

Alexandria University Faculty of Medicine

Alexandria Journal of Medicine





Gorham—Stout disease or new entity on the basis of vasculopathy



H. Hasan Yeter*

Hacettepe University, Internal Medicine, Ankara, Turkey

Received 24 December 2015; revised 11 February 2016; accepted 29 March 2016 Available online 25 May 2016

KEYWORDS

Gorham–Stout disease; Vanishing bone disease; Vasculitis; Thrombosis **Abstract** Gorham–Stout disease (GSD) is a rare osteolytic bone disease also known as vanishing bone disease. The pathogenesis of GSD is not well understood. Studies showed that lymphatic and blood endothelial cells in addition to macrophages secrete TNF α and IL-6 that stimulate osteoclast formation with osteolysis. Also TNF α secretion inhibits osteoblast differentiation and new bone formation. It is known that cytokines such as TNF α , IL-1 and complement system activation are responsible for inflammation and necrosis in the vessel wall at vasculitis. Both diseases have similar pathogenesis. Here, we presented a case of Gorham's disease with involvement of bilateral humerus, systemic arterial thrombosis and mesenteric vasculitis.

© 2016 Alexandria University Faculty of Medicine. Production and hosting by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

1. Introduction

Gorham–Stout disease (GSD) is a rare osteolytic bone disease also known as vanishing bone disease. It is an musculoskeletal disorder characterized by spontaneous and progressive osteolysis, angiomatous proliferation and soft tissue swelling without new bone formation, although first description of a humerus osteolysis in 1838 by Jackson. Moreover, in 1955, Gorham and Stout published a paper, which correlated the massive osteolysis noted in the disease with hemangiomatosis. The pathogenesis of GSD is not well understood. Studies showed that lymphatic and blood endothelial cells in addition to macrophages secrete TNF α , IL-1 β and IL-6 that stimulate osteoclast formation with osteolysis. Also TNF α secreted

E-mail address: hasanyeter@hotmail.com

Peer review under responsibility of Alexandria University Faculty of Medicine.

by macrophage and endothelial cells inhibits osteoblast differentiation and new bone formation. Additionally, the signaling pathway of the receptor of the lymphangiogenic growth factor Platelet Derived Growth Factor BB(PDGFR-b) and vascular endothelial growth factor (VEGF) family has the critical role that could play in the pathogenetic mechanism of the disease.^{4,5} Besides the bone disease, also lymphatic vascular malformations involving the skin and the soft tissues can be adjacent to the diseased bone. It can affect any part of the skeleton. The sites most affected are the cranium, shoulder and pelvis. 6 GSD shows no preference for gender or race and occurs more often in children and young adults. Although dissolution, fracture, fragmentation and disappearance of a part of a bone are typical at X-ray diagnosis of GSD is based on exclusion of other causes of osteolysis, such as infection, cancer, inflammatory and endocrine disorders.

Herein, we report a 50 years old female patient who had been presented with osteolysis of bilateral humerus, systemic thrombosis and mesenteric vasculitis.

^{*} Tel.: +90 5542397449.

2. Case report

A 50 year-old woman was admitted to the hospital having abdominal pain and anemia to find etiology of osteolysis of humerus. Her shoulder pain had started five years earlier with progressive loss of range of motion. She did not have any previous history of trauma but she was treated because of soft tissue infection. There were no document neither tissue nor blood culture positivity.

At the objective examination she presented with pain upon palpation of bilateral shoulder. She also could not raise left upper arm and right upper arm was limited to internal and external rotation, abduction and adduction. At her abdominal examination she had diffuse defence without rebound.

She underwent X-ray examination with the right and left shoulder (Fig. 1). X-ray showed the right humerus radiolucent foci (osteolytic lesions) and loss of tissue and left humerus proximal resorption of bone. MRI on bilateral humerus (Fig. 2) also showed complete bone resorption of left humerus head and neck and scapula's glenoid process and partial bone resorption of right humerus. Bilateral shoulder muscles widespread lipoid atrophy was seen. Her blood chemistry, parathyroid and thyroid hormone levels, serum immunoglobulins, complement levels, and tumor markers were all normal (Table 1). She had iron deficiency anemia and mild leukocytosis. Sedimentation rate was high and also she had anti nuclear anticor (ANA) positivity (1/320 titer). Lupus anticoagulants and hereditary thrombophilia tests were negative except for PAI heterozygosity (Table 2). Her Schirmer test was negative and salivary gland biopsy was normal. Her upper and lower endoscopy was normal except for gastritis.

Thorax CT showed thrombus in right subclavian artery and aorta and third costa focal bone destruction with normal pulmonary parenchyma. Abdominal CT angiography showed thrombus in left common iliac artery, mesenteric vasculitis, ileal wall thickness and partial intestinal obstruction. To exclude hematologic malignancy bone marrow aspiration and biopsy were performed. It was negative for malignancy and dysplasia.

She underwent humerus biopsy and culture. Her biopsy was reported nonspecific minimal chronic inflammatory process. Tissue tuberculosis and aerobe culture were both negative.

