

African Journal of Biotechnology Vol. 10(38), pp. 7541-7547, 25 July, 2011
Available online at <http://www.academicjournals.org/AJB>
DOI: 10.5897/AJB10.2260
ISSN 1684-5315 © 2011 Academic Journals

Full Length Research Paper

Effects of dietary supplementation of multi-enzyme complex on the energy utilization in rooster and performance of broiler chicks

Fatemeh Shirmohammad and Morteza Mehri*

Islamic Azad University, Shahr-e-Qods Branch, Department of Animal Science, Tehran, Iran.

Accepted 26 April, 2011

Two experiments were conducted to determine the effects of dietary supplementation of REAP[®] enzyme into corn-soybean diet on the energy utilization in poultry and performance of broiler chicks. In the first experiment, a total of 16 50 weeks adult roosters (ISA-Brown) were divided into 4 groups with 4 birds per replicate and the experimental diets contained the two levels of energy (2650 and 2759 kcal TME/kg diet) with 0 or 0.1% REAP[®] and were subjected to assay of apparent metabolizable energy (AME). In the second experiment, 360 3 days old male broiler chicks (Ross) were divided into 4 groups with 3 replicates of 30 birds per replicate and were assigned at random to one of the four experimental diets containing the two levels of energy (3100 and 2980 kcal TME/kg diet) with 0 or 0.1% REAP[®]. AME value of the high energy groups were significantly higher ($p < 0.05$) than that of the low energy groups when measured at 28 days. The body weight gain of the birds fed the low energy diet with 0% REAP was lower significantly than those of the other groups ($p < 0.05$). There were no significant differences in feed intake and feed conversion rate among the treatments. The breast muscle weights of the low energy diet birds were higher than those of the high ones and those of the lower energy group with 0.1% REAP were the highest ($p < 0.05$). The relative abdominal fat weight was reduced by the dietary REAP ($p < 0.05$). Percentage duodenum weights of high energy group were higher than those of the low energy group. The intestinal lengths (cm/100 g BW) of low energy diet group without REAP were lower than those of the others ($P < 0.05$). The results demonstrated that, dietary REAP improved body weight gain and reduced abdominal fats. Therefore, it can be concluded that, dietary supplementation of REAP improves nutritive value of corn-soybean diet in the broiler chicks.

Key words: REAP[®], broiler chicks, corn-soybean, apparent metabolizable energy (AME).

INTRODUCTION

Corn-soybean meal poultry feed is considered to be favorable because of its high nutritional value but soybean meal contains oligosaccharides that have been shown to decrease bird health and growth (Iji and Tivey, 1998) and in corn cell wall, arabinoxylans, β -glucan and cellulose are present. There is strong evidence that, some nutrients in corn are not completely digested in the small intestine and that considerable amounts of starch and protein escape digestion, reach the midgut and undergo fermentation with a relatively low energy yield

(Noy and Sklan, 1995). Among potential factors reducing nutrient bioavailability are the non-starch polysaccharides (NSP). In most of the studies, with adding an enzyme or enzyme mixture to corn-soybean diets, a significant increase in NSP digestibility was observed, indicating that the enzymatic degradation of cell wall polysaccharides is possible despite the complex nature of these polymers. (Meng et al., 2005). Enzymes have been added to broiler diets for more than 30 years. Supplementing corn-soybean meal diets with an enzyme or enzyme mixture that possess a broad-spectrum range of activities may improve the digestibility and as a result, growth performance (Odetallah et al., 2002). Several studies have demonstrated some beneficial effect on the AME and NSP digestibility of soybean diets, depending on the

*Corresponding author. E-mail: mortezamehri@gmail.com
Tel/Fax: +982612706168 or +985112740420.

Table 1. Ingredients of rations in different treatments (%).

