

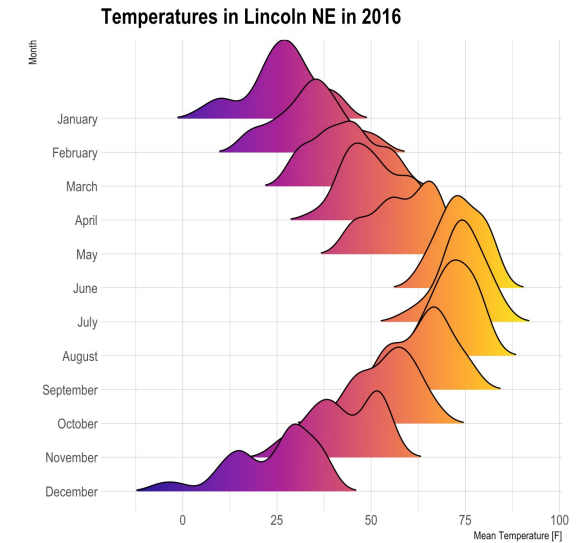
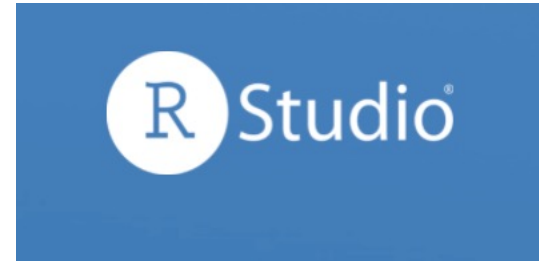
Statistical Analysis of Eye Tracking Data

Krzysztof Krejtz



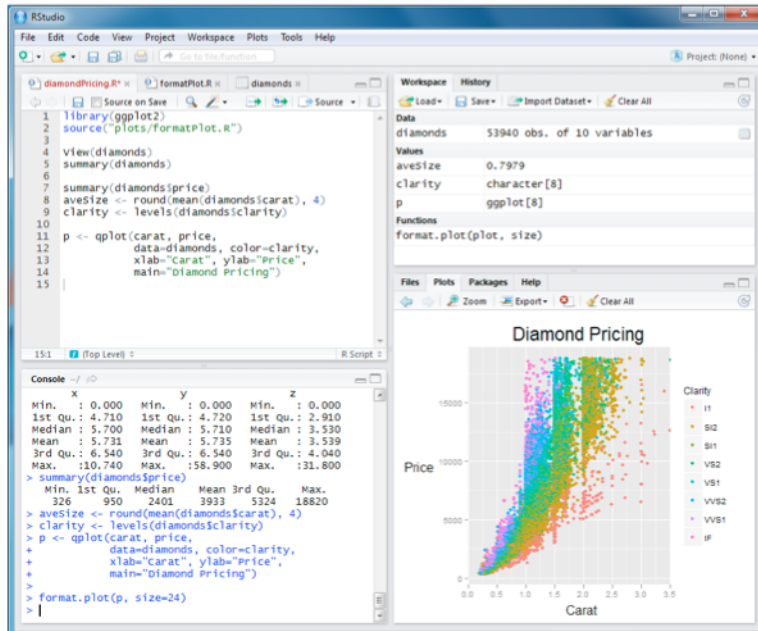
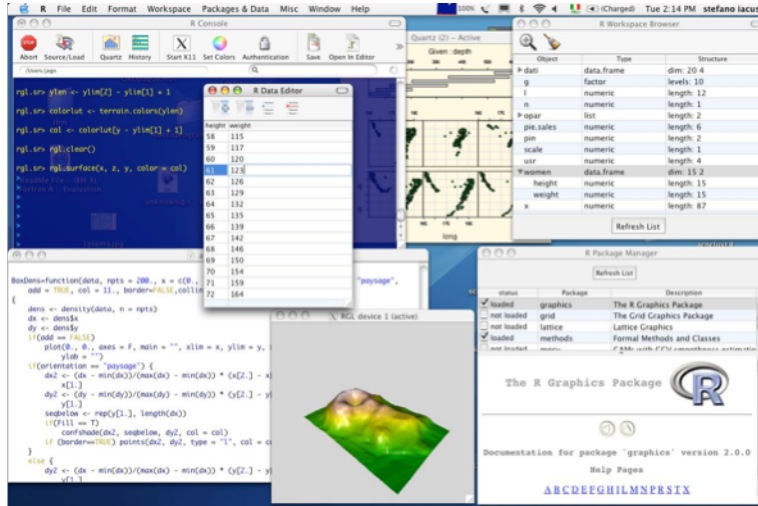
Agenda

