

POLYLACTIC ACID/THERMOPLASTIC TAPIOCA STARCH BLEND
INCORPORATED WITH SELECTED ESSENTIAL OILS FOR ACTIVE FOOD
PACKAGING

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ABSTRACT

The environmental effect of petroleum-based polymers and the activity of food spoilage microorganisms made researchers work on alternative bioactive packaging materials. This work aimed at producing sustainable thermoplastic tapioca starch (TPTS) and its polylactic acid (PLA) blend with introduction of antimicrobial (AM) activity via incorporation of lemongrass (LG), lemon balm (LB) and pandan (PA) essential oils (EOs) on the films. The plasticising effect of glycerol and water and compatibilising effect of vinegar loadings on the properties of TPTS were tested. So also the impact of TPTS loading on the PLA/TPTS blend properties. Furthermore, the AM activity of TPTS incorporated with the EOs on *E. coli*, *B. cereus* and *S. marcescens* was evaluated by inhibitory zone; and packaging effect of PLA/TPTS films coated with the EOs on death rate of microorganisms inoculated on *Bahulu* cake was investigated. The TPTS was formed by tape casting method, while PLA/TPTS film by hot pressing. The results indicated an increase in crystallinity and elongation at break with increase in plasticiser loading in TPTS and percentage TPTS loading in PLA/TPTS blend; both were accompanied by a decrease in T_g and tensile strength. Continuous surface morphology was seen at high plasticiser loading in TPTS, while high TPTS loading manifested a phase separation in PLA/TPTS blend. The results show that 125 °C processing temperature, 15 wt% glycerol, 45 wt% water and 3.5 wt% vinegar loading were optimum parameters for TPTS production. Whereas, 185°C processing temperature and 25 wt% TPTS loading are optimum parameters for PLA/TPTS blend production. Citral was found as the common active compound among the EOs. LG and LB were active against all the tested microorganisms, while PA was very mild at higher concentration. For active TPTS and PLA/TPTS films activity, *E. coli*: LB>LG>PA; *B. cereus*: LG>LB>PA; *S. marcescens*: LB>LG>PA. The TPTS and PLA/TPTS active films produced have shown improved properties for food packaging and effective AM activity against the selected microorganisms.

ABSTRAK

Kesan alam sekitar daripada polimer berasaskan petroleum dan aktiviti mikroorganisma yang membinasakan makanan membuat penyelidik bertumpu pada bahan pembungkusan bioaktif alternatif. Kajian ini bertujuan menghasilkan adunan lestari termoplastik kanji ubi kayu (TPTS) dan asid polilaktik (PLA) dengan aktiviti antimikrob (AM) yang diperbaiki menerusi gabungan minyak pati (EOs) *serai* (LG), *lemon balm* (LB) dan *pandan* (PA). Kesan pengekstrakan gliserol dan air dan kesan keserasian kandungan cuka ke atas sifat-sifat TPTS telah diuji. Begitu juga kesan kandungan TPTS pada sifat gabungan PLA/TPTS. Tambahan pula, aktiviti AM TPTS yang digabungkan dengan EOs pada *E. coli*, *B. cereus* dan *S. marcescens* dinilai oleh zon penghalang; dan kesan pembungkusan filem PLA/TPTS yang disalut dengan EOs pada kadar kematian mikroorganisma yang diumpukkan pada kek Bahulu telah disiasat. TPTS terhasil menerusi kaedah tuangan pita, manakala filem PLA/TPTS terhasil menerusi penekanan panas. Hasil dapatan menunjukkan peningkatan dalam kehabluran dan pemanjangan patah dengan pertambahan kandungan pemplastik dalam TPTS serta peratus kandungan TPTS dalam adunan PLA/TPTS, kedua-duanya dituruti dengan pengurangan T_g dan kekuatan tegangan. Morfologi permukaan yang berterusan dilihat pada kandungan pemplastik yang tinggi dalam TPTS, manakala kandungan TPTS yang tinggi menunjukkan pemisahan fasa dalam adunan PLA/TPTS. Keputusan menunjukkan bahawa pada suhu pemprosesan 125°C , 15 wt% berat gliserol, 45 wt% air dan 3.5 wt% cuka adalah parameter optimum untuk pengeluaran TPTS. Sementara itu, suhu pemprosesan 185°C dan 25 wt% TPTS adalah parameter optimum untuk pengeluaran PLA/TPTS. Citral dijumpai sebagai sebatian aktif yang biasa di kalangan EOs. LG dan LB aktif terhadap semua mikroorganisma yang diuji, sementara PA kurang berkesan pada kepekatan yang lebih tinggi. Untuk aktiviti filem aktif TPTS dan PLA/TPTS, *E. coli*: LB>LG>PA; *B. cereus*: LG>LB>PA; *S. marcescens*: LB>LG>PA. Filem aktif TPTS dan PLA/TPTS yang terhasil telah menunjukkan sifat yang dipertingkatkan sebagai pembungkus makanan dan aktiviti AM berkesan terhadap mikroorganisma terpilih.

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LIST OF SYMBOLS AND ABBREVIATIONS

μ	-	death rate
α	-	Alpha crystalline structure
ε	-	Elongation at break (%)
σ	-	Tensile stress (MPa)
$^{\circ}\text{C}$	-	Degree Celsius
μm	-	Micrometre
ANOVA	-	Analysis of variance
$\text{Au} \times \text{nm}$	-	Area under curve \times nanometer
ASTM	-	American society for testing and materials
BCM	-	Bacillus cereus medium
CEN	-	European standardisation committee
DIN	-	German institute for standardisation
DMA	-	Dynamic mechanical analysis
DSC	-	Differential scanning calorimetry
DTG	-	Derivative thermogravimetry
E'	-	Storage elastic modulus
E''	-	Loss viscous modulus
EMB	-	Eosin methylene blue
FTIR	-	Fourier transform infrared
g	-	gram
GCMS	-	Gas chromatography mass spectrometry
HPMC	-	Hydroxypropyl methyl cellulose
ISO	-	International Standards Organisation
LAB	-	Lactic acid bacteria
MBC	-	Minimum bactericidal concentration
MIC	-	Minimum inhibition concentration
min	-	minute

mm	-	millimeter
N	-	Population
NA	-	Nutrient agar
NB	-	Nutrient Broth
ORCA	-	Organic reclamation and composting association
PDA	-	Potatoes dextrose agar
PLA	-	Polylactic acid
PVC	-	Polyvinyl chloride
rpm	-	Revolution per minute
s	-	second
SEM	-	Scanning electron microscopy
t	-	Time
Tan δ	-	Damping coefficient
T _g	-	Glass transition temperature
TGA	-	Thermogravimetric analysis
T _m	-	Melting temperature
TPS	-	Thermoplastic starch
TPTS	-	Thermoplastic tapioca starch
XLD	-	Xylose lysine deoxy-cholate
XRD	-	X-ray diffraction
UV	-	Ultraviolet



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CHAPTER 1

INTRODUCTION

1.1 Research background

The plastics have immense importance in modern life; its function is universal ranging from household articles, self-healing materials, clothes, shoes, packaging, up to auto parts. The primary source of plastics is the fossil fuel. The increased change of petroleum resource price in the 1970s extend the consciousness of researchers toward alternative sources for plastics, which is one of the significant products of fossil fuel (Wittcoff *et al.*, 2013). The rate of depletion of fossil fuel is among other factors of concern (Pervaiz *et al.*, 2014). This limited resource is bound to finish in the midst of an uneven price system. It is forecasted that fossil fuel will be depleted averagely by 2100 if the current rate of consumption is maintained (IEA, 2017). Secondly, the environmental pollution caused by the excessive volume of non-biodegradable plastic waste materials poses a threat to life and wellbeing of the environment. This issue is coupled with inadequate landfill space for disposal, the high maintenance cost of landfill, and non-degradability of the plastics for about a thousand years (Cressey, 2016).

The cumulative amount of plastics produced globally between 1950 and 2015 was around 7.82 billion tonnes. Most of these products were used in the field of packaging. One unique characteristic of plastic packaging is that it is used once after which it is discarded, particularly the food packaging where contamination by the microorganisms is involved (Geyer *et al.*, 2017). By 2015 the packaging sector accounted for 42 % (141 million tonnes) of global plastic produced. While during the same period, packaging constituted about 50 % of global plastic waste (Hannah & Max, 2018).

In Malaysia a similar trend was observed, packaging constituted 45 % of plastics produced. Also, about 24 % of municipal solid waste is made of plastic materials, even though, the percentage of packaging material was not quantified (Zainua & Songip, 2017). But research shows that about 22-55 billion shopping bags are sold per year in Malaysia (Sung, 2017). This activity constitutes a major environmental concern because only 5 % of the plastics are recycled, 42 % of the municipal waste is incinerated which brings about air pollution. On the other hand, the rest of the 53 % are dumped in landfills, and the plastics in this category does not degrade quickly, destroying the environment in open landfills (Zainua & Songip, 2017). Disposable food package provides the benefit and convenience of one time, sanitary use. Thin films used in such products are typically made from water-insoluble synthetic polymers or polymer blends. However, the disposal of these products is a concern due to limited landfill space. Incineration of such products is not desirable because of increasing alarm about greenhouse gases generation. Consequently, there is a need for bio-degradable products which may be quickly and conveniently disposed of without dumping or incineration (Gabor & Tita, 2012). It has been proven by series of researches (Iwate, 2015; Roshafima *et al.*, 2012; Seligra *et al.*, 2016) that petroleum-based plastics degrade eventually, but that process usually takes a very long time and contribute to global warming through the release of carbon dioxide and methane gas. Therefore, research in alternative renewable, biodegradable, eco-friendly and cost-effective raw material for the production of plastics flourish.

