Full Length Research Paper

# Study on relationship between microsatellite polymorphism and producing ability on litter size trait of Hu sheep in China

W. Sun<sup>1</sup>, H. Chang<sup>1</sup>, Hassan Hussein Musa<sup>2</sup> and Mingxing Chu<sup>3</sup>

<sup>1</sup>Animal Science and Technology College, Yangzhou University, Yangzhou 25009, China. <sup>2</sup>Faculty of Veterinary Science, University of Nyala, 155 Nyala, Sudan. <sup>3</sup>Key Laboratory of Farm Animal, Genetic Resources and Utilization of Ministry of Agriculture, Institute of Animal Science, Chinese Academy of Agricultural Sciences, Beijing 100193, China.

Accepted 5 October, 2010

Four microsatellite loci (OarAE101, BM1329, BM143 and OarHH55) linked to FecB gene on chromosome 6 and one microsatellite locus (OarHH55) on chromosome 4 were selected to study their correlation with litter size of Hu sheep breed. The results showed that the average polymorphism information content (PIC), heterozygosity (He) and effective allele number (Ne) were 0.7214, 0.7558 and 4.4094, respectively, and all 5 microsatellite loci reached high polymorphism (PIC > 0.5), and the five microsatellite loci can be used for genetic diversity evaluation in Hu sheep breed. The result of variance analysis showed that there were extreme significant differences among the different genotypes for the producing ability of litter size in OarAE101 locus and BM143 locus (P < 0.01), and OarHH35 locus (0.01 < P < 0.05). However, there was no significant difference among the different genotypes for the producing ability of litter size in BM1329 and OarHH55 loci (P > 0.05). Choosing by combining producing ability and special gene (gene type) will play an great role in improving selecting accuracy and breeding progress, and it is a wonderful and new analyzed thought in repeatable characters with low heritability which is affected by measuring frequencies. And this study will have an important role in MAS (marker-assistant selection) and molecular breeding in Hu sheep in future.

Key words: Hu sheep, microsatellite markers, litter size trait, producing ability of ewes.

# INTRODUCTION

The profits of sheep production is dependent on litter size, and it is very difficult to improve litter size by normal breeding methods since its heritability ranges between 0.05 - 0.15 (Wang et al., 1994). There are many sheep breeds with fecundity traits in the world, but only the Australian Booroola merino sheep has been clearly known. The fecundity gene was located on 6th chromosome (Piper et al., 1982; Davis et al., 1982, 1983). In 1989, it was named Fec<sup>B</sup> (Fec = fecundity, B = Booroola) (Gognosag, 1989), it was precisely located in the area of 10cM between OarAE101 and BM1329 (Montgomery et al., 1993, 1994; Lord et al., 1998). Fec<sup>B</sup> has been used as a marker assisted selection to improve sheep fecundity (Gootwine et al., 1998). The high fecundity Hu sheep in China gives 2 lambs per birth, except those that are in the first lambing period, 6 - 8 lambs per birth was recorded. The average lambing rate of Hu sheep's was 261.09% (Editorial section of "Records of Sheep and Goat Breeds in China", 1989).

Currently, research on molecular mechanism of fecundity in Hu sheep is very few. Sheep's fecundity is affected by physiological condition, nutrition level and germ quality and other exterior factors. Phenotypic value of fecundity could not really be inherited. Producing ability on fecundity of ewes, obtained from calculating repeatability of litter size, is a good genetic parameter for choosing ewes, which could measure potential inheritability of fecundity. Therefore, it would be very meaningful to improve choosing accuracy and breeding progress by combining producing ability on fecundity of ewes and alleles (gene

<sup>\*</sup>Corresponding author: E- mail: dkxmsunwei@163.com.

Marker	Primer sequences	Annealing temperature (℃)	MgCl₂ (μl)
OarHH55	5'-3'GTTATTCCATATTCTTTCCTCCATCATAAGC	TCCTCCATCATAAGC 63	
	3'-5'CCACACAGAGCAACTAAAACCCAGC		
OarHHHH35	5'-3'AATTGCATTCAGTATCTTTAAACATCTGGC	63	2.4
	3'-5'ATGAAAATATAAAGAGAATGAACCACACGG		
BM143	5'-3'ACCTGGGAAGCCTCCATATC	63	2.4
	3'-5'CTGCAGGCAGATTCTTTATCG		
BM1329	5'-3'TTGTTTAGGCAAGTCCAAAGTC	60	1.2
	3'-5'AACACCGCAGCTTCATCC		
OarAE101	5'-3'TTCTTATAGATGCACTCAAGCTAGG	59	2.4
	3'-5'TAAGAAATATATTTGAAAAAACTGTATCTCCC		

Table 1. Primer sequences, annealing temperature and MgCl<sub>2</sub> amount.

type). In this paper, we focused on fecundity trait of Hu sheep and we selected microsatellites closely linked with FecB gene reference to Booroola sheep.