- R language & R-Studio IDE basics
- Descriptive statistics
- Moderation analysis in eye tracking studies
 - Mixed-design Analysis of Variance (ANOVA) with interaction effect
 - Pairwise comparisons
 - Graphical representations of interaction effects
- Multiple regression with interaction effect
 - Simple slopes analysis
 - Visualization of interaction effect in regression



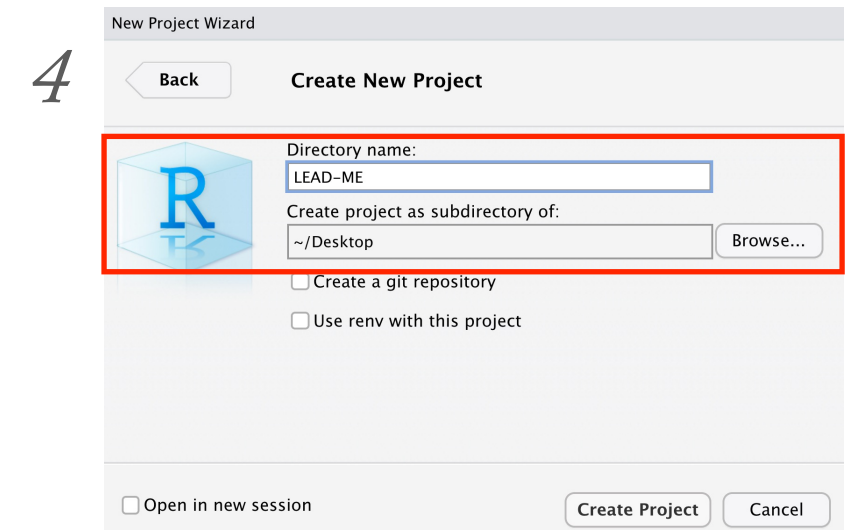
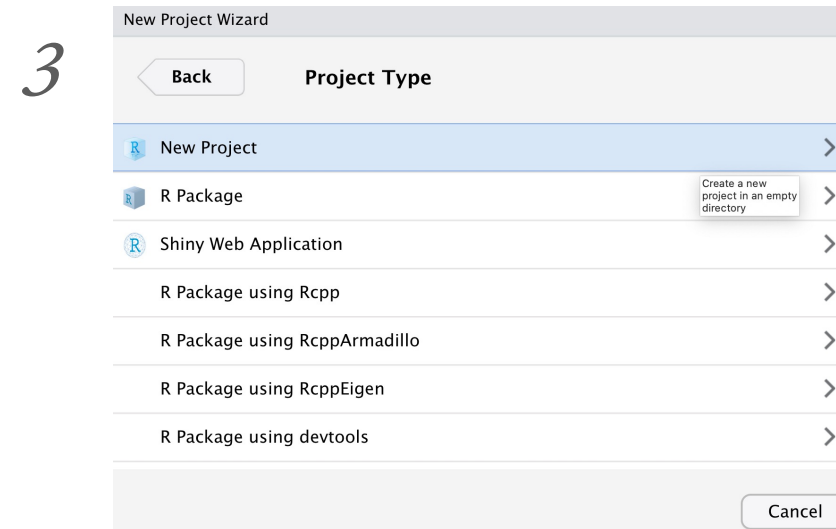
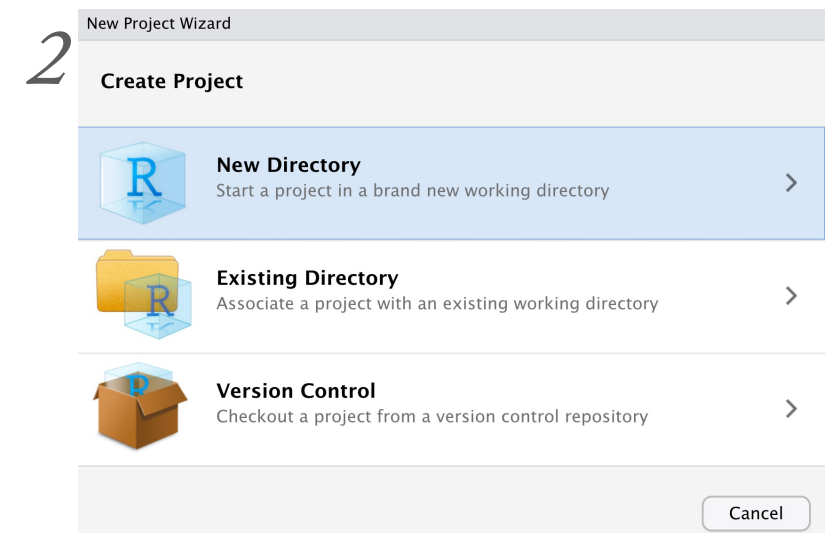
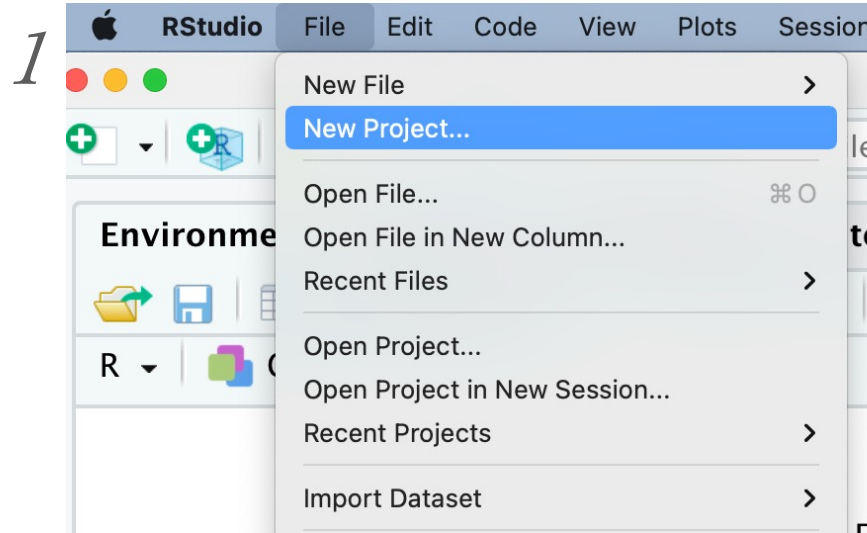
R & R Studio Basics

