

Journal of Science and Technology

Growth of Freshwater Microalga, *Botryococcus* sp. in Heavy Metal Contaminated Industrial Wastewater

Joan Iye Onalo^{1,*}, Hazel Monica Matias-Peralta¹, Norshuhaila Mohamed Sunar²

¹Department of Technology and Heritage, Faculty of Science Technology and Human Development Universiti Tun Hussein Onn Malaysia, 86400 Parit Raja, Batu Pahat, Johor, Malaysia.

²Department of Chemical Engineering Technology, Faculty of Chemical Technology Universiti Tun Hussein Onn Malaysia, 86400 Parit Raja, Batu Pahat, Johor, Malaysia.

**Corresponding email: joaniyeh@gmail.com*

Abstract

The aim of this study was to determine the growth and the bioremoval capacity of the green microalga, *Botryococcus* sp. grown in industrial wastewater contaminated with heavy metals. The freshwater green microalga, *Botryococcus* sp. was cultured in different concentrations of wastewater (25%, 50% and 100%) with an initial cell concentration of 1000 cells/ml for a period of 12 days. Bold basal medium and sterile distilled water were used as positive and negative control, respectively. The *Botryococcus* sp. grown in Bold's basal medium showed the highest ($P < 0.05$) average growth rate (7.8×10^6 cells/ml) after a period of 12 days, whereas, the lowest ($P < 0.05$) growth was observed in 50% concentration of wastewater (4.8×10^4 cells/ml). Similar results were obtained for the specific growth rate (μ /day) with an average of 1.93μ /day and 1.22μ /day for the positive control and the 50% concentration respectively. Highest reduction of heavy metals was achieved for chromium which is equivalent to 94%, followed by copper (45%), arsenic (9%) and cadmium (2%). The results of this study suggest the potential of *Botryococcus* sp. as bioremediator of wastewater contaminated with heavy metals.

Keywords: industrial wastewater, heavy metals, microalgae, *botryococcus* sp

1. INTRODUCTION

The introduction of untreated industrial wastewater containing toxic elements (such as the heavy metals) into the water bodies has become a great deal of concern to the environment [1]. Heavy metals contained in industrial effluents are dangerous particularly in human health perspective due to their toxicity and bio-accumulating effects [2,3,4]. Some of the conventional methods which are still used in the treatment of wastewaters such as ion exchange and chemical precipitation [5], do not produce significant or desired results when the concentration of heavy metal present is relatively high [6]. In addition, they are also expensive to manage and could sometimes lead to an unfavourable condition such as the formation of toxic chemical sludge as a result of the many chemicals involved in the process. The application of bioremediation in heavy metal removal from wastewater has offered some advantages over the use of conventional wastewater treatment technologies. These include cost effectiveness, high efficiency, renewability of materials and no generation of additional pollutants [1,7,8]. Microalgae have shown high heavy metal binding affinity primarily due to short-chain amino acids they possess which are linked by peptide bonds [9,10]. It has also been found that they possess a large surface area-to-volume ratio that tends to be accessible for contact with the environment within as well as their accumulation ability which makes them potential remediating agents [11]. Considering the toxicity of heavy metals, there is therefore a need to treat industrial effluents before discharging into the waterways with a cost effective and environmental friendly approach like the use of biosorbents (such as the microalgae). The aim of this study was to determine the growth efficiency and the bioremoval capacity of the green microalga, *Botryococcus* sp. in industrial wastewater contaminated with heavy metals.

2 MATERIALS AND METHODS

2.1 Industrial Wastewater

Industrial wastewater was obtained from an industry in Batu Pahat, (1° 51' 0" North, 102° 56' 0" East) Johor, Malaysia in acid washed sample bottles and was transferred to the laboratory. Immediately upon arrival in the laboratory, the water quality parameters of raw wastewater were measured (BOD, COD, Temp, pH, DO, Conductivity, TN, TP, TOC, TDS, TSS and heavy metals) following the standard method for water and wastewater [12]. The wastewater was kept in a cold room (4°C) before the commencement of the experiment. Prior to the experiment, the wastewater was filtered first with 0.45µm membrane filter (Whatman) and then diluted with distilled water at the desired concentrations (25%, 50% and 100%).