Biodegradable plastics are made out of ingredients that can be metabolised by naturally occurring micro-organisms in the environment. Bio-degradable films and coatings have been mainly considered in food preservation, because of their capability for improving global food quality (McKeen, 2012). Recently, innovative ways to inhibit microbial growth in the foods while maintaining quality, freshness, and safety have been investigated. One option is the use of packaging to provide protection, quality and longer shelf life (Priyaa *et al.*, 2014). Packaging is the most significant process aimed at giving the stable condition of food products for their storage, transportation, distribution, and consumption. The primary function of packaging is protection from mechanical damage and prevention or inhibition of chemical changes, biochemical changes and microbiological spoilage (Biron, 2007). Food products during packaging can be subjected to various type of contaminations including microbial contamination that is mainly caused by bacteria, yeasts and mould (Malhotra

et al., 2015; Prasad & Kochhar, 2014). Specifically, food products are sensitive and their shelf life is limited by their interaction with packaging materials that can alter their water activity, pH, added preservatives and also substances that can affect their temperature, relative humidity, light and gas composition (Lucera *et al.*, 2016; Miranda *et al.*, 2016). One of the significant possibility to extend the shelf life of the products is to develop packaging material with specific properties such as pH, water activity, respiration rate and existence of antimicrobial compounds which is referred to as bioactive packaging material and can be used to improve the quality of food products and to extend their shelf life (Du *et al.*, 2011). In food packaging, the goal is to use bioactive materials to get a desirable response, for example, the inhibition of microbial growth, barrier adjusting materials, as well as flavour maintenance and enhancing material properties. Most bio-based materials such as polysaccharides and protein-based polymers (starch, cellulose, alginate, collagen, gelatin, and proteins) are hydrophilic with a relatively high degree of crystallinity causing processing and performance difficulties. Therefore, antimicrobial (AM) packages made from such bio-based films demonstrate high moisture sensitivity, poor water barrier and poor mechanical properties compared to those made from synthetic polymers such as polyvinyl chloride (PVC), polypropylene (PP) and polyethylene terephthalate (PET) (Roshafima *et al.*, 2012). Packaging materials with suitable physicomechanical properties can be prepared from biopolymers such as starch-based materials when the contents are modified by physical, mechanical and chemical techniques or by blending with compatible plasticisers (McKeen, 2012).

The production of starch globally was estimated to be around 75 million tons in 2012, where maize, tapioca (cassava), wheat and potato were the major plants for starch production with an amount of rice and other starches being produced (Waterschoot *et al.*, 2015). Tapioca starch global production was estimated to reach a volume of 6.7 million tons, and the report forecast a 1.6 % growth annually where it will be 7.4 million tons by 2023 (IMARC, 2018). It was reported that Malaysia grows about 400,000 tonnes/year of tapioca, which is large enough to earn a sustainable income to farmers, and provide raw materials to the industries (Hillocks *et al.*, 2002; UFAO, 2012). But another report shows that Malaysia's tapioca production in 2016 stood at 77,980 tonnes declining from 400,000 tonnes in 1989 due to market and other factors (Factfish, 2017).

REFERENCES

- Abdellatif, F., Boudjella, H., Zitouni, A., & Hassani, A. (2014). Chemical Composition and antimicrobial activity of the essential oil from leaves of Algerian Melissa Officinalis. *EXCLI Journal*, 13, 772-781
- Acevedo-Fani, A., Salvia-Trujillo, L., Rojas-Graü, M. A., & Martín-Belloso, O. (2015). Edible films from essential-oil-loaded nanoemulsions: Physicochemical characterization and antimicrobial properties. *Food Hydrocolloids*, 47, 168-177
- Adinee, J., Piri, K., & Karami, O. (2009). Essential Oil Composition of Lemon Balm (Melissa officinalis L.) Leaves Grown in Hamadan Province, Iran. *Medicinal and Aromatic Plant Science and Biotechnology*
- Adukwu, E. C., Bowles, M., Edwards-Jones, V., & Bone, H. (2016). Antimicrobial activity, cytotoxicity and chemical analysis of lemongrass essential oil (Cymbopogon flexuosus) and pure citral. *Appl Microbiol Biotechnol*, 100(22), 9619-9627
- Adukwu, E. C., Bowles, M., Edwards-Jones, V., & Bone, H. (2016). Antimicrobial activity, cytotoxicity and chemical analysis of lemongrass essential oil (Cymbopogon flexuosus) and pure citral. *Applied Microbiology and Biotechnology*, 100(22), 9619-9627
- Ahmad, A., & Viljoen, A. (2015). The in vitro antimicrobial activity of Cymbopogon essential oil (lemon grass) and its interaction with silver ions. *Phytomedicine*, 22(6), 657-665
- Ahvenainen, P., Kontro, I., & Svedström, K. (2016). Comparison of sample crystallinity determination methods by X-ray diffraction for challenging cellulose I materials. *Cellulose*, 23(2), 1073-1086
- Akhila, A. (2010). Chemistry and Biogenesis of Essential Oils from the Genus Cymbopogon. *New York: Taylor and Francis group, LLC*, 69

- Akrami, M., Ghasemi, I., Azizi, H., Karrabi, M., & Seyedabadi, M. (2016). A new approach in compatibilization of the poly(lactic acid)/thermoplastic starch (PLA/TPS) blends. *Carbohydr Polym*, 144, 254-262
- Albayrak, S., Aksoy, A., Albayrak, S., & Sagdic, O. (2013). In vitro antioxidant and antimicrobial activity of some Lamiaceae species. *IJST, AI*, 1-9
- Alboofetileh, M., Rezaei, M., Hosseini, H., & Abdollahi, M. (2014). Antimicrobial activity of alginate/clay nanocomposite films enriched with essential oils against three common foodborne pathogens. *Food Control*, 36(1), 1-7
- Alrumman, S. A. (2016). Enzymatic saccharification and fermentation of cellulosic date palm wastes to glucose and lactic acid. *Braz J Microbiol*, 47(1), 110-119
- Alshehrei, F. (2017). Biodegradation of Synthetic and Natural Plastic by Microorganisms. *Journal of Applied & Environmental Microbiology*, 5(1), 8-19
- Ander, O., M Angeles Corcuera, Pen, C., Eceiza, A., & Arbelaiz, A. (2014). Bionanocomposites based on thermoplastic starch and cellulose nanofibers. *Journal of Thermoplastic Composite Materials & Design*, 1-16
- Anon. (1988). Glossary of packaging terms *The Packaging Institute International Stanford Connecticut*.
- Arboleda, G. A., Montilla, C. E., Villada, H. S., & Varona, G. A. (2015). Obtaining a Flexible Film Elaborated from Cassava Thermoplastic Starch and Polylactic Acid. *International Journal of Polymer Science*, 2015, 1-9
- ASTM-D-882. (2012). Standard test Method for Tensile properties of thin plastic sheeting significance and use.
- ASTM-D-3418. (1999). Standard Test Method for Transition Temperatures of Polymers By Differential Scanning Calorimetry.
- ASTM-D-5338-98. (2003). Standard Test Method for Determining Aerobic Biodegradation of Plastic Materials Under Controlled Composting Conditions. 1-6
- Attaran, S. A., Hassan, A., & Wahit, M. U. (2015). Materials for food packaging applications based on bio-based polymer nanocomposites. *Journal of Thermoplastic Composite Materials*, 30(2), 143-173
- Averous, L., & Boquillon, N. (2004). Biocomposites based on plasticized starch: thermal and mechanical behaviours. *Carbohydr Polym*, 56(2), 111-122

- Ayana, B., Suin, S., & Khatua, B. B. (2014). Highly exfoliated eco-friendly thermoplastic starch (TPS)/poly (lactic acid)(PLA)/clay nanocomposites using unmodified nanoclay. *Carbohydr Polym*, *110*, 430-439
- Bachrouri, M., Quinto, E. J., & Mora, M. T. (2002). Survival of Escherichia Coli O157:H7 During Storage of Yogurt at Different Temperatures. *Journal of Food Science*, *6*(5)
- Balakrishnan, P., Sreekala, M. S., Kunaver, M., Huskic, M., & Thomas, S. (2017). Morphology, transport characteristics and viscoelastic polymer chain confinement in nanocomposites based on thermoplastic potato starch and cellulose nanofibers from pineapple leaf. *Carbohydr Polym*, *169*, 176-188
- Bastarrachea, L. J., Dana, E. W., Maxine, J. R., Lin, Z., & Julie, M. G. (2015). Active packaging Coatings. *Coatings*, *5*, 771-791
- Bastioli, C. (1998). Biodegradable materials Present situation and future perspective. *Macromol. Symp.*, *135*, 193-204
- Bensebaa, F. (2013). Chapter 4 - Nanoparticle Assembling and System Integration. In F. Bensebaa (Ed.), *Interface Science and Technology* (Vol. 19, pp. 185-277): Elsevier.
- Bergo, P., Sobral, P. J. A., & Prison, J. M. (2010). Effect of Glycerol on Physical Properties of Cassava Starch Films. *Journal of Food Processing and Preservation*, *34*, 401-410
- Bergo, P. V. A., Carvalho, R. A., Sobral, P. J. A., dos Santos, R. M. C., da Silva, F. B. R., Prison, J. M., Solorza-Feria, J., & Habitante, A. M. Q. B. (2008). Physical properties of edible films based on cassava starch as affected by the plasticizer concentration. *Packaging Technology and Science*, *21*(2), 85-89
- Bhosle, S. M., & Friedrich, C. R. (2017). Facile Synthesis of Nanosilver-Incorporated Titanium Nanotube for Antibacterial Surfaces. *Journal of Bio- and Tribo-Corrosion*, *3*(3)
- Bibhu, P. S., Panchanan, G., & Chakrapani, P. (2016). Pandanus Odoratissimus Linn: Isolation of Stigmast-5, 22-dien-3 β -ol from Ethanolic Extract of Stem Bark and Study of Antimicrobial Activity. *Journal of Chemistry and Chemical Sciences*, *6*(6), 574-585
- Bilal, D., Ali, Y., & Hüseyin, E. (2011). Crystallization Behavior of PET Materials. *BAÜ Fen Bil. Enst. Dergisi Cilt*, *13*(1), 26-35

