

## Full Length Research Paper

# Assessment of the safety of aqueous extract of *Aloe vera* on haematology of Wistar rats

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*Aloe vera* is used both traditionally and packaged commercially in many regions of the world for several medicinal and or cosmetic purposes. It is claimed to have rejuvenating, moisturizing, healing or soothing properties on the skin and gastrointestinal tract. This study focused on assessment of the safety of *A. vera* on blood parameters: packed cell volume (PCV), red blood cell count (RBC), haemoglobin concentration, mean corpuscular volume, mean corpuscular haemoglobin, mean corpuscular haemoglobin concentration, white blood cell count (WBC), its differentials neutrophils, lymphocytes and platelet counts. Thirty Wistar rats were equally and randomly divided into 3 groups and *A. vera* extract solution was administered to 2 groups for 12 or 24 h respectively, for 7 days consecutively. The third group served as control for the experiment. Blood samples were collected on day 8 to determine changes in the haemogram as a basis for toxicity. Rats administered with *A. vera* extract, particularly for 24 h showed increased levels of PCV ( $47.42 \pm 4.32\%$ ), RBC ( $9.26 \pm 0.60 \times 10^6/\mu\text{L}$ ), WBC ( $12.61 \pm 0.45 \times 10^3/\mu\text{L}$ ) and its differentials. Platelet count was also significantly increased ( $150.25 \pm 4.77 \times 10^9/\text{L}$ ). The results from this study showed that *A. vera* stimulated increased production of all blood cell types. In conclusion, protracted consumption of the extract of *A. vera* cause stimulation of haematopoiesis which may induce or encourage the progression of haemoproliferative disorders.

**Key words:** *Aloe vera*, haematology, Wistar rat.

## INTRODUCTION

*Aloe vera* is a naturally occurring plant with succulent leaves, originating from Northern Africa (Akinyele and Odiyi, 2007). The whole leaves or juice from the leaves has been used in several cultures of the world dating back to the first century A.D. as herbal remedy for various skin conditions (Boudreau and Beland, 2006; Akinyele and Odiyi, 2007). It is being packaged and marketed alone or in combination with other substances in

commercially available lotions, creams, yogurt, beverages and as desert. It is claimed to have rejuvenating, moisturizing, healing or soothing properties on the skin and gastrointestinal tract (Davies et al., 1989; Hegggers et al., 1997; Vogler and Ernst, 1999; Boudreau and Beland, 2006). Preliminary reports have also been documented on its blood glucose and lipid lowering effects, suggesting possibility of its use as an anti-diabetic agent (Nassiff et

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al., 1993; Boudreau and Beland, 2006; Choudhary et al., 2014; Alinejad-Mofrad et al., 2015). Reduction of symptoms and inflammation in patients with ulcerative colitis has also been suggested amongst other medicinal uses (Langmead et al., 2004; Bottenberg et al., 2007).

*Aloe vera* is a stem-less or very short-stemmed succulent plant growing to 60–100 cm (24–39 in) in height and spreading by offsets. It has fleshy, thick leaves which are usually green to grey-green in colour, with some varieties showing white flecks on the upper and lower stem surfaces (Gao and Xiao, 1997; Wang et al., 2004). The margin of the leaf is serrated and has small white teeth. It has pendulous flowers which are produced in summer season and these may reach up to 90 cm (35 in) tall. The flowers have a yellow tubular corolla, 2–3 cm (0.8–1.2 in) in length. Like other *Aloe* species, *A. vera* forms arbuscular mycorrhiza, a symbiosis that allows the plant better access to mineral nutrients in soil (Gong et al., 2002).

Most evidences of the activities of *A. vera* cannot be substantiated as little scientific evidence exist on its effectiveness or safety for the medicinal or cosmetic purpose for which it is used (Cosmetic Ingredient Review Panel, 2007). The few scientific reports however, showed conflicting evidences (Vogler and Ernst, 1999; Ernst, 2000; Marshall, 2000; Boudreau and Beland, 2006). Some conflicting reports on its wound healing ability were documented by Heggars et al., (1997) and Davis et al. (1989) who reported that *A. vera* promoted wound healing, while Schmidt and Greenspoon (1991) and Kaufman et al. (1988) reported the contrary.

There are little or no reports on the effect of the plant on the blood, the vehicle of transportation of most substances. Some information of its effect on blood cells may give some insight on safety of the plant on blood cells and related organ tissues. This study was therefore designed to determine the effect of sub-chronic administration of *A. vera* on the haemogram using assessment of changes in the packed cell volume, various red and white blood cell indices and platelet counts.

