (Zhou and Broodbank, 2014; Pintado-Herrera et al., 2016; Omar et al., 2017). Various types of pollutants that are deposited and accumulated in the sediment, may disrupt the ecosystem directly or indirectly and impact the surrounding as sediment plays an extremely important role to maintain the food web and acts as a pool of pollutants for bioaccumulation and trophic transfer (Burton, 2002). The information about the level concentration of EDCs might be a key tool to evaluate the impact of human activities towards aquatic ecosystem although there is no data that has been published in Directive 2008/105/EC (Pintado-Herrera et al., 2016).

Furthermore, the information on level of EDCs pollutants in fish culture is important because the fish production from Pulau Kukup's mariculture site are exported to the other neighbor countries as mentioned in previous section. However, like other farming systems, aquaculture is plagued with disease problems resulting from its intensification and commercialization (Bonded-Reantaso et al., 2005). Natural and synthetic substances such as antibiotics, disinfectants, water and soil treatment compounds, pesticides, fertilizers, probiotic, and other feed additives have become crucial inputs to treat and prevent bacterial and parasitic disease, to improve water quality, to increase pond natural productivity and/or as growth promoters (Bonded-Reantaso et al., 2005; Ali et al., 2016). The usage of these EDCs can contribute to the improvement of the productivity and increased growth of the aquaculture sector. However, it also can cause negative impacts to human and environmental health (Ali et al., 2016).

Among the seafood product, fish is the most consumable protein for human nutrition because it has high source of nutrient such as variable type of vitamin, minerals, and fatty acids that are needed by the human body. For example, high levels of polyunsaturated omega-3 fatty acids cannot be generated by human body and only can be obtained from fish lipids through diet. Increasing of global consuming seafood since 1950s regardless of nature's limitation, the seafood intake still keeps growing till now (Cunha et al., 2017). Owing to the high demands of fish worldwide (Chatterjee et al., 2016) especially for Asian people, who consume large quantity of fish in their daily diet and the occurrence and distribution of organic emerging pollutants has become a hot issue in edible fish species, some precaution action need to be done for ecology and human health (Cheung et al., 2008). Therefore, the study on the quality of seafood by examine the level of targeted multi-class of EDCs in the most consumed fish species in Malaysia is important.

Due to the lack of information on the human health risk assessment for mariculture fish, a good human health risk assessment cannot be reported. Results on monitoring, human health assessment, and managing mariculture activity in relation to environmental protection and food safety cannot be provided due to the inadequate data obtained. Thus, the NABC (Needs, Approaches, Benefits, and Challenges) analysis was conducted in the present study to identify the knowledge gaps of the previous research that needed to be addressed regarding the monitoring studies of EDCs in mariculture ecosystem at Pulau Kukup, Johor (Table 1.1). In addition, the NABC analysis could also assist the present study to overcome the possible limit of the previous studies.

Table 1.1: Needs, Approaches, Benefits, and Challenges (NABC) analysis for recent monitoring studies of EDCs in estuarine water, sediment, and mariculture fish.

	Outputs
Needs	<ul style="list-style-type: none"> • Baseline records on EDCs contaminants in estuarine water, sediment, and culture fish in one of largest and active mariculture site in Malaysia. • Information on the EDCs concentration in mariculture fish is needed to evaluate the degree of EDCs contaminants and associated with human health risks assessment. • Continuous monitoring studies are needed in mariculture area in Pulau Kukup, Johor because aquaculture activity is one of the biggest economic contribution in Malaysia.
Approaches	<ul style="list-style-type: none"> • Determine the concentration of selected targeted EDCs in the estuarine water, sediment, and mariculture fish collected using solid-phase extraction, soxhlet extraction and ultrasonic extraction respectively. • Monitor the level of EDCs contaminants in the estuarine water, sediment, and mariculture fish by using analytical analyses of LC-MS/MS. • Assess the potential human health risk by calculating the human health risk assessment associated with the edible mariculture fish muscle.
Benefits	<ul style="list-style-type: none"> • Provide a better insights on the background concentration of EDCs pollution in aquatic ecosystem mainly in estuarine area. • Provide baseline data on EDCs contaminants status in mariculture site. • Create awareness about the toxicity of EDCs contaminant in the environmental mainly in aquatic ecosystem. • To provide the researchers and government the baseline of guideline on food safety using the result of human health risk assessment.
Challenges	<ul style="list-style-type: none"> • A simple, cheap, and green-based analytical method is needed to extract multi-residue of EDCs in three difference complex matrices (water, sediment, and fish). • A sensitive and accurate chromatogram analysis is needed to analyse and examine the multi-residue of EDCs in a single run time.

1.3 Objectives of the study

The research framework developed for the monitoring study of multi-class EDCs on the estuarine water, sediment, and mariculture fish and the human health risk assessment (Figure 1.2). Generally, the main objective of this study was to monitor the level of multi-class EDCs in estuarine water, sediment, and six different species of mariculture fish (*Trichinous blochii*, *Lutjanus campechanus*, *Lutjanus erythropterus*, *Lutjanus argentimaculatus*, *Carangoides armatu*, and *Lates calcarifer*) at Pulau Kukup, Johor. The specific research objectives are stated as follows:

- i. To modify and optimize the analytical method for simultaneous determination of multi-class EDCs (hormone, pharmaceutical, pesticides, and phenolic compounds) in estuarine water, sediment, and mariculture fish's samples using respectively extraction methods coupled with LC-MS/MS.
- ii. To identify and quantify the occurrence and distribution of multi-class EDCs in environmental matrices (estuarine water and sediment) and elucidate the relationship between multi-class EDCs in environmental matrices with environmental variables.
- iii. To identify and quantify the occurrence and distribution of multi-class EDCs in different part of mariculture fishes (muscle, liver, and reproductive organ) samples and elucidate the relationship between EDCs level with fish size (weight and length).
- iv. To assess the potential human health risk assessment associated with targeted multi-class EDCs contamination in muscle of six different fish species (*T. blochii*, *L. campechanus*, *L. erythropterus*, *L. argentimaculatus*, *C. armatu*, and *L. calcarifer*) from mariculture site.