- Biron, M. (2007). *Thermoplastics and Thermoplastic Composites Technical Information for Plastics Users*: Elsevier Science.
- Bonferoni, M. C., Sandri, G., Rossi, S., Usai, D., Liakos, I., Garzoni, A., Fiamma, M., Zanetti, S., Athanassiou, A., Caramella, C., & Ferrari, F. (2017). A novel ionic amphiphilic chitosan derivative as a stabilizer of nanoemulsions: Improvement of antimicrobial activity of *Cymbopogon citratus* essential oil. *Colloids Surf B Biointerfaces*, 152, 385-392
- Boyce, J. M. (2018). Alcohols as Surface Disinfectants in Healthcare Settings. *Infect Control Hosp Epidemiol*, 39(3), 323-328
- BSAC. (2014). BSAC-disc-susceptibility-testing-method. (13)
- Burt, S. (2004). Essential oils: their antibacterial properties and potential applications in foods--a review. *Int J Food Microbiol*, 94(3), 223-253
- Byun, Y., Zhang, Y., & Gen, X. (2014). Plasticization and Polymer Morphology, *Innovations in Food Packaging* (pp. 87-109): Elsevier Ltd.
- Carraher, C. E. J. (2011). *Polymer Chemistry* (8th ed.). US: CRC Press, Taylor & Francis.
- Cetin-Karaca, H., & Newman, M. C. (2015). Antimicrobial efficacy of plant phenolic compounds against *Salmonella* and *Escherichia Coli*. *Food Bioscience*, 11, 8-16
- Chang, Y. P., Cheah, P. B., & Seow, C. C. (2000). Plasticizing-Antiplasticizing Effects of Water on Physical Properties of Tapioca Starch Films in the Glassy State. *Journal of Food Science*, 65(3), 445-453
- Chauhan, S. K., Kumar, R., & Nadasabapathi, S. (2017). Turbidimetric Assay of Nisin in Tender Coconut Water. *Defence Life Science Journal*, 2(2), 212
- Chen, Q., Yu, H., Wang, L., ul Abidin, Z., Chen, Y., Wang, J., Zhou, W., Yang, X., Khan, R. U., Zhang, H., & Chen, X. (2015). Recent progress in chemical modification of starch and its applications. *RSC Advances*, 5(83), 67459-67474
- Chen, X. K., & Ge, F. H. (2014). Chemical components from essential oil of *Pandanus amaryllifolius* leaves. *Journal of Chinese Medicinal Materials*, 37(4), 616-620
- Chieng, B. W., Ibrahim, N. A., Then, Y. Y., & Loo, Y. Y. (2014). Epoxidized vegetable oils plasticized poly(lactic acid) biocomposites: mechanical, thermal and morphology properties. *Molecules*, 19(10), 16024-16038

- Combrzynski, M., Moscicki, L., Rejak, A., Wojtowicz, A., & Oniszczyk, T. (2013). Selected mechanical properties of starch film. *Teka Komisji Energetyki i Rolnictwa, 13*(2), 7-12
- Cortes, H., Edgar, V., Suárez, R., Castillo, J., Gonçalves, L. M., & Bueno, P. R. (2014). Pitahaya Aging Diagnostic by Impedance/Capacitance Spectroscopy. *Food Analytical Methods, 8*(1), 126-129
- Cressey, D. (2016). The Plastic Ocean (Vol. 536, pp. 263-265). London: Nature.
- Da Róz, A. L., Veiga-Santos, P., Ferreira, A. M., Antunes, T. C. R., Leite, F. d. L., Yamaji, F. M., & Carvalho, A. J. F. d. (2016). Water Susceptibility and Mechanical Properties of Thermoplastic Starch–Pectin Blends Reactively Extruded with Edible Citric Acid. *Materials Research, 19*(1), 138-142
- Dang, K. M., & Yoksan, R. (2015). Development of thermoplastic starch blown film by incorporating plasticized chitosan. *Carbohydr Polym, 115*, 575–581
- Dang, K. M., & Yoksan, R. (2016). Morphological characteristics and barrier properties of thermoplastic starch/chitosan blown film. *Carbohydr Polym, 150*, 40-47
- Dashipour, A., Razavilar, V., Hosseini, H., Shojaee-Aliabadi, S., German, J. B., Ghanati, K., Khakpour, M., & Khaksar, R. (2015). Antioxidant and antimicrobial carboxymethyl cellulose films containing Zataria multiflora essential oil. *Int J Biol Macromol, 72*, 606-613
- Day, B. P. F. (2001). Active Packaging brand – a fresh approach. *the journal of brand technology, 1* (1), 32–41
- Day, B. P. F. (2008). Active Packaging of Food. In J. Kerry & P. Butler (Eds.), *Smart Packaging Technologies for fast moving consumer goods* (pp. 1-17). UK: John Wiley & Sons, Ltd.
- Day, B. P. F., & Potter, L. (2011). Active Packaging. In C. Richard & K. Mark (Eds.), *Food and Beverage Packaging Technology* (Second ed.). UK: Blackwell Publishing Ltd. .
- de Moraes, J. O., Scheibe, A. S., A., B., C., M., & Laurindo, J. B. (2015). Conductive drying of starch-fiber films prepared by tape casting: Drying rates and film properties. *LWT - Food Science and Technology, 64*(1), 356-366
- de Moraes, J. O., Scheibe, A. S., Sereno, A., & Laurindo, J. B. (2013). Scale-up of the production of cassava starch based films using tape-casting. *Journal of Food Engineering, 119*(4), 800-808

- Detduangchan, N., Sridach, W., & Wittaya, T. (2014). Enhancement of the properties of biodegradable rice starch films by using chemical crosslinking agents. *International Food Research Journal*, 2(3), 1225-1235
- Ding, C., Zhang, M., & Li, G. (2015). Preparation and characterization of collagen/hydroxypropyl methylcellulose (HPMC) blend film. *Carbohydr Polym*, 119, 194–201
- Du, W.-X., Roberto, J. A.-B., Hua, S. S. T., & McHugh, T. H. (2011). Antimicrobial volatile essential oils in edible films for food safety. *Science against microbial pathogens: communicating current research and technological advances*, 1124-1135
- Dumaol, O. S. R., Alaras, L. B., Karen G. Dahilan, Depadua, S. A. A., & Pulmone, C. J. G. (2010). In Vitro Activity of Pandan (*Pandanus amaryllifolius*) Leaves Crude Extract Against Selected Bacterial Isolates. *JPAIR Multidisciplinary Journal*, 102-124
- Ebrahimi, H., Afshar Najafi, F. S., Shahabadi, S. I. S., & Garmabi, H. (2015). A response surface study on microstructure and mechanical properties of poly(lactic acid)/thermoplastic starch/nanoclay nanocomposites. *Journal of Composite Materials*, 50(2), 269-278
- Edwin, S. M., Leonard, G., & Elijah, G. (2012). Isolation And Identification Of Essential Oils From Cymbopogon Citratus (Stapf) DC Using GC-MS And FT-IR. *Chemistry and Materials Research*, 2(4), 13
- Emiroğlu, Z. K., Yemiş, G. P., Coşkun, B. K., & Candoğan, K. (2010). Antimicrobial activity of soy edible films incorporated with thyme and oregano essential oils on fresh ground beef patties. *Meat Science*, 86, 283–288
- Enil Klarmann, Vladimira, A. S., & Gates, L. W. (1933). The Alkyl Derivatives of Halogen Phenols and their Bactericidal Action. I. Chlorophenols. *J. Bacteriol.*, 17(437)
- Factfish. (2017). Malaysia: Cassava, production quantity (tons) <http://www.factfish.com/statistic-country/malaysia/cassava,+production+quantity>.
- Falcão, M. A., Fianco, A. L. B., Lucas, A. M., Pereira, M. A. A., Torres, F. C., Vargas, R. M. F., & Cassel, E. (2012). Determination of antibacterial activity of vacuum distillation fractions of lemongrass essential oil. *Phytochemistry Reviews*, 11(4), 405-412

- Fu, X., Feng, F., & Huang, B. (2006). Physicochemical characterization and evaluation of a microemulsion system for antimicrobial activity of glycerol monolaurate. *Int J Pharm*, 321(1-2), 171-175
- Fu, Z.-q., Wang, L.-j., Li, D., Wei, Q., & Adhikari, B. (2011). Effects of high-pressure homogenization on the properties of starch-plasticizer dispersions and their films. *Carbohydr Polym*, 86(1), 202-207
- Gabor, D., & Tita, O. (2012). Biopolymers used in Food Packaging: A Review. *Acta Universitatis Cibiniensis Series E: Food Technology*, 16(2), 1-17
- Gao, C., Pollet, E., & Averous, L. (2017). Innovative plasticized alginate obtained by thermo-mechanical mixing: Effect of different biobased polyols systems. *Carbohydr Polym*, 157, 669-676
- García, N. L., L., R., A., D., M., A., & S., G. (2011). Effect of glycerol on the morphology of nanocomposites made from thermoplastic starch and starch nanocrystals. *Carbohydr Polym*, 84(1), 203-210
- García, N. L., Ribba, L., Dufresne, A., Aranguren, I. M., & Goyanes, S. (2009). Physico-Mechanical Properties of Biodegradable Starch Nanocomposites. *Macromolecular Materials and Engineering*, 294(3), 169-177
- Gardini, D., Deluca, M., Nagliati, M., & Galassi, C. (2010). Flow properties of PLZTN aqueous suspensions for tape casting. *Ceramics International*, 36(5), 1687-1696
- Garlotta, D. (2001). A Literature Review of Poly(Lactic Acid). *Journal of Polymers and the Environment*, 9(2), 63-85
- Georgieva, R., Yocheva, L., Tserovska, L., Zhelezova, G., Stefanova, N., Atanasova, A., Danguleva, A., Ivanova, G., Karapetkov, N., Rumyan, N., & Karaivanova, E. (2015). Antimicrobial activity and antibiotic susceptibility of *Lactobacillus* and *Bifidobacterium* spp. intended for use as starter and probiotic cultures. *Biotechnol Biotechnol Equip*, 29(1), 84-91
- Geyer, R., Jambeck, J. R., & Law, K. L. (2017). Production, use and fate of all plastics ever made. *Science Advances*, 3(7)
- Ghabraie, M., Vu, K. D., Tata, L., Salmieri, S., & Lacroix, M. (2016). Antimicrobial effect of essential oils in combinations against five bacteria and their effect on sensorial quality of ground meat. *LWT - Food Science and Technology*, 66, 332-339

