



Effects of imidacloprid-sodium chloride association on survival and reproduction of the stink bug *Podisus nigrispinus*

Efectos de la asociación de imidacloprid-cloruro de sodio en la supervivencia y reproducción del chinche *Podisus nigrispinus*

Gabryele Silva Ramos¹, Paula Daiana de Paulo², Pedro F. S. Toledo³, Khalid Haddi^{4†},
Jose Cola Zanuncio⁵, Eugenio E. Oliveira⁶

ARTICLE DATA

¹ Ph.D. Plant Protection Department. Universidade Estadual de São Paulo, Botucatu, SP, Brasil. gabryele.sr@gmail.com

² Ph.D. Entomology Department. Universidade Federal de Viçosa, Viçosa, MG, Brasil. paula.paulo@ufv.br

³ MSc. Entomology Department, Universidade Federal de Viçosa, Viçosa, MG, Brasil. pedro.toledo@ufv.br

^{4†} Dr. Entomology Department, Universidade Federal de Lavras, Lavras, MG, Brasil. khalid.haddi@ufla.br

⁵ Ph.D. Entomology Department, Universidade Federal de Viçosa, Viçosa, MG, Brasil. zanuncio@ufv.br

⁶ Ph.D. Entomology Department, Universidade Federal de Viçosa, Viçosa, MG, Brasil. eugenio@ufv.br

Cite: Ramos, G., De Paulo, P., Toledo, P., Haddi, K., Zanuncio, J., Oliveira, E. (2019). Effects of imidacloprid-sodium chloride association on survival and reproduction of the stink bug *Podisus nigrispinus*. *Revista de Ciências Agrícolas*. 36(E): 71-81. doi: <https://doi.org/10.22267/rcia.1936E.108>

Received: October 10 2019.

Accepted: October 20 2019.



ABSTRACT

Pesticide effects on natural enemies in an agroecosystem are of paramount importance for integrated pest management programs. Natural enemies can be subject to direct and indirect exposure to insecticides and synergistic molecules (e.g., sodium chloride - NaCl) which are used to control various pests of agricultural crops such as soybean. Here, we evaluated the potential effects of imidacloprid and its interaction with NaCl as an enhancer on the survival and reproductive abilities of the non-target predator *Podisus nigrispinus* (Dallas) (Hemiptera: Pentatomidae). The insects were exposed to the stink bugs control field recommended dose of imidacloprid associated or not with the salt at the concentration of 0.5% (w/v). NaCl as a pesticide enhancer did not affect the survival of *P. nigrispinus* adults after 48 h of exposure (less than 12% of mortality was always recorded). However, the fifth instar nymph mortality was almost 50%. The effects of imidacloprid on the reproductive parameters of *P. nigrispinus* included a decrease in the oviposition, showing fewer eggs per day. However, the fertility was not affected. The NaCl addition, therefore, had no effect on the mortality, survival, and reproduction of the non-target predator. The use of NaCl associated to imidacloprid and other pesticides in the presence of *P. nigrispinus* demonstrated compatibility, however, it requires further evaluation to endorse the set of these pest control strategies.

Keywords: Asopinae, biological control, ecotoxicology, IPM, pest control, selectivity.

RESUMEN

Los efectos de los pesticidas sobre los enemigos naturales en un agroecosistema son de suma importancia para los programas de manejo integrado de plagas. Los enemigos naturales pueden estar sujetos a la exposición directa e indirecta de insecticidas y moléculas sinérgicas (por ejemplo, cloruro de sodio - NaCl) que se utilizan para controlar diversas plagas de cultivos agrícolas como la soja. Aquí, evaluamos los posibles efectos del imidacloprid y su interacción con NaCl como un potenciador de la supervivencia y las capacidades reproductivas del depredador no objetivo *Podisus nigrispinus* (Dallas) (Hemiptera: Pentatomidae). Los insectos fueron expuestos a una tasa de campo de imidacloprid, recomendado para controlar los chinches en campos de soja. La solución de imidacloprid fue mezclada (o no) con NaCl (0.5%, w/v). El NaCl como potenciador de pesticidas no afectó la supervivencia de los adultos de *P. nigrispinus* después de 48 h de exposición (siempre se registró menos del 12% de mortalidad).

Sin embargo, la mortalidad de la ninfa del quinto estadio fue casi del 50%. Los efectos de imidacloprid sobre los parámetros reproductivos de *P. nigrispinus* incluyeron una disminución en la oviposición, mostrando menos huevos por día. Sin embargo, la fertilidad no se vio afectada. La adición de NaCl, por lo tanto, no tuvo efecto sobre la mortalidad, supervivencia y reproducción del depredador no objetivo. El uso de NaCl asociado a imidacloprid y otros pesticidas en presencia de *P. nigrispinus* demostró compatibilidad, sin embargo, requiere una evaluación adicional para respaldar el conjunto de estas estrategias de control de plagas.

Palabras clave: Asopinae, control biológico, ecotoxicología, MIP, control de plagas, selectividad.

INTRODUCTION

The soybean, *Glycine max* L., presents great economic importance in Brazil and the productivity of these plants is severely affected by insect pest attacks. These insects, besides decreasing productivity, substantially raise the cost of production due to pest management strategies which are of unquestioned need (Ávila and Grigolli, 2014; Peterson *et al.*, 2018).