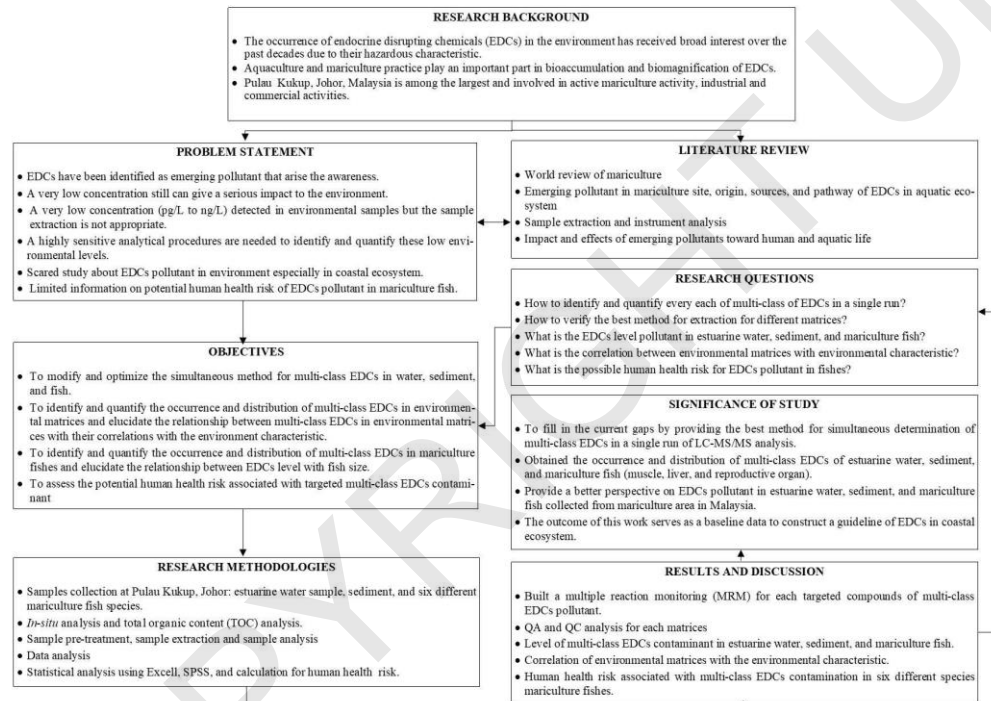


Figure 1.2: Research framework for the analytical study on estuarine water, sediment, and mariculture fish and risk assessment of Pulau Kukup, Johor, Malaysia.

1.4 Scopes of the study

This study generally covers the area as follows:

- i) Involved in the modification and optimization of the method for simultaneous determination of multi-class EDCs (pharmaceutical, phenolic compounds, hormone and pesticides) in estuarine water, sediment, and mariculture fishes samples using SPE extraction, soxhlet extraction, and sonicator extraction respectively.
- ii) Multiple reaction monitoring (MRM) for each targeted compound of multi-residues EDCs, quinalphos, diazinon, chlorpyrifos, caffeine, chloramphenicol, amoxicillin, propranolol, atorvastatin, diethylstilbestrol, nitrofurazone, dexamethasone, primidone, sulfamethoxazole, and diclofenac, testosterone, progesterone, 17 β -estradiol, estrone, 17 α -ethynylestradiol, and bisphenol A was conducted by optimizing the compound-dependent parameters such as declustering potential, entrance potential (EP), exit potential (EP), and collision energy (CE).
- iii) This study focused on the occurrence and distribution of multi-class EDCs in environmental matrices (estuarine water and sediment) and six different species of mariculture fish (*T. blochii*, *L. campechanus*, *L. erythropterus*, *L. argentimaculatus*, *C. armatu*, and *L. calcarifer*) from mariculture site.
- iv) This study also narrowed down to evaluate the relationship between level of multi-class EDCs in samples with environmental variable (environmental matrices) and fish size (mariculture fish).
- v) To assess the potential human health risk assessment associated with multi-class EDCs contamination in muscle of six different species from mariculture site.

1.5 Significance of the study

To date, the previous studies from the literatures have some gaps of knowledge on the multi-class of EDCs monitoring in the tropical estuarine and coastal ecosystem, especially in the mariculture area. The present study is conducted to fill in such information gaps by providing the best method for simultaneous determination of multi-class EDCs in a single run of LC-MS/MS analysis, and using that method, the occurrence and distribution of multi-class EDCs can be obtained for estuarine water, sediment, and mariculture fish (muscle, liver, and reproductive organ). This work uses environmental forensic approach by combining analytical and statistical analyses to trace and interpret the collected data. This study aims to provide the data on EDCs pollutant in estuarine ecosystem focusing on mariculture area at Pulau Kukup, Johor. Quantitative outputs from this study include i) the QA and QC analysis for each matrices collected from Pulau Kukup, Johor, ii) the concentration of EDCs in the estuarine water, sediment, and mariculture fish, and iii) the potential human health risk associated with multi-class EDCs contamination in six different species of mariculture fishes. The outcome of this work serves as a baseline data to construct a guideline of EDCs in water, sediment, and fish in estuarine ecosystem.

Meanwhile, for qualitative outputs include i) the distribution pattern of multi-class EDCs in estuarine ecosystem, ii) the EDCs pollution status of mariculture cages in Pulau Kukup, Johor, Malaysia, iii) the correlation of multi-class EDCs level between environmental matrices (estuarine water and sediment) with the environment characteristic, and iv) the correlation of multi-class EDCs level between mariculture fish with fish size (weight and length). This study can serve as a baseline study to provide a better perspective on EDCs pollutant in estuarine water, sediment, and mariculture fish collected from mariculture area in Malaysia. Finding from this study could also benefit the relevant agencies in setting priorities and allocating resources based on the organic pollution status. The estimated risk could also help to increase the public awareness of the organic pollutant in primary protein sources (mariculture fish).

1.5 Thesis outline

This thesis mainly consists of five chapters which are the Introduction, Literature Review, Methodologies, Results and Discussion, and Summary, Conclusion and Recommendations to provide monitoring data for level concentration of EDCs pollutants in the estuarine water, sediment and mariculture fish. These monitoring data will be the baseline data for constructing a new guideline for EDCs in the marine ecosystem. The chapters of this thesis are arranged as the following flow:

- i. Chapter 1 is an introduction with a brief description of study background, objectives, problem statements, scope and significance of the study.
- ii. Chapter 2 provided a comprehensive literature review related to the world review of aquaculture/mariculture activities, emerging pollutants in aquaculture/mariculture (origin, sources, and pathway of pollutant), types of analytical procedures to detect the pollutant in environment samples, impacts and effect the pollutant toward other organisms, and potential environmental risk.
- iii. Chapter 3 consists of the materials and method applied in this study. This chapter include sampling activities such sample collection and lab analysis like sample preservation, sample pre-treatment, sample extraction, and sample analysis.
- v. Chapter 4 elaborates on the results and discussion of multiple reaction monitoring (MRM) for each targeted compounds of multi-class EDCs pollutant, the occurrence and distribution of multi-class EDCs in samples (estuarine water, sediment, and mariculture fish), correlation of samples with the environment characteristic, and potential human health risk in mariculture fish.
- vi. Chapter 5 summarizes and concludes on the findings and recommendation made on appropriate measures for managing EDCs pollutants in estuarine ecosystem.