- Ghasemlou, M., Aliheidari, N., Fahmi, R., Shojaee-Aliabadi, S., Keshavarz, B., Cran, M. J., & Khaksar, R. (2013). Physical, mechanical and barrier properties of corn starch films incorporated with plant essential oils. *Carbohydr Polym*, 98(1), 1117-1126
- Gill, P., Tohidi Moghadam, T., & Ranjbar, B. ((2010)). Differential Scanning Calorimetry Techniques: Applications in Biology and Nanoscience. *Journal of Biomolecular Techniques*(21), 167–193
- Goldstein, J. I., Dale, E. N., Joseph, R. M., Nicholas, W. M. R., John, H. J. S., & David, C. J. (2017). *Scanning Electron Microscopy and X-Ray Microanalysis* (Vol. 4). Newyork, USA: Springer.
- Gontard, N., Guilbert, S., & Cuq, J.-L. (1993). Water and Glycerol as Plasticizers Affect Mechanical and Water Vapor Barrier Properties of an Edible Wheat Gluten Film. *Journal of Food Science*, 58(1), 206-212
- Guo, M., Jin, T. Z., & Yang, R. (2014). Antimicrobial Polylactic Acid Packaging Films against *Listeria* and *Salmonella* in Culture Medium and on Ready-to-Eat Meat. *Food and Bioprocess Technology*, 7(11), 3293-3307
- Gurmeet, S., & Amrita, P. (2015). Unique pandanus - Flavour, food and medicine. *Journal of Pharmacognosy and Phytochemistry*, 5(3), 8-14
- Gyawali, R., & Ibrahim, S. A. (2014). Natural products as antimicrobial agents. *Food Control*, 46, 412-429
- Han, J. H., & Floros, J. D. (1997). Casting Antimicrobial Packaging Films and Measuring Their Physical Properties and Antimicrobial Activity. *Journal of Plastic Film and Sheeting*, 13, 287-298
- Hannah, R., & Max, R. (2018). "Plastic Pollution". Retrieved 10/12/2018, from 'https://ourworldindata.org/plastic-pollution'
- Heras-Mozos, R., Muriel-Galet, V., López-Carballo, G., Catalá, R., Pilar, H.-M., & Rafael, G. (2019). Development and optimization of antifungal packaging for sliced pan loaf based on garlic as active agent and bread aroma as aroma corrector. *Int J Food Microbiol*, 290 42–48
- Hillocks, R. J., Thresh, J. M., & Bellotti, A. C. (2002). *Cassava Biology, Production and Utilization*, : CAB International Wallingford, UK
- Hou, C., Chen, Y., Chen, W., & Li, W. (2011). Microwave-assisted methylation of cassava starch with dimethyl carbonate. *Carbohydr Res*, 346(9), 1178-1181

- Ibrahim, N., Mohamad, K. A. W., Du, N. U., & Hanafi, I. (2017). Physical and Degradation Properties of Polylactic Acid and Thermoplastic Starch Blends – Effect of Citric Acid Treatment on Starch Structures. *BioResources*, 12(2), 3076-3087
- IEA. (2017). World Energy Outlook (WEO-2017) International Energy Agency <https://www.iea.org/statistics>.
- IMARC. (2018). Global Cassava (Tapioca) Starch Market Overview 2018, Demand by Regions, Share, Price Trends and Forecast to 2023.
- Imre, B., & Pukanszky, B. (2013). Compatibilization in bio-based and biodegradable polymer blends. *European Polymer Journal*, 49, 1215 –1233
- ISO-1666. (1996). Starch - Determination of moisture content - Oven drying method
- ISO-14855-1. (2012). Determination of the ultimate aerobic biodegradability of plastic materials under controlled composting conditions. Method by analysis of evolved carbon dioxide. General method.
- Issaadi, K., Habi, A., Grohens, Y., & Pillin, I. (2015). Effect of the montmorillonite intercalant and anhydride maleic grafting on polylactic acid structure and properties. *Applied Clay Science*, 107, 62-69
- Ivanic, F., Johec-Moskova, D., Janigova, I., & Chodak, I. (2017). Physical properties of starch plasticized by a mixture of plasticizers. *European Polymer Journal*, 93, 843-849
- Iwate, T. (2015). Biodegradable and Biobased Polymers: Future prospects of Eco-Friendly Plastics. *Angew. Chem. Int. Ed.*, 54, 2 – 8
- Jafari, N. K., & Sani, A. M. (2016). Chemical composition and antibacterial activity of essential oil from *Melissa officinalis* leaves. *ARPJ Journal of Agricultural and Biological Scienc*, 11(9), 367-372
- Jaramillo, C. M., Gutierrez, T. J., Goyanes, S., Bernal, C., & Fama, L. (2016). Biodegradability and plasticizing effect of yerba mate extract on cassava starch edible films. *Carbohydr Polym*, 151, 150-159
- Jeannine, B. L., Nivea, M. V., Rodolfo, M. C. D. S., Ana, M. Q. B. B., & Paulo, J. A. S. (2015). Mechanical properties of cassava starch films as affected by different plasticizers and different relative humidity conditions. *International Journal of Food Studies*, 4, 116–125
- Jeroen, J. G., Soest, v., Hulleman, S. H. D., Wit, D. d., & Vliegthart, J. F. G. (1996). Crystallinity in starch bioplastics. *Industrial Crops and Products*, 5, 11-22

- Jesionek, W., Majer-Dziedzic, B., & Choma, I. M. (2017). TLC–direct bioautography as a method for evaluation of antibacterial properties of *Thymus vulgaris* L. and *Salvia officinalis* L. essential oils of different origin. *Journal of Liquid Chromatography & Related Technologies*, 40(5-6), 292-296
- Jideani, V. A., & Vogt, K. (2016). Antimicrobial Packaging for Extending the Shelf Life of Bread-A Review. *Crit Rev Food Sci Nutr*, 56(8), 1313-1324
- Jiugao, Y., Ning, W., & Xiaofei, M. (2005). The Effects of Citric Acid on the Properties of Thermoplastic Starch Plasticized by Glycerol. *Starch - Stärke*, 57(10), 494-504
- Johnston, B., Jiang, G., Hill, D., Adamus, G., Kwiecien, I., Zieba, M., Sikorska, W., Green, M., Kowalczyk, M., & Radecka, I. (2017). The Molecular Level Characterization of Biodegradable Polymers Originated from Polyethylene Using Non-Oxygenated Polyethylene Wax as a Carbon Source for Polyhydroxyalkanoate Production. *Bioengineering (Basel)*, 4(3)
- Juansang, J., Puncha-Arnon, S., Uttapap, D., Puttanlek, C., Rungsardthong, V., & Watcharatewinkul, Y. (2017). Concentration of plasticizers applied during heat-moisture treatment affects properties of the modified canna starch. *Food Chem*, 221, 1587-1594
- Kaewtatip, K., & Thongmee, J. (2013). Effect of kraft lignin and esterified lignin on the properties of thermoplastic starch. *Materials & Design*, 49, 701-704
- Katz, F. (1998). New research in packaging. *Food Technol*, 52, 56
- Ke, T., & Sun, X. (2003). Melting Behavior and Crystallization Kinetics of Starch and Poly(lactic acid) Composites. *Journal of Applied Polymer Science*, 89, 1203–1210
- Khankruea, R., Pivsa-Art, S., Hiroyuki, H., & Suttiruengwong, S. (2014). Effect of chain extenders on thermal and mechanical properties of poly(lactic acid) at high processing temperatures: Potential application in PLA/Polyamide 6 blend. *Polymer Degradation and Stability*, 108, 232-240
- Kizkitza, G., Retegia, A., Alba, G., Arantxa, E., & Nagore, G. (2015). Starch and cellulose nanocrystals together into thermoplastic starch bionanocomposites. *Carbohydr Polym*, 117, 83–90
- Klangmuang, P., & Sothornvit, R. (2016). Barrier properties, mechanical properties and antimicrobial activity of hydroxypropyl methylcellulose-based

- nanocomposite films incorporated with Thai essential oils. *Food Hydrocolloids*, 61, 609-616
- Kodre, K., Attarde, S., Yendhe, P., Patil, R., & Barge, V. (2014). Differential Scanning Calorimetry: A Review. *Journal of Pharmaceutical Analysis*, 3(3), 11-22
- Kong, X., Zhu, P., Sui, Z., & Bao, J. (2015). Physicochemical properties of starches from diverse rice cultivars varying in apparent amylose content and gelatinisation temperature combinations. *Food Chemistry*, 172, 433-440
- Kumar, P. M., Irina, A., & Manjeet, S. C. (2002). Modeling water vapour, gas and solute transport through protein films. In A. Gennadios (Ed.), *Protein Based Films And Coatings* (pp. 329-337). New York, USA: CRC Press.
- Kumar, S. V., Sajeevkumar, V. A., & Sunny, K. (2018). The influence of bound water on the FTIR characteristics of starch and starch nanocrystals obtained from selected natural sources†. *Starch - Stärke*
- Kuorwel, K. K., Cran, M. J., Sonneveld, K., Miltz, J., & Bigger, S. W. (2011). Antimicrobial activity of biodegradable polysaccharide and protein-based films containing active agents. *J Food Sci*, 76(3), R90-R102
- Kuorwel, K. K., Cran, M. J., Sonneveld, K., Miltz, J., & Bigger, S. W. (2013). Migration of antimicrobial agents from starch-based films into a food simulant. *LWT - Food Science and Technology*, 50(2), 432-438
- Leire, S., Jordana, K. P., Mercedes, F., José Ignacio, E., Antxon, S., & Alejandro, J. M. (2016). Linear and non-linear rheological behavior of polypropylene_polyamide blends modified with a compatibilizer agent and nanosilica and its relationship with the morphology. *European Polymer Journal*, 83(10-21)
- Leon, P. B. M. J., & Leszek, M. (2009). Thermoplastic Starch A Green Material for Various Industries. In P. B. M. J. Leon & M. Leszek (Eds.). Germany: Wiley-VCH.
- Leroy, A., Ribeiro, S., Grossiord, C., Alves, A., Vestberg, R. H., Salles, V., Brunon, C., Gritsch, K., Grosogeat, B., & Bayon, Y. (2017). FTIR microscopy contribution for comprehension of degradation mechanisms in PLA-based implantable medical devices. *J Mater Sci Mater Med*, 28(6), 87
- Li, C., Hou, J., Huang, Z., Zhao, T., Xiao, L., Gao, G., Harnoode, C., & Dong, A. (2015). Assessment of 2,2,6,6-tetramethyl-4-piperidinol-based amine N-

