

Tropical Journal of Pharmaceutical Research, October 2009; 8 (5): 449-454

© Pharmacotherapy Group,
Faculty of Pharmacy, University of Benin,
Benin City, 300001 Nigeria.

All rights reserved.

Available online at <http://www.tjpr.org>

Research Article

Extractive Spectrophotometric Determination of Omeprazole in Pharmaceutical Preparations

Amol Bhandage¹, Ashok Bhosale¹, Ashok Kasture² and Vijaya Popatrao Godse^{1*}

¹Department of Pharmaceutical Analysis, S. G. R. S. College of Pharmacy, Saswad, Pune. (MS), 412301,

²Department of Pharmaceutical Sciences, Nagpur University, Nagpur, India.

Abstract

Purpose: To develop a simple, rapid and selective method for the extractive spectrophotometric determination of omeprazole using acidic dyes.

Methods: Extractive spectrophotometric determination of omeprazole was developed using acidic dyes - bromophenol blue and orange G - as ion-pairing agents in aqueous medium (pH 7.0 and 6.0, respectively). The ion pair chromogen formed, which was extracted with chloroform, was measured quantitatively at 408 nm and 508 nm, respectively. The developed method was used to analyse commercial omeprazole tablets.

Results: Using bromophenol blue and orange G dyes, the ion-paired formed obeyed Beer's law in the ranges 5 - 30 µg/ml and 50 - 250 µg/ml at 408nm and 503nm, respectively, with molar absorptivities of $1.712 \times 10^3 \text{ L mol}^{-1} \text{ cm}^{-1}$ and $2.095 \times 10^3 \text{ L mol}^{-1} \text{ cm}^{-1}$, respectively, for omeprazole,. The purity of omeprazole obtained was 98.1 ± 0.9 and 99.7 ± 0.3 , respectively. Standard deviation (S.D.), % relative standard deviation (% R.S.D.) and standard error were 0.001 - 0.013, 0.94 - 1.07 % and 4×10^{-4} , respectively. The complexes formed were stable for approx. 3 h.

Conclusion: Recovery studies gave satisfactory results indicating that none of the major additives/excipients interfered with the assay method. Therefore, a simple, rapid and selective method was developed for extractive spectrophotometric determination of omeprazole. This method may be useful for routine laboratory analysis of omeprazole.

Keywords: Extractive spectrophotometry, Omeprazole, Bromophenol blue, Orange G.

Received: 5 March 2009

Revised accepted: 12 July 2009.

*Corresponding author: **E-mail:** vijayagodse2002@gmail.com, **Tel:** +91-9923265133

INTRODUCTION

Omeprazole (Fig 1) is a member of benzimidazole class of drugs. It is an important benzimidazole derivative which is used in the treatment of gastric and duodenal ulcers, and reflux oesophagitis [1]. Its efficacy as an antiulcer and antisecretory agent has been well established.

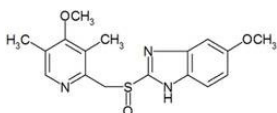


Fig. 1: Omeprazole

Modern pharmaceutical analysis demands separation of the desired component from a complex dosage formulation followed by its instrumental determination. The importance of omeprazole prompted the development of many methods for its detection and determination. Apart from official methods based on UV spectrometry [2,3], visible spectrometry [2,3], non-aqueous titrations [2-3], and TLC [2-5], a variety of analytical techniques involving HPTLC [5], HPLC [6], electrochemical [7] and polarographic [8,9] techniques have been reported in the literature. Amongst optical methods for drug analysis, visible spectrophotometry seems to be the most attractive analytical approach because it provides a simple precise and accurate measurement of suitable analytes.

Visible spectrophotometric methods used for the determination of omeprazole are based on extraction of the analyte [10-14] into a non-aqueous solvent from a complex dosage formulation. A marked advantage of this method is that it can be applied to the determination of an individual component in the presence of routine excipients and filling materials. An important advantage of the extractive spectrophotometric method is that it can be applied to the determination of an individual component in the presence of routine excipients and filling materials. This aspect of spectrophotometric analysis is of great interest since it offers distinct

possibilities for the assay of a particular component in a complex dosage formulation.

Survey of the literature revealed that only four reagents, viz, suprachen violet 3B (SV 3B), tropaeolin ooo (TP ooo), bromocresol green (BCG) and azocarmin G (AG) [15], have been reported for the extractive spectrophotometric determination of omeprazole. To the best of our knowledge, bromophenol blue and orange G, which are economical and easily available dyes, have not been used for the extractive spectrophotometric determination of omeprazole.

