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Floristic diversity in areas of sandy soil grasslands in Southwestern Rio Grande do Sul, Brazil

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Received: July 25 2009

Accepted: November 17 2009

Available online at <http://www.ufrgs.br/seerbio/ojs/index.php/rbb/article/view/1316>

ABSTRACT: (Floristic diversity in areas of sandy soil grasslands in Southwestern Rio Grande do Sul, Brazil). Extensive natural grasslands areas in southwestern Rio Grande do Sul occur on sandy soils, which are susceptible to the sandy patch process. This process leads to the formation of large sandy patches, which constitutes a large-scale environmental problem. In grassland areas undergoing sandy patch processes in Alegrete, Manoel Viana and São Francisco de Assis municipalities, we recorded the occurrence of 343 species, 195 genera and 52 botanical families, with two new records for Rio Grande do Sul State (*Eragrostis articulata* (Schrank) Nees and *Eragrostis leucosticta* Nees ex Döll) and one for Brazil (*Croton lorentzii* Müll. Arg. ex Griseb.). The families with higher specific richness were Asteraceae, Poaceae, Fabaceae, Euphorbiaceae, Cyperaceae, Rubiaceae and Myrtaceae. From the whole species set, 19 species show a restricted distribution area, seven of these are exclusive to Rio Grande do Sul's grasslands and 16 are considered as extinction threatened.

Key words: sandy patch process, grassland species, threatened species, restrict occurrence.

RESUMO: (Diversidade florística dos campos em áreas de solos arenosos do sudoeste do Rio Grande do Sul, Brasil). Extensas áreas de campo do sudoeste do Rio Grande do Sul ocorrem sobre solos arenosos, suscetíveis ao processo de arenização. Esse processo conduz à formação de grandes manchas de areia, constituindo um problema ambiental de grande escala. Em campos, com o processo de arenização nos municípios de Alegrete, Manoel Viana e São Francisco de Assis, foi registrada a ocorrência de 343 espécies, 195 gêneros e 52 famílias botânicas, com duas novas citações para o estado do Rio Grande do Sul (*Eragrostis articulata* (Schrank) Nees e *Eragrostis leucosticta* Nees ex Döll) e uma para o Brasil (*Croton lorentzii* Müll. Arg. ex Griseb.). As famílias com maior riqueza foram Asteraceae, Poaceae, Fabaceae, Euphorbiaceae, Cyperaceae, Rubiaceae e Myrtaceae. Do total de espécies, 19 apresentam área de dispersão restrita, sendo sete exclusivas dos campos do Rio Grande do Sul e 16 consideradas ameaçadas de extinção.

Palavras-chave: arenização, espécies campestres, espécies ameaçadas, ocorrência restrita.

INTRODUCTION

Plant landscapes are largely determined by soil, relief and climate conditions (Kern 1991). However, although climatic conditions in the South-Brazilian region are typical of forest-covered regions, extensive grassland-covered areas occur. The distribution of the South-Brazilian extant flora is not in agreement with present climate conditions, such as with the annual rainfall and temperature data (Kern 1991), being grasslands the prevailing formations in areas that have wet climate conditions, favoring the forest expansion (Behling *et al.* 2005). These grasslands, formations that had prevailed since 22 thousand years before the present, consist in relictual evidences of a dryer early climate that changed into a more humid one after 5170 years before the present (Behling *et al.* 2005). The existence of plants with attributes related to arid environments is relictual evidence of these changes and of the resulting vegetation adaptation (Marchiori 1995).

In the present time, grasslands are distributed in determined spots of the landscape due to the spatial variation of soil type and water-retaining capacity, usually

with above-average water shortage (Cunha 1992).

In Rio Grande do Sul State (RS), grasslands are included in two biomes: Pampa (southern and western halves of the state) and Mata Atlântica (northeastern area, associated to *Araucaria* forests). These grasslands present high floristic diversity, with ca. 3000 spermatophytes (Boldrini 2002), and few grassland regions in the world have diversity such as the one found in the Brazilian subtropics.

The grasslands of RS southwestern region, inserted on Pampa biome, show deep, sandy and permeable soils, where the sandy patch process takes place (Suertegaray 1995, 1998). The sandy patch process is the result of wet climate processes reworking sandstone or superficial sandy deposits through the constant mobility of the sediments. Consequently, the vegetation of extensive areas of these grasslands is buried or removed, leading to the formation of sand-covered areas that are designated as sandy patches (Suertegaray 1995, 1998, Verдум 2004). Therefore, it is a natural process, and its expansion is the consequence of nature dynamics and inadequate use of the ecosystem, particularly through intensive grazing

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(Suertegaray 1995, 1998). Starting from the decade of 1970, with the implementation of the mechanized extensive monoculture and the consequent reduction in grazing areas, there was an increase in the number of areas undergoing sandy patch process (Verdum 2004). Nowadays, spots undergoing sandy patch process occur in Alegrete, Cacequi, Itaqui, Maçambará, Manoel Viana, Quarai, Rosário do Sul, São Borja, São Francisco de Assis and Unistalda municipalities (Suertegaray *et al.* 2001).

These grasslands, locally named as “campo limpo”, show a flat landscape distinguished by the presence of smooth hills and testimonial (tabular) hills, with sandstone substratum (Verdum 2004). With xeromorphic attributes derived from strong edaphic influences, these grasslands have witnessed a semiarid period that underwent a humidification that started in the Mid Holocene (Kern 1991, Suertegaray 1995, 1998, Medeiros *et al.* 1995). Vegetation is characterized by species with abundant hairiness in leaves, flowers and other organs, flowers in large and/or colorfull inflorescences, presence of volatile oils that grant a strong scent to their green organs, short leaves and the presence of xylopodium or other subterraneous organs (Lindman 1906).

The sandy patch process, alongside the surge in agriculture, the reduction of graze-destined areas, the growing implementation of cultivated forests and the flexible use of environmental laws has progressively contributed to the reduction of biological diversity in grassland formations. The destruction of the Pampa biome is no longer a threat, but a reality, because ca. 50% of the original vegetation cover has been destroyed or deeply altered (Hasenack *et al.* 2007).

The present paper aimed to describe the floristic diversity of natural sandy soil grassland areas in the southwest of Rio Grande do Sul state, at Alegrete, Manoel Viana and São Francisco de Assis municipalities.

MATERIAL AND METHODS

The study site, located in the southwest of RS State,

between the latitudes 29°00'S and 31°00'S and the longitudes 54°30'W and 58°45'W, is characterized by the occurrence of sandy patches. The study covered natural grassland areas present in sandy soils with traces of sandy patch process, with and without cattle grazing, at Alegrete, Manoel Viana and São Francisco de Assis municipalities (Fig. 1).

According to Köppen classification, climate in the region is Cfa type, with 1400 mm mean annual rainfall (Nimer 1979). However, rainfall is poorly distributed, with the occurrence of reduced rainfall periods interposed by heavy rains, often concentrated in only one day (Freitas *et al.* 2009). The region shows great hydric limitation due to the atmosphere high evaporative uptake, mainly between November and February (Berlato *et al.* 2006). During this period, the probability of hydric deficiency (relationship between rainfall and evapotranspiration) is 41% in November, rising up to 60% in December and decreasing to 47% and 36% in January and February, respectively (Leivas *et al.* 2006). Mean annual temperature is 14.3°C in winter and 26.3°C in summer (Nimer 1979), while maximum and minimum absolutes can reach 40°C and -4°C, respectively (Cordeiro & Soares 1975).

The landscape is smoothly rippled, permeated by flat areas, configuring a surface of unconsolidated arenous deposits cradled over the Botucatu Formation sandstone. The flat areas contrast with the silicified sandstone plateaus in the regional landscape (Suertegaray 1995, 1998). The soil, classified as a typical Orthic Quartzarenic Neosol (Streck *et al.* 2008), shows arenous and silt-arenous texture, acid pH, lack of phosphorus and potassium, aluminum surplus (Verdum 2004), strong natural limitations in fertility, high erosion susceptibility and low cation-exchanging capacity (Azevedo & Kaminski 1995). Besides that, the soil shows water-retaining difficulties, which leads to hydric shortage even in short dry periods (Klamt & Schneider 1995).

For the floristic survey, field expeditions were

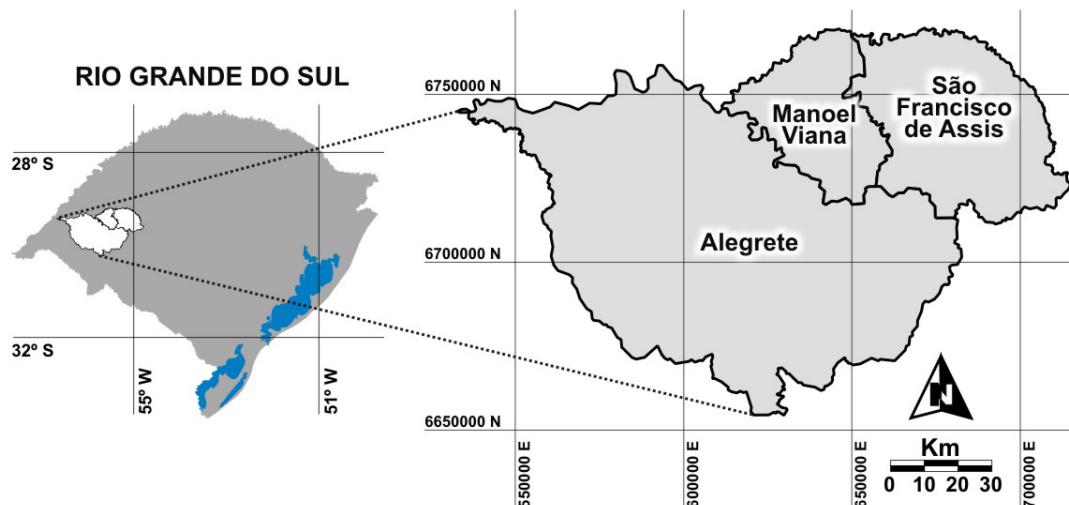


Figure 1. Location of Alegrete, Manoel Viana and São Francisco de Assis municipalities in southwestern Rio Grande do Sul, Brazil.

conducted between 2006 and 2009, covering all months. We collected fertile botanic material of the phanerogamous species present in the grassland areas, including herbs, vines, shrubs and trees occurring in scattered patterns along the grasslands. The collected material was herborized, identified and included in the ICN Herbarium collection of the Biosciences Institute of Universidade Federal do Rio Grande do Sul (UFRGS). Botanical families were considered according to APG II (2003). Some species were identified in the field but were not included in the herbarium collection, because they were in vegetative phase. Furthermore, we added to our floristic list the species found in a survey carried out in a grassland area at São Francisco de Assis municipality during 2004 and 2005. The collected botanical material for this previous survey can be found in the HVAT herbarium (Vale do Taquari Herbarium) of the Natural Sciences Museum, Univates University Center, Lajeado municipality, RS.

