Title: Strontium substituted hydroxyapatite promotes primary human osteoblast maturation

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Abstract

Strontium-substituted hydroxyapatite (SrHAp) materials are known to actively promote bone formation. However, the optimum level of Sr inclusion needed to elicit a physiologically relevant response from bone cells is unclear and can vary dependent on the fabrication process employed. In this work hydroxyapatite (HAp), SrHAp powders (2, 5 and 10 wt% (i.e., 1, 2 and 5 at%) with respect to [Sr/(Sr+Ca) ·100]), were synthesized with the purity and Sr-substitution was confined in the range of 1-8 wt% (1-4 at%). All SrHAp samples contained rod-like crystals (<106nm in length), which decreased in length with increasing Sr content, and exhibited larger flatter crystals (>300nm in length). TEM-EDX confirmed the presence of Sr and maintenance of the HAp lattice structure for both types of crystals. Qualitative *in vitro* evaluation using primary human ostesoblast cells (HOBC) cultured in contact with the SrHAp over 28 days showed that the presence of Sr (in particular with the highest Sr content) directly promotes the maturation of osteoblasts into osteocytes as compared to the response observed for HAp. As these materials contain no additives other than Sr, the effects observed here can only be attributed to the physiologically important levels of Sr in the samples.

Keywords: strontium substitution, hydroxyapatite, osteoblast, osteocyte, collagen I

1. Introduction

Effective bone regeneration remains a significant clinical challenge and advantageous strategies to enhance bone defect healing are still urgently needed. Hydroxyapatite (HAp) has been widely used in the biomedical fields in applications such as implant coatings, fillers for bone substitutes and as drug carriers for controlled release. Its widespread application is due to its biocompatibility, bioactivity and similarity to bone mineral component [1]. Today bone graft materials are expected not only to facilitate the growth of as replacement tissue after disease, injury, or failure due to ageing, but also to provide for the delivery of biologically active substances, which act against the causative disease of bone loss [2]. Extensive work has been carried out on the development of biomaterials combined with components that specifically promote bone growth, e.g. growth factors and bone morphogenetic proteins. However, the need for high concentrations to maintain a therapeutic efficiency increases costs of the therapy and generates serious concerns over long-term complications associated with the high dosages of the biological molecules [3]. Thus, the substitution of other more biologically active ions such as Na⁺, Mg²⁺, K⁺, Sr²⁺, Zn²⁺, Ba²⁺, Al³⁺ etc. for Ca²⁺ into bone graft materials, including HAp, provides a feasible solution for bone defect healing [4]-[6]. Substitutions for PO_4^{3-} and OH^- ions are also possible, for example silicon (SiO₄⁴⁻) and carbonate (CO₃²⁻) [7]. The advantages of such a strategy are lower costs, greater simplicity, higher stability, and more efficacies at low concentrations [8]. Strontium (Sr) is a unique biologically active trace element that simultaneously stimulates bone formation while inhibiting bone resorption [9]. It has been reported that Sr increases pre-osteoblast proliferation, osteoblast differentiation, type I collagen synthesis and bone matrix mineralization and inhibits osteoclast differentiation and activation [9], [10].

Typically, Sr is found in bone at very low concentrations of between 0.008-0.01 at% but can be found at concentrations of between 3 and 7 at% where bone is actively metabolising [7], [11]. The latest advances in Sr based regenerative therapies have been reviewed by Jimenez *et al* [10]. Given the positive biological properties of Sr in bone remodelling, it has been used in many studies as an additive to various biomaterials, including bioactive glasses [12], [13] and calcium phosphates (CaP) [12], [14]. Sr incorporation into different CaP-based biomaterials are highly promising for providing appropriate support to a defective site and delivering biologically active components to encourage bone tissue regeneration. Moreover, in vivo studies showed that Srdoped CaP cements and bioceramic materials promote bone regeneration [15], [16]. Previously, the drug strontium ranelate (SrRan) was used to treat osteoporosis, however, it was subsequently withdrawn from use in 2017 in Europe due to concerns it increased the chances of myocardial infarction in patients [17]. As such, there is a clear need to provide more localised delivery of Sr into bone defects, to administer the required therapeutic effect. Further to this, there is still disagreement in the literature as to the appropriate (and minimum) concentration of Sr to be effective without provoking any negative consequences [18], [19].

The aim of this work was to manufacture a range of Sr-substituted HAp materials with physiologically relevant levels of Sr and to study their effect on primary human osteoblast cells (HOBC). Specifically, the work has focused on low concentrations of Sr substitution of the HAp to determine an effective minimum dosage to inform future work. The physicochemical properties and level of ionic substitution of each sample were compared using atomic absorption spectrometry (AAS), Fourier transform infrared spectroscopy (FTIR), X-ray photoelectron spectroscopy (XPS), X-ray diffraction (XRD), scanning electron microscopy, energy dispersive X-Ray (EDX) analysis and transmission electron microscopy (TEM). Studies have shown that

biocompatibility is very dependent on the cell type involved in the tests. This work utilizes primary human osteoblasts cells (HOBC) which provide more clinically relevant results and brings the translational process closer as the interspecies differences are circumvented [20].

2. Materials and methods

2.1. Preparation

HAp powders with 4 different Sr concentrations, namely, pure HAp (HAp), HAp with 2 wt% Sr in respect to total content of cations, namely, Ca and Sr (2SrHAp), HAp with 5 wt% Sr (5SrHAp) and HAp with 10 wt% Sr (10SrHAp) were synthesised via aqueous precipitation described in detail previously [21]. Briefly, the main steps are shown in Figure 1.

The powders were uniaxially pressed at 10 kN into cylindrical moulds using 0.3 g of powder per disk (13 mm x 1 mm) and were sintered at 600 °C for 2 h using heating rate at 2 °C/min up to 600 °C, followed by cooling rate at 2 °C/min to room temperature.

