

## RESEARCH ARTICLE

# Ocean acidification alters properties of the exoskeleton in adult Tanner crabs, *Chionoecetes bairdi*

Gary H. Dickinson<sup>1,†</sup>, Shai Bejerano<sup>1</sup>, Trina Salvador<sup>1</sup>, Christine Makdisi<sup>1</sup>, Shrey Patel<sup>1</sup>, W. Christopher Long<sup>2</sup>, Katherine M. Swiney<sup>\*2</sup>, Robert J. Foy<sup>2</sup>, Brittan V. Steffel<sup>3</sup>, Kathryn E. Smith<sup>4</sup> and Richard B. Aronson<sup>3</sup>

## ABSTRACT

Ocean acidification can affect the ability of calcifying organisms to build and maintain mineralized tissue. In decapod crustaceans, the exoskeleton is a multilayered structure composed of chitin, protein and mineral, predominately magnesian calcite or amorphous calcium carbonate (ACC). We investigated the effects of acidification on the exoskeleton of mature (post-terminal-molt) female southern Tanner crabs, *Chionoecetes bairdi*. Crabs were exposed to one of three pH levels – 8.1, 7.8 or 7.5 – for 2 years. Reduced pH led to a suite of body region-specific effects on the exoskeleton. Microhardness of the claw was 38% lower in crabs at pH 7.5 compared with those at pH 8.1, but carapace microhardness was unaffected by pH. In contrast, reduced pH altered elemental content in the carapace (reduced calcium, increased magnesium), but not the claw. Diminished structural integrity and thinning of the exoskeleton were observed at reduced pH in both body regions; internal erosion of the carapace was present in most crabs at pH 7.5, and the claws of these crabs showed substantial external erosion, with tooth-like denticles nearly or completely worn away. Using infrared spectroscopy, we observed a shift in the phase of calcium carbonate present in the carapace of pH 7.5 crabs: a mix of ACC and calcite was found in the carapace of crabs at pH 8.1, whereas the bulk of calcium carbonate had transformed to calcite in pH 7.5 crabs. With limited capacity for repair, the exoskeleton of long-lived crabs that undergo a terminal molt, such as *C. bairdi*, may be especially susceptible to ocean acidification.

**KEY WORDS:** Biomineralization, Climate change, Cuticle, Calcite, Amorphous calcium carbonate (ACC), Crustacea

## INTRODUCTION

Decapod crustaceans possess a multifunctional exoskeleton, which serves roles in feeding, defense, desiccation resistance and muscle attachment (Meyers et al., 2013; Meyers and Chen, 2014). The exoskeleton, or cuticle, is a multilayered, composite structure

(Chen et al., 2008; Fabritius et al., 2011, 2016; Meyers and Chen, 2014). From interior to exterior, the cuticle is composed of four structural layers: the membranous layer, the endocuticle, the exocuticle and the epicuticle (Travis, 1963; Roer and Dillaman, 1984). The membranous layer sits atop the hypodermis and is not mineralized (Roer and Dillaman, 1984; Fabritius et al., 2012). The endocuticle and exocuticle comprise the vast majority of the cuticle. These layers are composed of alpha-chitin chains, which are wrapped in protein and grouped into fibrils (Giraud-Guille, 1984; Sachs et al., 2006; Chen et al., 2008; Fabritius et al., 2011). Multiple fibrils bundle into chitin–protein fibers, which are then assembled into planes. Within the endocuticle and exocuticle, planes of fibers are stacked on top of one another, with each plane offset slightly with respect to the last, resulting in a Bouligand, or twisted-plywood, structure (Bouligand, 1972; Giraud-Guille, 1984; Raabe et al., 2006). Both the endocuticle and exocuticle layers are embedded with calcium salts, typically nanocrystalline magnesian calcite or amorphous calcium carbonate (Roer and Dillaman, 1984; Dillaman et al., 2005; Boßelmann et al., 2007). The outermost epicuticle is composed primarily of waxes and protein, interspersed with mineral aggregates (Hegdahl et al., 1977; Roer and Dillaman, 1984; Fabritius et al., 2012). The entire cuticle is shed periodically and replaced with newly formed cuticle during the process of ecdysis, which enables growth (Travis, 1963; Roer and Dillaman, 1984). In a portion of decapod species, juveniles undergo a terminal molt to maturity, after which time full replacement of the cuticle no longer occurs (Vogt, 2012).

Structure, elemental composition and mechanical properties of the decapod cuticle can vary among body regions (Boßelmann et al., 2007; Chen et al., 2008; Lian and Wang, 2011; Coffey et al., 2017; Steffel et al., 2019), among species (Boßelmann et al., 2007; Steffel et al., 2019; Rosen et al., 2020) and with environmental conditions (Taylor et al., 2015; Coffey et al., 2017; Glandon et al., 2018; Bednaršek et al., 2020). For example, in blue and red king crabs (*Paralithodes platypus* and *Paralithodes camtschaticus*, respectively), hardness of the claw is about twice that of the carapace, and calcium content is elevated in the claw in both species (Coffey et al., 2017). Long-term exposure to seawater with reduced pH (7.8 or 7.5) led to a 40% reduction in hardness of the claw endocuticle in blue king crabs and a 45% reduction in claw endocuticle hardness in red king crabs (Coffey et al., 2017). Hardness of the carapace was not affected by reduced pH, but exocuticle thickness was reduced in blue king crabs.

Sensitivity to the environment is particularly relevant within the context of ocean acidification (OA), the global-scale reduction in seawater pH that has resulted from elevated atmospheric  $P_{CO_2}$ . Since the Industrial Revolution, atmospheric  $P_{CO_2}$  has risen from ~280 ppm to over 410 ppm (IPCC, 2001; Raven, 2005; <http://www.esrl.noaa.gov/gmd/ccgg/trends/global.html>). Dissolution of  $CO_2$  in the world's oceans has reduced the pH of global surface

<sup>1</sup>Department of Biology, The College of New Jersey, 2000 Pennington Rd, Ewing, NJ 08628, USA. <sup>2</sup>NOAA, National Marine Fisheries Service, Alaska Fisheries Science Center, Resource Assessment and Conservation Engineering Division, Kodiak Laboratory, 301 Research Ct, Kodiak, AK 99615, USA. <sup>3</sup>Department of Ocean Engineering and Marine Sciences, Florida Institute of Technology, Melbourne, FL 32901, USA. <sup>4</sup>The Marine Biological Association, The Laboratory, Citadel Hill, Plymouth, PL1 2PB, UK.

\*Present address: NOAA, National Marine Fisheries Service, Southwest Fisheries Science Center, Fisheries Resources Division, 8901 La Jolla Shores Drive, La Jolla, CA 92037, USA.

†Author for correspondence (dickinga@tcnj.edu)

© G.H.D., 0000-0003-1073-1483; C.M., 0000-0002-4299-319X; W.C.L., 0000-0002-7095-1245; K.M.S., 0000-0001-5482-2710; B.V.S., 0000-0003-0477-8883; R.B.A., 0000-0003-0383-3844

waters by ~0.1 pH units since the Industrial Revolution, and based on projected CO<sub>2</sub> emissions scenarios, pH will drop an additional 0.3–0.5 units by the year 2200 (Caldeira and Wickett, 2003; Orr et al., 2005; Doney et al., 2009). At high latitudes, changes in seawater chemistry associated with OA are likely to be more extreme than at lower latitudes because of the higher solubility of CO<sub>2</sub> in colder waters and ocean mixing patterns (Fabry et al., 2009).