REFERENCES

- Abozaid, H., Wessels, S., & Hörstgen-Schwark, G. (2011). Effect of rearing temperatures during embryonic development on the phenotypic sex in zebrafish (*Danio rerio*). *Sexual Development*, 5(5), 259-265.
- Achary, M. S., Satpathy, K. K., Panigrahi, S., Mohanty, A. K., Padhi, R. K., Biswas, S., Prabhu, R.K., Vijayalakshmi, S., & Panigrahy, R. C. (2017). Concentration of heavy metals in the food chain components of the nearshore coastal waters of Kalpakkam, southeast coast of India. *Food control*, 72, 232-243.
- Agusa, T., Kunito, T., Sudaryanto, A., Monirith, I., Kan-Atireklap, S., Iwata, H., Ismail, A., Sanguansin, J., Muchtar, M., Tana, T.S., & Tanabe, S. (2007). Exposure assessment for trace elements from consumption of marine fish in Southeast Asia. *Environmental Pollution*, 145(3), 766-777.
- Ahn, Y. G., Seo, J., Shin, J. H., Khim, J., & Hong, J. (2006). Development of new cleanup method of polychlorinated dibenzo-p-dioxins/dibenzofurans in fish by freezing-lipid filtration. *Analytica chimica acta*, 576(1), 31-36.
- Ahn, Y. G., Shin, J. H., Kim, H. Y., Khim, J., Lee, M. K., & Hong, J. (2007). Application of solid-phase extraction coupled with freezing-lipid filtration clean-up for the determination of endocrine-disrupting phenols in fish. *Analytica chimica acta*, 603(1), 67-75.
- Alexander, K. A., Potts, T. P., Freeman, S., Israel, D., Johansen, J., Kletou, D., Rebours, C., Shorten, M., & Angel, D. L. (2015). The implications of aquaculture policy and regulation for the development of integrated multi-trophic aquaculture in Europe. *Aquaculture*, 443, 16-23.
- Ali, H., Rico, A., Murshed-e-Jahan, K., & Belton, B. (2016). An assessment of chemical and biological product use in aquaculture in Bangladesh. *Aquaculture*, 454, 199-209.
- Álvarez-Muñoz, D., Rodríguez-Mozaz, S., Maulvault, A. L., Tediosi, A., Fernández-Tejedor, M., Van den Heuvel, F., Kotterman, M., Marques, A., & Barceló, D. (2015). Occurrence of pharmaceuticals and endocrine disrupting compounds in macroalgae, bivalves, and fish from coastal areas in Europe. *Environmental research*, 143, 56-64.
- Anumol, T., Vijayanandan, A., Park, M., Philip, L., & Snyder, S. A. (2016). Occurrence and fate of emerging trace organic chemicals in wastewater plants in Chennai, India. *Environment International*, 92, 33-42.
- Aris, A. Z., Shamsuddin, A. S., & Praveena, S. M. (2014). Occurrence of 17 α -ethynylestradiol (EE2) in the environment and effect on exposed biota: a review. *Environment international*, 69, 104-119.
- Ashfield, L. A., Pottinger, T. G., & Sumpter, J. P. (1998). Exposure of female juvenile rainbow trout to alkylphenolic compounds results in modifications to growth and

- ovosomatic index. *Environmental Toxicology and Chemistry: An International Journal*, 17(4), 679-686.
- Baken, K. A., Sjerps, R. M., Schriks, M., & van Wezel, A. P. (2018). Toxicological risk assessment and prioritization of drinking water relevant contaminants of emerging concern. *Environment international*, 118, 293-303.
- Barreiros, L., Queiroz, J. F., Magalhães, L. M., Silva, A. M., & Segundo, M. A. (2016). Analysis of 17- β -estradiol and 17- α -ethinylestradiol in biological and environmental matrices—a review. *Microchemical Journal*, 126, 243-262.
- Bashaar, M., Thawani, V., Hassali, M. A., & Saleem, F. (2017). Disposal practices of unused and expired pharmaceuticals among general public in Kabul. *BMC Public Health*, 17(1), 45.
- Bay, K., Asklund, C., Skakkebaek, N. E., & Andersson, A. M. (2006). Testicular dysgenesis syndrome: possible role of endocrine disruptors. *Best Practice & Research Clinical Endocrinology & Metabolism*, 20(1), 77-90.
- Bayen, S., Estrada, E. S., Juhel, G., Kit, L. W., & Kelly, B. C. (2016). Pharmaceutically active compounds and endocrine disrupting chemicals in water, sediments and mollusks in mangrove ecosystems from Singapore. *Marine Pollution Bulletin*, 109(2), 716-722.
- Bayen, S., Yi, X., Segovia, E., Zhou, Z., & Kelly, B. C. (2014). Analysis of selected antibiotics in surface freshwater and seawater using direct injection in liquid chromatography electrospray ionization tandem mass spectrometry. *Journal of chromatography A*, 1338, 38-43.
- Bayen, S., Zhang, H., Desai, M. M., Ooi, S. K., & Kelly, B. C. (2013). Occurrence and distribution of pharmaceutically active and endocrine disrupting compounds in Singapore's marine environment: influence of hydrodynamics and physical-chemical properties. *Environmental pollution*, 182, 1-8.
- Beck, I. C., Bruhn, R., Gandrass, J., & Ruck, W. (2005). Liquid chromatography-tandem mass spectrometry analysis of estrogenic compounds in coastal surface water of the Baltic Sea. *Journal of Chromatography A*, 1090(1), 98-106.
- Béné, C., Arthur, R., Norbury, H., Allison, E. H., Beveridge, M., Bush, S., Campling, L., Leschen, W., Little, D., Squires, D. & Thilsted, S. H. (2016). Contribution of fisheries and aquaculture to food security and poverty reduction: assessing the current evidence. *World Development*, 79, 177-196.
- Boas, M., Feldt-Rasmussen, U., Skakkebaek, N. E., & Main, K. M. (2006). Environmental chemicals and thyroid function. *European Journal of Endocrinology*, 154(5), 599-611.
- Bondad-Reantaso, M. G., Subasinghe, R. P., Arthur, J. R., Ogawa, K., Chinabut, S., Adlard, R., Tan, Z. & Shariff, M. (2005). Disease and health management in Asian aquaculture. *Veterinary parasitology*, 132(3-4), 249-272.
- Brion, F., Tyler, C. R., Palazzi, X., Laillet, B., Porcher, J. M., Garric, J., & Flammarion, P. (2004). Impacts of 17 β -estradiol, including environmentally relevant

concentrations, on reproduction after exposure during embryo-larval-, juvenile-and adult-life stages in zebrafish (*Danio rerio*). *Aquatic Toxicology*, 68(3), 193-217.

