# AUTOMATED PLATFORM FOR HISTOLOGICAL RACE AND SEX COMPARISON OF HUMAN CORTICAL BONE

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A thesis submitted in fulfillment of the requirement for the award of the Degree of Doctor of Philosophy in Electrical Engineering

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JUNE 2019

I would like to dedicate this work to the poor and needy people of the world who face neglect from the society. May Allah give me the courage and power to support them.

#### ACKNOWLEDGEMENT

I would like to thanks Almighty Allah for blessing me with the knowledge, courage and all the means to perform my research. Truly, Allah is the greatest planner of all. I was blessed with a great supervisor, Assoc. Prof. Dr. Muhammad Mahadi Abdul Jamil who was more like a guardian in my stay in Malaysia. I would like to thank Assoc. Prof. Dr. Muhammad Mahadi Abdul Jamil for all his support during my research. Without his efforts and guidelines our research would not have researched its goals. My co-supervisor Assoc. Prof. Dr. Faridah Mohd Nor not only provided us human bone specimens but also taught us the slides preparation methods and basic bone histology. I would like to thank Assoc. Prof. Dr. Faridah Mohd Nor for the multiple learning sessions despite her busy schedule in Hospital Universiti Kebangsaan Malaysia. I would also like to thank Dr. Mohd Shamian Bin Zainal for his extensive assistance.



I am indebted to Dr. Ijaz Khan for always being by my side in hard times. His encouragement and support helped me build solid foundations for the research. I can never forget my friends for their immense support and care especially during the days I was hospitalized for surgery. I would like to thank Dr. Mohamed Zaltum for his support during my PhD.

I wish to thank my parents for their support, love and encouragement, without whom I would never have enjoyed so many opportunities. I would also like to thank my wife Ms. Hira, who decided to stay in Pakistan and took great care of my old parents during my stay in Malaysia. Her sacrifice for my PhD is beyond any measures. I am particularly thankful of my brother Mr. Hammad Abdullah whose efforts provided great support for my journey.

Finally I am thankful to Universiti Tun Hussein Onn Malaysia and Office for Research, Innovation, Commercialization and Consultancy Management (ORICC), for providing GIPS GRANT vote number (U280) and support for research conferences.

#### ABSTRACT

Research on histological bone variation in population is in its early stages in Malaysia and limited information is available about age graded race and sex comparison. This research performed race and sex comparison of histological cortical bone parameters in the Malaysian population and presented an automated system which could be used as assistance tool by forensic experts. Human bone specimen were collected from Hospital Universiti Kebangsaan Malaysia Medical Centre (UKMMC), Kuala Lumpur, Malaysia. Haversian canals were measured and five parameters were calculated for comparison. Comparison test (t-test/u-test) showed that the size of Haversian canals were significantly greater (p<0.05) in females (HCM fifth, sixth decade: 5955.8  $\mu$ m<sup>2</sup>, 5788.0  $\mu$ m<sup>2</sup>) than males (HCM fifth, sixth decade: 4117.6  $\mu$ m<sup>2</sup>, 3965.1  $\mu$ m<sup>2</sup>). In race comparison, total area covered by Haversian canals (bone porosity) was significantly greater (p<0.05) in Indian samples (HCA: 0.457mm<sup>2</sup>) compared to Chinese samples (HCA: 0.385mm<sup>2</sup>) in the second decade. However in fifth decade, total area covered by Chinese samples (HCA: 0.894mm<sup>2</sup>) was significantly greater (p<0.05) than Indian samples (HCA: 0.570mm<sup>2</sup>). Three main steps of histological comparison were focused for automation i.e. parameter calculation, data management and statistical comparisons. The system was designed with GUI which utilizes aforementioned automation step. Validation of the system was divided into two main parts. In first part, parameter measurement and calculation performed by the system were compared with existing tools in terms of percentage error in measurement (DinoCapture: 5.3%, Lmeasure: 5.1%, ImageJ: 4.7%, designed system: 4.0%) and consumed time for measurement (DinoCapture: 15-20min, L-measure: 15-20min, ImageJ: 20-25min, designed system: 1-2min). Similarly automated race and sex comparison performed by the system were compared with comparisons performed manually using SPSS software. Significance and t/z values showed no differences and did not change overall hypothesis of the comparison tests. Which implies that the automated system is efficient for histological race and sex comparisons.