Among other insect pests, the herbivory attacks delivered by a “stink bug complex”, which includes over three species of stink bugs, is considered one of the major barriers to the development of soybean crop fields in Brazil and other countries (Da Graça *et al.*, 2016). The management of such pests, for instance, the Neotropical stink bug, *Euchistus heros* (Fabr.) (Hemiptera: Pentatomidae) heavily relies on the use of synthetic pesticides, which includes, organophosphates, pyrethroids, organochlorides (endosulfan) and neonicotinoids (Panizzi *et al.*, 2012; Tuelher *et al.*, 2018).

The neonicotinoids, acting as agonists of acetylcholine receptors (Buckingham *et al.*, 1997; Nauen *et al.*, 2001), are among the most used pesticides in soybean fields and are frequently used by Brazilian farmers due to their high efficiency (Pazini *et al.*, 2019; Castellanos *et al.*, 2019). Furthermore, a common practice in Brazilian soybean fields consisting of adding sodium chloride (NaCl) at the concentration of 0.5% (w/v) to

insecticide solutions is, supposedly, believed to decrease the quantities of pesticide by half (50%) without losing efficiency (Corso, 1990; Panizzi *et al.*, 2007; Ávila and Grigolli, 2014).

It is believed that NaCl acts like a phagostimulant or arrestant molecule, which increases the insect contact (exposure period) with the pesticide solution or dry residues (Corso, 1990; Sosa-Gómez *et al.*, 1993; Niva and Panizzi, 1996; Corso and Gazzoni, 1998). The existence of salt action on the pentatomid stink bugs nervous system was confirmed by the presence of neuroreceptors sensitive to low concentrations of sodium chloride, causing a simultaneous attraction (Gr5a receptors) and deterrence (Gr66a receptors) behavior (Yarmolinsky *et al.*, 2009).

A couple of recent works tried to better elucidate the actions of imidacloprid (Haddi *et al.*, 2016; Santos *et al.*, 2016), but the synergic effects of NaCl added to some pesticides are not fully explained and still need further investigations that address the reasons underlying the aforementioned synergism as well as the reasons why it occurs only with some pesticides.

The use of biological control and integrated pest management programs in soybean fields, especially for the control of *Anticarsia gemmatalis* (Hübner, 1818) (Lepidoptera: Noctuidae), and some other stink bugs have also had favorable results (Ramiro *et al.*, 1986). Predatory stink bugs, e.g., *Podisus nigrispinus*, Hemiptera: Pentatomidae,

naturally occur in the Neotropical region (in many crop fields, including soybean) and are considered very important natural enemies of defoliator caterpillars (Pires *et al.*, 2006, Holtz *et al.*, 2019).

These predatory insects delivery their free of costs ecological services aiding in the management of soybean pests and have the potential to be introduced to enhance the power of other pest control strategies, in cases of augmentative biological control uses (Desneux *et al.*, 2007; Batalha *et al.*, 2012; He *et al.*, 2012; Naranjo *et al.*, 2015; Bueno *et al.*, 2017). However, the impacts of commonly used conventional pesticides and their enhancers (e.g., NaCl) have been completely neglected for predatory stink bugs. Hence, it is of extreme importance to study the potential selectivity/compatibility of these compounds to natural enemies.

Thus, this study aimed to investigate the potential effects including side effects on the reproductive abilities of the neonicotinoid imidacloprid associated to NaCl which is highly recommended for controlling the Neotropical stink bug *E. heros* on the predatory stink bug *P. nigrispinus*.

MATERIAL AND METHODS

Insect rearing

***Podisus nigrispinus*.** Eggs of *Podisus nigrispinus* were field-collected and kept in Petri dishes (9cm diameter) with distilled water (soaked cotton wool) to provide humidity. After egg hatching, immature stages were transferred to cages (30x15cm) made by wood (frame) and veil cloth (coverage). They were fed with *Tenebrio molitor* L. (Coleoptera: Tenebrionidae) pupae. Additionally, *Eucalyptus* spp. leaves were provided by setting a small branch inside the

cage. The stem of the branches was kept inside a glass flask (15 mL of volume capacity) filled with water to keep them hydrated. The stock colony was kept under controlled conditions of temperature ($25 \pm 2^\circ\text{C}$), photoperiod (12h scotophase), and humidity ($70 \pm 10\%$) (Lemos *et al.*, 2003).

Bioassays

Acute toxicity against the predatory stink bug *Podisus nigrispinus*. Newly emerged fifth instar nymphs and adult females (< 72 hours) were exposed to dry residues of the recommended field rate ($4.2\mu\text{g a.i./cm}^2$) of imidacloprid to control *E. heros* in soybean fields (MAPA, 2018) with or without the addition of NaCl 0.5% (w/v). Distilled water added or not with NaCl was used as the control. Insecticide exposure was achieved by contact with glass surface impregnated with dried residues. Briefly, 2mL of insecticide solution of distilled water were applied to 250mL transparent glass jars (EME Equipment, Paulicéia, SP, Brazil), which were maintained under rotation until dry using a heavy-duty rotator (Roto-Torque model 7637, ColeParmer, Vernon Hills, IL, USA) to coat the inner walls of the jars with insecticide residue. The upper portion of each container was coated with Teflon PTFE (DuPont, Wilmington, DE, USA) to prevent insects from escaping. Ten insects, nymphs or adult females, were set inside the glass flasks and the exposure period was 48 hours. Insects were considered dead when did not move or cannot walk a distance corresponding to double the size of their own body for one minute. The experiments were kept under laboratory-controlled conditions of temperature $25 \pm 2^\circ\text{C}$ relative humidity $60 \pm 10\%$ and L:D photoperiod of 14:10h. Was performed at least nine replicates for each *P. nigrispinus* stage and treatment.