- halamine-labeled silica nanoparticles as potent antibiotics for deactivating bacteria. *Colloids Surf B Biointerfaces*, 126, 106-114
- Li, X., Wang, C., Fei, L., Lili, Z., Qiang, Y., Jing, M., & Xinhua, L. (2015). Physicochemical properties of corn starch isolated by acid liquid and L-cysteine. *Food Hydrocolloids*, 44, 353-359
- Linyao, Z., Guiyan, Z., & Wei, J. (2016). Mechanical properties of biodegradable polylactide/poly(ether-block-amide)/thermoplastic starch blends: Effect of the crosslinking of starch. *Journal of Applied Polymer Science*, 1-7
- Liu, H., Xie, F., Yu, L., Chen, L., & Li, L. (2009). Thermal processing of starch-based polymers. *Progress in Polymer Science*, 34, 1348-1368
- Lomeli-Ramirez, M. G., Kestur, S. G., Manriquez-Gonzalez, R., Iwakiri, S., de Muniz, G. B., & Flores-Sahagun, T. S. (2014). Bio-composites of cassava starch-green coconut fiber: part II-Structure and properties. *Carbohydr Polym*, 102, 576-583
- Lopes, L. Q., Santos, C. G., Gende, L., Raffin, R. P., de Almeida Vaucher, R., & Santos, R. C. (2016). Evaluation of antimicrobial activity of glycerol monolaurate nanocapsules against American foulbrood disease agent and toxicity on bees. *Microb Pathog*, 97, 183-188
- López, O., Castillo, L., Zaritzky, N., Barbosa, S., Villar, M., & García, M. A. (2015). Talc Nanoparticles Influence on Thermoplastic Corn Starch Film Properties. *Procedia Materials Science*, 8, 338-345
- Lopez, O., Garcia, M. A., Villar, M. A., Gentili, A., Rodriguez, M. S., & Albertengo, L. (2014). Thermo-compression of biodegradable thermoplastic corn starch films containing chitin and chitosan. *LWT - Food Science and Technology*, 57(1), 106-115
- Lu, D. R., Xiao, C. M., & Xu, S. J. (2009). Starch-based completely biodegradable polymer materials. *Express Polymer Letters*, 3(6)
- Lu, Z., Dang, W., Zhao, Y., Wang, L., Zhang, M., & Liu, G. (2017). Toward high-performance poly(para-phenylene terephthalamide) (PPTA)-based composite paper via hot-pressing: the key role of partial fibrillation and surface activation. *RSC Advances*, 7(12), 7293-7302
- Lucera, A., Conte, A., & Del Nobile, M. A. (2016). Volatile Compounds Usage in Active Packaging Systems. 319-327

- Madivoli, E. S., Gitu, L., & Gumba, M. E. (2012). Isolation And Identification Of Essential Oils From *Cymbopogon Citratus* (Stapf) Dc Using Gc-MS And Ft-IR. *Chemistry and Materials Resear*, 2(4), 13-24
- Mahieua, A., Terriéa, B. C., & Youssef, B. (2015). Thermoplastic starch films and thermoplasti oxygen scavenging Influence of water content. *Industrial Crops and Products*, 72, 192–199
- Maizura, M., Fazilah, A., Norziah, M. H., & Karim, A. A. (2007). Antibacterial Activity and Mechanical Properties of Partially Hydrolyzed Sago Starch–Alginate Edible Film Containing Lemongrass Oil. *Journa of Food Science*, 72(6)
- Malhotra, B., Keshwani, A., & Kharkwal, H. (2015). Antimicrobial food packaging: potential and pitfalls. *Front Microbiol*, 6(611), 1-9
- Mali, S., Sakanaka, L. S., Yamashita, F., & Grossmann, M. V. E. (2005). Water sorption and mechanical properties of cassava starch films and their relation to plasticizing effect. *Carbohydr Polym*, 60(3), 283-289
- Maran, J. P., Sivakumar, V., Sridhar, R., & Thirugnanasambandham, K. (2013). Development of model for barrier and optical properties of tapioca starch based edible films. *Carbohydr Polym*, 92(2), 1335-1347
- Maree, J., Kamatou, G., Gibbons, S., Viljoen, A., & Van Vuuren, S. (2014). The application of GC–MS combined with chemometrics for the identification of antimicrobial compounds from selected commercial essential oils. *Chemometrics and Intelligent Laboratory Systems*, 130, 172-181
- Martin, A. H., Ferrer, A., Tyagi, P., Yin, Y., Salas, C., Pal, L., & Rojas, O. J. (2017). Nanocellulose in Thin Films, Coatings, and Plies for Packaging Applications: A Review. *BioResources*, 12(1), 2143-2233
- Martin, O., & Averous, L. (2001). Poly(lactic acid): plasticisation and properties of bidegradable multiphase systems. *Polymer*, 42, 6209-6219
- Martina, S. d. L., & Giovanni, F. (2016). Effects of nanoparticles on the morphology of immiscible polymer blends Challenges and opportunities. *European Polymer Journal*, 79, 198–218
- Martins, A. B., & Santana, R. M. (2016). Effect of carboxylic acids as compatibilizer agent on mechanical properties of thermoplastic starch and polypropylene blends. *Carbohydr Polym*, 135, 79-85

- Martucci, J. F., Gende, L. B., Neira, L. M., & Ruseckaite, R. A. (2015). Oregano and lavender essential oils as antioxidant and antimicrobial additives of biogenic gelatin films. *Industrial Crops and Products*, 71, 205-213
- Masmoudi, F., Atef, B., Mohamed, D., Jaziri, M., & Emna, A. (2016). Biodegradable packaging materials conception based on starch and polylactic acid (PLA) reinforced with cellulose. *Environ Sci Pollut Res*, 1-11
- McKeen, L. W. (2012). *Introduction to Plastics and Polymers*: Elsevier Inc.
- Medina, J. C., Gutierrez, T. J., Goyanes, S., Bernal, C., & Fama, L. (2016). Biodegradability and plasticizing effect of yerba mate extract on cassava starch edible films. *Carbohydr Polym*, 151, 150-159
- Mehyar, G. F., & Han, J. H. (2006). Physical and Mechanical Properties of High-amylose Rice and Pea Starch Films as Affected by Relative Humidity and Plasticizer. *Journal of Food Science*, 1-7
- Meimoun, J., Wiatz, V., RenSaint-Loup, Parcq, J., Favrelle, A., Bonnet1, F., & Zinck, P. (2017). Modification of Starch graft copolymerization. *Starch - Stärke*, 69, 1-23
- Menard, K. P. (1999). *Dynamic mechanical analysis, A Practical Introduction*. United States of America: CRC Press.
- Menner, A. S. M. (1996). Assessment of Biodegradability of Plastics under Simulated Composting Conditions in a Laboratory Test System. *International Biodeterioration and Biodegradation*, 85-92
- Menzel, C., Olsson, E., Plivelic, T. S., Andersson, R., Johansson, C., Kuktaite, R., Jarnstrom, L., & Koch, K. (2013). Molecular structure of citric acid cross-linked starch films. *Carbohydr Polym*, 96(1), 270-276
- Mesias, R., & Murillo, E. (2018). Hyperbranched polyester polyol modified with polylactic acid as a compatibilizer for plasticized tapioca starch/polylactic acid blends. *Polímeros*, 28(1), 44-52
- Mir, S. A., Shah, M. A., Naik, H. R., & Zargar, I. A. (2016). Influence of hydrocolloids on dough handling and technological properties of gluten-free breads. *Trends in Food Science & Technology*, 51, 49-57
- Miranda, J. M., Mondragón, A. C., Lamas, A., Roca-Saavedra, P., Ibarra, I. S., Rodríguez, J. A., Cepeda, A., & Franco, C. M. (2016). Effect of Packaging Systems on the Inactivation of Microbiological Agents. 107-116
- Mistler, R. E., & Twiname, E. R. (2000). *Tape Casting: Theory and Practice*: Wiley.