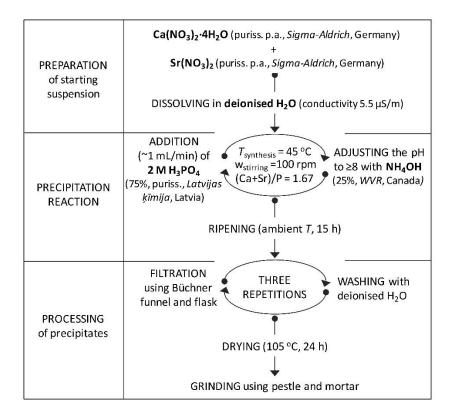


Figure 1. The main steps and technological parameters of the HAp powders synthesis.

2.2. Physico-chemical characterization

Atomic absorption spectrometry (AAS, Varian SpektrAA 880, Australia) was performed using electrothermal atomization according to the LVS ISO 11047:1998 standard test method.

X-ray photoelectron spectroscopy (XPS) was performed using an AXIS Ultra delay-line detector (DLD) spectrometer (Kratos, UK). Spectra were analysed using monochromated Al Kα X-rays (hv=1486.6 eV) operating at 10 kV and 15 mA (150 W). Wide energy survey scans (WESS) were recorded at a pass energy of 160 eV. High resolution spectra for C1s, O1s, Ca2p, P2p/Sr3d, Sr3p and P2s were recorded at pass energy of 40 eV. A Kratos charge neutraliser system was used for all samples. Sample charging effects on the calculated binding energy (BE) positions were adjusted by setting the lowest BE component of the C1s spectral envelope to 285.0 eV.

Photoelectron spectra were further processed by subtracting a Tougaard background and using the peak area for the most intense spectral line of each detected elemental species to determine the atomic%. In the case of the Sr-substituted samples, the P2p and Sr3d peaks overlap at 133/134 eV therefore, for these samples the P2s and Sr3p peaks were used in order to provide the appropriate quantification results. In total 3 areas were analysed from each sample.

Scanning electron microscopy (SEM) with energy dispersive X-ray (EDX) analysis was performed using a Hitachi SU5000 field emission instrument equipped with an X-Max^N 80 mm² silicon drift detector (Oxford Instruments, UK). For SEM and EDX analysis, an accelerating voltage of 10 kV was used. To make the surface of the samples conductive for the electron microscopy, they were coated with a thin layer (20 nm) of Au/Pd (60/40 ratio). The coating was performed using an Emitech K500X sputtering deposition with Argon gas.

Fourier transform infrared (FTIR) spectra were recorded using spectrometer (Varian 800 FT-IR, Scimitar Series, USA) in attenuated total reflectance (ATR, GladiATRTM, Pike technologies, USA) mode. The FTIR spectra were obtained at 4 cm⁻¹ resolution co-adding 50 scans over a range of wavenumbers from 400 cm⁻¹ to 4000 cm⁻¹. Before every measurement a background spectrum was taken (50 scans) and deducted from the sample spectrum.

X-Ray diffraction (XRD) analysis were performed using system (X'Pert PRO, PANalytical, the Netherlands) operating at 40 KV and 30 mA using a Cu K α_1 radiation (λ =1.5406 Å). Patterns were obtained over the range of 2-Theta from 10° to 70° with an angular step interval of 0.0334°. The X'Pert Data Viewer and X'Pert HighScore software were used for processing and analyzing the XRD data. For the phase identification the International Centre for Diffraction Data (ICDD) was used (PDF-2/2005 card #01-072-1243 for HAp). Computer structural analysis of XRD diffractograms was performed by the *Rietveld* method using the software Profex-BGMN-Bundle3.8.0 [22]. Rietveld's refinement for HAp was done based on data from with the hexagonal space group P63/m using PDF card #01-074-0565 as structural model for HAp. R_{wp} , R_{exp} and X^2 for refinements indicate the quality of the fits. The observed, calculated and differential patterns are shown in Figure 2.

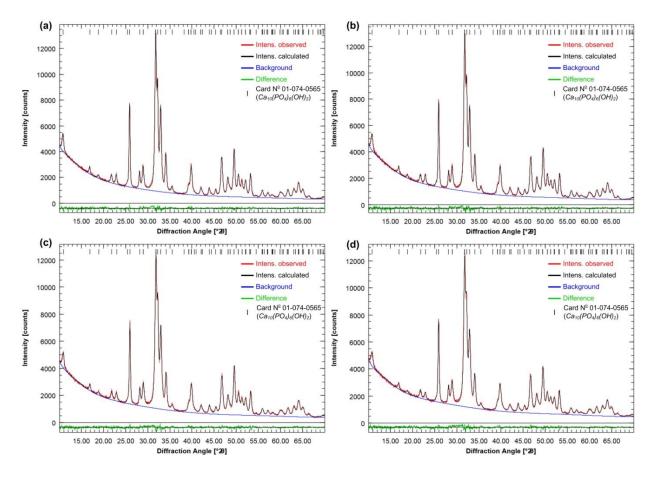


Figure 2. Observed (red), calculated (black) and differential (green) patterns for (a) HAp, (b) 2SrHAp, (c) 5SrHAp and (d) 10SrHAp calcined at 600 °C for 2 h.

Transmission electron microscopy (TEM) observations were made using a JEOL 2100F microscope operating at 200 kV bias voltage. The samples for TEM analysis were prepared by roughly grinding the sintered disks followed by sonication in ethanol. The sonicated solutions were drop-casted on a Cu grid with 3 nm carbon films and the images were recorded using a charged

coupled device detector. Scanning transmission electron microscopy-energy dispersive X-ray analysis (STEM-EDX) was performed using Oxford instruments X-Max detector coupled with the same JOEL 2100F system at 1.5 nm spot size. The STEM images were recorded using a JEOL high-angle annular dark field (HAADF) detector. ImageJ software was utilised to measure the dimensions of 60 nanocrystals from each sample. The results are reported as the mean \pm standard deviation (SD).

