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Effects of *roxazyme-G* on growth indices and haematological variables of broilers fed maize offal-based diets

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ABSTRACT

Influence of *roxazyme-G* on the utilization of maize offal was investigated using 420 broiler chicks and 210 (5-week old) growing broilers at starter and finisher phases, respectively. A basal diet that met the nutrient requirement for each phase was formulated and used for the trial. Basal diet meant for starter phase contained 529 g kg⁻¹ maize while basal diet for finisher phase contained 569 g kg⁻¹ maize. For each phase, 25% or 50% maize component of each basal diet were replaced with maize offal. Thereafter, the maize offal-based diets were divided into 3 equal parts and designated diets 2, 3 and 4 for diets in which its 25% maize component was replaced with maize offal and diets 5, 6 and 7 for those with their 50% maize components replaced with maize offal. *Roxazyme-G* was added to the diets at levels of 100, 200 and 300 mg kg⁻¹ in diets 2 & 5, 3 & 6 and 4 & 7, respectively. The birds were fed their respective diets for 21 days. At the close of the starter (2-4 weeks of age) and finisher (5-8 weeks of age) phases 10 chicks and 10 chickens per replicate, respectively were sacrificed conventionally and their blood collected for blood analysis. Of the growth indices measured only the final weight of chicks was significantly ($P \leq 0.05$) influenced by the dietary treatment. The entire haematological indices measured were not significantly ($P \geq 0.05$) influenced by dietary treatment in both starter and finisher birds. At starter phase, the optimum level of maize offal inclusion could be achieved at 132.3 g kg⁻¹ + 200 mg kg⁻¹ *roxazyme-G* while 142.3 g kg⁻¹ maize offal + 200 mg kg⁻¹ *roxazyme-G* was for finishers. Conclusively, the use of maize offal and *roxazyme-G* at these optimal levels could lead to more of maize offal being included in broiler diets in tropical countries.

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Keywords: Finisher, maize offal, starter, *roxazyme-G*.

INTRODUCTION

Report by Nnenna et al. (2006) and recently by Aro et al. (2010) showed that agricultural by-products have in recent years become important dietary component for non-

ruminant animals in most tropical countries, including Nigeria. Maize offal is a by-product obtained from maize grain after wet or dry milling of maize. Its potential as filler or energy diluents in non-ruminant

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feeding has been reported by Uko et al. (1999) and Iyayi (2004). Birds fed on such feeds have poor feed consumption and a concomitant reduced weight gain and, this according to Agbede et al. (2005) could be attributed to high dietary fibre content.

Conceivably, improving the nutritive value of this fibrous ingredient could lead to more of it been incorporated in non-ruminant diets in tropical countries. Improved utilization of maize offal by non-ruminant could be achieved if such maize offal-based diets are supplemented with exogenous enzymes such as *roxazyme-G*. *Roxazyme-G* is an enzyme produced by fungus (*Trichoderma*). It's potential in reducing the viscosity of intestinal content with an improved feed utilization after consumption of non-starch polysaccharides (NSP) by non-ruminant have been reported by Pitson et al. (1993) and Agbede et al. (2002). However, to our knowledge, it appears that there are limited studies conducted to examine the influence of *roxazyme-G* on the utilization of maize offal by broilers. Consequently, this study was conducted to examine the utilization of maize offal by broiler chickens as influenced by *roxazyme-G* using performance and blood indices as response criteria.

MATERIALS AND METHODS

Experimental diets

Two basal diets were formulated to meet NRC (1994) requirements for broiler starter and finisher (Table 1). At both starter and finisher phases, the two basal diets contained 529 g kg⁻¹ and 569 g kg⁻¹ maize, respectively without enzyme or maize offal. For each phase, 25% or 50% maize component of each basal diet were replaced with maize offal. Each diet with their maize

replaced at 25 or 50 % maize offal was mixed in one batch, divided to three equal portions and designated diets 2,3 & 4 for diets in which its 25% maize component was replaced with maize offal and diets 5,6 & 7 for those with their 50% maize components replaced with maize offal. *Roxazyme-G* was added to the diets at and above the manufacturer recommended levels of 100, 200 and 300 mg kg⁻¹ in diets 2 & 5, 3 & 6 and 4 &7, respectively.