- Mittal, V., Akhtar, T., & Matsko, N. (2015). Mechanical, Thermal, Rheological and Morphological Properties of Binary and Ternary Blends of PLA, TPS and PCL. *Macromolecular Materials and Engineering*, 300(4), 423-435
- Mlalila, N., Hilonga, A., Swai, H., Devlieghere, F., & Ragaert, P. (2018). Antimicrobial packaging based on starch, poly(3-hydroxybutyrate) and poly(lactic-co-glycolide) materials and application challenges. *Trends in Food Science & Technology*, 74, 1-11
- Mohammadi, N. A., Moradpour, M., Saeidi, M., & Alias, A. K. (2013). Thermoplastic starches: Properties, challenges, and prospects. *Starch - Stärke*, 65(1-2), 61-72
- Müller, C. M. O., Laurindo, J. B., & Yamashita, F. (2009). Effect of cellulose fibers addition on the mechanical properties and water vapor barrier of starch-based films. *Food Hydrocolloids*, 23(5), 1328-1333
- Müller, C. M. O., Pires, A. T. N., & Yamashita, F. (2012). Characterization of Thermoplastic Starch/Poly(Lactic Acid) Blends Obtained by Extrusion and Thermopressing. *J. Braz. Chem. Soc.*, 23(3), 426-443
- Muller, J., González-Martínez, C., & Chiralt, A. (2017). Poly(lactic) acid (PLA) and starch bilayer films, containing cinnamaldehyde, obtained by compression moulding. *European Polymer Journal*, 95, 56-70
- Müller, P., Bere, J., Fekete, E., Moczo, J., Nagy, B., Kally, M., Gyarmati, B., & Pukanszky, B. (2016). Interactions, structure and properties in PLA plasticized starch blends. *Polymer*, 103, 9-18
- Müller, P., Imre, B., Bere, J., Móczó, J., & Pukánszky, B. (2015). Physical ageing and molecular mobility in PLA blends and composites. *Journal of Thermal Analysis and Calorimetry*, 122(3), 1423-1433
- Muller, P., Kapin, E., & Fekete, E. (2014). Effects of preparation methods on the structure and mechanical properties of wet conditioned starch/montmorillonite nanocomposite films. *Carbohydr Polym*, 113, 569-576
- Muriel-Galet, V., Cran, M. J., Bigger, S. W., Hernández-Muñoz, P., & Gavara, R. (2015). Antioxidant and antimicrobial properties of ethylene vinyl alcohol copolymer films based on the release of oregano essential oil and green tea extract components. *Journal of Food Engineering*, 149, 9-16
- Muthui, Z. W., Kamweru, P. K., Nderitu, F. G., Hussein, S. A. G., Ngumbu, R., & Njoroge, G. N. (2015). Poly lactic acid viscoelastic properties and their

- degradation compared with those of polyethylene. *International Journal of Physical Sciences*, 10(2), 568-575
- Nafchi, A. M., Moradpour, M., Saeidi, M., & Alias, A. K. (2013). Thermoplastic starches: Properties, challenges, and prospects. *Starch/Stärke*, 65, 61-72
- Naik, M. I., Fomda, B. A., Jaykumar, E., & Bhat, J. A. (2010). Antibacterial activity of lemongrass (*Cymbopogon citratus*) oil against some selected pathogenic bacteria. *Asian Pacific Journal of Tropical Medicine*, 535-538
- Nasri-Nasrabadi, B., Behzad, T., & Bagheri, R. (2014). Preparation and characterization of cellulose nanofiber reinforced thermoplastic starch composites. *Fibers and Polymers*, 15(2), 347-354
- NatureWorks-LLC. (2015). Sheet-Extrusion Processing Guide. 1-8
- Nawapat, D., & Thawien, W. (2013). Effect of UV-treatment on the properties of biodegradable rice starch films. *International Food Research Journal*, 20(3), 1313-1322
- NCCLS. (2003). Susceptibility Testing of Mycobacteria, Nocardiae, and Other Aerobic Actinomycetes; Approved Standard. *M24-A*, 23(18)
- Nezamzadeh, S. A., Ahmadi, Z., & Taromi, F. A. (2017). From microstructure to mechanical properties of compatibilized polylactide/thermoplastic starch blends. *Journal of Applied Polymer Science*, 1-9
- Nossa, T. S., Belgacem, N. M., Gandini, A., & Carvalho, A. J. F. (2015). Thermoreversible crosslinked thermoplastic starch. *Polymer International*, 64(10), 1366-1372
- Nur Ain, A. H., Zaibunnisa, A. H., Halimahton Zahrah, M. S., & Norashikin, S. (2013). An experimental design approach for the extraction of lemongrass (*Cymbopogon citratus*) oleoresin using pressurised liquid extraction (PLE). *International Food Research Journal*, 20(1), 451-455
- Nurzyńska-Wierdak, R., Bogucka-Kocka, A., & Szymczak, G. (2014). Volatile constituents of *Melissa officinalis* leaves determined by plant age. *Nat Prod Commun.*, 9(5), 703-706
- Ojeda, J. s. J., & Maria, D. (2012). Fourier Transform Infrared Spectroscopy for Molecular Analysis of Microbial Cells. *Methods in Molecular Biology.*, 881, 187-213

- Olivato, J. B., Grossmann, M. V. E., Bilck, A. P., & Yamashita, F. (2012). Effect of organic acids as additives on the performance of thermoplastic starch/polyester blown films. *Carbohydr Polym*, *90*(1), 159-164
- Olivato, J. B., Grossmann, M. V. E., Yamashita, F., Eiras, D., & Pessan, L. A. (2012). Citric acid and maleic anhydride as compatibilizers in starch/poly(butylene adipate-co-terephthalate) blends by one-step reactive extrusion. *Carbohydr Polym*, *87*(4), 2614-2618
- Olivato, J. B., Müller, C. M. O., Yamashita, F., Grossmann, M. V. E., & Nobrega, M. M. (2013). Study of the compatibilizer effect in the properties of starch / polyester blends. *Polímeros Ciência e Tecnologia*, *23*(3), 346-351
- Olsson, E., Hedenqvist, M. S., Johansson, C., & Jarnstrom, L. (2013). Influence of citric acid and curing on moisture sorption, diffusion and permeability of starch films. *Carbohydr Polym*, *94*(2), 765-772
- Ortega-Toro, R., Muñoz, A., Talens, P., & Chiralt, A. (2016). Improvement of properties of glycerol plasticized starch films by blending with a low ratio of polycaprolactone and/or polyethylene glycol. *Food Hydrocolloids*, *56*, 9-19
- Otgonzul, O. (2010). *Bioactive polymeric systems for food and medical packaging applications*. (Ph.D Doctoral Thesis), Tomas Bata University, Zlin, Czech Republic.
- Pallab, D., & Pankaj, T. (2017). Thermal degradation kinetics of plastics and model selection. *Thermochimica Acta*, *654* 191–202
- Park, J. W., Im, S. S., Kim, S. H., & Kim, Y. H. (2000). Biodegradable Polymer Blends of Poly(L-actic acid) and Gelatinized Starch. *Polymer Engineering and Sciece*, *40*(12), 2539-2551
- Peelman, N., Ragaerta, P., Meulenaer, B. D., Adons, D., Peeters, R., Ludwig Cardond, Impe, F. V., & Devlieghere, F. (2013). Application of bioplastics for food packaging. *Food Science and Technology*, *32*, 128-141
- Pérez-Pérez, C., Regalado-González, C., Barbosa-Rodríguez, J. R., Rodríguez-Rodríguez, C. A., & Villaseñor-Ortega, F. (2006). Incorporation of antimicrobial agents in food packaging films and coatings. *Advances in Agricultural and Food Biotechnology*, *2006*., 193-216
- Pervaiz, M., Oakley, P., & Sain, M. (2014). Extrusion of Thermoplastic Starch: Effect of “Green” and Common Polyethylene on the Hydrophobicity Characteristics. *Materials Sciences and Applications*, *05*(12), 845-856

- Peter, J. G., John, H., & Richard, B. (2001). *Electron Microscopy and Analysis* (Third edition ed.): Taylor and Francis.
- Pizzoli, A. P. d. O., Yamashita, F., Gonçalves, O. H., Shirai, M. A., & Leimann, F. V. (2017). The effect of gelatin amount on the properties of PLA/TPS/gelatin extruded sheets. *Polímeros*, 27(1), 27-34
- Prasad, P., & Kochhar, A. (2014). Active Packaging in Food Industry: A Review. *IOSR Journal of Environmental Science, Toxicology and Food Technology*, 8(5), 1-7
- Priyaa, B., Guptab, V. K., Pathaniab, D., & Singhac, A. S. (2014). Synthesis, characterization and antibacterial activity of biodegradable starch PVA composite films reinforced with cellulosic fibre. *Carbohydr Polym*, 109, 171–179
- Pushpadass, H. A., Marx, D. B., Wehling, R. L., & Hanna, M. A. (2009). Extrusion and Characterization of Starch Films. *Cereal Chemistry Journal*, 86(1), 44-51
- Raeisi, M., Tajik, H., Yarahmadi, A., & Sanginabadi, S. (2015). Antimicrobial Effect of Cinnamon Essential Oil Against Escherichia Coli and Staphylococcus aureus. *Health Scope*, 4(4)
- Ramírez, M. G. L., Muniz, G. I. B. d., Satyanarayana, K. G., Tanobe, V., & Iwakiri, S. (2010). Preparation and characterization of biodegradable composites based on Brazilian cassava starch, corn starch and green coconut fibers. *Revista Matéria*, 15(2), 330-337
- Ramirez, N. V., Sanchez-Soto, M., Illescas, S., & Gordillo, A. (2009). Thermal Degradation of Polyoxymethylene Evaluated with FTIR and Spectrophotometry. *Polymer-Plastics Technology and Engineering*, 48, 470–477
- Ranitha, M., Nour, A. H., Sulaiman, Z. A., Nour, A. H., & Thana Raj, S. (2014). A Comparative Study of Lemongrass (*Cymbopogon Citratus*) Essential Oil Extracted by Microwave-Assisted Hydrodistillation (MAHD) and Conventional Hydrodistillation (HD) Method. *International Journal of Chemical Engineering and Applications*, 5(2), 104-108
- Rasal, R. M., Janorkar, A. V., & Hirt, D. E. (2010). Poly(lactic acid) modifications. *Progress in Polymer Science*, 35(3), 338-356
- Ratnayake, W. S., Hoover, R., & Warkentin, T. (2002). Pea Starch: Composition, Structure and Properties – A Review. *Starch/Stärke*, 54, 217–234