#### ABSTRAK

Penyelidikan mengenai variasi tulang histologi dalam perbandingan kaum dan jantina dalam populasi di Malaysia merupakan di peringkat awal dan maklumat adalah terhad. Kajian ini ialah pembinaan sistem automatik mengenai perbandingan parameter tulang kortikal histologi kaum dan jantina populasi Malaysia yang boleh digunakan sebagai alat bantuan pakar forensik. Spesimen tulang manusia telah dikumpulkan dari Pusat Perubatan Hospital Universiti Kebangsaan Malaysia (UKMMC), Kuala Lumpur, Malaysia. Terusan Haversian telah diukur dan lima parameter dikira sebagai perbandingan. Ujian perbandingan (t-test / u-test) menunjukkan bahawa saiz terusan Haversian lebih besar (p <0.05) pada wanita (HCM kelima, dekad keenam: 5955.8  $\mu$ m<sup>2</sup>, 5788.0  $\mu$ m<sup>2</sup>) daripada lelaki (HCM kelima, dekad keenam: 4117.6  $\mu$ m<sup>2</sup>, 3965.1  $\mu$ m<sup>2</sup>). Dalam perbandingan kaum, jumlah kawasan terusan Haversian (porositas tulang) jauh lebih besar (p <0.05) dalam sampel kaum India (HCA: 0.457 mm<sup>2</sup>) berbanding sampel kaum Cina (HCA: 0.385 mm<sup>2</sup>) dalam dekad kedua. Walau bagaimanapun pada dekad kelima, sampel kaum Cina (HCA: 0.894 mm<sup>2</sup>) jauh lebih tinggi (p <0.05) berbanding sampel kaum India (HCA: 0.570 mm<sup>2</sup>). Tiga langkah utama parameter automasi iaitu, pengiraan data, pengurusan data dan perbandingan statistik. Sistem ini direka dengan GUI yang menggunakan langkah automasi yang disebutkan di atas. Pengesahan sistem dibahagikan kepada dua bahagian utama, yaitu di bahagian pertama, pengukuran dan pengiraan parameter sistem dibandingkan dengan alat yang sedia ada dari segi kesilapan peratusan dalam pengukuran (DinoCapture: 5.3%, ukuran L: 5.1%, ImageJ: 4.7%, sistem yang dirancang: 4.0%) dan penggunaan masa untuk pengukuran (DinoCapture: 15-20min, L-ukuran: 15-20min, ImageJ: 20-25min, sistem yang dirancang: 1-2min). Perbandingan sistem automatik kaum dan seks yang dibandingkan dengan kaedah secara manual menggunakan perisian SPSS. Nilai penting dan t / z tidak menunjukkan perbezaan dan juga tidak mengubah hipotesis keseluruhan ujian. Ini menunjukkan bahawa sistem automatik adalah cekap untuk histologi perbandingan kaum dan jantina.