Ingredient	High energy		Low energy	
	-	+	-	+
Corn	56.2	56.2	60.1	60.1
Soybean meal	34.5	34.5	33.2	33.2
Fish meal	3	3	2.8	2.8
Soybean oil	3.5	3.5	1.2	1.2
Dicalcium phosphate	1.3	1.3	1.2	1.2
Common salt	0.4	0.4	0.37	0.37
Mineral premix	0.5	0.5	0.5	0.5
Vitamin premix	0.5	0.5	0.5	0.5
DL-methionine	0.11	0.11	0.12	0.12
TME _n (kcal)	3100	3100	2980	2980

(-) Indicates non-addition of REAP and (+) indicates the addition of 0.1% of REAP.

enzyme preparation used (Zhou et al., 2009; Kocher et al., 2002; Fuente et al., 1995). Hesselman and Aman (1986) proposed that, β -glucanase breaks down cell wall and releases nutrients from cellular contents of digestion. Recently, *in vitro* studies in a laboratory (Saleh et al., 2003a, b) showed that, digestibility was higher when protease was excluded from the mixture of enzymes. Zhou et al. (2009) reported that, supplementing a corn-soybean broiler starter diet with an enzyme preparation containing a mixture of xylanase, protease and amylase resulted in improvements in AME value of diets in the starter, grower and finisher phases. REAP[®] is a multi-enzyme with β -glucanase, protease and cellulase activity. Therefore, this study was conducted to investigate whether REAP[®] could affect the AME and improve the performance of poultry.

MATERIALS AND METHODS

Two experiments were conducted to determine the effects of dietary supplementation of REAP[®] enzyme in corn-soybean diet on the energy utilization in poultry and performance of broiler chicks. The enzyme preparation used in this study was a commercial multi-enzyme complex with β -glucanase, protease and cellulase. In the first experiment, a total of 1650 weeks adult roosters (ISA-Brown) of uniform body weight (BW) (2.4 to 2.8 kg) were obtained from a commercial farm and divided into 4 groups with 4 birds per replicate and subjected to assay of apparent metabolizable energy (AME) (Farrell, 1978). They were housed in cages; 1 per cage that permitted collection of excreta on plastic. Experimental period was 28 days. The birds were fasted for 24 h before the introduction of assay diets. Four treatment groups which included high energy groups (with 2759 kcal/kg TME_n and 0% or 1% REAP[®]) and low energy groups (with 2650 kcal/kg TME_n and 0% or 1% REAP[®]) were used.

All the samples of excreta were collected separately for each rooster. These were weighed, oven dried at 100°C and ground to pass through 0.3 mm mesh sieve. The gross energy (GE) content of each feedstuff and its excreta was determined with the help of Parr adiabatic oxygen bomb calorimeter (Nukamp, 1965). Samples of feedstuffs tested for TME contents were subjected to proximate

analysis (AOAC, 1990). The AME and TME_n per kg of feed were calculated by the following formula:

$$\text{AME (Kcal/kg)} = \frac{\text{GE intake} - \text{GE excreta}}{\text{Intake}} \times 1000 \quad (1)$$

$$\text{TME}_n \text{ (Kcal/kg)} = \frac{\text{GE intake} - (\text{GE excreta} - \text{GE endogenous}) - 8.22 \times \text{nitrogen balance (g)}}{\text{Intake}} \times 1000 \quad (2)$$

In the second experiment, 360 3 days old male broiler chicks (Ross) were divided into 4 groups with 3 replicates of 30 birds per replicate and were assigned at random to one of the four experimental diets containing the two levels of energy (3100 and 2980 kcal TME_n/kg diet) with 0 or 0.1% REAP[®] (Table 1). The birds were housed in an environmentally controlled room and feed and water were provided *ad libitum* from 3 to 23 days of age. Feed consumption, body weight gain and feed conversion rate were measured weekly. Body composition, the length and weight of the intestine (duodenum, jejunum, and ileum) were determined at 23 days of age.

Statistical analysis of the data as a completely randomized design was accomplished using the General Linear Models procedure of SAS (SAS Institute, 1990). Differences between means were determined using the least significant difference mean separation procedure.

RESULTS

The enzyme activity of α -galactosidase in REAP[®] was 13,708 unit/g. Although there were no significant differences, the AME value of the dietary supplementation of REAP[®] tended to be increased at the 14th day. The AME value of the high energy groups were significantly higher ($p < 0.05$) than the low energy groups when measured at the 28th day (Table 2). However, the dietary supplementation of REAP[®] did not affect AME values of the diets for the experimental period.

The body weight gain of birds fed the low energy diet with 0% REAP was lower significantly than the other groups ($p < 0.05$) (Table 3). There were no significant

Table 2. Effects of REAP[®] supplementation on metabolizable energy in roosters.