2.3. *In vitro* cell studies

2.3.1. Cytoskeletal and collagen I

Primary human osteoblast cells (HOBC) were isolated and characterised using previously described method [23]. The SrHAp and HAp control samples were seeded with 2x10⁵ cells/well in 1 mL medium (Dulbecco's modified eagle medium (DMEM) + 5% fetal calf serum (FCS), Sigma-Aldrich, UK) for 1, 7 and 28 days. Fresh medium was added twice a week. At the predesignated time points (1, 7 and 28 days), the cell culture supernatant was removed from the wells and samples were gently washed once with phosphate-buffered saline (PBS) without Ca²⁺ and Mg²⁺ at room temperature. Samples were then fixed with 4% formaldehyde, 2% sucrose solution in PBS and incubated for 15 min at 37 °C, 5% CO₂ followed by further washing. Fixed samples were permeabilized with 0.05% Triton X100 solution (Sigma-Aldrich, UK) for 5 min at 4 °C, followed by washing with PBS for 5 min at 37 °C. Samples were incubated with the collagen I primary antibody (1:500 dilution with 1% bovine serum albumin (BSA):PBS, Sigma-Aldrich, UK) overnight at 4 °C, washed with 0.1% Tween 20 solution (ICN Biomedicals, USA) for 5 min at room temperature, followed by incubation with the Rhodamine conjugated goat IgG fraction to mouse IgG (Invitrogen, USA) secondary antibody at a concentration of 1 µg/mL in 1% BSA:PBS for 1 h at 37 °C. The samples were washed with Tween 20 wash solution and incubated with 5 μg/mL Oregon-green phalloidin (Molecular Probes, USA) for 30 min at 4 °C. The samples were then placed onto glass slides and mounted with a fluorescence stabilising mounting medium with DAPI (Vectashield, Vector Laboratories Inc., UK). The samples were viewed immediately by confocal laser scanning microscopy (Zeiss 510, UK).

2.3.2. Alkaline phosphatase enzymatic activity assay

At each experimental endpoint (1, 7 and 28 days), the supernatant was removed from the wells. The cells were washed twice in ice-cold PBS and lysed in 0.1% Triton-X. 100 μ L of the lysate was transferred to a 96-well plate, 100 μ L of para-Nitrophenylphosphate (p-NPP) solution was added to the sample, allowed the reaction to take place at 37 °C for 40 min, and then 50 μ L of 3 M NaOH was added to terminate the reaction. The absorbance at 405 nM within 15 min of terminating the reaction was read. The values obtained were normalized to the control measurements and expressed as a percentage of change from control, i.e., alkaline phosphatase (ALP) activity on the HAp samples at day 1. Results are expressed as an average value of the three assessments per series of samples \pm SD. The differences were tested for statistical significance using a one-way ANOVA followed by Tukey post hoc test for multiple comparisons.

3. **Results**

3.1. Physico-chemical characterization

The chemical composition of all the samples was determined using AAS, EDX and XPS. The nominal and experimental amounts of Sr measured by AAS were close to the nominal values (Table 1). This indicates that no significant Sr losses have occurred during the synthesis. EDX of the pressed HAp disk gave a stoichiometry of 1.67 ± 0.02 , equivalent to that of pure HAp. The corresponding Ca/P ratios of the Sr-substituted HAp materials were higher than expected, with values reported between 1.74 ± 0.01 and 1.76 ± 0.02 , as shown in Table 2 and Figure 3. The corresponding (Ca+Sr)/P ratios were shown to be slightly higher than the Ca/P ratio, with the Sr/(Ca+Sr) atomic ratios increasing with the increasing levels of Sr in the samples. The additional EDX peaks correspond to either Au/Pd coating or adventitious carbon on the surface of the samples (with peaks at 0.28 KeV (C K_a), 2.12 keV (Au M_a) and 2.83 keV (Pd L_a)). No other impurities were detected from the EDX analysis.

Sample		Nominal Sr COI		nental Sr itent AAS)	Nominal Sr/(Ca + Sr)		Experimental Sr/(Ca + Sr)	
	wt%	at%	wt%	at%	atomic ratio	weight ratio	atomic ratio	weight ratio
НАр	-	-	-	-	-	_	-	-
2SrHAp	0.78	9.00	0.56 ± 0.06	6.44 ± 0.69	0.01	0.02	0.01 ± 0.00	0.01 ± 0.00
5SrHAp	2.07	24.00	1.41 ± 0.14	16.29 ± 1.62	0.02	0.05	0.02 ± 0.01	0.04 ± 0.01
10SrHAp	4.09	48.00	3.10 ± 0.30	36.15 ± 3.50	0.05	0.10	0.04 ± 0.01	0.08 ± 0.01

Table 1. Nominal and AAS quantification results from the powders calcined at 600 °C for 2 h.

Table 2. EDX quantification results from the surfaces of the discs sintered at 600 °C for 2 h.

Sample	Ca/P	(Ca+Sr)/P	Sr/(Ca+Sr)	
НАр	1.67 ± 0.02	1.67 ± 0.02	-	
2SrHAp	1.75 ± 0.02	1.77 ± 0.02	0.01 ± 0.01	
5SrHAp	1.76 ± 0.02	1.81 ± 0.03	0.02 ± 0.01	
10SrHAp	1.74 ± 0.01	1.81 ± 0.01	0.04 ± 0.01	

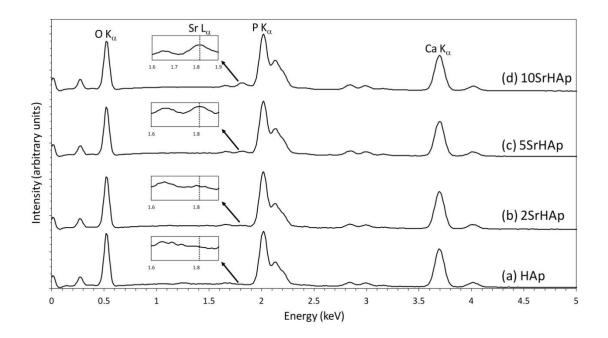


Figure 3. EDX spectra for (a) HAp, (b) 2SrHAp, (c) 5SrHAp, (d) 10SrHAp discs sintered at 600 °C for 2 h.