RESULTS AND DISCUSSION

Floristic diversity

In the floristic survey, 343 species were found, pertaining to 194 genera and 52 botanical families (angiosperms and gymnosperms) (Table 1). Families with the highest species richness were Asteraceae (77 species), Poaceae (58 species), Fabaceae (31 species), Euphorbiaceae (15 species), Cyperaceae and Rubiaceae (11 species each) and Myrtaceae (10 species) (Fig. 2). Besides these families, Malvaceae (nine species), Amaranthaceae, Apocynaceae and Convolvulaceae (eight species each) also stand out. The eleven families (21.2%) with the highest species richness correspond to 71.6% of the total species number. Of all the families, 21 (40.4%) are represented by only one species, and eight by two species. The genera with the highest species number were *Eupatorium*, *Baccharis*, *Eragrostis*, *Senecio*, *Vernonia*, *Aristida*, *Axonopus*, *Croton*, *Eryngium*, *Oxalis*, *Paspalum*, *Eugenia*, *Pterocaulon* and *Sisyrinchium* (Fig. 3).

For the South-Brazilian grasslands, Boldrini (2002) pointed out Asteraceae (ca. 600 species), Poaceae (400 species) and Fabaceae (150 species) as the most representative families. In the present survey, besides these families, other ones such as Apocynaceae, Euphorbiaceae and Myrtaceae figure among the most species-rich families, making these grasslands floristically different from the remaining grasslands in southern Brazil.

Previous studies focusing on the grassland vegetation distributed in other areas of RS, when compared to our results, indicate differences in floristic composition, mainly concerning the most numerous families (Boldrini & Eggers 1996, Boldrini *et al.* 1998, Boldrini *et al.* 2008). The floristic inventory of the grassland vegetation at São Pedro hill, which presents the higher proportions of dry grassland formations at the granitic hills of Porto Alegre

municipality (RS), pointed out the occurrence of 497 native species pertaining to 66 botanical families. Considering the dry grassland areas, the most representative families were Asteraceae (97 spp.), Poaceae (63 spp.), Fabaceae (44 spp.), Rubiaceae (16 spp.), Verbenaceae (13 spp.), Apiaceae (11 spp.), Malvaceae and Cyperaceae (10 spp.) (Setúbal & Boldrini 2009). In these studies, the families Apocynaceae, Euphorbiaceae and Myrtaceae, when represented, showed reduced species number.

The reason for the existing differences in floristic composition between the sandy soil grasslands and the grasslands covering other areas of RS state may be the result of adaptations to edaphic and climatic factors existing in the first ones since dryer climatic times, when grasslands were the prevailing formations in the region. Nowadays, these formations are predominantly found on soils of low fertility, high acidity and high susceptibility to hydric and aeolian erosion, associated to high probability of hydric shortage, summer rains and extreme temperatures. Besides that, the direct anthropic action, through cattle treading and grazing, contributes for the selection of grassland species (Boldrini & Eggers 1996).

There is a certain similarity between the most numerous botanical families at the Cerrado biome and at the sandy soil grasslands of RS. In a study accomplished by Silva *et al.* (2002) at Parque Estadual da Serra de Caldas Novas, Goiás state, Apocynaceae and Myrtaceae figured between the families with highest species number. Tannus & Assis (2004), studying the floristic composition of a Cerrado area at Itapiranga region, São Paulo state, verified that Euphorbiaceae and Myrtaceae were among the richest families. Such resemblance may also be the consequence of climatic and edaphic conditions found at the Cerrado region (rainy periods followed by dry ones, light temperatures during the year and intemperized soils with lack of nutrients and high aluminum concentration) (Klink & Machado 2005).

Studies concerning sandy soil grasslands at RS state are scarce. Trindade *et al.* (2008) evaluated the floristic composition of grassland areas located at the edge of 11 sandy patches at São Francisco de Assis, Manoel Viana and Alegrete municipalities, registering the occurrence of 53 species and 16 families. The most representative families were Poaceae, Asteraceae and Fabaceae. Rovedder *et al.* (2005) also evaluated the floristic composition in the edges and in the core of a sandy patch in a grassland area at Alegrete municipality, southwestern RS, identifying the occurrence of 42 species and 13 families, with Poaceae as the richest family, followed by Fabaceae, Rubiaceae, Asteraceae and Myrtaceae. Euphorbiaceae and Cyperaceae registered few species, probably because the survey focused on edges and cores of sandy patches, whereas our survey encompassed all the sandy soil grassland area. Freitas *et al.* (2009), in a floristic and phytosociological survey of a grassland undergoing sandy patch process at São Francisco de Assis municipality, registered the occurrence of 102 species

Table 1. Families and species of sandy soil grasslands in southwestern Rio Grande do Sul state, with respective vouchers.

Family/Species	Voucher
ACANTHACEAE <i>Justicia axillaris</i> (Nees) Lindau	<i>Freitas s.n.</i> (HVAT1444)
<i>Stenandrium diphylum</i> Nees	<i>Freitas 576</i> (ICN)
<i>Stenandrium dulce</i> (Cav.) Nees	<i>Freitas s.n.</i> (HVAT1721)
<i>Ruellia bulbifera</i> Lindau	<i>Freitas 579</i> (ICN)
<i>Ruellia</i> sp.	<i>Freitas 645</i> (ICN)
ALLIACEAE <i>Nothoscordum bonariense</i> (Pers.) Beauverd	<i>Freitas 386</i> (ICN)
AMARANTHACEAE <i>Alternanthera hirtula</i> (Mart.) R.E. Fr.	<i>Bruisma 002</i> (ICN)
<i>Alternanthera paelonga</i> A. St.-Hil.	<i>Bruisma 069</i> (ICN)
<i>Froelichia tomentosa</i> (Mart.) Moq.	<i>Freitas 055</i> (ICN)
<i>Gomphrena celosioides</i> Mart.	<i>Mundeleski 020</i> (ICN)
<i>Gomphrena graminea</i> Moq.	<i>Freitas 096</i> (ICN)
<i>Gomphrena perennis</i> L.	<i>Freitas 378</i> (ICN)
<i>Pfaffia gnaphaloides</i> (L. f.) Mart.	<i>Freitas 056</i> (ICN)
<i>Pfaffia tuberosa</i> (Sprengel) Hicken	<i>Freitas 335</i> (ICN)
ANACARDIACEAE <i>Schinus weinmannifolius</i> Engl.	<i>Freitas 656</i> (ICN)
APIACEAE <i>Apium leptophyllum</i> (Pers.) Muell. ex Benth.	<i>Freitas 365</i> (ICN)
<i>Centella hirtella</i> Nannf.	<i>Freitas 649</i> (ICN)
<i>Eryngium ciliatum</i> Cham. & Schldl.	<i>Freitas 451</i> (ICN)
<i>Eryngium elegans</i> Cham. & Schldl.	<i>Freitas 091</i> (ICN)
<i>Eryngium eriophorum</i> Cham. & Schldl.	<i>Mundeleski 051</i> (ICN)
<i>Eryngium horridum</i> Malme	<i>Bruisma 008</i> (ICN)
<i>Eryngium nudicaule</i> Lam.	<i>Freitas 413</i> (ICN)
APOCYNACEAE <i>Asclepias campestris</i> Vell.	<i>Freitas 387</i> (ICN)
<i>Asclepias mellodora</i> A. St.-Hil.	<i>Freitas 652</i> (ICN)
<i>Blepharodon lineare</i> (Decne.) Decne.	<i>Freitas 637</i> (ICN)
<i>Macrosiphonia longiflora</i> (Desf.) Müll. Arg.	<i>Freitas 087</i> (ICN)
<i>Oxypetalum glomeratum</i> E. Fourn.	<i>Freitas 516</i> (ICN)
<i>Oxypetalum</i> sp.	<i>Freitas 333</i> (ICN)
<i>Oxypetalum solanoides</i> Hook. & Arn.	<i>Freitas 417</i> (ICN)
<i>Tabernaemontana australis</i> Müll. Arg.	<i>Freitas 385</i> (ICN)
ARECACEAE <i>Butia lallemandii</i> Deble & Marchiori	<i>Freitas 098</i> (ICN)
ARISTOLOCHIACEAE <i>Aristolochia sessilifolia</i> (Klotzsch) Duch.	<i>Freitas 415</i> (ICN)
ASTERACEAE <i>Acanthospermum australe</i> (Loefl.) Kuntze	<i>Freitas 392</i> (ICN)
<i>Achyrocline marchiorii</i> Deble	<i>Muller s.n.</i> (ICN152811)
<i>Achyrocline saturaeoides</i> (Lam.) DC.	<i>Freitas 283</i> (ICN)
<i>Ambrosia tenuifolia</i> Spreng.	<i>Bruisma 063</i> (ICN)
<i>Aspilia montevidensis</i> (Spreng.) Kuntze	<i>Freitas 080</i> (ICN)
<i>Asteropsis megapotamica</i> (Spreng.) Marchesi, Bonifacino & Sancho	<i>Mundeleski 040</i> (ICN)
<i>Baccharis albolanosa</i> A.S. Oliveira & Deble	<i>Freitas 506</i> (ICN)
<i>Baccharis coridifolia</i> DC.	<i>Freitas 489</i> (ICN)
<i>Baccharis dracunculifolia</i> DC.	<i>Bruisma 048</i> (ICN)
<i>Baccharis leptophylla</i> DC.	<i>Freitas 287</i> (ICN)
<i>Baccharis multifolia</i> A.S.Oliveira, Deble & Marchiori	<i>Bruisma 051</i> (ICN)
<i>Baccharis pseudotenuifolia</i> Malag.	<i>Mundeleski 032</i> (ICN)
<i>Baccharis punctulata</i> DC.	<i>Bruisma 051</i> (ICN)
<i>Baccharis riograndensis</i> Malag. & J.E. Vidal	<i>Mundeleski 022</i> (ICN)
<i>Baccharis trimera</i> (Less) DC.	<i>Freitas s.n.</i> (HVAT1479)
<i>Calea clematidea</i> Baker	<i>Freitas 513</i> (ICN)
<i>Calea uniflora</i> Less.	<i>Freitas 101</i> (ICN)
<i>Centhratherum camporum</i> (Hassl.) Malme	<i>Freitas 514</i> (ICN)
<i>Chaptalia integriflora</i> (Vell.) Burk.	<i>Freitas 501</i> (ICN)
<i>Chaptalia nutans</i> (L.) Pol.	<i>Freitas 288</i> (ICN)
<i>Chaptalia sinuata</i> (DC.) Baker	<i>Heberle s.n.</i> (ICN157298)
<i>Chevreulia sarmentosa</i> (Pers.) S.F. Blake	<i>Freitas 556</i> (ICN)
<i>Conyza bonariensis</i> var. <i>microcephala</i> (Cabrera) Cabrera	<i>Freitas 448</i> (ICN)
<i>Conyza primulifolia</i> (Lam.) Cuatrec. & Lourteig	<i>Bruisma 064</i> (ICN)
<i>Elephantopus mollis</i> Kunth	<i>Freitas s.n.</i> (HVAT1385)
<i>Eupatorium angusticeps</i> Malme	<i>Freitas 180</i> (ICN)
<i>Eupatorium calyculatum</i> Hook. & Arn.	<i>Freitas 641</i> (ICN)
<i>Eupatorium commersonii</i> (Cass.) Hieron.	<i>Freitas 509</i> (ICN)
<i>Eupatorium inulifolium</i> Kunth	<i>Freitas 493</i> (ICN)
<i>Eupatorium laevigatum</i> Lam.	<i>Bruisma 049</i> (ICN)
<i>Eupatorium macrocephalum</i> Less.	<i>Bruisma 062</i> (ICN)
<i>Eupatorium squarrulosum</i> Hook. & Arn.	<i>Bruisma 042</i> (ICN)
<i>Eupatorium subhastatum</i> Hook. & Arn.	<i>Freitas 099</i> (ICN)
<i>Eupatorium tanacetifolium</i> Gillies ex Hook. & Arn.	<i>Freitas 562</i> (ICN)
<i>Eupatorium tweedianum</i> Hook. & Arn.	<i>Mundeleski 041</i> (ICN)
<i>Facelis retusa</i> (Lam.) Sch. Bip.	<i>Freitas 397</i> (ICN)
<i>Gamochaeta americana</i> (Mill.) Wedd.	<i>Heberle s.n.</i> (ICN157277)
<i>Gamochaeta falcata</i> (Lam.) Cabrera	<i>Freitas 329</i> (ICN)
<i>Gamochaeta spicata</i> Cabrera	<i>Freitas 391</i> (ICN)