R language and R Studio IDE



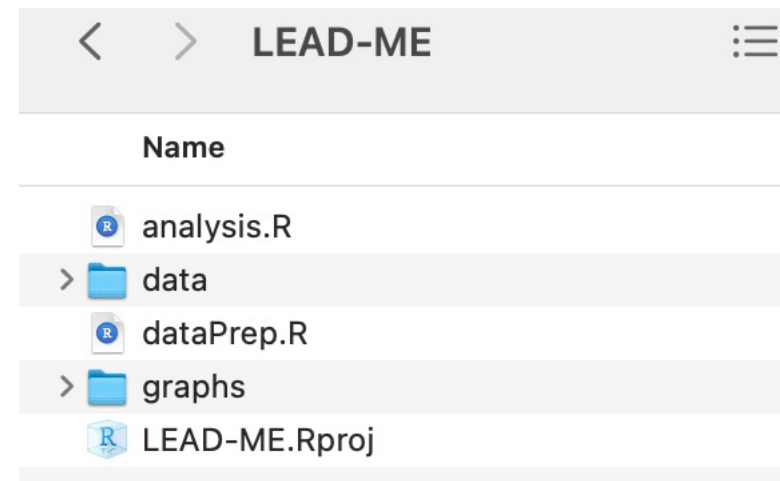
- R is a free software environment for statistical computing and graphics with large worldwide community.
 - <http://www.r-project.org/>
 - R Core Team (2015). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria.
- RStudio is a free and open source Integrated Development Environment (IDE) for R.
 - <http://www.rstudio.com/ide/>