The survivor insects that came from the 48-hour exposure were kept individualized in

Petri dishes (diameter: 90 mm) bottom-side covered with filter paper. Insects were fed with pupae of *T. molitor*, *Eucalyptus* spp. leaves (7cm stem containing three or four leaves) and distilled water (soaked cotton wool). The experiments were kept under laboratory-controlled conditions of temperature $25 \pm 2^\circ\text{C}$ relative humidity $60 \pm 10\%$ and L:D photoperiod of 14:10h. Was performed 88 replicates each insect was considered one replicate.

Sublethal effects on the reproduction of *Podisus nigrispinus* females. Newly emerged adults of *P. nigrispinus* (24h) were exposed individually to dry residues of the recommended field rate ($4.2\mu\text{g a.i./cm}^2$) of imidacloprid to control *E. heros* in soybean fields (MAPA, 2018) with or without the addition of sodium chloride at the concentration of 0.5% (m/v) as described earlier. Distilled water was added with sodium chloride or not as control treatments. After 48 hours of exposure, couples of this predator were formed and set in Petri dish (\emptyset 90 mm x 15 mm of depth), fed every two days with one *Eucalyptus* spp. leaflet, two *T. molitor* pupae and distilled water on soaked cotton wools. Twelve replicates (each couple was considered one replicate) were performed per treatment. The number of eggs laid per female was daily evaluated and the eggs removed and placed in new Petri dishes with distilled water (soaked cotton wool) to provide humidity. The emergence of nymphs was also daily evaluated until the female death. The bioassay was kept under laboratory-controlled conditions of temperature $25 \pm 2^\circ\text{C}$ relative humidity $60 \pm 10\%$ and L:D photoperiod of 14:10h.

Statistical Analysis. The mortality data obtained were corrected for that occurred in the control treatment. The data were subject to a 1-way analysis of variance and when the assumptions of residual normality

and variance homogeneity were not met, the mortality was subjected to Kruskal-Wallis test at 5% probability. Survival analysis was subjected to Kaplan-Meier test (LogRank method) and the reproductive parameters data (daily egg and nymphs numbers) were respectively submitted to Tukey and Kruskal-Wallis tests ($P < 0.05$) using SigmaPlot version 12.5 software (Systat Software, San Jose, CA, USA).

RESULTS AND DISCUSSION

Toxicity to the fifth instar of *Podisus nigrispinus*. The exposure of fifth instar nymphs of *P. nigrispinus* to $4.2\mu\text{g a.i./cm}^2$ (which is equivalent to the recommended field rate to control *E. heros*) with or without the addition of NaCl differed from the control group ($H = 21.35$; $P < 0.001$). Nymph mortality occasioned by the neonicotinoid imidacloprid (associated or not to NaCl) was lower than 50%. The average mortalities by treatments with the field dose and the field dose plus salt were $36.3 \pm 13.49\%$ and $42.3 \pm 14.38\%$, respectively. Treatments containing only water and water plus salt (0.5% c/c) caused mortality of $5.7 \pm 0.07\%$ and $10.1 \pm 0.08\%$, respectively (Figure 1A).

Toxicity to adult females of *Podisus nigrispinus*. The mortality rate of adult females of *P. nigrispinus* exposed to $4.2\mu\text{g a.i./cm}^2$ of imidacloprid (associated or not to NaCl) did not differ from the control group ($H = 2.3$; $P = 0.51$). Mortality of adult females of *P. nigrispinus* was lower than 12% in all treatments. The mortality average in the treatments containing the field dose and the field dose plus salt was $9.0 \pm 4.65\%$ for both. Treatment containing only water caused mortality of $6.3 \pm 2.42\%$ and water plus salt (0.5% c/c) $4.8 \pm 2.42\%$ (Figure 1B).

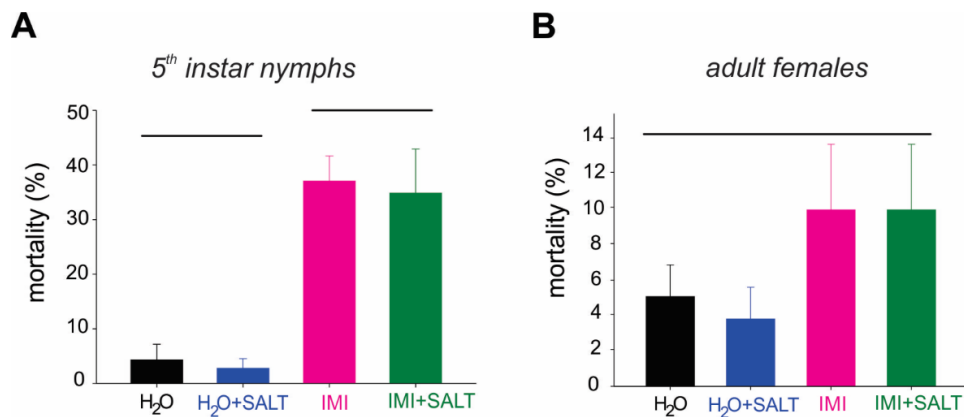


Figure 1. Mortality of fifth instar nymphs **(A)** and adult females **(B)** of *Podisus nigrispinus* after 48-hour of exposure to 4.2 µg a.i./cm² of imidacloprid (associated or not to NaCl).

Survival of exposed *Podisus nigrispinus*.