Table 1. cont.

Family/Species	Voucher
<i>Gochnatia cordata</i> Less.	<i>Freitas</i> 508 (ICN)
<i>Hieracium commersonii</i> Monnier	<i>Freitas</i> 559 (ICN)
<i>Hypochaeris chillensis</i> (Kunth) Britton	<i>Heberle s.n.</i> (ICN157310)
<i>Hypochaeris megapotamica</i> Cabrera	<i>Freitas</i> 618 (ICN)
<i>Hysterionica filiformis</i> (Spreng.) Cabrera	<i>Freitas</i> 553 (ICN)
<i>Isostigma peucedanifolium</i> (Spreng.) Less.	<i>Freitas</i> 624 (ICN)
<i>Lucilia acutifolia</i> (Poir.) Cass.	<i>Bruisma</i> 018 (ICN)
<i>Lucilia nitens</i> Less.	<i>Freitas</i> 400 (ICN)
<i>Mikania thapsoides</i> DC.	<i>Freitas</i> 496 (ICN)
<i>Mikania trachypleura</i> B. L. Rob.	<i>Freitas</i> 426 (ICN)
<i>Noticastrum acuminatum</i> (DC.) Cuatrec.	<i>Freitas</i> 502 (ICN)
<i>Noticastrum diffusum</i> (Pers.) Cabrera	<i>Bruisma</i> 041 (ICN)
<i>Noticastrum gnaphaloides</i> (Baker) Cuatrec.	<i>Freitas</i> 086 (ICN)
<i>Orthopappus angustifolius</i> (Sw.) Gleason	<i>Freitas</i> 168 (ICN)
<i>Pterocaulon alopecuroides</i> (Lam.) DC.	<i>Freitas</i> 174 (ICN)
<i>Pterocaulon angustifolium</i> DC.	<i>Freitas</i> 178 (ICN)
<i>Pterocaulon lorentzii</i> Malme	<i>Freitas</i> 108 (ICN)
<i>Pterocaulon polypterum</i> (DC.) Cabrera	<i>Freitas</i> 640 (ICN)
<i>Senecio brasiliensis</i> (Spreng.) Less. var. <i>brasiliensis</i>	<i>Freitas</i> s.n. (HVAT1636)
<i>Senecio cisplatinus</i> Cabrera	<i>Freitas</i> 020 (ICN)
<i>Senecio grisebachii</i> var. <i>schyzotus</i> Cabrera	<i>Freitas</i> s.n. (HVAT1633)
<i>Senecio leptolobus</i> DC.	<i>Freitas</i> 014 (ICN)
<i>Senecio oxyphyllus</i> DC.	<i>Freitas</i> 021 (ICN)
<i>Senecio riograndensis</i> Matzenbacher	<i>Freitas</i> s.n. (HVAT1557)
<i>Senecio selloi</i> (Spreng.) DC.	<i>Freitas</i> 568 (ICN)
<i>Solidago chilensis</i> Meyen	<i>Mundeleski</i> 037 (ICN)
<i>Soliva pterosperma</i> (Juss.) Less.	<i>Freitas</i> 435 (ICN)
<i>Stenachaenium riedelii</i> Baker	<i>Freitas</i> 548 (ICN)
<i>Tagetes ostenei</i> Hicken	<i>Bruisma</i> 066 (ICN)
<i>Trixis pallida</i> Less.	<i>Freitas</i> 511 (ICN)
<i>Trixis praestans</i> (Vell.) Cabrera	<i>Freitas</i> 574 (ICN)
<i>Trixis verbascifolia</i> (Gardner) S.F. Blake	<i>Freitas</i> 492 (ICN)
<i>Vernonia brevifolia</i> Less.	<i>Freitas</i> 370 (ICN)
<i>Vernonia cognata</i> Less.	<i>Freitas</i> 176 (ICN)
<i>Vernonia macrocephala</i> Less.	<i>Freitas</i> 436 (ICN)
<i>Vernonia megapotamica</i> Spreng.	<i>Freitas</i> 507 (ICN)
<i>Vernonia nudiflora</i> Less.	<i>Freitas</i> 267 (ICN)
<i>Vernonia sellowii</i> Less.	<i>Spellmeier</i> 094 (ICN)
BORAGINACEAE	
<i>Euploca salicoides</i> (Cham.) J.I.M. Melo & Semir	<i>Freitas</i> 646 (ICN)
<i>Cordia verbenacea</i> DC.	<i>Freitas</i> 557 (ICN)
BRASSICACEAE	
<i>Lepidium aletes</i> J.F. Macbr.	<i>Freitas</i> 577 (ICN)
BROMELIACEAE	
<i>Dyckia vicentensis</i> T. Strehl	<i>Freitas</i> 390 (ICN)
CACTACEAE	
<i>Cereus hildmannianus</i> K. Schuman	<i>Freitas</i> 650 (ICN)
<i>Echinopsis oxygona</i> (Link) Zucc.	<i>Freitas</i> 626 (ICN)
<i>Parodia ottonis</i> (Lehmann) N. P. Taylor	<i>Freitas</i> 587 (ICN)
<i>Frailea cataphracta</i> (Dams) Britton & Rose	Sem testemunho
CAMPANULACEAE	
<i>Lobelia hederacea</i> Cham.	<i>Freitas</i> 518 (ICN)
<i>Wahlenbergia linarioides</i> (Lam.) A. DC.	<i>Freitas</i> 456 (ICN)
CARYOPHYLLACEAE	
<i>Cardionema ramosissima</i> (Weinm.) A. Nelson & J.F. Macbr.	<i>Freitas</i> 035 (ICN)
<i>Cerastium commersonianum</i> DC.	<i>Freitas</i> 011 (ICN)
<i>Paronychia brasiliiana</i> DC.	<i>Freitas</i> 357 (ICN)
<i>Polycarpon tetraphyllum</i> (L.) L.	<i>Freitas</i> s.n. (HVAT1637)
<i>Silene gallica</i> L.	<i>Heberle s.n.</i> (ICN157287)
<i>Stellaria media</i> (L.) Vill.	<i>Freitas</i> 332 (ICN)
CISTACEAE	
<i>Helianthemum brasiliense</i> (Lam.) Pers.	<i>Heberle s.n.</i> (ICN157286)
COMMELINACEAE	
<i>Commelinia erecta</i> L.	<i>Freitas</i> 424 (ICN)
<i>Commelinia rufipes</i> var. <i>glabrata</i> (D.R.Hunt) Faden & D.R. Hunt	<i>Freitas</i> 079 (ICN)
<i>Tradescantia umbraculifera</i> Hand.-Mazz.	<i>Freitas</i> 657 (ICN)
CONVOLVULACEAE	
<i>Cuscuta xanthochortos</i> Mart.	<i>Freitas</i> 078 (ICN)
<i>Dichondra sericea</i> Swartz	<i>Freitas</i> 434 (ICN)
<i>Evolvulus glomeratus</i> Nees & C. Mart.	<i>Freitas</i> 423 (ICN)
<i>Evolvulus sericeus</i> Swartz	<i>Santos</i> s.n. (ICN148690)
<i>Evolvulus sericeus</i> f. <i>pedunculatus</i> Ooststr.	<i>Freitas</i> 069 (ICN)
<i>Ipomoea kunthiana</i> Meisn.	<i>Freitas</i> 615 (ICN)
<i>Ipomoea malvaeoides</i> var. <i>lineariloba</i> Hallier	<i>Freitas</i> 449 (ICN)
<i>Ipomoea nitida</i> Griseb.	<i>Freitas</i> 616 (ICN)
CYPERACEAE	
<i>Bulbostylis capillaris</i> var. <i>elatior</i> Osten	<i>Freitas</i> 351 (ICN)
<i>Bulbostylis major</i> Palla	<i>Freitas</i> 659 (ICN)
<i>Bulbostylis sphaerocephala</i> (Boeck.) C.B. Clarke	<i>Freitas</i> 289 (ICN)
<i>Carex phalaroides</i> Kunth	<i>Freitas</i> 552 (ICN)