The aims of this study were to study the relationship between microsatellites and producing ability on fecundity trait of Hu sheep, and to provide scientific basis to further study the mechanism of fecundity and marker assisted selection.

#### MATERIALS AND METHODS

#### **Experimental animals**

The litter traits (lambs orn/ewe lambing) were collected from the records of 65 Hu sheep ewes from Suzhou sheep breeding farm in Jiangsu province, China. Blood samples were collected and DNA was isolated using deposit leukocyte phenol/chloroform extraction method (Sun et al., 2008).

#### Microsatellite analysis

Primers were selected based on the Boroola sheep data and synthesized by Sangon Biotech Company, Shanghai, China (Table 1). Four microsatellites (OarAE101, BM1329, BM143 and OarHH55) linked with fecundity gene on the 6th chromosome and one microsatellite (OarHH55) linked with production gene on the 4th chromosome were used. PCR were performed in 20 µl reaction volume containing 2  $\mu$ l of 50 ng genomic DNA, 2  $\mu$ l of 10×buffer, 1.2 - 2.4 μl 25 mmol/l MgCl<sub>2</sub>, 0.4 μl 10 mmol dNTP, 1 μl 8 μmol/μl upper and lower primers, 0.2 µl of 5 U Tag DNA polymerase and ddH<sub>2</sub>O to 20 µl as described by (Buchanan, 1992a, 1992b, 1993). The PCR protocol consisted of an initial denaturation at 94 °C for 5 min. 30 cycles of 1 min at 94°C, anneal for 1 min and extended 1 min at 72℃, followed by a final extension at 72℃ for 10 min. PCR products were electrophoreses on denaturing polyacrylamide gel, silver stained and visualized under ultraviolet light using transilluminator. The length of amplified fragment relevant to pBR322/MspI marker was calculated by Kodak Digital Science ID Image Analysis Software.

#### Statistical analysis

#### Genetic polymorphism analysis

Allele frequencies were computed by gene counting method.

Heterozygosity (He) (Nei, 1978), polymorphism information content (PIC) (Botstein et al., 1980) and effective allele number (Ne) (Kimura and Crow, 1974) were calculated using SAS package.

#### Assessment of producing ability of ewes

Repeatability:

$$r_e = \frac{MS_B - MS_W}{MS_B + (n-1)MS_W}$$

If there is difference in each population in observation number, weighted value  $n_0$  should be taken into consideration:

$$n_0 = \frac{1}{D-1} (N - \sum \frac{n_i^2}{N})$$

 $n_0$  = weighted mean of measurement frequency of individuals, N = sum of measurement frequencies, D = number of individuals, and  $n_j$  = measurement frequency of each individual.

#### Producing ability of ewes:

Producing ability of ewes can be calculated earlier when we have the evaluation of repeatability, and when litter records are different from individuals, the mean repeatability should be calculated with more records  $r_{e(n)}.$ 

$$r_{e (n)} = \frac{V_{G} + V_{Eg}}{V_{p(n)}} = \frac{V_{G} + V_{Eg}}{[1 + (n \Box 1)r_{e}] \cdot V_{p}/n}$$
$$= \frac{nr_{e}}{1 + (n \Box 1)r_{e}}$$

The producing ability on special trait of individual can be calculated by the following formula (Wang et al., 1994).

$$7P_X = (P_n - \bar{P})r_{e(n)} + \bar{p} = (p_n - \bar{p})\frac{nr_e}{1 + (n-1)r_e} + \bar{p}$$

