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EFFECT OF METHODS OF EXTRACTION ON PHYTOCHEMICAL CONSTITUENTS AND ANTIBACTERIAL PROPERTIES OF tetracarpidium conophorum SEEDS

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ABSTRACTS

Effect of methods of extraction on phytochemical constituents and antibacterial properties of the extracts of the seeds of *tetracarpidium conophorum* was studied. Different solvents extraction methods yielded varying amounts of the phytochemical constituents and the antibacterial screening showed that all the extracts except the water extracts have antibacterial activity. The inhibition of bacterial growth showed that the extracts were dose dependent since no inhibition occurred at lower concentration. The study therefore showed that the extracts of the seeds of *tetracarpidium conopharum* are dependent upon the solvent and methods of extraction, and that the extract possesses antibacterial properties, which could be useful in treatment of bacterial infections and or other related diseases.

KEY WORDS: Extraction methods, Chemical constituents, medicinal properties, *tetracarpidium conophorum*.

INTRODUCTION

Tetracarpidium conophorum (African walnut) belongs to the family of Euphorbiacea. It is a perennial woody climber commonly found in low bush especially in Africa, America, Europe and the Asia (Hutchinson and Dalzier, 1987). The leaves are globrous, ovate, long and margin toothed. The bases of the leaves are broaded and rounded up to the 5×3 inches with slender petioles up to 2 inches long. The fruits are four winged, ridged between wings and up to 3 inches in diameter. The seeds are edible even when raw. They have a bitter taste and a tonic effect like Kola. They can be eaten raw, cooked or with roasted corn. (Nuhu et al, 2002; Okerulu and Ani, 2001). The plant is known in Africa especially in Nigeria for its antibacterial efficacy, Decoction of the leaves and seeds serve as beverages, relieves abdominal pains and fever. The water extracts of the roots provides soothing beverage for fever and malaria infection (Nuhu et al, 2000; Balch and Balch, 2000). The method of extraction could affect the phytochemical contents and antibacterial properties of this medicinal plant. Therefore, the present study focused on the evaluation of the effects of extraction method on phytochemical constituents as well as the antibacterial properties of the seeds extracts.

MATERIALS AND METHODS

Collection and treatment of samples

The seeds of teracarpidium conophorum plant were obtained at the Marian Market (Calabar, Nigeria). The seeds were sun dried for seven days, shells removed, and ground into fine powder using an electric grinder. Then 100g of the powdered mass obtained was stored in a clean sterile bottle at room temperature, and used for the various extractions. The soxhlet ethanolic extract was obtained by soxhlet extraction of 20g in 100ml of 95% ethanol at 78°C using soxhlet apparatus. The extract was then concentrated to 10ml on a water bath, and dried at room temperature. The n-hexane extract was obtained by dissolving 20g of the powdered mass in 100ml n-hexane in a conical flask. The mixture was stirred, covered, allowed to stand for 24hrs, and filtered using sterile Whatman No.42 filter paper. The bright yellow filtrate (extract) was concentrated to 10ml on a water bath, and evaporated to dryness at room temperature.

The ethylacetate, 95% ethanolic, and distilled water extracts were obtained by repeating the above procedure for n-hexane extract. The n-hexane, ethanol and soxhlet extracts were light brown in colour while the water extract was pale yellow. The various extracts obtained were then used for determination of phytochemical contents (i.e. steroids, alkaloids, tannins, saponnins and flavonoids) as well as antibacterial analysis.

Estimation of phytochemical contents

Qualitative test for steroids

2.0ml of acetic anhydride was added to 0.15g of each sample extract, mixed and then 2.0ml of 1N H_2SO_4 was added. The colour changed from violet to blue in the sample, indicating absence of steroids.