We started 1 mg/kg/day metilprednizolone as a treatment of vasculitis. After one month follow-up she had no abdominal pain and her acute phase reactants were normal. The second month of discharge she admitted to emergency department with acute abdominal pain and diffuse abdominal defence and rebound. Her abdominal CT showed intestinal perforation. After surgery she died because of septic shock.

3. Discussion

GSD is a rare condition that is characterized by loss of bone matrix, which is replaced by fibrotic tissue and proliferation of vascular and lymphatic canals. There is no association between GSD and vasculitis and thrombosis in the literature. We can also not explain the bone findings of our patient with vasculitis. Patient's X-ray and MRI findings are appropriate with GSD but lymphatic malformation and nonmalignant hyperproliferation of small vessels have not seen in bone biopsy taken from the patient. Presence of multiple thrombosis, mesenteric vasculitis and ANA positivity was considered systemic lupus erythematosus (SLE) but not enough to diagnose SLE to Systemic Lupus International collaborating clinics classification criteria for systemic lupus erythematosus.8 In SLE, lupus vasculopathy is found in up to 40% patient especially woman and typically characterized by inflammation and necrosis in the vessel wall. It is known that cytokines such as TNFα. IL-1 and complement system activation are responsible for inflammation and necrosis in the vessel wall. 10 Also TNFα, IL-1β and IL-6 stimulate osteoclast formation with osteolysis and the other way is TNFa inhibits osteoblast activation and new bone formation at GSD.^{2,3} Experimental animal models showed that, TNF α infusion was rapidly followed by thrombin generation and the study with the colorectal cancer patients also showed association between TNFa levels and venous thromboembolism risk. 11 Chronic inflammation or





Figure 1 Bilateral humerus X-ray. X-ray showed that right humerus radiolucent foci(osteolytic lesions) and loss of tissue and left humerus proximal resorption of bone.

Gorham–Stout disease 195



Figure 2 Bilateral humerus MRI of patients. MRI showed complete bone resorption of left humerus head and neck and scapula's glenoid process and partial bone resorption of right humerus. Bilateral shoulder muscles widespread lipoid atrophy was seen.

	Results	Normal ranga
	Results	Normal range
ALT ^a	7 μ/L	< 33
AST ^a	13 μ/L	< 31
GGT ^a	24 μ/L	< 36
ALP ^a	76 μ/L	< 104
T. Bilirubin	0.4 mg/dl	0.1-1.2
Albumin ^a	3.24 g/dl	3.4-4.8
Creatinine	0.52 mg/dl	0.5-0.9
CK ^a	141 μ/L	26-192
LDH ^a	$175 \mu/L$	240-480
TSH ^a	$1.7~\mu IU/L$	0.27 - 4.2
Hemoglobin	10.4 g/dl	11.7-15.5
MCV ^a	89.7 fl	80–95
Leukocyte	$14 \times 10^3 \mu L$	$4-11.2 \times 10^3$
Neutrophil	$12.9 \times 10^3 \mu$ L	$1.8-6.4 \times 10^3$
Thrombocyte	$352 \times 10^3 \mu$ L	$159-388 \times 10^3$
Sedimentation	66 mm/h	0–25
Ca 125 ^a	4.5	0–35 U/ml
Ca 19-9 ^a	32.5	0–35 U/ml
Ca 15-3 ^a	6.1	0-31.3 U/ml
CEA ^a	1.37	0-3 ng/mL

^a ALT: Alanine transaminase; AST: Aspartate aminotransferase; GGT: Gamma-glutamyl transferase; ALP: Alkaline phosphatase; CK: Creatine kinase; LDH: Lactate dehydrogenase; TSH: Thyroid stimulating hormone; MCV: Mean corpuscular volume; Ca: carbohydrate antigen; CEA: Carcinoembryonic antigen.

autoinflammatory diseases are also possible cause of thrombosis. 12

In a study conducted by Franchi et al. to find a diagnostic marker for GSD, CD105 expression was found to be significantly higher in GSD vessels compared to control group.¹³ Similarly, CD105 (endoglin) expression was significantly

Table 2 Immunologic tests.			
	Results	Normal range	
C-rp ^a	2.18	0–1 mg/dl	
ANA ^a	1/320	Negative	
Anti ds-DNA	Negative	Negative	
ENA-SsA ^a	Positive	Negative	
ENA-SsB	Negative	Negative	
ENA-Jo1	Negative	Negative	
ENA-Scl70	Negative	Negative	
Anti phospholipid IgM	Negative	Negative	
Anti phospholipid IgG	Negative	Negative	
Beta-2 glycoprotein IgM	Negative	Negative	
Beta-2 glycoprotein IgG	Negative	Negative	
Anti cardiolipin Antibody IgM	Negative	Negative	
Anti cardiolipin Antibody IgG	Negative	Negative	
ANCA profile ^a	Negative	Negative	
Anti HBc total	Negative	Negative	
Anti HBs	14n IU/ml	0–10	
Anti HCV	Negative	Negative	
TPHA ^a	Negative	Negative	
Brucella agglutination	Negative	Negative	
C3 ^a	128 mg/dl	79–152	
C4 ^a	25 mg/dl	16-38	
IgA	361	82-453 mg/dl	
IgG	1170	751–1560 mg/dl	
IgM	64	46–304 mg/dl	