Survival of *P. nigrispinus* adults did not differ between unexposed and imidacloprid-exposed insects (*Log Rank* = 7.5; *P* = 0.06). The median survival time ranged from 15.1 to 20.7 days. The survival averages

of *P. nigrispinus* adults were 20.7 ± 2.43; 20.3 ± 2.24; 16.1 ± 1.4 and 14.9 ± 0.98 days in the treatments using the field dose of imidacloprid, imidacloprid with the salt addition, only water and water with salt (0.5% w/v), respectively (Figure 2).

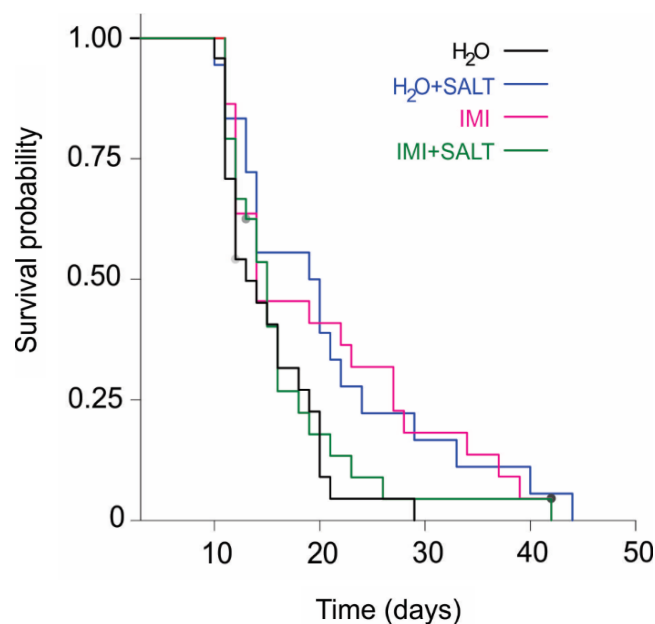


Figure 2. Survival curves of *Podisus nigrispinus* adults after 48 hours of exposure to 4.2 µg a.i./cm² of imidacloprid (associated or not to NaCl) using the Kaplan-Meier method and compared using the log-rank test (*Log Rank* _{df=3} = 7.51; *P* = 0.057).

Sublethal effects on the reproduction of *Podisus nigrispinus* females. Fecundity and fertility of *P. nigrispinus* females were similar between treatments without insecticide (Figures 3 and 4). On the contrary, imidacloprid associated or not to NaCl reduced the total (Figure 3A) and daily

(Figure 3B) fertility of this natural enemy ($H = 21.4$; $P < 0.001$). This reduction in fecundity caused by the pesticide exposure affected the total (Figure 4A) and daily nymph hatching (Figure 4B) which differed from the control with water ($H = 9.80$; $P = 0.014$) and water plus salt ($H = 9.80$; $P = 0.016$).

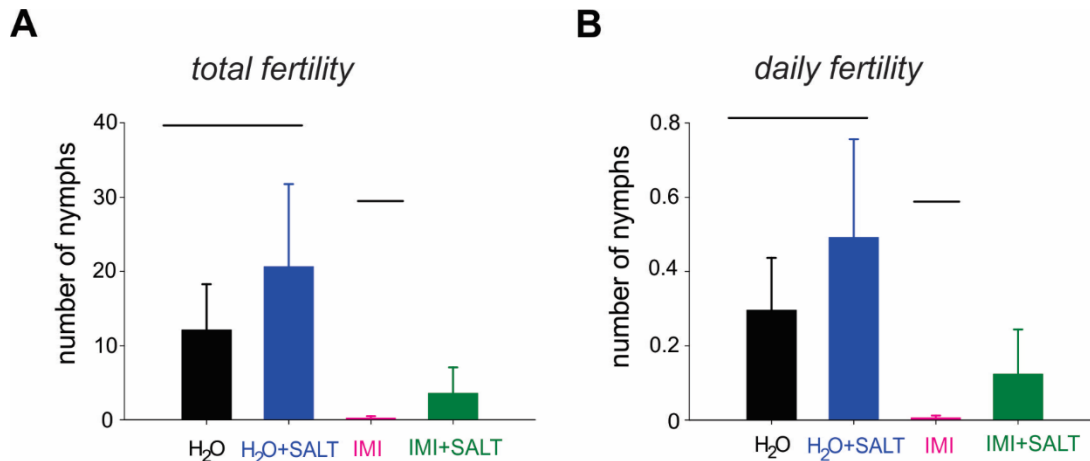


Figure 3. Fertility of adult females of *Podisus nigrispinus* after 48-hour exposure to 4.2 µg i.a./cm² of imidacloprid (associated or not to NaCl). **(A)** Fertility graphic based on the total number of eggs and **(B)** Fertility graphic of daily number of eggs hatched.