Estimation of alkaloids contents

The alkaloid content of each extract was determined by the method of Harborne (1973). About 5.0g of each sample was weighed into a 250ml beaker, and a 200ml of 10% acetic acid in ethanol was added, and allowed to stand for 4hrs. This was filtered using whatmann No.42 filtered paper, concentrated in water bath to $\frac{1}{4}$ (50ml) of the original volume. Then, concentrated NH₄OH was added dropwise to each

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extract until the precipitate was complete. The suspension was allowed to settle and the precipitate was collected, washed with NH_4OH and then filtered. The residue was dried and weighed. The percentage alkaloid was then calculated.

Estimation of tannin content

Tannin content was determined by the method of Van Burden and Robinson (1981). 500mg of each sample extract was weighed into a 50ml plastic bottle,50ml distilled water added, shaken for 1hr in a mechanical shaker, filtered into a 50ml volumetric flask, and made up to the mark with distilled water. Then 5.0ml of each sample was mixed with 2.0ml of 0.1M FeCl₃ in 0.1N HCl and 0.008M potassium ferricynide was added, allowed to stand for 10 minutes and the absorbance read at 120nm in 6400/6400 spectrophotometer(Jenway,Essex,England).

Estimation of saponin content

Saponin content was determined by the method of Obadoni and Chuko (2001). About 2.0g of each extract was mixed with 100ml of 20% ethanol, and were incubated in a water bath at 55°C for 4hrs with stirring. The mixture was filtered and the extract was reextracted with 200ml of 20% ethanol. The combined extract was concentrated to 40ml in a water bath at 90°C. The concentrate was then transferred into a 20ml separatory funnel and 20ml diethyl ether was added, and shaken vigorously. The aqueous layer was recovered while the ether layer was discarded, the purification process was heated and 60ml of n-butanol added. The combined n-butanol extracts were washed twice with 10ml of 5% aqueous NaCl, and the remaining solution was heated in a water bath. After evaporation, the samples were dried in the oven to a constant weight, and the percentage saponnin content was calculated.

Estimation of flavonoid content

The flavonoid content of the extracts was estimated by the method of Boham and Kopicalabyazen (1974). About 1.0g of each sample was extracted repeatedly with 100ml of 80% methanol. The whole solution was then filtered through Whatmann No.42 filtered paper. The filtrate was transferred into a crucible and evaporated to dryness on a water bath, and weighed. The percentage flavonoid content was then calculated.

Estimation of Glycosides

The glycosides content of the extracts was determine by dissolving 10.0g of the extracts in 100ml of 50% H_2SO_4 in test tubes. The mixture was heated in boiling water for 15minutes, and 10ml of Fehling solution added, and the mixture boiled. A red precipitate in each extract tested, indicated the presence of glycosides. The percentage glycoside was calculated.

Antibacterial screening

The antibacterial analysis was carried out by the disc diffusion method of Kirby Bauer described by Duguid et al, (1989). This method determines the antibacterial activity of the extracts. The method involves preparation of nutrient medium, culture and inoculation. The inoculated extracts were then diluted serially and

examined for inhibition zones. The presence of zones of inhibition indicates activity.

Preparation of the nutrient medium

Nutrient agar medium was prepared by dissolving 2.8g of nutrient agar in 100ml distilled water. The solution was sterized in an autoclave at 121°C at 1.1N pressure for 15min. the suspension was cooled and poured into sterile Petri-dishes to solidify. The agar depth of the medium was 4mm.

Preparation of cultures and inoculation

Pure cultures of coliform bacillus, staphylococcus epidermidis and streptococcus viridians obtained from the microbiology laboratory, Cross River University of Technology, Calabar, Nigeria, were separately used to inoculate the Petri-dishes. This was done by streaking the surface of the plates in a zig zag manner until the entire surface was covered. The inoculated plates were then incubated at room temperature for 24hrs.