^a CRP: C-reactive protein; RF: Rheumatoid factor; ANA: Anti nuclear anticor; C3: Complement 3; C4: Complement 4; ENA: extractable nuclear antigens; ANCA: antineutrophil cytoplasmic antibodies; TPHA: *Treponema pallidum* hemagglutination assay.

higher in systemic vasculitis and the expression of it correlates with disease activity. $^{14-16}$

H.H. Yeter

As a major molecule in angiogenesis, VEGF also contributes to the regulation of the immune system. ¹⁷ Significantly increased levels of VEGF were demonstrated in activation of vasculitis as well as in GSD. ^{2,3,18}

4. Conclusion

196

TNF α , IL-1 β , IL-6, VEGF and CD105 have pathophysiological role both in the vasculitis and in the GSD. This common pathway increases the risk of coexistence of both diseases. Thrombosis may be the result of direct effect of cytokines such as TNF α or chronic inflammation or autoinflammation. To our knowledge this report is the first combination of GSD, arterial thrombosis and vasculitis in the literature. Perhaps it is a new entity on the basis of vasculopathy including vasculitis, thrombosis and GSD like disease.

5. Conflict of interest

The authors declare that there is no conflict of interest.

References

- 1. Liu Y, Zhong D-R, Zhou P-R, et al. Gorham–Stout disease: radiological, histological, and clinical features of 12 cases and review of literature. *Clin Rheumatol* 2014;1–11.
- Faruqi T, Dhawan N, Bahl J, et al. Molecular, phenotypic aspects and therapeutic horizons of rare genetic bone disorders. *BioMed Res Int* 2014.
- Colucci S, Taraboletti G, Primo L, et al. Gorham–Stout syndrome: a monocyte-mediated cytokine propelled disease. *J Bone Miner Res* 2006;21(2):207–18.
- Hagendoorn J, Padera TP, Yock TI, et al. Platelet-derived growth factor receptor-β in Gorham's disease. Nat Clin Pract Oncol 2006;3(12):693-7.
- Venkatramani R, Ma NS, Pitukcheewanont P, et al. Gorham's disease and diffuse lymphangiomatosis in children and adolescents. *Pediatr Blood Cancer* 2011;56(4):667–70.

 Sá P, Marques P, Oliveira C, et al. Gorham's disease: clinical case. Revista Brasileira de Ortopedia (English Edition) 2015;50 (2):239–42.

- Nikolaou VS, Chytas D, Korres D, et al. Vanishing bone disease (Gorham–Stout syndrome): a review of a rare entity. World J Orthoped 2014;5(5):694.
- 8. Petri M, Orbai AM, Alarcón GS, et al. Derivation and validation of the systemic lupus international collaborating clinics classification criteria for systemic lupus erythematosus. *Arthritis Rheum* 2012;**64**(8):2677–86.
- Veller M. 16: Pathophysiology and principles of management of vasculitides and Raynaud's syndrome. Mech Vasc 2011;295.
- Belmont HM, Abramson SB, Lie J. Pathology and pathogenesis of vascular injury in systemic lupus erythematosus. Interactions of inflammatory cells and activated endothelium. *Arthritis Rheum* 1996;39(1):9–22.
- Roselli M, Ferroni P, Rolfo C, et al. TNF-α gene promoter polymorphisms and risk of venous thromboembolism in gastrointestinal cancer patients undergoing chemotherapy. *Ann Oncol* 2013, mdt251.
- La Regina M, Orlandini F, Manna R. Autoinflammatory diseases: a possible cause of thrombosis? *Thrombosis J* 2015;13(1):19.
- Franchi A, Bertoni F, Bacchini P, et al. CD105/endoglin expression in Gorham disease of bone. J Clin Pathol 2009;62(2):163–7.
- Erdbruegger U, Grossheim M, Hertel B, et al. Diagnostic role of endothelial microparticles in vasculitis. *Rheumatology* 2008;47 (12):1820-5.
- Brogan P, Shah V, Brachet C, et al. Endothelial and platelet microparticles in vasculitis of the young. *Arthritis Rheum* 2004;50 (3):927–36.
- Brogan PA, Dillon MJ. Endothelial microparticles and the diagnosis of the vasculitides. *Int Med* 2004;43(12):1115–9.
- Hamamichi Y, Ichida F, Yu X, et al. Neutrophils and mononuclear cells express vascular endothelial growth factor in acute Kawasaki disease: its possible role in progression of coronary artery lesions. *Pediatr Res* 2001;49(1):74–80.
- Song GG, Kim J-H, Lee YH. Vascular endothelial growth factor gene polymorphisms and vasculitis susceptibility: a meta-analysis. *Hum Immunol* 2014;75(6):541–8.