Locus	Genotype (number of individuals)		Size of alleles (bp)	Allele frequency		
0arAE101	103/113 (4)	101/111 (3)	103	0.091 7		
	107/113 (8)	113/113 (10)	101	0.083 3		
	107/107 (9)	107/111 (2)	107	0.266 7		
	101/101 (2)	103/107 (3)	109	0.016 7		
	101/107 (1)	103/103 (2)	113	0.275 0		
	115/115 (4)	115/117 (1)	115	0.075 0		
	111/111 (7)	101/113 (1)	111	0.158 3		
	101/125 (1)	125/125 (1)	125	0.025 0		
	109/109 (1)		117	0.008 3		
<i>BM</i> 1329	146/146 (12)	146/162 (15)	146	0.416 7		
	164/164 (13)	158/158 (3)	164	0.233 3		
	146/158 (9)	162/162 (3)	158	0.133 3		
	148/148 (2)	148/158 (1)	148	0.041 7		
	146/164 (2)		162	0.175 0		
OarHH35	139/139 (7)	125/125 (5)	139	0.475 0		
	139/141 (4)	137/125 (3)	141	0.033 3		
	127/127 (2)		127	0.225 0		
	139/127 (23)		125	0.241 7		
	139/125 (16)		137	0.025 0		
<i>BM</i> 143	118/118 (3)	116/104 (1)	118	0.266 7		
	118/116 (8)	112/114 (2)	116	0.116 7		
	102/104 (10)	112/118 (1)	102	0.141 7		
	102/106 (2)	112/102 (3)	104	0.125 0		
	118/106 (12)	118/112 (1)	106	0.175 0		
	118/102 (1)	112/104 (1)	110	0.016 7		
	118/108 (1)	116/112 (4)	108	0.041 7		
	108/108 (1)	108/102 (1)	112	0.100 0		
	106/104 (3)	116/106 (1)	114	0.016 7		
	106/106 (1)	108/106 (1)				
OarHH55	131/131 (1)	147/127 (11)	131	0.058 3		
	131/129 (3)	147/135 (2)	129	0.266 7		
	129/129 (3)	147/125 (4)	135	0.033 3		
	135/131 (1)	127/125 (4)	147	0.325 0		
	147/147 (4)	127/129 (8)	127	0.216 7		
	147/129 (13)	127/127 (1)	125	0.100 0		
	147/131 (1)	125/125 (2)				
	135/129 (1)	129/127 (1)				

Table 2. Amplication of five microsatellite markers in Hu sheep.

 $\mathsf{P}_x$  = producing ability on special trait of individual x, P = mean value of traits in the populaton, and  $\mathsf{P}_n$  = mean value of n records on special trait.

# Relationship analysis between marked gene type and producing ability of Hu sheep in litter size

The difference between marked gene type and producing ability of Hu sheep litter size was compared with General Linear Model procedure using SAS package according to the following model:  $y_{ij}$  = Producing ability of Hu sheep in litter size;  $\mu$  =population mean value;  $M_i$  = Fixed effect of i marked gene type;  $e_{ij}$  = random error effect.

### RESULTS

#### Amplications analysis of microsatellitemarkers

Allele distribution and frequencies are list in Table 2. From the tables, 17 genotypes and 9 alleles that ranged

 $y_{ij} = \mu + M_i + e_{ij}$ 

Locus	Polymorphism information content (PIC)	Heterozygosity (He)	Ne
OarAE101	0.7803	0.8062	5.1611
BM1329	0.6788	0.7218	3.5945
OarHH35	0.6073	0.6636	2.9726
BM143	0.8169	0.8366	6.1210
OarHH55	0.7236	0.7618	4.1978
Mean Value	0.7214	0.758	4.4094

**Table 3.** Polymorphism information content (mean), mean heterozygosity (H) and mean effective number of alleles (Ne) of microsatellite DNA sites in Hu sheep.

between 101 to 125 bp were found in OarAE101. The dominant genotypes are 107/113, 107/107, 111/111 and 113/113, and the dominant alleles are 107, 113, 111. 9 genotypes and 5 alleles that ranged between 101 and 125 bp were found in BM1329. The dominant genotypes are 146/146, 164/164, 146/158 and 146/162, and the dominant alleles are 146, 164, 158 and 162. 7 genotypes and 5 alleles that ranged between 125 and 141 bp are found in OarHH35. The dominant genotypes are 139/139, 139/127 and 139/125, and the dominant alleles are 139, 127 and 125. Twenty genotypes and nine alleles that ranged between 102 and 118 bp are found in BM143. The dominant genotypes are 116/118, 102/104, 118/106, and the dominant alleles are 118, 116, 106, 104 and 102. 15 genotypes and 6 alleles that ranged between 125 to 147 bp are found in Oarhh55. The dominant genotypes are 147/129, 147/127 and 127/129, and the dominant alleles are 147, 129 and 127.