Figure 4 show typical XPS WESS for (a) HAp and (b) 10SrHAp. For the HAp sample, all the expected peaks for O1s, C1s Ca2p, P2p and P2s were detected, as shown in Table 4. However, the Ca/P ratio was slightly lower than the expected 1.67 for stoichiometric HAp, at 1.60 ± 0.03 as highlighted in Table 3. For all three Sr-substituted samples, the WESS showed the presence of Sr, as evidenced by the Sr3p_{3/2} peak around 269.0eV (Table 4 and Figures 4(b) for the 10SrHAp sample). The corresponding high resolution Sr3p_{3/2} peaks, as shown in Figures 5(d, g and j) for 2SrHAp, 5SrHAp and 10SrHAp, respectively, confirm the presence and successful substitution of Sr for Ca within each substituted material. No CO₃²⁻ peaks were detected in the C1s spectral envelope (typically observed around 289.0-290.0 eV) [24]. The Ca/P ratios for the Sr-substituted samples were calculated at 1.44 ± 0.07 for 2SrHAp, 1.41 ± 0.05 for 5SrHAp and 1.29 ± 0.06 for 10SrHAp (as shown in Table 3), all of which are lower than 1.67 for stoichiometric HAp. As all

three Sr-substituted samples contain Sr (as determined by AAS, EDX and XPS), it should be noted that the spectral envelope observed around 133.4-133.8 eV comprises an overlap of the Sr3d and P2p peaks as the $Sr3d_{5/2}$ (133 ± 0.5 eV), $Sr3d_{3/2}$ (135 ± 0.5 eV) and the P2p (132-133 eV) lines are positioned very close together. Therefore, in order to quantify the P and Sr at% values in the Sr-substituted samples, the P2s and Sr3p peaks are used instead of the P2p and Sr3d peaks. The main peaks recorded for all Sr-substituted samples shown in Table 4 were therefore Ca2p_{3/2} (347.5-347.7 eV), P2s (190.9-190.2 eV), O1s (530.4 eV) and Sr3p_{3/2} (269.2-269.7 eV).

The (Ca+Sr)/P ratios for each Sr-substituted sample determined by XPS (see Table 3), are as follows: 1.45 ± 0.07 for 2SrHAp, 1.43 ± 0.05 for 5SrHAp and 1.35 ± 0.06 for 10SrHAp. EDX analyse approximately 1 μ m depth of the disk samples, whereas XPS only analyse the top 5-10 nm of the disks sample surface [25]. There is a high probability that the total area of analysis in XPS and EDS is different which gives rise to the difference in Ca/P and (Ca+Sr)/P ratios observed. However, it can be seen that for each sample the ratio is slightly larger for (Ca+Sr)/P in comparison to the Ca/P ratios alone, highlighting that for each Sr-substituted sample Sr has been successfully substituted for Ca within HAp. The corresponding Sr/(Ca+Sr) ratios found in Table 3 are interesting as they show a clear increase in Sr substitution, therefore, 0.01 ± 0.00 for 2SrHAp, 0.02 \pm 0.01 for 5SrHAp, and 0.04 \pm 0.01 for 10SrHAp. This data indicates that as the amount of Sr is increased in the following sequence 2SrHAp→5SrHAp→10SrHAp as highlighted by AAS in Table 1 for each Sr-substituted samples, the subsequent percentage ratios of atomic concentration also increase accordingly, confirming that as a larger percentage concentration of Sr is added at the beginning of the reaction, a larger amount of Sr is substituted for Ca within the SrHAp sample. It should be noted that the Sr/(Ca+Sr) values as determined by XPS correlate strongly with the Sr/(Ca+Sr) results from the AAS and the EDX, as shown in Table 1 (AAS), Table 2 (EDX) and

Table 3 (XPS). No impurities were detected from the XPS analysis. This suggests that the aqueous precipitation synthesis employed here successfully substitutes Sr within the HAp lattice as amounts of the Sr source are increased.

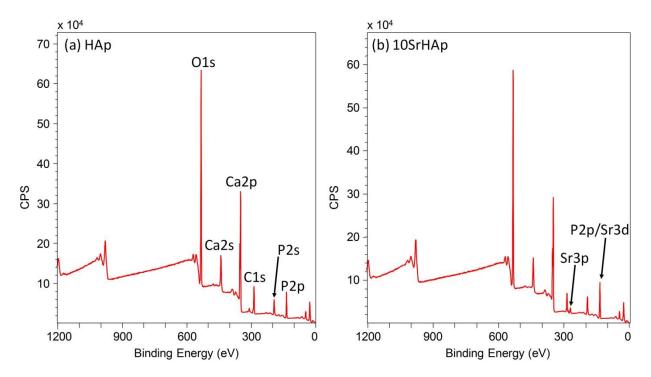


Figure 4. XPS WESS of (a) HAp and (b) 10SrHAp discs sintered at 600 °C for 2 h.

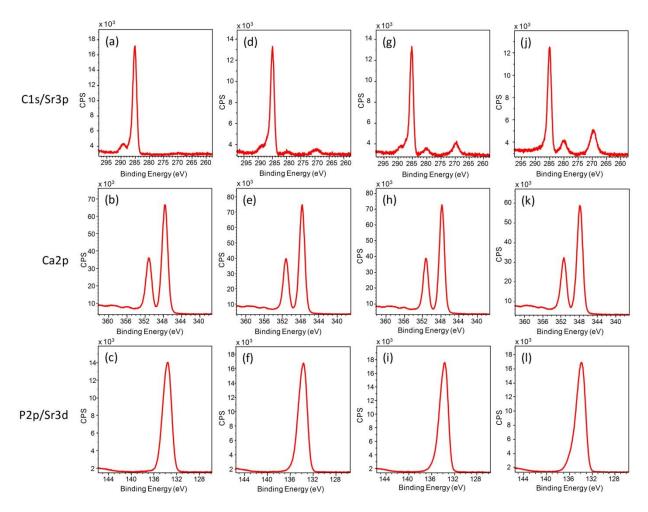


Figure 5. XPS of (a-c) HAp, (d-f) 2SrHAp, (g-i) 5SrHAp, (j-l) 10SrHAp discs sintered at 600 °C

for 2 h.

Table 3. XPS quantification results from the surface of the discs sintered at 600 °C for 2 h.

Sample	Ca/P	(Ca+Sr)/P	Sr/(Ca+Sr)
НАр	1.60 ± 0.03	-	-
2SrHAp	1.44 ± 0.07	1.45 ± 0.07	0.01 ± 0.00
5SrHAp	1.41 ± 0.05	1.43 ± 0.05	0.02 ± 0.01
10SrHAp	1.29 ± 0.06	1.35 ± 0.06	0.04 ± 0.01

Table 4. Peak positions (eV) as determined by XPS.

