

Tropical Journal of Pharmaceutical Research November 2015; 14 (11): 1969-1974

ISSN: 1596-5996 (print); 1596-9827 (electronic)

© Pharmacotherapy Group, Faculty of Pharmacy, University of Benin, Benin City, 300001 Nigeria.

All rights reserved.

Available online at <http://www.tjpr.org><http://dx.doi.org/10.4314/tjpr.v14i11.3>

Original Research Article

Cytotoxic T-lymphocyte Antigen-4 Binding to SHP2 Interacting Transmembrane Adapter Protein by Phosphorylation in T-Cell

Sung-Kyu Lee and Hyun Kang*

Department of Medical Laboratory Science, College of Health Science, Dankook University, Cheonan-si, Chungnam, 330-714, Republic of Korea

*For correspondence: **Email:** hyunbio@gmail.com, hkang@dankook.ac.kr; **Tel:** 82-41-550-1452; **Fax:** 82-41-559-7934

Received: 9 July 2015

Revised accepted: 11 October 2015

Abstract

Purpose: To investigate potential cytotoxic T-lymphocyte antigen-4 (CTLA-4) binding partners and assess whether potential binding partners affect the full function of CTLA-4.

Methods: The down-regulation effects of CTLA-4 and SIT were assessed by culturing CD3 stimulated T-cells. CTLA-4 and SIT proteins were measured by immunoblot analysis and production of interleukin-2 transcription activity evaluated by luciferase assay.

Results: CTLA-4 inhibited the interleukin-2 production capacity of CD3-stimulated T cells. CTLA-4 interaction with SHP2 interacting transmembrane adapter protein (SIT) in the down-regulation of the transcription of Interleukin-2 required CTLA-4 binding to SIT tyrosine motifs. The SIT tyrosine mutants were significantly lower (25 – 75 %) after phosphorylation compared with WT-SIT (transfected cells, $p < 0.05$) and untreated control. The remaining 90 % phosphorylation in the F188ANS mutant can be explained by phosphorylation of other tyrosines in the sequence of SIT ($p < 0.05$). For interleukin-2 transcription, F188ANS single mutant and double F148SEV mutant, increased NF-AT activity by 35 % compared with the wild type ($p < 0.05$).

Conclusion: The findings imply that SIT transmembrane adaptor (SIT) protein, binds to CTLA-4 and thus potentiates the inhibitory role of this co-receptor. This phenomenon may lead to the development of new treatment strategies for autoimmune diseases and graft rejection.

Keywords: Cytotoxic T-lymphocyte antigen-4, Interleukin-2, Nuclear factor of activated T-cells/Activator protein-1, SHP2 interacting transmembrane adapter protein, Autoimmune diseases, Graft rejection

Tropical Journal of Pharmaceutical Research is indexed by Science Citation Index (SciSearch), Scopus, International Pharmaceutical Abstract, Chemical Abstracts, Embase, Index Copernicus, EBSCO, African Index Medicus, JournalSeek, Journal Citation Reports/Science Edition, Directory of Open Access Journals (DOAJ), African Journal Online, Bioline International, Open-J-Gate and Pharmacy Abstracts

INTRODUCTION

CTLA-4 signaling is well-documented in its ability to downregulate T-cell proliferation and cytokine production [1,2]. Studies using CTLA-4 antibodies have shown that CTLA-4 blocks T-cell proliferation, IL-2 production and NFAT translocation into the nucleus, implying that its general effects downplay the immune response [3]. An *in vivo* study of CTLA-4 knockout mice

had tissue destruction by the immune system and extensive lymphadenopathy [4].

The full activation of naïve T-cells depends on two signals that ignite proliferation, differentiation and cytokine secretion [2]. The first is provided by the T-cell receptor (TcR) upon recognition of a specific antigenic peptide in complex with a MHC class II molecule. The second signal is delivered by the T-cell's co-stimulatory receptors upon ligation of their counterparts by antigen

presenting cells (APC). The balance between signaling of the major co-stimulatory molecules CD28 (the positive co-receptor) and CTLA-4 (the negative co-receptor) determines the outcome of immune response [5].

