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Research Article

Acute and Sub-chronic (28-day) Oral Toxicity Studies of Hydroalcohol Leaf Extract of *Ageratum conyzoides* L (Asteraceae)

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Abstract

Purpose: Ageratum conyzoides is an annual herbaceous plant commonly used in African traditional medicine as a purgative, antipyretic, anti-ulcer and wound dressing agent. The objective of this study was to investigate the acute and sub-chronic toxicity of A. conyzoides leaves in Wistar rats.

Methods: In the acute test, the limit test dose of 5000 mg/kg was administered to Wistar rats and then observed individually 1 h post-dosing, and at least once daily for 14 days. Sub-chronic toxicity was evaluated after administering daily oral doses of 500 and 1000 mg/kg body wt., for 28 days to the rats, Biochemical and haematological assessments as well as body and relative organ weights of the rats were carried out

Results: The limit dose of 5000 mg/kg did not cause any mortality or signs of acute toxicity in the rats tested during the observation period. In the sub-chronic tests, the results did not show any treatment-related abnormalities in terms of haematological and biochemical parameters. However, urea was significantly (p < 0.05) lower in the group treated with 500 mg/kg of A. conyzoides extract. The weekly body and organ weight of the rats showed no significant differences between the control and the rats treated with the extract except for liver where there was a significant increase (p < 0.05) in rats that received 1000 mg/kg, i.e., 3 ± 0.2 g as against 2.5 ± 0.1 g for the control.

Conclusion: Our results suggest that the hydroalcohol extract of A. conyzoides is relatively safe when administered orally in rats.

Keywords: Ageratum conyzoides, Acute and sub-chronic toxicity, Biochemical parameters, Haematological analysis, Wistar rats.

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INTRODUCTION

Ageratum conyzoides is an erect, annual herbaceous plant. It is known to possess a broad spectrum of pharmacological and therapeutic properties [1]. In African traditional medicine, A. conyzoides has been used as a purgative, antipyretic, anti-ulcer and wound dressing agent [2]. In Togo, it is used to treat fever, measles and snake bites [3] while in Nigeria this plant is used for skin diseases, wound healing, diarrhoea, pain associated with navel in children [4], and in the treatment of HIV/AIDS [5].

Phytochemical investigations on Α. convzoides have identified a number of secondary metabolites such us flavonoids, coumarins and alkaloidal compounds [6-8]. Horie et al [7] reported the presence of hexametoxyflavone and 1-2 benzopirone while Moore et al [8] identified several 1.2-desifropirrolizidinic alkaloids including and licopsamine which are hepatotoxic [8]. Other compounds such as tannins have also been detected [9,10]. However, the toxicity, including hepatotoxicity, of this plant has not been extensively studied [4,11]. In view of the presence of hepatotoxic alkaloids in A. conyzoides and the lack of toxicity study on the leaf extract, the objective of this study was to investigate the acute and sub-chronic toxicity of *A. conyzoides* leaves in Wistar rats.

EXPERIMENTAL

Collection and extraction of plant materials

A. conyzoides was collected from Djagblé, Togo, in July 2007. It was identified by Prof Kouami Kokou from the Botany Department of University of Lome, Lome, Togo and a voucher specimen was kept in the herbarium of the Laboratory of Botany and Plant Ecology, Faculty of Science, University of Lome with reference no. 212 of Tchala.

The leaves of the plant were air-dried to a constant weight and ground to a coarse

powder. The dried powder (250 g) was soaked in 2 L of ethanol-water (90:10) for 72 h with occasional agitation. The solution was filtered and evaporated using a rotary evaporator, giving a dry extract yield of 15.9 %. Appropriate concentrations of the extract were made in 2 % aqueous Tween 80 and used in the animal experiments.

Animals

Wistar rats of either sex (150 - 200 g), provided by the Department of Animal Physiology, were used. They were housed in a standard environmental condition and fed with rodent standard diets and water *ad libitum*. Animal care and handling conformed to accepted guidelines [12,13]. Ethical approval was obtained from the institutional Ethical Committee for Teaching and Research (ref no. CNCB- CEER 2801/2010).