OA affects the ability of many calcifying marine organisms to build and maintain mineralized tissue (Doney et al., 2009; Kroeker et al., 2010, 2013; Sokolova et al., 2016). Reduced shell growth, shell dissolution, alterations in structure, and compromised biomechanical properties have been observed in a wide range of taxa (Orr et al., 2005; Ries et al., 2009; Byrne and Fitzer, 2019; Fitzer et al., 2019; Gaylord et al., 2019). Such changes may result from reduced pH and associated changes in acid–base homeostasis, and from the reduction in calcium carbonate saturation states ( $\Omega$ ) associated with OA (Ries et al., 2009; Roleda et al., 2012; Cyronak et al., 2016; Sokolova et al., 2016; Waldbusser et al., 2016). Within this body of literature, crustaceans are often reported to be less susceptible to OA than other mineralizing taxa (Ries et al., 2009; Kroeker et al., 2010, 2013; Sokolova et al., 2016; Byrne and Fitzer, 2019). Relatively high metabolic rates and iono/osmoregulatory capacity, protection of the site of mineralization by a waxy epicuticle, and the ability of crustaceans to employ bicarbonate within the mineralization process have all been cited as contributing to their success in tolerating OA (Wickins, 1984; Melzner et al., 2009; Ries et al., 2009; Whiteley, 2011; Sokolova et al., 2016). Systematic assessments of the effects of OA on the decapod cuticle, however, are relatively rare; most studies limit their assessments to gross calcification rates or calcium content (e.g. Ries et al., 2009; Page et al., 2017).

The southern Tanner crab, *Chionoecetes bairdi*, is an ecologically and commercially important brachyuran decapod that inhabits the North Pacific shelf, from Oregon to the Bering Sea in Alaska. After ~3 months as larvae, juveniles settle into benthic habitats and take ~5–6 years to reach maturity (Donaldson et al., 1981). Females have a terminal molt to maturity, after which they mate and extrude their first clutch of eggs. They then exhibit an annual reproductive cycle, hatching larvae in the late spring and extruding a new clutch shortly thereafter (Paul and Adams, 1984; Donaldson and Adams, 1989; Swiney, 2008). As there are no direct methods for determining the age of a decapod crustacean, it is not known how long females live after their terminal molt; however, in one study, 33% of the mature females in Cook Inlet had barnacles on them that were 3–4 years old, suggesting that many females live at least 5 years after the terminal molt (Paul and Paul, 1986). Because Tanner crabs live from the subtidal down to 440 m (Jadamec et al., 1999), the carbonate chemistry that crabs are exposed to *in situ* almost certainly varies considerably among individuals and stocks. In the Bering Sea, the pH at 70 m depth fluctuates seasonally from a high of about 8.2 from the autumn to the spring, to summer lows around 7.5 (Mathis et al., 2014). Crabs that live in shallower, seasonally less stratified waters, however, likely experience less dramatic pH swings. Previous OA studies with juvenile *C. bairdi* found a reduction in carapace width by 28% and an 11% reduction in calcium content of the carapace in individuals held at reduced pH (7.5) compared with crabs held under ambient pH (~8.0) (Long et al., 2013b). In adult Tanner crabs (the life stage assessed in the current study), exposure to pH 7.5 for 2 years resulted in a ~29% reduction in carapace calcium, compared with crabs at ambient pH (~8.1), and the carapaces of pH 7.5 crabs were ‘noticeably more pliable’ than those of crabs held at higher pH (Swiney et al., 2016).

The goal of this study was to assess the effect of OA on properties of the cuticle in mature southern Tanner crabs, *C. bairdi*. Crabs were held at one of three pH levels, ~8.1 (ambient), 7.8 or 7.5, for 2 years. Given that these crabs were past their terminal molt when the exposure began, potential differences in cuticle properties reflected the crabs’ ability to maintain or repair mineralized tissue. Specifically, we quantified cuticle micromechanical properties, thickness, structural integrity, elemental content and the phase or polymorph of calcium carbonate [i.e. whether calcite or amorphous calcium carbonate (ACC) was present]. Assessments for each individual crab were conducted separately in the carapace, which protects the internal organs, and right claw, which is employed in feeding and defense. This approach allowed us to determine whether the response to OA varies among body regions. Although mechanical properties of the decapod cuticle are sensitive to hydration (Hepburn et al., 1975; Joffe et al., 1975; Chen et al., 2008; Fabritius et al., 2011), the majority of studies on the decapod cuticle that have assessed mechanical properties at the micrometer scale have tested samples when dry (e.g. Chen et al., 2008; Sachs et al., 2006; Coffey et al., 2017). Hence, a secondary objective was to determine whether the hydration state of the cuticle affects micro-mechanical responses to OA. Differences in the mechanics, structure, elemental content or mineralogy of the cuticle after long-term exposure to reduced pH could affect cuticle functionality in these long-lived crabs because the post-terminal-molt cuticle is never fully replaced.

## MATERIALS AND METHODS

### Animal collection and experimental exposure

Collection of crabs, experimental exposures and seawater acidification are described in detail in Long et al. (2016) and Swiney et al. (2016). A total of 48 multiparous female adult southern Tanner crabs (*Chionoecetes bairdi* M. J. Rathbun 1924), of carapace width 98.7±4.8 mm (mean±s.d.), were caught in Chiniak Bay, Kodiak, AK, USA (57°43.25′N, 152°17.5′W; depth ~80 m) over a 5 week period in May and June of 2011. Crabs were held in ambient incoming seawater until the beginning of the experiment. Throughout the holding period, crabs were fed *ad libitum* on a diet of fish and squid. Crabs were randomly assigned to one of three pH levels, ~8.1 (unmodified surface-ambient), 7.8 and 7.5, for 2 years, from June 2011 to July 2013. The duration of the exposure was dictated by the need to capture two full reproductive cycles to examine both direct and carryover effects on the embryos and larvae (Long et al., 2016; Swiney et al., 2016); it represents an exposure time that is a substantial portion of the mature crab’s life expectancy.

Exposures were conducted at the Alaska Fisheries Science Center’s Kodiak Laboratory. Crabs were placed individually in 68 l tubs with 1 l min<sup>-1</sup> flow of water. Water temperature was allowed to vary to mimic seasonal conditions, except that it was chilled to 9°C during the warmest months of the summer to keep it within the range experienced by crabs *in situ*. Salinity was 31.22±0.47 psu (mean ±s.d.). Seawater was acidified using the method described by Long et al. (2013a). The method involved mixing ambient water (pumped into the laboratory from the Trident Basin at 15–26 m depth) with water from a super-acidified tank (pH 5.5, acidified via bubbling of CO<sub>2</sub>) within a head-tank for each treatment. Mixing within the pH 7.8 and 7.5 head-tanks was controlled using Honeywell controllers and Durafet III pH probes. The ambient treatment head-tank contained only ambient water with no input from the super-acidified tank. pH<sub>F</sub> (pH on the free proton scale) and temperature were measured daily in each tub using a Durafet III pH probe (precision ±0.03) calibrated daily with Tris buffer (Millero, 1986). Best

practice in carbonate chemistry measurements (Dickson et al., 2007) was followed throughout. Total alkalinity and dissolved inorganic carbon (DIC) were measured on water samples weekly as described in Swiney et al. (2016) per the methods in Dickson et al. (2007) and DOE (1994). Other carbonate chemistry variables were calculated in R (v2.14.0) using the seacarb package and the default constants (<http://CRAN.R-project.org/package=seacarb>). Target pH levels were achieved throughout the exposure (Table 1). Saturation state with respect to calcite ( $\Omega_{\text{Calcite}}$ ) decreased with decreasing pH and was <1 at pH 7.5.