Transmembrane adaptor proteins (TRAPS) facilitate precise signal transduction by forming specific protein complexes [5]. Unlike kinases, which use enzymatic and transcriptional activity to promote signaling, TRAPS act as scaffolds and recruit molecules for detailed intracellular signaling complexes [6]. Recently, our laboratory found that a small TRAP, SIT, binds to CTLA-4 and enhances CTLA-4 surface expression [6]. SIT is a disulfide-linked homodimeric polypeptide which is exclusively expressed in lymphocytes. In contrast to the other transmembrane adaptor proteins known so far, SIT is a heavily glycosylated polypeptide [13]. This could indicate that SIT possesses an external ligand that modulates its function. This study was conducted to understand the molecular mechanisms leading to tyrosine phosphorylation of CTLA4/SIT complex.

EXPERIMENTAL

Cells and reagents

Jurkat T-cells (ATCC, American Type Culture Collection, VA, USA) were cultured in RPMI 1640 supplemented with 5 % fetal calf serum (FCS), 2 mM L-glutamine and 100 U/ml penicillin/streptomycin [7]. Anti-CD3 (OKT3) was obtained from the ATCC and anti-human CTLA-4 mAb was provided by Dr. Broeker (Greifswald, Germany). Anti-HA antibody was purchased from Covance Research Product Inc.

Plasmid DNA cloning

Full-length human SIT-R cDNA PCR primer sequences were 5-gctctagagccaccatg-3 sense and 5-tgacacgccaagcttggtac-3 antisense. PCR contained five cycles of 60 s at 94 °C, 30 s at 37 °C and 60 s at 72 °C and a further 25 cycles within an annealing temperature of 45 °C. The amplified products were ligated into the TA cloning vector (Invitrogen) and confirmed by DNA sequencing.

Immunoprecipitation and immunoblotting

Cells were lysed in an ice-cold lysis buffer containing 1 % Triton X-100 in 20 mM Tris-HCl, pH 8.3, 150 mM NaCl, protease inhibitors, and phosphatase inhibitors. Post-nuclear lysates were incubated for 1 h with the indicated

antibody. Protein A-Sepharose beads (30 μ l, Amersham Pharmacia) were added and incubated for 1 h at 4 °C. The eluted proteins were separated by 10 % SDS-PAGE and transferred to a nitrocellulose membrane for immunoblotting. The membranes were blocked with 5 % milk in TBS (10 mM Tris-HCl, pH 7.6, 150 mM NaCl) and incubated with their corresponding antibodies. A secondary antibody, provided by Amersham (NJ, USA), allowed the first antibody to be observed by enhanced chemiluminescence (ECL).

Interleukin-2 luciferase assay

Jurkat T-cells were co-transfected with 10 μ g of corresponding cDNA and combined with 2 μ g of pNFAT3-Luc plasmid or 0.1 μ g of a control reporter plasmid (pRL-TK from Promega). Cells were pulsed using BTX Gene Pulser at 260 V, 960 F in 10 % Fetal Calf Serum. Cells (1×10^7 cells) were aliquoted in a 96-well plate 18 hrs after transfection and cultured in a final volume of 200 μ L of RPMI 1640 growth medium. After 5 h stimulation by CD3 (145-2C11; 2 μ g/mL), CD3/rabbit anti-mouse (2 μ g/mL) antibodies, or rabbit anti-mouse antibodies alone (which served as negative controls), cells were lysed in 100 μ L of lysis buffer (Promega kit, Promega, WI, USA). Luciferase activity was quantified using a luminometer (MicroLumat, EG7G Berthold) immediately after addition of 100 μ L luciferase substrate (Promega kit) and a Stop and Go reaction to control amounts of reporter plasmid (dual luciferase system kit from Promega, WI, USA). Luciferase units of the experimental vector were normalized to the level of the control vector in each sample.

Statistical analysis

Data are presented as mean \pm SEM ($n = 3$). Statistical analysis was performed with a statistical software (SAS Institute, Cray, NC, USA) using one-way analysis of variance, followed by Dunnett's multiple range tests. $P < 0.05$ was considered statistically significant.

RESULTS

Identification of the SIT tyrosine phosphorylation

Fig 1A shows the amino acid sequence of a portion the potential tyrosine phosphorylation motifs that would be suited for SH2 domain binding. The cytoplasmic domain of SIT carries a total of five potential sites of tyrosine phosphorylation. These are Y⁹⁰GNL, Y¹²⁸TSL, Y¹⁴⁸SEV, Y¹⁶⁸ASV and Y¹⁸⁸ANS. To assess

which of these five tyrosines represent phosphorylation sites, we generated a series of SIT mutants.