In this study, a method for the determination of omeprazole is described. The proposed method is based on the formation of an ion pair between omeprazole and the acidic dyes, bromophenol blue and orange G, in aqueous buffers - Clark and Lubs buffer (pH 7), Walpole buffer (acetic acid + sodium acetate (pH 4) and Sorenson buffer (pH 6), respectively.

EXPERIMENTAL

Apparatus

UV-Visible spectrophotometer Shimadzu (1601-A), fitted with a computer and a 1.0 cm matched cell, was used.

Reagents and solutions

All the chemicals used were of analytical grade. The solvents were of spectroscopic grade and were procured from Research Laboratories, Pune, India. Aqueous bromophenol blue (100 µg/ml) and orange G (100 µg/ml) solution were used.

The buffers used were: Sorenson buffer (disodium hydrogen citrate + sodium hydroxide, pH range 4.96 – 6.33), Walpole buffer (acetic acid + sodium acetate, pH range 3.72 – 5.57) and Clark and Lubs buffer (potassium dihydrogen phosphate + sodium hydroxide, pH range 5.8 – 8.0).

A stock solution of omeprazole (Torrent Pharmaceuticals, Ahmedabad) was prepared in 0.05M HCl (100 mg of the drug in 100 ml). Working solutions of the drug were prepared by dilution of the stock solution. The marketed capsule of omeprazole used in the determination was Pirazole[®] with a labeled strength of 20 mg and manufactured by Nicholas Piramal India Limited, Parel, Mumbai, India.

Optimisation of experimental conditions

While studying the effect of pH, the drug-dye ion-pair formation was found to be critically dependent on the pH of the aqueous phase; at pH values higher than 7.5, no ion-pair was formed. In this method, the ion-pairs formed with bromophenol blue (BPB) and orange G at pH 6.0 and 7.0 were extracted into chloroform and the absorbance was measured at 408 nm and 503 nm, respectively.

Prior to this experiment, several organic solvents such as benzene, toluene, cyclohexane, carbon tetrachloride, in addition to chloroform, were examined for their ability to extract the drug-dye ion-pair. Chloroform was found to be the most suitable solvent in terms of extraction efficiency and hence, it was used in this experiment.

Optimum conditions for quantitative extraction of the drug-dye ion-pair were also investigated. For omeprazole-BPB and omeprazole-orange G ion-pairs, the absorbances were maximal and constant over the pH range 6.6 - 7.4 and 5.8 - 6.2, respectively; hence, pH 7.0 and 6.0, respectively, were selected for the study and Clark and Lubs buffer and Sorenson buffer were used in the determinations. These buffers were found to provide appropriate media for efficient extraction. A single extraction with chloroform and 5 ml of buffer was found to be adequate for the concentration range investigated. Shaking times, ranging from 0.5 to 3.0 min, produced no change in absorbance; therefore, a 1 min

shaking time was selected. The absorbance of the chloroform phase was found to be stable for 2-3 h. Absorbance was measured periodically at an interval of 15 min for 3 h. The colour intensity of the organic layer had shown a very gradual increase during the first hour but subsequently became stable for 2-3 h.

The experiments were repeated with varying concentrations of dye in the range 0.01 mg to 0.2 mg per ml. The dye concentration of 0.1 mg per ml was found to give maximal absorbance.

Preparation of calibration curve

Standard solutions of omeprazole in 0.05M HCl, having final concentrations in the range 0.01 mg per 2 ml to 0.06 mg per 2 ml, were taken in separatory funnels. To each separatory funnel, 2 ml of dye solution (0.1 mg per ml), 2 ml buffer solution of pH 7 and 5 ml chloroform were added, and the mixtures were thoroughly shaken. The organic layer was allowed to separate and then collected in a dry test tube. The absorbance of each chloroform extract was measured at 408 nm against a blank similarly prepared by replacing drug solution with 0.05M HCl.

Assay of a commercial pharmaceutical preparation

Standard solutions of pure omeprazole of 10 and 150 µg/ml for bromophenol blue dye and orange G dye, respectively) were prepared.

Twenty capsules were weighed and their contents mixed thoroughly. An accurately weighed portion of the powder equivalent to the labelled strength (20mg) of the omeprazole capsules was dissolved in 10 ml of 0.05M HCl and filtered through a Whatman filter paper no. 2. The filtrate was transferred to a volumetric flask containing 0.05M HCl. An aliquot of this solution was diluted with 0.05M HCl to obtain a concentration of 10 µg/ml for bromophenol blue dye and 150 µg/ml for orange G dye.