	High energy		Low energy	
	- ¹	+ ¹	-	+
14 days	kcal/kg			
Gross energy	3643	3643	3513	3513
Fecal energy	2722	2592	2693	2333
AME	2922±38	2958±82	2792±20	2892±21
28 days				
Gross energy	3643	3643	3513	3513
Fecal energy	2349	2517	2674	2433
AME	2992±3 ^a	2982±4 ^a	2809±11 ^b	2869±12 ^b

^{a,b}Means ±SE within a row with no common superscripts differ significantly (P < 0.05). ¹ A (-) indicates non-addition of REAP, and a (+) indicates the addition of REAP.

Table 3. Effect of REAP supplementation on daily weight gain in broiler chicks.

	High energy		Low energy	
	- ¹	+ ¹	-	+
	g/day/bird			
1 week	20.8±0.5	21.0±0.7	20.4±0.3	20.9±0.1
2 weeks	43.3±1.0 ^{ab}	45.7±0.4 ^a	41.04±0.0 ^b	43.2±0.7 ^{ab}
3 weeks	59.1±0.4 ^a	58.1±0.8 ^{ab}	55.7±0.6 ^b	58.0±0.4 ^{ab}
Mean	41.1±0.3 ^a	41.5±0.4 ^a	39.1±0.4 ^b	40.6±0.1 ^a

^{a,b}Means ± SE within a row with no common superscripts differ significantly (P < 0.05). ¹ A (-) indicates non-addition of REAP and a (+) indicates the addition of REAP.

Table 4. Effect of REAP[®] supplementation on feed intake in broiler chicks.

	High energy		Low energy	
	- ¹	+ ¹	-	+
	g/day/bird			
1 week	31.6±0.8	31.1±0.2	31.7±0.6	32.3±0.1
2 weeks	70.5±0.5	69.9±0.7	71.6±1.1	71.1±1.3
3 weeks	95.6±0.3 ^a	91.8±0.7 ^b	91.9±0.1 ^b	93.2±0.8 ^{ab}
Mean	65.9±0.5	64.7±0.7	65.06±0.4	65.4±0.2

^{a,b}Means ± SE within a row with no common superscripts differ significantly (P < 0.05). ¹ A (-) indicates non-addition of REAP[®] and a (+) indicates the addition of REAP[®].

differences in the feed intake (Table 4) and feed conversion rate (Table 5) among the treatments.

The relative sizes (percent body weight) of the liver and leg muscle were not different significantly among the treatments. The breast muscle weights of the low energy

diet birds were higher than those of the high ones and those of the lower energy group with 0.1% REAP were the highest. The relative abdominal fat weight was reduced by the dietary REAP (Table 6).

No significant differences in the relative jejunum and

Table 5. Effect of REAP supplementation on feed conversion rate in broiler chicks.

Week	High energy		Low energy	
	-	+	-	+
	Feed/gain			
1 week	1.52±0.01	1.49±0.04	1.55±0.05	1.54±0.01
2 weeks	1.63±0.05 ^{ab}	1.53±0.00 ^b	1.74±0.03 ^a	1.65±0.06 ^{ab}
3 weeks	1.62±0.02	1.58±0.03	1.65±0.02	1.61±0.02
Mean	1.62±0.02	1.58±0.03	1.65±0.02	1.61±0.02

^{a,b}Means ± SE within a row with no common superscripts differ significantly (P < 0.05). (-) Indicates non-addition of endopower and a (+) indicates the addition of REAP.

Table 6. Effects of REAP supplementation on relative organ weights in broiler chicks.

Organ	High energy		Low energy	
	-	+	-	+
	g/100 g body weight (BW)			
Liver	2.55±0.03	2.82±0.10	2.64±0.11	2.68±0.06
Abdominal fat	1.73±0.08 ^a	1.57±0.07 ^{ab}	1.73±0.18 ^a	1.17±0.14 ^b
Breast muscle	5.53±0.15 ^{bc}	5.32±0.10 ^c	6.06±0.30 ^{ab}	6.32±0.17 ^a
Leg muscle	8.75±0.16	9.08±0.20	8.99±0.20	8.84±0.07

^{a-c}Means ± SE within a row with no common superscripts differ significantly (P < 0.05). (-) Indicates non-addition of REAP and (+) indicates the addition of REAP.

Table 7. Effects of REAP supplementation on weight and length of small intestine in broiler chickens.

