



**UNIVERSITI PUTRA MALAYSIA**

***IN VITRO* EVALUATION OF ANTI-DENGUE ACTIVITY OF SELECTED  
SYNTHETIC SCHIFF BASES AND TRADITIONAL CHINESE MEDICINAL  
PLANTS EXTRACTS**

**MARYAM MAQSOOD**

**FPSK(p) 2019 39**



***IN VITRO* EVALUATION OF ANTI-DENGUE ACTIVITY OF SELECTED  
SYNTHETIC SCHIFF BASES AND TRADITIONAL CHINESE MEDICINAL  
PLANTS EXTRACTS**

By

**MARYAM MAQSOOD**

**Thesis Submitted to the Graduate School of Management Universiti Putra Malaysia,  
in Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

**July 2018**

## **COPYRIGHT**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



## **DEDICATION**

This was and will always be for Raiha and Hamza



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

***IN VITRO* EVALUATION OF ANTI-DENGUE ACTIVITY OF SELECTED SYNTHETIC SCHIFF BASES AND TRADITIONAL CHINESE MEDICINAL PLANTS EXTRACTS**

By

**MARYAM MAQSOOD**

July 2018

**Chairman : Chee Hui Yee, PhD**  
**Faculty : Medicine and Health Sciences**

During the last few decades, dengue virus (DENV) has emerged as the major virus spread by mosquito that can cause life threatening disease. Recently, it has spread to more than a hundred countries around the globe and still lacks a specific treatable medication. Hospitals provide only supportive and symptomatic treatment hence the development of a safe and effective antiviral is an urgent need. To identify new antivirals against DENV, realms of both synthetic compounds and ethnomedicine were explored.

A library of eighty-five dithiocarbamate (DTC)-derived Schiff bases and their metal complexes including twelve plant extracts were analyzed in a primary antiviral evaluation. Hits displaying 50% or more anti-dengue activity were selected and evaluated through foci forming unit reduction assay, serotype based analysis and real time quantitative RT-PCR based time-of-addition analysis.

During antiviral evaluation of synthetic compounds, ten anti-dengue compounds were identified with promising antiviral activity. Secondary evaluation presented two lead Ni complexes with SI values 86.3 and 80.4. Ni complexes were the strongest contestants and inhibited all DENV serotypes equally. Lead compounds inhibited against all stages of virus replication cycle but the activity was strongest during early stages. It was demonstrated *in silico* that Ni complexes were binding at domain III of DENV E-glycoprotein involved during early stages of replication. In conclusion, Schiff bases and their metal complexes are a whole new horizon for anti-dengue development and can be studied further to develop safe, effective, stable, and affordable anti-dengue in future.

Discovery and development of modern medicine relies on long and rigorous clinical trials until they are available for public use. Meanwhile, new classes of chemical compounds are being explored for innovative anti-dengue options, Traditional Chinese Medicinal (TCM) plants are frequently being used by dengue endemic populations as Contemporary and Alternate Medicine (CAM). These traditional herbs will continue to be an alternative for dengue treatment as disease burden is rapidly increasing every year. Present study also evaluated the anti-dengue potential of twelve TCM plant extracts designated as cool herbs used for the diseases with high fever.

During antiviral evaluation of TCM plants, four anti-dengue plants were identified. Secondary evaluation proved two lead plants extract *Dryopteris crassirhizoma* (DC) and *Morus alba* (MA) with SI = 4.21 and 4.62 respectively, which inhibited DENV serotypes equally. In general, the plants were not equally inhibiting against all stages of viral replication cycle. DC was identified as potential anti-dengue plant which was active at late stages of virus replication. MA was also inhibiting but the dose was high, however its safety profile was better than DC and the plant was active at early stages of viral replication. Our research elucidated and identified the anti-dengue activity of the TCM plants in practice that have been time-tested to reduce the illness. Similar research can be planned for other traditional anti-dengue practices. The present times of integrated treatments brings together conventional medicine with safe and effective complementary medicine. Combined research approaches from different healthcare disciplines may lead to identification of new therapeutic options.

Abstrat tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PENILAIAN AKTIVITI ANTI-DENGGI *IN VITRO* DI KALANGAN BES SCHIFF SINTETIK DAN TUMBUHAN UBAT TRADISIONAL CINA**

Oleh

**MARYAM MAQSOOD**

July 2018

**Pengerusi : Chee Hui Yee, PhD**

**Fakulti : Perubatan dan Sains Kesihatan**

Dalam beberapa dekad yang lalu, virus denggi (DENV) yang disebarkan oleh nyamuk telah muncul sebagai virus utama yang boleh mengancam nyawa manusia. Virus ini telah merebak melebihi seratus buah negara di seluruh dunia dan masih tiada ubat yang boleh menyembuhkan penyakit ini. Hospital hanya boleh memberikan rawatan sokongan dan mengurangkan simptom pesakit. Justeru itu, pembangunan antivirus yang selamat dan berkesan terhadap DENV telah menjadi keperluan yang mendesak kini. Dalam kajian ini, kami telah menerokai kedua-dua sebatian sintetik dan etnoperubatan untuk tujuan ini.

Analisis telah dijalankan terhadap 85 jenis bes Schiff terbitan ditiokarbamat (DTC) dan kompleks logamnya serta 12 ekstrak tumbuhan untuk menilai aktiviti antivirus utama. Sebatian yang menunjukkan aktiviti anti-denggi melebihi 50% telah dipilih dan dinilai melalui cara *foci forming unit reduction assay*, analisis berasaskangolongan serotaip serta *real-time quatitative (qRT)-PCR based time-of-addition analysis* sehingga tahap perencatan virus.

Sepuluh sebatian telah didapati mempunyai aktiviti anti-denggi yang baik semasa penilaian antivirus sebatian sintetik. Pengesahan kedua telah menunjukkan bahawa dua kompleks yang mengandungi nikel (Ni) dengan nilai SI 86.3 dan 80.4 masing-masing merupakan sebatian utama dalam aktiviti anti-denggi, di mana kedua-dua kompleks ini mempunyai aktiviti perencatan terhadap semua serotaip DENV yang paling ketara. Secara umum, kompleks-kompleks ini didapati merencatkan kitaran replikasi virus di semua peringkat tetapi perencatan yang paling ketara berlaku di peringkat awal kitaran replikasi virus. Kajian *in silico* telah mendapati bahawa kompleks-kompleks ini berikat pada domain III E-glikoprotein DENV yang terlibat semasa peringkat awal replikasi. Kesimpulannya, bes Schiff serta kompleks

logamnya merupakan ufuk yang baru dalam penyelidikan dan pembangunan anti-denggi yang selamat, berkesan, stabil, dan terjangkau pada masa depan.

Penemuan dan perkembangan perubatan moden bergantung pada ujian klinikal yang mengambil masa sehingga boleh digunakan oleh orang awam. Sementara itu, sebatian kimia kelas baru sedang dieksplorasi sebagai pilihan inovatif untuk anti-denggi. Perubatan tradisional Cina (TCM) yang berasaskan tumbuhan sering digunakan oleh penduduk endemik denggi sebagai perubatan kontemporari dan alternatif (CAM). Herba tradisional ini akan terus menjadi rawatan alternatif untuk penyakit denggi kerana beban penyakit denggi yang semakin meningkat setiap tahun. Penyelidikan ini juga menilai potensi anti-denggi untuk dua belas jenis ekstrak tumbuhan TCM yang digunakan secara tradisional sebagai herba penyejuk demam tinggi.

Empat jenis tumbuhan telah didapati mempunyai aktiviti anti-denggi. Melalui pengesahan kedua, ekstrak tumbuhan *Dryopteris crassirhizoma* (DC) dan *Morus alba* (MA) merupakan tumbuhan utama yang mempunyai aktiviti anti-denggi, di mana masing-masing mempunyai nilai SI = 4.21 dan 4.62. Ekstrak-ekstrak ini didapati merencatkan kesemua serotaip DENV sama sekali. Secara umumnya, ekstrak daripada tumbuhan-tumbuhan ini tidak merencatkan semua peringkat dalam kitaran replikasi virus. DC telah dikenal pasti sebagai tumbuhan anti-denggi yang berpotensi serta aktif di peringkat akhir kitaran replikasi virus. MA juga didapati merencatkan DENV tetapi hanya pada dos yang tinggi. Namun demikian, profil keselamatan MA adalah lebih baik daripada DC dan tumbuhan ini adalah efektif di peringkat awal kitaran replikasi virus. Penyelidikan kami telah menjelaskan dan mengenal pasti aktiviti tumbuhan TCM dalam melawan denggi. Rawatan bersepadu kini telah menggabungkan perubatan konvensional dengan perubatan komplementari yang selamat dan efektif. Penggabungan penyelidikan-penyelidikan dari disiplin berlainan akan membantu pengenalan pilihan terapeutik yang baru.



## ACKNOWLEDGEMENTS

From sincere depths of my heart, Praise be to Allah (SWT) to bestow me with strength and capability to proceed on a righteous path with his gracious and merciful help and guidance. Writing this dissertation was one of the most challenging academic endeavors I have ever experienced, and without the support and inspiration from so many people around me, it would never have been accomplished.

Foremost, an honest gratitude to my advisor and my friend Dr. Chee Hui Yee, whose commitment to the highest standards is inspirational. Her expert guidance, warm encouragement, thoughtful direction, and immense knowledge had transformed me over the time. I pay my sincere gratitude for her motivation, enthusiasm, critical comments, and continuous support for the research and for the writing of this thesis with patience. I could not imagine having a better advisor and mentor for the Ph.D. as a milestone in my life.

Besides my advisor, I would like to thank the rest of my thesis committee: Dr. Mohamad Ibrahim Mohamad Tahir and Dr. Ching Siew Mooi, for spirited encouragements, insightful comments, and moral support which kept me going. I pay my sincere thanks also to Prof Dr. Jhonsan Stanslas for introducing me to pharmacology and helping me in this diverse and exciting field. My deepest gratitude to Dr. Tan Sang Loon for helping me with stereochemistry and coordination chemistry and I am also grateful to Dr. Mohammad Alif Mohammad Latif for helping and guiding me through the computational chemistry and molecular docking.

I must thank my fellow lab mates in Virology lab, for the stimulating discussions, for being part of my sorrows and my happy moments and for all the fun we have had in the last four years. My working experience in Virology laboratory with all the friends has been a remarkable expedition. I am also grateful to my friends and colleagues in University Tunku Abdul Rahman for their support and unconditional assistance. I was very lucky to come to Malaysia, I enjoyed the hospitality of the Malaysian people and I loved Malaysian food.

Special thanks to SBK University for believing in me and offering me the needed scholarship, without which my Ph.D. would never have been possible. My special appreciation goes to all my friends and colleagues in SBKW University particularly Prof. Dr. Rukhsana Jabeen, Dr. Muzaffar Khan and the staff of my department, for their consideration and facilitation.

Last but not the least; my words cannot be enough to thank my family. My utmost love and respect for my parents to bear me since my birth to this day and for giving me a unique, magical and a carefree childhood and humble upbringing, they always stood next to me and supported me throughout my life. My love and affection for my siblings for the special bond we share. I give my heart-breaking appreciation to my in-laws for supporting me whenever I needed. A very special thanks to my husband indeed, it is his immeasurable patience and understanding my struggle that helped me stand up every day. My days began with his smile and ended with his love and his hand on my shoulder. My kids Hamza and Raiha deserve the most of my appreciation because they are the most who suffered.