To determine whether the tyrosine base motifs are sites of phosphorylation, wild-type SIT (WT SIT) and SIT mutants were expressed in Jurkat T cells, the cell were activated 2 min with pervanadate and were assessed for phosphorylation by anti-phosphotyrosine blotting (Fig. 1B, upper panel). The F⁹⁰GNL and F¹⁶⁸ASV single mutants were significantly less phosphorylated when compared with WT SIT (lane 3 and 6 vs 2). Phosphorylation was reduced even further (generally < 90 % of WT) in the F¹⁸⁸ANS (lane 7; also see histogram in Fig. 1C). As an internal control for expression, anti-SIT Ab detected similar levels of expression for SIT and the mutants (Fig. 1B, lower panel). These data demonstrate that F⁹⁰GNL, F¹⁶⁸ASV and F¹⁸⁸ANS are major sites of phosphorylation by pervanadate. The remaining 90 % phosphorylation in the F¹⁸⁸ANS mutant can be explained by protein tyrosine kinase phosphorylation of other tyrosines in the sequence of SIT.

SIT tyrosine based motifs are necessary for CTLA-4 binding

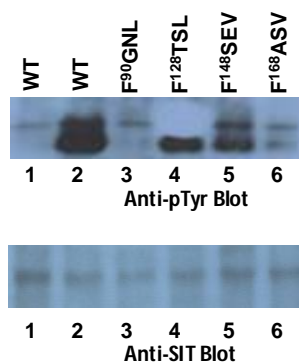
To assess whether the tyrosine sites are responsible for binding to CTLA-4, the SIT mutants (F⁹⁰GNL, F¹²⁸TSL, F¹⁴⁸SEV, F¹⁶⁸ASV and F¹⁸⁸ANS) were co-expressed with CTLA-4 in Jurkat T cells followed by immuno-precipitation with anti-CTLA Ab and anti-SIT blotting. The expression levels of SIT and the mutants and CTLA-4 were similar as shown in the whole-cell lysates (Fig. 2A, right upper and lower panels, respectively). As expected, CTLA-4 co-precipitated WT SIT (left panel, lane 1), whereas the mutant F⁹⁰GNL, F¹⁴⁸SEV, F¹⁸⁸ANS fail to co-precipitate with CTLA-4.

The single mutations resulted in significant reductions in CTLA-4 binding, with Y¹⁸⁸ANS having a greater effect (90 % reduction) than Y¹⁴⁸SEV (85 % reduction) (left panel, lanes 5 and 6 also see Fig 2B: densitometric readings in histogram).

A

```
MNQADPRLRAVCLWLTSAAMSRGDNCTDLLALGIPSITQAWGLWVLLGAVTLLFLISLAAHLSQWTRGR
SRSHPGQGRSGESVEEVPLY90GNLHYLQTGRSLQDPEPDQQDPTLGGPARAAEEVMCY128TSLQLRPP
QGRIPGPGTPVKY148SEVLDSEPKSQASGPEPELY168ASVCAQTRRARASFPDQAY188ANSQPAAS
```

B



C

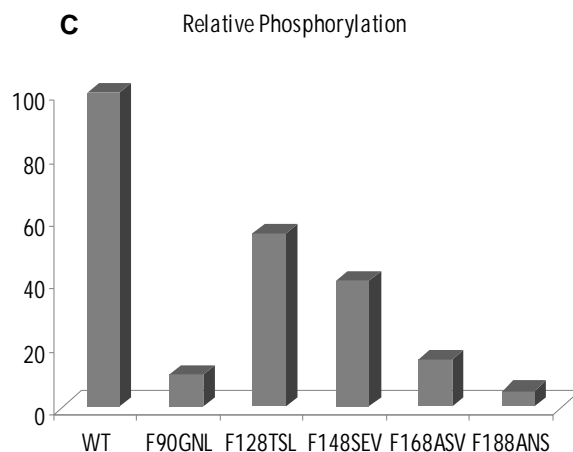


Figure 1: Identification of major sites of SIT phosphorylation by pervanadate in T cell. The tyrosines 128 and 148 are phosphorylated by pervanadate in Jurkat cells. A =The mutants of SIT created for these studies with phenylalanine (F) replacing tyrosine (Y) in the underlined region. B = cDNA wild type SIT (lanes 1 and 2) and SIT mutants (lanes 3–6) were transfected alone into Jurkat cells. Then, 18 h after transfection, lysates were prepared in 1% Triton X-100 lysis buffer, subjected to SDS-PAGE, and immunoblotted with anti-pTyr Ab (upper panel) or anti-SIT Ab (lower panel). Positions of molecular mass markers (kDa) are indicated. C, Densitometric analysis using the Scantjet laser scanner (Hewlett-Packard) of anti-pTyr binding to SIT (B, upper panel). Values were standardized with anti-SIT binding to SIT (B, lower panel). The data were normalized to the control cells. Data are expressed as the mean \pm SD (n = 5). Transfected cells (p < 0.05) compared to untreated control