Table 1. cont.

Family/Species	Voucher
<i>Carex sororia</i> Kunth	<i>Freitas 500</i> (ICN)
<i>Cyperus aggregatus</i> (Willd.) Endl.	<i>Freitas 272</i> (ICN)
<i>Cyperus</i> sp.	<i>Spellmeier 090</i> (ICN)
<i>Kyllinga odorata</i> Vahl	<i>Heberle s.n.</i> (ICN157276)
<i>Kyllinga vaginata</i> Lam.	<i>Freitas 376</i> (ICN)
<i>Lipocarpha humboldtiana</i> Nees	<i>Freitas 175</i> (ICN)
<i>Rhynchospora rugosa</i> (Vahl) Galé	<i>Heberle s.n.</i> (ICN157275)
DROSERACEAE	
<i>Drosera brevifolia</i> Pursh	<i>Freitas 550</i> (ICN)
ERICACEAE	
<i>Agarista eucalyptoides</i> (Cham. & Schtdl.) G. Don	<i>Freitas 639</i> (ICN)
EUPHORBIACEAE	
<i>Acalypha communis</i> Müll. Arg.	<i>Freitas 427</i> (ICN)
<i>Croton echinulatus</i> (Griseb.) Croizat	<i>Freitas 555</i> (ICN)
<i>Croton glandulosus</i> L.	<i>Freitas 200</i> (ICN)
<i>Croton lorentzii</i> Müll. Arg. ex Griseb.	<i>Bruisma 006</i> (ICN)
<i>Croton parvifolius</i> Müll. Arg.	<i>Schneider 1581</i> (ICN)
<i>Croton subpannosus</i> Müll. Arg. ex Griseb.	<i>Freitas 049</i> (ICN)
<i>Euphorbia caecorum</i> Mart. ex Boiss.	<i>Freitas 422</i> (ICN)
<i>Euphorbia papillosa</i> A. St.-Hil.	<i>Freitas 349</i> (ICN)
<i>Euphorbia selloi</i> (Klotzsch & Gärcke) Boiss.	<i>Freitas 082</i> (ICN)
<i>Euphorbia</i> sp.	<i>Freitas 663</i> (ICN)
<i>Jatropha isabelliae</i> Müll. Arg.	<i>Freitas 377</i> (ICN)
<i>Manihot</i> sp.	<i>Freitas 564</i> (ICN)
<i>Sapium haematospermum</i> Müll. Arg.	<i>Santos s.n.</i> (ICN156290)
<i>Sebastiania hispida</i> var. <i>graciliramea</i> Pax & K. Hoffm.	<i>Freitas 092</i> (ICN)
<i>Sebastiania hispida</i> var. <i>interceedens</i> (Müll. Arg.) Pax	<i>Freitas 445</i> (ICN)
FABACEAE	
<i>Aeschynomene histrix</i> var. <i>incana</i> (Vogel) Benth.	<i>Freitas 044</i> (ICN)
<i>Arachis burkartii</i> Handro	<i>Freitas 402</i> (ICN)
<i>Chamaecrista flexuosa</i> (L.) E. Greene var. <i>flexuosa</i>	<i>Spellmeier 089</i> (ICN)
<i>Chamaecrista repens</i> (Vogel) H.S. Irwin & Barneby	<i>Bruisma 005</i> (ICN)
<i>Chamaecrista rotundifolia</i> (Pers.) Greene	<i>Freitas 189</i> (ICN)
<i>Clitoria nana</i> Benth.	<i>Freitas 188</i> (ICN)
<i>Crotalaria tweediana</i> Benth.	<i>Freitas 421</i> (ICN)
<i>Desmanthus virgatus</i> (L.) Willd.	<i>Freitas 443</i> (ICN)
<i>Desmanthus</i> sp.	<i>Muller s.n.</i> (ICN159196)
<i>Desmodium barbatum</i> (L.) Benth.	<i>Freitas 638</i> (ICN)
<i>Desmodium incanum</i> DC.	<i>Santos s.n.</i> (ICN148691)
<i>Eriosema tacuarembense</i> Arechav.	<i>Freitas 094</i> (ICN)
<i>Galactia benthamiana</i> Micheli	<i>Freitas 623</i> (ICN)
<i>Galactia gracillima</i> Benth.	<i>Freitas 648</i> (ICN)
<i>Lupinus albescens</i> Hook. & Arn.	<i>Freitas 317</i> (ICN)
<i>Lupinus bracteolaris</i> Desr.	<i>Freitas 595</i> (ICN)
<i>Lupinus lanatus</i> Benth.	<i>Heberle s.n.</i> (ICN158799)
<i>Macropitilium heterophyllum</i> (Humb. & Bonpl. ex Willd.) Maréchal & Baudet	<i>Bruisma 072</i> (ICN)
<i>Macropitilium prostratum</i> (Benth.) Urb.	<i>Freitas 116</i> (ICN)
<i>Mimosa bifurca</i> Benth.	<i>Freitas 575</i> (ICN)
<i>Pomaria pilosa</i> (Vogel) B.B. Simpson & G.P. Lewis	Sem testemunho
<i>Rhynchosia corylifolia</i> Mart. ex Benth.	<i>Freitas 084</i> (ICN)
<i>Rhynchosia lineata</i> Benth.	<i>Freitas 085</i> (ICN)
<i>Senna pallifera</i> (Vogel) H.S. Irwin & Barneby	<i>Freitas 455</i> (ICN)
<i>Senna scabriuscula</i> (Vogel) H. S. Irwin & Barneby	<i>Lima s.n.</i> (ICN151412)
<i>Stylosanthes guianensis</i> var. <i>subviscosa</i> Benth.	<i>Spellmeier 099</i> (ICN)
<i>Stylosanthes montevideensis</i> Vogel	<i>Freitas 383</i> (ICN)
<i>Stylosanthes</i> sp.	<i>Freitas s.n.</i> (HVAT1402)
<i>Vigna peduncularis</i> var. <i>clitoroides</i> (Mart. ex Benth.) Maréchal, Mascherpa & Stainier	<i>Freitas 617</i> (ICN)
<i>Zornia reticulata</i> Sm.	<i>Freitas 393</i> (ICN)
<i>Zornia</i> sp.	<i>Freitas 596</i> (ICN)
GESNERIACEAE	
<i>Sinningia stricta</i> (Hook. & Arn.) Wiehler	<i>Freitas 642</i> (ICN)
HYPERICACEAE	
<i>Hypericum connatum</i> Lam.	<i>Freitas 177</i> (ICN)
IRIDACEAE	
<i>Cypella herbacea</i> Kook.	<i>Bruisma 057</i> (ICN)
<i>Sisyrinchium hasslerianum</i> Baker	<i>Freitas 356</i> (ICN)
<i>Sisyrinchium micranthum</i> Cav.	<i>Freitas 322</i> (ICN)
<i>Sisyrinchium ostenianum</i> Beauv.	<i>Heberle s.n.</i> (ICN159211)
<i>Sisyrinchium vaginatum</i> Spreng.	<i>Freitas 573</i> (ICN)
JUNCACEAE	
<i>Juncus capillaceus</i> Lam.	<i>Lima s.n.</i> (ICN157268)
<i>Juncus scirpoideus</i> Lam.	<i>Spellmeier 110</i> (ICN)
LAMIACEAE	
<i>Hesperozygis ringens</i> Benth.	<i>Bruisma 007</i> (ICN)
<i>Hyptis mutabilis</i> (Rich.) Briq.	<i>Freitas 494</i> (ICN)
<i>Marsypianthes chamaedrys</i> (Vahl) Kuntze	<i>Santos s.n.</i> (ICN159202)
<i>Marsypianthes hassleri</i> Briq.	<i>Freitas 062</i> (ICN)
<i>Peltodon longipes</i> Kunth ex Benth.	<i>Mundeleski 055</i> (ICN)
<i>Salvia ovalifolia</i> A. St.-Hil.	<i>Freitas 498</i> (ICN)

Table 1. cont.