## MATERIALS AND METHODS

### Preparation of *Aloe vera* juice

Fresh leaves of *A. vera* were plucked daily and washed. The juice was expressed by gentle milking downwards. Daily water intake of the rats was determined during the acclimatization period to be approximately 40 ml per rat per day. The fresh juice was reconstituted to 46.20 mg/ml in fresh drinking water and served to rats unprocessed. This study was carried out in September, 2012 in South West, Nigeria.

### Experimental animals

Thirty male Wistar rats (140 – 160g) were obtained from the Experimental Animal Unit of the Faculty of Veterinary Medicine. The animals were housed in 12 h light: dark condition and maintained on standard rat diet. Clean water was provided *ad libitum*. The

animals were stabilized for 4 weeks before commencement of the experiment. All the rats were humanely managed and the study protocols were in compliance with the Faculty of Veterinary Medicine guidelines for the use of laboratory animals.

### Experimental protocol

Thirty rats were randomly and equally divided into 3 groups of one control and two treatment groups. The rats in the control group were allowed free access to clean water throughout the course of the experiment. Clean water was withdrawn from rats in treatment groups 1 and 2, and replaced with the *A. vera* solution for 12 and 24 h respectively, for 7 days consecutively. For the 12 h exposure group, half of the daily water requirement was reconstituted with *A. vera* and offered for 12 h, while fresh clean drinking water was offered for the remaining 12 h.

### Sample collection

On day 8, the rats were anaesthetized using anaesthetic ether and blood samples were collected from each rat via the retro-orbital sinus. About 3 ml of blood was collected into Lithium heparinized bottles for haematological analysis by Cole's method (Cole, 1986).

### Statistical analysis

All values are expressed as mean  $\pm$  S.E.M. Data obtained were analyzed using one-way analysis of variance (ANOVA), followed by Tukey post-test. Differences between means were considered significantly different when values  $p < 0.05$  were obtained using Graph-Pad Prism software Version 5 (2007).

## RESULTS

### Packed cell volume (PCV)

An increase in the PCV of rats administered with the extract of *A. vera* was observed with a significant ( $p < 0.05$ ) increase in rats administered with the extract for 24 h ( $47.42 \pm 4.32\%$ ) when compared to the control rats ( $41.75 \pm 2.17\%$ ) (Table 1).

### Red blood cell indices

Red blood cell count (RBC) of rats administered with the extract increased from  $8.22 \pm 1.57 \times 10^6/\mu\text{L}$  observed in the control rats to  $8.97 \pm 0.16$  and  $9.26 \pm 0.60 \times 10^6/\mu\text{L}$  observed in rats administered with the extract for 12 and 24 h respectively. Increases were also observed in the haemoglobin concentration of the treated rats with a significant ( $p < 0.05$ ) increase in rats treated for 24 h ( $16.31 \pm 0.68$  g/dl) compared to the control rats ( $13.88 \pm 0.89$  g/dl). Other red cell indices also increased accordingly (Table 1).

### White blood cell indices

White blood cell count (WBC) and the differential cell

**Table 1.** Packed cell volume and red blood cell indices obtained from rats administered with *Aloe vera* for 12 or 24 h of seven consecutive days.

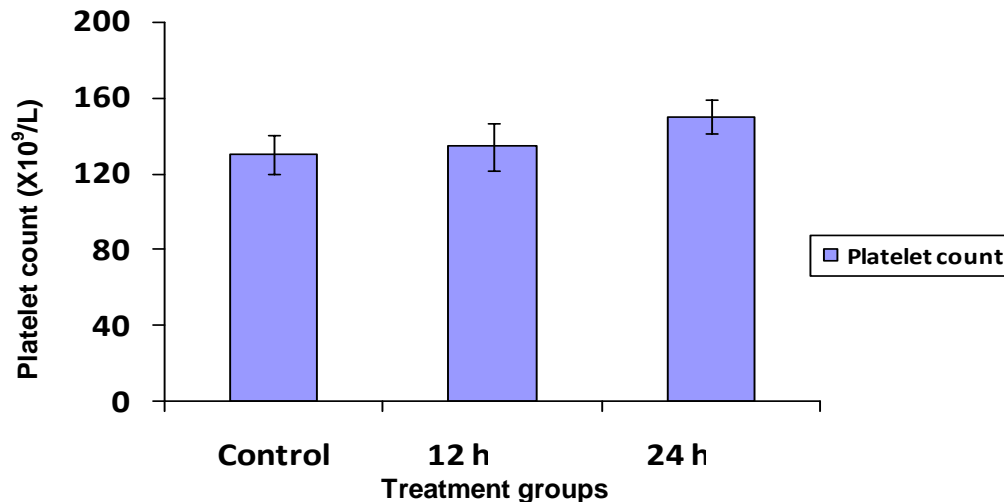
Haematological parameters	Control	12 h	24 h
PCV (%)	41.75±2.17	43.30±1.43	47.42±4.32*
RBC (X10 <sup>6</sup> /μL)	8.22±1.57	8.97±0.16	9.26±0.60
Hb (g/dl)	13.88±0.89	14.25±0.41	16.31±0.68*
MCV (fl)	50.79±4.05	48.27±2.00*	51.21±1.21
MCH (pg)	16.89±0.65	16.38±0.34	17.61±0.29
MCHC (g/dl)	33.25±0.41	30.39±0.26	34.39±0.22