General procedure

Several standard solutions of omeprazole in 0.05M HCl were taken in individual separatory funnels. To each separatory funnel, 2 ml of dye solution (0.1 mg/ml), 2 ml buffer solution and 5 ml chloroform were added and the mixtures were thoroughly shaken. The chloroform layers were allowed to separate, collected in separate dry test tubes and the absorbance was measured at λ max 408nm and 503nm for omeprazole-BPB complex and omeprazole-orange-G complex, respectively, against a blank similarly prepared by replacing drug solution with 0.05M HCl. The concentration of omeprazole in each test tube was obtained by interpolating the corresponding absorbance value from Beer's plot of standard omeprazole solutions.

Recovery studies

Standard solution

An accurately weighed quantity of omeprazole, equivalent to about 50 mg, was dissolved in 10 ml of 0.05M HCl and diluted to 50 ml with the same medium. It was then diluted further to a concentration of 10 μ g/ml.

Sample solution

An accurately weighed quantity of the capsule contents, equivalent to about 50 mg, was transferred to 50 ml volumetric flask and about 50 mg of accurately weighed omeprazole powder was added to it. The mixed contents were dissolved in 0.05M HCl. The solution was filtered and the final volume was made to 50 ml with the same medium. The solution was further diluted to a concentration of 20 μ g/ml.

Determination of omeprazole content

From absorbance values, concentration of the reference and the test samples were calculated, and the omeprazole content was calculated as in Eq. 1

$$\text{Weight (Q)} = \text{concentration} \times \text{dilution factor} \dots(1)$$

The weight of omeprazole contributed by capsule powder (Q) was deducted from total omeprazole and the amount of drug after deduction was assumed to be recovered from the added quantity of omeprazole. From the weight of omeprazole in standard and test sample, recovery (R) was calculated using the standard IUPAC formula provided -

$$R (\%) = \frac{Q_{\text{sample}} - Q_{\text{standard}}}{Q_{\text{standard}}} \times 100 \dots\dots(2)$$

RESULTS

The factors affecting color development, reproducibility, sensitivity and adherence to Beer's law were analysed from the data. Beer's law limit, molar absorptivity, regression data, correlation coefficient and precision data for the various omeprazole preparations are given in Table 1.

Table 1: Optical characteristics of omeprazole complexes

Parameter	Omeprazole with BPB	Omeprazole with orange G
λ max (nm)	408	503
Stability (h)	3	3
Beer's law (μ g/ ml)	5-30	50-250
Molar absorptivity (ϵ L mol ⁻¹ cm ⁻¹)	1.712 \times 10 ³	2.0954 \times 10 ³
Regression data		
Slope (a)	0.005	0.0064
Intercept (b)	-0.0003	-0.039
Correlation coefficient (r)	0.9986	0.9984
RSD (%) ^a	1.0669	0.93776

^aNumber of independent analyses, n =5.

The regression equation and regression coefficient for omeprazole-BPB complex are $Y = 0.005X - 0.0003$ and 0.9986, while for omeprazole-orange-G complex, they are $Y = 0.0064X - 0.039$ and 0.9984, respectively.

Table 2: Determination of omeprazole in commercial preparation

Preparation (mg/capsule)	Label claim method	Proposed added (found %) ^a	Analyte (%) ^a (mg) ^b	Recovery (found %) ^a	Official method (USP 24)
BPB ^c	10	98.1±0.9	10	100.7±0.7	100±1
Orange G	10	99.7±0.3	10	98.0±0.7	100±1

^aMean ± SD (n = 5); ^bAdded to the known volume of the standard solution; ^cBromophenol blue

As the results in Table 2 show, the developed method compared favorably with those obtained by USP 24 method [3].

DISCUSSION

Ion-pair extractive spectrometry has attracted considerable attention for quantitative analyses of many pharmaceutically active compounds. In the present investigation, bromophenol blue (BPB) and orange G, being anionic dyes, formed a coloured ion-pair with omeprazole at pH 7 and 6 respectively; each of the chromogen formed was soluble in chloroform and their absorbances were measurable at 408 and 503 nm, respectively.

Optimisation of the spectrophotometric conditions were intended to take into account the various goals of method development and to weigh each goal accurately. Analytical conditions were optimised via a number of preliminary experiments. Among various organic solvents studied - carbon tetrachloride, benzene and chloroform - chloroform was preferred because of its more efficient extraction of the drug-dye ion-pair from the aqueous phase. Only one extraction was sufficient to achieve quantitative recovery of the complex. Shaking times ranging from 0.5 to 3 min produced no change in absorbance, and so a one min shaking time was selected. The absorbance of the chloroform phase was stable for 2 - 3 h. The absorbance was measured periodically at an interval of 15 min for 3 h. The colour intensity of the organic layer showed a gradual increase during the 1st

hour but subsequently became stable for 2 - 3 h. The experiments were repeated with varying concentrations of dye in the range 0.01 mg to 0.2 mg per ml of dye solution but the dye concentration of 0.1 mg/ml was found to give maximal absorbance for both dyes.