Family/Species	Voucher
LAURACEAE <i>Ocotea pulchella</i> (Nees) Mez	<i>Heberle s.n.</i> (ICN158803)
LOGANIACEAE <i>Spigelia stenophylla</i> Progel	Sem testemunho
LYTHRACEAE <i>Cuphea racemosa</i> (L.f.) Spreng.	<i>Bruisma 060</i> (ICN)
MALPIGHIAEAE <i>Galphimia</i> sp.	<i>Freitas 605</i> (ICN)
<i>Heteropterys</i> sp.	<i>Freitas 446</i> (ICN)
MALVACEAE <i>Aenia mansfeldiana</i> (Herter) Herter & Cristobal	<i>Spellmeier 092</i> (ICN)
<i>Hochreutineria hasslerana</i> (Hochr.) Krapov.	<i>Freitas 171</i> (ICN)
<i>Krapovickasia flavescentia</i> (Cav.) Fryxell	<i>Freitas 043</i> (ICN)
<i>Krapovickasia urticifolia</i> (A. St.-Hil.) Fryxell	<i>Freitas 660</i> (ICN)
<i>Sida rhombifolia</i> L.	<i>Freitas 037</i> (ICN)
<i>Sida vespertina</i> Ekman	<i>Mundeleski 058</i> (ICN)
<i>Sida viarum</i> A. St.-Hil.	<i>Bruisma 014</i> (ICN)
<i>Sida</i> sp.	<i>Freitas 619</i> (ICN)
<i>Waltheria douradinha</i> A. St.-Hil.	<i>Freitas 064</i> (ICN)
MELASTOMATACEAE <i>Tibouchina gracilis</i> (Bonpl.) Cogn.	<i>Bruisma 067</i> (ICN)
MOLLUGINACEAE <i>Mollugo verticillata</i> L.	<i>Freitas 431</i> (ICN)
MORACEAE <i>Dorstenia brasiliensis</i> Lam.	<i>Freitas 416</i> (ICN)
MYRTACEAE <i>Blepharocalyx salicifolius</i> (Kunth) O. Berg	<i>Freitas 653</i> (ICN)
<i>Campomanesia aurea</i> O. Berg.	<i>Freitas 352</i> (ICN)
<i>Eugenia arenosa</i> Mattos	<i>Freitas 581</i> (ICN)
<i>Eugenia hiamalis</i> Cambess.	<i>Freitas 572</i> (ICN)
<i>Eugenia pitanga</i> (O. Berg) Kiaersk.	<i>Freitas 105</i> (ICN)
<i>Eugenia plurisepala</i> Barb. Rodr. ex Chodat & Hassl.	<i>Freitas 042</i> (ICN)
<i>Hexachlamys humilis</i> O. Berg	<i>Freitas 319</i> (ICN)
<i>Myrcia verticillaris</i> O. Berg	<i>Freitas 265</i> (ICN)
<i>Myrciagenia campestris</i> (DC.) D. Legrand & Kausel	<i>Freitas 644</i> (ICN)
<i>Psidium luridum</i> (Spreng.) Burret	<i>Freitas 197</i> (ICN)
ONAGRACEAE <i>Ludwigia sericea</i> (Cambess.) Hara	<i>Spellmeier 111</i> (ICN)
<i>Oenothera molissima</i> L.	<i>Freitas 419</i> (ICN)
OROBANCHACEAE <i>Agalinis communis</i> (Cham. & Schldl.) D'Arcy	<i>Freitas 661</i> (ICN)
<i>Buchnera longifolia</i> Kunth	<i>Freitas 613</i> (ICN)
OXALIDACEAE <i>Oxalis lasiopetala</i> Zucc.	<i>Freitas 488</i> (ICN)
<i>Oxalis conorrhiza</i> Jacq.	<i>Freitas 560</i> (ICN)
<i>Oxalis eriocarpa</i> DC.	<i>Freitas 009</i> (ICN)
<i>Oxalis hispidula</i> Zucc.	<i>Freitas s.n.</i> (HVAT1529)
<i>Oxalis perdicaria</i> (Molina) Bertero	<i>Freitas 490</i> (ICN)
PLANTAGINACEAE <i>Angelonia integrifolia</i> Spreng.	<i>Freitas 077</i> (ICN)
<i>Plantago tomentosa</i> Lam.	<i>Freitas 394</i> (ICN)
<i>Scoparia ericacea</i> Cham. & Schldl.	<i>Freitas 389</i> (ICN)
<i>Scoparia montevidensis</i> (Kuntze) R.E. Fr.	<i>Freitas 551</i> (ICN)
POACEAE <i>Agrostis montevidensis</i> Spreng. ex Nees var. <i>montevidensis</i>	<i>Bruisma 059</i> (ICN)
<i>Andropogon lateralis</i> Nees	<i>Spellmeier 085</i> (ICN)
<i>Andropogon sellianus</i> (Hackel) Hackel	<i>Freitas 053</i> (ICN)
<i>Andropogon ternatus</i> (Spreng.) Nees	<i>Freitas 440</i> (ICN)
<i>Aristida circinalis</i> Lindm.	<i>Freitas 274</i> (ICN)
<i>Aristida condylifolia</i> Caro	<i>Spellmeier 091</i> (ICN)
<i>Aristida filifolia</i> (Arecav.) Herter	<i>Muller s.n.</i> (ICN153139)
<i>Aristida glaziowii</i> Hack. ex Henrard	<i>Freitas 277</i> (ICN)
<i>Aristida jubata</i> (Arecav.) Herter	<i>Freitas 109</i> (ICN)
<i>Axonopus affinis</i> Chase	<i>Freitas s.n.</i> (HVAT1639)
<i>Axonopus argentinus</i> Parodi	<i>Freitas 342</i> (ICN)
<i>Axonopus fissifolius</i> (Raddi) Kuhlm.	<i>Freitas 344</i> (ICN)
<i>Axonopus siccus</i> (Nees) Kuhlm.	<i>Freitas 182</i> (ICN)
<i>Axonopus suffultus</i> (Mikan ex Trin.) Parodi	<i>Santos s.n.</i> (ICN148682)
<i>Briza subaristata</i> Lam.	<i>Freitas 569</i> (ICN)
<i>Cenchrus echinatus</i> L.	<i>Freitas s.n.</i> (HVAT1723)
<i>Cynodon dactylon</i> (L.) Pers.	<i>Mundeleski 027</i> (ICN)
<i>Dichanthelium sabulorum</i> (Lam.) Gould et C.A. Clark var. <i>sabulorum</i>	<i>Freitas 345</i> (ICN)
<i>Digitaria ciliaris</i> (Retz.) Koeler	<i>Freitas 491</i> (ICN)
<i>Digitaria eriantha</i> subsp. <i>pentzii</i> (Stent) P.D.F.Kok	<i>Mundeleski 040</i> (ICN)
<i>Digitaria insularis</i> (L.) Fedde	<i>Freitas 437</i> (ICN)
<i>Eleusine tristachya</i> (Lam.) Lam.	<i>Mundeleski 028</i> (ICN)
<i>Elionurus</i> sp.	<i>Freitas 338</i> (ICN)
<i>Eragrostis articulata</i> (Schrank) Nees	<i>Freitas 051</i> (ICN)
<i>Eragrostis bahiensis</i> Schrad. ex Schult.	<i>Freitas s.n.</i> (HVAT1473)

Table 1. cont.