Parameter	High energy		Low energy	
	-	+	-	+
	g/100 g body weight (BW)			
Weight				
Duodenum	1.42±0.07 ^{ab}	1.52±0.06 ^a	1.22±0.07 ^b	1.26±0.06 ^b
Jejunum	1.58±0.10	1.90±0.17	1.64±0.05	1.62±0.08
Ileum	1.61±0.09	1.72±0.23	1.53±0.13	1.48±0.10
Length				
	cm/100g BW			
Duodenum	2.62±0.09 ^a	2.54±0.06 ^a	2.25±0.10 ^b	2.58±0.11 ^a
Jejunum	6.18±0.25 ^{ab}	6.94±0.27 ^a	5.52±0.29 ^b	6.27±0.22 ^{ab}
Ileum	5.84±0.25 ^{ab}	6.50±0.55 ^a	5.42±0.08 ^b	5.81±0.17 ^{ab}

^{a,b}Means ± SE within a row with no common superscripts differ significantly (P < 0.05). (-) Indicates non-addition of REAP and (+) indicates the addition of REAP.

ileum weights were found, but percentage duodenum weights of high energy group were higher than those of the low energy group. The intestinal lengths (cm/100 g

BW) of the low energy diet group without REAP were lower than those of the others (Table 7). The results demonstrated that, dietary REAP improved body weight

gain and reduced abdominal fats. Therefore, it can be concluded that, dietary supplementation of REAP improved the nutritive value of the corn-soybean diet in the broiler chicks.

DISCUSSION

The body weight gain of birds fed the low energy diet with 0% REAP[®] was lower significantly than the other groups ($p < 0.05$) but among the groups, there were no significant differences in the feed conversion ratio. This result are in agreement with that reported by Greenwood et al. (2002) who showed that supplementation of a corn-soybean broiler starter diet with a mixture of xylanase, protease and amylase improved the BW at 14 and 42 days of age with no significant effects on the feed conversion ratio (FCR).

FCR of birds fed the low energy diet with 0% enzyme was higher than those of the other group, although, not significantly. Addition of the enzyme to the birds diet showed trend towards improving the BWG significantly ($P < 0.05$) and FCR numerically.

Pertilla et al. (2001) suggested that, the β -glucans present in corn-soybean meal diet may have affected the digestibility of nutrients. A possible explanation for this could be additional β -glucanase protease and cellulose activity contained in this enzyme preparation which might enhance the digestion of NSP and protein because by disruption of the cell wall, encapsulated intracellular nutrient may be released. Furthermore, β -glucans has been shown to decrease ileal digesta viscosity (Almirall et al., 1995; Esteve-Garcia et al., 1997; Fuente et al., 1998). Some studies indicated that, inclusion of supplemental protease, α -amylase, β -glucanases and mixed enzymes might have a positive influence on animal growth (Merstad and McNab, 1975; Moss et al., 1977; Pettersson and Aman, 1989) and increase the availability of nutrients (Walsh et al., 1993). Improvement is often attributed to the degradation of NSP (Odetallah, 2002) and improvement of the digestion and absorption of nutrients, such as amino acids and energy (Oloffs et al., 1999; Mathlouthi et al., 1999; Rutkowski et al., 1999), protein (Oloffos et al., 1999) and fats (Francesch et al., 1999). The development of secretion of digestive enzymes in the post hatched chick could also be a limiting factor in digestion (Krogdahl and Sell, 1989; Noy and Sklan, 1995; Sklan, 2001). A variety of complex proteins may not be easily digested by the young chick due to the rapid food rate and the deficiency of the necessary innate enzyme (Uni et al., 1999) leading to inefficient growth by birds, poor feed conversion ratio, and poor livability. The insufficient enzyme activity for early chicks may possibly be complemented through exogenous enzyme supplementation to promote digestion and utilization of diets. In contrast, Kocher et al. (2002) did not show significant improvements in the

BWG of broilers by supplementing a commercial enzyme containing mainly hemicellulase, pectinase, β -glucanase and some protease activities. Studies with similar multienzyme preparation (Energex) also failed to bring improvement in BWG (Mohamed and Hamza, 1991; Marsman et al., 1997)

It was shown that cellulose activity was significantly reduced after incubation with protease (Saleh et al., 2004). The lack of improvement in the performance reported by those researchers could be attributed to the protease activity in their enzyme preparations.