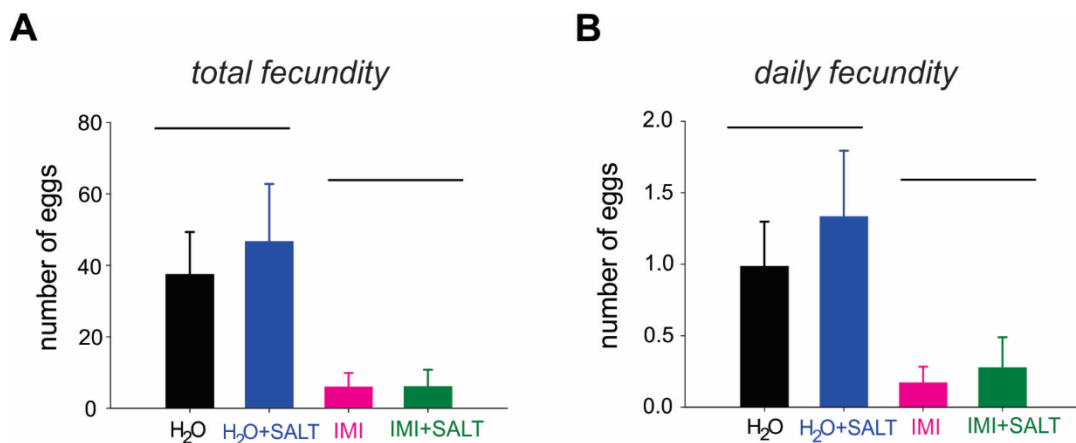


Figure 4. Fecundity of *Podisus nigrispinus* adults after 48 hours of exposure to 4.2 µg i.a./cm² of imidacloprid (associated or not to NaCl). **(A)** Fecundity graphic of total nymphs' number hatched and **(B)** Fecundity graphic of daily nymphs' number hatched.

Studying the potential impacts of conventional pesticides and their synergists (e.g., NaCl) against beneficial organisms is of unquestioned need. Although neonicotinoid insecticides have been thriving on the market due to their power to control insect pests, they have received attention regarding their side effects on non-target organisms e.g., pollinating bees and natural enemies and it becomes a more pessimistic scenario in the presence of pesticide enhancers.

In the present study, we have demonstrated that imidacloprid solutions (regardless of salt addition) increased the mortality of fifth instar nymphs of the predatory stink bug *P. nigrispinus*. On the other hand, those pesticide solutions neither caused mortality nor affected the survival of *P. nigrispinus* adults. A higher survival of *P. nigrispinus* adults compared to its nymphal stages fourth and fifth instars after exposure to pesticides such as imidacloprid, pyriproxyfen and neem oil has been demonstrated elsewhere (Zanuncio *et al.*, 2016). The increased susceptibility of insect immature stages may reflect a lower concentration of detoxifying enzymes (esterases) as well as the cuticle composition (Guedes *et al.*, 1992; Zanuncio *et al.*, 2001). Hence, our results reinforced that immature stages of this predatory stink bug are more susceptible to pesticides.

The mixture of thiamethoxan, cypermethrin, and NaCl as an enhancer allowed the control of *Piezodorus guildinii*, *Nezara viridula* and *E. heros* species for longer periods (up to 15 days) (Ramiro *et al.*, 2005). The NaCl is a synergistic agent in insecticide solutions due to the preference of pest insects for areas treated with low NaCl concentrations (Corso and Gazzoni, 1998; Marcomini *et al.*, 2016). This synergist causes shifts on the behavior of insect pests through an arresting effect which makes them more dynamic on the plant, increasing insect exposure to insecticides

(Corso *et al.*, 1990). These effects may result of a neuronal response in the expression of Gr5a receptors stimulated on the presence of low NaCl concentrations and Gr66a which results in aversion to NaCl (Yarmolinsky *et al.*, 2009). Such response may be absent in *P. nigrispinus* due to its different life history traits the differential confirmation of its site of action, as well as the presence of tarsal sensilla that may alter the toxicodynamic of the NaCl (Focks *et al.*, 2018).

Toxic effects caused by imidacloprid may affect the nutrient uptake, disaggregation, digestive physiology and reproductive abilities and behavior of the non-target predator (Magalhães *et al.*, 2002; Malaquias *et al.*, 2014; Martínez *et al.*, 2019). The neonicotinoids had been reported as molecules that did not affect the oviposition and egg hatch of treated female adults of *Podisus* sp. and showing lower mortality and survival impacts compared with to residual toxicity of pyrethroid molecules (De Cock *et al.*, 1996; Tillman and Mullinix, 2004; Torres and Ruberson, 2004; Pereira *et al.*, 2005). These results suggest that the viability of the eggs was not affected by the imidacloprid with or without salt but the fertility of this predator was reduced in agreement with results for pesticides reducing mature oocytes and increasing the maturation period of *P. nigrispinus* (Reis *et al.*, 2018).

The use of salt as a synergist favors a reduction in the amount of insecticide applied to soybean crops, reducing the farms expenditure on inputs (i.e., increasing profit), as well as causing a lower environmental impact. Interestingly, the addition of the enhancer (i.e., NaCl) did not impact any of the parameters evaluated here for *P. nigrispinus*. The fecundity data show that the non-target predator is susceptible to imidacloprid, but not to the addition of salt. The treatments that contained the field dose

of this insecticide presented lower number of nymphs compared to that with water or water with salt. Even though imidacloprid increases the mortality and impacts the fertility and fecundity of *P. nigrispinus*, the addition of NaCl which supposedly increase the strength of pesticides against stink bug pests did not impact the evaluated parameters. Sublethal doses of imidacloprid showed effects on the medium intestine of *P. nigrispinus*, affecting severally the digestion capacity consequently the predatory potential (Martínez *et al.*, 2019).

CONCLUSIONS

In summary, we assessed the lethal effects of the field dose of imidacloprid (with and without association with NaCl) used to control *E. heros* towards the pentatomid predator *P. nigrispinus*. Therefore, the synergistic agent NaCl may be used against pests in the presence of *P. nigrispinus* as the ingestion of this Asopinae predator was not affected by the salt but sublethal effects deserves attention. Thus, our results reinforce that the tandem use of NaCl as an enhancer with imidacloprid do not increase the problems associated with the pesticide against the predatory stink bug, *P. nigrispinus*. However, this interaction should be carefully investigated to endorse its use in more sustainable soybean IPM programs.