2.2 Preparation of *Botryococcus* sp. inoculum

The *Botryococcus* sp. isolates was obtained from microbiology laboratory of the Faculty of Science, Technology and Human Development, University Tun Hussein Onn Malaysia. Prior to the experiment, the isolate was cultured for 10 days in 1 litre Bold's basal media [13,14]. Algal cells were harvested with centrifugation set at 5000 rpm for 10 minutes followed by the determination of cell concentration. The cells of *Botryococcus* were counted using the Neubauer haemocytometer with the aid of a compound microscope.

2.3 Experimental Setup

A total of 15 Erlenmeyer flasks (500ml capacity) filled with 125ml of distilled water, growth media and wastewater diluted at different concentrations were used for this experiment. The growth media was used as the positive control and distilled water was used as the negative control. Three treatments corresponding to different concentrations of wastewater, 25% (25% wastewater and 75% distilled water), 50% (50% wastewater and 50% distilled water) and 100% (pure wastewater). All treatments were replicated three times. The treatment flasks were inoculated with the same initial microalgal cell concentrations (1000 cells/ml) at the start of the experiment and were covered with sterile cotton plugs and kept at room temperature all throughout the experimental period. Microalgal sample collection for growth monitoring started at day one and daily until day 12. Collections of samples were done under sterile condition. Daily growth rate of the microalga was monitored by counting the density under compound microscope using haemocytometer.

2.4 Data Analysis

2.4.1 Specific Growth Rate (μ /day)

The specific growth rate (μ /day) of the microalga was calculated using the following equation [42].

$$SGR(\mu / day) = \frac{\ln X_2 - \ln X_1}{t_2 - t_1}$$

Where;

X_2 = final algal concentration

X_1 = initial algal concentration

t_2 = final time (day 4)

t_1 = initial time (day 0)

2.4.2 Statistical Analysis

The statistical analysis using ANOVA with Turkey's-b test were performed on the bioassay data to determine significant differences between treatments at $p < 0.05$. All analysis was done using Statistical Package for Social Sciences version 20.0.

3. RESULTS AND DISCUSSION

3.1 Wastewater Parameters

There are established standards for effluents before discharging into the water bodies [15]. These standards are to be met in the level of effluents that are released and are ready to be discharged to the environment which is the responsibility of either the company or the treatment plants. In this study the concentrations of heavy metals arsenic (21.99 mg/L) and cadmium (0.081 mg/L) were found to be above permissible limits for water effluent standard (Table 1). Similarly, two of the most important water parameters which are the Chemical oxygen demand (COD) and Biochemical oxygen demand (BOD) at the concentration of 3847 mg/L and 167.16 mg/L, respectively were found to be above the permissible limits (Table 1). In addition, total suspended solids (240 mg/L) were also found to be above permissible limits (Table1).

The results obtained from the analysis of the raw wastewater in this study were a clear indication and in fact evident that wastewater coming from the industry has to be first treated before releasing into the environment. In particular, the heavy metals arsenic and cadmium which were found present in the wastewater could post harm to the receiving ecosystem due to their toxic effect. Heavy metal pollution in the aquatic biota has severe effects that threaten all the living organisms in the receiving water bodies [16,17,18]. Specifically, their persistence in the environment which is as a result of non-biodegradable nature and their bioaccumulation potential in the food chain make heavy metals very dangerous [19,20,21]. According to Rani et al (2010), toxic metallic elements such as cadmium, chromium and copper are exceptionally toxic to biological and ecological activities.) [22]. Heavy metals are considered the most important form of pollution particularly the arsenic and cadmium which are exceptionally toxic to humans [23]. It is said that heavy metal bioaccumulation is dependent on the type of metal, however, all heavy metals are known to disseminate throughout the trophic level of a specific ecosystem [24]. Heavy metals are regarded pervasive, highly soluble and are therefore promptly taken in by aquatic life [25]. According to Mansour and Sidky (2002), in the aquatic food chain, fishes are most times at the top and they can accumulate significant concentrations of these heavy metals in their tissues [26]. Considering the regularity of fish in human diet can bring about exposure which may have detrimental effect as heavy metals accumulate in the system [27,23].