I certify that a Thesis Examination Committee has met on 20 July 2018 to conduct the final examination of Maryam Maqsood on her thesis entitled "*In Vitro* Evaluation of Anti-Dengue Activity of Selected Synthetic Schiff Bases and Traditional Chinese Medicinal Plant Extracts" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

**Vasanth Kumari Neela, PhD**

Associate Professor  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Chairman)

**Zamperi bin Sekawi, PhD**

Professor  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Internal Examiner)

**Intan Safinar Ismail, PhD**

Associate Professor  
Faculty of Science  
Universiti Putra Malaysia  
(Internal Examiner)

**Ramar Krishnamurthy, PhD**

Professor  
Faculty of Science  
Uka Tarsadia University  
India  
(External Examiner)



---

**ROBIAH BINTI YUNUS, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 22 October 2019

This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

**Chee Hui Yee, PhD**

Senior Lecturer  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Chairman)

**Mohamed Ibrahim Bin Mohamed Tahir, PhD**

Senior Lecturer  
Faculty of Science  
Universiti Putra Malaysia  
(Member)

**Ching Siew Mooi, PhD**

Senior Lecturer  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Member)

**ROBIAH BINTI YUNUS, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:

## Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: Maryam Date: \_\_\_\_\_

Name and Matric number: Maryam Maqsood, GS39961

## Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: \_\_\_\_\_  
Name of Chairman  
of Supervisory  
Committee: Dr. Chee Hui Yee

Signature: \_\_\_\_\_  
Name of Member  
of Supervisory  
Committee: Dr. Mohamed Ibrahim Bin Mohamed Tahir

Signature: \_\_\_\_\_  
Name of Member  
of Supervisory  
Committee: Dr. Ching Siew Mooi

## TABLE OF CONTENTS

	<b>Page</b>
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	iii
<b>ACKNOWLEDGEMENTS</b>	v
<b>APPROVAL</b>	vi
<b>DECLARATION</b>	viii
<b>LIST OF TABLES</b>	xiii
<b>LIST OF FIGURES</b>	xv
<b>LIST OF ABBREVIATIONS</b>	xix
<b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	<b>1</b>
1.1 Problem statement	3
1.2 Significance of the study	3
1.3 Objectives	3
1.3.1 General objectives	3
1.3.2 Specific objectives	4
<b>2 LITERATURE REVIEW</b>	<b>5</b>
2.1 Dengue virus	5
2.1.1 Virus structure and replication	5
2.1.2 Virus epidemiology and pathology	8
2.1.3 Virus prevention and control	10
2.1.4 Antiviral research and development	11
2.1.5 Virus entry as antiviral target	15
2.2 Schiff bases	17
2.2.1 S-substituted dithiocarbazate (DTC) derived Schiff bases	17
2.2.2 Schiff base metal complexes and metalloterapeutics	18
2.2.3 Chirality and drug action	19
2.3 Traditional Chinese Medicinal (TCM) plants	20
2.3.1 TCM plants for antiviral evaluation	21
2.3.2 TCM plants with established anti-dengue potential	23
2.4 Virus titration assays	26
<b>3 MATERIAL AND METHODOLOGY</b>	<b>27</b>
3.1 Cell lines	27
3.2 Virus stocks	27
3.3 Positive controls	28
3.4 Synthetic Schiff bases and metal complexes	28
3.5 Crude aqueous plant extracts	30
3.6 TCID <sub>50</sub> assay	31
3.6.1 Result analysis	32
3.7 CPE based antiviral assay	32
3.7.1 Result analysis	32

3.8	MTT cytotoxicity assay	33
3.8.1	Result analysis	33
3.9	Foci Forming Unit Reduction Assay (FFURA)	35
3.9.1	Result analysis	35
3.10	Time-of-addition analysis	36
3.10.1	Prophylactic treatment	36
3.10.2	Anti-adsorption treatment	36
3.10.3	Post adsorption treatment	36
3.10.4	Virus inactivation treatment	37
3.11	Total viral RNA extraction	37
3.12	Real-time Quantitative Reverse Transcriptase Polymerase Chain Reaction (qRT-PCR)	37
3.12.1	Result analysis	38
3.13	Molecular Docking (MD)	38
3.14	HPLC/MS analysis	41
<b>4</b>	<b>RESULTS</b>	<b>42</b>
4.1	DENV stocks for antiviral assays	42
4.2	Evaluation of cytotoxic effects of DMSO dilutions	42
4.3	Determination of anti-dengue potential of DTC-derived Schiff bases and metal complexes	43
4.3.1	CPE reduction based primary antiviral analysis	43
4.3.2	MTT-based cytotoxicity analysis	49
4.3.3	Foci forming unit reduction based secondary antiviral analysis	51
4.3.4	DENV serotypes based antiviral analysis for lead anti-dengue compounds	60
4.3.5	Time-of-addition analysis through quantitative real-time reverse transcriptase PCR	68
4.3.6	Determination of putative receptor-ligand binding interactions	73
4.4	Determination of anti-dengue potential of crude aqueous TCM plant extracts	79
4.4.1	CPE reduction based primary antiviral analysis	79
4.4.2	MTT-based cytotoxicity analysis	83
4.4.3	Foci forming unit reduction based secondary antiviral analysis	85
4.4.4	DENV serotype based antiviral analysis for lead anti-dengue plants	91
4.4.5	Time-of-addition analysis through quantitative real-time reverse transcriptase PCR	99
4.4.6	Determination of putative compounds in active plant extracts	106
<b>5</b>	<b>DISCUSSION</b>	<b>111</b>
5.1	Anti-dengue potential of DTC-derived Schiff bases and their metal complexes	111
5.2	Anti-dengue potential of TCM plant extracts	118

<b>6</b>	<b>SUMMARY, CONCLUSIONS, AND FUTURE RECOMMENDATIONS</b>	125
6.1	Limitations of the study	125
6.2	Schiff bases and their metal complexes as anti-dengue option	125
6.3	TCM plant extracts as anti-dengue option	126
	<b>REFERENCES</b>	128
	<b>APPENDICES</b>	149
	<b>BIODATA OF STUDENT</b>	163
	<b>LIST OF PUBLICATIONS</b>	164





## LIST OF TABLES

Table	Page	
2.1	The DENV structural and non-structural proteins and their properties	7
2.2	Ideal profile for dengue therapeutic target product [51]	12
2.3	List of pharmacological interventions and drugs for DENV treatment in the phases of clinical optimization	14
2.4	Assortment of chiral drugs and the desired therapeutic activity in their two enantiomers with their examples from the recognized medicines	20
2.5	Literature review of some TCM plants with established anti-dengue potential	24
3.1	The sample IDs and molecular weights (M.W) of thirteen compounds consisting of eight starting reactants and five metal salts	29
3.2	The sample IDs and molecular weights (M.W) of 72 S-substituted DTC-derived Schiff bases and their metal complexes	30
3.3	Scientific names, common names, Chinese names, and parts of the plants used as crude aqueous extracts for primary <i>in vitro</i> antiviral evaluation	31
3.4	Sample IDs, molecular weights (M.W.), and molecular structures of six <i>in vitro</i> anti-dengue Ni complexes	40
4.1	The degree of CPE reduction in DENV 2-infected Vero cells upon treatment with twelve selected ligand Schiff base compounds	44
4.2	The degree of CPE reduction in DENV 2-infected Vero cells upon treatment with twelve selected Ni complex compounds	44
4.3	CC50 doses of ten potential <i>in vitro</i> anti-dengue hit compounds determined based on MTT cytotoxicity assay experiments were presented as means and standard error of mean.	50
4.4	IC50 doses of ten potential <i>in vitro</i> anti-dengue Schiff bases and Ni complexes determined based on FFUR assay	59
4.5	Selectivity index (SI) for ten potential <i>in vitro</i> anti-dengue Schiff bases and Ni complexes. SI is calculated as a ratio between CC50 dose and IC50 dose and higher SI means more potent antiviral agent.	59

4.6	IC50 doses calculated from serotype based anti-dengue inhibition of DENV infected Vero cell treated with SMRCM Ni and SMSCM Ni. Vero cells infected with DENV serotypes individually were treated with SMRCM Ni and SMSCM Ni	66
4.7	IC50 doses calculated from time-of-addition analysis for SMRCM Ni and SMSCM Ni treatments against DENV 2-infected Vero cells	71
4.8	AutoDock Vina results for docking between six anti-dengue Ni complexes and three models of DENV E-glycoprotein	74
4.9	Protein-ligand interaction profile for six docked protein-ligand complexes with top binding affinity	75
4.10	The degree of CPE reduction in DENV 2-infected Vero cells upon treatment with twelve crude aqueous plant extracts	83
4.11	CC50 doses of twelve aqueous plant extracts determined based on MTT cytotoxicity assay	84
4.12	IC50 doses of four potential anti-dengue plant extracts determined based on FFUR assay	90
4.13	Selectivity index (SI) of four potential <i>in vitro</i> anti-dengue plant extracts. SI is calculated as a ratio between CC50 dose and IC50 dose and higher SI means more potent antiviral agent	91
4.14	IC50 doses calculated from serotype based anti-dengue inhibition of DENV infected Vero cells treated with <i>Dryopteris crassirhizoma</i> (DC) extracts and <i>Morus alba</i> (MA) extracts. Vero cells infected with four DENV serotypes individually and treated with DC extracts and MA extracts	97
4.15	IC50 doses calculated from time-of-addition analysis for <i>Dryopteris crassirhizoma</i> (DC) extracts and <i>Morus alba</i> (MA) extracts treatments against DENV 2-infected Vero cells	104
4.16	Spectrometric data of putative compounds found in aqueous extracts of <i>Dryopteris crassirhizoma</i> (DC)	108

## LIST OF FIGURES

Figure		Page
2.1	Global dengue incidence and disease burden [1-4]	9
3.1	Methodology work flow for the CPE reduction based antiviral assay and MTT cytotoxicity assay	34
4.1	The conventional reverse transcriptase polymerase chain reaction (RT-PCR) agarose gel electrophoresis analysis for four DENV serotypes	42
4.2	Viable cells (%) at different concentrations of DMSO solution calculated through MTT assay	43
4.3	CPE reduction of DENV 2-infected Vero cells treated with different dilution of compounds SMRCM Ni and SMSCM Ni	46
4.4	CPE reduction of DENV 2-infected Vero cells treated with different dilution of compounds SBRCM Ni and SBSCM Ni	47
4.5	CPE reduction of DENV 2-infected Vero cells treated with different dilution of compounds SMRCQ Ni and SMSCQ Ni	48
4.6	Comparative bar diagrams for CC50 doses calculated through MTT cytotoxicity based analysis for twelve ligand Schiff bases (A) and respective twelve Ni complexes (B)	51
4.7	The anti-dengue effects of ribavirin against DENV 2-infected Vero cells	52
4.8	The anti-dengue effects of SMRCM and SMSCM against DENV 2-infected Vero cells	54
4.9	The anti-dengue effects of SBRCM and SBSCM against DENV 2-infected Vero cells	55
4.10	The anti-dengue effects of SMRCM Ni and SMSCM Ni against DENV 2-infected Vero cells	56
4.11	The anti-dengue effects of SBRCM Ni and SBSCM Ni against DENV 2-infected Vero cells	57
4.12	The anti-dengue effects of SMRCQ Ni and SMSCQ Ni against DENV 2-infected Vero cells	58

4.13	Comparative bar chart of serotype based anti-dengue inhibition of DENV infected Vero cells treated with different concentrations of ribavirin	61
4.14	Anti-dengue effects of SMRCM Ni and SMSCM Ni against DENV 1-infected Vero cells	62
4.15	Anti-dengue effects of SMRCM Ni and SMSCM Ni against DENV 2-infected Vero cells	63
4.16	Anti-dengue effects of SMRCM Ni and SMSCM Ni against DENV 3-infected Vero cells	64
4.17	Anti-dengue effects of SMRCM Ni and SMSCM Ni against DENV 4-infected Vero cells	65
4.18	Two-way ANOVA Bonferroni post-test analysis for the difference of IC50 doses between four DENV serotypes treated individually with SMRCM Ni and SMSCM Ni	67
4.19	Reduction (%) in DENV RNA copy number during prophylactic treatment with SMRCM Ni and SMSCM Ni against DENV 2 infection	69
4.20	Reduction (%) in DENV RNA copy number during anti-adsorption treatment with SMRCM Ni and SMSCM Ni against DENV 2 infection	69
4.21	Reduction (%) in DENV RNA copy number during post adsorption treatment with SMRCM Ni and SMSCM Ni against DENV 2 infection	70
4.22	Reduction (%) in DENV RNA copy number during virus inactivation treatment with SMRCM Ni and SMSCM Ni against DENV 2 infection	70
4.23	Two-way ANOVA Bonferroni post-test analysis between IC50 doses calculated for four time-of-addition of treatments with SMRCM Ni and SMSCM Ni	72
4.24	Top docked pose for lead anti-dengue Ni complex docked with DENV E-glycoprotein (1K4R)	73
4.25	3D structures of ligand SMRCM Ni and SMSCM Ni obtained from PyMol and PLIP server, showing interactions with receptor residues	76
4.26	3D structures of ligand SBRCM Ni and SBSCM Ni obtained from PyMol and PLIP server, showing interactions with receptor residues	77