Throughout the experimental exposure, each crab was examined daily and fed fish and squid in excess twice a week. Each of the three pH treatments included 16 randomly assigned crabs. Ultimately, 10 survived in the ambient treatment, six in the pH 7.8 treatment, and seven in the pH 7.5 treatment (for an analysis of the survival data, see Swiney et al., 2016). At the end of the 2 year exposure, the surviving crabs were killed by rapidly removing the carapace from the rest of the body. The right claw and a  $\sim 2.5 \text{ cm}^2$  portion of the carapace, cut from the posterior margin, were immediately frozen at  $-80^\circ\text{C}$  and shipped on dry ice to The College of New Jersey (TCNJ) for analysis. Four crabs that had died within the last 6 weeks of exposure but did not show any visible signs of exoskeletal decay were also included in the analyses; all were in the pH 7.8 treatment. All cuticle samples remained frozen during transit and, upon arrival, were kept at  $-70^\circ\text{C}$  until analysis.

### Sample preparation

To prepare samples for analysis, frozen samples were first cut to size using a water-cooled diamond band-saw (Gryphon C-40). Samples were placed on ice and kept as cold as possible during cutting. Carapace samples were cut into four strips, each about  $5 \times 25 \text{ mm}$ , for use in the assessments described below. For claw samples, the dactylus (movable finger) and pollex (fixed finger) were first cut from the manus of each claw. The entire dactyl was embedded in epoxy resin (see below), which was used for micromechanical and cuticle-thickness assessments. The pollex was further cut along its short axis to produce two segments. The first segment, consisting of  $\sim 4 \text{ mm}$  from the manus into the pollex, was used for  $\text{CaCO}_3$  polymorph assessments, whereas the remainder of the pollex was used for structural and elemental analysis. For all samples, any visible tissue adhering to the cuticle after cutting was carefully removed with forceps. Cut samples were lyophilized on a Yamato DC41-A lyophilizer for  $\sim 18 \text{ h}$  and then stored in a desiccator until use.

### Micromechanical properties

Samples of cuticle were embedded in epoxy resin, ground and polished for micromechanical assessments. Polishing of samples is

necessary to achieve the completely level and scratch-free surface necessary for microhardness testing; for irregularly shaped cuticle samples, this is only possible when the samples are embedded in epoxy. Embedding and polishing followed the method described by Coffey et al. (2017). Individual samples were affixed to the bottom of a 3.2 cm cylindrical mounting cup. Carapace samples were oriented in such a way that grinding and polishing would reveal a cross-section along the anterior–posterior axis, and they were positioned in the mounting cup using a plastic coil-clip. Dactyl samples were positioned with the long axis parallel to the bottom of the mounting cup using a small amount of cyanoacrylate glue (Loctite® Control Gel), producing a cross-section along the longitudinal axis upon grinding and polishing. Embedding cups were filled with a two-part epoxy (Allied High Tech, EpoxySet) and left to cure at room temperature for at least 18 h. Grinding and polishing were conducted on a manual grinding/polishing machine (Allied High Tech, M-Prep 5). Each sample was ground using a series of silicon carbide papers (180, 320, 600 and 800 grit) and then polished with a  $1 \mu\text{m}$  diamond suspension and a  $0.04 \mu\text{m}$  colloidal silica suspension. Samples were checked after polishing under a Jenco MET-233 metallurgical microscope and were repolished if necessary until completely flat and free of scratches. Polished samples were stored in a desiccator until testing.

Vickers microhardness was measured on a Mitutoyo HM-200 microhardness tester. Each sample was first tested dry and then hydrated and tested again when wet. For each sample and each hydration condition, a total of 12 indents were made within the endocuticle. During the initial round of testing (dry condition), indents were spread roughly evenly along the length of the cross-section, with a spacing of at least  $500 \mu\text{m}$  between indents. When samples were retested (hydrated condition), indents were placed in between those made during the first round of testing, resulting in a final spacing of at least  $250 \mu\text{m}$  between indents. To avoid potential edge-effects, indents were placed at least  $200 \mu\text{m}$  away from layer boundaries and other structural features. This spacing was only possible within the endocuticle layer. For dactyl samples, grinding/polishing of the roughly cone-shaped dactyl resulted in a V-shaped cross-section, with the upper and lower portion of the cuticle converging at the tip. Indents were made in both the upper and lower portion of the cuticle, but, as the dactyl tips were visibly damaged in some crabs, indents in the tip region were avoided. All indents were made at 20 g load, 5 s dwell time. Individual indents were measured directly on the hardness tester in two dimensions, and Vickers microhardness values were automatically calculated. Replicate indentations within the same sample and hydration condition were averaged to determine the mean microhardness for each sample.

Once all samples were tested in the dry condition, samples were hydrated by soaking in artificial seawater. Embedded samples were

**Table 1. Seawater chemistry parameters**

	pH 8.1 (ambient)	pH 7.8	pH 7.5
pH <sub>F</sub>	8.09±0.07	7.80±0.03	7.50±0.03
Temperature (°C)	5.00±1.54	4.94±1.54	4.93±1.53
$P_{\text{CO}_2}$ ( $\mu\text{atm}$ )	391.9±65.74	781.17±31.13	1597.15±62.76
DIC ( $\mu\text{mol kg}^{-1}$ SW)	2010.76±34.21	2082.18±38.29	2156.84±38.74
$\text{HCO}_3^-$ ( $\mu\text{mol kg}^{-1}$ SW)	1895.17±41.68	1989.7±37.25	2045.45±36.1
$\text{CO}_3^{2-}$ ( $\mu\text{mol kg}^{-1}$ SW)	94.72±16.26	50.89±2.99	26.3±1.54
Total alkalinity ( $\mu\text{mol kg}^{-1}$ SW)	2135.38±30.3	2119.25±36.29	2112.47±35.98
$\Omega_{\text{Calcite}}$	2.31±0.39	1.24±0.07	0.64±0.04

pH and temperature were measured daily ( $N=728$  per treatment). Dissolved inorganic carbon (DIC) and alkalinity were measured weekly ( $N=101$ – $104$  per treatment). Other parameters were calculated (see Materials and Methods). pH<sub>F</sub>, pH on the free proton scale;  $\Omega_{\text{Calcite}}$ , calcium carbonate saturation; SW, sea water. Data are means±s.d.

placed in a single layer in a plastic food storage container. The container was filled with artificial seawater (Instant Ocean, 35 psu) and the samples were soaked for ~72 h before testing in the wet condition. Samples were removed from seawater one at a time and briefly rinsed with deionized water to remove salts; visible droplets of water were removed from the sample surface using compressed air, and a series of 12 additional indents were made as described above. Indentations were made as quickly as possible once the sample was removed from the water (typically within 10 min) to prevent dehydration. Soaking of samples was conducted in small batches (4–6 samples per batch) to ensure that the amount of time in seawater was consistent among samples. Note that it was not possible to test microhardness in cuticle samples that had never been dried, because of the need to embed samples in moisture-sensitive epoxy (see above). In mineralized tissue samples, where direct comparisons have been made between samples that were rehydrated (as described here) and those that were never dried, no differences in micromechanical properties were observed between conditions (Baldassarri et al., 2008).

### Cuticle thickness and structural assessment

Total cuticle thickness, which includes thickness of the endocuticle, exocuticle and epicuticle (if visible), was quantified on the same embedded samples used for the micromechanical assessments. Each sample was imaged under a reflected light microscope (Zeiss AxioScope A1 with a Zeiss AxioCam 105 color camera). Thickness measurements were made on digital images using the camera's analysis software (Zeiss Zen 2), and at least 15 independent thickness measurements were made on each image. To determine measurement locations, a 350  $\mu\text{m}^2$  grid was placed on the digital image, and measurements were made each time the vertical grid lines crossed the sample. As in microhardness testing, replicate thickness measurements were made in both the upper and lower portions of the dactyl cross-section but were not made in the tip region. Replicate thickness measurements within the same sample were averaged to determine the mean total cuticle thickness for each sample.