- Burton Jr, G. A. (2002). Sediment quality criteria in use around the world. *Limnology*, 3(2), 65-76.
- Calisto, V., & Esteves, V. I. (2009). Psychiatric pharmaceuticals in the environment. *Chemosphere*, 77(10), 1257-1274.
- Camilleri, J., & Vulliet, E. (2015). Determination of steroid hormones in sediments based on quick, easy, cheap, effective, rugged, and safe (modified-QuEChERS) extraction followed by liquid chromatography-tandem mass spectrometry (LC-MS/MS). *Analytical Methods*, 7(22), 9577-9586.
- Campbell, C. G., Borglin, S. E., Green, F. B., Grayson, A., Wozel, E., & Stringfellow, W. T. (2006). Biologically directed environmental monitoring, fate, and transport of estrogenic endocrine disrupting compounds in water: a review. *Chemosphere*, 65(8), 1265-1280.
- Ccancapa, A., Masiá, A., Navarro-Ortega, A., Picó, Y., & Barceló, D. (2016). Pesticides in the Ebro River basin: occurrence and risk assessment. *Environmental pollution*, 211, 414-424.
- Chatterjee, N. S., Utture, S., Banerjee, K., Shabeer, T. A., Kamble, N., Mathew, S., & Kumar, K. A. (2016). Multiresidue analysis of multiclass pesticides and polyaromatic hydrocarbons in fatty fish by gas chromatography tandem mass spectrometry and evaluation of matrix effect. *Food chemistry*, 196, 1-8.
- Chen, K. Y., & Chou, P. H. (2016). Detection of endocrine active substances in the aquatic environment in southern Taiwan using bioassays and LC-MS/MS. *Chemosphere*, 152, 214-220.
- Chen, S., Yu, X., He, X., Xie, D., Fan, Y., & Peng, J. (2009). Simplified pesticide multiresidues analysis in fish by low-temperature cleanup and solid-phase extraction coupled with gas chromatography/mass spectrometry. *Food Chemistry*, 113(4), 1297-1300.
- Cheung, K. C., Zheng, J. S., Leung, H. M., & Wong, M. H. (2008). Exposure to polybrominated diphenyl ethers associated with consumption of marine and freshwater fish in Hong Kong. *Chemosphere*, 70(9), 1707-1720.
- Chien, L. C., Hung, T. C., Choang, K. Y., Yeh, C. Y., Meng, P. J., Shieh, M. J., & Han, B. C. (2002). Daily intake of TBT, Cu, Zn, Cd and As for fishermen in Taiwan. *Science of the total environment*, 285(1-3), 177-185.
- Choi, K., Kim, Y., Park, J., Park, C. K., Kim, M., Kim, H. S., & Kim, P. (2008). Seasonal variations of several pharmaceutical residues in surface water and sewage treatment plants of Han River, Korea. *Science of the Total Environment*, 405(1-3), 120-128.
- Clotfelter, E. D., Bell, A. M., & Levering, K. R. (2004). The role of animal behaviour in the study of endocrine-disrupting chemicals. *Animal behaviour*, 68(4), 665-676.

- Cockburn, N., Pradhan, A., Taing, M. W., Kisely, S., & Ford, P. J. (2017). Oral health impacts of medications used to treat mental illness. *Journal of affective disorders*, 223, 184-193.
- Comhaire, F. H., Mahmoud, A. M., & Schoonjans, F. (2007). Sperm quality, birth rates and the environment in Flanders (Belgium). *Reproductive Toxicology*, 23(2), 133-137.
- Costa, E. M. F., Spritzer, P. M., Hohl, A., & Bachega, T. A. (2014). Effects of endocrine disruptors in the development of the female reproductive tract. *Arquivos Brasileiros de Endocrinologia & Metabologia*, 58(2), 153-161.
- Cunha, S. C., Oliveira, C., & Fernandes, J. O. (2017). Development of QuEChERS-based extraction and liquid chromatography-tandem mass spectrometry method for simultaneous quantification of bisphenol A and tetrabromobisphenol A in seafood: fish, bivalves, and seaweeds. *Analytical and bioanalytical chemistry*, 409(1), 151-160.
- De Coster, S., & Van Larebeke, N. (2012). Endocrine-disrupting chemicals: associated disorders and mechanisms of action. *Journal of environmental and public health*, 2012.
- Deblonde, T., Cossu-Leguille, C., & Hartemann, P. (2011). Emerging pollutants in wastewater: a review of the literature. *International journal of hygiene and environmental health*, 214(6), 442-448.
- DeMatteo, R., Keith, M. M., Brophy, J. T., Wordsworth, A., Watterson, A. E., Beck, M., Rochon, A., Michael, F., Jyoti, G., Magali, P., & Dayna, R. (2013). Chemical exposures of women workers in the plastics industry with particular reference to breast cancer and reproductive hazards. *New Solutions: A Journal of Environmental and Occupational Health Policy*, 22(4), 427-448.
- Desai, M., Jellyman, J. K., & Ross, M. G. (2015). Epigenomics, gestational programming and risk of metabolic syndrome. *International journal of obesity*, 39(4), 633.
- Dorabawila, N., & Gupta, G. (2005). Endocrine disrupter- estradiol- in Chesapeake Bay tributaries. *Journal of Hazardous Materials*, 120 (1-3), 67-71
- Englert, B. (2007). Method 1694: pharmaceuticals and personal care products in water, soil, sediment, and biosolids by HPLC/MS/MS. *US Environmental Protection Agency (EPA)*, 1-72.
- Errico, S., Nicolucci, C., Migliaccio, M., Micale, V., Mita, D. G., & Diano, N. (2017). Analysis and occurrence of some phenol endocrine disruptors in two marine sites of the northern coast of Sicily (Italy). *Marine pollution bulletin*, 120(1-2), 68-74.
- Esteban, S., Gorga, M., González-Alonso, S., Petrovic, M., Barceló, D., & Valcárcel, Y. (2014). Monitoring endocrine disrupting compounds and estrogenic activity in tap water from Central Spain. *Environmental Science and Pollution Research*, 21(15), 9297-9310.
- Esteban, S., Gorga, M., Petrovic, M., González-Alonso, S., Barceló, D., & Valcárcel, Y. (2014). Analysis and occurrence of endocrine-disrupting compounds and estrogenic

- activity in the surface waters of Central Spain. *Science of the Total Environment*, 466, 939-951.
- Esteve, C., Herrero, L., Gómara, B., & Quintanilla-López, J. E. (2016). Fast and simultaneous determination of endocrine disrupting compounds by ultra-high performance liquid chromatography–tandem mass spectrometry. *Talanta*, 146, 326-334.
- Falconer, I. R., Chapman, H. F., Moore, M. R., & Ranmuthugala, G. (2006). Endocrine-disrupting compounds: A review of their challenge to sustainable and safe water supply and water reuse. *Environmental Toxicology: An International Journal*, 21(2), 181-191.
- FAO. (2014). *The State of World Fisheries and Aquaculture 2014*. Rome.
- FAO. (2015). *Fisheries and aquaculture software. FishStat- software for fishery statistical time series*. In: FAO Fisheries and Aquaculture Department (online). Rome. Updated 16 February 2015.
- Fiedler, J. L., Lividini, K., Drummond, E., & Thilsted, S. H. (2016). Strengthening the contribution of aquaculture to food and nutrition security: the potential of a vitamin A-rich, small fish in Bangladesh. *Aquaculture*, 452, 291-303.
- Gaw, S., Thomas, K. V., & Hutchinson, T. H. (2014). Sources, impacts and trends of pharmaceuticals in the marine and coastal environment. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 369(1656), 20130572.
- Geens, T., Aerts, D., Berthot, C., Bourguignon, J. P., Goeyens, L., Lecomte, P., aghuin-Rogister, G., Pironnet, A.M., Pussemier, L., Scippo, M.L., & Van Loco, J. (2012). A review of dietary and non-dietary exposure to bisphenol-A. *Food and chemical toxicology*, 50(10), 3725-3740.
- Gilannejad, N., Dorafshan, S., Heyrati, F. P., Soofiani, N. M., Asadollah, S., Martos-Sitcha, J. A., Prat, F., Yúfera, M., & Martínez-Rodríguez, G. (2016). Vitellogenin expression in wild cyprinid *Petroleuciscus esfahani* as a biomarker of endocrine disruption along the Zayandeh Roud River, Iran. *Chemosphere*, 144, 1342-1350.
- Giulivo, M., de Alda, M. L., Capri, E., & Barceló, D. (2016). Human exposure to endocrine disrupting compounds: Their role in reproductive systems, metabolic syndrome and breast cancer. A review. *Environmental research*, 151, 251-264.
- Gong, J., Xu, L., Yang, Y., Chen, D. Y., & Ran, Y. (2011). Sequential ASE extraction of alkylphenols from sediments: occurrence and environmental implications. *Journal of hazardous materials*, 192(2), 643-650.
- Gregoraszczyk, E. L., Rak, A., Ludewig, G., & Gasińska, A. (2008). Effects of estradiol, PCB3, and their hydroxylated metabolites on proliferation, cell cycle, and apoptosis of human breast cancer cells. *Environmental toxicology and pharmacology*, 25(2), 227-233.
- Grover, D. P., Balaam, J., Pacitto, S., Readman, J. W., White, S., & Zhou, J. L. (2011). Endocrine disrupting activities in sewage effluent and river water determined by chemical analysis and in vitro assay in the context of granular activated carbon upgrade. *Chemosphere*, 84(10), 1512-1520.