ACKNOWLEDGMENT

To the Brazilian agencies “Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq)”, Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES- Finance Code 001)”, “Fundação de Amparo à Pesquisa do Estado de Minas Gerais (FAPEMIG)”, and “Programa Cooperativo sobre Proteção Florestal (PROTEF) do Instituto de Pesquisas e Estudos Florestais (IPEF)” for financial support.

Conflict of interest: The authors declare that they have no conflict of interest.

BIBLIOGRAPHIC REFERENCES

- Ávila, C.B. & Grigolli, J.F.J. (2014) Pragas de soja e seu controle. Tecnologia e Produção: Soja 2013/2014. *Fundação MS*. Recovered from <http://www.fundacaoms.org.br/tecnologia-producao-soja-2013-2014>.
- Batalha, V.C., Zanuncio, J.C., Picanco, M. & Guedes, R.N.C. (2012). Selectivity of insecticides to *Podisus nigrispinus* (Heteroptera: Pentatomidae) and its prey *Spodoptera frugiperda* (Lepidoptera: Noctuidae). *Ceiba*, 38(1), 19-22.
- Buckingham, S., Lapied, B., Corronc, H. & Sattelle, F. (1997). Imidacloprid actions on insect neuronal acetylcholine receptors. *The Journal of Experimental Biology*, 200(Pt 21), 2685–2692. Recovered from <http://www.ncbi.nlm.nih.gov/pubmed/9326496>.
- Bueno, A. de F., Carvalho, G.A., Dos Santos, A.C., Sosa-Gómez, D.R. & Da Silva, D.M. (2017). Pesticide selectivity to natural enemies: challenges and constraints for research and field recommendation. *Ciência Rural*, 47(6): 2-10. doi: <https://doi.org/10.1590/0103-8478cr20160829>
- Castellanos, N.L., Haddi, K., Carvalho, G.A., De Paulo, P.D., Hirose, E., Guedes, R.N.C., Smagghe, G. & Oliveira, E.E. (2019). Imidacloprid resistance in the Neotropical brown stink bug *Euschistus heros*: selection and fitness costs. *Journal of Pest Science*, 92(2): 847–860. doi: <https://doi.org/10.1007/s10340-018-1048-z>
- Corso, I.C. (1990). Uso do sal de cozinha na redução da dose de inseticida para controle de percevejos. *Embrapa Soja - Comunicado Técnico (INFOTECA-E)*, 7p. Recovered from <https://www.infoteca.cnptia.embrapa.br/infoteca/bitstream/doc/445306/1/45.pdf>
- Corso, I.C. & Gazzoni, D.L. (1998). Sodium chloride: An insecticide enhancer for controlling pentatomids on soybeans. *Pesquisa Agropecuária Brasileira*, 33(10): 1563 - 1571.