Structural integrity of the cuticle was assessed semi-quantitatively using a stereomicroscope (Leica S8Apo with a Leica EC1 color camera). An unembedded segment of the carapace (cut as described in 'Sample preparation', above) and the pollex region of the claw were used for structural assessments. Images of the interior and exterior surfaces of the carapace and of the exterior surface of the pollex were taken of each sample at a range of magnifications. Images were compared side-by-side among treatments, and deviations among samples were documented. Specifically, on the interior of the carapace, the presence or absence of erosion was assessed. The carapace interior was typically smooth and pearly white, but in a portion of samples the interior was uneven with translucent patches, which appeared dark gray under the stereomicroscope and suggested erosion of the mineralized cuticle (see Results). On the carapace exterior, discoloration and broken, uneven or rough regions were documented. Signs of wear, resulting from prolonged abrasion, were documented and scored for pollex samples. Broken, damaged and pitted surfaces, including on the tooth-like denticles of the pollex, were noted. Four independent evaluators assessed images of the pollex from each crab and scored each as displaying minimal, moderate or extensive damage as defined in Fig. S1. Images were scored without the evaluators having knowledge of the exposure pH. Scores for each crab were averaged among the four evaluators.

### Elemental content

Calcium, magnesium and strontium content were quantified using inductively coupled plasma optical emission spectrometry (ICP-

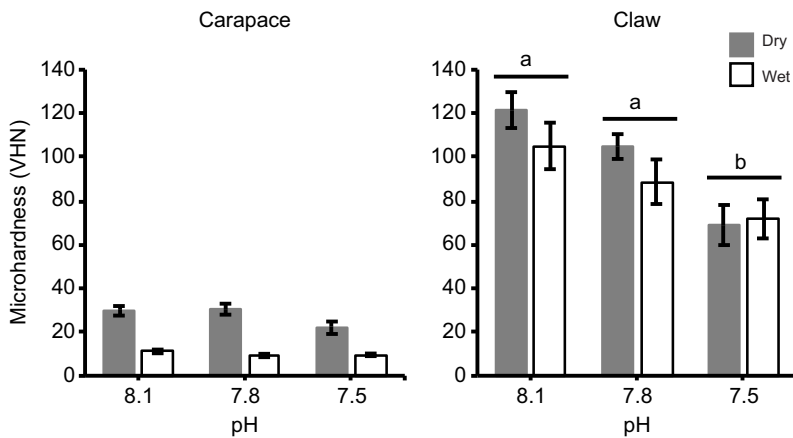
OES) at the US Geological Survey's Coastal and Marine Science Center, St Petersburg, FL, USA. Assessments were conducted using a portion of the carapace and the distal portion of the pollex, cut and lyophilized as described above (see 'Sample preparation'). Methods followed Gravinese et al. (2016) and Steffel et al. (2019). Briefly, whole samples were subjected to two rounds of oxidation, which consisted of sonication in a 1:1 mixture of 30%  $\text{H}_2\text{O}_2$  and 0.1 mol  $\text{l}^{-1}$  NaOH, followed by sonication in Milli-Q water. After oxidation, samples were dried overnight at 90°C and then ground to a fine powder using a mortar and pestle. Powdered samples were then subjected to an additional round of oxidation treatment (as described above), followed by drying at 90°C for at least 3 h.  $\text{Ca}^{2+}$ ,  $\text{Mg}^{2+}$  and  $\text{Sr}^{2+}$  content were measured on powdered and oxidized samples using a PerkinElmer 7300 dual-view ICP-OES. Individual samples were weighed and acidified in 2%  $\text{HNO}_3$  to obtain a target concentration of 20 ppm  $\text{Ca}^{2+}$ , which is compatible with the linear calibration of the instrument. Mass-percentages for each element were calculated by multiplying concentration by the volume of  $\text{HNO}_3$  added prior to ICP-OES analysis, and then dividing by the dry mass of the sample using the conversion 1 ppm=1 mg  $\text{l}^{-1}$  (Long et al., 2013b).

### FTIR spectroscopy: $\text{CaCO}_3$ polymorphs

Fourier transform infrared (FTIR) spectroscopy was used to assess the phase or polymorph of calcium carbonate present in cuticle samples. A portion of the carapace and the proximal portion of the pollex, cut and lyophilized as described above, were used for FTIR. Each sample was ground to a fine powder using a mortar and pestle. Spectra were collected using a PerkinElmer Spectrum Two spectrometer. Powdered samples were placed directly on the instrument's ATR (attenuated total reflectance) crystal and compressed with a uniform force by a built-in anvil. Spectra were taken at 4-wavenumber resolution, with 32 scans per sample. Spectra were normalized and baseline-corrected within the 700–900  $\text{cm}^{-1}$  region, which includes the  $\nu_2$  and  $\nu_4$  peaks characteristic of  $\text{CaCO}_3$  (Beniash et al., 1997; Khouzani et al., 2015).  $\nu_2$ -peak position was determined using the spectrometer's analysis software (PerkinElmer Spectrum 10).

### Statistical analysis

Statistical analyses were conducted using SPSS (v.24, IBM Analytics) or R 3.1.2 (<http://www.R-project.org/>). Prior to analyses, outliers were calculated for all metrics as values greater than three times the interquartile range below or above the first or third quartile, respectively, and were removed from the dataset. Outliers were rare throughout the dataset, with no outliers identified for most assessments and a maximum of two per pH treatment for  $\nu_2$ -peak position. Within the pH 7.8 treatment only, data for calcium content (carapace and claw) and  $\nu_2$ -peak position (carapace) from four crabs that had died just before the conclusion of the exposure period (see 'Animal collection and experimental exposure', above) were excluded from the dataset. For these specific metrics, a slight difference between crabs that were killed at the conclusion of the experiment and those that had died just before the conclusion of the exposure period was observed (decreased carapace and calcium content, increased carapace  $\nu_2$ -peak position in the crabs that died early; Mann–Whitney  $U$ -test:  $P<0.05$ ). For all other metrics, there was no difference between crabs that were killed at the conclusion of the experiment and those that had died just before the conclusion of the exposure period. Microhardness data were analyzed using a mixed-model analysis of variance (ANOVA) at the 5% significance level. This allowed assessment of the interaction of pH (between-subject variable) and hydration (within-subject variable) on microhardness, as well as the main effects of pH and hydration



**Fig. 1. Vickers microhardness of the carapace and claw of Tanner crab, *Chionoecetes bairdi*.** Adult crabs were exposed to one of three pH conditions (8.1, 7.8 and 7.5) for 2 years. Microhardness measurements are means $\pm$ s.e.m. Among pH levels, groups marked with different letters are significantly different as shown by Tukey HSD *post hoc* analysis.  $N=6-10$ ; specific values of  $N$  can be found in Table S1.

individually. Mixed-model ANOVA assumptions of sphericity and equal variance were assessed using the Mauchly and Levene's tests, respectively. Other quantitative metrics – total cuticle thickness,  $\text{Ca}^{2+}$ ,  $\text{Mg}^{2+}$  and  $\text{Sr}^{2+}$  content, pollex damage and  $v_2$ -peak position – were assessed using one-way ANOVA followed by Tukey HSD *post hoc* testing. The carapace and claw data were assessed separately. Datasets were analyzed for normality and homogeneous variance with Kolmogorov–Smirnov and Levene's tests, respectively, and data were log-transformed if necessary to meet these assumptions. If assumptions of normality or equal variance could not be met after log-transforming the data, a non-parametric Kruskal–Wallis test was used in place of the parametric ANOVA. For structural integrity of the carapace, the probability of carapace erosion was fitted to two models in R 3.1.2, using maximum likelihood and assuming a binomial distribution of errors, one in which the probability of erosion did not differ among the treatments and one in which it differed among all treatments. Akaike's information criterion corrected for sample size,  $\text{AIC}_c$ , was calculated for each model, and the most parsimonious model was selected. Models whose  $\text{AIC}_c$  differed by  $<2$  were considered to explain the data equally well (Burnham and Anderson, 2002).