- Grześkowiak, T., Czarczyńska-Goślińska, B., & Zgoła-Grześkowiak, A. (2016). Current approaches in sample preparation for trace analysis of selected endocrine-disrupting compounds: focus on polychlorinated biphenyls, alkylphenols, and parabens. *TrAC Trends in Analytical Chemistry*, 75, 209-226.
- Gu, Y., Yu, J., Hu, X., & Yin, D. (2016). Characteristics of the alkylphenol and bisphenol A distributions in marine organisms and implications for human health: A case study of the East China Sea. *Science of the Total Environment*, 539, 460-469.
- He, X., Dai, K., Li, A., & Chen, H. (2015). Occurrence and assessment of perfluorinated compounds in fish from the Danjiangkou reservoir and Hanjiang river in China. *Food chemistry*, 174, 180-187.
- Hibberd, A., Maskaoui, K., Zhang, Z., & Zhou, J. L. (2009). An improved method for the simultaneous analysis of phenolic and steroidal estrogens in water and sediment. *Talanta*, 77(4), 1315-1321.
- Hong, J., Kim, H. Y., Kim, D. G., Seo, J., & Kim, K. J. (2004). Rapid determination of chlorinated pesticides in fish by freezing-lipid filtration, solid-phase extraction and gas chromatography–mass spectrometry. *Journal of Chromatography A*, 1038(1-2), 27-35.
- Hu, D. P., Hu, W. Y., Xie, L., Li, Y., Birch, L., & Prins, G. S. (2016). Actions of estrogenic endocrine disrupting chemicals on human prostate stem/progenitor cells and prostate carcinogenesis. *The Open Biotechnology Journal*, 10(1).
- Huang, P. C., Tien, C. J., Sun, Y. M., Hsieh, C. Y., & Lee, C. C. (2008). Occurrence of phthalates in sediment and biota: relationship to aquatic factors and the biota-sediment accumulation factor. *Chemosphere*, 73(4), 539-544.
- Iliyasu, A., Mohamed, Z. A., & Terano, R. (2016). Comparative analysis of technical efficiency for different production culture systems and species of freshwater aquaculture in Peninsular Malaysia. *Aquaculture Reports*, 3, 51-57.
- Ismail, N. A. H., Wee, S. Y., & Aris, A. Z. (2017). Multi-class of endocrine disrupting compounds in aquaculture ecosystems and health impacts in exposed biota. *Chemosphere*, 188, 375-388.
- Ismail, N. A. H., Wee, S. Y., & Aris, A. Z. (2018). Bisphenol A and alkylphenols concentrations in selected mariculture fish species from Pulau Kukup, Johor, Malaysia. *Marine pollution bulletin*, 127, 536-540.
- Jobling, S., Casey, D., Rodgers-Gray, T., Oehlmann, J., Schulte-Oehlmann, U., Pawlowski, S., Baunbeck, T., Turner, A.P., & Tyler, C. R. (2003). Comparative responses of molluscs and fish to environmental estrogens and an estrogenic effluent. *Aquatic toxicology*, 65(2), 205-220.
- Kabir, E. R., Rahman, M. S., & Rahman, I. (2015). A review on endocrine disruptors and their possible impacts on human health. *Environmental toxicology and pharmacology*, 40(1), 241-258.
- Kawasaki, N., Kushairi, M. R. M., Nagao, N., Yusoff, F., Imai, A., & Kohzu, A. (2016). Release of nitrogen and phosphorus from aquaculture farms to Selangor River,

Malaysia. *International Journal of Environmental Science and Development*, 7(2), 113.

- Kasprzyk-Hordern, B., Dinsdale, R. M., & Guwy, A. J. (2008). The effect of signal suppression and mobile phase composition on the simultaneous analysis of multiple classes of acidic/neutral pharmaceuticals and personal care products in surface water by solid-phase extraction and ultra performance liquid chromatography–negative electrospray tandem mass spectrometry. *Talanta*, 74(5), 1299-1312.
- Kidd, K. A., Blanchfield, P. J., Mills, K. H., Palace, V. P., Evans, R. E., Lazorchak, J. M., & Flick, R. W. (2007). Collapse of a fish population after exposure to a synthetic estrogen. *Proceedings of the National Academy of Sciences*, 104(21), 8897-8901.
- Kim, M. S., Park, K. S., Pyo, H. S., & Hong, J. K. (2008). Rapid determination of chlorostyrenes in fish by freezing-lipid filtration, solid-phase extraction and gas chromatography-mass spectrometry. *Bulletin of the Korean Chemical Society*, 29(2), 352-356.
- Kim, S. C., & Carlson, K. (2005). LC–MS2 for quantifying trace amounts of pharmaceutical compounds in soil and sediment matrices. *TrAC Trends in Analytical Chemistry*, 24(7), 635-644.
- King Heiden, T. C., Spitsbergen, J., Heideman, W., & Peterson, R. E. (2009). Persistent adverse effects on health and reproduction caused by exposure of zebrafish to 2, 3, 7, 8-tetrachlorodibenzo-p-dioxin during early development and gonad differentiation. *Toxicological Sciences*, 109(1), 75-87.
- Kirby, M. F., Allen, Y. T., Dyer, R. A., Feist, S. W., Katsiadaki, I., Matthiessen, P., Scott, A.P., Smith, A., Stentiford, G.D., Thain, J.E., & Thomas, K. V. (2004). Surveys of plasma vitellogenin and intersex in male flounder (*Platichthys flesus*) as measures of endocrine disruption by estrogenic contamination in United Kingdom estuaries: temporal trends, 1996 to 2001. *Environmental Toxicology and Chemistry: An International Journal*, 23(3), 748-758.
- Kraak, G. V. D., Hewitt, M., Lister, A., McMaster, M. E., & Munkittrick, K. R. (2001). Endocrine toxicants and reproductive success in fish. *Human and Ecological Risk Assessment: An International Journal*, 7(5), 1017-1025.
- Lacey Jr, J. V., Devesa, S. S., & Brinton, L. A. (2002). Recent trends in breast cancer incidence and mortality. *Environmental and molecular mutagenesis*, 39(2-3), 82-88.
- Lapane, K. L., Hume, A., Ulbricht, C., & Gambassi, G. (2016). Safety of psychotropic drugs in the elderly. In *Pharmacovigilance in Psychiatry* 285-297. Springer International Publishing.
- Lee, M. K., & Yoo, S. H. (2014). The role of the capture fisheries and aquaculture sectors in the Korean national economy: An input–output analysis. *Marine Policy*, 44, 448-456.
- Legler, J., Fletcher, T., Govarts, E., Porta, M., Blumberg, B., Heindel, J. J., & Trasande, L. (2015). Obesity, diabetes, and associated costs of exposure to endocrine-