- Da Graça, J.P., Ueda, T.E., Janegitz, T., Vieira, S.S., Salvador, M.C., De Oliveira, M.C.N., Zingaretti, S.M., Powers, S.J., Pickett, J.A., Birkett, M.A. & Hoffmann-Campo, C. B. (2016). The natural plant stress elicitor cis-jasmone causes cultivar-dependent reduction in growth of the stink bug, *Euschistus heros* and associated changes in flavonoid concentrations in soybean, *Glycine max*. *Phytochemistry*, 131: 84–91. doi: <https://doi.org/10.1016/j.phytochem.2016.08.013>.
- De Cock, A., De Clercq, P., Tirry, L. & Degheele, D. (1996). Toxicity of Diafenthiuron and Imidacloprid to the Predatory Bug *Podisus maculiventris* (Heteroptera: Pentatomidae). *Environmental Entomology*, 25(2): 476–480. doi: <https://doi.org/10.1093/ee/25.2.476>.
- Desneux, N., Decourtye, A. & Delpuech, J.M. (2007). The Sublethal Effects of Pesticides on Beneficial Arthropods. *Annual Review of Entomology*, 52(1): 81–106. doi: <https://doi.org/10.1146/annurev.ento.52.110405.091440>.
- Focks, A., Belgers, D., Boerwinkel, M.C., Buijse, L., Roessink, I. & Van den Brink, P.J. (2018). Calibration and validation of toxicokinetic-toxicodynamic models for three neonicotinoids and some aquatic macroinvertebrates. *Ecotoxicology*, 27(7): 992–1007. doi: <https://doi.org/10.1007/s10646-018-1940-6>.
- Guedes, R.N.C., Lima, J.O.G. & Zanuncio, J.C. (1992) Seletividade dos inseticidas deltametrina, fenvalerato e fenitrotiom para *Podisus connexivus* Bergroth, 1891 (Heteroptera: Pentatomidae). *Anais da Sociedade Entomologica do Brasil*, 21: 339-346.
- Haddi, K., Mendes, M.V., Barcellos, M.S., Lino-Neto, J., Freitas, H.L., Guedes, R.N.C. & Oliveira, E. E. (2016). Sexual Success after stress? Imidacloprid-Induced Hormesis in Males of the Neotropical Stink Bug *Euschistus heros*. *PLoS ONE*, 11(6):e0156616. doi: <https://doi.org/10.1371/journal.pone.0156616>.
- He, Y., Zhao, J., Zheng, Y., Desneux, N. & Wu, K. (2012). Lethal effect of imidacloprid on the coccinellid predator *Serangium japonicum* and sublethal effects on predator voracity and on functional response to the whitefly *Bemisia tabaci*. *Ecotoxicology*, 21(5): 1291–1300. doi: <https://doi.org/10.1007/s10646-012-0883-6>.
- Holtz, A.M., Marinho-Prado, J.S., Pallini, A., Pires, A.A., Cofler, T.P., Rocha, C.M., & Pazianotto, R.A.A. (2019). Host plant and the predator *Podisus nigrispinus*: when the defense compounds of the plant affect the third trophic level. *Entomologia Experimentalis et Applicata*, 167(4): 306 -312. doi: <https://doi.org/10.1111/eea.12774>
- Lemos, W.P., Ramalho, F.S., Serrao, J.E. & Zanuncio, J.C. (2003). Effects of diet on development of *Podisus nigrispinus* (Dallas) (Het., Pentatomidae), a predator of the cotton leafworm. *Journal of Applied Entomology*, 127(7): 389 - 395. doi: <https://doi.org/10.1046/j.1439-0418.2003.00765.x>
- Magalhães, L.C., Guedes, R.N.C., Oliveira, E.E., & Tuelher, E.S. (2002). Desenvolvimento e Reprodução do Predador *Podisus distinctus* (Stal) (Heteroptera: Pentatomidae) frente a doses subletais de permetrina. *Neotropical Entomology*, 31(3): 445-448. doi: <https://doi.org/10.1590/S1519-566X2002000300015>.
- Malaquias, J.B., Ramalho, F.S., Omoto, C., Godoy, W.A.C. & Silveira, R.F. (2014). Imidacloprid affects the functional response of predator *Podisus nigrispinus* (Dallas) (Heteroptera: Pentatomidae) to strains of *Spodoptera frugiperda* (J.E. Smith) on Bt cotton. *Ecotoxicology*, 23(2): 192–200. doi: <https://doi.org/10.1007/s10646-013-1162-x>.
- Marcomini, M.C., Cremonez, P.S.G & Neves, P.M.O.J. (2016). Efeito do NaCl no comportamento alimentar de *Euschistus heros* (Hemiptera: Pentatomidae). *Anais Do Encontro Anual De Iniciação Científica Da UEL - ISSN 2447-4118*. Recovered from http://www.uel.br/eventos/eaic/anais/?content=2016/anais_resumo.php&cod_artigo=564.
- Martínez, L.C., Plata-Rueda, A., Gonçalves, W.G., Freire, A.F.P.A., Zanuncio, J.C., Bozdoğan, H. & Serrão, J.E. (2019). Toxicity and cytotoxicity of the insecticide imidacloprid in the midgut of the predatory bug, *Podisus nigrispinus*. *Ecotoxicology and Environmental Safety*, 167: 69 - 75. doi: <https://doi.org/10.1016/j.ecoenv.2018.09.124>.
- Ministério Da Agricultura, Pecuária E Abastecimento – MAPA. (2018). *AGROSTAT - Estatísticas de Comércio Exterior do*