4.27	3D structures of ligand SMRCQ Ni and SMSCQ Ni obtained from PyMol and PLIP server, showing interactions with receptor residues	78
4.28	CPE reduction of DENV 2-infected Vero cells treated with different dilution of plants extracts <i>Andrographis paniculata</i> (AP), <i>Artemisia annua</i> (AA) and <i>Lonicera japonica</i> (LJ)	80
4.29	CPE reduction of DENV 2-infected Vero cells treated with different dilution of plants extracts <i>Dryopteris crassirhizoma</i> (DC), <i>Lithospermum erythrorhizone</i> (LP), <i>Morus alba</i> (MA) and <i>Smilax glabra</i> (SG)	82
4.30	Comparative bar chart between CC50 doses and the CPE reduction based anti-dengue activity of the twelve plant extracts	85
4.31	Anti-dengue effects of <i>Artemisia annua</i> (AA) extracts against DENV 2-infected Vero cells	86
4.32	Anti-dengue effects of <i>Dryopteris crassirhizoma</i> (DC) extracts against DENV 2-infected Vero cells	87
4.33	Anti-dengue effects of <i>Lithospermum erythrorhizon</i> (LE) extracts against DENV 2-infected Vero cells	88
4.34	Anti-dengue effects of <i>Morus alba</i> (MA) extracts against DENV 2-infected Vero cells	89
4.35	Anti-dengue effects of <i>Smilax glabra</i> (SG) extracts against DENV 2-infected Vero cells	90
4.36	Comparative bar chart of serotype based anti-dengue inhibition of DENV infected Vero cell treated with <i>Artemisia annua</i> (AA) extracts	92
4.37	Anti-dengue effects of <i>Dryopteris crassirhizoma</i> (DC) extracts and <i>Morus alba</i> (MA) extracts with DENV 1-infected Vero cells	93
4.38	Anti-dengue effects of <i>Dryopteris crassirhizoma</i> (DC) extracts and <i>Morus alba</i> (MA) extracts with DENV 2-infected Vero cells	94
4.39	Anti-dengue effects of <i>Dryopteris crassirhizoma</i> (DC) extracts and <i>Morus alba</i> (MA) extracts with DENV 3-infected Vero cells	95
4.40	Anti-dengue effects of <i>Dryopteris crassirhizoma</i> (DC) extracts and <i>Morus alba</i> (MA) extracts with DENV 4-infected Vero cells	96
4.41	Two-way ANOVA Bonferroni post-test analysis for the difference of IC50 doses between four DENV serotypes treated individually with <i>Dryopteris crassirhizoma</i> (DC) extracts and <i>Morus alba</i> (MA) extracts	98

4.42	Reduction (%) in DENV RNA copy number of prophylactic treatment with <i>Dryopteris crassirhizoma</i> (DC) extracts and <i>Morus alba</i> (MA) extracts against Vero cells	100
4.43	Reduction (%) in DENV RNA copy number of anti-adsorption treatment with <i>Dryopteris crassirhizoma</i> (DC) extracts and <i>Morus alba</i> (MA) extracts together with DENV 2 infection	101
4.44	Reduction (%) in DENV RNA copy number of treatment with <i>Dryopteris crassirhizoma</i> (DC) extracts and <i>Morus alba</i> (MA) extracts in a post adsorption treatment against DENV 2	102
4.45	Reduction (%) in DENV RNA copy number of virus inactivation treatment with <i>Dryopteris crassirhizoma</i> (DC) extracts and <i>Morus alba</i> (MA) extracts against DENV 2 particles	103
4.46	Two-way ANOVA Bonferroni post-test analysis between IC50 doses calculated for four time-of-additions of treatments with <i>Dryopteris crassirhizoma</i> (DC) extracts and <i>Morus alba</i> (MA) extracts activity was not detected within the range of tested concentration	105
4.47	The HPLC/MS results for the aqueous extracts of <i>Dryopteris crassirhizoma</i> (DC)	107
4.48	The HPLC/MS results for the aqueous extracts of <i>Morus alba</i> (MA)	109

## LIST OF ABBREVIATIONS

AA	<i>Artemisia annua</i>
ADE	Antibody-dependent enhancement
AP	<i>Andrographis paniculata</i>
CAM	Complementary and alternative medicine
Cd	Cadmium
Co	Cobalt
C protein	Core Protein
CPE	Cytopathic effect
Cu	Copper
DC	<i>Dryopteris crassirhizoma</i>
DENV	Dengue Virus
DF	Dengue fever
DHF	Dengue Hemorrhagic fever
DMSO	Dimethyl sulfoxide
DTC	Dithiocarbazate
DSS	Dengue shock syndrome
E protein	Envelope protein
EMEM	Eagles minimal essential media
FBS	Fetal bovine serum
HCV	Hepatitis C virus
HPV	Human papilloma virus
IT	<i>Isatis tinctorial</i>
JEV	Japanese Encephalitis Virus
LCMS	Liquid chromatography –mass spectrometry
LE	<i>Lithospermum erythrorhizone</i>
LJ	<i>Lonicera japonica</i>
MA	<i>Morus alba</i>
MD	Molecular docking
MTT	3-(4,5-dimethylethiaza-2-yl)-2,5-diphenyltetrazolium bromide
m/z	Mass-to-charge ratio

PBS	Phosphate buffer saline
PC	<i>Phellodendron chinense</i>
QSAR	Quantitative structure-activity relationship
RO	<i>Rheum officinale</i>
RT	Room temperature
RCM	<i>R</i> -camphor
RCQ	<i>R</i> -camphorquinone
RCV	<i>R</i> -carvone
SBDTC	<i>S</i> -benzyl dithiocarbazate
SI	Selectivity index
SG	<i>Smilax glabra</i>
SCM	<i>S</i> -camphor
SCQ	<i>S</i> -camphorquinone
SCV	<i>S</i> -carvone
SMDTC	<i>S</i> -methyl dithiocarbazate
SN	<i>Scrophularia ningpoensis</i>
TEM	Transmission electron microscopy
TCM	Traditional Chinese medicines
TCID50	Tissue culture infectious dose 50
VY	<i>Viola yedoensis</i>
WNV	West Nile Virus
Zn	Zinc



## CHAPTER 1

### INTRODUCTION

Dengue virus (DENV) is a positive single-stranded RNA virus of the *Flaviviridae* family that causes dengue fever (DF) with and without warning signs and severe dengue. It has four serotypes DENV 1, DENV2, DENV3, and DENV4 that are genetically and antigenically distinct and epidemiologically similar. Infection with one serotype leads to all-time protection against homologous re-infection but only brief protection against heterologous confront. DENV infection and immune system interactions may result in either immunopathology leading to severe forms of the disease or recovery from infection [1]. DENV is emerging as one of the biggest threats to the human population in the tropical and subtropical region, affecting 3.9 billion people worldwide every year. It is spreading rapidly due to global demographic changes, rapid and unrestrained urbanization, population growth and global ease of travelling [2]. In a recent study, the total cost for the management of the disease caused by DENV was estimated globally which turned out to be US\$8.9 billion per year. The DENV transmission has also been shown in 141 countries around the globe [3]. Currently, there is no therapeutic medicine for DENV and according to WHO, vector management policies are the only alternative control [4]. Besides all the measures taken for the vector control, the increasing number of dengue cases around the globe shows the failure of vector management measures. Treatment is largely supportive and symptomatic, using common antipyretics and analgesics, oral or intravenous rehydration and in-patient monitoring. Appropriate medical attention reduces the fatality rate of severe disease to <1%. Currently, there are many dengue vaccine candidates and the leading candidate is CYD-TDV (Dengvaxia, Sanofi Pasteur), which manage to get licence from a few national regulatory authorities. However, DENV vaccine is intricate because of antibody-dependent enhancement (ADE) of the disease leading to Dengue Haemorrhagic Fever (DHF) and Dengue Shock Syndrome (DSH) [5].

The disease has already grown around the globe and the symptomatic patients are increasing every year, so a suitable and effective antiviral is a better therapeutic solution for the ones who are sick and seeking treatment. Similarly, a prophylactic therapeutic option can help people living in high risk areas, to decrease dengue symptoms during dengue epidemics. Searching for a therapeutic solution for DENV is a matter of rigorous investigation and examining both synthetic and naturally available options. The advancement in high through-put target based approach for drug evaluation has led to the discovery of less number of approved drugs than the era of classical phenotypic target-free drug discovery approach [6]. Therefore, many diseases are still lacking a specific treatment because the ideal disease target is yet to be identified. Consequently, phenotypic screening is gaining new momentum in drug screening. Phenotypic antiviral screening in resource-limited settings is convenient for the disease endemic countries because they can examine the locally infecting clinical isolates of the pathogen to identify effective antiviral options. Targets can then be identified for the potential *in vitro* or *in vivo* antiviral therapeutics.

Schiff bases have been long known for their remarkable biological activities such as antimicrobial, antihistaminic, antipyretic, antifungal, anthelmintic, antiviral as well as they have been widely explored for their industrial applications. Schiff bases are the huge class of compounds and are easily derived from a wide range of derivatives while based on their types of derivative they may have varied biological activity. Schiff base complexes are among the most important stereochemical models and metal coordination chemistry due to their preparative accessibility and structural variety, and the metal complexes have enhanced biological activities [7]. Schiff bases and metal complexes are widely studied owing to their ease of preparation and diverse pharmacological potential. Medicinal chemists across the world have done immense work on Schiff bases and developed agents with better medicinal activity and low toxicity profiles. However, the antiviral activity of these compounds deserves further investigation. Present study focuses on S-substituted dithiocarbamate (DTC)-derived Schiff bases and their metal complexes for their antiviral potential. Other than the fact that these compounds were easily and economically synthesized locally by the Department of Chemistry, Universiti Putra Malaysia, selection of this compound library was for two main reasons;

1. This synthetic compound library is proved to have anticancer properties while cancers are sometimes found associated with virus. Therefore, it was suggested that if these compounds are anticancer they might have antiviral potential as well.
2. Metal complexes were aimed for the metallotherapeutics, the principle of metal based therapeutics is that metal ions can alter the virus structure by metal chelation because virus nucleic acid and proteins are effective chelating agents.

The whole compound library was a series of structurally related compounds and their respective metal complexes, which were systematically subjected to *in vitro* antiviral evaluation to find out any pattern of viral inhibition that could be related to the structure or the addition of the metal atom to the compounds.

Medicinal herbs are being progressively documented as useful complementary treatments for many ailments. Many studies have reported the beneficial as well as safer effects of herbal medicines on patients suffering from common to life threatening diseases. Studies even support the idea of enhanced therapeutics when these herbal medicines are used in combination with conventional antiviral medicines [8]. For diseases like dengue, traditional herbs are a safe alternative approach as modern medicine has not managed to solve the problem yet. There are many herbs that are being used in routine as practical and clinical alternative medicine against DENV infection by the local communities, that have been time-tested to reduce the illness [9]. These traditional herbs will continue to be an alternative for dengue treatment as the discovery and development of modern medicine relies on long and rigorous clinical trials to test the effectiveness of a drug. This process may involve years of research, meanwhile, if the scientific research proves the effectiveness of these medicinal plants, patients can always benefit from

them. Physicians may recommend alternative therapies to their patients for the diseases lacking specific medicinal treatment which might work better than expensive drugs at treating life threatening diseases like dengue. Chinese Herbal Medicine (CHM) is one of the great herbal systems of the world and it is the most important components of Traditional Chinese Medicine (TCM). It has been reported to cure infectious diseases, in the form of hot water extracts, for almost 2,000 years with 100,000 recorded recipes for 10,000 herbal medicines in ancient TCM literature [10]. These TCM and other Complementary and Alternative Medicine (CAM) with their plants and plant-based products will continue to provide treatments for many diseases including dengue for which modern medicine cannot cater. Therefore, present study also focused on the anti-dengue potential of TCM herbal medicines.

### **1.1 Problem statement**

Over the past few decades, there has been a dramatic increase in cases of dengue infection thus, proving that vector management policies alone are not enough to curb the problem. Though vaccine is a better option for any viral disease but dengue vaccine is complicated because of antibody-dependent enhancement which compromises its efficacy. Dengue is a rapidly increasing health problem contributing immensely toward health economics in endemic countries and patient with symptomatic cases are on the rise, seeking medical treatment. Whereas, need for the development of effective dengue therapeutic and prophylactic is still not fulfilled.