\*Significant (p<0.05) difference compared to control value.

**Table 2.** White blood cell indices and platelet count of rats administered with *Aloe vera* for 12 or 24 h of seven consecutive days.

Haematological parameters	Control	12 h	24 h
WBC (X10 <sup>3</sup> /μL)	8.70±0.27	9.48±0.67*	12.61±0.45*
Lymphocytes (X10 <sup>3</sup> /μL)	4.77±0.49	5.53±0.31	8.35±0.12*
Neutrophils (X10 <sup>3</sup> /μL)	2.23±0.25	2.93±0.14	4.34±0.13*
Eosinophils (X10 <sup>3</sup> /μL)	0.13±0.01	0.19±0.05	0.28±0.05*
Neutrophil/ Lymphocyte ratio	0.47±0.01	0.53±0.05	0.52±0.02

\*Significant (p<0.05) difference compared to control value.

**Figure 1.** Mean platelet count of rats administered with aqueous extract of *Aloe vera* for a 12 or 24 h period of 7 consecutive days.

count of rats administered with *A. vera* extract increased compared to those of the control rats. Notably, WBC in rats treated for the 24 h period ( $12.61 \pm 0.45 \times 10^3/\mu\text{L}$ ) was significantly ( $p < 0.05$ ) higher than that of control rats ( $8.70 \pm 0.27 \times 10^3/\mu\text{L}$ ). The same significant ( $p < 0.05$ ) pattern was observed for the differential cell count of these rats treated for 24 h (Table 2).

### Platelet count

Platelet counts were non-significantly ( $p > 0.05$ ) increased in the rats treated for 12 h ( $134.2 \pm 1.24 \times 10^9/\text{L}$ ) but significantly ( $p < 0.05$ ) increased in rats treated for 24 hours ( $150.25 \pm 4.77 \times 10^9/\text{L}$ ) compared to the control rats ( $130.01 \pm 2.31 \times 10^9/\text{L}$ ) (Figure 1).

## DISCUSSION

In this study, rats administered with the aqueous extract of *A. vera* had increased values of the packed cell volume (PCV), red blood cell counts and other red cell indices. A significant ( $p < 0.05$ ) increase in PCV was observed in rats administered with the extract for the period of 24 h. This increase in PCV was not due to haemoconcentration because there was a generalized increase in red and white blood cells, but can be attributed to stimulation of haematopoiesis. This can further be related to the result of the red cell indices; increased MCV, MCH and MCHC, which showed that immature red cells were present in circulation, indicative of stimulation of production of immature erythrocytes, also known as reticulocytes. Morphologically, reticulocytes are characterised by increases in the size of red cells in circulation and it is usually observed as the initial response to stimulation of the haematopoietic system during active blood regeneration (Saba et al., 2009). *Telfaria occidentalis* leaves which are consumed in soups in several regions of West Africa had also been reported to have haematopoietic stimulatory ability and it is used traditional for treatment of anaemia (Alada, 2000; Dina et al., 2000).