Animal management and experimental Layout

Starter phase

A batch of 420 day-old Arbor Acre broiler chicks were obtained from Ajanla farms, Ibadan, Nigeria. The chicks were electrically brooded together on the floor for the first three days during which they were sexed as described by Laseinde and Oluyemi (1997). The chicks were thereafter transferred into metal metabolism cages which have facilities for faecal collection. During the pre-experimental period, the chicks were fed on commercial feed (Livestock feed: 230 g kg⁻¹ crude protein; 11.97 MJkg⁻¹ ME). After the first week, the chicks were randomly divided into seven groups of sixty (60) such that the groups mean weights were similar in a completely randomized design. The treatments were replicated thrice with 20 birds (10 males + 10 females) per replicate. The birds were fed for 21 days with their respective experimental diets. The routine veterinary practices as outlined by the Teaching and Research farm of the Federal University of Technology, Akure, were followed.

Finisher phase

Two hundred and ten 5 weeks old growing broiler birds left from the starter

phase were used for this trial. Prior to the commencement of the trial, the birds were fed on commercial feed (Livestock feed: 21 g kg⁻¹ crude protein; 12.6 MJkg⁻¹ ME). The birds were randomly divided into seven groups of thirty (30) each in a completely randomized design. The treatments were replicated thrice with 10 birds (5 males and 5 females) per replicate. This phase also lasted for 21 days.

Blood collection

Three hours to the close of the finisher phase, the birds were starved so as to empty the crops and weighed. Thereafter, five males and five females were randomly selected per replicate in each phase and all were sacrificed conventionally by severing the jugular veins with sharp knives. The blood was then allowed to flow freely into labeled bijou bottles which contained a speck of Ethylene Diamine Tetra Acetic Acid (EDTA) for analysis.

Analyses

The proximate composition of the ingredients used before the feed formulation and the test diet samples were analyzed by the methods of A.O.A.C (1990). The micro-Kjeldahl method was used for crude protein determination (% N x 6.25) while the soxhlet extraction method was used for fat determination. The crude fibre was determined by alkali and acid hydrolysis of the feed samples. The packed cell volume (PCV) was estimated by spinning about 75 μ l of each blood sample in heparinized capillary tubes in a haematocrit micro centrifuge for 5 minutes while the total red blood cell count (rbc) was determined using normal saline as the diluting fluid. The haemoglobin concentration (Hbc) was estimated using cyanomethaemoglobin

method while the erythrocyte sedimentation rate (ESR) was determined as described by Lamb (1981).

Calculation and statistical analyses

The performance indices were calculated per unit bird. The weight gain was calculated by subtracting the initial weight of birds from the final weight. The average weekly feed consumption was calculated as the total feed consumed for the experimental days divided by the number of weeks while the feed conversion ratio (FCR) was the ratio of average feed consumed to average weight gain. The mean corpuscular haemoglobin concentration (MCHC), mean corpuscular haemoglobin (MCH) and the mean corpuscular volume (MCV) were calculated as described by Lamb (1981) as follows:

$$\text{MCHC} = \frac{\text{Haemoglobin concentration} \times 100}{\text{PCV}} \%$$

$$\text{MCV} = \frac{\text{PCV} \times 10 (\mu^3)}{\text{RBC}}$$

Data collected were subjected to statistical analysis using analysis of variance (ANOVA) of SPSS 15 (2006) package. The significant treatment means were compared using the Duncan option of the same software.