Table 1: Characteristics of raw wastewater

Effluent Parameter	Value present in wastewater	Standard for effluent parameter (Environmental Quality Act, 1974)	
		A	B
Dissolved Oxygen (DO)	3.98 mg/L	-	-
Temperature	18oC	40oC	40oC
Conductivity	51.2 Ms/cm	-	-
pH	9.94	6.90-9.0	5.5-9.0
Total Phosphorus (PO43-)	10.20 mg/L	-	-
Total Nitrogen (TN)	655.3 ppm	-	-
Total Organic Carbon (TOC)	210.5 ppm	-	-
Chemical Oxygen Demand (COD)	3847 mg/L	50mg/L	100mg/L
Biochemical Oxygen Demand (BOD5)	167.16 mg/L	20mg/L	50mg/L
Turbidity	46 NTU	-	-
Total Dissolved Solids (TDS)	31, 550 mg/L	-	-
Total Suspended Solids (TSS)	240 mg/L	50mg/L	100mg/L
Total Solids (TSS)	31, 790 mg/L	-	-
Inorganic Carbon	1.503 ppm	-	-
Total Carbon	212.0 ppm	-	-
Chromium	0.018 mg/L	0.05mg/L (Chromium Hexavalent)	0.05mg/L (Chromium Hexavalent)
		0.02mg/L (Chromium Trivalent)	1.0 mg/L (Chromium Trivalent)
Cadmium	0.081 mg/L	0.01 mg/L	0.02 mg/L
Copper	0.099 mg/L	0.20 mg/L	1.0 mg/L
Arsenic	21.99 mg/l	0.05 mg/L	0.10 mg/L

3.2 Microalgal Ggrowth in Wastewater

The growth pattern of the microalga in wastewater and in the control was the same which peak on the 4th day (Figure 1). The growth rate of *Botryococcus* sp. was found highest in the positive control (BBM), with an average density of 7.8×10^6 cells/ml whereas, the lowest growth rate was found in treatment 2 (50% wastewater) with an average cell density of 4.8×10^4 cells/ml. Generally, since the nutrient required by the algal cell for normal growth are provided in the growth media, the microalga growth rate is expected to be highest in comparison to wastewater medium [28]. Although the growth rate of *Botryococcus* sp in wastewater treatment were lower, the fact that pattern of growth was similar with that of the control, suggest that this microalga was able to utilize both the nutrients and heavy metals present in the wastewater. This result proved the potential of this microalgal species in bioremediation.

The specific growth rate (SGR) of microalgae ranged from 1.20 μ /day - 1.43 μ /day being highest ($p < 0.05$) in BBM, while lowest ($p < 0.05$) in 50% concentration (Figure 2). The high SGR in BBM was expected as this media is supposed to provide the necessary nutrients for the normal growth of microalgal cell. On the other hand, the SGR of *Botryococcus* sp. was found to be most affected by the 50% wastewater concentrations as show in the lowest SGR value (Figure 2). In fact, the SGR was even higher ($p < 0.05$) in pure wastewater as compared to 50% wastewater. Perhaps in this case, higher nutrient present in the undiluted wastewater provided an advantage over those that are diluted. Higher nutrient concentrations were readily available for microalgal utilization, hence rapid growth which peaked on the 4th day of culture was observed in this study. According to De la Noue et al (1992), wastewater is regarded as a potential and sustainable media of growth for algal feedstock [29]. Many microalgae, specifically *Botryococcus braunii* has been effectively grown in municipal sewage wastewater, agricultural wastewater and industrial wastewater [30,31,32]. Moreover, microalgae have also shown high adaptability to a variety of environmental conditions such as industrial and domestic effluent dumping site [11] which probably explained their ability to grow even in the medium containing toxic heavy metals.

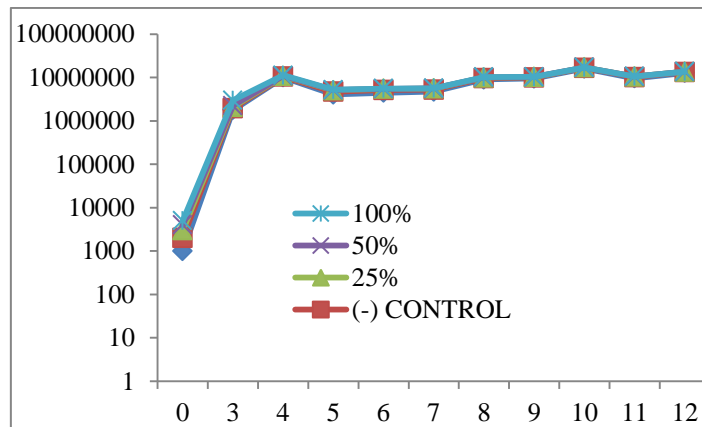


Figure 1: Daily growth rate (x 10³) cells/ml of *Botryococcus* sp. in different concentration of the wastewater (25%, 50% and 100%), distilled water and BBM.