Acute toxicity test

The limit test dose of 5000 mg/kg was used as stipulated in Organization for Economic Cooperation Development (OECD) guidelines [12]. Three female rats, each sequentially dosed at intervals of 48 h, were used for the test. The animals were observed individually for acute toxicity signs and behavioural changes 1 h post-dosing, and at least once daily for 14 days.

Sub-chronic toxicity test

Repeat-dose oral toxicity study was carried out according to OECD guideline 407 [13]. The animals were divided into three groups of 8 animals each (4 males and 4 females). Group 1 received 10 ml/kg body weight of distilled water and served as control. Groups 2 and 3 received extract doses of 500 and 1000 mg/kg body wt, respectively. The extract was administered daily for 28 days the same time daily and observed at least twice daily for morbidity and mortality. Body weights of the animals were evaluated weekly.

On the 29th day, after an overnight fast, the rats were anaesthetized with ether and blood sample for haematological and biochemical analysis were collected into tubes with and without EDTA, respectively. Haemoglobin, haematocrit, red blood cell count, white blood cell count, mean corpuscular haemoglobin concentration (MCHC), mean corpuscular haemoglobin (MCH), mean corpuscular volume (MCV) and platelet count were determined using an automatic counter (Sysmex K21, Tokyo, Japan). Biochemical analysis was performed on serum obtained after centrifugation of total blood (without anticoagulant) at 2500 rpm for 15 min. Standardized diagnostic kits (Labkit®) were used for spectrophotometric determination of the following biochemical parameters: alanine aminotransferase (ALT), aspartate amino-(AST), creatinine, transferase alkaline phosphatase, glucose, total proteins and urea.

Necroscopy of all animals was carried and the organ weights (heart, liver, kidney and spleen) were recorded.

Statistical analysis

The results are expressed as mean \pm standard error of the mean (SEM). Statistical analysis was performed by one-way analysis of variance (ANOVA) with Tukey test to evaluate significant differences between groups. Values of p < 0.05 were considered significant. All statistical analyses were carried out using the Instat statistical package (Graph Pad Software, Inc., USA).

RESULTS

Acute toxicity

The limit dose of 5 g/kg did not cause mortality or any sign of acute toxicity in the three rats dosed for a short period (48 h) and long period (14 days).

Sub-chronic toxicity

No behavioural changes and death were observed at the end of the treatment. Similarly, no significant differences in body weight were observed between control and treated groups during this period (see Table 1).

Table 1: Mean body weight of rats after 28 days treatment with hydroalcohol leaf extract of *A. conyzoides*

	Mean body weight (g, ± SEM)			
Week	Control	Extract		
WCCK		500 mg/kg	1000 mg/kg	
0	132 ± 4.8	135 ± 5.2	129 ± 4.5	
1	139 ± 5.4	139 ± 4.1	133 ± 4.2	
2 3	142 ± 5.1 148 ± 4.8	144 ± 3.9 151 ± 4.4	137 ± 4.7 143 ± 5.4	
4	152 ± 5.5	156 ± 5.2	147 ± 6.4	

Fig 2 shows that the weight of liver in the group treated with 1000 mg/kg significantly higher (p < 0.05) than that of the control group. For other organs and at both doses tested, there were no changes in organ weight

Table 2: Mean organ weight of rats after 28 days treatment with hydroalcohol leaf extract of *A. conyzoides*

	Mean weight (g, ± SEM)			
Organ	Control	Extract dose		
		500 mg/kg	1000 mg/kg	
Heart	0.34±0.02(8)	0.33±0.01(8)	0.34±0.01(7)	
Liver	2.50±0.12(8)	2.70±0.11(8)	3.00±0.15(7)*	
Spleen	0.17±0.02(8)	0.18±0.018(8)	0.23±0.03(7)	
Kidney	0.55±0.08(8)	0.54±0.020(8)	0.60±0.03(7)	
Testis	1.30±0.08(4)	1.00±0.26(4)	1.00±0.23(4)	

No. of rats/group indicated in parenthesis; * p <0.05, control group vs extract

Tables 3 and 4 show the haematological and biochemical parameters for the rats after subchronic toxicity dosing

Table 3: Haematological parameters for rats after 28 days treatment with hydroalcoholic extract of *A. conyzoides* leaves.