## RESULTS

### Micromechanical properties

Vickers microhardness was measured within the endocuticle when samples were dry and again when rehydrated. In the carapace, hydration led to a significant reduction in endocuticle hardness (Fig. 1, Table 2; Table S1), with an average reduction of 60%. Hardness of the carapace was not affected by treatment pH, and the interaction of hydration and pH was not significant. In the claw, the opposite response was observed: hydration did not affect endocuticle hardness, but pH did (Fig. 1, Table 2; Table S1). Hardness of the claw for crabs held at pH 7.5 was, on average, 38% lower than for those held at pH 8.1 (ambient) and 27% lower than for those held at pH 7.8. The interaction of hydration and pH was not significant in the claw. Both when dry and when wet, hardness of the claw was substantially higher than that of the carapace, with claw samples about four times harder than the carapace when dry and nearly 10 times harder when wet.

### Cuticle thickness and structural assessment

Total cuticle thickness was affected by exposure pH in both the carapace and claw (Fig. 2; Table S2). The carapace of crabs exposed to pH 7.5 was on average 15% thinner than that of crabs at pH 8.1 (ambient). Erosion was visible on the interior of the carapace of most crabs (57%) held at pH 7.5 but never in crabs held at ambient

pH (Fig. 3; Table S2). The model in which the probability of carapace erosion differed among all pH treatments was a better fit and more parsimonious than the model where erosion did not differ among treatments, and was therefore selected ( $\Delta\text{AIC}_c=4.1$ ). Crabs at pH 7.8 had intermediate measures of total cuticle thickness and internal erosion; total cuticle thickness did not differ significantly from that of the pH 8.1 (ambient) or pH 7.5 crabs, and internal carapace erosion was identified in 22% of crabs. The exterior of all carapace samples assessed showed signs of wear (discolored, broken, uneven or rough regions), and the frequency of wear did not differ among pH treatments.

For the claw, the cuticle was 31% thinner in crabs exposed to pH 7.5 compared with those at the ambient pH of 8.1 (Fig. 2; Table S2); cuticle thickness in crabs exposed to pH 7.8 was intermediate to, and did not differ from, that at either ambient pH or pH 7.5. Although patterns of wear in the form of broken, worn or pitted surfaces were visible on the exterior of all pollex samples, the extent of pollex damage was far greater in crabs exposed to reduced pH (Fig. 3; Table S2). Particularly in pH 7.5 crabs, the contact surface of the pollex, which displays the tooth-like denticles, was completely worn down, with the denticles barely visible (Fig. 3). In contrast, the pollex of crabs at ambient pH showed a relatively smooth appearance, with prominent denticles. Semi-quantitative assessment of pollex damage confirmed these observations, with the extent of damage greater in the pH 7.5 and 7.8 crabs compared with those at ambient pH (Table S2).

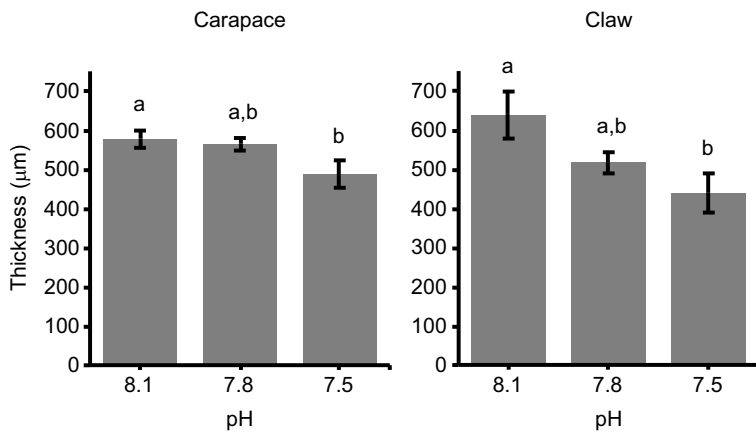
### Elemental content

Calcium content of the cuticle, measured per unit dry mass, was lower in animals held at reduced pH (7.5) in the carapace but not in the claw (Table 3). In the carapace, calcium content was reduced on average by 11% in animals exposed to pH 7.5 compared with those

**Table 2. Mixed-model ANOVA table, assessing the effect of hydration and pH on the cuticle microhardness of Tanner crab, *Chionoecetes bairdi***

	d.f.	<i>F</i>	<i>P</i>
Carapace			
Hydration	1, 23	150.2	<b>0.000</b>
pH	2, 23	2.123	0.143
Hydration <p>×</p> pH	2, 23	2.616	0.095
Claw			
Hydration	1, 21	2.797	0.109
pH	2, 21	7.654	<b>0.003</b>
Hydration <p>×</p> pH	2, 21	1.101	0.351

Significant *P*-values are shown in bold.



**Fig. 2. Cuticle thickness of the carapace and claw of Tanner crab, *C. bairdi*.** Adult crabs were exposed to one of three pH conditions (8.1, 7.8 and 7.5) for 2 years. Cuticle thickness measurements are means±s.e.m. Groups marked with different letters are significantly different as shown by Tukey HSD *post hoc* analysis.  $N=7-10$ ; specific values of  $N$  can be found in Table S2.

held at ambient pH. Calcium content for crabs at pH 7.8 did not differ from that at ambient pH. Treatment pH also exerted a significant effect on magnesium content of the carapace, but, in contrast to calcium, magnesium content increased by 17% in crabs held at pH 7.5 compared with ambient pH and by 15% compared with that of crabs held at pH 7.8. Magnesium content did not vary significantly among pH levels in the claw. Strontium content was not affected by treatment pH in the carapace or claw (Table 3).

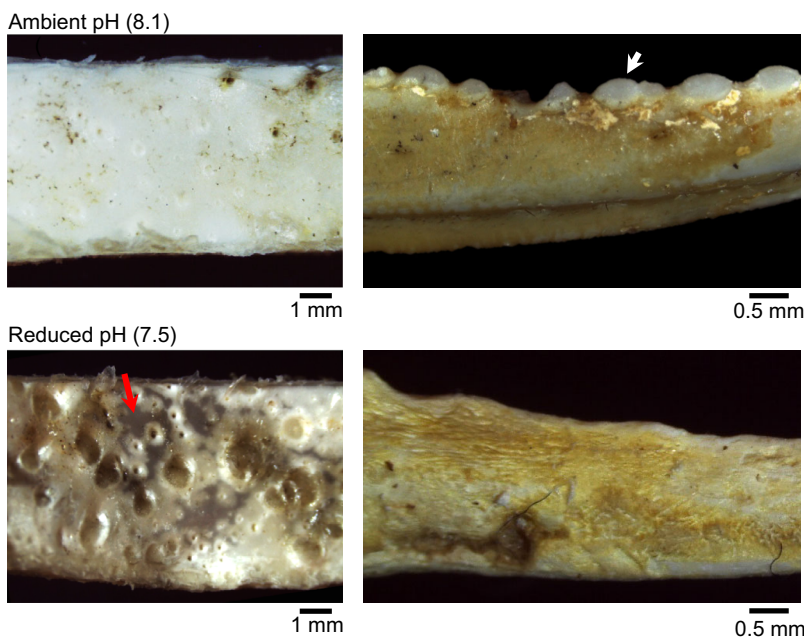
#### FTIR spectroscopy: $\text{CaCO}_3$ polymorphs