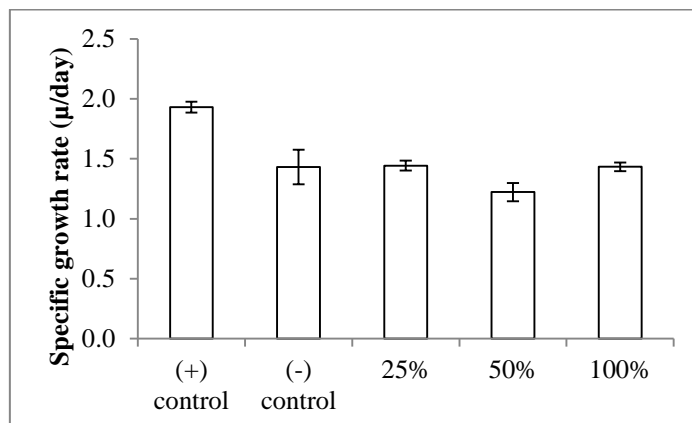


Figure 2: Specific growth rate (μ/day)

3.3 Heavy Metal Bioremoval

The heavy metal concentration was determined before and after the growth study of *Botryococcus* sp. in the industrial wastewater containing heavy metals to observe the reduction in the concentration of the elements. Four different heavy metals were studied in the course of these experiments which include cadmium, copper, arsenic and chromium (Figure 3). *Botryococcus* sp. efficiently reduced chromium from 0.018 to 0.001 which is equivalent to 94% and copper from 0.099 to 0.054 equal to 45% (Figure 3). Whereas, arsenic and cadmium were reduced to less than 10% (Figure 3). According to Jordanova et al (1999) and Bajguz (2000), different species of microalgae differ in their heavy metal accumulation potential in as much as many parameters have roles to play in the bioaccumulation process [33,34]. The use of active metabolic *Botryococcus braunii* in the removal of copper from solution has been reported by Areco et al (2013) [35]. Similarly, utilization of *Botryococcus* as an effective biological agent in the reduction of heavy metals in industrial wastewater has been reported earlier [32,36].

The application of biological system in the treatment of wastewater containing heavy metals is regarded as pertinent, cost effective and an efficient approach [37]. In particular, the use of microalgae in the removal of contaminants from industrial wastewater can go a long way in helping to lessen pollutant load in the environment [38]. The bioaccumulation potential of microalgae can be explained by their resistance mechanism against heavy metal toxicity [11]. According to Mehta and Gaur (2005), the most common and widely used resistance mechanism by microalgae to counter the toxic effects of heavy metals even at the dominance of ion exchange is the complexation and microprecipitation of the metallic ions [6]. Other known mechanisms include the regulation of metal concentration within the cells by the development of energy-driven flux pump, enzymatic and intracellular conversion of toxic metal to a non or less toxic form [6].

Although it has been recognised for many years that microalgae are indeed capable of absorbing metals from the environment, and in fact, microalgae are known to play a major role in controlling metal concentration in lakes and oceans [39,40]. However, it has to be understood as well that the tolerance of microalgae to heavy metal toxicity depend greatly on their defence responses [41] and that different species of microalgae have different bioaccumulation capacity against different heavy metal species.