There were no significant differences in the feed intake among the groups. The addition of REAP[®] failed to show any significant effect on the feed intake. This result is in agreement with that reported by Tahir et al. (2005). After partial degradation, some of the insoluble NSP becomes soluble NSP with a high-molecular weight (Castanon et al. 1997). The high molecular weight NSP increases digesta viscosity (Chesson, 2001), which reduces the availability of nutrients by reducing the ingestinal passage (Campbell et al., 1989; Rotter et al., 1989; Choct and Annison, 1992). However, Dänicke et al. (1999) reported that, feed intake was higher for enzyme supplemented diets, since use of enzyme decreases mean retention time of digesta in the gizzard and large intestine and increases gut motility. Digesta viscosity and microbial fermentation decrease nutrient digestibility and the rate of absorption are increased so that more feed can be eaten. Pertilla et al. (2001) reported that, the actual differences in feed intake were relatively minor with their effect on rate of passage and digestibility likely to be negligible. An increase in the carcass weight is a typical response to an increased calorie: protein ratio (Mabray and Waldroup, 1981; Donaldson et al., 1985). Sibbald (1980) showed that the variation in the TME values of the different foodstuffs was more dependent on the rate of passage.

Some studies reported an increase in the AME of a corn-soybean diet by supplementing commercial enzyme preparations (Zanella et al., 1999; Douglas et al., 2000; Kocher et al., 2002; Gracia et al., 2003) but in the present study, the dietary supplementation of REAP[®] did not affect the values of the diets. However, digestibility coefficients measured with older animals were not universally applicable for the optimization of young broiler diets (Pertilla et al., 2001). β -glucans are less significant in older animals (Pettersson et al., 1999). Digestive enzyme activities (units/ kg of BW) measured in the pancreas and intestinal contents increased with age (Nitsan et al., 1991). In mature animals, the digestive tract is large and therefore, the rate of digesta passage is slower and enzymes and microbes have more time to digest food. Consequently, the effect of β -glucan is expected to diminish and the digestibility of nutrients to improve with age (Almirall et al., 1995).

Results of a study by Pertilla et al. (2001) showed that, β -glucans increased viscosity and decreased the apparent ileal nutrient digestibility (AID) of amino acid in

broilers. A mixture of enzymes could increase the protein digestibility of broiler feed. The enzymes could also save dietary protein (Tahir et al., 2008). A protein-sparing effect by Versazyme (VZ) was suggested by Odetallah et al. (2005). Numerical improvements in the body weight and breast meat yield in broiler chickens was reported with enzyme supplementation to diets but most prominently when applied to low ME diets with calculated ME in the range of 2964 to 3185 kcal / kg (Sims et al., 2001).

In this study, the breast muscle weight of the low energy diet birds (2980 kcal / kg) with 0.1% REAP[®] was significantly higher than that of the high energy group without enzyme ($p < 0.05$). The abdominal fat was reduced significantly by the enzyme ($P < 0.05$). This result is in agreement with that reported by Tahir et al. (2005). The increase in abdominal fat was reported to be a typical response to an increase in energy: protein ration (Mabray and Waldroup, 1981; Donaldson, 1985). The degradation of cellulose into smaller molecules, such as cellobiose, by cellulose might affect intestinal microflora, thereby changing the lipid metabolism (Tahir et al., 2005). It has been reported that, alternations of colonic microflora influence serum lipid levels (Jenkins et al., 1999). Viscous polysaccharides cause physiological and morphological changes to the digestive system in various species (Brown et al., 1979; Cassidy et al., 1981; Jacobs, 1983).

In this study, the enzyme treatment did not affect the relative weights of the digestive system and liver. The results are in close agreement with those reported by Tahir et al. (2005) who showed that enzyme treatments (cellulose and hemicellulase) did not affect the relative weights of the digestive system and liver. Gracia et al. (2003) also reported that, enzyme has no effect on the relative weights of digestive organs. But the addition of enzyme to the low energy diet group increased the duodenum length significantly ($p < 0.05$). Duodenum plays an important role in nutrient absorption.

Silva and Smithard (2002) suggested that the absorption of nutrients may be impeded by an increase in the thickness of the unstirred layer in the small intestine. It may be concluded that the antinutritive effect of NSP is related to their ability to increase digesta viscosity which in turn causes changes in gut morphology and in the efficiency of nutrient utilization by the chicken (Mehri et al., 2010).

The data presented indicated that, dietary REAP improved body weight gain and reduced abdominal fats. Therefore, it can be concluded that, dietary supplementation of REAP improves nutritive value of corn-soybean diet in broiler chicks.