- Ray, S. S., & Masami, O. (2003). Biodegradable Polylactide and Its Nanocomposites: Opening a New Dimension for Plastics and Composites. *Macromol. Rapid Commun.*, 24, 815-840
- Richard, A. G., & Kalra, B. (2017). Biodegradable polymers for the environment. *sciencemag.org*, 297
- Rindlav, A., H.D., S., Hullernan, & Gatenholm, P. (1997). Formation of starch films with varying crystallinity *Carbohydr Polym*, 34 25-30
- Robert, S., & Ing, K. (2012). Thermoplastic Starch. In P. A. El-Sonbati (Ed.), *Thermoplastic Elastomers*. Croatia: In Tech.
- Rocha, M., Ferreira, F. A., Souza, M. M., & Prentice, C. (2013). Antimicrobial films: a review. *Formatex*, 23-32
- Rodriguez-Gonzalez, F. J., Ramsay, B. A., & Favis*, B. D. (2004). Rheological and thermal properties of thermoplastic starch with high glycerol content. *Carbohydr Polym*, 58, 139-147
- Rodriguez, A., Batlle, R., & Nerin, C. (2007). The use of natural essential oils as antimicrobial solutions in paper packaging. Part II. *Progress in Organic Coatings*, 60, 33–38
- Rooney, M. (1995). Overview of active food packaging. In R. ML (Ed.), *Active food packaging*. Glasgow: Blackie Academic & Professional
- Roshafima, R. A., AbdulRahman, W. A. W., Kasmani, R. M., & Ibrahim, N. (2012). Starch Based Biofilms-for-Green-Packaging. *World Academy of Science, Engineering and Technology*, 6, 508-513
- Rostami, H., Kazemi, M., & Shafiei, S. (2012). Antibacterial activity of *Lavandula officinalis* and *Melissa officinalis* against some human pathogenic bacteria. *Asian Journal of Biochemistry*, 7(3), 133-142
- Salvia-Trujillo, L., Rojas-Graü, A., Soliva-Fortuny, R., & Martín-Belloso, O. (2015). Physicochemical characterization and antimicrobial activity of food-grade emulsions and nanoemulsions incorporating essential oils. *Food Hydrocolloids*, 43, 547-556
- Sánchez-González, L., Cháfer, M., Hernández, M., Chiralt, A., & González-Martínez, C. (2011). Antimicrobial activity of polysaccharide films containing essential oils.
- Sanyang, M. L., Sapuan, S. M., Jawaid, M., Ishak, M. R., & Sahari, J. (2016). Effect of plasticizer type and concentration on physical properties of biodegradable

- films based on sugar palm (*arenga pinnata*) starch for food packaging. *J Food Sci Technol*, 53(1), 326-336
- Schmitt, H., Guidez, A., Prashatha, K., Soulestin, J., Lacrampe, M. F., & Krawczak, P. (2015). Studies on the effect of storage time and plasticisers on the structural variations in thermoplastic starch. *Carbohydr Polym*, 115, 365-372
- Schmitz, C. S., de Simas, K. N., Santos, K., Joao, J. J., de Mello Castanho Amboni, R. D., & Amante, E. R. (2006). Cassava starch functional properties by etherification - hydroxypropylation. *International Journal of Food Science and Technology*, 41(6), 681-687
- Schultz, J. M. (2016). Overview and Prospects of Polymer Morphology *Polymer Morphology* (pp. 1-13): John Wiley & Sons, Inc.
- Schultz, J. M. (2016). Overview and prospects of polymer morphology. In Q. Guo (Ed.), *Polymer Morphology Principles, Characterisation and Processing* (pp. 1-13). Hoboken, New Jersey, USA: JohnWiley & Sons, Inc., .
- Seligra, P. G., Jaramillo, C. M., Fama, L., & Goyanes, S. (2016). Biodegradable and non-retrogradable eco-films based on starch-glycerol with citric acid as crosslinking agent. *Carbohydr Polym*, 138, 66-74
- Seo, M.-K., Lee, J.-R., & Park, S.-J. (2005). Crystallization kinetics and interfacial behaviors of polypropylene composites reinforced with multi-walled carbon nanotubes. *Materials Science and Engineering: A*, 404(1-2), 79-84
- Seo, M., Lee, J., & Park, S. (2005). Crystallization kinetics and interfacial behaviors of polypropylene composites reinforced with multi-walled carbon nanotubes. *Materials Science and Engineering: A*, 404(1-2), 79-84
- Seow, Y. X., Yeo, C. R., Chung, H. L., & Yuk, H. G. (2014). Plant essential oils as active antimicrobial agents. *Crit Rev Food Sci Nutr*, 54(5), 625-644
- Seydim, A. C., & Sarikus, G. (2006). Antimicrobial activity of whey protein based edible films incorporated with oregano, rosemary and garlic essential oils. *Food Research International*, 39, 639-644
- Shirahase, T. K., Yoichi Tominaga, Yoichi Asai, Shigeo , & Masao, S. (2006). Miscibility and hydrolytic degradation in alkaline solution of poly(L-lactide) and poly(methyl methacrylate) blends. *Polymer*, 47, 4839-4844
- Shirai, M. A., Grossmann, M. V., Mali, S., Yamashita, F., Garcia, P. S., & Muller, C. M. (2013). Development of biodegradable flexible films of starch and

- poly(lactic acid) plasticized with adipate or citrate esters. *Carbohydr Polym*, 92(1), 19-22
- Shirai, M. A., Juliana, B. O., Maria, V. E. G., Ivo, M. D., Carmen, M. O., Müller, & Fabio, Y. (2016). Poly lactic acid and thermoplastic starch sheets: effect of adipate esters on the morphological mechanical and barrier properties. *Polímeros*
- Shojaee-Aliabadi, S., Hosseini, H., Mohammadifar, M. A., Mohammadi, A., Ghasemlou, M., Hosseini, S. M., & Khaksar, R. (2014). Characterization of kappa-carrageenan films incorporated plant essential oils with improved antimicrobial activity. *Carbohydr Polym*, 101, 582-591
- Shojaee-Aliabadi, S., Hosseini, H., Mohammadifar, M. A., Mohammadi, A., Ghasemlou, M., Ojagh, S. M., Hosseini, S. M., & Khaksar, R. (2013). Characterization of antioxidant-antimicrobial kappa-carrageenan films containing Satureja hortensis essential oil. *Int J Biol Macromol*, 52, 116-124
- Silverajah, V. S., Ibrahim, N. A., Zainuddin, N., Yunus, W. M., & Hassan, H. A. (2012). Mechanical, thermal and morphological properties of poly(lactic acid)/epoxidized palm olein blend. *Molecules*, 17(10), 11729-11747
- Smith, B. (1999). *Infrared Spectral Interpretation A Systematic Approach*: CRC Press LLC.
- Sonkhaskar, Y. M., Choubey, A., Bhamra, A., Singhal, R., & Anurag, S. (2014). New-Design-Of-A-Plastic-Bottle-Crusher. *International journal of Scientific and Technoogy research*, 3(7), 61-63
- Souza, A. C., Benze, R., Ferrão, E. S., Ditchfield, C., Coelho, A. C. V., & Tadini, C. C. (2012). Cassava starch biodegradable films: Influence of glycerol and clay nanoparticles content on tensile and barrier properties and glass transition temperature. *Food Science and Technology*, 46 110-117
- Stading, M. (1998). Dynamic Mechanical Analysis of Biopolymer Films. *Annual Transactions of The Nordic Rheology Society*, 6, 147-150
- Su, Y.-l., Wang, J., & Liu, H.-z. (2002). FTIR Spectroscopic Study on Effects of Temperature and Polymer Composition on the Structural Properties of PEO-PPO-PEO Block Copolymer Micelles. *langmuir*, 18, 5370-5374
- Suet-Yen, S., Sin, L. T., Tee, T.-T., Bee, S.-T., Rahmat, A. R., Rahman, W. A. W. A., Tan, A.-C., & Vikhraman, M. (2013). Antimicrobial agents for food packaging applications. *Trends in Food Science & Technology*, 33(2), 110-123

- Sun, S., Liu, P., Ji, N., Hou, H., & Dong, H. (2017). Effects of low polyhydroxyalkanoate content on the properties of films based on modified starch acquired by extrusion blowing. *Food Hydrocolloids*, 72, 81-89
- Sung, C. T. B. (2017). Malaysians use 3 billion plastic shopping bags per year, so why is limiting or even banning their use still a grossly inadequate strategy <http://www.christopherteh.com/blog/2017/02/malaysia-plastic-shopping-bags/>.
- Supanchaiyamat, N., Shuttleworth, P. S., Yue, H.-B., Fernandez-Blazquez, J. P., Budarin, V. L., Hunt, A. J., Sikhon, C., & Chaengkham, S. (2017). Bio-based carbonaceous composite materials from epoxidised linseed oil, bio-derived curing agent and starch with controllable functionality. *RSC Adv.*, 7(7), 24282–2429
- Suppakul, P. (2004). *study of Antimicrobial Polymeric packaging Films Containing Basil Extracts*. Victoria University.
- Suppakul, P., Sonneveld, K., Bigger, S. W., & Miltz, J. (2011). Diffusion of linalool and methylchavicol from polyethylene-based antimicrobial packaging films. *LWT - Food Science and Technology*, 44(9), 1888-1893
- Szymońska, J., Targosz-Korecka, M., & Krok, F. (2009). Characterization of starch nanoparticles. *Journal of Physics: Conference Series*, 146, 012027
- Tako, M., Tamaki, Y., Teruya, T., & Takeda, Y. (2014). The Principles of Starch Gelatinization and Retrogradation. *Food and Nutrition Sciences*, 05(03), 280-291
- Tapia-Blácido, D. R., do Amaral Sobral, P. J., & Menegalli, F. C. (2011). Optimization of amaranth flour films plasticized with glycerol and sorbitol by multi-response analysis. *LWT - Food Science and Technology*, 44(8), 1731-1738
- Tawakkal, I. S. M. A., Cran, M. J., & Bigger, S. W. (2016). Release of thymol from poly(lactic acid)-based antimicrobial films containing kenaf fibres as natural filler. *LWT - Food Science and Technology*, 66, 629-637
- TC-WI. (2003). Chemical Analyses – Determination of dry matter and water content on a mass basis in sediment, sludge, soil, and waste – Gravimetric method.
- Tegene, D. Z., Segne, T. A., & Chebude, Y. (2016). Synthesis and Characterization of Starch Vernolates in Organic Solvents. *American Journal of Applied Chemistry*, 4(6), 212