Sample	Peak Positions – BE (eV)					
Sample	C1s	O1s	Ca2p3/2	P2p/Sr3d5/2	5/2 P2s Sr3p3/2	
НАр	285.0	531.4	347.5	133.4	190.9	-

2SrHAp	285.0	531.4	347.5	132.8	190.9	269.3
5SrHAp	285.0	531.4	347.5	133.7	191.0	269.7
10SrHAp	285.0	531.4	347.7	133.8	191.2	269.2

A typical FTIR spectrum for SrHAp is shown in Figure 6 with absorbance bands at 1090, 1022, 963, 560, 600 and 472 cm⁻¹ attributed to phosphate [PO₄] vibrations and at 3569 and 631 cm⁻¹ attributed to characteristic [OH] bands [26]. The only difference observed in the spectra was that with an increase of Sr content, the intensity of the [OH] groups' absorption band at 631 cm⁻¹ decreased. This is in good agreement with previous studies suggesting that substituting Ca²⁺ ions with ionic radius 0.100 nm with larger Sr²⁺ ions with radius of 0.118 nm results in perturbation of the HAp lattice structure [27]. No CO₃²⁻ peaks (typically observed around 1550-1400 cm⁻¹ and 875 cm⁻¹) were detected in any of the samples from the FTIR analyses [28].

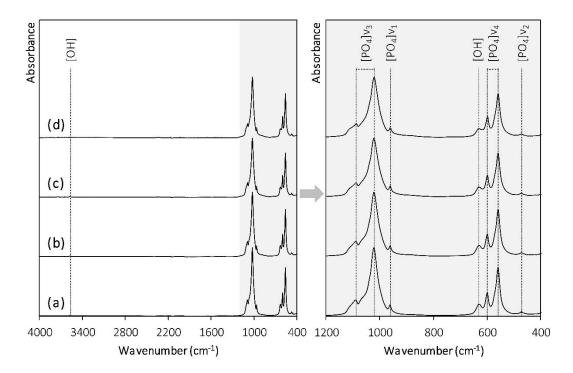
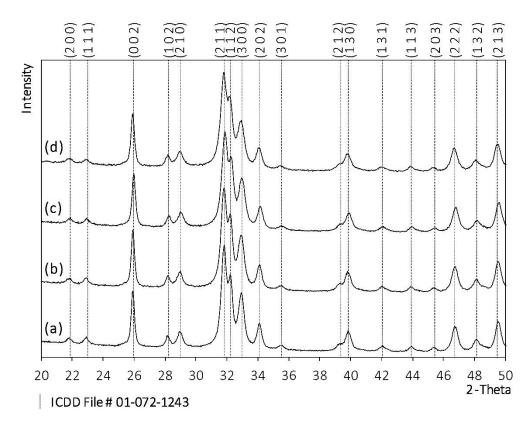


Figure 6. FTIR spectra of (a) HAp, (b) 2SrHAp, (c) 5SrHAp, (d) 10SrHAp calcined at 600 °C



The XRD patterns shown in Figure 7 indicate that regardless of the chemical composition all samples had a hexagonal lattice structure with space group P6₃/m typical to HAp [29]. Displacement of the peaks, which according to analogous studies would confirm the incorporation of Sr into the HAp structure [30], was observed for all the Sr substituted materials (Table 5). At the same time, peaks indicating the presence of other Sr-containing compounds were not detected. Slight broadening and merging of peaks as Sr content increases was observed, specifically, for peaks (211) and (112). The full width half maximum (FWHM) for the representative peaks are shown in Table 6. This is consistent with studies reporting that incorporation of Sr causes deformation of the HAp crystalline structure, which leads to a decrease in crystallinity [31]. The broadening of the peaks can also be a consequence of nano-sized crystals, i.e. reduction of the crystallite size [32]. Although the broadening and shift in the peak positions are very small, they are very consistent with the amount of Sr.



for 2 h.

Plane	НАр	2SrHAp	5SrHAp	10SrHAp
(002)	25.8961	25.8938	25.8839	25.8615
(211)	31.7668	31.7560	31.7530	31.7323
(130)	39.7873	39.7727	39.7520	39.7296
(222)	46.7074	46.6946	46.6795	46.6298

Table 5. 2θ Position (degrees) values for the peaks of various samples.

Table 6. FWHM (in degrees) values for the peaks of various samples as determined from the

Plane	2θ (degrees)	НАр	2SrHAp	5SrHAp	10SrHAp
(002)	25.8	0.2446	0.2540	0.2602	0.2660
(211)	31.7	0.4698	0.5045	0.5527	0.5739
(130)	39.7	0.5014	0.5393	0.5944	0.6021
(222)	46.7	0.4342	0.4435	0.4522	0.4634

TEM analyses.

The above observations regarding changes in crystallite size with increasing Sr content were confirmed by Rietveld analysis. Refined lattice parameters, crystallite size and *R*-values obtained for all samples are listed in Table 7. The low *R*-values confirm the successful and reliable performed refinements. The *c* and *a* axis parameters values show little effect with the different Sr content, i.e. slightly increases with increasing Sr content suggesting the larger radius ions entered the apatite lattice and induced the hexagonal lattice expansion. This is in a good agreement with the results obtained by Chen *et al.* [33]. All the samples have larger crystallite size along the [001] plane compared to that along the [100] plane, irrespective of the Sr content. In general, the size of crystallites decreases by increasing the Sr content in the hexagonal HAp structure.

		НАр	2SrHAp	5SrHAp	10SrHAp
Lattice parameters (Å)	a	9.42854	9.43273	9.43623	9.44355
Lature parameters (A)	С	6.88281	6.88329	6.88650	6.89267
Crystallite size (nm)	[001]	60.30 ± 2.50	61.60 ± 2.70	56.40 ± 2.20	56.20 ± 2.50
Ci ystainte size (iiii)	[100]	24.64 ± 0.37	22.98 ± 0.31	21.88 ± 0.28	19.82 ± 0.29
	R_{exp} (%)	2.44	2.38	2.36	2.27
<i>R</i> -Values (%)	R_{wp} (%)	3.20	3.23	3.18	3.07
	X^2	1.72	1.84	1.82	1.83

Table 7. Lattice parameters and *R*-values for HAp and SrHAp samples obtained by

Rietveld refinement.