SIT and CTLA-4 interaction is required to downregulate IL-2 transcription activity

To address the role of CTLA-4 binding to SIT in T cell function, Jurkat T cells were transiently transfected with SIT mutants or a control vector together with CTLA-4 and a luciferase reporter construct driven by an IL-2 NF-AT/AP-1 promoter. The combined expression of F¹⁸⁸ANS SIT mutant /CTLA-4 potentiated IL-2 transcription by about 35-fold beyond vector-transfected control (Fig 3).

By contrast, the F¹²⁸TSL and F¹⁴⁸SEV had little effect on with CTLA-4. WT experiments showed no effect. This limited interaction may be sufficient to mediate the down-regulation of TCR-stimulated NF-AT/AP-1 activity.

Analysis of the SIT mutants indicate that SIT-CTLA-4 interaction is crucial in the down-regulation of T cell pathway, as a marked enhancing of NF-AT/AP-1 activity was observed in transfectants expressing the mutants compared with the WT FYB (Fig. 3).

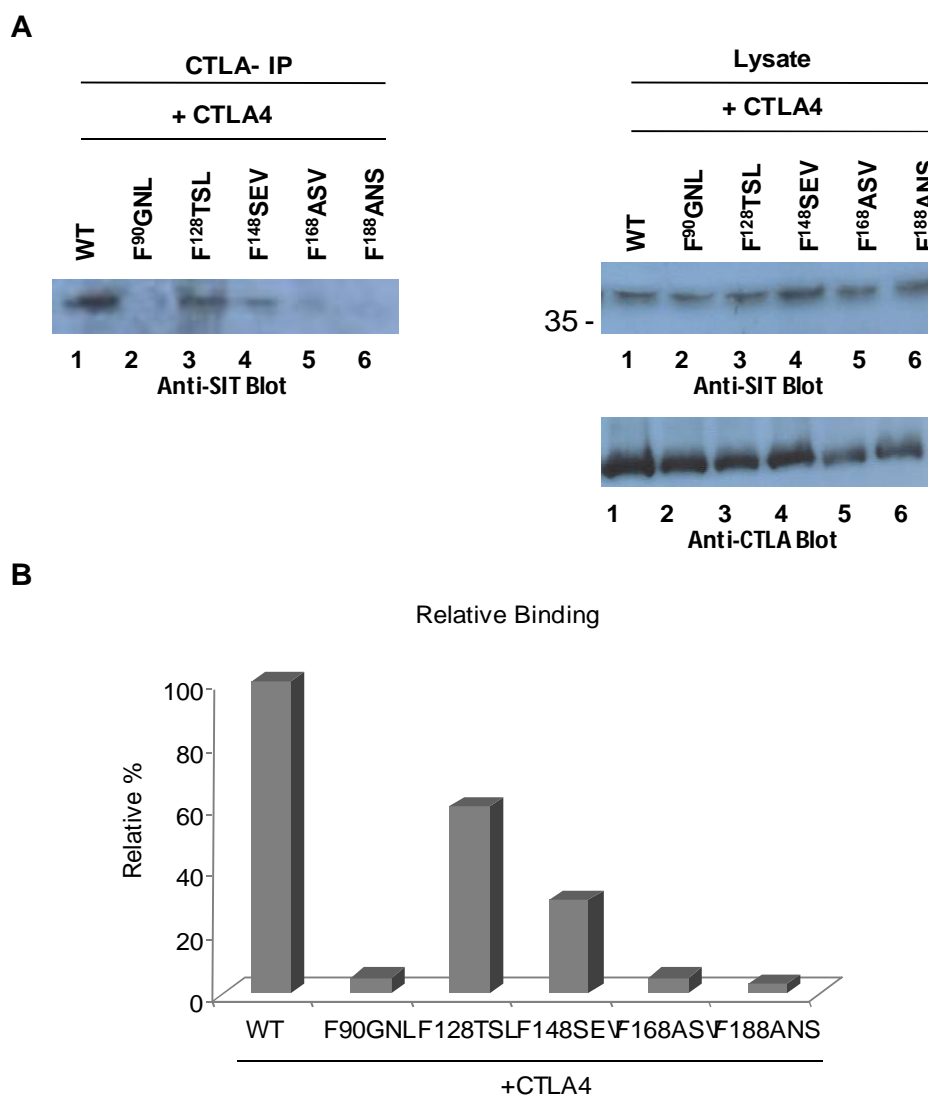


Figure 2: (A) Comparison binding analysis of = SIT mutants with CTLA-4. Then, 18 h after transfection, cells were lysed and precipitated with anti-SIT Ab. Anti-SLP-76 immune complexes (lanes 1–6) and cell lysates (A. left panel, lanes 1–6) and were subjected to SDS-PAGE and blotted with anti-SIT and anti-CTLA-4. Positions of molecular mass markers (kDa) are indicated. Lanes 1, cells transfected with WT SIT; lanes 2, F⁹⁰GNL; lanes 3, F¹²⁸TSL; lanes 4, F¹⁴⁸SEV; lanes 5, F¹⁶⁸ASV; lanes 6, F¹⁸⁸ANS. (B) Densitometric analysis using the Scantjet laser scanner (Hewlett-Packard) of SIT bound to CTLA-4 (A, left panel). The data were normalized to the control cells. Data are expressed as the mean ± SD (n = 5). Transfected cells (p < 0.05) compared to untreated control

The enhancing was greatest with the F¹⁸⁸ANS single mutant and the double F¹⁴⁸SEV mutant, enhancing the NF-AT activity by 30–35 % compared with the wild type (Fig 3).

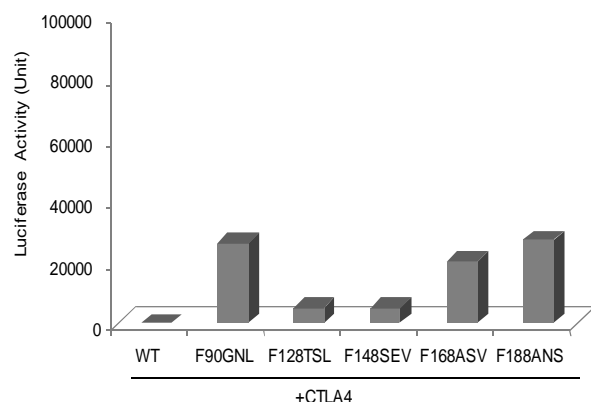