Assay of extracts

The extracts were serially diluted to obtain 1.0%, 0.5%.0.25% and 0.125% solutions of the extracts in sterile test tubes. Sterilized 9mm filter paper discs soaked in the dilute extracts were placed on the plates and incubated for 24hrs. at room temperature. The plates were examined for clear zones of inhibition. Presence of zones of inhibition indicated activity. The zones were measured.

Estimation of Glycosides RESULTS

presents the results Table 1 of the of phytochemical constituents the seeds of tetracarpidium conophorum. The results showed that alkaloids, tannins, saponnins, flavanoids and glycosides are present in the extracts. The results showed that the soxhlex extraction methods has the highest content of tannins, flavonoids, alkaloids, saponins and glycosides (42.2±0.42. 41.21±0.41,40.34±0.51,31.28±0.34 and 20.17±0.17±0.24) respectively. The ethanolic extraction method came second in content of tannins alkaloids, glycosides flavonoids. saponins and (33.21±0.38,32.83±0.21,32.23±0.34,30.11±0.32,and 20.13±0.20)respectively. This was followed by the ethyl acetate extraction methods with higher values in glycosides and saponins. Water extraction methods came fourth with higher values in glycosides and saponin content. Generally there was no significant difference (p>0.05) in the glycosides contents in the extracts obtained from all the different solvent extraction methods.

Table 2 presents the results of antibacterial activity. The results showed that all the extracts except the water extract have antibacterial activity. Table 3 presents the results of inhibition of bacterial growth by the extracts. The results showed that the effects of the extracts are dose dependent since no activity was observed at very low concentrations. Also the results showed that higher dose of the extract will be useful in the treatment of bacterial infections. Further, the steroid extracts of these seeds can be useful in the regulation of reproductive

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system particularly as sex hormones and d could be useful

useful in the correction of impotence.

Extract	Alkaloids (%)	Tannins (%)	Flavonoids	Saponnins	conophorum Glycosides	
			(%)	(%)	(%)	
n-Hexane	8.36±0.24	7.14±0.22	8.12±0.25	12.24±0.22	19.23±0.21	
Ethyl acetate	9.54±0.21	9.13±0.20	9.21±0.24	12.23±0.26	19.24±0.18	
Ethanol	32.83±0.21	33.21±0.38	32.23±0.34	30.11±0.32	20.13±0.20	
Soxhlet	40.34±0.54	42.21±0.42	41.21±0.41	31.28±0.34	20.17±0.24	
Water	8.93±0.18	8.13+0.21	9.23+0.24	13.20+0.23	21,23+0,20	

Table 2: The antibacterial activity of the extracts of the seeds of tetracarpidium conophorum

Test organism	n-Hexane	Ethylacetate	Ethanol	Soxhlet	Water	
Coliform bacillus	-	+	+	+	_	
Straphylococcus epidermis	+	+	+	+	-	
Streptococcus viridans	+	+	+	+	-	

+ = active

- = not active.

Table 3: Inhibition of bacterial growth by the extracts of the seeds of tetracarpidium conophorum

Test organism	Dilution	Zone of inhibition (mm)				
	(%)	n-hexane	Ethyl acetate	Ethanol	Soxhlet	Water
Coliform bacillus	1.00	_	4.0	5.0	6.0	_
	0.50	_	2.5	3.0	4.5	_
	0.25	_	_	_	_	_
	0.125	_	_	_	_	_
Straphylococcus epidermis	1.00	3.00	5.0	5.5	7.0	_
	0.50	1.0	3.0	3.5	4.0	
	0.25	_	1.5	_		
	0.125	_	_	_	_	_
Streptococcus viridans	1.00	6.0	5.5	6.0	6.5	
	0.50	3.5	3.0	3.0	3.5	
	0.25		_			
	0.125					