The high-resolution TEM images shown in Figure 8 indicate that the average length of the HAp crystals is 106 ± 23 nm, with crystals measuring from 69 to 168 nm. In comparison, the average length of the Sr-substituted materials is on the nanoscale, with values reported for the 2SrHAp of 82 ± 22 nm (range of 44-157 nm), 5SrHAp of 67 ± 16 nm (range of 32-115 nm) and 10SrHAp of 51 \pm 11 nm (range of 28-88 nm). This decrease in the dimensions of the measured crystals with increasing Sr content would be in line with expectations, as Sr substitution of HAp inhibits crystal growth [34]. This is in accordance with the XRD observations where the broadening increases with the increased levels of Sr substitution. Further to this, for all the Srsubstituted samples, larger flatter crystals of varying dimensions are observed (typically up to 300 nm in length), which are not seen in the HAp sample. The SEM micrographs of the surface of the HAp and SrHAp disks corroborated the results of the TEM, as shown in Figure 9. Two distinct morphologies were observed, with predominantly small rod-like features observed for the HAp, with larger flatter features clearly shown for the 2SrHAp, 5SrHAp and 10SrHAp disk sample surfaces, as shown in Figures 9(b-d), respectively. In addition, as the Sr content increased, the amount of the larger flatter particles increased. Based on analogous studies, this could indicate that non-miscible phases of Ca-rich HAp and Sr-rich HAp have formed [35], [36]. However, the indepth STEM-EDX (as shown in Figure 10) and TEM (Figure 11) analysis confirmed the presence of Sr in both the rod-like crystals and the larger flatter crystals. STEM-EDX mapping of the distribution of Ca, P, O and Sr highlighted that these elements were homogeneously distributed across both the rod-like and larger flatter crystals, as observed in Figure 10(a). As shown in Figure 10(b), the peak ratio of Sr/Ca for both rod-shaped and flatter crystals were very similar indicating no major change in the Sr ratio in the two types of crystallites.

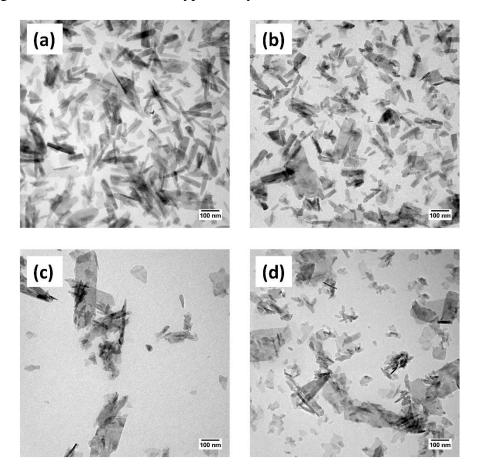


Figure 8. TEM micrographs of (a) HAp, (b) 2SrHAp, (c) 5SrHAp, (d) 10SrHAp calcined at 600

^oC for 2 h (scale bar 100 nm).

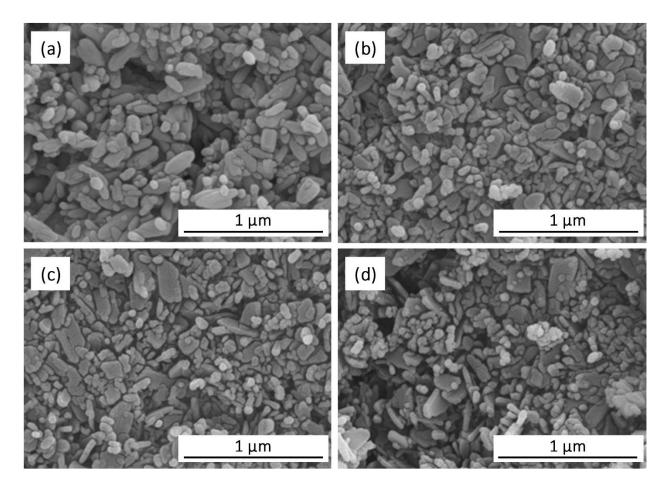


Figure 9. SEM surface micrographs of (a) HAp, (b) 2SrHAp, (c) 5SrHAp, (d) 10SrHAp discs sintered at 600 °C for 2 h.

Further detailed analyses of the TEM results clearly highlights that for the SrHAp samples both the rod-like crystals and flatter larger crystals observed (as shown in Figure 11) have lattice spacings corresponding to the (002) plane of HAp. This indicates that the flat regions, although not observed in pure HAp samples, have maintained the native lattice structure even upon the substitution of Ca^{2+} with Sr^{2+} . This concurs with the STEM-EDX observations highlighted in Figure 10. The selected area electron diffraction (SAED) patterns from the TEM analyses (Figure 12) show the different planes of the HAp and 10SrHAp samples, which correspond to those observed earlier in the XRD analyses (Figure 7).

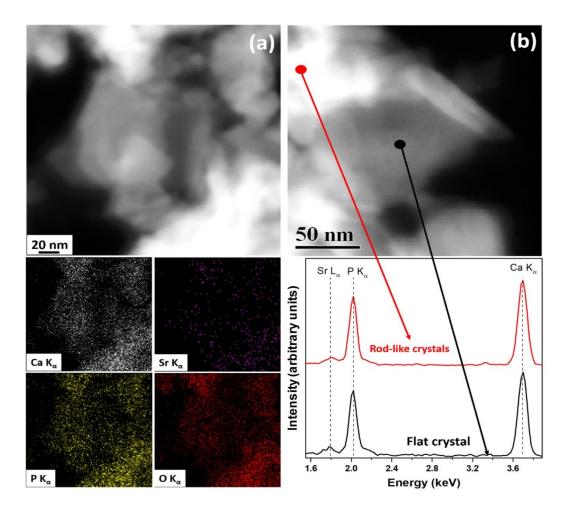


Figure 10. (a) STEM-EDX results of 10SrHAp calcined at 600 °C for 2 h showing the presence of Sr in both flat and rod-like crystals and (b) shows the EDX spectra from points on flatter and rod-like crystals.