Figure 3: CTLA-4/SIT-R complex completely inhibited IL-2 gene transcription. Luciferase units of the experimental vector were normalized to the level of the control plasmid in each sample. The data were normalized to the control cells. Data are expressed as the mean \pm SD (n = 5). Transfected cells (p < 0.05) compared to untreated control

DISCUSSION

CTLA-4 plays an important role in mediating T-cell function, but little is known regarding the mechanisms behind this response. Thus, a better understanding of the molecular mechanism of CTLA-4 will allow researchers to develop therapeutic approaches to reverse the adverse impact of CTLA-4 dysfunction and harness the immune response.

Reported mechanisms include competition for CD28 binding to CD80 and CD86 [8], disruption of CD28 localization at the immunological synapse [9], binding to phosphatases PP2A and SHP-2 and interference with lipid raft expression [10]. In contrast to CD28, which is found on resting and activated cells, CTLA-4 is only found on activated cells. It is primarily an intracellular antigen whose surface expression is tightly regulated [11]. This tight regulation is achieved by controlled trafficking to the cell surface and rapid internalization. Even at its highest amounts, surface expression of CTLA-4 is only at about 3 percent of the time CD28 (the positive co-receptor) is expressed [12].

During the past few years, several TRAPs involved in the relay signaling have been identified including LAT (linker for activation of T-cells), SIT (SHP2 interacting transmembrane adaptor protein), LAX (Ick interacting membrane

protein), LIME (Ick interacting membrane protein), NTAL/LAB (T-cell activation linker/linker for activation of B-cells) and PAG/Cbp (phosphoprotein associated with glycosphingolipid-enriched microdomains/CSK binding protein) [13]. Recently, the small transmembrane adaptor protein T cell receptor-interacting molecule (TRIM) was found to bind to CTLA-4 leading to enhanced CTLA-4 surface expression [7].