White blood cells on the other hand showed significant ( $p < 0.05$ ) increases, particularly in rats administered with the extract for 24 h. About 2-fold increment in lymphocyte and neutrophil counts were observed in these rats. Lymphocytosis may be associated with increased immunological response to an antigenic stimulation, while the neutrophilia may be traced to increased inflammatory response in the body (Guyton and Hall, 2006a, b). Increased circulating neutrophils are usually as a result of mobilization of neutrophils into circulation in response to an antigenic stimulation. Such stimulants include trauma and bacteria endotoxins (Zekonis and Zekonis, 2004; Tang et al., 2010), of which the extract is neither. From the result obtained for this study, it can be postulated that *A. vera* may contain bioactive substances which are capable of mobilizing all blood cell types into circulation, and or stimulate haematopoiesis resulting in increased production/ release of blood cells into circulation. Our argument favours the haematopoietic theory more, considering the fact that neutrophil: lymphocyte ratios, a marker of subclinical inflammation, were approximately 0.5 in control and test groups, which were clinically and statistically non-significantly different (Sen et al., 2013; Wang, 2014).

Platelet counts were non-significantly ( $p > 0.05$ ) increased in the rats administered with the extract for 12 h, but a significant ( $p < 0.05$ ) increase was observed in rats administered the extract for the 24 h period. Thus, it can be inferred that blood clotting mechanisms may not be affected by *A. vera* extract, but this corroborates our theory in favour of indiscriminate stimulation of blood cell production.

Administration of the extract for the 12 h period showed

minimal haemopoietic ability compared to rats administered with the extract for the 24 h period. A cumulative dose-dependent pattern was established from this study which discourages the continuous consumption of the extract as it is administered for certain traditional uses. The indiscriminate stimulation of blood cells may be detrimental to the body with depletion of haematopoietic stem cells in bone marrow and may eventually trigger or encourage uncontrolled stimulation of haematopoiesis which can be seen in cases of myeloproliferative disorders (Tefferi and Vainchenker, 2011; Barbui et al., 2013).

## Conflict of Interest

The authors disclose that they do not have any conflict of interest.

## REFERENCES

- Akinyele BO, Odiyi AC (2007). Comparative study of the vegetative morphology and the existing taxonomic status of *Aloe vera* L. *J. Plant Sci.* 2(5):558-563.
- Alada ARA (2000). The haematological effect of *Telfaria occidentalis* diet preparation. *Afr J. Biomed. Res.* 3(1):186.
- Alinejad-Mofrad S, Foadoddini M, Saadatjoo SA, Shayesteh M (2015). Improvement of glucose and lipid profile status with *Aloe vera* in pre-diabetic subjects: a randomized controlled-trial. *J Diabetes Metab Disord.* 14:22
- Barbui T, Finazzi G, Falanga A (2013). Myeloproliferative neoplasms and thrombosis. *Blood* 122 (13):2176-2184.
- Bottenberg MM, Wall GC, Harvey RL, Habib S (2007). Oral aloe vera-induced hepatitis. *Ann. Pharmacother.* 41 (10):1740-1743.
- Boudreau MD, Beland FA (2006). An evaluation of the biological and toxicological properties of *Aloe barbadensis* (Miller), *Aloe vera*. *J. Environ. Sci. Health Part C* 24:103-154.
- Choudhary M, Kochhar A, Sangha J (2014). Hypoglycemic and hypolipidemic effect of *Aloe vera* L. in non-insulin dependent diabetics. *J Food Sci. Technol.* 51(1):90-96.
- Cole EH (1986). *Veterinary clinical pathology*, 4th ed W.B Saunders Publishers.
- Cosmetic Ingredient Review Expert Panel (2007): Final report on the safety assessment of *Aloe andongensis* extract, *Aloe andongensis* leaf juice, *Aloe arborescens* leaf extract, *Aloe arborescens* leaf juice, *Aloe arborescens* leaf protoplasts, *Aloe barbadensis* flower extract, *Aloe barbadensis* leaf, *Aloe barbadensis* leaf extract, *Aloe barbadensis* leaf juice, *Aloe barbadensis* leaf polysaccharides, *Aloe barbadensis* leaf water, *Aloe ferox* leaf extract, *Aloe ferox* leaf juice, and *Aloe ferox* leaf juice extract. *Int. J. Toxicol.* 26 (Suppl 2), 1–50.
- Davis RH, Leitner MG, Russo JM, Byrne ME (1989). Wound healing. Oral and topical activity of *Aloe vera*. *J. Am. Podiatr. Med. Assoc.* 79 (11):559–62.
- Dina OA, Adedapo AA, Oyinloye OP, Saba AB (2000). Effect of *Telfaria occidentalis* extract on experimentally induced anaemia in domestic rabbits. *Afr. J. Biomed. Res.* 3 (3):181-183.
- Ernst E (2000). Adverse effects of herbal drugs in dermatology. *Br. J. Dermatol.* 143:923–929
- Gao W, Xiao P (1997): Peroxidase and soluble protein in the leaves of *Aloe vera* L. var. *Chinensis* (Haw.) Berger. *China J. Chin. Mat. Med.* 22 (11):653–654.
- Gong M, Wang F, Chen Y (2002): Study on application of arbuscular-mycorrhizas in growing seedlings of *Aloe vera*. *J. Chin. Med. Mat.* 25 (1):1–3.
- Guyton AC, Hall JE (2006a). Resistance of the body to infection: II. Immunity and allergy. In: *Textbook of Medical Physiology*, 11th edn Guyton AC and Hall JE (Editors). Saunders Publishers, Philadelphia. . p. 440.