FTIR spectroscopy, which is sensitive to the phase of calcium carbonate present in a material, was conducted on powdered carapaces and the pollex region of the claws. Calcite is characterized by a sharp  $\nu_2$  peak at  $874\text{ cm}^{-1}$  and a well-defined  $\nu_4$  peak at  $713\text{ cm}^{-1}$ , whereas ACC shows a broad  $\nu_2$  peak at  $866\text{ cm}^{-1}$  and no  $\nu_4$  peak (Beniash et al., 1997; Khouzani et al., 2015). Fig. 4 shows representative FTIR spectra for cuticle samples from crabs held at ambient (8.1) or reduced (7.5) pH, along with reference spectra for synthetic ACC (Kimmel Center for Archaeological Science Infrared Standards Library, Weizmann Institute of Science, Rehovot, Israel) and biogenic calcite (from barnacle shell; Nardone et al., 2018). In all cases, spectra were consistent with a mix of calcite and ACC. In

the carapace, there was a statistically significant shift in the position of the  $\nu_2$  peak in crabs held at pH 7.5 compared with those at ambient pH (Fig. 4; Table S2): the  $\nu_2$  peak was positioned at  $866.3\pm 1.3\text{ cm}^{-1}$  (mean±s.e.m.) for animals held at ambient pH, but at  $872.4\pm 0.1\text{ cm}^{-1}$  for those held at pH 7.5. This shift, combined with the reduced width of the  $\nu_2$  peak at pH 7.5 and the initial formation of a  $\nu_4$  peak, suggests a transition from ACC to calcite in crabs held at pH 7.5. FTIR spectra of the claw showed a sharp  $\nu_2$  peak and a well-defined  $\nu_4$  peak, suggesting the predominance of calcite. The position of the  $\nu_2$  peak in the claw showed a very slight but statistically significant shift in crabs held at pH 7.5, compared with those at ambient pH (Fig. 4; Table S2): the  $\nu_2$  peak was positioned at  $871.8\pm 0.1$  for animals held at ambient pH, but at  $872.2\pm 0.1$  for those at pH 7.5. For both the carapace and the claw, the position of the  $\nu_2$  peak of crabs held at intermediate pH (7.8) did not differ significantly from that of crabs held at ambient pH.

#### DISCUSSION

The decapod cuticle is a multifunctional, composite structure that is central to the animal's success in feeding, defense and resistance to desiccation (Meyers et al., 2013; Meyers and Chen, 2014). For animals that are past their terminal molt, functionality of the cuticle



**Fig. 3. Representative light microscopy images of cuticles of Tanner crab, *C. bairdi*, exposed to either ambient or reduced pH for 2 years.** Left: carapace interior. Visible erosion (e.g. the darkened region marked by the red arrow, lower left) was apparent on most animals held at reduced pH (7.5), but was not observed in animals held at ambient pH (8.1). Right: pollex exterior. Denticles (white arrow, upper right) were prominent in animals held at ambient pH but were highly worn in animals held at pH 7.5.

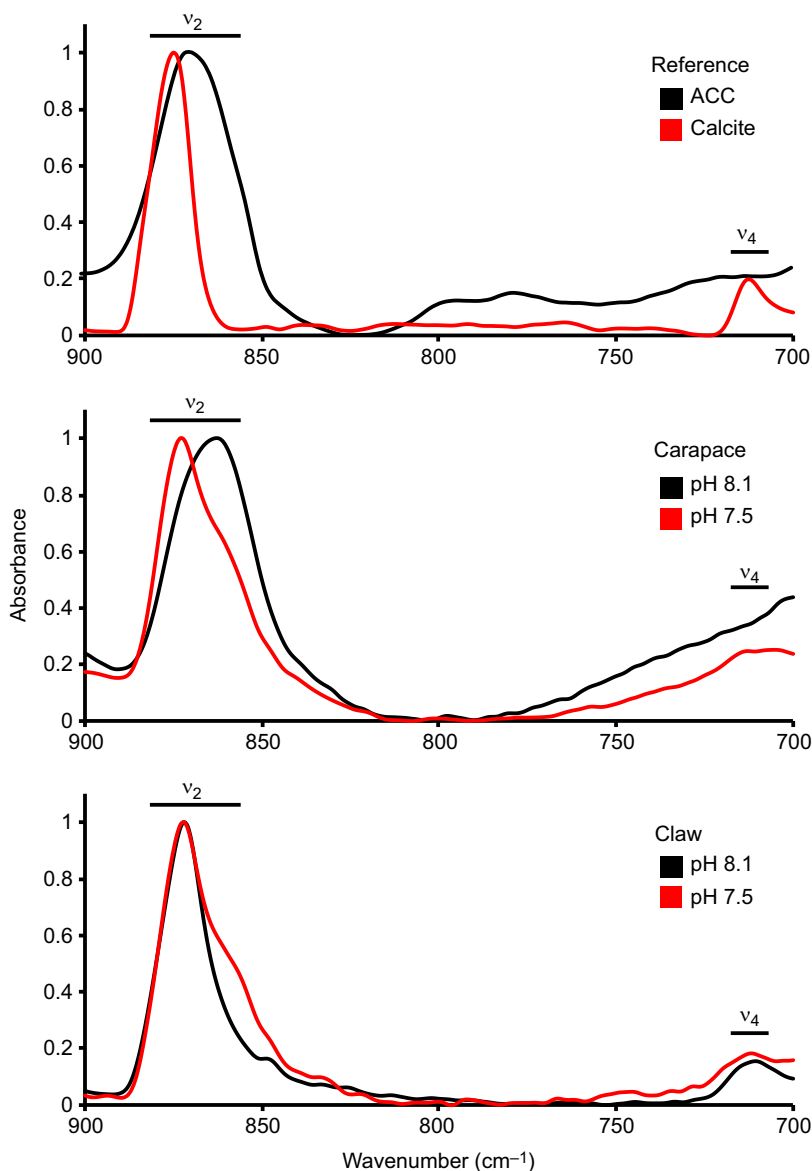
**Table 3. Elemental content for the cuticle of Tanner crab, *C. bairdi***

Parameter	8.1		7.8		7.5		ANOVA ( <i>F</i> ) or Kruskal–Wallis ( <i>H</i> )
	Mean±s.e.m.	<i>N</i>	Mean±s.e.m.	<i>N</i>	Mean±s.e.m.	<i>N</i>	
<b>Carapace</b>							
Ca (% dry mass)	20.7±0.5 <sup>A</sup>	10	21.2±0.5 <sup>A</sup>	10	18.5±0.4 <sup>B</sup>	7	$F_{2,22}=7.2$ , <b><i>P</i>=0.004</b>
Mg (% dry mass)	2.40±0.07 <sup>A</sup>	10	2.44±0.06 <sup>A</sup>	10	2.81±0.13 <sup>B</sup>	7	$F_{2,26}=6.6$ , <b><i>P</i>=0.005</b>
Sr (% dry mass)	0.42±0.01	10	0.42±0.01	10	0.49±0.03	7	$H_2=4.3$ , <i>P</i> =0.115
<b>Claw</b>							
Ca (% dry mass)	28.5±0.7	10	28.4±0.4	5	27.0±0.9	7	$F_{2,1}=1.3$ , <i>P</i> =0.289
Mg (% dry mass)	0.89±0.06	10	1.07±0.08	9	0.94±0.05	7	$H_2=5.4$ , <i>P</i> =0.066
Sr (% dry mass)	0.37±0.01	10	0.37±0.01	9	0.35±0.01	7	$F_{2,5}=2.9$ , <i>P</i> =0.074