- Teixeira, E. d. M., Curvelo, A. A. S., Corrêa, A. C., Marconcini, J. M., Glenn, G. M., & Mattoso, L. H. C. (2012). Properties of thermoplastic starch from cassava bagasse and cassava starch and their blends with poly (lactic acid). *Industrial Crops and Products*, 37(1), 61-68
- Tharazi, I., Sulong, A. B., Muhamad, N., Haron, C. H. C., Tholibon, D., Ismail, N. F., Radzi, M. K. F. M., & Razak, Z. (2017). Optimization of Hot Press Parameters on Tensile Strength for Unidirectional Long Kenaf Fiber Reinforced Polylactic-Acid Composite. *Procedia Engineering*, 184, 478-485
- Thomas, S., Grohens, Y., & Jyotishkumar, P. (2015). *Characterization of Polymer blend* Singapore: Markono PrintMedia Pte Ltd.
- Thunwall, M., Antal Boldizar, & Mikael, R. (2006). Compression Molding and Tensile Properties of Thermoplastic Potato Starch Materials. *Biomacromolecules*, 7, 981-986
- Tong, S. Y., Lim, P. N., Wang, K., & Thian, E. S. (2018). Development of a functional biodegradable composite with antibacterial properties. *Materials Technology Advanced Performance Materials*, 1-7
- Tongdeesoontorn, W., Mauer, L. J., Wongruong, S., Pensiri, S., & Pornchai, R. (2012). Mechanical and Physical Properties of Cassava Starch-Gelatin Composite Films. *International Journal of Polymeric Materials*, 61, 778-792
- Trombetta, D., Saija, A., Bisignano, G., Arena, S., Caruso, S., Mazzanti, G., Uccella, N., & Castelli, F. (2002). Study on the mechanisms of the antibacterial action of some plant a,b-unsaturated aldehydes. *Letters in Applied Microbiology*, 35, 285-290
- Tsai, W. C., Hedenqvist, M. S., Laiback, Å., Melin, H., Ngo, M., Trollsås, M., & Gedde, U. W. (2016). Physical changes and sorption/desorption behaviour of amorphous and semi-crystalline PLLA exposed to water, methanol and ethanol. *European Polymer Journal*, 76, 278-293
- Tsai, Y. C., Yu, M. L., El-Shazly, M., Beerhues, L., Cheng, Y. B., Chen, L. C., Hwang, T. L., Chen, H. F., Chung, Y. M., Hou, M. F., Wu, Y. C., & Chang, F. R. (2015). Alkaloids from *Pandanus amaryllifolius*: Isolation and Their Plausible Biosynthetic Formation. *J Nat Prod*, 78(10), 2346-2354
- Turner, R. J. (2017). Metal-based antimicrobial strategies. *Microb Biotechnol*, 10(5), 1062-1065

- Twinaime, E. R., & Mistler, R. E. (2001). Tape Casting and Lamination. In K. H. J. Buschow, R. W. Cahn, M. C. Flemings, B. Ilshner, E. J. Kramer, S. Mahajan & P. Veyssi re (Eds.), *Encyclopedia of Materials: Science and Technology* (pp. 9083-9088). Oxford: Elsevier.
- UFAO. (2012). Cassava Production in Malaysia (United Nations Food and Agriculture) https://www.quandl.com/data/UFAO/CR_CASS_MYS-Cassava-Production-in-Malaysia.
- Ullah, A., & Wu, J. (2012). Feather Fiber-Based Thermoplastics: Effects of Different Plasticizers on Material Properties. *Macromol. Mater. Eng*, 1-10
- van-Soest, J. J. G., Hulleman, S. H. D., de-Wit, D., & Vilegenthart, J. F. G. (1996). Changes in the mechanical properties of thermoplastic potato starch in relation with changes in B-type crystallinity. *Carbohydr Polym*, 29, 225-232
- Vermeiren, L., Devliegher, F., Van Beest, M., de Kruijf, N., & Debevere, J. (1999). Developments in the active packaging of foods. *Trends in Food Science & Technology*, 10, 77-86
- Versino, F., L pez, O. V., & Garc a, M. A. (2015). Sustainable use of cassava (*Manihot esculenta*) roots as raw material for biocomposites development. *Industrial Crops and Products*, 65, 79-89
- Wang, L.-F., & Rhim, J.-W. (2016). Grapefruit seed extract incorporated antimicrobial LDPE and PLA films: Effect of type of polymer matrix. *LWT - Food Science and Technology*, 74, 338-345
- Wang, N., Yu, J., Chang, P. R., & Ma, X. (2008). Influence of formamide and water on the properties of thermoplastic starch/poly(lactic acid) blends. *Carbohydr Polym*, 71(1), 109-118
- Wang, N., Yu, J., & Ma, X. (2008). Preparation and characterization of compatible thermoplastic dry starch/poly(lactic acid). *Polymer Composites*, 29(5), 551-559
- Wang, S., Zhang, X., Wang, S., & Copeland, L. (2016). Changes of multi-scale structure during mimicked DSC heating reveal the nature of starch gelatinization. *Sci Rep*, 6, 28271
- Wang, Y., Zhang, L., Li, X., & Gao, W. (2011). Physicochemical Properties of Starches from Two Different Yam (*Dioscorea Opposita* Thunb.) Residues. *Braz. Arch. Biol. Technol*, 54, 243-251

- Wang, Y., Zheng, Q., Li, W., Ma, Y., Zhao, X., & Zhang, C. (2018). Measurement of free water in foods by secondary derivative thermogravimetry. *Journal of food*, *16*(1), 438–443
- Waterschoot, J., Gomand, S. V., Fierens, E., & Delcour, J. A. (2015). Production, structure, physicochemical and functional properties of maize, cassava, wheat, potato and rice starches. *Starch - Stärke*, *67*(1-2), 14-29
- Wattanakornsiri, A., Pachana, K., Kaewpirom, S., Sawangwong, P., & Migliaresi, C. (2013). Green composites of thermoplastic corn starch and recycled paper cellulose fibers. *Songklanakarin J. Sci. Technol.*, *33*(4), 461-467
- Wiegand, I., Hilpert, K., & Hancock, R. E. W. (2008). Agar and broth dilution methods to determine the minimal inhibitory concentration (MIC) of antimicrobial substances. *Nature Protocols*, *3*, 163
- Wittcoff, H. A., Reuben, B. G., & Plotkin, J. S. (2013). *Industrial Organic Chemicals* (Third Edition ed.). Hoboken, New Jersey: John Wiley & Sons, Inc.
- Wu, S., Lv, G., & Lou, R. (2012). Applications of Chromatography Hyphenated Techniques in the Field of Lignin Pyrolysis.
- Yan qun, L., De-xin, K., & Hong, W. (2013). Analysis and evaluation of essential oil components of cinnamon barks using GCMS and FTIR spectroscopy. *Industrial Crops and Products* *41* (2013) 269– 278, 41
- Yang, Y., Tang, Z., Xiong, Z., & Zhu, J. (2015). Preparation and characterization of thermoplastic starches and their blends with poly lactic acid. *Int J Biol Macromol*, *77* 273–279
- Yu, T., Jie, R., Shumao, L., Hua, Y., & Yan, L. (2010). Effect of fiber surface-treatments on the properties of poly(lactic acid)/ramie composites. *Composites Part B: Engineering*, *41*, 499-505
- Zain, A. H. M., Kahar, A. W. M., & Noriman, N. Z. (2016). Chemical-Mechanical Hydrolysis Technique of Modified Thermoplastic Starch for Better Mechanical Performance. *Procedia Chemistry*, *19*, 638-645
- Zainua, Z. A., & Songip, A. R. (2017). Policies, challenges and strategies for Municipal waste management in Malaysia. *JOSTIP*, *3*(1), 18-23
- Zhang, B., Fengwei, X., Tianlong, Z., Ling, C. L., Rowan, W. T., Peter, J. H., Julia, L. S., Tony, M., & Robin, D. R. (2016). Different characteristic effects of ageing on starch-based films plasticised by 1-ethyl-3-methylimidazolium acetate and by glycerol. *Carbohydr Polym*, *146*, 67–79

- Zhang, Y., Liu, X., Wang, Y., Jiang, P., & Quek, S. (2016). Antibacterial activity and mechanism of cinnamon essential oil against *Escherichia coli* and *Staphylococcus aureus*. *Food Control*, 59, 282-289
- Zhang, Y., Liu, Z., & Han, J. (2008). Starch-based edible films *Environmentally compatible food packaging* (pp. 108-136). USA.
- Zhang, Y., & Rempel, C. (2012). Retrogradation and Antiplasticization of Thermoplastic Starch.
- Zhang, Y., Rempel, C., & Liu, Q. (2014). Thermoplastic starch processing and characteristics-a review. *Crit Rev Food Sci Nutr*, 54(10), 1353-1370
- Zhen-Xiu, Z., Tao, Z., Xin, Z., Zhenxiang, X., Xu, D., & Prakashan, K. (2016). Mechanically stable superhydrophobic polymer films by a simple hot press lamination and peeling process. *RSC Advances*, 6(15), 12530-12536
- Zhu, F. (2015). Review Composition structure physicochemical properties and modifications of cassava starch. *Carbohydr Polym*, 122, 456-480
- Zia, K. M., Noreen, A., Zuber, M., Tabasum, S., & Mujahid, M. (2016). Recent developments and future prospects on bio-based polyesters derived from renewable resources: A review. *Int J Biol Macromol*, 82, 1028-1040
- Zullo, R., & Lannace, S. (2009). The effects of different starch sources and plasticizers on film blowing of thermoplastic starch: Correlation among process, elongational properties and macromolecular structure. *Carbohydrate Polymers*, 77, 376-383