REFERENCES

- Almirall M, Francesch M, Perez-Vendrell AM, Brufau J, Esteve-Garcia E (1995). The differences in intestinal viscosity produced by barley and b -glucanase alter digesta enzyme activities and ileal nutrient digestibilities more in chicks than cocks. *J. Nutr.*, 125: 947-955.
- AOAC (1990). Official Methods of Analysis of the Association of Official Analytical Chemist. 15th ed. Arlington, Virginia.
- Brown, RC, Kelleher J, Losowsky MS (1979). The effect of pectin on the structure and function of the rat small intestine. *Br. J. Nutr.*, 42: 357-365.
- Campbell GL, Rosnagel BF, Classen HL, Thacker PA (1989). Genotypic and environmental differences in extract viscosity of barley and their relationship to its nutritive value for broiler chickens. *Anim. Feed Sci. Technol.*, 26: 221-230.
- Cassidy MM, Lightfoot FG, Grau LH, Story JA, Kritchevsky D, Vahouny GH (1981). Effect of chronic intake of dietary fibers on the ultra structural topography of rat jejunum and colon: A scanning electron microscopy study. *Am. J. Clin. Nutr.*, 34: 218-228.
- Castanon JIR, Flores MP, Pettersson D (1997). Mode of degradation of non-starch polysaccharides by feed enzymes preparations. *Anim. Feed Sci. Technol.*, 68: 361-365.
- Chesson A (2001). Non-starch polysaccharide degrading enzymes in poultry diets: influence of ingredients on the selection of activities. *World's Poult. Sci. J.*, 57: 251-263.
- Choct M, Annison G (1992). Anti-nutritive effect of wheat pentosans in broiler chickens: Roles of viscosity and gut microflora. *Br. Poult. Sci.*, 33: 821-834.
- Dänicke S, Dusbl G, Jeroch H, Kluge H (1999). Factors affecting efficiency of NSP-degrading enzymes in rations for pigs and poultry. *Agribiol. Res.*, 52: 1-24.
- Donaldson WE (1985). Lipogenesis and body fat in chicks: effects of calorie-protein ratio and dietary fat. *Poult. Sci.*, 64: 1195-1204.
- Donaldson WE, Combs GF, Romoser GL (1985). Studies on energy levels in poultry rations. 1. Effect of calorieprotein ratio of the ration on growth, nutrient utilization, and body composition of chicks. *Poult. Sci.*, 35: 1100-1105.
- Douglas MW, Parsons CM, Bedford MR (2000). Effect of various soybean meal sources and Avizyme on chick growth performance and ileal digestible energy. *J. Appl. Poult. Res.*, 9: 74-80.
- Esteve-Garcia E, Brufau J, Perez-Vendrell AM, Miquel A, Duven K (1997). Bioefficacy of enzyme preparations containing b -glucanase and xylanase activities in broiler diets based on barley or wheat, in combination with flavomycin. *Poult. Sci.*, 76: 1728-1737.
- Francesch M, Perez-Moya S, Bodiola I, Brufau J (1999). Effects of cereal and feed enzyme on water consumption, dietary metabolizable energy and nutrient digestibility in broiler chickens. In Proceedings of the 12th European Symposium on Poultry Nutrition. WPSA-Dutch Branch, Veldhoven, The Netherlands, p. 258.
- Fuente JM, Perezdeayala P, Flores A, Villamide MJ (1998). Effect of storage time and dietary enzyme on the metabolizable energy and digesta viscosity of barley-based diets for poultry. *Poult. Sci.*, 7: 90-97.
- Fuente JM, Pkrez de Ayala P, Villamide MJ (1995). Effect of dietary enzyme on the metabolizable energy of diets with increasing levels of barley fed to broilers at different ages. *Anim. Feed Sci. Technol.* 56: 45-53.
- Gracia MI, Aranibar MJ, Lazaro R, Medel P, Mateos GG (2003). Alpha-amylase supplementation of broiler diets based on corn. *Poult. Sci.*, 82: 436-442.
- Greenwood MW, Fritts CA, Waldroup PW (2002). Utilization of Avizyme 1502 in corn-soybean meal diets with and without antibiotics. *Poult. Sci.*, 81(Suppl. 1): 25 (Abstr.).
- Hesselman K, Aman P (1986). The effect of b -glucanase on the utilisation of starch and nitrogen by broiler chickens fed on barley and low- or high-viscosity. *Anim. Feed Sci. Technol.*, 15: 83-93.
- Iji PA, Tivey DR (1998). Natural and synthetic oligosaccharides in broiler chicken diets. *World's Poult. Sci. J.*, 54: 129- 143
- Jacobs LR (1983). Effects of dietary fiber on mucosal growth and cell proliferation in the small intestine of rat: A comparison of oat bran, pectin, and guar with total fiber deprivation. *Am. J. Clin. Nutr.*, 37: 954-960.
- Jenkins DJ, Vuksan V, Rao AV, Vidgen E, Kendall CW, Tariq N, Wursch P, Koellreutter B, Shiwnarain N, Jeffcoat R (1999). Colonic bacterial activity and serum lipid risk factors for cardiovascular disease. *Metabolism*, 48: 264-268.
- Kocher A, Choct M, Porter MD, Broz J (2002). Effects of feed enzymes on nutritive value of soybean meal fed to broilers. *Br. Poult. Sci.*, 43,