Data, sample size (*N*) and ANOVA results are shown. Groups marked with different letters are significantly different as shown by Tukey HSD *post hoc* analysis. Significant *P*-values are shown in bold. Subscripts in the right-most column refer to degrees of freedom.

depends on the maintenance of cuticle structural and mechanical integrity on scales ranging from the microscopic to the macroscopic. In decapods, the cuticle is very much a ‘living tissue’ (Roer and Dillaman, 1984). It sits atop a multi-layered hypodermis, and cytoplasmic extensions of the outer epithelial layer of the hypodermis extend into the cuticle via pore canals (Travis, 1963;

Roer and Dillaman, 1984; Cameron, 1989; Kunkel, 2013). Such intimate contact with the hypodermis may permit modification of the mineral and protein portions of the cuticle even during intermolt or after the terminal molt (Halcrow and Steel, 1992; Kunkel, 2013). Here, we aimed to assess the properties of the cuticle in the southern Tanner crab, *C. bairdi*, a long-lived inhabitant of the North Pacific



**Fig. 4. Representative FTIR spectra of powdered cuticle samples from the carapace and claw of Tanner crab, *C. bairdi*.** Reference spectra for synthetic amorphous calcium carbonate (ACC) and biogenic calcite are shown for comparison.

shelf, in the face of reduced seawater pH (OA) and a concomitant decrease in calcite saturation state. *Chionoecetes bairdi* inhabits a geographic region where the pH and calcium carbonate saturation state are already seasonally low (Long et al., 2016; Punt et al., 2016) and where future changes in ocean chemistry are likely to occur more rapidly than at lower latitudes (Fabry et al., 2009). A 2 year exposure of *C. bairdi* to reduced pH (7.5) led to a reduction in microhardness of the claw, alterations in the mineral content of the carapace, thinning of both the claw and carapace, internal dissolution of the carapace, a loss of the tooth-like denticles on claws, and a shift in the phase or polymorph of calcium carbonate present in the carapace. These changes occurred despite the fact that decapod crustaceans are often reported to be more resilient to OA than other marine calcifiers (Ries et al., 2009; Kroeker et al., 2010, 2013; Whiteley, 2011; Byrne and Fitzner, 2019).

Microhardness is a measure of a material's resistance to mechanical (plastic) deformation. Assessment of microhardness within the *C. bairdi* endocuticle revealed two general patterns. First, microhardness of the claw was consistently higher than that of the carapace, regardless of seawater pH, a pattern previously observed in a number of other decapod crab species (Lian and Wang, 2011; Steffel et al., 2019). Second, long-term exposure of crabs to reduced-pH seawater resulted in a body region-specific reduction in microhardness. Although a significant reduction in endocuticle microhardness was observed in the claw, the effect of reduced pH on microhardness of the carapace was not significant. The body region-specific response to seawater pH observed here is consistent with previous assessments of juvenile blue and red king crabs, *P. platypus* and *P. camtschaticus*, in which a reduction in endocuticle microhardness was also observed for the claw, but not the carapace (Coffey et al., 2017). In the Coffey et al. (2017) study, crabs had molted several times during experimental exposure. Together with our results, these findings suggest that exposure to reduced-pH seawater induces a similar pattern of changes in cuticle mechanical properties, whether the cuticle is newly deposited during ecdysis or is pre-existing when exposure begins.

The harder endocuticle of the claw compared with the carapace may result from an elevated calcium content (Sachs et al., 2006; Waugh et al., 2006; Boßelmann et al., 2007; Page et al., 2017): on average, claw samples contained ~40% more calcium than those from the carapace. The phase of calcium carbonate present within these cuticle regions may also contribute. The proportion of calcite versus ACC is greater in the claw than the carapace, and calcite tends to be harder than ACC (Bentov et al., 2016a). Neither calcium content nor the phase of calcium carbonate, though, can adequately explain the reduction in hardness observed in the claws of crabs exposed to reduced pH. Calcium content of the claw did not differ significantly among pH treatments, and claws from all pH levels were composed primarily of calcite. This observation is again consistent with the work of Coffey et al. (2017) on *P. platypus* and *P. camtschaticus*. Despite a reduction in endocuticle hardness in both species at reduced pH, calcium content was not affected by exposure pH in *P. platypus*, and in *P. camtschaticus*, calcium content of the claw was actually greater at reduced pH. A number of other properties can influence cuticle hardness in decapods, including: the packing density of twisted plywood structures; phosphate content (including the presence of calcium phosphate); cross-linking and other modifications of the protein portion; density of pore canals; and the orientation, density and structural integrity of mineralized protein–chitin fibers (Melnick et al., 1996; Chen et al., 2008; Fabritius et al., 2011; Lian and Wang, 2011; Fabritius et al., 2012; Bentov et al., 2016a,b; Rosen et al., 2020). It remains to be

determined which, if any, of these properties are driving the observed reduction in claw microhardness at reduced seawater pH seen here and by Coffey et al. (2017).

The decapod cuticle is hydrated in its natural state (Hepburn et al., 1975; Cameron and Wood, 1985; Boßelmann et al., 2007; Neues et al., 2007). Cameron and Wood (1985) estimated that 26.5% of the *Callinectes sapidus* carapace was water, based on wet and dry mass. Using thermogravimetry, Boßelmann et al. (2007) identified 11.8% of the carapace of *Cancer pagurus* as water, whereas the dactylus of the claw contained only 1% water. The difference in hydration of the carapace versus the dactylus, along with the elevated calcium content in the claw, may explain the difference in sensitivity to hydration observed here (Vincent, 2002; Fabritius et al., 2011). The microhardness of carapace samples when tested dry was about three times higher than when the same samples were tested wet, whereas microhardness of the dactylus was not affected by hydration. Importantly, the response of the cuticle in terms of microhardness to reduced pH was not affected by hydration state (i.e. the interaction of pH and hydration within the repeated measures ANOVA was not statistically significant for the carapace or claw; see Table 2). Hence, the structural or chemical properties of the cuticle that drive body region-specific changes in microhardness with pH do not appear to be affected by hydration.

Long-term exposure of *C. bairdi* to reduced pH resulted in a suite of potentially interrelated alterations in the cuticle of the carapace. At pH 7.5, calcium content was reduced, while magnesium content was elevated, implying a higher  $Mg^{2+}:Ca^{2+}$  ratio. Thickness of the carapace was reduced at pH 7.5, and the majority of pH 7.5 carapace samples showed internal erosion. Solubility of calcite tends to increase with elevated  $Mg^{2+}:Ca^{2+}$  ratio (Morse et al., 2006; Andersson et al., 2008), which may have left the cuticle more susceptible to internal dissolution (Bednaršek et al., 2020). Observed internal dissolution could in turn have driven the reduction in thickness of the carapace cuticle. It is possible that the reduction in carapace calcium results from mobilization of  $Ca^{2+}$  and  $HCO_3^-$  from the cuticle, as a mechanism to buffer hemolymph pH (DeFur et al., 1980; Henry et al., 1981; Cameron, 1985; Spicer et al., 2007; Page et al., 2017; Bednaršek et al., 2020). Indeed, when Meseck et al. (2016) assessed extracellular hemolymph pH ( $pH_e$ ) in the same *C. bairdi* assessed here,  $pH_e$  was maintained at ~8.09 even in crabs held at the lowest seawater pH. It is important to note, however, that the contribution of carapace ions to hemolymph buffering in other crab species appears to be minor compared with the uptake of ions from external seawater (Cameron, 1985; Spicer et al., 2007).

