





## Complete Genome Sequences of Five Bluetongue Virus (BTV) Vaccine Strains from a Commercial Live Attenuated Vaccine, a BTV-4 Field Strain from South Africa, and a Reassortant Strain Isolated from Experimentally Vaccinated Cattle

Carien van den Bergh, Peter Coetzee, Dalan J. Guthrie, Misha le Grange, Estelle H. Venter

Department of Veterinary Tropical Diseases, Equine Research Centre, Faculty of Veterinary Science, University of Pretoria, Onderstepoort, South Africa

This is a report of the complete genome sequences of plaque-selected isolates of each of the five virus strains included in a South African commercial trivalent bluetongue virus (BTV) attenuated live virus vaccine, a BTV-4 field strain isolated from Rustenburg, South Africa, in 2011, and a bluetongue reassortant (bluetongue virus 4 strain 4/O. aries-tc/ZAF/11/OBP-115) isolated from experimentally vaccinated cattle. Full-genome sequencing and phylogenetic analyses show that the bluetongue virus 9 strain 9/B. taurus-tc/ZAF/15/Onderstepoort\_B02b is a reassortant virus containing segments from both BTV-9 and BTV-8.

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luetongue virus (BTV) is a noncontagious disease of ruminants that is transmitted by *Culicoides* biting midges (Diptera: Ceratopogonidae) (1) (genus Orbivirus, family Reoviridae). The genome of BTV is composed of 10 segments of double-stranded RNA (dsRNA) that collectively encode seven structural proteins and five nonstructural proteins (2-4). In South Africa, a polyvalent BTV-modified live virus (MLV) vaccine is manufactured by Onderstepoort Biological Products (OBP) Ltd. This vaccine is supplied in three separate vials, each of which contains different BTV serotypes: bottle A contains serotypes 1, 4, 6, 12, and 14, bottle B contains serotypes 3, 8, 9, 10, and 11, while bottle C contains serotypes 2, 5, 7, 13, and 17. Here, we report the fullgenome sequences of BTV-3, BTV-8, BTV-9, BTV-10, and BTV-11, which were isolated from bottle B of the BTV-MLV vaccine (batch 115; OBP Ltd., Onderstepoort, South Africa), a BTV-4 field strain isolated from a clinically infected ewe in Rustenburg, and a reassortant strain between BTV-9 and BTV-8 segment 8 isolated from experimentally vaccinated cattle. The individual serotypes were independently isolated from the vaccine bottle and/or infected blood using plaque selection on Vero cells (5). Each of these viruses was then passaged on monolayers of Vero African green monkey cells grown in 25-cm<sup>2</sup> tissue culture flasks. BTV dsRNA was extracted from virus-infected cells using TRIzol reagent (Life Technologies, Johannesburg, South Africa). Sequencing templates were prepared using full-length amplification of cDNAs (6). Amplicons were sequenced on an Illumina MiSeq sequencer (Inqaba Biotechnical Industries [Pty] Ltd., Pretoria, South Africa) using the Nextera XT DNA sample preparation kit and 300-bp paired-end V3 Illumina chemistry. The Illumina sequence reads were analyzed using Geneious 8.1.5. A combination of de novo

assembly, followed by mapping to reference genomes available on GenBank, was used to obtain the full-length genome sequences of the seven viruses.

Pairwise nucleotide sequence identities between the genome sequences derived from the isolated BTV-MLVs and MLV sequences already available on GenBank were as follows: bluetongue virus 3 strain3/Labstr/ZAF/14/OBP-115 was 99% identical to BTV-3 isolate RSArrrr/03, bluetongue virus 8 strain8/Labstr/ZAF/14/OBP-115 was 99% identical to BTV-8 isolate RSArrrr/08, bluetongue virus 9 strain9/Labstr/ZAF/14/OBP-115 was 99 to 100% identical to BTV-9 isolate RSArrrr/09, bluetongue virus 10 strain10/Labstr/ZAF/14/OBP-115 was 99% identical to BTV-10 isolate 2627, and bluetongue virus 11 strain11/Labstr/ZAF/14/OBP-115 was 99% identical to BTV-11 strain BTV-11 VAC.

Nucleotide sequence accession numbers. The full-genome sequences of bluetongue virus 3 strain 3/Labstr/ZAF/14/OBP-115, bluetongue virus 8 strain 8/Labstr/ZAF/14/OBP-115, bluetongue virus 9 strain9/Labstr/ZAF/14/OBP-115, bluetongue virus 10 strain10/Labstr/ZAF/14/OBP-115, and bluetongue virus 11 strain11/Labstr/ZAF/14/OBP-115 have been deposited in GenBank under the accession numbers KT317675 to KT317684, KT317685 to KT317694, KT885055 to KT885064, KT317695 to KT317704, and KT885065 to KT885074, respectively. The fullgenome sequences of the wild-type strain of BTV-4 (bluetongue virus 4 strain 4/O. aries-tc/ZAF/11/OBP-115) are available on GenBank as accession numbers KT317665 to KT317674. The fullgenome sequences of the bluetongue virus 9 strain 9/B. taurus-tc/ ZAF/14/Onderstepoort\_B02b that is a reassortant between BTV-9 and BTV-8 has been deposited in GenBank as accession numbers KT885075 to KT885084.

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## **REFERENCES**

- Verwoerd DW, Erasmus BJ. 2004. Bluetongue, p 1201–1220. In Coetzer JAW, Tustin RC (ed.), Infectious diseases of livestock, 2nd ed. Oxford University Press, Cape Town, South Africa.
- Verwoerd DW, Louw H, Oellermann RA. 1970. Characterization of bluetongue virus ribonucleic acid. J Virol 5:1–7.
- 3. Ratinier M, Caporale M, Golder M, Franzoni G, Allan K, Nunes SF, Armezzani A, Bayoumy A, Rixon F, Shaw A, Palmarini M. 2011. Iden-

- tification and characterization of a novel non-structural protein of bluetongue virus. PLoS Pathogenesis 7:e1002477. http://dx.doi.org/10.1371/journal.ppat.1002477.
- 4. Ligisa DM, Perez Aguirreburualde MS, Gonzalez FN, Marin-Lopez A, Ruiz V, Wigdorovitz A, Martinez-Escribano JA, Ortego J, Du Santos MJ. 2015. An experimental subunit vaccine based on bluetongue virus 4 VP2 protein fused to an antigen-presenting cells single antibody elicit cellular and humoral immune response in cattle, guinea pigs and IFNAR(-/-) mice. J Vaccine 33:2614–2619.
- Howell PG, Kümm NA, Botha MJ. 1970. The application of improved techniques to the identification of strains of bluetongue virus. Onderstepoort J Vet Res 37:59–66.
- Potgieter AC, Page NA, Liebenberg J, Wright IM, Landt O, Van Dijk AA. 2009. Improved strategies for sequence-independent amplification and sequencing of viral double-stranded RNA genomes. J Gen Virol 90: 1423–1432. http://dx.doi.org/10.1099/vir.0.009381-0.