- 54-63.
- Krogdahl A, Sell JL (1989). Influence of age on lipase, amylase, and protease activities in pancreatic tissue and intestinal contents of young turkeys. *Poult. Sci.* 68: 1561-1568.
- Mabray CJ, Waldroup PW (1981). The influence of dietary energy and amino acid levels on abdominal fat pad development in the broiler chick. *Poult. Sci.*, 60: 151-159.
- Marsman GJ, Gruppen H, Van der Poel AF, Kwakkel RP, Verstegen MW, Voragen AG (1997). The effect of thermal processing and enzyme treatments of soybean meal on growth performance, ileal nutrient digestibilities, and chyme characteristics in broiler chicks. *Poult. Sci.*, 76: 864-872.
- Mathlouthi N, Larbier M, Lessire M (1999). Effect of xylanase and β -glucanase supplementation on laying hen performances. In Proceedings of the 12th European Symposium on Poultry Nutrition. WPSA-Dutch Branch, Veldhoven, The Netherlands, p. 269.
- Mehri M, Adibmoradi M, Samie AH, Shivazad M (2010). Effects of β -Mannanase on broiler performance, gut morphology and immune system. *Afr. J. Biotechnol.*, 9(17).
- Meng X, Slominski BA, Nyachoti CA, Campbell LD, Guenter W (2005). Degradation of cell wall polysaccharides by combinations of carbohydrase enzymes and their effect on nutrient utilization and broiler chicken performance. *Poult. Sci.*, 84: 37-47.
- Merstad O, McNab JM (1975). The effect of heat treatment and enzyme supplementation on the nutritive value of barley for broiler chicks. *Br. Poult. Sci.*, 16: 1-8.
- Mohamed MA, Hamza AS (1991). Using enzyme preparation in corn-soybean meal broiler rations. *Egypt J. Anim. Prod.*, 28: 245-254.
- Moss BR, Beechler AF, Newman CW, Ei-Negoumy AM (1977). Enzyme supplementation of broiler rations. *Poult. Sci.*, 56: 1741.
- Nitsan Z, Ben-Avraham G, Zoref Z, Nir I (1991). Growth and development of the digestive organs and some enzymes in broiler chicks after hatching. *Br. Poult. Sci.*, 32: 515-523.
- Noy Y, Sklan D (1995). Digestion and absorption in the young chick. *Poult. Sci.*, 74: 366-373.
- Nukamp HJ (1965). Some remarks about the determination of the heat of combustion and the carbon content of urine. In: Blaxter KL (ed.). *Fide EAAP, Publi. Academic Press, London, New York*, 11: 147-157.
- Odetallah NH (2002). Enzymes in corn-soybean diets. In Proceedings of the 29th Annual Carolina Poultry Nutrition Conference. North Carolina State University, Raleigh, NC, pp. 37-50.
- Odetallah NH, Wang JJ, Garlich JD, Shih JCH (2002). Keratinase in starter diets improves growth of broiler chicks. *Poult. Sci.*, 81(In Review).
- Odetallah NH, Wang JJ, Garlich JD, Shih JCH (2005). Versazyme supplementation of broiler diets improves market growth performance. *Poult. Sci.*, 84: 858-864.
- Oloffs K, Samli E, Jeroch H (1999). Effect of supplementation of non-starch-polysaccharide (NSP) hydrolysing enzymes and wheat variety on prececal and faecal digestibility as well as metabolizable energy of laying hens. In Proceedings of the 12th European Symposium on Poultry Nutrition. WPSA-Dutch Branch, Veldhoven, The Netherlands, p. 256
- Pertilla S, Valaja J, Partanen K, Jalava T, Kiiskinen T, Palander S (2001). Effects of preservation method and β -glucanase supplementation on ileal amino acid digestibility and feeding value of barley for poultry. *Br. Poult. Sci.*, 42: 218-229.