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### REFERENCES

- Abdullah, M. B. (2014). Ethnic Composition, Current Population Estimates, Malaysia, 2014 - 2016. Department of statictics Malaysia, Official portal: Department of statictics Malaysia.
- Abu Bakar, S. N., Aspalilah, A., AbdelNasser, I., Nurliza, A., Hairuliza, M. J., Swarhib, M., Mohd Nor, F. (2017). Stature Estimation from Lower Limb Anthropometry using Linear Regression Analysis: A Study on the Malaysian Population. Clin Ter, 168(2), e84-e87. doi: 10.7417/ct.2017.1988
- Aiello, L. C., & Molleson, T. (1993). Are Microscopic Ageing Techniques more accurate than Macroscopic Ageing Techniques? Journal of Archaeological Science, 20(6), 689-704. doi: http://dx.doi.org/10.1006/jasc.1993.1043
- Albu, I. G., R.Georoceanu, M. (1990). The canal system in the diaphysial compacta of the femur in some mammals (Vol. 170).
- Alias, A., Ibrahim, A., Abu Bakar, S. N., Shafie, M. S., Das, S., & Nor, F. (2017).Morphometric and morphological study of mental foramen in the malaysian population: Anatomy and forensic implications (Vol. 16).
- Bell, K. L., Loveridge, N., Reeve, J., Thomas, C. D., Feik, S. A., & Clement, J. G. (2001). Super-osteons (remodeling clusters) in the cortex of the femoral shaft: influence of age and gender. Anat Rec, 264(4), 378-386.
- Bertelsen, P. K., Clement, J. G., & Thomas, C. D. L. (1995). A morphometric study of the cortex of the human femur from early childhood to advanced old age. Forensic Sci Int, 74(1), 63-77. doi: https://doi.org/10.1016/0379-0738(95)01738-5
- Black, S. (2001). A Tour of Mile Canyon: The University of Texas at Austin and College of Liberal Arts.
- Boivin, G. (2007). The hydroxyapatite crystal: A closer look. Medicographia, 29, 126-132.

- Britz, H. M., Thomas, C. D., Clement, J. G., & Cooper, D. M. (2009). The relation of femoral osteon geometry to age, sex, height and weight. Bone, 45(1), 77-83. doi: 10.1016/j.bone.2009.03.654
- Bruce Martin, R., Burr, D., Sharkey, N., & Fyhrie, D. (2015). Skeletal Tissue Mechanics.
- Burr, D. B., Ruff, C. B., & Thompson, D. D. (1990). Patterns of skeletal histologic change through time: comparison of an archaic native American population with modern populations. Anat Rec, 226(3), 307-313. doi: 10.1002/ar.1092260306
- Carneiro, L. J. a. J. (2005). Basic Histology: Text and Atlas (A. L. Mescher Ed. 14 ed.): McGraw-Hill Medical.
- contributors, W. C. (2018). Bone cross-section: Wikimedia Commons, the free media repository.

Cormack, H. (1987). Ham's Histology: Philadelphia: J. B. Lippincott.

- Cosgriff-Hernandez, M.-T. J. (2012). Histomorphometric Estimation of Age at Death Using the Femoral Cortex: A Modification of Established Methods. The Ohio State University. Retrieved from http://rave.ohiolink.edu/etdc/view?acc\_num=osu1338361172
- Cunningham, C., Scheuer, L., & Black, S. (2016). Chapter 3 Bone Development. InC. Cunningham, L. Scheuer & S. Black (Eds.), Developmental JuvenileOsteology (Second Edition) (pp. 19-35). San Diego: Academic Press.
- De Boer, Aarents, & Maat. (2012). Staining ground sections of natural dry bone tissue for microscopy. International Journal of Osteoarchaeology, 22(4), 379-386. doi: doi:10.1002/oa.1208
- Dupras, T. L., Schultz, J. J., Wheeler, S. M., & Williams, L. J. (2011). Forensic Recovery of Human Remains: Archaeological Approaches, Second Edition: Taylor & Francis.
- Ericksen, M. F. (1991). Histologic estimation of age at death using the anterior cortex of the femur. Am J Phys Anthropol, 84(2), 171-179.
- Faridah Mohd, N., Robert, F. P., & Holger, S. (2013). Age at death estimation from bone histology in Malaysian males. Medicine, Science and the Law, 54(4), 203-208. doi: 10.1177/0025802413506573
- Forwood, M. R., & Turner, C. H. (1995). Skeletal adaptations to mechanical usage: results from tibial loading studies in rats. Bone, 17(4 Suppl), 197s-205s.