### **1.2 Significance of the study**

This study focuses on both the naturally existing and synthetic therapeutic options for the development of safe and effective dengue therapeutics. The natural options are the medicinal plants used as Traditional Chinese Medicine (TCM) for centuries to treat numerous common illnesses as well as dengue fever. Synthetic library includes perceptively designed compound scheme of Schiff bases for chemotherapeutics and their metal complexes for more recent metallotherapeutics. Prospective antivirals evaluated through a blind target-free phenotypic analysis method, an original drug evaluation paradigm, for the hit identification. Various virological methods employed for hit-to-lead and earlier lead optimization of the drug discovery process. The leading prospective anti-dengue agents can be made available for future *in vivo* studies and clinical trials.

### **1.3 Objectives**

#### **1.3.1 General objectives**

To evaluate potential anti-dengue activity of synthetic compounds and crude plant extracts and make them available for further research.

### 1.3.2 Specific objectives

1. To systematically evaluate synthetic compounds and crude plant extracts against DENV 2 to identify the anti-dengue “hits”.
2. To determine the cytotoxic dose (CC50) of the potential anti-dengue compounds and plant extracts.
3. To optimize the “lead” through Foci Forming Unit Reduction Assay (FFURA) and to measure the difference of inhibition against four serotypes of DENV.
4. To perform time-of-addition analysis by measuring the reduction in DENV RNA copy number through quantitative real-time RT-PCR and deduce stage of virus inhibition.



## REFERENCES

- [1.] Halstead, S.B., *Pathogenesis of dengue: challenges to molecular biology*. Science, 1988. **239**(4839): p. 476.
- [2.] Bhatt, S., et al., *The global distribution and burden of dengue*. Nature, 2013. **496**(7446): p. 504.
- [3.] Shepard, D.S., et al., *The global economic burden of dengue: a systematic analysis*. The Lancet Infectious Diseases, 2016. **16**(8): p. 935-941.
- [4.] Gubler, D.J., *The economic burden of dengue*. The American journal of tropical medicine and hygiene, 2012. **86**(5): p. 743-744.
- [5.] Ruche, G.I., et al., *First two autochthonous dengue virus infections in metropolitan France, September 2010*. Eurosurveillance, 2010. **15**(39) : p. 73-74.
- [6.] Zheng, W., N. Thorne, and J.C. McKew, *Phenotypic screens as a renewed approach for drug discovery*. Drug discovery today, 2013. **18**(21): p. 1067-1073.
- [7.] Abu-Dief, A.M. and I.M. Mohamed, *A review on versatile applications of transition metal complexes incorporating Schiff bases*. Beni-suef university journal of basic and applied sciences, 2015. **4**(2): p. 119-133.
- [8.] Hozumi, T., et al., *Antiviral agent containing crude drug*. 1995, Google Patents.
- [9.] Handel, A.S., et al., *Knowledge, attitudes, and practices regarding dengue infection among public sector healthcare providers in Machala, Ecuador*. Tropical Diseases, Travel Medicine and Vaccines, 2016. **2**(1): p. 8.
- [10.] Lin, L.L., et al., *Application of Traditional Chinese Medical Herbs in Prevention and Treatment of Respiratory Syncytial Virus*. Evidence-Based Complementary and Alternative Medicine, 2016. **204**(2): p. 199-133.
- [11.] Holmes, E.C. and S.S. Twiddy, *The origin, emergence and evolutionary genetics of dengue virus*. Infection, genetics and evolution, 2003. **3**(1): p. 19-28.
- [12.] Gubler, D.J., *Dengue and dengue hemorrhagic fever*. Clinical microbiology reviews, 1998. **11**(3): p. 480-496.
- [13.] Gubler, D.J., *COMMENTARY: Ashburn PM, Craig CF. Experimental Investigations Regarding the Etiology of Dengue. J Infect Dis 1907; 4: 440–75*. Journal of Infectious Diseases, 2004. **189**(9): p. 1744-1783.

- [14.] Sabin, A.B., *Research on dengue during World War II*. American journal of tropical medicine and hygiene, 1952. **1**(1): p. 30-50.
- [15.] Hammon, W.M., A. Rundnick, and G. Sather, *Viruses associated with epidemic hemorrhagic fevers of the Philippines and Thailand*. Science, 1960. **131**(3407): p. 1102-1103.
- [16.] Halstead, S., *Arboviruses of the Pacific and Southeast Asia*. Textbook of pediatric infectious diseases, third edition. Philadelphia, PA: WB Saunders, 1992: p. 1468-75.
- [17.] Kuhn, R.J., et al., *Structure of dengue virus: implications for flavivirus organization, maturation, and fusion*. Cell, 2002. **108**(5): p. 717-725.
- [18.] Halstead, S.B., *Dengue*. Vol. 5. 2008: World Scientific.
- [19.] Alen, M.M. and D. Schols, *Dengue virus entry as target for antiviral therapy*. Journal of tropical medicine, 2012. **204**(2): p. 119-178.
- [20.] Gebhard, L.G., C.V. Filomatori, and A.V. Gamarnik, *Functional RNA elements in the dengue virus genome*. Viruses, 2011. **3**(9): p. 1739-1756.
- [21.] Alcaraz-Estrada, S.L., M. Yocupicio-Monroy, and R.M. del Angel, *Insights into dengue virus genome replication*. Future Virology, 2010. **5**(5): p. 575-592.
- [22.] Ma, L., et al., *Solution structure of dengue virus capsid protein reveals another fold*. Proceedings of the National Academy of Sciences of the United States of America, 2004. **101**(10): p. 3414-3419.
- [23.] Yu, I.-M., et al., *Structure of the immature dengue virus at low pH primes proteolytic maturation*. Science, 2008. **319**(5871): p. 1834-1837.
- [24.] Lin, S.-R., et al., *The helical domains of the stem region of dengue virus envelope protein are involved in both virus assembly and entry*. Journal of virology, 2011. **85**(10): p. 5159-5171.
- [25.] Libraty, D.H., et al., *High circulating levels of the dengue virus nonstructural protein NS1 early in dengue illness correlate with the development of dengue hemorrhagic fever*. Journal of Infectious Diseases, 2002. **186**(8): p. 1165-1168.
- [26.] Akey, D.L., et al., *Flavivirus NS1 structures reveal surfaces for associations with membranes and the immune system*. Science, 2014. **343**(6173): p. 881-885.
- [27.] Modhiran, N., et al., *Dengue virus NS1 protein activates cells via Toll-like receptor 4 and disrupts endothelial cell monolayer integrity*. Science translational medicine, 2015. **7**(304): p. 304-142.

- [28.] Xie, X., et al., *Membrane topology and function of dengue virus NS2A protein*. Journal of virology, 2013. **87**(8): p. 4609-4622.
- [29.] Muñoz-Jordán, J.L., et al., *Inhibition of interferon signaling by dengue virus*. Proceedings of the National Academy of Sciences, 2003. **100**(24): p. 14333-14338.
- [30.] Erbel, P., et al., *Structural basis for the activation of flaviviral NS3 proteases from dengue and West Nile virus*. Nature structural & molecular biology, 2006. **13**(4): p. 372-373.
- [31.] Lescar, J., et al., *Towards the design of antiviral inhibitors against flaviviruses: the case for the multifunctional NS3 protein from Dengue virus as a target*. Antiviral research, 2008. **80**(2): p. 94-101.
- [32.] Miller, S., et al., *The non-structural protein 4A of dengue virus is an integral membrane protein inducing membrane alterations in a 2K-regulated manner*. Journal of Biological Chemistry, 2007. **282**(12): p. 8873-8882.
- [33.] Umareddy, I., et al., *Dengue virus NS4B interacts with NS3 and dissociates it from single-stranded RNA*. Journal of general virology, 2006. **87**(9): p. 2605-2614.
- [34.] Zhao, Y., et al., *A crystal structure of the dengue virus NS5 protein reveals a novel inter-domain interface essential for protein flexibility and virus replication*. PLoS Pathog, 2015. **11**(3): p. e1004682.
- [35.] Halstead, S.B., *Controversies in dengue pathogenesis*. Paediatrics and international child health, 2012. **32**(sup1): p. 5-9.
- [36.] Murray, N.E.A., M.B. Quam, and A. Wilder-Smith, *Epidemiology of dengue: past, present and future prospects*. 2013. **4**(2): p. 119-133.
- [37.] Wong, L.P., S. AbuBakar, and K. Chinna, *Community knowledge, health beliefs, practices and experiences related to dengue fever and its association with IgG seropositivity*. PLoS Negl Trop Dis, 2014. **8**(5): p. e2789.
- [38.] Organization, W.H., *Update on the dengue situation in the Western Pacific Region*. Update, 2014(455) **4**(2): p. 119-133.
- [39.] Kittigul, L., et al., *The differences of clinical manifestations and laboratory findings in children and adults with dengue virus infection*. Journal of Clinical Virology, 2007. **39**(2): p. 76-81.
- [40.] Ishak, I.H., et al., *Contrasting patterns of insecticide resistance and knockdown resistance (kdr) in the dengue vectors Aedes aegypti and Aedes albopictus from Malaysia*. Parasites & vectors, 2015. **8**(1): p. 181.

- [41.] SANGKAWIBHA, N., et al., *Risk factors in dengue shock syndrome: a prospective epidemiologic study in Rayong, Thailand I. The 1980 outbreak*. American journal of epidemiology, 1984. **120**(5): p. 653-669.
- [42.] Guzmán, M.G., et al., *Epidemiologic studies on Dengue in Santiago de Cuba, 1997*. American journal of epidemiology, 2000. **152**(9): p. 793-799.
- [43.] Guzman, M.G. and E. Harris, *Dengue*. The Lancet, 2015. **385**(9966): p. 453-465.
- [44.] Benelli, G. and H. Mehlhorn, *Declining malaria, rising of dengue and Zika virus: insights for mosquito vector control*. Parasitology research, 2016. **115**(5): p. 1747-1754.
- [45.] Elias, M., et al., *Biological control of mosquito larvae by Guppy fish*. Bangladesh Medical Research Council Bulletin, 1995. **21**(2): p. 81-86.
- [46.] Iturbe-Ormaetxe, I., T. Walker, and S. LO'Neill, *Wolbachia and the biological control of mosquito-borne disease*. EMBO reports, 2011. **12**(6): p. 508-518.
- [47.] Takada, A. and Y. Kawaoka, *Antibody-dependent enhancement of viral infection: molecular mechanisms and in vivo implications*. Reviews in medical virology, 2003. **13**(6): p. 387-398.
- [48.] Mairuhu, A., et al., *Dengue: an arthropod-borne disease of global importance*. European journal of clinical microbiology and infectious diseases, 2004. **23**(6): p. 425-433.
- [49.] Beesetti, H., N. Khanna, and S. Swaminathan, *Drugs for dengue: a patent review (2010–2014)*. Expert opinion on therapeutic patents, 2014. **24**(11): p. 1171-1184.
- [50.] Kuno, G., *Emergence of the severe syndrome and mortality associated with dengue and dengue-like illness: historical records (1890 to 1950) and their compatibility with current hypotheses on the shift of disease manifestation*. Clinical microbiology reviews, 2009. **22**(2): p. 186-201.
- [51.] Whitehorn, J., et al., *Dengue therapeutics, chemoprophylaxis, and allied tools: state of the art and future directions*. PLoS Negl Trop Dis, 2014. **8**(8): p. e3025.
- [52.] Blacksell, S.D., et al., *Evaluation of six commercial point-of-care tests for diagnosis of acute dengue infections: the need for combining NSI antigen and IgM/IgG antibody detection to achieve acceptable levels of accuracy*. Clinical and Vaccine Immunology, 2011. **18**(12): p. 2095-2101.
- [53.] Rothman, A.L., *Immunity to dengue virus: a tale of original antigenic sin and tropical cytokine storms*. Nature reviews. Immunology, 2011. **11**(8): p. 532.