- Agronegócio Brasileiro*. Recovered from <http://agrostat.agricultura.gov.br>.
- Naranjo, S.E., Ellsworth, P.C. & Frisvold, G.B. (2015). Economic Value of Biological Control in Integrated Pest Management of Managed Plant Systems. *Annual Review of Entomology*, 60(1): 621 - 645. doi: <https://doi.org/10.1146/annurev-ento-010814-021005>.
- Nauen, R., Ebbinghaus-Kintscher, U. & Schmuck, R. (2001). Toxicity and nicotinic acetylcholine receptor interaction of imidacloprid and its metabolites in *Apis mellifera* (Hymenoptera: Apidae). *Pest Management Science*. 57(7): 577-586. doi: <https://doi.org/10.1002/ps.331>.
- Niva, C.C. & Panizzi, A.R. (1996). Efeitos do cloreto de sódio no comportamento de *Nezara viridula* (L.) (Heteroptera: Pentatomidae) em vagens de soja. *Anais da Sociedade Entomológica do Brasil*. 25: 251-257.
- Panizzi, A.R., Bueno, A.F. & Silva, F.A.C. (2012). Insetos que atacam vagens e grãos. In: Bueno, A. de F., Pomari, A., Panizzi, A., Corrêa-Ferreira, B., Omoto, C., De Oliveira, C.M., Hoffmann-Campo, C.B., Ávila, C.J., Sosa-Gómez, D.R., Gazzoni, D.L., Hirose, E., Da Silva, F.A.C., Moscardi, F., Marcelino-Guimarães, F.C., Lorini, I., Corso, I.C., Salvadori, J.R., Oliveira, L.J., De Carvalho, M.C. da C., Fernandes, P.M., Bueno, R.C.O. de F., Roggia, S. & Yano, S.A.C. *Soja: manejo integrado de insetos e outros artrópodes-praga*. pp. 335-420. Brasil: Embrapa.
- Panizzi, A.R., Kogan, M. & Jepson, P. (2007). Nutritional ecology of plant feeding arthropods and IPM. In: Kogan, M. & Jepson, P. *Perspectives in Ecological Theory and Integrated Pest Management*. pp. 170-222. Cambridge, UK: Cambridge University Press.
- Pazini, J. de B., Padilha, A.C., Cagliari, D., Bueno, F.A., Rakes, M., Zotti, M.J., Martins, J.F. da S. & Grützmaier, A.D. (2019). Differential impacts of pesticides on *Euschistus heros* (Hem.: Pentatomidae) and its parasitoid *Telenomus podisi* (Hym.: Platygasteridae). *Scientific Reports*, 9(1): 6544. doi: <https://doi.org/10.1038/s41598-019-42975-4>
- Pereira, A.I. de A., Ramalho, F. de S. & Zanuncio, J.C. (2005). Susceptibility of *Podisus nigrispinus* (Dallas) (Heteroptera: Pentatomidea) to gamma-cyhalothrin under laboratory conditions. *Scientia Agricola*. 62(5): 478-482. doi: <https://doi.org/10.1590/S0103-90162005000500012>
- Peterson, R.K.D., Higley, L.G. & Pedigo, L.P. (2018). Whatever Happened to IPM? *American Entomologist*. 64(3): 146 - 150. doi: <https://doi.org/10.1093/ae/tmy049>.
- Pires, E.M., Pinto, R., Lacerda, M.C., Zanuncio, J.C. & Fialho, M. do C.Q. (2006). Potencial reprodutivo horário do predador de lagartas desfolhadoras do eucalipto: *Podisus nigrispinus* (Heteroptera: Pentatomidae). *Revista Árvore*. 30(6): 1039-1044. doi: <https://doi.org/10.1590/S0100-67622006000600020>.
- Ramiro, Z.A., Batista Filho, A. & Cintra, E.R.R. (2005). Eficiência do inseticida Actara Mix 110 + 220 CE (thiamethoxam + cipermetrina) no controle de percevejos-pragas da soja. *Arquivos Do Instituto Biológico*. 72(2): 239-247.
- Ramiro, Z.A., Batista Filho, A. & Machado, L.A. (1986). Ocorrência de pragas e inimigos naturais em soja no Município de Orlândia, SP. *Anais da Sociedade Entomológica do Brasil*. 15 (2): 239-246.
- Reis, T.C., Soares, M.A., Dos Santos, J.B., Dos Santos, C.A., Serrão, J.E., Zanuncio, J.C. & Ferreira, E.A. (2018). Atrazine and nicosulfuron affect the reproductive fitness of the predator *Podisus nigrispinus* (Hemiptera: Pentatomidae). *Anais Da Academia Brasileira de Ciências*. 90(4): 3625-3633. doi: <https://doi.org/10.1590/0001-3765201820170748>
- Santos, M.F., Santos, R.L., Tomé, H.V.V., Barbosa, W.F., Martins, G.F., Guedes, R.N.C. & Oliveira, E.E. (2016). Imidacloprid-mediated effects on survival and fertility of the Neotropical brown stink bug *Euschistus heros*. *Journal of Pest Science*. 89(1): 231-240. doi: <https://doi.org/10.1007/s10340-015-0666-y>
- Sosa-Gómez, D.R., Takachi, C.Y. & Moscardi, F. (1993). Determinação de sinergismo e suscetibilidade diferencial de *Nezara viridula* (L.) e *Euschistus heros* (F.) (Hemiptera: Pentatomidae) à inseticidas em mistura com cloreto de sódio. *Anais da Sociedade Entomológica do Brasil*. 22: 569-576.

- Tillman, P.G. & Mullinix, B.G. (2004). Comparison of susceptibility of pest *Euschistus servus* and predator *Podisus maculiventris* (Heteroptera: Pentatomidae) to selected insecticides. *Journal of Economic Entomology*, 97(3): 800-806. doi: [https://doi.org/10.1603/0022-0493\(2004\)097\[0800:cosope\]2.0.co;2](https://doi.org/10.1603/0022-0493(2004)097[0800:cosope]2.0.co;2)
- Torres, J.B., & Ruberson, J.R. (2004). Toxicity of thiamethoxam and imidacloprid to *Podisus nigrispinus* (Dallas) (Heteroptera: Pentatomidae) nymphs associated to aphid and whitefly control in cotton. *Neotropical Entomology*. 33(1): 99-106. doi: <https://doi.org/10.1590/S1519-566X2004000100017>
- Tuelher, E.S., Da Silva, É.H., Rodrigues, H.S., Hirose, E., Guedes, R.N.C. & Oliveira, E.E. (2018). Area-wide spatial survey of the likelihood of insecticide control failure in the neotropical brown stink bug *Euschistus heros*. *Journal of Pest Science*. 91(2): 849 -859. doi: <https://doi.org/10.1007/s10340-017-0949-6>.
- Yarmolinsky, D.A., Zuker, C.S. & Ryba, N.J.P. (2009). Common Sense about Taste: From Mammals to Insects. *Cell*, 139(2): 234-244. doi: <https://doi.org/10.1016/j.cell.2009.10.001>
- Zanuncio, J.C., Molina-Rugama, A.J., Serrao, J. & Pratisoli, D. (2001). Nymphal Development and Reproduction of *Podisus nigrispinus* (Heteroptera: Pentatomidae) fed with combinations of *Tenebrio molitor* (Coleoptera: Tenebrionidae) pupae and *Musca domestica* (Diptera: Muscidae) larvae. *Biocontrol Science and Technology*. 11(3): 331-337. doi: <https://doi.org/10.1080/09583150120055736>.
- Zanuncio, J.C., Mourão, S.A., Martínez, L.C., Wilcken, C.F., Ramalho, F.S., Plata-Rueda, A., Soares, M.A. & Serrão, J. E. (2016). Toxic effects of the neem oil (*Azadirachta indica*) formulation on the stink bug predator, *Podisus nigrispinus* (Heteroptera: Pentatomidae). *Scientific Reports*. 6(1): 30261. doi: <https://doi.org/10.1038/srep30261>