RESULTS AND DISCUSSION

Performance indices

Table 2 shows that only the final weight (FW) of the chicks were significantly ($P \leq 0.05$) influenced by the dietary treatment. Chicks on the basal diet had the highest FW (547.2 \pm 1.0g) and this was significantly ($P \leq 0.05$) higher than those fed the test diets with chicks on 50% maize offal + 200 mg kg⁻¹ roxazyme-G-based diet having the

least FW (461.1 ± 3.0 g). Also, the chicks fed on 25% and 50% maize offal + enzyme-based diets had 7.7 – 10.7% and 10.4 – 15.7% respectively, lesser weight than those on the basal diet. This observation could be attributed to the inability of the chicks to effectively digest and utilize the dietary fibre of maize offal-based diets despite the *roxazyme-G* inclusion. While this observation is consistent with earlier report by (Ojewola et al., 2003), it is contrary to the reports by Vranjes and Wenk, (1995), Alp et al. (1999) and Agbede et al. (2002) that *roxazyme-G* enhances growth performance in poultry. Furthermore, chicks on diets containing 25% maize offal in most cases had higher FW, average weight gain (AWG) and average feed consumption (AFC) than those on 50% maize offal-based diets suggesting that the optimum level of maize offal substitution for maize in starter diet could be achieved at 25% but with *roxazyme-G* inclusion level at 200 mg kg⁻¹.

At the finisher phase, the FW, AWG, AFC and feed conversion ratio (FCR) of chickens on the basal diet were not significantly ($P \geq 0.05$) different from those fed on the test diets (Table 2). The non significant differences observed could be attributed partly to *roxazyme-G* inclusion which led to improve digestibility, absorption and utilization of non starch polysaccharide (NSP) by chickens on maize offal-based diets. This is achievable by unlocking the nutrients in maize offal-based diets through reduction in gut viscosity in the intestinal lumen by the enzyme. Also, it could be attributed to the ability of adult birds to tolerate higher dietary fibre than the chicks. This could be due to the well developed gizzard of the adult birds (Agbede et al., 2005). Birds on 25% Maize offal + *roxazyme-G* diets had higher FW than those fed 50% Maize offal + *roxazyme-G* when compared on level to level of enzyme supplementation

(1790 vs 1660 g), (1800 vs 1710 g) and (1770 vs 1750 g) for 100 mg, 200 mg and 300 mg kg⁻¹, respectively, but this was not significant. However, this further confirms that the optimum level of maize offal inclusion in broiler finisher diets is 25% with 100 or 200 mg kg⁻¹ *roxazyme-G* supplementation.

Haematological studies

Table 3 shows that the haematological variables of the chicks were not significantly ($P \geq 0.05$) influenced by the dietary treatments. However, the PCV ranged: 25.3- 28.3%, RBC $1.9-3.0 \times 10^6$ mm³, Hbc 4.0-5.3 g/100 ml, MCHC 15.4-19.4%, MCV 90.7-147.4 μ^3 m and ESR 2.5-4.3 mmhr⁻¹. Also at the finisher phase (Table 4), all the haematological variables evaluated were not significantly ($P \geq 0.05$) influenced by dietary treatments but the group fed on 50% maize offal + 300 mg kg⁻¹ *roxazyme-G* had in most cases the least values for the haematological variables (except MCV and ESR).

The haematological values obtained for the finished birds were in most cases lower than those reported for broilers fed maggot-based diets by Awoniyi et al. (2004). For instance, a range of 29.7-34% PCV, 2.2-2.7 RBC $\times 10^6$ mm³ and 7.5-8.0 g/100ml Hbc was report by Awoniyi et al. (2004). The values obtained for PCV, RBC and Hbc in birds fed the test diets were similar to those fed the basal diets at both phases, thus indicating the potential of *roxazyme-G* at improving the nutritive value of maize offal-based diets in such away that they promoted identical blood synthetic activities in the experimental birds. Similarly, the values of MCHC and MCV for chickens fed on the basal and test diets were similar. This could imply that all the blood samples collected and analyzed had similar

Table 1: Basal composition of experimental diets (g kg⁻¹).