In the claw, exposure to reduced pH resulted in thinning of the cuticle without a corresponding change in elemental content. Internal dissolution could not be readily assessed in claw samples, but extensive erosion of the exterior of the pollex was observed in crabs held at pH 7.5, with nearly complete loss of the tooth-like denticles in these crabs. This occurred despite the fact that the captive crabs were fed soft foods and hence did not experience the high levels of abrasion they might have experienced in the field from consuming heavily calcified foods and interacting with other crabs. The waxy epicuticle in decapods protects the underlying mineral from changes in seawater chemistry (Ries et al., 2009). As shown by Kunkel et al. (2012), removal of the epicuticle leads to an increase in ion flux from the mineralized cuticle. Waugh et al. (2006) and Rosen et al. (2020) documented in multiple crab species that normal wear on the denticles results in loss of the epicuticle, as well as the exocuticle, from the denticle surface, which could leave the mineralized endocuticle susceptible to dissolution. The presence of epicuticle on the denticle surface was not assessed before



exposure in our study, but given that the crabs used here were already past their terminal molt when collected from the field, it is likely that the epicuticle covering the denticles was absent at the start of the experimental exposure. Damage to the claw, and particularly to the tooth-like denticles, can lead to a reduction in the crabs' prey-capture efficiency (Juanes and Hartwick, 1990).

The interior dissolution of the *C. bairdi* carapace and exterior dissolution and wear of the claw show promise as ecosystem indicators (*sensu* Kershner et al., 2011) for OA effects in Alaska. Scoring of both could be done on a semi-quantitative scale and could easily be incorporated into existing annual surveys that target *C. bairdi*, an economically important species. These measures are correlated with other significant negative outcomes such as embryonic mortality and decreased female survival (Swiney et al., 2016) that are harder to measure or estimate on an annual basis. As a next step in developing these metrics as ecosystem indicators, future work should examine variation in cuticle dissolution and wear in natural populations to determine whether they are correlated with natural environmental gradients.

Multiple mineral forms and phases are found within the decapod cuticle, with the mineral component being predominately nanocrystalline calcite and ACC (Roer and Dillaman, 1984; Dillaman et al., 2005; Fabritius et al., 2012). After molting, calcium carbonate is initially deposited as ACC, and some (but not all) of the ACC is transformed to calcite in the days following initial mineral deposition. Stabilization of ACC (i.e. the inhibition of calcite nucleation) may involve protein components of the cuticle, specific ions (magnesium, phosphorus and silicon), and glycolytic intermediates (PEP and 3PG) (Coblentz et al., 1998; Addadi et al., 2003; Weiner et al., 2003; Sato et al., 2011; Roer and Dillaman, 2018). Given that ACC is highly unstable (Weiner and Addadi, 1997; Addadi et al., 2003; Weiner et al., 2003), a slight change in conditions within the cuticle could result in calcite nucleation.

FTIR spectroscopy of the *C. bairdi* carapace suggests a shift in the phase of calcium carbonate from ACC to calcite. To the best of our knowledge, this is the first report of a shift in mineral phase in a crustacean (Ries, 2011). Benefits of the use of ACC have been discussed in depth (Addadi et al., 2003; Weiner et al., 2003; Neues et al., 2007; Bentov et al., 2016a); ACC is isotropic and fracture resistant, and it can serve as a readily soluble  $\text{Ca}^{2+}$  pool. The functional implications of this shift in mineral phase remain to be determined. At least at the micrometer scale, the shift toward calcite did not appear to affect hardness (i.e. microhardness was not affected by exposure pH), but isotropy and fracture resistance were not directly assessed. Continued quantification of cuticle mechanical properties at a range of spatial scales (from the nanometer scale to the scale of the entire carapace) and temporal scales (from short to extended times since molting and durations of pH exposure), as well as assessment of the role of carapace ions in hemolymph buffering, may help to resolve the functional consequences of the observed shift in mineralogy.

## Conclusions

Variations in mechanical, elemental, structural and mineralogical properties of the decapod crab exoskeleton lead to differences in functionality. This is clearly evident when comparing the carapace with the claw, which is the primary feeding and active defensive structure. Compared with the carapace, the claw is substantially harder, and it contains more calcium but less magnesium. A greater proportion of calcium carbonate in the claw is present as crystalline calcite as opposed to ACC, and the cuticle of the claw is less

sensitive to hydration. These differences set the stage for the body region-specific response to OA observed in *C. bairdi*. Exposure to reduced pH led to a reduction in microhardness of the claw, but not the carapace. There was no change in elemental content at reduced pH in the claw, but in the carapace, calcium content was reduced and magnesium content increased. Calcium carbonate in the claw was already predominantly in the form of calcite, whereas in the carapace, calcium carbonate was primarily ACC at ambient pH but shifted to calcite in crabs exposed to pH 7.5.

Assessment of the structural integrity of the cuticle suggests that long-lived crabs that display determinate growth may be particularly susceptible to OA. *Chionoecetes bairdi* held at a reduced pH of 7.5 displayed internal dissolution of the carapace, as well as extensive erosion of the claw, with nearly complete loss of tooth-like denticles. At the functional level, the loss of denticles could inhibit feeding ability and efficiency, as has been shown in other crabs (Juanes and Hartwick, 1990). Although direct assessments of the effect of degraded claws on feeding in *C. bairdi* are still needed, impaired feeding could lead to energy limitation with potential consequences for reproductive output. The thinner, eroded cuticle observed in both body regions may also break more readily, diminishing its protective functionality. Although cuticle repair is possible after the terminal molt (Halcrow and Steel, 1992), the cuticle is never fully replaced as occurs during molting. Even under current oceanic conditions, cuticle damage tends to accumulate over time, leading to a decrease in shell condition with age (Ernst et al., 2005; Fonseca et al., 2008; Vogt, 2012). Furthermore, in *C. bairdi*, the hemocytes responsible for cuticle repair (granular and semi-granular cells) show reduced intracellular pH ( $\text{pH}_i$ ), which may limit their functionality in the cuticle-repair process (Meseck et al., 2016). Altogether, the results presented here demonstrate that OA can alter exoskeleton properties in *C. bairdi*, which may affect the success of this ecologically and economically important species in coming years.

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## Competing interests

The authors declare no competing or financial interests.

## Author contributions

Conceptualization: G.H.D., W.C.L., R.J.F.; Methodology: G.H.D., W.C.L., B.V.S., K.E.S., R.B.A.; Validation: G.H.D.; Formal analysis: G.H.D., W.C.L.; Investigation: G.H.D., S.B., T.S., C.M., S.P., W.C.L., K.M.S., B.V.S.; Resources: G.H.D., W.C.L., R.J.F., R.B.A.; Data curation: G.H.D.; Writing - original draft: G.H.D., W.C.L.; Writing - review & editing: G.H.D., S.B., T.S., C.M., S.P., W.C.L., K.M.S., R.J.F., B.V.S., K.E.S., R.B.A.; Visualization: G.H.D.; Supervision: G.H.D., W.C.L., R.J.F., R.B.A.; Project administration: G.H.D., W.C.L., R.J.F., R.B.A.; Funding acquisition: G.H.D., W.C.L., R.J.F., R.B.A.

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## Data availability

Data are available from the Dryad digital repository (Dickinson et al., 2021): dryad.5mkkwh74w

## Supplementary information

Supplementary information available online at  
<https://jeb.biologists.org/lookup/doi/10.1242/jeb.232819.supplemental>

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