- [54.] Póvoa, T.F., et al., *The pathology of severe dengue in multiple organs of human fatal cases: histopathology, ultrastructure and virus replication*. PLoS one, 2014. **9**(4): p. e83386.
- [55.] Limonta, D., et al., *Dengue virus identification by transmission electron microscopy and molecular methods in fatal dengue hemorrhagic fever*. Infection, 2012. **40**(6): p. 689-694.
- [56.] Alomar, M.J., *Factors affecting the development of adverse drug reactions*. Saudi Pharmaceutical Journal, 2014. **22**(2): p. 83-94.
- [57.] Tricou, V., et al., *A randomized controlled trial of chloroquine for the treatment of dengue in Vietnamese adults*. PLoS Negl Trop Dis, 2010. **4**(8): p. e785.
- [58.] Tam, D.T., et al., *Effects of short-course oral corticosteroid therapy in early dengue infection in Vietnamese patients: a randomized, placebo-controlled trial*. Clinical infectious diseases, 2012: p. cis655.
- [59.] Whitehorn, J., et al., *Lovastatin for adult patients with dengue: protocol for a randomised controlled trial*. Trials, 2012. **13**(1): p. 203.
- [60.] Low, J.G., et al., *Efficacy and safety of celgosivir in patients with dengue fever (CELADEN): a phase 1b, randomised, double-blind, placebo-controlled, proof-of-concept trial*. The Lancet infectious diseases, 2014. **14**(8): p. 706-715.
- [61.] Nguyen, N.M., et al., *A randomized, double-blind placebo controlled trial of balapiravir, a polymerase inhibitor, in adult dengue patients*. Journal of Infectious Diseases, 2013. **207**(9): p. 1442-1450.
- [62.] Rajapakse, S., et al., *Corticosteroids in the treatment of dengue shock syndrome*. Infection and drug resistance, 2014. **7**: p. 137.
- [63.] Salgado, D., et al., *Use of pentoxifylline in treatment of children with dengue hemorrhagic fever*. The Pediatric infectious disease journal, 2012. **31**(7): p. 771-773.
- [64.] Frias-Staheli, N., et al., *Utility of humanized BLT mice for analysis of dengue virus infection and antiviral drug testing*. Journal of virology, 2014. **88**(4): p. 2205-2218.
- [65.] Pugach, P., et al., *Neutralizing antibody and anti-retroviral drug sensitivities of HIV-1 isolates resistant to small molecule CCR5 inhibitors*. Virology, 2008. **377**(2): p. 401-407.
- [66.] Lalezari, J.P., et al., *Enfuvirtide, an HIV-1 fusion inhibitor, for drug-resistant HIV infection in North and South America*. New England Journal of Medicine, 2003. **348**(22): p. 2175-2185.
- [67.] Modis, Y., et al., *Structure of the dengue virus envelope protein after membrane fusion*. Nature, 2004. **427**(6972): p. 313-319.

- [68.] De La Guardia, C. and R. Lleonart, *Progress in the identification of dengue virus entry/fusion inhibitors*. BioMed research international, 2014. **2014**.
- [69.] Watterson, D., B. Kobe, and P.R. Young, *Residues in domain III of the dengue virus envelope glycoprotein involved in cell-surface glycosaminoglycan binding*. Journal of General Virology, 2012. **93**(1): p. 72-82.
- [70.] Butrapet, S., et al., *Amino acid changes within the E protein hinge region that affect dengue virus type 2 infectivity and fusion*. Virology, 2011. **413**(1): p. 118-127.
- [71.] Hrobowski, Y.M., R.F. Garry, and S.F. Michael, *Peptide inhibitors of dengue virus and West Nile virus infectivity*. Virology journal, 2005. **2**(1): p. 49.
- [72.] Lok, S.-M., et al., *Release of dengue virus genome induced by a peptide inhibitor*. PLoS One, 2012. **7**(11): p. e50995.
- [73.] Schmidt, A.G., P.L. Yang, and S.C. Harrison, *Peptide inhibitors of flavivirus entry derived from the E protein stem*. Journal of virology, 2010. **84**(24): p. 12549-12554.
- [74.] Modis, Y., et al., *A ligand-binding pocket in the dengue virus envelope glycoprotein*. Proceedings of the National Academy of Sciences, 2003. **100**(12): p. 6986-6991.
- [75.] Kaptein, S.J., et al., *A derivate of the antibiotic doxorubicin is a selective inhibitor of dengue and yellow fever virus replication in vitro*. Antimicrobial agents and chemotherapy, 2010. **54**(12): p. 5269-5280.
- [76.] Wang, Q.-Y., et al., *A small-molecule dengue virus entry inhibitor*. Antimicrobial agents and chemotherapy, 2009. **53**(5): p. 1823-1831.
- [77.] Liao, M. and M. Kielian, *Domain III from class II fusion proteins functions as a dominant-negative inhibitor of virus membrane fusion*. J Cell Biol, 2005. **171**(1): p. 111-120.
- [78.] Alhoot, M.A., et al., *Inhibition of dengue virus entry into target cells using synthetic antiviral peptides*. International journal of medical sciences, 2013. **10**(6): p. 719-729.
- [79.] Hidari, K.I., et al., *Structure and anti-dengue virus activity of sulfated polysaccharide from a marine alga*. Biochemical and biophysical research communications, 2008. **376**(1): p. 91-95.
- [80.] Chen, Y., et al., *Dengue virus infectivity depends on envelope protein binding to target cell heparan sulfate*. Nature medicine, 1997. **3**(8): p. 866-871.

- [81.] Talarico, L., et al., *The antiviral activity of sulfated polysaccharides against dengue virus is dependent on virus serotype and host cell*. Antiviral research, 2005. **66**(2): p. 103-110.
- [82.] Vervaeke, P., et al., *Sulfated Escherichia coli K5 polysaccharide derivatives inhibit dengue virus infection of human microvascular endothelial cells by interacting with the viral envelope protein E domain III*. PLoS One, 2013. **8**(8): p. e74035.
- [83.] Lin, L.-T., et al., *Broad-spectrum antiviral activity of chebulagic acid and punicalagin against viruses that use glycosaminoglycans for entry*. BMC microbiology, 2013. **13**(1): p. 187.
- [84.] Premaratna, R., et al., *A clinical guide for early detection of dengue fever and timing of investigations to detect patients likely to develop complications* ☆. Transactions of the Royal Society of Tropical Medicine and Hygiene, 2009. **103**(2): p. 127-131.
- [85.] Raman, N., S. Johnson Raja, and A. Sakthivel, *Transition metal complexes with Schiff-base ligands: 4-aminoantipyrine based derivatives—a review*. Journal of Coordination Chemistry, 2009. **62**(5): p. 691-709.
- [86.] Prakash, A. and D. Adhikari, *Application of Schiff bases and their metal complexes-A Review*. Int. J. Chem. Tech. Res, 2011. **3**(4): p. 1891-1896.
- [87.] Anwer, J., et al., *Synthesis, characterization, semi-empirical study and biological activities of homobimetallic complexes of tranexamic acid with organotin (IV)*. Journal of Coordination Chemistry, 2013. **66**(7): p. 1142-1152.
- [88.] Tian, Y.-P., et al., *Crystal structures, spectroscopic and nonlinear optical properties of metal complexes of Schiff-base ligands containing nitrogen and sulfur donors*. Transition metal chemistry, 1997. **23**(1): p. 17-20.
- [89.] Low, M.L., *Synthesis, characterization and bioactivities of dithiocarbamate Schiff base ligands and their metal complexes*. 2014, Université Pierre et Marie Curie-Paris VI.
- [90.] Kumar, S., D.N. Dhar, and P. Saxena, *Applications of metal complexes of Schiff bases-A review*. 2009.
- [91.] Tarafder, M., et al., *Coordination chemistry and bioactivity of some metal complexes containing two isomeric bidentate NS Schiff bases derived from S-benzylthiocarbamate and the X-ray crystal structures of S-benzyl-β-N-(5-methyl-2-furylmethylene) dithiocarbamate and bis [S-benzyl-β-N-(2-furylmethylketone) dithiocarbamate] cadmium (II)*. Polyhedron, 2002. **21**(27): p. 2691-2698.

- [92.] Maia, P.I.d.S., et al., *Dithiocarbazate complexes with the [M (PPh 3)]<sup>2+</sup> (M= Pd or Pt) moiety: Synthesis, characterization and anti-Tripanosoma cruzi activity*. Journal of inorganic biochemistry, 2010. **104**(12): p. 1276-1282.
- [93.] Ravoof, T.B., et al., *Synthesis, characterization and bioactivity of mixed-ligand Cu (II) complexes containing Schiff bases derived from S-benzylthiocarbazate and saccharinate ligand and the X-ray crystal structure of the copper-saccharinate complex containing S-benzyl-β-N-(acetylpyrid-2-yl) methylenedithiocarbazate*. Polyhedron, 2007. **26**(6): p. 1159-1165.
- [94.] Manan, M.A.F.A., et al., *The crystal structure and cytotoxicity of centrosymmetric copper (II) complex derived from S-methylthiocarbazate with isatin*. Journal of Chemical Crystallography, 2011. **41**(12): p. 1866-1871.
- [95.] Manan, M.A.F.A., et al., *Synthesis, characterization and cytotoxic activity of S-benzylthiocarbazate Schiff bases derived from 5-fluoroisatin, 5-chloroisatin, 5-bromoisatin and their crystal structures*. Journal of Chemical Crystallography, 2011. **41**(11): p. 1630.
- [96.] Tarafder, M.T.H., et al., *Complexes of a tridentate ONS Schiff base. Synthesis and biological properties*. Transition Metal Chemistry, 2000. **25**(4): p. 456-460.
- [97.] Zhang, L., et al., *Biological activities of pyridine-2-carbaldehyde Schiff bases derived from S-methyl- and S-benzylthiocarbazate and their zinc (II) and manganese (II) complexes. Crystal Structure of the Manganese (II) complex of pyridine-2-carbaldehyde S-benzylthiocarbazate*. Russian Journal of Coordination Chemistry, 2011. **37**(5): p. 356-361.
- [98.] Liu, Y.-T., et al., *Synthesis and antimicrobial activity of some novel ferrocene-based Schiff bases containing a ferrocene unit*. Research on Chemical Intermediates, 2012. **38**(3-5): p. 1043-1053.
- [99.] Liu, Y.-T., et al., *Synthesis, characterization and biological activity of ferrocene-based Schiff base ligands and their metal (II) complexes*. Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy, 2013. **100**: p. 131-137.
- [100.] Carrillo-Infante, C., et al., *Viral infections as a cause of cancer (review)*. International journal of oncology, 2007. **30**(6): p. 1521.
- [101.] McAllister, R.M., *Viruses and Cancer—A Review*. California medicine, 1965. **102**(5): p. 344.
- [102.] Sartorelli, A.C., *Effect of chelating agents upon the synthesis of nucleic acids and protein: inhibition of DNA synthesis by 1-formylisoquinoline thiosemicarbazone*. Biochemical and biophysical research communications, 1967. **27**(1): p. 26-32.

- [103.] Ali, M.A. and S. Livingstone, *Metal complexes of sulphur-nitrogen chelating agents*. Coordination Chemistry Reviews, 1974. **13**(2-3): p. 101-132.
- [104.] McConathy, J. and M.J. Owens, *Stereochemistry in drug action*. Prim Care Companion J Clin Psychiatry, 2003. **5**(2): p. 70-73.
- [105.] ANDERSON, R.J. and A. TODD, *Stereochemistry and drug action*. Pharmaceutical Chemistry, 2013. **4**(2): p. 119-133.
- [106.] Hutt, A.J., *The development of single-isomer molecules: why and how*. CNS spectrums, 2002. **7**(S1): p. 14-22.
- [107.] Lima, J.J. and H. Boudoulas, *Stereoselective effects of disopyramide enantiomers in humans*. Journal of cardiovascular pharmacology, 1987. **9**(5): p. 594-600.
- [108.] Banitt, E.H., J.R. Schmid, and R.A. Newmark, *Resolution of flecainide acetate, N-(2-piperidylmethyl)-2, 5-bis (2, 2, 2-trifluoroethoxy) benzamide acetate, and antiarrhythmic properties of the enantiomers*. Journal of medicinal chemistry, 1986. **29**(2): p. 299-302.
- [109.] Hughes, A., S. Hering, and T. Bolton, *Evidence that agonist and antagonist enantiomers of the dihydropyridine PN 202-791 act at different sites on the voltage-dependent calcium channel of vascular muscle*. British journal of pharmacology, 1990. **101**(1): p. 3-5.
- [110.] Mannhold, R., et al., *Molecular biology in medicinal chemistry*. Vol. 21. 2006: John Wiley & Sons **4**(2): p. 119-133..
- [111.] Kato, R., et al., *Electrophysiologic effects of the levo-and dextrorotatory isomers of sotalol in isolated cardiac muscle and their in vivo pharmacokinetics*. Journal of the American College of Cardiology, 1986. **7**(1): p. 116-125.
- [112.] Ruffolo, R. and K. Messick, *Effects of dopamine, (+/-)-dobutamine and the (+)-and (-)-enantiomers of dobutamine on cardiac function in pithed rats*. Journal of Pharmacology and Experimental Therapeutics, 1985. **235**(3): p. 558-565.
- [113.] Yuan, R. and Y. Lin, *Traditional Chinese medicine: an approach to scientific proof and clinical validation*. Pharmacology & therapeutics, 2000. **86**(2): p. 191-198.
- [114.] Kadir, S.L.A., H. Yaakob, and R.M. Zulkifli, *Potential anti-dengue medicinal plants: a review*. Journal of natural medicines, 2013. **67**(4): p. 677-689.
- [115.] Mukhtar, M., et al., *Antiviral potentials of medicinal plants*. Virus research, 2008. **131**(2): p. 111-120.