- Pettersson D, Aman P (1989). Enzyme supplementation of a poultry diet containing rye and wheat. *Br. J. Nutr.*, 62: 139-149.
- Pettersson D, Graham H, Aman P (1991). The nutritive value for broiler chickens of pelleting and enzyme supplementation of a diet containing barley, wheat and rye. *Anim. Feed Sci. Technol.*, 33: 1-14.
- Rotter BA, Marquardt RR, Guenter W, Biliaderis C, Newman CW (1989). In vitro viscosity measurements of barley extracts as predictors of growth responses in chicks fed barley-based diets supplemented with a fungal enzyme preparation. *Can. J. Anim. Sci.*, 69: 431-439.
- Rutkowski A, Mand AJ, Bremmers RPM (1999). Effect of enzyme supplementation to wheat and triticale based diets on digestibility and energetic value for chickens. In Proceedings of the 12th European Symposium on Poultry Nutrition. WPSA-Dutch Branch, Veldhoven, The Netherlands, p. 267.
- Saleh F, Ohtsuka A, Tanaka T, Hayashi K (2003a). Effect of enzymes of microbial origin on in vitro digestibilities of dry matter and crude protein in soybean meal. *Anim. Sci.*, 74: 23-29.
- Saleh F, Ohtsuka A, Tanaka T, Hayashi K (2003b). Effect of enzymes of microbial origin on in vitro digestibilities of dry matter and crude protein in maize. *Poult. Sci.*, 40: 274-281.
- SAS Institute (1990). *SAS/STAT User's guide: Statistics*. SAS Inst. Inc., Cary, NC.
- Saleh F, Ohtsuka A, Tanaka T, Hayashi K (2004). Carbohydrases are digested by proteases present in enzyme preparation during *in vitro* digestion. *Poult. Sci.*, 41: 229-235.
- Sibbald IR (1980). The clearance time and rate of passage of feed residues. *Poult. Sci.*, 59: 374-377.
- Silva SSP, Smithard RR (2002). Effect of enzyme supplementation of a rye-based diet on xylanase activity in the small intestine of broilers, on intestinal crypt cell proliferation and on nutrient digestibility and growth performance of the birds. *Br. Poult. Sci.*, 43: 274-282.
- Sims MD, Blair M, Hooge DM (2001). Live performance, caloric efficiency, carcass characteristics, and cost/gain of broiler chickens fed corn-soy-poultry byproduct diets with or without the enzyme Rovabio Excel™. *Poult. Sci.*, 80(Suppl.1): p. 168.
- Sklan D (2001). Development of the digestive system of poultry. *World's Poult. Sci. J.*, 57: 415-428.
- Tahir M, Saleh F, Ohtsuka A, Hayashi K (2005). Synergistic effect of cellulose and hemicellulase on nutrient utilization and performance in broilers fed a corn-soybean meal diet. *J. Anim. Sci.*, 76: 559-565.
- Tahir M, Saleh F, Ohtsuka A, Hayashi K (2008). An effective combination of carbohydrases that enables reduction of dietary protein in broilers: importance of hemicellulase. *Poult. Sci.*, 87: 713-718.
- Uni Z, Noy Y, Sklan D (1999). Posthatch development of small intestinal function in the poult. *Poult. Sci.*, 78: 215-222.
- Walsh GA, Power RF, Headon DR (1993). Enzymes in the animal-feed industry. *Trends Biotechnol.*, 11: 424-430.
- Zanella I, Sakomura NK, Silversides FG, Figueirodo A, Pack M (1999). Effect of enzyme supplementation of broiler diets based on corn and soybeans. *Poult. Sci.*, 78: 561-568.
- Zhou Y, Jiang Z, Lu D, Wang T (2009). Improved energy-utilizing efficiency by enzyme preparation supplement in broiler diets with different metabolizable energy levels. *Poult. Sci.*, 88: 316-322.