## PIC, Ne and He of microsatellite sites

Information of 5 microsatellite markers on PIC, Ne and He is list in Table 3. From Table 7, PIC of OarAE101, BM1329, OarHH35, BM143 and OarHH55 are 0.7803, 0.6788, 0.6073, 0.8169 and 0.7236, respectively, with an average of 0.7214. He of OarAE101, BM1329, OarHH35, BM143 and OarHH55 are 0.8062, 0.7218, 0.6636, 0.8366 and 0.7618, respectively, with 0.758 as mean value. Ne of OarAE101, BM1329, OarHH35, BM143 and OarHH55 are 5.1611, 3.5945, 2.9726, 6.1210 and 4.1978, respectively, with 4.4094 as mean value.

The result shows that polymorphism exists in 5 microsatellite markers, which can be used to evaluate genetic diversity of Hu sheep in reproduction.

# Association between microsatellite markers and producing ability of ewe in litter size

Result showed that there are extreme significant differences among the different genotypes for the producing ability of litter size in OarAE101 locus and BM143 locus (P < 0.01), there are significant differences among the different genotypes for the producing ability of litter size in

OarHH35 locus (0.01 < P < 0.05), however, no significant difference among the different genotypes for the producing ability of litter size in BM1329 and OarHH55 loci (P > 0.05).Those genotype whose indivi-dual number are less than 2 are not included in our analysis. Least square means ± standard error of producing ability of ewe in litter size in 5 microsatellite markers with multiple comparisons was listed in the Table 4.

Table 4 showed that least squares mean for the producing ability of litter size for OarAE101 107/113 was the largest, and it is not significantly higher than those for OarAE101 103/113, 107/107, 101/111, 113/113, 107/111 and 103/103 (P > 0.05), however it is significantly higher than those for OarAE101 101/101 and 103/107(0.01 < P < 0.05), and it is extremely significantly higher than those for OarAE101 115/115 and 111/111 (P < 0.01) in Hu sheep.

Table 5 shows that least square means of producing ability of ewe in 162/162 of BM1329 is the largest, but no significance with other markers. Table 6 showed that the least squares mean for the producing ability of litter size for OarHH35 139/139 was the highest, however it was not significantly higher than those for OarHH35 139/144, 139/127 and 137/125 (P > 0.05). It was significantly higher than those for OarHH35 127/127 and 125/125 (0.01 < P < 0.05), and extremely significantly higher than those for OarHH35 139/125 (P < 0.01) in Hu sheep.

Table 7 showed that least squares mean for the producing ability of litter size for BM143 118/118 was the highest, however it is not significantly higher than those for BM143, 118/106, 118/110, 106/104 and 112/114 (P > 0.05), it is significantly higher than those for BM143 118/ 116 and 102/106 (0.01 < P < 0.05), and it is extremely significantly higher than those for BM143 102/104, 112/102 and 116/112 (0.01 < P < 0.05) in Hu sheep. Table 8 shows that least square means of producing ability of ewe in 147/135 of OarHH55 are the largest, but no significance with other markers.

# DISCUSSION

# Microsatellite polymorphism

Ne, PIC and He are all used to evaluate population genetic

Treat	OarAE101 genotypes	Number of individuals	Least square means (LSMs)
1	103/113	4	2.572 5±0.036 2 <sup>abAB</sup>
2	107/113	8	2.595 0±0.025 6 <sup>aA</sup>
3	107/107	9	2.585 6±0.024 1 <sup>aAB</sup>
4	101/101	2	2.460 0±0.051 2 <sup>bcABC</sup>
5	115/115	4	2.475 0±0.036 2 <sup>bcBC</sup>
6	111/111	7	2.430 0±0.027 3 <sup>cC</sup>
7	101/111	3	2.516 7±0.041 8 <sup>abcABC</sup>
8	113/113	10	2.588 0±0.022 9 <sup>aAB</sup>
9	107/111	2	2.490 0±0.051 2 <sup>abcABC</sup>
10	103/107	3	2.473 3±0.041 8 <sup>bcABC</sup>
11	103/103	2	2.555 0±0.051 2 <sup>abcABC</sup>

Table 4. Least square means of OarAE101 genotypes for the producing ability of litter size of ewes in Hu sheep.