- [116.] Newman, D.J. and G.M. Cragg, *Natural Products as Sources of New Drugs over the Last 25 Years*. Journal of natural products, 2007. **70**(3): p. 461-477.
- [117.] Canard, B., *Antiviral research and development against dengue virus*. Available: Accessed, 2012. **9**.
- [118.] Baell, J. and M.A. Walters, *Chemical con artists foil drug discovery*. Nature, 2014. **513**(7519): p. 481.
- [119.] Dragland, S., et al., *Several culinary and medicinal herbs are important sources of dietary antioxidants*. The Journal of nutrition, 2003. **133**(5): p. 1286-1290.
- [120.] Bensky, D., A. Gamble, and T.J. Kaptchuk, *Chinese herbal medicine: materia medica*. 1993: Eastland Press.
- [121.] Dharmananda, S., *The Nature of Ginseng from Traditional Use to Modern Research*. 2002: ITM.
- [122.] Chang, S.-H., et al., *Dryopteris crassirhizoma has anti-cancer effects through both extrinsic and intrinsic apoptotic pathways and G0/G1 phase arrest in human prostate cancer cells*. Journal of ethnopharmacology, 2010. **130**(2): p. 248-254.
- [123.] Byung-Sun, M., et al., *Kaempferol acetylramnosides from the rhizome of Dryopteris crassirhizoma and their inhibitory effects on three different activities of human immunodeficiency virus-1 reverse transcriptase*. Chemical and pharmaceutical bulletin, 2001. **49**(5): p. 546-550.
- [124.] Bensky, D. and R. Barolet, *Formulas and Strategies*. Eastland, Seattle, 1990.
- [125.] Tsai, H.-H., S.-M. Hwang, and P.-C. Kung, *Use of plant extracts for treatment of HIV, HCV and HBV infections*. 1998, Google Patents.
- [126.] Han, J., X. Weng, and K. Bi, *Antioxidants from a Chinese medicinal herb—Lithospermum erythrorhizon*. Food chemistry, 2008. **106**(1): p. 2-10.
- [127.] Chen, X., et al., *Shikonin, a component of Chinese herbal medicine, inhibits chemokine receptor function and suppresses human immunodeficiency virus type 1*. Antimicrobial agents and chemotherapy, 2003. **47**(9): p. 2810-2816.
- [128.] Mei, M., et al., *In vitro pharmacokinetic characterization of mulberroside A, the main polyhydroxylated stilbene in mulberry (Morus alba L.), and its bacterial metabolite oxyresveratrol in traditional oral use*. Journal of agricultural and food chemistry, 2012. **60**(9): p. 2299-2308.

- [129.] Ma, Y., et al., *Establishment of an Anti-HIV/AIDS Agents Screening Technique Based on Human CXCR4 Promoter*. LifeScienceJournal, 2005. **7**: p. 1.
- [130.] Park, S.-D., Y.-S. Lai, and C.-H. Kim, *Immunopotentiating and antitumor activities of the purified polysaccharides from Phellodendron chinese SCHNEID*. Life sciences, 2004. **75**(22): p. 2621-2632.
- [131.] Kim, J.-H., et al., *Inhibitory effects of an aqueous extract from Cortex Phellodendri on the growth and replication of broad-spectrum of viruses in vitro and in vivo*. BMC Complementary and Alternative Medicine, 2016. **16**(1): p. 265.
- [132.] Yang, F., et al., *Preparative isolation and purification of hydroxyanthraquinones from Rheum officinale Baill by high-speed counter-current chromatography using pH-modulated stepwise elution*. Journal of Chromatography A, 1999. **858**(1): p. 103-107.
- [133.] Cai, Y., et al., *Antioxidant phenolic constituents in roots of Rheum officinale and Rubia cordifolia: structure– radical scavenging activity relationships*. Journal of agricultural and food chemistry, 2004. **52**(26): p. 7884-7890.
- [134.] Wang, Z., et al., *Anti-herpes virus action of ethanol-extract from the root and rhizome of Rheum officinale Baill*. China journal of Chinese materia medica, 1996. **21**(6): p. 364-366.
- [135.] Xu, C., L. Luo, and R.X. Tan, *Antidepressant effect of three traditional Chinese medicines in the learned helplessness model*. Journal of ethnopharmacology, 2004. **91**(2): p. 345-349.
- [136.] Díaz, A.M.a., et al., *Phenylpropanoid glycosides from Scrophularia scorodonia: in vitro anti-inflammatory activity*. Life sciences, 2004. **74**(20): p. 2515-2526.
- [137.] Gao, Y., et al., *Mitochondrial apoptosis contributes to the anti-cancer effect of Smilax glabra Roxb*. Toxicology letters, 2011. **207**(2): p. 112-120.
- [138.] Sa, F., et al., *Anti-proliferative and pro-apoptotic effect of Smilax glabra Roxb. extract on hepatoma cell lines*. Chemico-biological interactions, 2008. **171**(1): p. 1-14.
- [139.] Xie, C., et al., *Flavone C-glycosides from Viola yedoensis Makino*. Chemical and pharmaceutical bulletin, 2003. **51**(10): p. 1204-1207.
- [140.] Chang, R.S. and H. Yeung, *Inhibition of growth of human immunodeficiency virus in vitro by crude extracts of Chinese medicinal herbs*. Antiviral research, 1988. **9**(3): p. 163-175.

- [141.] Wang, C.K., et al., *Anti-HIV cyclotides from the Chinese medicinal herb Viola yedoensis*. Journal of natural products, 2007. **71**(1): p. 47-52.
- [142.] Okhwarobo, A., et al., *Harnessing the medicinal properties of Andrographis paniculata for diseases and beyond: a review of its phytochemistry and pharmacology*. Asian Pacific Journal of Tropical Disease, 2014. **4**(3): p. 213-222.
- [143.] Tang, L.I., et al., *Screening of anti-dengue activity in methanolic extracts of medicinal plants*. BMC complementary and alternative medicine, 2012. **12**(1): p. 1.
- [144.] Steele, R.L., et al., *Artemisinin-based combination therapy for treating viral mediated disease*. 2014, Google Patents **4**(2): p. 119-133..
- [145.] Rangarajan, P.N., *The 2015 Nobel Prize in physiology or medicine*. Resonance, 2016. **21**(4): p. 315-326.
- [146.] Eng-Chong, T., et al., *Boesenbergia rotunda: from ethnomedicine to drug discovery*. Evidence-Based Complementary and Alternative Medicine, 2012. **204**(2): p. 119-133.
- [147.] Kiat, T.S., et al., *Inhibitory activity of cyclohexenyl chalcone derivatives and flavonoids of fingerroot, Boesenbergia rotunda (L.), towards dengue-2 virus NS3 protease*. Bioorganic & medicinal chemistry letters, 2006. **16**(12): p. 3337-3340.
- [148.] Shah, G., et al., *Scientific basis for the therapeutic use of Cymbopogon citratus, stapf (Lemon grass)*. Journal of advanced pharmaceutical technology & research, 2011. **2**(1): p. 3.
- [149.] Huang, L., S. Chen, and M. Yang, *Euphorbia hirta (Feiyangcao): A review on its ethnopharmacology, phytochemistry and pharmacology*. Journal of Medicinal Plants Research, 2012. **6**(39): p. 5176-5185.
- [150.] Srivastava, M. and V. Kapoor, *Seed galactomannans: an overview*. Chemistry & Biodiversity, 2005. **2**(3): p. 295-317.
- [151.] Ono, L., et al., *In vitro and in vivo antiviral properties of sulfated galactomannans against yellow fever virus (BeH111 strain) and dengue 1 virus (Hawaii strain)*. Antiviral research, 2003. **60**(3): p. 201-208.
- [152.] Bensky, D. and R. Barolet, *Chinese Herbal Medicine: Formulas & Strategies (1990)*. Eastland Press, Inc., Seattle, Washington.
- [153.] Lee, Y.-R., et al., *Honeysuckle aqueous extract and induced let-7a suppress dengue virus type 2 replication and pathogenesis*. Journal of Ethnopharmacology, 2017. **4**(2): p. 119-133.



- [154.] Grover, J. and S. Yadav, *Pharmacological actions and potential uses of Momordica charantia: a review*. Journal of ethnopharmacology, 2004. **93**(1): p. 123-132.
- [155.] Jassim, S. and M.A. Naji, *Novel antiviral agents: a medicinal plant perspective*. Journal of Applied Microbiology, 2003. **95**(3): p. 412-427.
- [156.] Gutiérrez, R.M.P., S. Mitchell, and R.V. Solis, *Psidium guajava: a review of its traditional uses, phytochemistry and pharmacology*. Journal of ethnopharmacology, 2008. **117**(1): p. 1-27.
- [157.] Jadhav, R. and B. Jadhav, *Evaluation of Antimicrobial principles of Rhizophora species along Mumbai Coast*. Journal of Advanced Scientific Research, 2012. **34**(2): p. 119-133.
- [158.] Smither, S.J., et al., *Comparison of the plaque assay and 50% tissue culture infectious dose assay as methods for measuring filovirus infectivity*. Journal of virological methods, 2013. **193**(2): p. 565-571.
- [159.] Medina, F., et al., *Dengue virus: isolation, propagation, quantification, and storage*. Current protocols in microbiology, 2012: p. 15D. 2.1-15D. 2.24.
- [160.] Yang, C.-F., et al., *Surveillance and Molecular Characterization on Dengue Viruses in Taiwan, 2013*. Epidemiology Bulletin, 2014. **30**(19): p. 157-170.
- [161.] Tang, L.I., et al., *Screening of anti-dengue activity in methanolic extracts of medicinal plants*. BMC complementary and alternative medicine, 2012. **12**(1): p. 3.
- [162.] Merquiol, E., et al., *HCV causes chronic endoplasmic reticulum stress leading to adaptation and interference with the unfolded protein response*. PLoS one, 2011. **6**(9): p. e24660.
- [163.] Lindenbach, B.D., *Measuring HCV infectivity produced in cell culture and in vivo*. Hepatitis C: Methods and Protocols, 2009. **4**(2): p. 119-133..
- [164.] Zandi, K., et al., *Extract of Scutellaria baicalensis inhibits dengue virus replication*. BMC complementary and alternative medicine, 2013. **13**(1): p. 91.
- [165.] Kudi, A. and S. Myint, *Antiviral activity of some Nigerian medicinal plant extracts*. Journal of ethnopharmacology, 1999. **68**(1): p. 289-294.
- [166.] Zandi, K., et al., *Novel antiviral activity of baicalein against dengue virus*. BMC complementary and alternative medicine, 2012. **12**(1): p. 1.
- [167.] Zandi, K., et al., *In vitro antiviral activity of fisetin, rutin and naringenin against dengue virus type-2*. Journal of Medicinal Plants Research, 2011. **5**(23): p. 119-133..