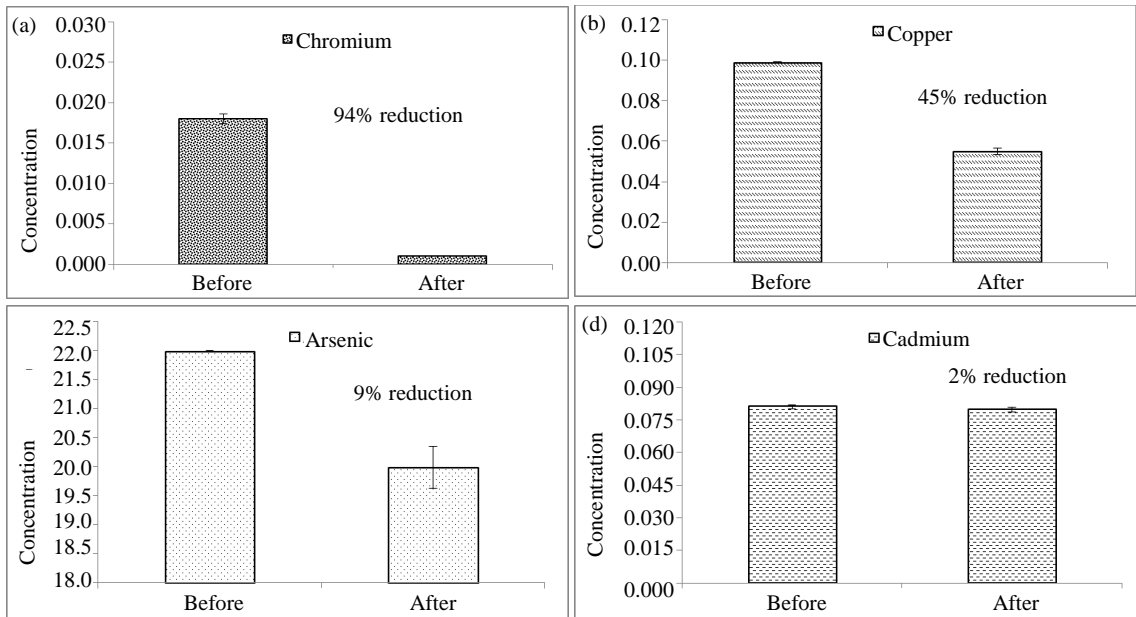


Figure 3: Heavy metal (a) Chromium, (b) Copper, (c) Arsenic and (d) Cadmium concentration before and after the treatment with *Botryococcus* sp

4. CONCLUSION

This study demonstrated the applicability of a freshwater microalga, *Botryococcus* sp. as a potential bioremediator of industrial wastewater contaminated with heavy metals. *Botryococcus* sp grown in different concentrations of wastewater showed similar growth pattern with those grown in synthetic growth media BBM. In addition, *Botryococcus* sp. significantly reduced the concentration of heavy metal chromium to up to 94% and 45% of the heavy metal copper.

REFERENCES

- [1] Hameed, M.S.A. (2006). "Continuous Removal and Recovery of Lead by Alginate Beads, Free and Alginate-Immobilized *Chlorella vulgaris*" African Journal of Biotechnology, Vol 5. No. 19 pp. 1819-1823.
- [2] Igwe, J.C. and Abia, A.A. (2003). "Maize Cob and Husk as Adsorbents for Removal of Cd, Pb and Zn ions from Wastewater". The Physical Sci., Vol. 2, pp. 83-94.
- [3] Igwe, J.C. and Abia, A.A. (2006). "A Bioseparation Process for Removing Heavy Metals from Waste Water Using Biosorbents" African Journal of Biotechnology, Vol. 5. No. 12 pp. 1167-1179.
- [4] Horsefall, M. Jnr. And Spiff, A. I. (2005). "Effects of Temperature on the Sorption of Co^{2+} and Cd^{2+} from Aqueous Solution by *Caladium Bicolour* (wild cocoyam) Biomass" Electronic Journal of Biotechnology [online]. Vol. 8. No. 2.
- [5] Eccles, H. (1999). "Treatment of Metal Contaminated waters: Why select a Biological Process?" Trends in Biotechnol., Vol. 17. No. 12 pp. 462-465.
- [6] Mehta, S.K. and Gaur, J.P. (2005). "Use of Algae for Removing Heavy Metal Ions from Wastewater: Progress and Prospects" Critical reviews in Biotechnol., Vol. 25. No. 3 pp. 113-152.
- [7] Spinti, M., Zhuang, H. and Trujillo, E.M. (1995). "Evaluation of Immobilized Biomass Beads for Removing Heavy Metals from Wastewaters" Water Environ. Res., Vol. 67. Pp. 943-435.
- [8] Srinath, T., Verma, T.P., Ramteke, W. and Garg, S.K. (2002). "Chromium (VI) Biosorption and Bioaccumulation by Chromate Resistant Bacteria" Chemosphere, Vol. 48. Pp. 427-435.
- [9] Jeon C, Park JY, Yoo YJ (2001). "Biosorption Model for Binary Adsorption Sites" J.Microbiol. Biotechnol, Vol 11. Pp. 781-787.
- [10] Perales-Vela, H.V, Peña-Castro, J.M & Cañizares-Villanueva . (2006). "Heavy Metal Detoxification in Eukaryotic Microalgae" Chemosphere, Vol. 64. No. 1 pp.1-10.
- [11] Cristina, M., Monteiro. And Castro, P.M.L. (2012) "Metal Uptake by Microalgae: Underlying Mechanisms and Practical Applications" American Institute of Chemical Engineers, Vol. 28. No. 2 pp. 299-311.