Family/Species	Voucher
<i>Eragrostis curvula</i> (Schrad.) Nees	<i>Freitas 314</i> (ICN)
<i>Eragrostis leucosticta</i> Nees ex Döll	<i>Freitas 089</i> (ICN)
<i>Eragrostis lugens</i> Nees	<i>Freitas 347</i> (ICN)
<i>Eragrostis mexicana</i> (Hornem.) Link subsp. <i>mexicana</i>	<i>Freitas 263</i> (ICN)
<i>Eragrostis neesii</i> Trin.	<i>Freitas 047</i> (ICN)
<i>Eragrostis plana</i> Nees	<i>Freitas 186</i> (ICN)
<i>Eragrostis polytricha</i> Nees	<i>Freitas 275</i> (ICN)
<i>Eustachys distichophylla</i> (Lag.) Nees	<i>Freitas 531</i> (ICN)
<i>Eustachys retusa</i> (Lag.) Kunth	<i>Freitas 075</i> (ICN)
<i>Gymnopogon spicatus</i> (Spreng.) Kuntze	<i>Freitas 280</i> (ICN)
<i>Leptocoryphium lanatum</i> (Kunth) Nees	<i>Freitas 268</i> (ICN)
<i>Melica eremophila</i> M. A. Torres	<i>Freitas 504</i> (ICN)
<i>Melinis repens</i> (Willd.) Zizka	<i>Freitas 012</i> (ICN)
<i>Microchloa indica</i> (L. F.) P. Beauv.	<i>Freitas s.n.</i> (HVAT1562)
<i>Panicum olyroides</i> Kunth	<i>Freitas 602</i> (ICN)
<i>Panicum tricholaenoides</i> Steud.	<i>Freitas 102</i> (ICN)
<i>Pappophorum macrosperrnum</i> Roseng., B.R. Arrill. & Izag.	<i>Freitas 264</i> (ICN)
<i>Paspalum leptum</i> Schult. (= <i>Paspalum nicorae</i> Parodi)	<i>Freitas 033</i> (ICN)
<i>Paspalum notatum</i> Flüggé	<i>Freitas 185</i> (ICN)
<i>Paspalum plicatulum</i> Michx.	<i>Freitas 312</i> (ICN)
<i>Paspalum polypodium</i> Nees ex Trin.	<i>Freitas 534</i> (ICN)
<i>Paspalum stellatum</i> Humb. & Bonpl. ex Flüggé	<i>Freitas 497</i> (ICN)
<i>Piptochaetium montevideense</i> (Spreng.) Parodi	<i>Freitas 346</i> (ICN)
<i>Poa lanigera</i> Nees	<i>Freitas 561</i> (ICN)
<i>Schizachyrium imberbe</i> (Hack.) A. Camus	<i>Freitas 594</i> (ICN)
<i>Schizachyrium microstachyum</i> (Desv. ex Ham.) Roseng. B.R. Arrill. & Izag.	<i>Santos s.n.</i> (ICN152719)
<i>Schizachyrium spicatum</i> (Spreng.) Herter	<i>Freitas 036</i> (ICN)
<i>Schizachyrium</i> sp.	<i>Freitas 660</i> (ICN)
<i>Setaria parviflora</i> (Poir.) Kerguélen	<i>Freitas 066</i> (ICN)
<i>Sporobolus indicus</i> (L.) R. Br. var. <i>indicus</i>	<i>Freitas s.n.</i> (HVAT1466)
<i>Steinchisma hians</i> (Elliott) Nash	<i>Freitas s.n.</i> (HVAT1406)
<i>Stipa melanosperma</i> J. Presl	<i>Freitas 313</i> (ICN)
<i>Trachypogon montufarii</i> (Kunth) Nees var. <i>montufarii</i>	<i>Freitas 337</i> (ICN)
<i>Vulpia austriaca</i> (Nees ex Steud.) C. H. Blom	<i>Heberle s.n.</i> (ICN157283)
PODOCARPACEAE	
<i>Podocarpus lambertii</i> Klotzsch ex Endl.	<i>Freitas 103</i> (ICN)
POLYGALACEAE	
<i>Monnieria cuneata</i> A. St.-Hil.	<i>Freitas 592</i> (ICN)
<i>Monnieria tristaniana</i> A. St.-Hil.	<i>Mundeleski 052</i> (ICN)
<i>Polygala molluginifolia</i> A. St.-Hil. & Moq.	<i>Mundeleski 056</i> (ICN)
<i>Polygala pulchella</i> A. St.-Hil. & Moq.	<i>Freitas 578</i> (ICN)
PORTULACACEAE	
<i>Portulaca grandiflora</i> Hook.	<i>Freitas 208</i> (ICN)
<i>Portulaca</i> sp.	<i>Freitas 432</i> (ICN)
RUBIACEAE	
<i>Galianthe fastigiata</i> Griseb.	<i>Spellmeier 123</i> (ICN)
<i>Galium atherodes</i> Spreng.	<i>Bruisma 061</i> (ICN)
<i>Galianthe verbenoides</i> (Cham. & Schlcht.) Griseb.	<i>Fleig 138</i> (ICN)
<i>Mitracarpus megapotamicus</i> (Spreng.) Kuntze	<i>Freitas 167</i> (ICN)
<i>Richardia brasiliensis</i> Gomes	
<i>Richardia humistrata</i> (Cham. & Schltdl) Steud.	<i>Freitas 371</i> (ICN)
<i>Richardia stellaris</i> (Cham. & Schltdl.) Steud.	<i>Freitas 398</i> (ICN)
<i>Spermacoce brachystemonoides</i> Cham. & Schltdl.	<i>Freitas 040</i> (ICN)
<i>Spermacoce poaya</i> A. St.-Hil.	<i>Freitas 425</i> (ICN)
<i>Spermacoce tenella</i> Kunth	<i>Freitas 520</i> (ICN)
<i>Staelia thymoides</i> Cham. & Schltdl.	<i>Santos s.n.</i> (ICN159144)
RUTACEAE	
<i>Zanthoxylum rhoifolium</i> Lam.	<i>Freitas 654</i> (ICN)
SAPOTACEAE	
<i>Chrysophyllum marginatum</i> (Hook. & Arn.) Radlk.	<i>Freitas 535</i> (ICN)
SOLANACEAE	
<i>Calibrachoa thymifolia</i> (A. St.-Hil.) Stehmann & Semir	<i>Freitas 107</i> (ICN)
<i>Petunia integrifolia</i> Schinz & Thell.	<i>Bruisma 054</i> (ICN)
<i>Solanum americanum</i> Mill.	<i>Freitas 285</i> (ICN)
<i>Solanum hasslerianum</i> Chodat	<i>Freitas 038</i> (ICN)
TURNERACEAE	
<i>Piriqueta suborbicularis</i> (A. St.-Hil. & Naudin) Arbo	<i>Freitas 519</i> (ICN)
<i>Turnera sidoides</i> subsp. <i>integrifolia</i> (Griseb.) Arbo	<i>Heberle s.n.</i> (ICN159199)
VERBENACEAE	
<i>Aloysia gratissima</i> (Gillies & Hook.) Tronc.	<i>Freitas 173</i> (ICN)
<i>Glandularia peruviana</i> (L.) Small	<i>Freitas 583</i> (ICN)
<i>Glandularia tenera</i> (Spreng.) Cabrera	<i>Freitas 010</i> (ICN)
<i>Glandularia thymoides</i> (Cham.) N. O'Leary	<i>Freitas 503</i> (ICN)
<i>Lantana camara</i> L.	<i>Freitas 571</i> (ICN)
<i>Lippia arechavaletae</i> Moldenke	<i>Freitas 081</i> (ICN)
<i>Verbena litoralis</i> Kunth.	<i>Mundeleski 044</i> (ICN)
VIOLACEAE	
<i>Hybanthus bicolor</i> (A. St.-Hil.) A. St.-Hil. & Baill.	<i>Freitas 045</i> (ICN)

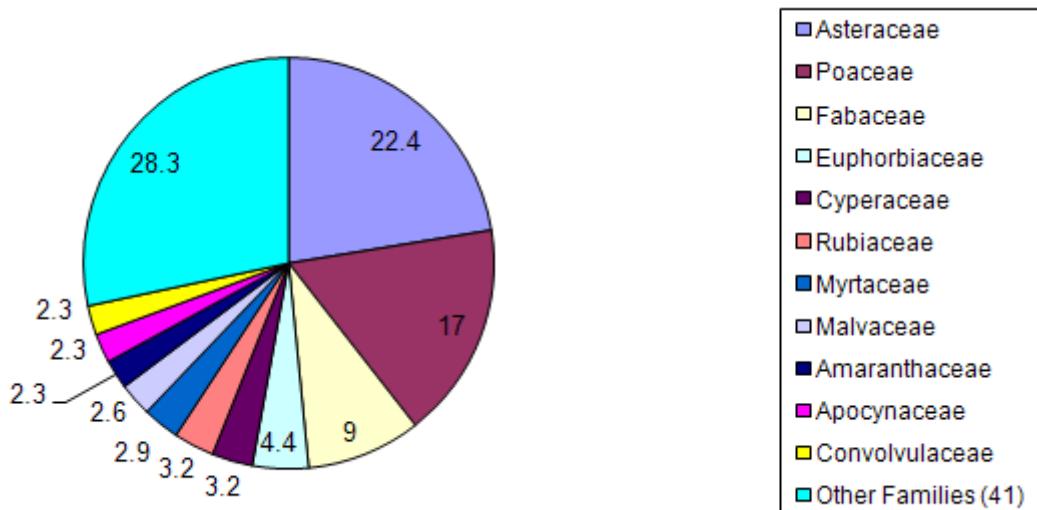


Figure 2. Percentual distribution of the families with higher richness values in sandy soil grasslands at Alegrete, Manoel Viana and São Francisco de Assis municipalities, RS.

distributed in 25 families. Among the richest families were Poaceae (29 species), Asteraceae (18 species) and Myrtaceae (six species).

Some species found in the study area are more resistant, surviving the soil removal or the sand burying processes: *Croton subpannosus*, *Elionurus* sp., *Eugenia pitanga*, *E. plurisepala*, *E. arenosa*, *Hexachlamys humilis*, *Psidium luridum*, *Paspalum lepton* (= *Paspalum nicorae*), *P. notatum* and *Vernonia brevifolia*.

Paspalum lepton and *P. notatum* are prostrate rhizomatous species, which allows their vegetative propagation when exposed to cattle treading. Besides that, they show resistance to sand burying processes when the sand is wind-driven. Both species are abundant in grassland areas undergoing sandy patch process where grazing is present. Besides these species, *Elionurus* sp., commonly known as ‘capim-limão’ (‘lemon-grass’) (Fig. 4A), *Vernonia brevifolia* (Fig. 4B) and *Croton subpannosus* (Fig. 4C, 4D) were frequently observed at edges and cores of sandy patches, evidencing resistance to erosive and sand burying processes. *Vernonia brevifolia* shows a well developed lignified base, favouring its

survival in adverse environments such as the sandy patches.

Senecio cisplatinus (Fig. 5A, 5B) is another species with marked presence in the grassland areas of southwestern RS. It frequently occurs in edges and cores of sandy patches, and was scarcely found in areas not undergoing sandy patch process. Due to the presence of wax, sand grains lifted by the wind adhere to the plant’s aerial structure. On the other hand, *Senecio selloi* (Fig. 5C, 5D), also frequent in these grasslands, was observed only in grassland areas distant from the sandy patches, showing no evident resistance to the sandy patch process.

Due to the abundance of individuals in their populations, *Vernonia macrocephala* (Fig. 5E, 5D) and *Axonopus argentinus* also stand out in the study area. Both species are frequent, constituting large populations and covering broad grassland areas.

Myrtaceae species, especially *Hexachlamys humilis* (Fig. 6A), *Eugenia arenosa* (Fig. 6B, 6C), *E. pitanga* (Fig. 6D) and *E. plurisepala* (Fig. 6E), probably favoured by their deep roots and coriaceous leaves, survive amidst sandy patches. They were frequently observed in spots with sparse vegetation cover (Fig. 6F).