- Frost, H. M. (1969). Tetracycline-based histological analysis of bone remodeling. Calcif Tissue Res, 3(3), 211-237.
- Frost, H. M. (1999). Why do bone strength and "mass" in aging adults become unresponsive to vigorous exercise? Insights of the Utah paradigm. J Bone Miner Metab, 17(2), 90-97.
- G. Robling, A., & Stout, S. (2007). Histomorphometry of Human Cortical Bone: Applications to Age Estimation.
- GeneralHelghast. (2013). The Glorious Races of the World map: Digital Art, GeneralHelghast.
- Granke, M., Makowski, A. J., Uppuganti, S., & Nyman, J. S. (2016). Prevalent role of porosity and osteonal area over mineralization heterogeneity in the fracture toughness of human cortical bone. J Biomech, 49(13), 2748-2755. doi: 10.1016/j.jbiomech.2016.06.009
- Gray, H. (1918). Anatomy of the Human Body (Vol. 1): Bartleby.
- Hall, B. K. (2005). Bones and cartilage: developmental and evolutionary skeletal biology. Elsevier Academic Press (USA) Ltd.
- Ijaz, K., Muhammad Mahadi Bin Abdul, J., & Faridah Mohd, N. (2018). Age regression for Malaysian males using cortical bone Histomorphometry. Journal of Physics: Conference Series, 1019(1), 012011.

Imaging, S. (2000). Spot Cameras. In S. I. C. cameras (Ed.).

- John.R., Mig G., Pind M., Vick D. Brown R., & Mj, A. (2001). Manual preparation of ground sections for the microscopy of natural bone tissue: update and modification of Frost's 'rapid manual method'. International Journal of Osteoarchaeology, 11(5), 366-374. doi: doi:10.1002/oa.578
- Jamil, M. M. A., Khan, I., Abdullah, H., & Nor, F. M. (2018). Microscopic analysis of bone microstructures with increasing age in malaysian females. 2018, 10(6S), 12. doi: 10.4314/jfas.v10i6s.38
- Jaworski, Z. F. (1984). Coupling of bone formation to bone resorption: a broader view. Calcif Tissue Int, 36(5), 531-535.
- Junqueria LC, C. J. (2003). Basic Histology: Text and Atlas. Bone. In: FoltinJ, Lebowitc H, Boyle PJ10ed. McGraw-Hill companies, Inc, p, 141-149.
- Kashi, A., & Saha, S. (2017). Chapter 11 Ethical Issues in Biomaterials Research A2
  Bose, Susmita. In A. Bandyopadhyay (Ed.), Materials for Bone Disorders (pp. 493-503): Academic Press.

- Katz, D., & Suchey, J. M. (1986). Age determination of the male os pubis. Am J Phys Anthropol, 69(4), 427-435. doi: 10.1002/ajpa.1330690402
- Kerley, E. R. (1965). The microscopic determination of age in human bone. Am J Phys Anthropol, 23(2), 149-163. doi: 10.1002/ajpa.1330230215
- Khan, I., Jamil, M. M. A., Ibrahim, T. N. T., & Nor, F. M. (2016a, 25-27 Nov. 2016). Analysis of age-related changes in Haversian canal using image processing techniques. Paper presented at the 2016 6th IEEE International Conference on Control System, Computing and Engineering (ICCSCE).
- Khan, I., Jamil, M. M. A., Ibrahim, T. N. T., & Nor, F. M. (2016b, 25-27 Nov. 2016). Automated human age estimation at death via bone microstructures. Paper presented at the 2016 6th IEEE International Conference on Control System, Computing and Engineering (ICCSCE).
- Khan, I., Jamil, M. M. A., Ibrahim, T. N. T., & Nor, F. M. (2017). Analysis of agerelated changes in Haversian canal using image processing techniques. Paper presented at the Proceedings - 6th IEEE International Conference on Control System, Computing and Engineering, ICCSCE 2016.
- Khan, I., Jamil, M. M. A., & Nor, F. M. (2017). Evaluation and reliability of bone histological age estimation methods. 2017, 9(4S), 18. doi: 10.4314/jfas.v9i4s.38
- Lanyon, L. E., Sugiyama, T., & Price, J. S. (2009). Regulation of bone mass: Local control or systemic influence or both? IBMS BoneKEy, 6(6), 218-226.
- Leeson, R. C., Leeson, T. S., Paparo, A. A. (1985). Specialized connective tissue:cartilage and bone. Textbook of Histology, 5 edn, W.B. Saunders Co., Japan,, 125-149.
- Li, D., Liu, J., Guo, B., Liang, C., Dang, L., Lu, C., . . . Zhang, G. (2016). Osteoclastderived exosomal miR-214-3p inhibits osteoblastic bone formation. Nature Communications, 7, 10872. doi: 10.1038/ncomms10872 https://www.nature.com/articles/ncomms10872#supplementary-information