- [12] APHA, (2012). Standard Methods for the Examination of Water and Wastewater. 22nd ed. American Public Health Association, 800 I Street, NW, Washington DC.
- [13] Nichols, H.W. and Bold, H.C. (1965). "Trichosarcina polymorpha gen. Et sp. Nov" J. Phycol., Vol. 1. Pp. 34-38.
- [14] Nichols, H.W. (1973). Growth Media-freshwater. In Stein, J. (Ed.) Handwork of Phycological Methods. Culture Methods and Growth Measurements, Camb. Univ. Press. Pp. 7-24.
- [15] Environmental Quality Act (1974). Department of Environment, Malaysia.
- [16] Eisler, R. (1988). "Hazards to Fish, Wildlife and Invertebrates: A Synoptic Review. United States Fish and Wildlife Service". Biological Reports, Vol. 85, No. 1.12 pp. 82-92.
- [17] De Filippis, L.F. and Pallaghy, C.K. (1994). Heavy Metals: Sources and Biological Effects. In: RAI, L.C.; GAUR, J.P. and SOEDER, C.J. eds. Advances in Limnology Series: Algae and Water Pollution, E.Scheizerbartsche Press, Stuttgart, pp. 31-77.
- [18] Nagasse, H., Inthorn, D., Isaji, Y, Oda, A., Hirata, K. and Miyamoto, K. (1997). "Selective Cadmium Removal from Hard Water using NaOH-Treated Cells of the Cyanobacterium Tolypothrix tenuis". J. Fermentation and Bioeng., Vol. 84, No 2 pp. 580-584.
- [19] Zhang, L., Zhao, L., Yu, Y. and Chen, C. (1998). "Removal of Lead from Aqueous Solution by Non-Living Rhizopus nigricans". Water Research, Vol. 32, No. 5 pp. 1437-1444.
- [20] Bai, R.S. and Abraham, A.E. (2003). "Studies on Chromium (VI) Adsorption-Desorption using Immobilized Fungal Biomass". Bioresource Technology, Vol. 87, pp. 17-26.
- [21] Mane, P.C. and Bhosle, A.B. (2012). "Potential of Nostoc muscrurom and Anabaena subcylindrica for the Bioremoval of some Metals from Aqueous Solution". Journal of Environmental Research and Development, Vol. 6, No. 3A pp. 702-708.
- [22] Rani, J.R., Hemambika, B., Hemapriya, J. and Rajeshkannan, V. (2010). "Comparative Assessment of Heavy Metal Removal by Immobilized and Dead Bacterial Cells: A Biosorption Approach". Global Journal of Environmental Research, Vol. 4, No. 1 pp. 23-30.