Other species showed potential for regeneration of grassland areas undergoing sandy patch process. Among them, *Acanthospermum australe* (Fig. 7A, 7B) and *Lupinus albescens* (Fig. 7C, 7D) deserve special attention. The first, due to its capacity to grow on small sand mounds, contributes to the stabilization of the sandy patch process. The latter, occurring only in the grasslands of RS state in Brazil (Pinheiro & Miotto 2001), has been mentioned as an alternative to sandy patch revegetation. It shapes dense populations in areas undergoing advanced sandy patch process (Fig. 7E, 7F), acting as a natural colonizer of this environment, which verifies its adaptation to the region’s unfriendly edaphic conditions (Rovedder et al. 2005). This species

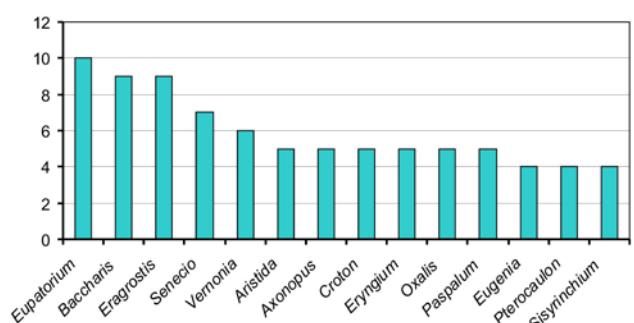


Figure 3. Genera with higher species richness, with the respective species number, of the sandy soil grasslands at Alegrete, Manoel Viana and São Francisco de Assis municipalities, RS.

shows high potential for use in recuperation initiatives concerning degraded areas, due to its high seed production and ease in occupying opened communities. Besides these, *Panicum tricholaenoides*, *Pappophorum macrospermum* and *Paspalum polyphyllum* are cespitose-rhizomatous grasses common in sandy areas, contributing for sand fastening.

Among the species registered for the study site, according to Schneider (2007) and Souza & Lorenzi (2005) nine are naturalized: *Cardionema ramosissima*, *Cerastium commersonianum* (Fig. 8A), *Polycarpon tetraphyllum*, *Silene gallica*, *Stellaria media*, *Digitaria ciliaris*, *Digitaria eriantha* subsp. *pentzii*, *Eragrostis plana* and *Melinis repens*. Among these, *Eragrostis plana*

is also characterized as an invasive plant, probably the most aggressive one in RS state due to its high capacity of seed production, allowing dispersion to great distances (Schneider 2007). *Melinis repens* is also mentioned by the author as one of the most aggressive species in the state's herbaceous flora.

New records

Among the species found in the area, *Croton lorentzii* (Fig. 8B) appears as a new record for Brazil. It is a shrub with distribution records for central and NE Argentina and for Uruguay (Burkart & Bacigalupo 2005). In the study area, the species was found in grasslands undergoing sandy patch process at São Francisco de Assis



A



C



B



D

Figure 4. Species frequently found at edges and cores of sandy patches. A. *Elionurus* sp.. B. *Vernonia brevifolia*. C. *Croton subpannosus* (habit). D. *C. subpannosus* (inflorescence detail).

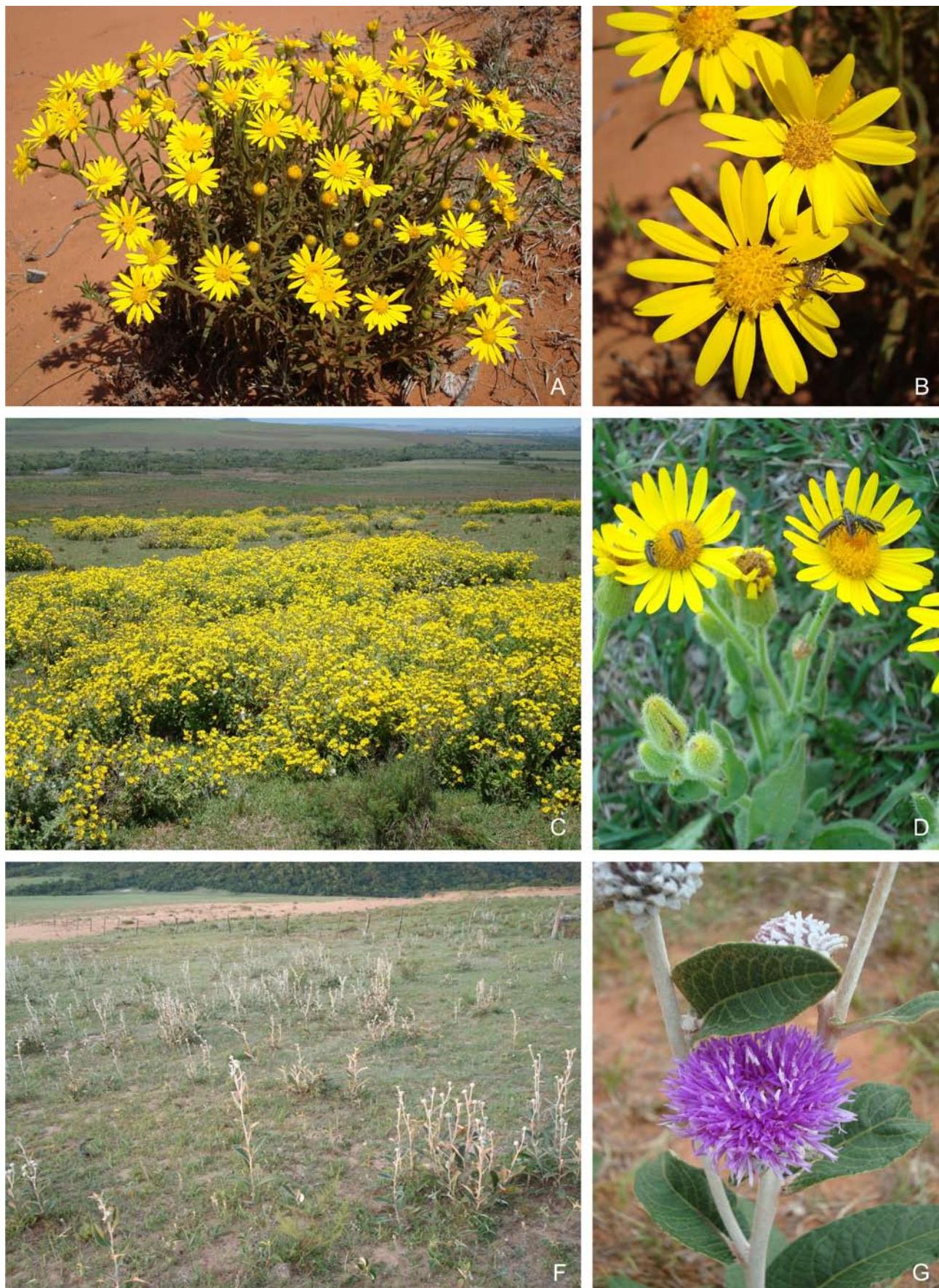


Figure 5. A, B. *Senecio cisplatinus*, occurring in the areas most severely hit by the sandy patch process. C, D. *Senecio selloi*, occurring in grassland portions still untouched by the sandy patch process. E. *Vernonia macrocephala*, represented by a high number of individuals in a grassland area at São Francisco de Assis municipality. F. *V. macrocephala* inflorescence.

municipality, Taquari district, Passo do Nagel locality.

Eragrostis articulata and *Eragrostis leucosticta* appear as new records for RS state. *E. articulata*, an annual cespitose grass is recorded for Argentina, Bolivia, Brazil and Paraguay. In Brazil it is recorded for many states and so that its southernmost record is for Paraná state. The rocky fields and the cerrado are the predominant habitats for this species (Boechat & Longhi-Wagner

2001). In the study area, it was found in dry and sandy grasslands undergoing sandy patch process, at Manoel Viana municipality, Lajeado locality. *E. leucosticta*, a cespitose perennial grass, has occurrence records for many states to Brazil, as a typical species of the cerrado biome (Boechat & Longhi-Wagner 2001). In the study area, it was collected in dry and sandy grasslands undergoing sandy patch process, at Alegrete municipality, Cerro do Tigre locality.



Figure 6. A. *Hexachlamys humilis*. B. *Eugenia arenosa*. C. *E. arenosa* individuals in a grassland areas undergoing sandy patch process at Jacaquá, São Francisco de Assis municipality. D. *Eugenia pitanga*. E. *E. plurisepala*. F. Grassland area with sandy patch process in Jacaquá, São Francisco de Assis, with specimens of *E. pitanga*, *E. arenosa* and *H. humilis*.



Figure 7. Species with potential attributes for recovering grassland areas undergoing sandy patch process. A. *Acanthospermum australe* on exposed sand. B. *A. australis* in the edge of a sandy patch at São Francisco de Assis municipality. C. *Lupinus albescens* (habit). D. *L. albescens* inflorescence. E, F. *L. albescens* individuals in a grassland area undergoing sandy patch process at Cerro do Tigre locality, Alegrete municipality.



Figure 8. A. *Cerastium commersonianum*, exotic species naturalized in Brazil. B. *Croton lorentzii*, new record for Brazil, found at São Francisco de Assis municipality, Taquari district, Passo do Nagel locality. C. *Froelichia tomentosa*, cited as extinction threatened, frequently observed in the study area. D. *F. tomentosa* (inflorescence). E. *Butia lallemandii*, exclusive to the sandy grasslands found in the southwestern RS state. F. *Baccharis multifolia*, exclusive to the sandy grasslands found in the southwestern RS state.

Table 2. Extinction-threatened species of Rio Grande do Sul state with records for grassland areas undergoing sandy patch process at Alegrete, Manoel Viana and São Francisco de Assis municipalities.