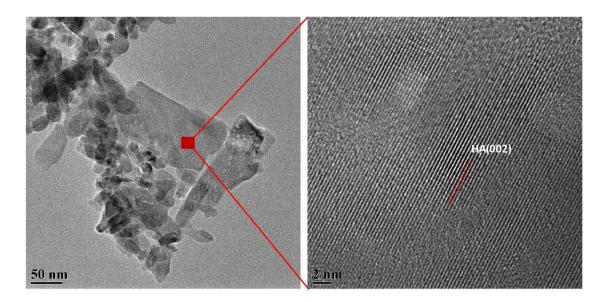


Figure 11. TEM image (left - scale bar 50 nm) and corresponding high-resolution TEM image (right - scale bar 2 nm) from 10SrHAp samples.

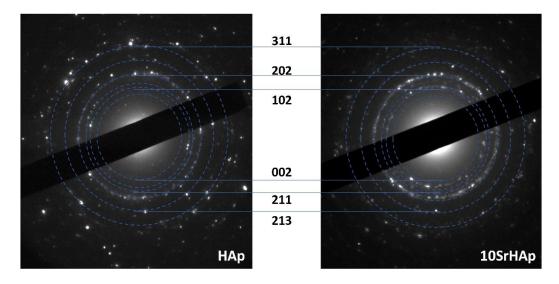


Figure 12. SAED pattern from (left) HAp and (right) 10SrHAp. The diffraction rings corresponding to various planes (miller indices) are highlighted in the centre of the Figure.

3.2. In vitro characterization

Qualitative *in vitro* evaluation (Figure 13) showed that throughout the test period all SrHAp samples maintained viable cell adhesion throughout the 28-day test period and were positive for collagen I at day 1, which increased by day 28. The HAp control sample supported minimal viable cell adhesion and, viable cells were not observed by day 28. In addition to positive collagen I staining, cells cultured in contact with the 2SrHAp, 5SrHAp and 10SrHAp materials demonstrated a high level of actin fibre organisation by day 28 and an enhanced expression of collagen I compared to earlier time points, with a denser and more organised expression observed for the 10SrHAp sample compared to other samples in the series (or HAp), as shown in Figure 13. Moreover, the reorganisation of the matrix as the osteoblast becomes an osteocyte is signalled by a reduction in ALP activity, which is clearly seen in Figure 14 [37].

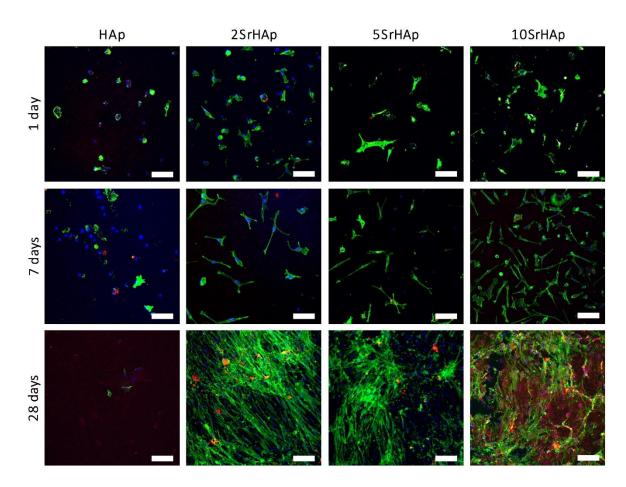


Figure 13. Qualitative immunohistochemistry for collagen I, when cells were cultured in contact with HAp, 2SrHAp, 5SrHAp and 10SrHAp discs sintered at 600 °C for 2 h (samples were stained with DAPI (blue), f-actin (green) and collagen I (red); scale bar 100 μm).

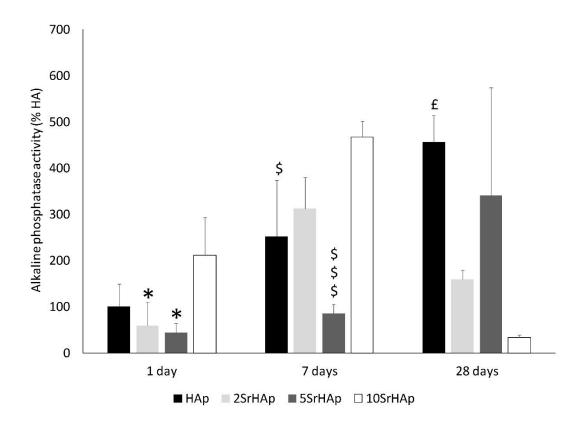


Figure 14. ALP activity of HOBC cultured in contact with HAp (control), 2SrHAp, 5SrHAp and 10SrHAp samples at 1, 7 and 28 days. *: p<0.05 compared to 10SrHAp at day 1; \$: p<0.05, \$\$\$:<0.001 compared to 10SrHAp at day 7; £: p<0.05 compared to 10SrHAp at day 28.

4. Discussion

A range of SrHAp powders with nominal compositions comprising 1, 2 and 5 at% Sr (with respect to [Sr/(Sr+Ca)]·100]) and a pure HAp control have been synthesised using an aqueous precipitation method. Characterisation using AAS, FTIR, XRD, SEM/EDX, STEM/EDX and XPS showed the actual Sr contributions to be 0.56 ± 0.06 , 1.41 ± 0.14 and 3.10 ± 0.30 wt% (6.44 ± 0.69 , 16.29 ± 1.62 and 36.15 ± 3.50 at%), respectively. The stoichiometry of the SrHAp samples and HAp control were 1.67 ± 0.02 and 1.60 ± 0.03 , as determined by EDX and XPS, respectively. SEM and TEM indicate the formation of rod-like crystals of average length 106 ± 23 nm in all