Parameter	Control	Extract dose	
		500 mg/kg	1000 mg/kg
WBC $(10^3/\mu I)$	9.7±0.9 (8)	9.9±0.9 (7)	8.7±1.3 (7)
RBC $(10^6/\mu l)$	7.4±0.1 (8)	6.9±0.4 (7)	7.0±0.2 (7)
Haemoglobin (g/dL)	14.0±0.4 (8)	14.0±0.4 (7)	13.0±0.4 (7)
Haematocrit (%)	42.0±0.9 (8)	40.0±2.2 (7)	41.0±1.4 (7)
MCV (fl)	57.0±0.6 (8)	57.0±0.6 (7)	56.0±0.9 (7)
MCH (pg)	18.0±0.4 (8)	20.0±1.5 (7)	19.0±0.4 (7)
MCHC (%) Platelet (10 ³ /μL)	32.0±0.6 (8) 926±45 (8)	35.0±2.4 (7) 801±64 (7)	33.0±0.8 (7) 966±43 (7)

No. of rats/group indicated in parenthesis

Table 4: Biochemical parameters for rats after 28 days treatment with hydroalcohol extract of *A. conyzoides* leaves.

Davamatav		Extract dose	
Parameter	Control	500 mg/kg	1000 mg/kg
ASAT (U/L)	241±25 (8)	206±25 (8)	229±16 (7)
ALAT (U/L)	69±11 (8)	66±5 (8)	60±7 (7)
Total proteins (g/dL)	6.0±2.2 (8)	6.2±1.8 (8)	6.5±1.4 (7)
Creatinine (mg/dL)	8.5±0.7 (8)	7.9±0.4 (8)	7.4±0.9 (7)
Urea (mg/dL)	45±4 (8)	29±2 (8)*	35±4 (7)
Alkaline phosphatase (U/L)	273±52 (8)	215±45 (8)	225±56 (7)

No. of rats/group indicated in parenthesis; *Statistically significant (p <0.05, control group vs extract)

Only urea showed any significant change (p <0.05) being lower in the group treated with 500 mg/kg of the extract than in the control.

Other biochemical and haematological parameters were similar for the two groups.

DISCUSSION

The results of the acute toxicity study indicate that the LD_{50} of the extract of $A.\ conyzoides$ extract is more than 5000 mg/kg. The limit test is primarily used in situations where the investigator has information indicating that the test material is likely to be non-toxic or of low toxicity [12]. This finding, therefore, suggests that the extract at the limit dose tested is essentially non-toxic and safe in oral formulation. This result is in line with previous data from Moura et al [11] who reported that $A.\ conyzoides\ LD_{50}$ in mice is more than 10,000 mg/kg.

After 28 days of treatment, there were no treatment-related changes in haematological parameters between control and treated groups, indicating that the extract was not toxic to circulating red cells, nor interfered with their production and that of platelets. The haematopoietic system is one of the most sensitive targets of toxic compounds and is an important index of physiological and pathological status in man and animals [14]. addition. most of the biochemical parameters were not also altered by the extract. The lack of significant alterations in the levels of ALT, AST, alkaline phosphatase, glucose and creatinine, which are good indicators of liver and kidney functions, suggests that sub-chronic administration of extract neither altered hepatocytes and kidneys of rats nor the normal metabolism of the animals. The extract produced significant decrease in urea levels at the lower dose (500 mg/kg); the reason is not exactly known but one possibility could be that the different active principles present in the extract are acting differently at the doses tested and moreover. some drugs are known to decrease blood urea levels [15].

The change in the relative weight of the liver and the presence of several alkaloids, including 1.2-desifropirrolizidinic and licopsa-

mine, which can have hepatotoxic activity [13] indicate that the extract might have toxic potential on liver with increasing dose. However, it could be argued that these changes may not be toxicologically significant, as they were not corroborated by the biochemical findings (ALT, AST, alkaline phosphatase and glucose). histological study and more specific assays of toxicity could furnish more information regarding the hepatotoxicity of the extract.

CONCLUSION

Our results have demonstrated that the hydroalcohol leaf extract of *A. conyzoides* is relatively safe when administered orally in rats.

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