LSMs are expressed with their standard error values. Means with the different superscripts within the same column differ significantly (0.01 < P < 0.05); means with the different capital superscripts within the same column differ extremely significantly (P < 0.01).

Table 5. Least square means of BM1329 genotypes for the Producing ability of litter size of ewes in Hu sheep.

Treat	BM1329 genotypes	Number of individuals	Least square means (LSMs)
1	146/146	12	2.540 0±0.024 2 <sup>a</sup>
2	164/164	13	2.483 8±0.023 2 <sup>a</sup>
3	146/158	9	2.561 1±0.028 0 <sup>a</sup>
4	148/148	2	2.460 0±0.059 3 <sup>a</sup>
5	146/164	2	2.460 0±0.059 3 <sup>a</sup>
6	146/162	15	2.548 7±0.021 7 <sup>a</sup>
7	158/158	3	2.556 7±0.048 4 <sup>a</sup>
8	162/162	3	2.576 7±0.048 4 <sup>a</sup>

LSMs are expressed with their standard error values. Means with the same superscripts within the same column differ not significantly (P >0.05).

diversity. Ne of OarAE101, BM1329, OarHH35, BM143 and OarHH55 are 5.1611, 3.5945, 2.9726, 6.1210 and 4.1978 respectively, with average value of 4.4094. Montgomery et al. (1994) detected OarAE101, OarHH55 and BM143, and found 7, 11 and 11 alleles, respectively, on the markers. Chu et al. (2001) found 4 - 5 and 4 alleles, respectively on OarAE101 and BM1329. Such differences may result from the difference of populations and samplings.

PIC can be used to describe variation of microsatellite markers. Botstein et al. (1980) thought PIC was an index to measure genetic valuation. When PIC >0.5, the locus is of high polymorphism. When 0.25 < PIC < 0.5, the locus is of moderate polymorphism. When PIC <0.25, the locus is of low polymorphism. In this way, OarHH35, BM1329, OarHH55, OarAE101 and BM143 are all of high polymorphism. In genetic linkage analysis, microsatellite markers with PIC over 0.70 are the better markers (Barker et al., 1994). Under this condition, parents are usually heterozygous on the locus but alleles are separated in offsprings. PIC is high in this study, and average value is 0.7214, showing genetic abundance in fecundity

trait of Hu Sheep.

The gene diversity (He) reflects population variation in multiple gene loci. Generally speaking, He is considered as the best genetic parameters. In our study, of 5 microsatellite markers, He of OarHH35 is the highest, and He of BM1329 is the lowest. The average He of Hu sheep is 0.758. Arranz et al. (1998) detected the genetic variation of 5 Spanish sheep breeds and Awassi sheep, and their result shows that the average He of these sheep is 0.713 - 0.771. Our study is close to the result.

## Producing ability of ewe

Litter size is a character with very low heritability, about 0.1. Therefore, it is hard to use normal breeding methods to improve litter size, and the maker-assisted selection may accelerate the sheep breeding process. However, currently, few molecular researches on litter size of Hu sheep have been conducted, and only some lateral researches were done in the condition of Hu sheep as control population. Moreover, most researchers follow the

Treat	OarHH35 genotypes	Number of individuals	Least square means (LSMs)
1	139/139	7	2.592 9±0.030 6 <sup>aA</sup>
2	139/141	4	2.540 0±0.040 5 <sup>abAB</sup>
3	127/127	2	2.460 0±0.0573 0 <sup>bB</sup>
4	139/127	23	22.550 8±0.016 9 <sup>aAB</sup>
5	139/125	16	2.484 3±0.020 2 <sup>bB</sup>
6	125/125	5	2.481 7±0.033 1 <sup>bB</sup>
7	137/125	3	2.536 7±0.046 7 <sup>abAB</sup>

 Table 6. Least square means of OarHH35 genotypes for the producing ability of litter size of ewes in Hu sheep.

LSMs are expressed with their standard error values. Means with the different superscripts within the same column differ significantly (0.01 < P < 0.05); means with the different capital superscripts within the same column differ extreme significantly (P < 0.01).