- disrupting chemicals in the European Union. *The Journal of Clinical Endocrinology & Metabolism*, 100(4), 1278-1288.
- Li, W. C. (2014). Occurrence, sources, and fate of pharmaceuticals in aquatic environment and soil. *Environmental Pollution*, 187, 193-201.
- Liu, R., Zhou, J. L., & Wilding, A. (2004). Simultaneous determination of endocrine disrupting phenolic compounds and steroids in water by solid-phase extraction-gas chromatography-mass spectrometry. *Journal of Chromatography A*, 1022(1-2), 179-189.
- Liu, J., Wang, R., Huang, B., Lin, C., Wang, Y., & Pan, X. (2011). Distribution and bioaccumulation of steroidal and phenolic endocrine disrupting chemicals in wild fish species from Dianchi Lake, China. *Environmental pollution*, 159(10), 2815-2822.
- Liu, S., Xu, X. R., Qi, Z. H., Chen, H., Hao, Q. W., Hu, Y. X., Zhao, J. L., & Ying, G. G. (2017). Steroid bioaccumulation profiles in typical freshwater aquaculture environments of South China and their human health risks via fish consumption. *Environmental pollution*, 228, 72-81.
- Locatelli, M., Sciascia, F., Cifelli, R., Malatesta, L., Bruni, P., & Croce, F. (2016). Analytical methods for the endocrine disruptor compounds determination in environmental water samples. *Journal of Chromatography A*, 1434, 1-18.
- Looi, L. J., Aris, A. Z., Johari, W. L. W., Yusoff, F. M., & Hashim, Z. (2013). Baseline metals pollution profile of tropical estuaries and coastal waters of the Straits of Malacca. *Marine pollution bulletin*, 74(1), 471-476.
- Lorenzen, A., Hendel, J. G., Conn, K. L., Bittman, S., Kwabiah, A. B., Lazarovitz, G., Masse, D., McAllister, T. A., & Topp, E. (2004). Survey of hormone activities in municipal biosolids and animal manures. *Environmental Toxicology: An International Journal*, 19(3), 216-225.
- Luzio, A., Santos, D., Fontainhas-Fernandes, A. A., Monteiro, S. M., & Coimbra, A. M. (2016). Effects of 17 α -ethinylestradiol at different water temperatures on zebrafish sex differentiation and gonad development. *Aquatic Toxicology*, 174, 22-35.
- Magi, E., & Di Carro, M. (2018). Marine environment pollution: The contribution of mass spectrometry to the study of seawater. *Mass spectrometry reviews*, 37(4), 492-512.
- Makinwa, T., & Uadia, P. (2017). Occurrence of Bisphenol A (BPA) in Ponds, Rivers and Lagoons in South-Western Nigeria and Uptake in Cat Fish Evidence of Environmental Contamination. *Food and Public Health*, 7(1), 1-6.
- Mamaug, R. E. P. (2016). Supporting ASEAN good aquaculture practices: Utilization of alternative protein sources for aquafeed to minimize pressure on fishery resources. *Fish for the People*, 14(2), 83-89.
- Maqbool, F., Mostafalou, S., Bahadar, H., & Abdollahi, M. (2016). Review of endocrine disorders associated with environmental toxicants and possible involved mechanisms. *Life sciences*, 145, 265-273.

- Matozzo, V., Gagné, F., Marin, M. G., Ricciardi, F., & Blaise, C. (2008). Vitellogenin as a biomarker of exposure to estrogenic compounds in aquatic invertebrates: a review. *Environment international*, *34*(4), 531-545.
- Matuszewski, B. K., Constanzer, M. L., & Chavez-Eng, C. M. (2003). Strategies for the assessment of matrix effect in quantitative bioanalytical methods based on HPLC–MS/MS. *Analytical chemistry*, *75*(13), 3019-3030.
- Meijide, F. J., Vázquez, G. R., Piazza, Y. G., Babay, P. A., Itria, R. F., & Nostro, F. L. (2016). Effects of waterborne exposure to 17 β -estradiol and 4-tert-octylphenol on early life stages of the South American cichlid fish *Cichlasoma dimerus*. *Ecotoxicology and environmental safety*, *124*, 82-90.
- Melvin, S. D. (2017). Effect of antidepressants on circadian rhythms in fish: Insights and implications regarding the design of behavioural toxicity tests. *Aquatic Toxicology*, *182*, 20-30.
- Mijangos, L., Bizkarguenaga, E., Prieto, A., Fernández, L. A., & Zuloaga, O. (2015). Simultaneous determination of a variety of endocrine disrupting compounds in carrot, lettuce and amended soil by means of focused ultrasonic solid–liquid extraction and dispersive solid-phase extraction as simplified clean-up strategy. *Journal of Chromatography A*, *1389*, 8-18.
- Mills, L. J., & Chichester, C. (2005). Review of evidence: are endocrine-disrupting chemicals in the aquatic environment impacting fish populations?. *Science of the total environment*, *343*(1-3), 1-34.
- Mita, L., Bianco, M., Viggiano, E., Zollo, F., Bencivenga, U., Sica, V. Monaco, G., Portaccio, M., Diano, N., Colonna, A., & Lepore, M. (2011). Bisphenol A content in fish caught in two different sites of the Tyrrhenian Sea (Italy). *Chemosphere*, *82*(3), 405-410.
- Nugegoda, D., & Kibria, G. (2017). Effects of environmental chemicals on fish thyroid function: Implications for fisheries and aquaculture in Australia. *General and comparative endocrinology*, *244*, 40-53.
- Nunes, J. P., Ferreira, J. G., Bricker, S. B., O'Loan, B., Dabrowski, T., Dallaghan, B., Hawkins, A. J. S., O'Connor, B., & O'Carroll, T. (2011). Towards an ecosystem approach to aquaculture: assessment of sustainable shellfish cultivation at different scales of space, time and complexity. *Aquaculture*, *315*(3-4), 369-383.
- Oberdörster, E., & Cheek, A. O. (2001). Gender benders at the beach: endocrine disruption in marine and estuarine organisms. *Environmental Toxicology and Chemistry*, *20*(1), 23-36.
- Olujimi, O. O., Fatoki, O. S., Odendaal, J. P., & Okonkwo, J. O. (2010). Endocrine disrupting chemicals (phenol and phthalates) in the South African environment: a need for more monitoring. *Water Sa*, *36*(5).
- Omar, T. F. T., Ahmad, A., Aris, A. Z., & Yusoff, F. M. (2016). Endocrine disrupting compounds (EDCs) in environmental matrices: Review of analytical strategies for pharmaceuticals, estrogenic hormones, and alkylphenol compounds. *TrAC Trends in Analytical Chemistry*, *85*, 241-259.