Family/Species	Threat rank
AMARANTHACEAE	
<i>Alternanthera hirtula</i> (Mart.) R.E. Fr.	Endangered
<i>Alternanthera praelonga</i> A. St.-Hil.	Critically endangered
<i>Froelichia tomentosa</i> (Mart.) Moq.	Critically endangered*
<i>Gomphrena graminea</i> Moq.	Vulnerable
<i>Gomphrena perennis</i> L.	Vulnerable
<i>Pfaffia gnaphaloides</i> (L. f.) Mart.	Vulnerable
ASTERACEAE	
<i>Eupatorium angusticeps</i> Malme	Probably extinct
<i>Gochnati cordata</i> Less.	Endangered
<i>Trixis pallida</i> Less.	Endangered
CACTACEAE	
<i>Echinopsis oxygona</i> (Link) Zucc.	Vulnerable
<i>Parodia ottonis</i> (Lehmann) N. P. Taylor	Vulnerable
LAMIACEAE	
<i>Hesperozygis ringens</i> Benth.	Endangered
MALVACEAE	
<i>Hochreutinera hasslerana</i> (Hochr.) Krapov.	Vulnerable
<i>Waltheria douradinha</i> A. St.-Hil.	Vulnerable
MORACEAE	
<i>Dorstenia brasiliensis</i> Lam.	Vulnerable
MYRTACEAE	
<i>Eugenia arenosa</i> Mattos	Endangered

* Species not cited in the Final List of the Threatened Flora – RS (State decree nº 42.099; December 31st 2002).

Threatened and restricted species

Fifteen taxa found in the study area (Table 2) are included in the Final List of the Threatened Flora – RS (State decree nº 42.099; December 31st 2002).: eight are categorized as ‘vulnerable’, five as ‘endangered’, one as ‘critically endangered’ (*Alternanthera praelonga*) and one as ‘probably extinct’ (*Eupatorium angusticeps*) (SEMA 2003). *Eupatorium angusticeps* was found in only one grassland area, in a population with few individuals. Beside these species, *Froelichia tomentosa* (Fig. 8C, 8D) was cited by Marchiorotto et al. (2005) as ‘critically endangered’. However, this species was frequently observed in all the areas studied, chiefly in grassland areas heavily affected by the sandy patch process.

From all the species, 20 show a restricted distributional area (Table 3). Of these, seven are exclusive to the sandy grasslands found in the southwestern RS state: *Butia lallemantii* (Fig. 8E), *Achyrocline marchiorii*, *Baccharis albolanosa*, *B. multifolia* (Fig. 8F), *B. riograndensis*, *Senecio riograndensis* and *Hesperozygis ringens*. Among the families with higher number of restricted species, Asteraceae stands out with 11 species.

Butia lallemantii, also known as ‘butiá-anão’ due to its reduced height, is among the species that stand out in sandy patches. The species occurs only in the sandy grasslands of southwestern RS, where it usually constitutes populations with large number of individuals, standing out in the landscape (Fig. 9A) (Deble & Marchiori 2006). It was observed in several grassland areas among the surveyed municipalities, where we also found strong interaction between the species and the local fauna.

Achyrocline marchiorii is mentioned as endemic to the ‘Campanha Gaúcha’ region (the southwest of RS state), where it dwells in sandy and rocky grasslands (Deble 2005). In the study area, it was found only at Cerro o Tigre farm, Alegrete municipality, the same place where the species’ type was collected.

Baccharis albolanosa, to the present moment, has occurrence records only for the sandy grasslands of the southwestern RS state, at Manoel Viana and São Francisco de Assis municipalities (Oliveira & Deble 2006). In the grassland areas surveyed during the present study, the species was collected only in an area located near the state road RS 377, km 351.

Baccharis multifolia is another species that, until this moment, is mentioned only for the southwest region of RS state, in sandy grassland areas (Oliveira & Marchiori 2006). The species was observed at Alegrete municipality, Cerro do Tigre locality, and at São Francisco de Assis municipality, Joaquim Paz farm, where its population is numerous, dominating all the grassland area during the blooming phase (Fig. 9C).

Baccharis riograndensis is considered endemic to RS, with occurrence recorded for dry or rocky grasslands, notably in the southernmost half of the state (Heiden & Schneider 2008). In the study area, the species was found in a grassland undergoing sandy patch process at São Francisco de Assis municipality, Joaquim Paz farm.

Senecio riograndensis is recorded only for the RS state, in coastal grasslands, Central Depression, Campanha and Planalto Médio regions (Matzenbacher 1996). Only a small population was found in a grassland area undergoing sandy patch process at São Francisco de Assis municipality, near Cerro da Esquina locality.

Table 3. Restricted species registered for grassland areas in southwestern RS State.

Family/Species	Geographic distribution
APIACEAE <i>Eryngium ciliatum</i>	Uruguay and dry grasslands of RS state (Central Depression, Missões and Campanha regions).
ARECACEAE <i>Butia lallemantii</i>	Exclusive to sandy soil grasslands of southwestern RS.
ASTERACEAE <i>Achyrocline marchiorii</i>	Endemic to sandy and rocky grasslands of the Campanha Gaúcha region.
<i>Asteropsis megapotamica</i>	Sandy and rocky grasslands of RS state and Uruguay.
<i>Baccharis albolanosa</i>	Sandy soil grasslands of southwestern RS.
<i>Baccharis multifolia</i>	Sandy soil grasslands of southwestern RS.
<i>Baccharis riograndensis</i>	Endemic to RS, in dry or rocky grasslands, especially in the state's southernmost half.
<i>Gochnativa cordata</i>	RS, in sandy soil grasslands, Uruguay and NE Argentina up to Entre Ríos province.
<i>Hysterionica filiformis</i>	Southern Brazil, Uruguay and NE Argentina.
<i>Noticastrum acuminatum</i>	Sandy soil grasslands and coastal dunes in southern Brazil, Uruguay and NE Argentina.
<i>Noticastrum diffusum</i>	Dry and rocky grasslands of southern Brazil, Uruguay and central/NE Argentina.
<i>Senecio cisplatinus</i>	Sandy patches in RS, Uruguay and NE Argentina.
<i>Senecio riograndensis</i>	Endemic to RS, in Coastal grasslands, Central Depression, Campanha and Planalto Médio regions.
CONVOLVULACEAE <i>Ipomoea malvaeoides</i> var. <i>lineariloba</i>	Dry grasslands of RS, Paraguay, Argentina and Uruguay.
<i>Ipomoea nitida</i>	RS (Missões and Campanha regions) and Argentina.
EUPHORBIACEAE <i>Croton lorentzii</i>	Central/NE Argentina and Uruguay. In Brazil, is found in southwestern RS.
FABACEAE <i>Lupinus albescens</i>	Uruguay (W), Argentina and Paraguay. In Brazil, only in RS (Litoral, Campanha and Missões regions).
LAMIACEAE <i>Hesperozygis ringens</i>	Endemic to the southern grasslands of RS (Serra do Sudeste and Missões regions).
MYRTACEAE <i>Myrcia verticillaris</i>	Clean and stony grasslands of Uruguay and RS (Campanha, Central Depression, Missões and Serra do Sudeste regions).
POACEAE <i>Pappophorum macrospermum</i>	Sandy soil grasslands of Central/N Uruguay, southern RS and NE Argentina.

Besides being present in the threatened flora list, *Hesperozygis ringens* (Fig. 9B) is endemic to the grasslands of RS, with distribution in the Serra do Sudeste and the south of the Missões, in sandy and rocky soils at Caçapava do Sul, Alegrete and São Francisco de Assis municipalities. Successive cleaning and burning of pastures, associated with the low gene flow between populations, led the species to the threatened list (Fracaro 2006). In the study area, only two populations were observed, and they were in greatly separated grassland areas (ca. 30 km from each other), which may complicate or even block gene flow between them.

The remaining species considered restricted (Tab. 3) reach grassland areas in RS state, Argentina, Uruguay and, occasionally, Paraguay. Among them, *Asteropsis megapotamica* (Fig. 9D, 9E) is restricted to the sandy soil grasslands of RS and rocky grasslands of Uruguay, where it dwells between scattered shrubs and *Butia paraguayensis* (Barb. Rodr.) L.H. Bailey (Bonifacino *et al.* 2009). In the study site, it was found in sandy soils at Manoel Viana municipality, among shrubs, cespitous grasses and *Butia lallemantii*. *Gochnativa cordata* grows on sandy soils in RS state, Paraguay, Uruguay and NE Argentina as far as Entre Ríos province (Burkart 1974). Sobral (2003) mentioned *Myrcia verticillaris* as restricted to the clean and stony grasslands of Uruguai and RS state. *Eryngium ciliatum* is described as endemic to Uruguay and to the dry and graminous grasslands of RS,

in the Depressão Central, Missões e Campanha regions (Rambo 1957, Irgang 1974). *Ipomoea malvaeoides* var. *lineariloba* (Fig. 9F), restricted to sandy grasslands in RS, also has registers for Paraguay, Argentina and Uruguay. On the other hand, *Ipomoea nitida* occurs in Argentina and Brazil, being restricted to RS state in the latter (Ferreira 2009). In our survey, both species of *Ipomoea* were found at only one location. *Pappophorum macrospermum* is restricted to the sandy grasslands of northern Uruguay, southern RS and northeastern Argentina (Corrientes province) (Pensiero 1986).

The grasslands from southwestern RS have a peculiar flora with significant number of species, including several threatened and endemic species. In addition, many of them present morpho-physiological adaptations that allow their survival in the extreme local environmental conditions.

ACKNOWLEDGEMENTS

Our thanks to Embrapa Pecuária Sul – Bagé, for funding our research. To Dr. José Pedro Pereira Trindade for support in the search for financial resources and in contacting the local farmers. To colleagues and friends that helped in the field expeditions. To the friends and colleagues Rafael Rodrigo Eckhardt and Úrsula Arend, for map and figures designing. To specialists that helped in the identification of some species. To the local farmers that gently allowed access to their properties.



Figure 9. A. *Butia lallemantii* specimens in a sandy patch at Manoel Viana municipality. B. *Hesperozygis ringens*, endemic to the grasslands of RS. C. Grassland area at São Francisco de Assis municipality with a large population of *Baccharis multifolia*. Species with restrict occurrence: D. *Asteropsis megapotamica* (habit). E. *A. megapotamica* (inflorescence). F. *Ipomoea malvaeoides* var. *lineariloba*.

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