**Table 7.** Least square means of BM143 genotypes for the producing ability of litter size of ewes in Hu sheep.

Treat	BM143 genotypes	Number of individuals	Least square means (LSMs)
1	118/118	3	2.640 0±0.040 7 <sup>aA</sup>
2	118/116	8	2.514 4±0.023 5 <sup>bcdABC</sup>
3	102/104	10	2.454 0±0.022 2 <sup>cdC</sup>
4	102/106	2	2.490 0±0.049 8 <sup>bcdABC</sup>
5	118/106	12	2.586 3±0.021 2 <sup>aAB</sup>
6	118/110	2	2.590 0±0.049 8 <sup>abABC</sup>
7	106/104	3	2.536 7±0.040 7 <sup>abcdABC</sup>
8	112/114	2	2.555 0±0.049 8 <sup>abcdABC</sup>
9	112/102	3	2.450 0±0.040 7 <sup>dC</sup>
10	116/112	4	2.475 0±0.035 2 <sup>bcdBC</sup>

Means with the different superscripts within the same column differ significantly (0.01 < P < 0.05; means with the different capital superscripts within the same column differ extremely and significantly (P < 0.01).

Table 8. Least square	means of OarHH55	genotypes for	or the	Producing	ability of	litter si	ize of	ewes	in
Hu sheep.									

Treat	OarHH55 genotypes	Number of individuals	Least square means (LSMs)
1	131/129	3	2.533 3±0.050 0 <sup>a</sup>
2	129/129	3	2.450 0±0.050 0 <sup>a</sup>
3	147/147	4	2.572 5±0.043 3 <sup>a</sup>
4	147/129	13	2.547 7±0.024 0 <sup>a</sup>
5	147/127	11	2.520 9±0.026 1 <sup>a</sup>
6	147/135	2	2.590 0±0.061 2 <sup>a</sup>
7	147/125	4	2.525 0±0.043 3 <sup>a</sup>
8	127/125	4	2.492 5±0.043 3 <sup>a</sup>
9	127/129	9	2.485 6±0.028 9 <sup>a</sup>
10	125/125	2	2.525 0±0.061 2 <sup>a</sup>

LSMs are expressed with their standard error values. Means with the same superscripts within the same column do not differ significantly (P > 0.05).

thought of doing selection or predicting production according to the correction between special marker alleles (gene type) and phenotypic value (Chu et al., 2002a; Wang et al., 2003; Lei et al., 2003; Ji et al., 2003). However, litter size of sheep is affected by many exterior factors as physiological condition, nutrition level and sperm quality of ram. Phenotypic value of fecundity do not pass down truly, thus it is not very efficient to use correction between marker and phenotypic value.

Producing ability is ewe's genetic potentiality on a repeatable character, which may be obtained by multiple measurement of litter size, and it is indeed an important reference to select excellent ewe. The data of producing ability of ewe can be used to evaluate ewe's potentiality on litter size. Thus, choosing by combining producing ability and special gene (gene type) will play a great role in improving selecting accuracy and breeding progress, and is a wonderful and new analyzed thought in repeatable characters with low heritability which is affected by measuring frequencies.

# Association between microsatellite markers and producing ability of ewe in litter size of Hu sheep

In this study, there are extreme significant difference among the different genotypes for the producing ability of litter size in OarAE101 locus and BM143 locus (P < 0.01). and there are significant difference among the different genotypes for the producing ability of litter size in OarHH35 locus (0.01 < P < 0.05), however, no significant difference among the different genotypes for the producing ability of litter size in BM1329 and OarHH55 loci (P > 0.05). In the locus of OarAE101, the least squares mean for the producing ability of litter size of 107/113 genotype was the highest, but it was not significantly higher than genotype of 103/113, 107/107, 101/111, 113/113, 107/111 and 103/103 (P > 0.05), and the producing ability of litter size of 111/111 genotype was the least. But in the research of Chu et al, the allele of 107 and genotype of 107/111 showed a positive significant relationship with the litter size trait of Small Han tail Han sheep, two alleles (109 and 109) showed a negative relationship with the litter size trait (Chu et al., 2001). In the locus of BM1329, there was no significant difference in producing ability of litter size among the different genotype, but in the further study, we could find that least square means of producing ability of ewe in 162/162, 158/158 and 146/158 were the highest (2.5567 - 2.5767), and the least square means of producing ability of ewe in 148/148 and 146/164 are lowest. But in the research of Chu et al. the allele of 146 and genotype of 146/158 showed a positive significant relationship with the litter size trait of Small Han tail Han sheep, the allele (148) showed a negative relationship with the litter size trait (Chu et al., 2001). In comparison with our study and the precious study (Chu et al., 2001) we could find, that there were some same or similar result, while there were also some different result. In our study, there were too many genotype and alleles, which maybe because of different sample scale or sample area or different polyacrylamide gel system.