- [23] Nath, S. and Bhoumik, M. (2013). "Levels of Toxic Metals in Edible Fish from a Wetland of India". *International Journal of Environmental Sciences*, Vol. 3, No. 5 pp. 1509-1515.
- [24] Arunakumara, K.K.I.U. and Xuecheng, Z. (2008). "Heavy Metal Bioaccumulation and Toxicity with Special Reference to Microalgae". *J. Ocean Univ. Chin.*, Vol. 7, pp. 60-64.
- [25] Alam, M.G.M., Tanaka, A., Allinson, G., Laurenson, L.J.B., Stagnitti, F. and Snow, E.T. (2002). "A Comparison of Trace Element Concentrations in Cultured and Wild Carp (*Cyprinus carpio*) of Lake Kasumigaura, Japan".
- [26] Mansour, S.A. and Sidky, M.M. (2002). "Ecotoxicological Studies: 3 Heavy Metals Contaminating Water and Fish from Fayoum Gov. Egypt". *Food Chemistry*, Vol. 78, pp. 15-22.
- [27] Özparlak, H., Arslan, G. and Arslan, E. (2012). "Determination of some Metal Levels in Muscle Tissue of Nine Fish Species from Beyşehir Lake, Turkey". *Turk. J. Fish. Aquat. Sci.*, Vol. 12, pp. 761-770.
- [28] Toyub, M.A., Miah, M.I., Habib, M.A.B. & Rahman, M.M. (2008). "Growth Performance and Nutritional Value of *Scenedesmus obliquus* Cultured in Different Concentrations of Sweetmeat Factory Waste Media" *Bang. J. Anim. Sci.*, Vol 37. No. 1 pp. 86-93.
- [29] De la Noue, J., Laliberte, G. and Proulx, D. (1992). "Algae and Waste Water". *Journal of Applied Phycology*, Vol. 4, pp. 247-254.
- [30] An, J.N., Sim, S.N., Lee, J.S. and Kim, B.W. (2003). "Hydrocarbon Production from Secondary Treated Piggery Wastewater by the Green Algae *Botryococcus braunii*" *J. Appl. Phycol.*, Vol.15. pp. 185-191.
- [31] Órpez, R., Martínez, M.E., Hodaifa G, El Yousfi F, Jbari N, Sánchez. S (2009) "Growth of the Microalga *Botryococcus braunii* in Secondary Treated Sewage" *Desalination*, Vol. 36. Pp. 623-630.
- [32] Chinnasamy, S., Bhatnager, A., Hunt, R.W. and Das, K.C. (2010). "Microalgae Cultivation in a Wastewater Dominated by Carpet Mill Eluents for Biofuel Applications". *J. Biotech.*, Vol. 101, pp. 3097-3105.
- [33] Jordanova, A., Strezov, A., Ayranov, M., Petkov, N. and Stoilova, T. (1999). "Heavy Metal Assessment in Algae, Sediments and Water from the Bulgarian Black Sea Coast". *Water Sci. Tech.*, Vol. 39, pp. 207-212.

- [34] Bajguz, A. (2000). "Blockade of Heavy Metals Accumulation in *Chlorella vulgaris* Cells by 24-Epibrassinolide". *Plant Physiol. Biochem.*, Vol. 38, pp. 797-801.
- [35] Areco, M.M., Cainzos, V. and Curutchet, G. (2013). "Copper Removal by *Botryococcus braunii* with Associated Production of Hydrocarbons". *Advanced Materials Research*, Vol. 825, pp. 528-534.
- [36] Peña-Castro, J.M., Martínez-Jerónimo, F., Esparza-García, F. and Cañizares-Villanueva, R.O. (2004). "Heavy Metals Removal by the Microalga *Scenedesmus incrassatulus* in Continuous Cultures". *Bioresource Technology*, Vol. 94, No. 2 pp. 219–222
- [37] Balaria, A., Schiewer, S. and Trainor, T. (2005). Biosorption of Pb (II) Onto Citrus Pectin: Effect of Process Parameter on Metal Binding Equilibrium and Kinetics. *World Water Congress. Impacts of Global Climate Changes. World Water and Environmental Resources Congress 2005*. Raymond Walton-Editor, May 15-19, 2005 Anchorage, Alaska, USA.
- [38] Kothari, R., Pathak, V.V., Kumar, V. and Singh, D.P. (2012). "Experimental Study for Growth Potential of Unicellular Alga *Chlorella pyrenoidosa* on Dairy Wastewater: An Integrated Approach for Treatment and Biofuel Production" *Bioresource Technology*, Vol 116. Pp. 466-470.
- [39] Sigg, L. (1985). Metal Transfer Mechanism in Lake: The role of settling particles. In W. Stumm (ed). *Chemical Processes in Lakes*. John Wiley and Sons, New York. Pp. 285-310.
- [40] Sigg, L., Sturn, M. and Kistler D. (1987). "Vertical Transport of Heavy Metals by Settling Particles in Lake Zurich. *Limnology and Oceanography*, Vol. 32, pp. 112-130.
- [41] Pinto, E., Sigaud-Kutner, T.C.S., Leita~o, M.S.A., Okamoto, O.K. and Morse, D. (2003). "Heavy Metal-Induced Oxidative Stress in Algae". *J. Phycol.*, Vol. 39, pp. 1008-1018.
- [42] Clesceri, L.S., Greenberg, A.E. and Trussel, R.R. (1989). *Standards Methods for the Examination of Water and Wastewater 17th Edn*. American Public Health Association, Washington, DC.