samples with the SrHAp samples also having flat crystals of 300 nm in length. The dimensions reported in this study are similar to those observed in previous work using an aqueous precipitation approach [32], [34]. Furthermore, the HAp material did not have any impurity phases or CO_3^{2-} substitutions (corroborated by FTIR and XPS analyses), corresponding to a chemical formula of $Ca_{10}(PO_4)_6(OH)_2$, as would have been expected. By comparison, the SrHAp samples all showed the presence of levels of Sr-substitution that were significantly lower that the nominal values, as highlighted by the EDX and XPS results. The EDX also highlighted higher than expected Ca/P and (Ca+Sr)/P ratios (>1.67), whereas the XPS showed Ca/P and (Ca+Sr)/P ratios lower than expected (<1.67). However, it should be noted that XPS is a surface analysis technique and only obtains information from the top 5-10 nm of any sample, whereas EDX in this case was analysing over 1 μ m into the sample [25]. As such, it is suggested here that the material in the uppermost surface of the samples is non-stoichiometric (with a lower set of Ca/P and Ca+Sr/P ratios) and differs from the bulk of the disks, which obviously show non-stoichiometry with higher than expected Ca/P and (Ca+Sr)/P ratios. However, the differences here could be attributed to experimental error within both the EDX and XPS techniques, which can be as much as 10% in each case. Despite this, the Sr/(Ca+Sr) ratios, observed in both the EDX and XPS results were in complete agreement, highlighting that the Sr levels increases from $2SrHAp \rightarrow 5SrHAp \rightarrow 10SrHAp$, as expected. Notably, the FWHM of the XRD peaks are seen to increase slightly with increasing levels of Sr (and are less well resolved), which indicates that the Sr-substituted materials are nanosized, as observed in previous work [32], [34]. These results are corroborated by the TEM analyses, showing the decrease in size of the crystallites with increasing Sr substitution in the samples. Previous research has highlighted that nanoscale HAp also has improved sintering and thermal densification properties, due to its increased surface area, which may improve its

mechanical performance [38], [39]. Larger flatter crystals were also observed in the TEM results for the SrHAp samples, similar to those observed previously by Bigi *et al.* [36]. However, further analysis utilised STEM/EDX of the SrHAp samples (both the rod-like crystals and the larger flatter crystals show the presence of Sr in both crystal types). Further to this, the XRD results show a small shift to lower 2θ values, which is indicative of larger Sr²⁺ ions substituting for Ca²⁺ in the HAp lattice [21], [40], [41]. This is corroborated by the dehydroxylation observed in the FTIR spectra and the Ca/P, (Ca+Sr)/P and Sr/(Ca+Sr) ratios observed in the AAS, EDX and XPS results. Therefore it would be suggested that the SrHAp samples manufactured here have the chemical formula Ca_{10-x}Sr_x(PO₄)₆(OH)₂. The purity and crystallinity of the SrHAp materials were largely as expected and confirmed the presence of low levels of Sr-substitution within the HAp lattice, as intended.

The higher ALP activity for 10SrHAp samples at earlier time points, i.e., 1 and 7 days, and significant reduction at 28 days suggest that osteoblasts are starting to form osteocytes by 28 days. This phenomenon, when Sr promotes the ultimate differentiation of human osteoblasts into osteocytes has previously been observed in a study with SrRan, an approved antiosteoporotic drug and is in line with the morphological changes observed [42]. ALP would be recognised as an early marker for osteogenic differentiation [43], as would collagen I [17]. Collagen I is an important initial bone matrix protein in new bone formation, and as shown here, the 10SrHAp sample has enhanced expression of collagen I, clearly undergoing significant reorganisation at 28 days. Indeed, Sr has been shown previously to increase collagen synthesis, without affecting matrix mineralisation, and the results obtained here for the 10SrHAp sample exhibit this phenomenon of increased collagen synthesis at 28 days [44]. Other reports in the literature also point out that although Sr has a positive effect on osteoblasts *in vitro*, it can inhibit mineralisation (by inhibiting

crystal growth, enhancing the SrHAp solubility and disturbing Ca metabolism) [44], [45]. Interestingly in this study, with increasing Sr content, the SrHAp samples exhibited on average smaller nanocrystals (as shown in the TEM results), which would be in line with these expectations. However, in this in vitro study we would not have expected the HOBC to mineralise [44]. It is obvious that the 10SrHAp sample here exhibited the more positive *in vitro* response with respect to the ALP expression at the earlier time-points of 1 and 7 days, along with enhanced collagen expression and matrix re-organisation at 28 days. The AAS, EDX and XPS results showed that this sample had the highest level of Sr substitution in the HAp lattice, also exhibiting the lowest Ca/P and (Ca+Sr)/P ratio in the uppermost surface of the disks, which may have facilitated enhanced dissolution of Sr into the surrounding media, resulting in the potential maturation of the osteoblasts into osteocytes. Further to this, as the crystal size decreases, so does the potential surface area, which facilitates enhanced dissolution of the materials. It is well documented that increased Sr substitution in the HAp lattice facilitates increased dissolution of the material [32], [34], and all the results here clearly show that this is optimised in the 10SrHAp sample. Importantly, as Sr content is the only variable between samples (given the phase purity and lack of other apparent ionic substitutions (namely CO_3^{2-})), the effects observed here can be attributed to Sr alone, which is a unique observation in the literature.

5. Conclusions

Hydroxyapatite with varying low levels of Sr substitution was synthesized and comprehensively characterized in order to evaluate the effect of these Sr concentrations on the physicochemical properties and interaction with primary human osteoblast cells. The results obtained clearly show the successful substitution of Ca^{2+} with Sr^{2+} ions in the hydroxyapatite lattice with levels significantly less than the nominal values. This has created a range of Srsubstituted hydroxyapatite materials with no other obvious impurities. *In vitro* results show that the samples with the highest actual Sr concentration have the highest early expression of alkaline phosphatase (days 1 and 7) and an enhanced expression of collagen I up to 28 days, clearly indicating the maturation of osteoblast cells into osteocytes indicative of the formation of new bone. Clearly the level of Sr substitution here is the discriminating factor that is driving the differentiation process of the osteoblasts. As such, these findings contribute to a better understanding of how the physicochemical properties Sr-substituted hydroxyapatite materials influence their *in vitro* (and *in vivo*) response in a manner that assists with the development of new therapeutic options.

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Data Availability

The raw/processed data required to reproduce these findings cannot be shared at this time as the data also forms part of an ongoing study.

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