Currently, there are a little over 100 million sheep in

China, ranking first in the world. However, number of slaughter is lower than world average. If we want to change the situation of low efficiency and low profit of sheep market in China, the most forceful method is to improve reproducing ability. This study builds a foundation for the maker-assistant selection in Hu sheep, and also provides new experiment thought in discussion of mechanism of reproductive ability of sheep.

# Conclusion

Five microsatellite markers: OarAE101, BM1329, BM143, OarHH55 and OarHH35 could be used to study genetics in Hu sheep diversity. The result of variance analysis showed that there were extreme significant difference among the different genotypes for the producing ability of litter size in OarAE101 locus and BM143 locus (P < 0.01), there were significant difference among the different genotypes for the producing ability of litter size in OarHH35 locus (0.01 < P < 0.05), and there were no significant difference among the different genotypes for the producing ability of litter size in BM1329 and OarHH55 loci (P > 0.05).

# ACKNOWLEDGEMENTS

This work was supported by National Modern Agriculture (Mutton Sheep and Goats) Industry Technology System Construction Program of China [MOA (2008) No.10]. State Scientific Basic Research platform Program of China (No. 2005DKA21101), International Cooperation Item of the National Natural Science Foundation of China (30410103150), China Postdoctoral Science Foundation special funded project (200902154), China Postdoctoral Science Foundation (No. 20080430470), Support Foundation of China during the 11th Five-Year Plan Period (No. 2006BAD13B08-07), Support Foundation of China during the 11th Five-Year Plan Period (No. 2008BADB2B04), Natural Science Foundation of Jiangsu Province of China (BK2007556), Basic Natural Science Foundation for Colleges and Universities Jiangsu Province (NK051039), Science and technology program of Suzhou City of Jiangsu Province of China (SNG0911), Northern Jiangsu technology development plan of China (BN2010004), Jiangsu Government Scholarship for Overseas Studies Project, Qing Lan Project of Colleges and Universities Jiangsu Province and the New Century Talent Project of Yangzhou University in China.

## REFERENCES

- Arranz JJ, Bayon Y, San Primitivo F (1998). Genetic relationships among Spanish sheep using microsatellites. Anim. Genet. 29: 435-440.
- Barker JSF (1994). A global protocol for determining genetic distances among domestic livestock breeds.Proc.5<sup>th</sup> World Congr. Genet. Appl.

Livest. Prod. 21: 501-508.