DISCUSSION

The phytochemical screening and quantitative estimation of the percentage crude vield of phytochemical constituents of the extracts of the seeds of tetracarpidium showed that the seeds are rich in alkaloids, flavonoids, tannins, saponnins and glycosides; and also possess antibacterial properties as well as physiological activity (Sofowora, 1993; Ekam and Ebong, 2007). Flavonoids are most commonly known for their antioxidant activity. They are modifiers which modify the body's reactions to allergens, viruses, and carcinogens. They show anti-allergic, anti-inflammatory, antimicrobical and anticancer activity (Balch and Balchi, 2000; Ekam and Ebong, 2007), and may be useful in therapeutic roles (Jisika et al, 1992). Alkaloids are organic compounds that contain nitrogen, and are physiologically active with sedative and analgesic properties. They are used in relieving pains, anxiety and depression (Jisika et al, 1992). Alkaloids are toxic due to their stimulatory effects, leading to excitation of cells and

neurological dysfunction (Obochi, 2006; Ekam and Ebong, 2007).

Glycosides are compounds containing a carbohydrate and non-carbohydrate residue (moiety) in the same molecule. In these compounds, the carbohydrate moiety is attached by an acetal linkage of carbon-1 to the non-carbohydrate residue (aglycone). They all contain steroid as aglycone component in combination with sugar molecules. They are important in medicine because of their action on heart and are used in cardiac insufficiency (Balch and Balch, 2000). Thus, cardiac glycosides are drugs and can be used in the treatment of congestive heart failure and cardiac arrhythmia. They work by inhibiting the Na⁺/Na⁺ pump, resulting in an increase in the levels of sodium ions in the myocytes, which then leads to a rise in the level of calcium ions. This inhibition increases the amount of Ca2+ ions available for concentration of the heart muscle, improves cardiac output and reduces distention of the heart (Bertorello et al, 1990; Clausen and Nielson, 1994; Beltowski et al, 1998). These glycosides are found

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as secondary metabolites in several plants and animals (Clausen, 1996; Beltowski et al, 1998).

However, some glycosides, such as ouabain, are toxic as it inhibits active transport of Na^+ in cardiac muscle (sodium pump inhibitor), resulting in inhibition of translocases during electron transport chain, and leading to death (Beltowski et al, 1998). Also phloridzin (toxic glycosides) blocks the transport of sugar across the mucosal cells of small intestine and also renal tubular epithelium; it displaces Na^+ from the binding sites of carrier protein and prevents the binding of sugar molecule and produces glycosuria (Clausen, 1994; Gloor, 1997; Beltowski et al, 1998).

Tannins (tannic acids) and saponnins are responsible for the antibacterial activity of the plant seed extracts (Gloor, 1997). Saponnins are used in veterinary vaccines as adjuvant (e.g. foot-and-mouth disease vaccines) helping to enhance immune response. They are also mild detergents and can be used commercially as well as for research (Belch and Belch, 2000). They can also be used in intracellular histochemistry staining to allow antibody access to intracellular proteins (Belch and Belch, 2000). Also, because of its ability to permeate cells without destroying cell morphology, it is used in laboratory applications to treat live cells in order to facilitate peptide or reagents such as antibodies entering cells instead of the detergents (Belch and Belch, 2000).

The results in this study have shown that different solvent extraction method affect changes in the content of the extracts depending on the solvent used as well as the method of extraction. The results for antibacterial screening have shown that all the extracts except the water extract have antibacterial activity. The antibacterial activity is due mainly to the presence of tannins and saponnins (Okerulu and Ani, 2001). The results of the inhibition of bacterial growth have shown that the effects of the extracts appear to be effective at higher concentration. The soxhlet extract was found to be the most effective, appears to be non-target specific; and can be beneficial in the treatment of bacterial infections.

In conclusion, this study has shown that the phytochemical contents of the extracts of the seeds of tetracarpidium conophorum are dependent upon the solvent and method of extraction. The results have also shown that these extracts possess antibacterial properties and that the inhibition of bacterial growth was dose dependent. The results of this study may suggest these extracts may be useful in the treatment of bacterial infections and other related diseases as well as enhancing immune responses.

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