Phases % Maize offal <i>Roxazyme-G</i>	Starter			Finisher		
	Basal 0	25% (diets 2-4) ← 100-300 mg kg ⁻¹ →	50% (diets 5-7)	Basal 0	25% (diets 2-4) ← 100-300 mg kg ⁻¹ →	50% (diets 5-7)
Maize	529.0	396.7	264.5	569.0	426.6	284.5
Maize offal	-	132.3	264.5	-	142.3	284.5
Soya bean meal	340.0	340.0	340.0	330.0	330.0	330.0
Fishmeal	30.0	30.0	30.0	-	-	-
Others*	131.0	131.0	131.0	101.0	101.0	101.0
Total	1000.0	1000.0	1000.0	1000.0	1000.0	1000.0
Calculated						
CP (g kg ⁻¹)	230.0	230.0	230.0	212.0	212.0	213.0
ME(MJ kg ⁻¹)	12.2	12.1	12.0	12.3	12.2	12.1
Cyst +Meth (%)	0.9	0.9	0.8	0.8	0.8	0.8
Leucine (%)	2.0	2.0	2.0	1.9	1.9	1.9
Analyzed						
CP (g kg ⁻¹)	233.0	233.0	230.0	213.0	210.0	211.0
Crude fibre “	31.0	43.0	52.0	42.0	55.0	63.0
Ether extract “	13.0	21.0	11.0	24.0	20.6	12.3

*Brewer's dried grain = 50g, palm oil = 10g, bone meal = 25g, oyster shell = 5g, DL Methionine = 1g, premix = 5g and salt = 5g kg⁻¹; CP = Crude protein

Table 2: Performance indices of broilers fed maize offal-based diets with *roxazyme*-G supplementation.

Diets	% maize replaced with maize offal	Enzyme inclusion (mgkg ⁻¹)	Initial weight	Final weight	Average weight gain/chick/week	Average Feed consumption (g/bird/week)	FCR
			(g)				
			← Starter		Phase →		
1	0	0	70.9±4.2	547.2 ^a ±1.0	182.7±3.5	302.4±9.1	1.7±0.1
2	25	100	70.8±8.3	501.4 ^b ±2.9	166.6±9.8	284.2±2.8	1.7±0.1
3	25	200	70.8±4.2	508.3 ^b ±2.7	169.4±9.1	284.9±23.1	1.7±0.1
4	25	300	70.8±4.2	488.9 ^c ±1.2	162.4±4.2	266.7±28.7	1.6±0.1
5	50	100	70.8±4.2	476.9 ^c ±3.5	158.2±11.9	280.0±7.0	1.8±0.1
6	50	200	70.8±4.2	461.1 ^d ±3.0	154.0±9.8	263.2±7.7	1.7±0.1
7	50	300	70.1±2.4	490.3 ^c ±4.8	163.8±1.4	280.7±7.7	1.7±0.1
			Finisher		Phase		
1	0	0	670±0.5	1790±0.9	352.1±47.6	915.6±9.1	2.6±0.2
2	25	100	670±0.5	1790±0.4	353.5±21.0	906.5±9.1	2.6±0.1
3	25	200	670±0.4	1800±0.4	374.5±8.4	954.8±1.8	2.6±0.1
4	25	300	680±0.2	1770±0.6	366.1±6.4	927.5±2.1	2.5±0.1
5	50	100	670±0.3	1660±0.7	329.7±14.7	927.5±1.9	2.8±0.1
6	50	200	670±0.2	1710±0.6	345.1±23.1	929.6±3.8	2.6±0.2
7	50	300	680±0.4	1750±0.5	359.8±18.9	953.4±1.5	2.7±0.2

Means with different superscript along the same column are significantly (P≤0.05) different; FCR = Feed conversion ratio.