The effect of pH was studied by extracting the coloured complex in the presence of various buffers of different acidic pH values. Buffers of pH 7 and 6 and bromophenol blue (0.1 mg/ml) and orange G (also 0.1 mg/ml) solutions, respectively, were the most suitable for quantitative formation of ion-pair. Being basic in nature, omeprazole formed ion-pairs with the acidic dyes, which are extractable into chloroform from the aqueous phase at the pH values provide optimal medium for the formation of ion-pair complex.

CONCLUSION

Omeprazole was estimated successfully by the developed extractive spectrophotometric method, both as a pure compound, and as a constituent of a capsule formulation. The method is simple, rapid, accurate, and does not involve any critical reaction conditions, or tedious sample preparation. It is unaffected by slight variations in experimental conditions such as pH, dye concentration, shaking time and temperature. The applicability of the new procedure for routine quality control of omeprazole in pharmaceutical formulations was established.

ACKNOWLEDGEMENT

The authors thank Torrent Pharmaceuticals, Ahmedabad, India, for supplying samples of omeprazole free of charge. The authors are

also grateful to Nagpur University, Nagpur, for permission to carry out this research.

REFERENCES

1. Doerge RF. *Wilson and Gisvold's Textbook of Organic Medical and Pharmaceutical Chemistry*, 8th edition, Lippincott, Philadelphia, 1982, pp 722.
2. *European Pharmacopoeia*, 5. European Directorate for the Quality of Medicines, Strasbourg, 2004, p 2146.
3. *United States Pharmacopoeia*, 24. USP Convention, Rockville, 2000, p 1358.
4. Dogrukol AK, Tunalier Z, Tuncel M. TLC densitometric determination of omeprazole in pharmaceutical preparations. *Pharmazie* 1998; 53: 272-273.
5. Ray S, Kumar P. HPTLC and TLC method for rapid quantification and identification of omeprazole. *Indian Drugs* 1994; 31: 543-547.
6. Cairns AM, Chiou RH, Rogers JD, Demetriades JL. Enantioselective high-performance liquid chromatographic determination of omeprazole in human plasma. *J Chromatogr* 1995; 666: 323-328.
7. Yan JL. Electrochemical behavior and the determination of omeprazole using glassy carbon electrode. *J Appl Sci* 2006; 6: 625-1627.
8. Dogrukol-Ak D, Tuncel M. Determination of omeprazole in capsules by certain polarographic techniques. *Pharmazie* 1995; 50: 701-702.
9. Ozaltin N, Temizer A. Differential pulse polarographic determination of omeprazole in pharmaceutical preparations. *Electroanal* 1994; 6: 799-803.
10. Sastry CS, Naidu PY, Murty SS. Spectrophotometric methods for the determination of omeprazole in bulk form and pharmaceutical formulations. *Talanta* 1997; 44: 1211-1217.
11. Castro D, Moreno MA, Torrado S, Lastres JL. Comparison of derivative spectrophotometric and liquid chromatographic methods for the determination of omeprazole in aqueous solution during stability studies. *J Pharm Biomed Anal* 1999; 21:291-298.
12. Ozaltin N, Kocer A. Determination of omeprazole in pharmaceuticals by derivative spectroscopy. *J Pharm Biomed Anal* 1997; 16: 337-342.
13. Lakshmi S, Anilkumar V, Venkatesan M, Raja TK. Simultaneous estimation of omeprazole and domperidone in solid oral dosage form using spectrophotometric method. *Indian drugs* 2003; 40: 589-591.
14. Mikio I, Masahiko S, Masayuki O, Takatoshi T, Ken-ichi H, Yasuyuki I. Simultaneous determination of omeprazole and its metabolite (5-hydroxyomeprazole and omeprazole sulphone) in human plasma by liquid chromatography tandem mass spectrometry. *J Liq Chromatograph and Related Tech.* 2007; 30(12): 1797-1810.
15. Sastry CSP, Naidu PY, Murthy SSN. Assay of omeprazole in pharmaceutical formulations by extraction spectrophotometry. *Ind J Pharm Sci* 1997; 59(3): 124-127.