- Guyton AC, Hall JE (2006b): Resistance of the body to infection: I. Leukocytes, granulocytes, the monocyte-macrophage system, and inflammation. *In: Textbook of Medical Physiology*, 11<sup>th</sup> edn. Guyton AC and Hall JE (Editors). Saunders Publishers, Philadelphia. pp. 431-434.
- Heggors JP, Elzaim H, Garfield R, Goodheart R, Listen-Garten D, Zhao J, Phillips LG (1997). Effect of the combination of *Aloe vera*, nitroglycerin, and L-NAME on wound healing in the rat excisional model. *J. Altern. Complement. Med.* 3 (2):149-53.
- Kaufman T, Kalderon N, Ullmann Y, Berger J (1988). *Aloe vera* gel hindered wound healing of experimental second-degree burns: a quantitative controlled study. *J. Burn Care Rehab.* 9 (2):156-159.
- Langmead L, Feakins RM, Goldthorpe S, Holt H, Tsironi E, De Silva A, Jewell DP, Rampton DS (2004). Randomized, double-blind, placebo-controlled trial of oral aloe vera gel for active ulcerative colitis. *Alim. Pharmacol. Therap.* 19 (7):739-747.
- Marshall JM (2000). *Aloe vera* gel: what is the evidence? *Pharm. J.* 244: 360-362.
- Nassiff HA, Fajardo F, Velez F (1993). Efecto del aloe sobre la hiperlipidemia en pacientes refractarios a la dieta. *Rev. Cuba Med. Gen. Integr.* 9:43-51
- Saba AB, Oridupa OA, Ofuegbe SO (2009). Evaluation of haematological and serum electrolyte changes in wistar rats administered with ethanolic extract of whole fruit of *Lagenaria breviflora* robert. *J. Med. Plants Res.* 3(10):758-762.
- Schmidt JM, Greenspoon JS (1991). *Aloe vera* dermal wound gel is associated with a delay in wound healing. *Obstet. Gynecol.* 78 (1):115-117.
- Sen BB, Rifaioğlu EN, Ekiz O, Inan MU, Sen T, Sen N (2013). Neutrophil to lymphocyte ratio as a measure of systemic inflammation in psoriasis. *Cutan. Ocul. Toxicol.* 33(3):223-237.
- Tang M, Zhao XG, Gu YJ, Chen CZ (2010). An in vitro model for studying neutrophil activation during cardiopulmonary bypass by using a polymerase chain reaction thermocycler. *Altern. Lab. Anim.* 38:213-219.
- Tefferi A, Vainchenker W (2011). Myeloproliferative neoplasms: molecular pathophysiology, essential clinical understanding, and treatment strategies. *J. Clin. Oncol.* 29 (5):573-582.
- Vogler BK, Ernst E (1999). *Aloe vera*: a systematic review of its clinical effectiveness. *Br. J. Gen. Pract.* 49(447):823-828.
- Wang H, Li F, Wang T, Li J, Li J, Yang X, Li J (2004). Determination of aloin content in callus of *Aloe vera* var. *Chinensis*. *J. Chin. Med. Mat.* 27 (9):627– 628.
- Wang X (2014). Neutrophil to lymphocyte ratio in relation to risk of all-cause mortality and cardiovascular events among patients undergoing angiography or cardiac revascularization: A meta-analysis of observational studies. *Atherosclerosis* 234 (1), 206-213
- Zekonis G, Zekonis J (2004): Effect of bacterial stimulants on release of reactive oxygen metabolites from peripheral blood neutrophils in periodontitis. *Medicina (Kaunas)* 40(3):260-264.