- [168.] Zandi, K., et al., *Novel antiviral activity of baicalein against dengue virus*. BMC complementary and alternative medicine, 2012. **12**(1): p. 214.
- [169.] Moghaddam, E., et al., *Baicalin, a metabolite of baicalein with antiviral activity against dengue virus*. Scientific reports, 2014. **4**: p. 5452.
- [170.] Salentin, S., et al., *PLIP: fully automated protein–ligand interaction profiler*. Nucleic acids research, 2015. **43**(W1): p. W443-W447.
- [171.] Lambeth, C.R., *Interactions between dengue type 3 viruses and human dendritic cells*. University of North Carolina, 2007. **4**(2): p. 119-133..
- [172.] Medina-Franco, J.L., et al., *Shifting from the single to the multitarget paradigm in drug discovery*. Drug discovery today, 2013. **18**(9): p. 495-501.
- [173.] Dang, V.T., K. Benkendorff, and P. Speck, *In vitro antiviral activity against herpes simplex virus in the abalone *Haliotis laevigata**. Journal of General Virology, 2011. **92**(3): p. 627-637.
- [174.] Chu, J. and P.L. Yang, *c-Src protein kinase inhibitors block assembly and maturation of dengue virus*. Proceedings of the National Academy of Sciences, 2007. **104**(9): p. 3520-3525.
- [175.] Cheng, H.-J., et al., *Anti-dengue virus nonstructural protein 1 antibodies recognize protein disulfide isomerase on platelets and inhibit platelet aggregation*. Molecular immunology, 2009. **47**(2): p. 398-406.
- [176.] Falkler, W., A. Diwan, and S. Halstead, *A lipid inhibitor of dengue virus in human colostrum and milk; with a note on the absence of anti-dengue secretory antibody*. Archives of Virology, 1975. **47**(1): p. 3-10.
- [177.] Kirschner, S., et al., *Anticancer and potential antiviral activity of complex inorganic compounds*. Journal of medicinal chemistry, 1966. **9**(3): p. 369-372.
- [178.] MOHAMED, G.G., M.M. Omar, and A.M. Hindy, *Metal complexes of Schiff bases: preparation, characterization, and biological activity*. Turkish Journal of Chemistry, 2006. **30**(3): p. 361-382.
- [179.] Mehvar, R., D.R. Brocks, and M. Vakily, *Impact of stereoselectivity on the pharmacokinetics and pharmacodynamics of antiarrhythmic drugs*. Clinical pharmacokinetics, 2002. **41**(8): p. 533-558.
- [180.] McConathy, J. and M.J. Owens, *Stereochemistry in drug action*. Primary care companion to the Journal of clinical psychiatry, 2003. **5**(2): p. 70.

- [181.] Niles, A.L., R.A. Moravec, and T.L. Riss, *Update on in vitro cytotoxicity assays for drug development*. Expert opinion on drug discovery, 2008. **3**(6): p. 655-669.
- [182.] Ren, S., et al., *Synthesis, biological evaluation, and quantitative structure– activity relationship analysis of new Schiff bases of hydroxysemicarbazide as potential antitumor agents*. Journal of medicinal chemistry, 2002. **45**(2): p. 410-419.
- [183.] Leandro, L.M., et al., *Chemistry and biological activities of terpenoids from copaiba (Copaifera spp.) oleoresins*. Molecules, 2012. **17**(4): p. 3866-3889.
- [184.] Bakkali, F., et al., *Biological effects of essential oils—a review*. Food and chemical toxicology, 2008. **46**(2): p. 446-475.
- [185.] Low, M.L., et al., *Conjugation of a new series of dithiocarbazate Schiff base copper (II) complexes with vectors selected to enhance antibacterial activity*. Bioconjugate chemistry, 2014. **25**(12): p. 2269-2284.
- [186.] Torrentes-Carvalho, A., et al., *Dengue-2 infection and the induction of apoptosis in human primary monocytes*. Memorias do Instituto Oswaldo Cruz, 2009. **104**(8): p. 1091-1099.
- [187.] Allard, P.-M., et al., *Alkylated flavanones from the bark of Cryptocarya chartacea as dengue virus NS5 polymerase inhibitors*. Journal of natural products, 2011. **74**(11): p. 2446-2453.
- [188.] Simões, L., et al., *Antiviral activity of Distictella elongata (Vahl) Urb.(Bignoniaceae), a potentially useful source of anti-dengue drugs from the state of Minas Gerais, Brazil*. Letters in applied microbiology, 2011. **53**(6): p. 602-607.
- [189.] Yin Low, J., et al., *Antiviral activity of emetine dihydrochloride against dengue virus infection*. J Antivir Antiretrovir, 2009. **1**: p. 062-000.
- [190.] Zhang, X.G., et al., *Antiviral activity of geneticin against dengue virus*. Antiviral research, 2009. **83**(1): p. 21-27.
- [191.] OhAinle, M., et al., *Dynamics of dengue disease severity determined by the interplay between viral genetics and serotype-specific immunity*. Science translational medicine, 2011. **3**(114): p. 114ra128-114ra128.
- [192.] Xu, M., et al., *A potent neutralizing antibody with therapeutic potential against all four serotypes of dengue virus*. npj Vaccines, 2017. **2**(1): p. 2.
- [193.] Yung, C.-F., et al., *Dengue serotype-specific differences in clinical manifestation, laboratory parameters and risk of severe disease in adults, Singapore*. The American journal of tropical medicine and hygiene, 2015. **92**(5): p. 999-1005.

- [194.] Keivan, Z., et al., *In vitro antiviral activity of fisetin, rutin and naringenin against dengue virus type-2*. Journal of Medicinal Plants Research, 2011. **5**(23): p. 5534-5539.
- [195.] Moghaddam, E., et al., *Baicalin, a metabolite of baicalein with antiviral activity against dengue virus*. Scientific reports, 2014. **4**.
- [196.] Flingai, S., et al., *Protection against dengue disease by synthetic nucleic acid antibody prophylaxis/immunotherapy*. Scientific reports, 2015. **5**.
- [197.] Food and D. Administration, *Guidance for Industry; Antiviral Product Development—Conducting and Submitting Virology Studies to the Agency*. Department of Health and Human Services, FDA June, 2006.
- [198.] Hung, S.-L., et al., *Analysis of the steps involved in dengue virus entry into host cells*. Virology, 1999. **257**(1): p. 156-167.
- [199.] Patil, R., et al., *Optimized hydrophobic interactions and hydrogen bonding at the target-ligand interface leads the pathways of drug-designing*. PloS one, 2010. **5**(8): p. e12029.
- [200.] Hansch, C. and T.E. Klein, [24] *Quantitative structure-activity relationships and molecular graphics in evaluation of enzyme-ligand interactions*, in *Methods in enzymology*. 1991, Elsevier. p. 512-543.
- [201.] Turner, J.V., D.J. Maddalena, and D.J. Cutler, *Pharmacokinetic parameter prediction from drug structure using artificial neural networks*. International journal of pharmaceutics, 2004. **270**(1-2): p. 209-219.
- [202.] Davis, A.M. and S.J. Teague, *Hydrogen bonding, hydrophobic interactions, and failure of the rigid receptor hypothesis*. Angewandte Chemie International Edition, 1999. **38**(6): p. 736-749.
- [203.] Panigrahi, S.K., *Strong and weak hydrogen bonds in protein-ligand complexes of kinases: a comparative study*. Amino Acids, 2008. **34**(4): p. 617-633.
- [204.] Kahraman, A., et al., *Shape variation in protein binding pockets and their ligands*. Journal of molecular biology, 2007. **368**(1): p. 283-301.
- [205.] Oliveira, A.S.d., et al., *NS3 and NS5 proteins: important targets for anti-dengue drug design*. Journal of the Brazilian Chemical Society, 2014. **25**(10): p. 1759-1769.
- [206.] Patkar, C.G. and R.J. Kuhn, *Yellow fever virus NS3 plays an essential role in virus assembly independent of its known enzymatic functions*. Journal of virology, 2008. **82**(7): p. 3342-3352.

- [207.] Ma, Y., et al., *NS3 helicase domains involved in infectious intracellular hepatitis C virus particle assembly*. Journal of virology, 2008. **82**(15): p. 7624-7639.
- [208.] Chiang, L.C., et al., *Antiviral activities of extracts and selected pure constituents of Ocimum basilicum*. Clinical and Experimental Pharmacology and Physiology, 2005. **32**(10): p. 811-816.
- [209.] Rasoanaivo, P., et al., *Whole plant extracts versus single compounds for the treatment of malaria: synergy and positive interactions*. Malaria Journal, 2011. **10**(1): p. S4.
- [210.] Mueller, M.S., et al., *The potential of Artemisia annua L. as a locally produced remedy for malaria in the tropics: agricultural, chemical and clinical aspects*. Journal of ethnopharmacology, 2000. **73**(3): p. 487-493.
- [211.] Gupta, O., *Nobel Prize 2015*. Journal of Mahatma Gandhi Institute of Medical Sciences, 2016. **21**(1): p. 75-75.
- [212.] Steele, R.L. and A.F. Musso, *Artemisinin-based combination therapy for treating viral mediated disease*. 2012, Google Patents.
- [213.] Lee, Y.-R., et al., *Honeysuckle aqueous extract and induced let-7a suppress dengue virus type 2 replication and pathogenesis*. Journal of ethnopharmacology, 2017. **198**: p. 109-121.
- [214.] Huang, K.C., *The pharmacology of Chinese herbs*. 1998: CRC press.
- [215.] Zheng, L., et al., *Inhibition of porcine reproductive and respiratory syndrome virus replication in vitro using DNA-based short antisense oligonucleotides*. BMC veterinary research, 2015. **11**(1): p. 199.
- [216.] Wang, J., et al., *Anti-Influenza Virus (H5N1) Activity Screening on the Phloroglucinols from Rhizomes of Dryopteris crassirhizoma*. Molecules, 2017. **22**(3): p. 431.
- [217.] Lee, J.S., et al., *Two new triterpenes from the rhizome of Dryopteris crassirhizoma, and inhibitory activities of its constituents on human immunodeficiency virus-1 protease*. Chemical and Pharmaceutical Bulletin, 2008. **56**(5): p. 711-714.
- [218.] Staniforth, V., et al., *Shikonins, phytochemicals from Lithospermum erythrorhizon, inhibit the transcriptional activation of human tumor necrosis factor  $\alpha$  promoter in vivo*. Journal of Biological Chemistry, 2004. **279**(7): p. 5877-5885.
- [219.] Younus, I., et al., *A review of ethnobotany, phytochemistry, antiviral and cytotoxic/anticancer potential of Morus alba Linn*. 2012. **73**(3): p. 487-493.

- [220.] Ooi, L.S., et al., *New mannose-binding lectin isolated from the rhizome of sarsaparilla Smilax glabra Roxb.(Liliaceae)*. Journal of agricultural and food chemistry, 2004. **52**(20): p. 6091-6095.
- [221.] Ooi, L.S., et al., *Antiviral and anti-proliferative glycoproteins from the rhizome of Smilax glabra roxb (Liliaceae)*. The American journal of Chinese medicine, 2008. **36**(01): p. 185-195.
- [222.] Montanha, J.A., et al., *Antiviral activity of Brazilian plant extracts*. Acta farmacéutica bonaerense, 2004. **23**(2): p. 183-186.
- [223.] Heim, K.E., A.R. Tagliaferro, and D.J. Bobilya, *Flavonoid antioxidants: chemistry, metabolism and structure-activity relationships*. The Journal of nutritional biochemistry, 2002. **13**(10): p. 572-584.
- [224.] Muhamad, M., et al., *Antiviral actions of flavanoid-derived compounds on dengue virus type-2*. International journal of biological sciences, 2010. **6**(3): p. 294.
- [225.] Wu, S.-F., et al., *Antiviral effects of an iminosugar derivative on flavivirus infections*. Journal of virology, 2002. **76**(8): p. 3596-3604.
- [226.] Williamson, E.M., *Synergy and other interactions in phytomedicines*. Phytomedicine, 2001. **8**(5): p. 401-409.
- [227.] Mukherjee, P.K. and P.J. Houghton, *Evaluation of herbal medicinal products*. Pharmaceutical Press 2009. **73**(3): p. 487-493.
- [228.] Srivastava, I.K. and A.B. Vaidya, *A mechanism for the synergistic antimalarial action of atovaquone and proguanil*. Antimicrobial agents and chemotherapy, 1999. **43**(6): p. 1334-1339.
- [229.] Saidu, Y., et al., *Toxicity studies of the crude aqueous root extract of albizzia chevalieri harms in albino rats*. Nigerian Journal of Basic and Applied Sciences, 2010. **18**(2) p. 487-493.
- [230.] Sebaugh, J., *Guidelines for accurate EC50/IC50 estimation*. Pharmaceutical statistics, 2011. **10**(2): p. 128-134.
- [231.] Thai, K.T., et al., *Clinical, epidemiological and virological features of Dengue virus infections in Vietnamese patients presenting to primary care facilities with acute undifferentiated fever*. Journal of Infection, 2010. **60**(3): p. 229-237.
- [232.] Bartenschlager, R. and S. Miller, *Molecular aspects of Dengue virus replication*. 2008.
- [233.] Azwanida, N., *A review on the extraction methods use in medicinal plants, principle, strength and limitation*. Med Aromat Plants, 2015. **4**(196): p. 2167-0412.1000196.