Malaysia, J. P. (2017). Population Quick Info.

Mann, H. B., & Whitney, D. R. (1947). On a Test of Whether one of Two Random Variables is Stochastically Larger than the Other. Ann. Math. Statist., 18(1), 50-60. doi: 10.1214/aoms/1177730491

- Mansourvar, M. (2014). Bone age assessment using hand and clavicle X-ray images. Faculty of computer science and information technology university of Malaya Kuala Lumpur.
- Mulhern, D. M., & Van Gerven, D. P. (1997). Patterns of femoral bone remodeling dynamics in a Medieval Nubian population. Am J Phys Anthropol, 104(1), 133-146. doi: 10.1002/(sici)1096-8644(199709)104:1<133::aid-ajpa9>3.0.co;2-s
- Nor, F., Pastor, R. F., & Schutkowski, H. (2015). Histological study to differentiate between human and non-human long bone (Vol. 22).
- Nor, F. M. (2010). A comparative microscopic study of human and non-human long bone histology. (Ph.D), University of Bradford, 2010-11-05. Retrieved from http://hdl.handle.net/10454/4463
- Nor, F. M., Pastor, R. F., & Schutkowski, H. (2014). Age at death estimation from bone histology in Malaysian males. Med Sci Law, 54(4), 203-208. doi: 10.1177/0025802413506573
- Ortner, D. J. (1975). Aging effects on osteon remodeling. Calcif Tissue Res, 18(1), 27-36.
- Petrtyl, M., Hert, J., & Fiala, P. (1996). Spatial organization of the haversian bone in man. J Biomech, 29(2), 161-169.
- Raisz, L. G. (1999). Physiology and pathophysiology of bone remodeling. Clin Chem, 45(8 Pt 2), 1353-1358.
- Ridgman, W. J. (2009). Statistical Methods, 8th edn, by G. W. Snedecor & amp; W. G.
  Cochran. xx + 503 pp. Ames: Iowa State University Press (1989). \$44.95 (hard covers). ISBN 0 8138 1561 6. The Journal of Agricultural Science, 115(1), 153-153. doi: 10.1017/S0021859600074104
- Ross, M. H., Romrell, L. J., Kaye, G. I. (1995). Bone. In: Histology: A Text and Atlas,3ed. Williams & Wilkens, Philadelphia,, 150-187.
- Samson C, B. K. (1987). A new method of estimating age at death from fragmentary and weathered bone. In Bodington A, Garland AN, Janaway RC, editors. Death Decay and Reconstruction Approaches to Archaeology and Forensic Science. Manchester: Manchester University Press, 101-108.
- Schaffler, M. B., & Burr, D. B. (1984). Primate cortical bone microstructure: relationship to locomotion. Am J Phys Anthropol, 65(2), 191-197. doi: 10.1002/ajpa.1330650211