- Omar, T. F. T., Aris, A. Z., Yusoff, F. M., & Mustafa, S. (2017). An Improved SPE-LC-MS/MS Method for Multiclass Endocrine Disrupting Compound Determination in Tropical Estuarine Sediments. *Talanta*.
- Omar, T. F. T., Aris, A. Z., Yusoff, F. M., & Mustafa, S. (2018). Risk assessment of pharmaceutically active compounds (PhACs) in the Klang River estuary, Malaysia. *Environmental geochemistry and health*, 1-13.
- Omar, T. F. T., Aris, A. Z., Yusoff, F. M., & Mustafa, S. (2019). Occurrence and level of emerging organic contaminant in fish and mollusk from Klang River estuary, Malaysia and assessment on human health risk. *Environmental Pollution*, 248, 763-773.
- Ottinger, M., Clauss, K., & Kuenzer, C. (2016). Aquaculture: relevance, distribution, impacts and spatial assessments—a review. *Ocean & Coastal Management*, 119, 244-266.
- Pal, A., Gin, K. Y. H., Lin, A. Y. C., & Reinhard, M. (2010). Impacts of emerging organic contaminants on freshwater resources: review of recent occurrences, sources, fate and effects. *Science of the total environment*, 408(24), 6062-6069.
- Patrick, L. (2009). Thyroid Disruption: Mechanisms and Clinical Implications in Human Health. *Alternative Medicine Review*, 14(4).
- Peng, X., Wang, Z., Yang, C., Chen, F., & Mai, B. (2006). Simultaneous determination of endocrine-disrupting phenols and steroid estrogens in sediment by gas chromatography–mass spectrometry. *Journal of chromatography A*, 1116(1-2), 51-56.
- Peng, X., Zheng, K., Liu, J., Fan, Y., Tang, C., & Xiong, S. (2018). Body size–dependent bioaccumulation, tissue distribution, and trophic and maternal transfer of phenolic endocrine-disrupting contaminants in a freshwater ecosystem. *Environmental toxicology and chemistry*.
- Pérez, R. A., Albero, B., Tadeo, J. L., & Sánchez-Brunete, C. (2016). Determination of endocrine-disrupting compounds in water samples by magnetic nanoparticle-assisted dispersive liquid–liquid microextraction combined with gas chromatography–tandem mass spectrometry. *Analytical and bioanalytical chemistry*, 408(28), 8013-8023.
- Pessoa, G. P., de Souza, N. C., Vidal, C. B., Alves, J. A., Firmino, P. I. M., Nascimento, R. F., & dos Santos, A. B. (2014). Occurrence and removal of estrogens in Brazilian wastewater treatment plants. *Science of the Total Environment*, 490, 288-295.
- Petrović, M., Eljarrat, E., de Alda, M. J. L., & Barceló, D. (2001). Analysis and environmental levels of endocrine-disrupting compounds in freshwater sediments. *TrAC Trends in Analytical Chemistry*, 20(11), 637-648.
- Pintado-Herrera, M. G., González-Mazo, E., & Lara-Martín, P. A. (2016). In-cell clean-up pressurized liquid extraction and gas chromatography–tandem mass spectrometry determination of hydrophobic persistent and emerging organic pollutants in coastal sediments. *Journal of Chromatography A*, 1429, 107-118.

- Prosser, R. S., & Sibley, P. K. (2015). Human health risk assessment of pharmaceuticals and personal care products in plant tissue due to biosolids and manure amendments, and wastewater irrigation. *Environment international*, *75*, 223-233.
- Rahman, M. F., Yanful, E. K., & Jasim, S. Y. (2009). Occurrences of endocrine disrupting compounds and pharmaceuticals in the aquatic environment and their removal from drinking water: Challenges in the context of the developing world. *Desalination*, *248*(1-3), 578-585.
- Ramaswamy, B. R., Kim, J. W., Isobe, T., Chang, K. H., Amano, A., Miller, T. W., Siringan, F.P., & Tanabe, S. (2011). Determination of preservative and antimicrobial compounds in fish from Manila Bay, Philippines using ultra high performance liquid chromatography tandem mass spectrometry, and assessment of human dietary exposure. *Journal of hazardous materials*, *192*(3), 1739-1745
- Reilly, M. P., & Rader, D. J. (2003) The metabolic syndrome. *Circulation* **108**(13): 1546-1551.
- Rice, C., Birnbaum, L. S., Cogliano, J., Mahaffey, K., Needham, L., Rogan, W. J., & vom Saal, F. S. (2003). Exposure assessment for endocrine disruptors: some considerations in the design of studies. *Environmental health perspectives*, *111*(13), 1683-1690.
- Robinson, T., Ali, U., Mahmood, A., Chaudhry, M. J. I., Li, J., Zhang, G., Jones, K. C., & Malik, R. N. (2016). Concentrations and patterns of organochlorines (OCs) in various fish species from the Indus River, Pakistan: a human health risk assessment. *Science of the Total Environment*, *541*, 1232-1242.
- Ros, O., Vallejo, A., Olivares, M., Etxebarria, N., & Prieto, A. (2016). Determination of endocrine disrupting compounds in fish liver, brain, and muscle using focused ultrasound solid–liquid extraction and dispersive solid phase extraction as clean-up strategy. *Analytical and bioanalytical chemistry*, *408*(21), 5689-5700.
- Saha, N., & Zaman, M. R. (2013). Evaluation of possible health risks of heavy metals by consumption of foodstuffs available in the central market of Rajshahi City, Bangladesh. *Environmental monitoring and assessment*, *185*(5), 3867-3878.
- Saha, N., Mollah, M. Z. I., Alam, M. F., & Rahman, M. S. (2016). Seasonal investigation of heavy metals in marine fishes captured from the Bay of Bengal and the implications for human health risk assessment. *Food Control*, *70*, 110-118.
- Salgueiro-González, N., Turnes-Carou, I., Besada, V., Muniategui-Lorenzo, S., Lopez-Mahia, P., & Prada-Rodríguez, D. (2015). Occurrence, distribution and bioaccumulation of endocrine disrupting compounds in water, sediment and biota samples from a European river basin. *Science of the Total Environment*, *529*, 121-130.
- Schäfers, C., Teigeler, M., Wenzel, A., Maack, G., Fenske, M., & Segner, H. (2007). Concentration- and time-dependent effects of the synthetic estrogen, 17 α -ethinylestradiol, on reproductive capabilities of the zebrafish, *Danio rerio*. *Journal of Toxicology and Environmental Health, Part A*, *70*(9), 768-779.
- Scheifes, A., de Jong, D., Stolker, J. J., Nijman, H. L., Egberts, T. C., & Heerdink, E. R. (2013). Prevalence and characteristics of psychotropic drug use in institutionalized