SIT-R is capable of binding to CTLA-4 in T-cell. This identification of SIT binding to CTLA-4 increases the number of proteins known to bind CTLA-4, and possibly suggests that SIT and SIT-R will act in a negative signaling cascade. From our observations, it appears that in T cells both Tyr¹⁴⁸ and Tyr¹⁸⁸ are essential for optimal binding of CTLA-4.

The regulation of CTLA-4 function by interaction with SIT may serve as a possible accessory immunomodulatory molecule for optimizing T-cell function. In addition, the absence of CTLA-4 has been shown to result in organ destruction, lymphoproliferative disease and numerous autoimmune diseases in human including diabetes. On the contrary, an excessive decrease in T-cell mediated immune reaction by CTLA-4/SIT complex leads to chronic infection.

CONCLUSION

The findings of this study show that TRAP, termed SIT transmembrane adaptor (SIT) protein, binds to CTLA-4 and potentiates the inhibitory role of this co-receptor. SIT-R plays an important role in the functions of CTLA-4, and hence, extensive studies of its properties *in vivo* is required.

ACKNOWLEDGEMENT

This research was supported by a grant from the Basic Science Research Program through the National Research Foundation of Korea (NRF) funded by Korean Ministry of Education, Science and Technology (no. 2010-0025105).

REFERENCES

1. Linsley PS. Distinct roles for CD28 and cytotoxic T lymphocyte-associated molecule-4 receptors during T cell activation. *J ExpMed*1995;182: 289-292.
2. Bluestone JA, Lechler RI. Transplantation. *Curr Opin Immunol* 1995; 7: 617-619.
3. Walunas TL, Lenschow DJ, Bakker CY, Linsley PS, Freeman GJ, Green JM, Thompson CB, Bluestone

- JA. CTLA-4 can function as a negative regulator of T cell activation. *Immunity* 1994; 1: 405-413.
4. Tivol EA, Borriello F, Schweitzer AN, Lynch WP, Bluestone JA, Sharpe AH. Loss of CTLA-4 leads to massive lymphoproliferation and fatal multiorgan tissue destruction, revealing a critical negative regulatory role of CTLA-4. *Immunity* 1995; 3: 541-547.
 5. Samelson LE. Signal transduction mediated by the T cell antigen receptor: the role of adapter proteins. *Annu. Rev. Immunol* 2002; 20: 371-394.
 6. Simeoni L, Lindquist JA, Smida M, Witte V, Arndt B, Schraven B. Control of lymphocyte development and activation by negative regulatory transmembrane adapter proteins. *Immunol Rev* 2008; 224: 215-228.
 7. Valk E, Leung R, Kang H, Kaneko K, Rudd CE, Schneider H. T cell receptor-interacting molecule acts as a chaperone to modulate surface expression of the CTLA-4 coreceptor. *Immunity* 2006; 25: 807-821.
 8. Masteller EL, Chuang E, Mullen AC, Reiner SL, Thompson CB. Structural analysis of CTLA-4 function in vivo. *J Immunol* 2000; 164: 5319-5327.
 9. Lee KM, Chuang E, Griffin M, Khattri R, Hong DK, Zhang W, Straus D, Samelson LE, Thompson CB, Bluestone JA. Molecular basis of T cell inactivation by CTLA-4. *Sci* 1998; 282: 2263-2266.
 10. Chikuma S, Bluestone JA. CTLA-4 and tolerance: the biochemical point of view. *Immunol Res* 2003; 28: 241-253.
 11. Alegre ML, Noel PJ, Eisfelder BJ, Chuang E, Clark MR, Reiner SL, Thompson CB. Regulation of surface and intracellular expression of CTLA4 on mouse T cells. *J Immunol* 1996; 157: 4762-4770.
 12. Lindsten T, Lee KP, Harris ES, Petryniak B, Craighead N, Reynolds PJ, Lombard DB, Freeman GJ, Nadler LM, Gray GS. Characterization of CTLA-4 structure and expression on human T cells. *J Immunol* 1993; 151: 3489-3499.
 13. Simeoni L, Lindquist JA, Smida M, Witte V, Arndt B, Schraven B. Control of lymphocyte development and activation by negative regulatory transmembrane adapter proteins. *Immunol Rev* 2008; 224: 215-228.
 14. Chambers CA, Kuhns MS, Egen JG, Allison JP. CTLA-4-mediated inhibition in regulation of T cell responses: mechanisms and manipulation in tumor immunotherapy. *Annu Rev Immunol* 2001; 19: 565-594.