Table 3: Haematological variables in starter-broiler fed maize offal-based diets with roxazyme-G supplementation.

Diets	% Maize replaced with maize offal	Enzyme inclusion (mg kg ⁻¹)	Packed Cell Volume (%)	Red blood Cell x 10 ⁶ mm ³	Haemoglobin (g/100ml)	MCHC (%)	MCV (μ ³ m)	ESR (mm/hr)
1	0	0	28.0±2.0	1.9±0.3	4.3±1.4	15.4±4.3	147.4±1.8	2.5±0.5
2	25	100	25.3±1.7	1.9±0.3	4.2±3.4	16.6±5.6	133.3±2.4	3.2±1.0
3	25	200	26.3±2.4	3.0±0.8	5.1±3.4	19.4±2.8	90.7±2.1	3.5±0.9
4	25	300	25.3±3.2	2.2±0.4	4.0±0.8	15.8±4.7	121.0±3.3	3.3±0.6
5	50	100	28.0±1.7	2.3±0.7	5.1±3.4	18.2±3.5	128.3±3.3	3.3±0.6
6	50	200	28.3±1.2	3.0±1.3	5.3±3.6	18.7±2.1	123.8±4.7	3.7±0.6
7	50	300	26.3±4.5	3.0±0.7	5.1±0.0	19.4±2.3	93.2±2.1	4.3±1.5

MCHC- Mean corpuscular haemoglobin concentration; MCV- Mean corpuscular volume; ESR - Erythrocyte sedimentation rate. Means with different superscript along the same column are significantly different ($P \leq 0.05$).

Table 4: Haematological variables in broiler - finishers fed maize offal-based diets with *roxazyme*-G supplementation.

Diets	Levels of maize replaced by maize offal (%)	Enzyme Inclusion Level (mg kg ⁻¹)	Packed Cell Volume (%)	Red Blood Cell (x10 ⁶ mm ³)	Haemoglobin (g/100ml)	MCHC (%)	MCV (µm ³)	ESR (mm/hr)
1	0	0	32.0±2.6	2.9±0.1	10.7±3.5	33.3±10.6	110.1±4.2	1.7±0.6
2	25	100	31.7±1.2	2.9±0.2	8.6±3.9	27.6±13.2	109.2±5.1	1.5±0.3
3	25	200	28.3±4.0	2.9±0.4	10.5±0.8	37.3±4.5	97.3±10.1	3.0±0.8
4	25	300	29.7±1.5	2.9±0.3	11.6±0.7	39.1±1.8	102.7±6.6	1.5±0.5
5	50	100	28.7±2.1	2.8±0.1	9.3±2.2	32.5±7.1	102.8±1.8	1.7±0.6
6	50	200	30.0±2.7	3.0±0.4	10.9±2.7	36.0±6.3	100.6±6.3	1.5±0.0
7	50	300	26.3±0.6	2.6±0.1	6.4±2.1	24.2±7.9	102.2±3.3	2.0±0.0

MCHC- Mean corpuscular haemoglobin concentration; MCV- Mean corpuscular volume; ESR - Erythrocyte sedimentation rate. Means with different superscript along the same column are significantly different (P ≤ 0.05).

cellular haemoglobin contents irrespective of the levels of maize offal and *roxazyme-G* inclusion in the diets. The relevance of MCHC measurement lies in its use in the diagnosis of anaemia and also as an index of the capacity of bone marrow to produce RBC (Aletor and Egberongbe, 1992).

From the forgoing, 132.3 g kg⁻¹ maize offal + 200 mg kg⁻¹ *roxazyme-G* and 142.3 g kg⁻¹ maize offal + 200 mg kg⁻¹ *roxazyme-G* appeared to be the optimal levels of inclusion for both maize offal and *roxazyme-G* supplementation for chicks and growing broilers diets, respectively.

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