- Botstein D, White RL, Skolnick M (1980). Construction of a genetic linkage map in man using restriction fragment length polymorphisms. Am. J. Hum. Genet. 32: 314-331.
- Buchanan FC, Crawford AM (1992a). Ovine dinucleotide repeat polymorphism at the MAF70 locus [J]. Anim. Genet. 23: p. 185.
- Buchanan FC, Crawford AM (1992b). Ovine dinucleotide repeat polymorphism at the MAF33 locus [J]. Anim. Genet. 23: p. 186.
- Buchanan FC, Crawford AM (1993). Ovine microsatellites at the OarFCB11, OarFCB128, OarFCB193, OarFCB266 and OarFCB304 loci. Anim. Genet. 24: p. 145.
- Chu MX, Cheng JH, Guo W (2001). Preliminary Studies of Microsatellite Markers OarAE101 and BM1329 in Five Sheep Breeds, Acta Genet. Sinica. 28(6): 510-517.
- Chu MX, Ji CL, Chen GH (2003b). Association between PCR-RFLP of melatonin receptor 1a gene and high prolificacy in Small Tail Han sheep [J]. Asian-Australasian J. Anim. Sci. 16(12): 1701-1704.
- Chu MX, Wang JZ, Wang AG, Li N, Fu JL (2002a). Cloning and Sequencing of Four Microsatellite Loci in Small Tail Han Sheep, Acta Genet. Sinica, 29(5): 402-405.
- Chu MX, Wang JZ, Wang AG, Li N, Fu JL (2002b). Genetic Polymorphisms of Five Microsatellite Loci in Small Tail Han Sheep. Acta Genet. Sinica, 29(6): 502-506.
- Chu MX, Wang JZ, Wang ÁG, Li N, Fu JL (2003a). Association analysis between five microsatellite loci and litter size in Small Tail Han sheep. Asian-Australasian J. Anim. Sci. 16(11): 1555-1559.
- Davis GH, Kelly RW (1983). Segregation of major gene influencing ovulation. NZ Soc. Anim. Prod. 43: 197-199.
- Davis GH, Montgomery GW, Allison AJ, Kelly RW, Bray AR (1982). Segregation of a major gene influencing fecundity in progeny of Booroola sheep. N.Z. J. Agric. Res. 25: 525-529.
- Editorial section of Records of Sheep and Goat Breeds in China (1989). Records of Sheep and Goat Breeds in China. Shanghai Scientific and Technical Publish House. pp. 29-58.
- Gognosag (1989). Standardized genetic nomenclature for sheep and goat. Lavoisier, Paris, pp. 49-52.
- Gootwine E, Yossefi S, Zenou A, Bor A (1998). Markers assisted selection for Fec<sup>B</sup> carrier in Booroola Awassi crosses [A]. In: Proceeding of the 6th World Congress on Genetics Applied to Livestock Production [C]. Armidale, NSW, Australia, 24: 161-164.
- Ji CL, Chu MX, Chen GH, Zhou GL, Zhu Y (2003). PolymorphismAnalysis for Exon 2 of Melatonin Receptor 1a Gene in Four Sheep Breeds by PCR-RFLP, J. Huazhong Agric. 22(3): 105-109.

- Kimura M, Crow JF (1974). The number of alleles that can be maintained in a finite population. Genetics, 49: 725-738
- Lei XQ, Chen H, Xu TS, Yuan ZF, Guo MC, Song SD, Lei CZ, Sun WB (2003). Microsatellite Markers on Fecundity Trait of Small-Tailed Han Sheep, Chinese J. Anim. Vet. Sci. 34(6): 530-535.
- Lord EA, Davis GH, Dodds KG, Henry HM, Lumsden JM, Montgomery GW (1998). Identification of Booroola carriers using microsatellite markers [A]. In: Proceeding of the 6th World Congress on Genetics Applied to Livestock Production [C]. Armidale, NSW, Australia, 27: 19-22.
- Montgomery GW, Crawford AM, Penty JM, Dodds KG, Ede AJ, Henry HM, Pierson CA, Lord EA, Galloway SM, Schmack AE, Sise JA, Swarbrick PA, Hanrahan V, Buchanan FC, Hill DF (1993). The ovine Booroola fecundity gene (FecB) is linked to markers from a region of human chromosome 4g. Nat. Genet. 4: 410-414.
- Montgomery GW, Lord EA, Penty JM, Dodds KG, Broad TE, Cambridge L, Sunden SL, Stone RT, Crawford AM(1994). The Booroola fecundity (FecB) gene maps to sheep chromosome 6. J. Genomics, 22: 148-153.
- Nei M (1978). Estimation of average heterozygosity and genetic distance from a small number of individuals, Genetics, 89: 583-590
- Piper LR, Bindon BM (1982). The Booroola merino and the performance of medium non-Peppin crosses at Amidale [A]. The Booroola merino [M] (eds Piper LR, Bindon BM, Nethery RD, Fahmy MH, CSIPO, Melboure, pp. 9-10.
- Sun W, Chang H, Liao XJ, Tsunoda K (2008). Extract DNA from Sheep Whole Blood using Deposit leukocyte-phenol/chloroform extraction Method. Jiangsu Agric. Sci. 1: 40-41.
- Wang GL, Mao XZ, George HD, Zhao ZS, Zhang LJ, Zeng YQ (2003). DNA tests in Hu sheep and Han sheep (small tail) showed the existence of Booroola (FecB) mutation, J. Nanjing Agric. Univ. 26(1): 104-106.
- Wang JY, Chen GH, Chen KW (1994). The principle and method of animal breeding. Southeast University Publish House, pp. 114-155, 225-227.