- children and adolescents with mild intellectual disability. *Research in developmental disabilities*, 34(10), 3159-3167.
- Segner, H., Caroll, K., Fenske, M., Janssen, C. R., Maack, G., Pascoe, D., Schäfers, C., Vandenberg, G. F., Watts, M., & Wenzel, A. (2003). Identification of endocrine-disrupting effects in aquatic vertebrates and invertebrates: report from the European IDEA project. *Ecotoxicology and environmental safety*, 54(3), 302-314.
- Silva, B., Costa, F., Neves, I. C., & Tavares, T. (2015). Pharmaceuticals in the environment: case study of psychiatric drugs. In *Psychiatric Pharmaceuticals as Emerging Contaminants in Wastewater* (pp. 19-46). Springer, Cham.
- Skakkebaek, N. E., Meyts, R. D., & Main, K. M. (2001). Testicular dysgenesis syndrome: an increasingly common developmental disorder with environmental aspects: Opinion. *Human reproduction*, 16(5), 972-978.
- Sodré, F. F., Pescara, I. C., Montagner, C. C., & Jardim, W. F. (2010). Assessing selected estrogens and xenoestrogens in Brazilian surface waters by liquid chromatography–tandem mass spectrometry. *Microchemical Journal*, 96(1), 92-98.
- Sosa-Ferrera, Z., Mahugo-Santana, C., & Santana-Rodríguez, J. J. (2013). Analytical methodologies for the determination of endocrine disrupting compounds in biological and environmental samples. *BioMed research international*, 2013.
- Tan, R., Liu, R., Li, B., Liu, X., & Li, Z. (2018). Typical Endocrine Disrupting Compounds in Rivers of Northeast China: Occurrence, Partitioning, and Risk Assessment. *Archives of environmental contamination and toxicology*, 75(2), 213-223.
- Tankiewicz, M., Fenik, J., & Biziuk, M. (2010). Determination of organophosphorus and organonitrogen pesticides in water samples. *TrAC Trends in Analytical Chemistry*, 29(9), 1050-1063.
- Teraoka, H., Dong, W., Ogawa, S., Tsukiyama, S., Okuhara, Y., Niiyama, M., Ueno, N., Peterson, R.E., & Hiraga, T. (2002). 2, 3, 7, 8-Tetrachlorodibenzo-p-dioxin toxicity in the zebrafish embryo: altered regional blood flow and impaired lower jaw development. *Toxicological Sciences*, 65(2), 192-199.
- Trasande, L., Attina, T. M., & Blustein, J. (2012). Association between urinary bisphenol A concentration and obesity prevalence in children and adolescents. *Jama*, 308(11), 1113-1121.
- Troell, M., Naylor, R. L., Metian, M., Beveridge, M., Tyedmers, P. H., Folke, C., Arrow, K. J., Barrett, S., Crépin, A. S., Ehrlich, P. R., & Gren, Å. (2014). Does aquaculture add resilience to the global food system?. *Proceedings of the National Academy of Sciences*, 111(37), 13257-13263.
- USEPA. US Environmental Protection Agency. (2016). Retrieved from <https://www.epa.gov/risk/human-health-risk-assessment>
- Vandenberg, L. N., & Prins, G. S. (2016). Clarity in the face of confusion: new studies tip the scales on bisphenol A (BPA). *Andrology*, 4(4), 561.

- Verlicchi, P., & Zambello, E. (2016). Predicted and measured concentrations of pharmaceuticals in hospital effluents. Examination of the strengths and weaknesses of the two approaches through the analysis of a case study. *Science of the Total Environment*, 565, 82-94.
- Vulliet, E., Berlioz-Barbier, A., Lafay, F., Baudot, R., Wiest, L., Vauchez, A., Lestremau, F., Botta, F., & Cren-Olivé, C. (2014). A national reconnaissance for selected organic micropollutants in sediments on French territory. *Environmental Science and Pollution Research*, 21(19), 11370-11379.
- Wang, B., Huang, B., Jin, W., Zhao, S., Li, F., Hu, P., & Pan, X. (2013). Occurrence, distribution, and sources of six phenolic endocrine disrupting chemicals in the 22 river estuaries around Dianchi Lake in China. *Environmental Science and Pollution Research*, 20(5), 3185-3194.
- Wang, M., Tong, Y., Chen, C., Liu, X., Lu, Y., Zhang, W., He, W., Wang, X., Zhao, S., & Lin, Y. (2018). Ecological risk assessment to marine organisms induced by heavy metals in China's coastal waters. *Marine pollution bulletin*, 126, 349-356.
- Waring, R. H., Harris, R. M. (2005). Endocrine disrupters: a human risk?. *Molecular and cellular endocrinology* 244(1): 2-9.
- Wee, S. Y., Omar, T. F. T., Aris, A. Z., & Lee, Y. (2016). Surface water organophosphorus pesticides concentration and distribution in the Langat River, Selangor, Malaysia. *Exposure and Health*, 8(4), 497-511.
- Wee, S. Y., & Aris, A. Z. (2017). Endocrine disrupting compounds in drinking water supply system and human health risk implication. *Environment international*, 106, 207-233.
- World Health Organization. (2001). *Water Quality: Guidelines, Standards and Health*.
- Wormley, D. D., Ramesh, A., & Hood, D. B. (2004). Environmental contaminant–mixture effects on CNS development, plasticity, and behavior. *Toxicology and applied pharmacology*, 197(1), 49-65.
- Wu, M., Pan, C., Yang, M., Xu, B., Lei, X., Ma, J., Cai, L., & Chen, J. (2016). Chemical analysis of fish bile extracts for monitoring endocrine disrupting chemical exposure in water: bisphenol A, alkylphenols, and norethindrone. *Environmental toxicology and chemistry*, 35(1), 182-190
- Ying, G. G., Kookana, R. S., & Ru, Y. J. (2002). Occurrence and fate of hormone steroids in the environment. *Environment international*, 28(6), 545-551.
- Yoon, Y., Ryu, J., Oh, J., Choi, B. G., & Snyder, S. A. (2010). Occurrence of endocrine disrupting compounds, pharmaceuticals, and personal care products in the Han River (Seoul, South Korea). *Science of the Total Environment*, 408(3), 636-643.
- Yu, R., Liu, Q., Liu, J., Wang, Q., & Wang, Y. (2016). Concentrations of organophosphorus pesticides in fresh vegetables and related human health risk assessment in Changchun, Northeast China. *Food Control*, 60, 353-360.

- Yu, Y., & Wu, L. (2015). Determination and occurrence of endocrine disrupting compounds, pharmaceuticals and personal care products in fish (*Morone saxatilis*). *Frontiers of Environmental Science & Engineering*, 9(3), 475-481.
- Zainol, M. A., Abas, Z., & Ariffin, A. S. (2016). Supply Chain Integration and Technological Innovation for Business Performance of Aquaculture Contract Farming in Malaysia: A Conceptual Overview. *Int. J Sup. Chain. Mgt Vol*, 5(3), 86.
- Zhang, J., Yang, G. P., Li, Q., Cao, X., & Liu, G. (2013). Study on the sorption behaviour of estrone on marine sediments. *Marine pollution bulletin*, 76(1-2), 220-226.
- Zhang, Y., Chu, C., Li, T., Xu, S., Liu, L., & Ju, M. (2017). A water quality management strategy for regionally protected water through health risk assessment and spatial distribution of heavy metal pollution in 3 marine reserves. *Science of The Total Environment*, 599, 721-731.
- Zheng, N., Wang, Q., Zhang, X., Zheng, D., Zhang, Z., & Zhang, S. (2007). Population health risk due to dietary intake of heavy metals in the industrial area of Huludao city, China. *Science of the Total Environment*, 387(1-3), 96-104.
- Zhou, J., & Broodbank, N. (2014). Sediment-water interactions of pharmaceutical residues in the river environment. *Water research*, 48, 61-70.