- [234.] Ramawat, K.G., S. Dass, and M. Mathur, *Herbal drugs: ethnomedicine to modern medicine*. Springer 2009. **73**(3): p. 487-493.
- [235.] DiMasi, J.A., H.G. Grabowski, and R.W. Hansen, *The cost of drug development*. New England Journal of Medicine, 2015. **372**(20): p. 1972-1972.
- [236.] Shetty, P., *Integrating modern and traditional medicine: Facts and figures*, Science and Development Network, 30 June 2010. Accessed November, 2010. **26**: p. 2011.
- [237.] Bham, Z. and E. Ross, *Traditional and western medicine: Cultural beliefs and practices of South African Indian Muslims with regard to stroke*. Ethnicity and Disease, 2005. **15**(4): p. 548.
- [238.] Chan, E., et al., *Interactions between traditional Chinese medicines and Western*. Curr Opin Drug Discov Devel, 2010. **13**(1): p. 50-65.
- [239.] Collier, A.C., et al., *Treatment of human immunodeficiency virus infection with saquinavir, zidovudine, and zalcitabine*. New England Journal of Medicine, 1996. **334**(16): p. 1011-1018.
- [240.] Qi, F., et al., *Chinese herbal medicines as adjuvant treatment during chemoradiotherapy for cancer*. Bioscience trends, 2010. **4**(6) p. 487-493.
- [241.] Cui, L. and X.-z. Su, *Discovery, mechanisms of action and combination therapy of artemisinin*. Expert review of anti-infective therapy, 2009. **7**(8): p. 999-1013.
- [242.] Organization, W.H., *SARS: Clinical trials on treatment using a combination of traditional Chinese medicine and western medicine*. Geneve, Switzerland (World Health Organization, 2004), 2009. **73**(3): p. 487-493.
- [243.] Bohin, M.C., et al., *Efficacy of food proteins as carriers for flavonoids*. Journal of agricultural and food chemistry, 2012. **60**(16): p. 4136-4143.
- [244.] Cho, N., et al., *Neuroprotective and anti-inflammatory effects of flavonoids isolated from Rhus verniciflua in neuronal HT22 and microglial BV2 cell lines*. Food and chemical toxicology, 2012. **50**(6): p. 1940-1945.
- [245.] Cho, J.-H., et al., *Eupatilin, a dietary flavonoid, induces G2/M cell cycle arrest in human endometrial cancer cells*. Food and Chemical Toxicology, 2011. **49**(8): p. 1737-1744.
- [246.] Bhourri, W., et al., *Evaluation of antioxidant and antigenotoxic activity of two flavonoids from Rhamnus alaternus L.(Rhamnaceae): Kaempferol 3-O- $\beta$ -isorhamninoside and rhamnocitrin 3-O- $\beta$ -isorhamninoside*. Food and chemical toxicology, 2011. **49**(5): p. 1167-1173.

- [247.] Xiao, J., et al., *Investigation of the mechanism of enhanced effect of EGCG on huperzine A's inhibition of acetylcholinesterase activity in rats by a multispectroscopic method*. Journal of agricultural and food chemistry, 2008. **56**(3): p. 910-915.
- [248.] Hwang, S.-L., P.-H. Shih, and G.-C. Yen, *Neuroprotective effects of citrus flavonoids*. Journal of Agricultural and Food chemistry, 2012. **60**(4): p. 877-885.
- [249.] Xie, Y., et al., *Antibacterial activities of flavonoids: structure-activity relationship and mechanism*. Current medicinal chemistry, 2015. **22**(1): p. 132-149.
- [250.] Orhan, D.D., et al., *Antibacterial, antifungal, and antiviral activities of some flavonoids*. Microbiological Research, 2010. **165**(6): p. 496-504.
- [251.] Li, Y.-L., et al., *Antiviral activities of flavonoids and organic acid from Trollius chinensis Bunge*. Journal of Ethnopharmacology, 2002. **79**(3): p. 365-368.
- [252.] Zakaryan, H., et al., *Flavonoids: promising natural compounds against viral infections*. Archives of virology, 2017. **162**(9): p. 2539-2551.
- [253.] Ono, K. and H. Nakane, *Mechanisms of inhibition of various cellular DNA and RNA polymerases by several flavonoids*. The Journal of Biochemistry, 1990. **108**(4): p. 609-613.
- [254.] Sanchez, I., et al., *Antiviral effect of flavonoids on the dengue virus*. Phytotherapy Research, 2000. **14**(2): p. 89-92.
- [255.] Cao, J., et al., *Characterization of flavonoids from Dryopteris erythrosora and evaluation of their antioxidant, anticancer and acetylcholinesterase inhibition activities*. Food and Chemical Toxicology, 2013. **51**: p. 242-250.
- [256.] Zhang, M., et al., *Flavonoid contents and free radical scavenging activity of extracts from leaves, stems, rachis and roots of Dryopteris erythrosora*. Iranian journal of pharmaceutical research: IJPR, 2012. **11**(3): p. 991.
- [257.] Han, Y., et al., *Design, synthesis, and antiviral activity of novel rutin derivatives containing 1, 4-pentadien-3-one moiety*. European journal of medicinal chemistry, 2015. **92**: p. 732-737.
- [258.] Keivan, Z., et al., *In vitro antiviral activity of fisetin, rutin and naringenin against dengue virus type-2*. Journal of Medicinal Plants Research, 2011. **5**(23): p. 5534-5539.
- [259.] Lagrota, M., et al., *Antiviral activity of lapachol*. Revista de Microbiología, 1983. **14**(1): p. 21-26.



- [260.] Zandi, K., et al., *Antiviral activity of four types of bioflavonoid against dengue virus type-2*. Virology journal, 2011. **8**(1): p. 560.
- [261.] Li, S., H. Yu, and C.T. Ho, *Nobiletin: efficient and large quantity isolation from orange peel extract*. Biomedical Chromatography, 2006. **20**(1): p. 133-138.
- [262.] Zhishen, J., T. Mengcheng, and W. Jianming, *The determination of flavonoid contents in mulberry and their scavenging effects on superoxide radicals*. Food chemistry, 1999. **64**(4): p. 555-559.
- [263.] Katsube, T., et al., *Effect of air-drying temperature on antioxidant capacity and stability of polyphenolic compounds in mulberry (Morus alba L.) leaves*. Food Chemistry, 2009. **113**(4): p. 964-969.
- [264.] Senthilvel, P., et al., *Flavonoid from Carica papaya inhibits NS2B-NS3 protease and prevents Dengue 2 viral assembly*. Bioinformation, 2013. **9**(18): p. 889.
- [265.] Júnior, I.I.d.S., et al., *Brazilian Morus nigra Attenuated Hyperglycemia, Dyslipidemia, and Prooxidant Status in Alloxan-Induced Diabetic Rats*. The Scientific World Journal, 2017. **73**(3): p. 487-493.
- [266.] Barnard, D.L., et al., *Evaluation of the antiviral activity of anthraquinones, anthrones and anthraquinone derivatives against human cytomegalovirus*. Antiviral Research, 1992. **17**(1): p. 63-77.
- [267.] van Cleef, K.W., et al., *Escape mutations in NS4B render dengue virus insensitive to the antiviral activity of the paracetamol metabolite AM404*. Antimicrobial agents and chemotherapy, 2016. **60**(4): p. 2554-2557.
- [268.] Tan Sang Loon., *Synthesis, characterization, cytotoxicity, and structure-activity relationship of chiral Schiff bases derived from dithiocarbazate and their metal complexes*. PhD thesis, Universiti Putra Malaysia: April 2014.

## BIODATA OF STUDENT

<b>Maryam Maqsood</b>	
Qualifications	MSc (Microbiology) CGPA: 3.87 <u>Gold Medal</u> year 2007
Age	32 years
Permanent address	Department of Animal Sciences SBK Women's University Brewery Road QUETTA, BALOCHISTAN, 87300 Pakistan
Present address	Virology lab, level 7 Department of Medical Microbiology and Parasitology Faculty of medicine and Health Sciences University Putra Malaysia Serdang, 43300, Selangor Malaysia
Permanent Contact number:	Cell: 092-321-8011233 Home: 092-81-2828623 Office: 092-81-9213301 Fax: 092-81-9213308
Email	<a href="mailto:maryam.umar1@yahoo.com">maryam.umar1@yahoo.com</a> (Facetime)
Facebook, LinkedIn, Research gate	Maryam Maqsood

## LIST OF PUBLICATIONS

### Publications:

Maryam U., Naheed S., Mehwish F., Shaheena R. (2008) Incidence of *Trichomonas Vaginalis* in the female patients of pelvic inflammatory disease in Quetta city, *Proceedings of Parasitology* 46; 127-133

### Award:

Second position: Three Minute Thesis (3MT) presentation competition, Universiti Putra Malaysia

### Conference paper presentation:

1. Poster presentation Genomics, Proteomics, Metabolomics: Recent Trends in Biotechnology University of Punjab, October 22, 23 2007, Lahore Pakistan

### Conference presentation:

As Speaker: Anti-dengue activity of traditional Chinese medicinal plant extracts

Inaugural FMHS Scientific Meeting 2017, 25-26 May 2017 University Tunku Abdulrahman Sugai Long.

### Poster Awards:

**Best Poster:** *In vitro* anti-dengue activity of Synthetic Schiff bases and their metal complexes

National Dengue and Arbovirus Infections Conference, 12-13 August 2017, Berjaya Times Square Hotel, Kuala Lumpur.

### Conference attended:

1. Infections 2015, 7 & 8 April 2015, IOI Marriot Hotel Putrajaya
2. Dengue 360, 11 May 2016, KLCC Convention center
3. National Dengue and Arbovirus Infections Conference 12-13 August 2017, Berjaya Times Square Hotel, Kuala Lumpur.
4. Inaugural FMHS Scientific Meeting 2017, 25-26 May 2017 University Tunku Abdulrahman Sugai Long.
5. Recent Trends in Biotechnology University of Punjab, October 22, 23 2007, Lahore Pakistan



**UNIVERSITI PUTRA MALAYSIA**

**STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT**

**ACADEMIC SESSION :** First Semester 2019/2020

**TITLE OF THESIS / PROJECT REPORT :**

IN VITRO EVALUATION OF ANTI-DENGUE ACTIVITY OF SELECTED SYNTHETIC  
SCHIFF BASES AND TRADITIONAL CHINESE MEDICINAL PLANTS EXTRACTS

**NAME OF STUDENT:** MARYAM MAQSOOD

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

1. This thesis/project report is the property of Universiti Putra Malaysia.
2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as :

\*Please tick (v )

**CONFIDENTIAL**

(Contain confidential information under Official Secret Act 1972).

**RESTRICTED**

(Contains restricted information as specified by the organization/institution where research was done).

**OPEN ACCESS**

I agree that my thesis/project report to be published as hard copy or online open access.

This thesis is submitted for :

**PATENT**

Embargo from \_\_\_\_\_ until \_\_\_\_\_  
(date) (date)

**Approved by:**

\_\_\_\_\_  
(Signature of Student)  
New IC No/ Passport No.:

Date :

\_\_\_\_\_  
(Signature of Chairman of Supervisory Committee)  
Name:

Date :

**[Note : If the thesis is CONFIDENTIAL or RESTRICTED, please attach with the letter from the organization/institution with period and reasons for confidentiality or restricted. ]**