- Schoenbuchner, S. M., Pettifor, J. M., Norris, S. A., Micklesfield, L. K., Prentice, A., & Ward, K. A. (2017). Ethnic Differences in Peripheral Skeletal Development
  Among Urban South African Adolescents: A Ten-Year Longitudinal pQCT
  Study. J Bone Miner Res, 32(12), 2355-2366. doi: 10.1002/jbmr.3279
- Shapiro, S. S., & Wilk, M. B. (1965). An Analysis of Variance Test for Normality (Complete Samples). Biometrika, 52(3/4), 591-611. doi: 10.2307/2333709
- Sommerfeldt, D. W., & Rubin, C. T. (2001). Biology of bone and how it orchestrates the form and function of the skeleton. Eur Spine J, 10 Suppl 2, S86-95. doi: 10.1007/s005860100283
- Starrett. (2018). Water Resistant Digital Caliper: The L. S. Starrett Company Limited.
- Steele, D. G. (1988). The Anatomy and Biology of the Human Skeleton: Texas A&M University Press.
- Stevens, D. D., & Levi, A. (2005). Introduction to Rubrics: An Assessment Tool to Save Grading Time, Convey Effective Feedback, and Promote Student Learning: Stylus Pub.
- Stout, S. D., & Lueck, R. (1995). Bone remodeling rates and skeletal maturation in three archaeological skeletal populations. Am J Phys Anthropol, 98(2), 161-171. doi: 10.1002/ajpa.1330980206
- Suchey, J. M. (1979). Problems in the aging of females using the Os pubis. Am J Phys Anthropol, 51(3), 467-470. doi: 10.1002/ajpa.1330510319
- Swee-Hock, S. (2015). The Population of Malaysia (Second Edition): Institute of Southeast Asian Studies.
- Teitelbaum, S. L. (2000). Bone resorption by osteoclasts. Science, 289(5484), 1504-1508.
- Thomas, C. D., Feik, S. A., & Clement, J. G. (2005). Regional variation of intracortical porosity in the midshaft of the human femur: age and sex differences. J Anat, 206(2), 115-125. doi: 10.1111/j.1469-7580.2005.00384.x
- Thomas, C. D. L., Feik, S. A., & Clement, J. G. (2005). Regional variation of intracortical porosity in the midshaft of the human femur: age and sex differences. Journal of Anatomy, 206(2), 115-125. doi: 10.1111/j.1469-7580.2005.00384.x
- Thompson, D. D. (1979). The core technique in the determination of age at death of skeletons. J Forensic Sci, 24(4), 902-915.

- Thompson, D. D., & Gunness-Hey, M. (1981). Bone mineral-osteon analysis of Yupik-inupiaq skeletons. Am J Phys Anthropol, 55(1), 1-7. doi: 10.1002/ajpa.1330550102
- Wheater PR, a. B. H. (1987). Functional Histology: A Text and Colour Atlas: Edinburgh: Churchill Livingstone.
- White, T. D., Black, M. T., & Folkens, P. A. (2012a). Chapter 4 Skull: Cranium and Mandible Human Osteology (Third Edition) (pp. 43-100). San Diego: Academic Press.
- White, T. D., Black, M. T., & Folkens, P. A. (2012b). Chapter 9 Arm: Humerus, Radius, and Ulna Human Osteology (Third Edition) (pp. 175-198). San Diego: Academic Press.
- White, T. D., Black, M. T., & Folkens, P. A. (2012c). Chapter 12 Leg: Femur, Patella, Tibia, and Fibula Human Osteology (Third Edition) (pp. 241-270). San Diego: Academic Press.
- Wu, K., Schubeck, K. E., Frost, H. M., & Villanueva, A. (1970). Haversian bone formation rates determined by a new method in a mastodon, and in human diabetes mellitus and osteoporosis. Calcif Tissue Res, 6(3), 204-219.