Start a new project in 4 steps

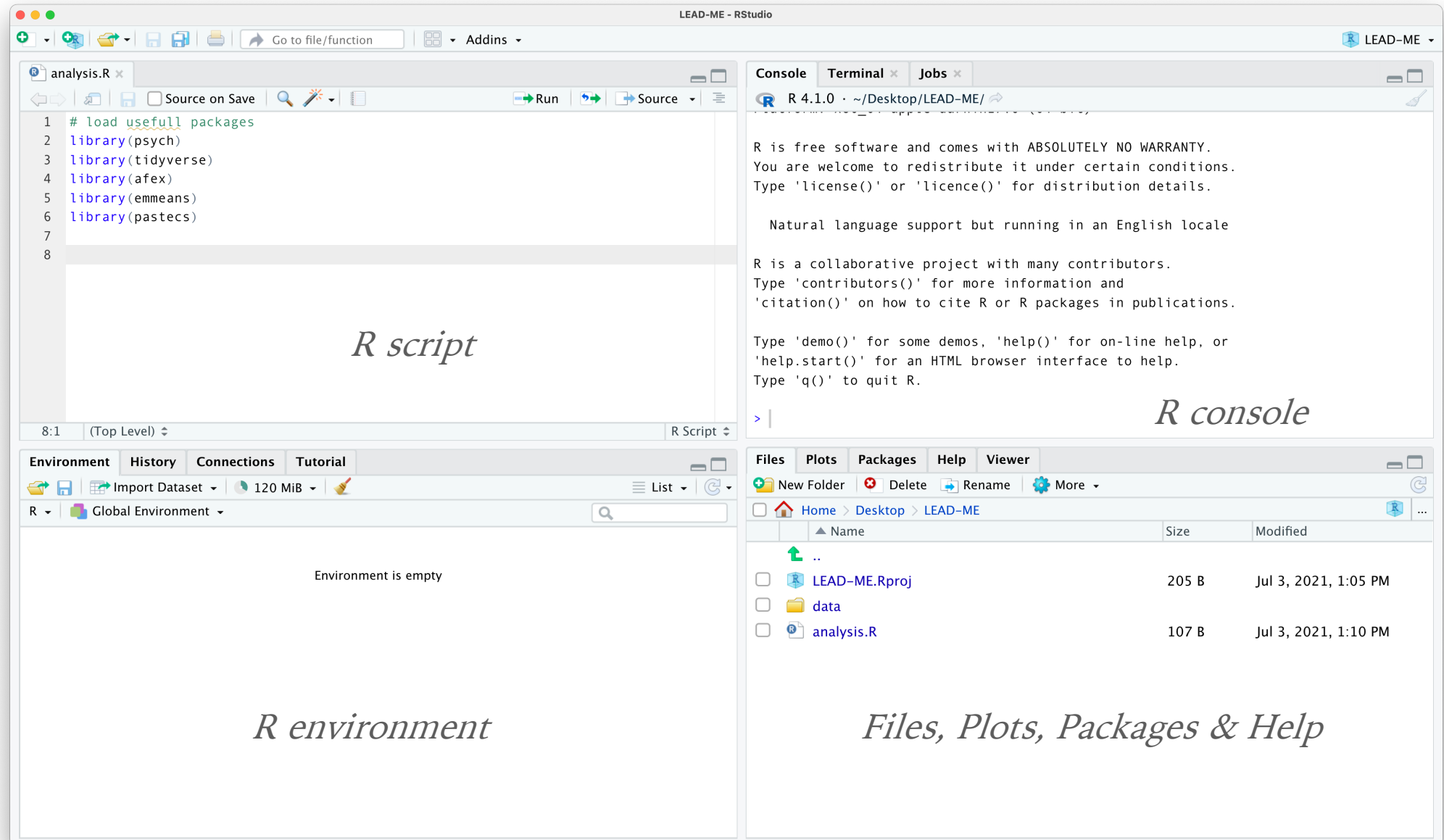


Outside RStudio

- Add data folder into your working folder
- Create 'graphs' folder in you working folder



Complete
 RStudio IDE
 ready for
 statistical
 analyses



The screenshot shows the RStudio IDE interface with the following components:

- Source Editor (analysis.R):** Contains R code for loading packages:


```

            1 # load usefull packages
            2 library(psych)
            3 library(tidyverse)
            4 library(afex)
            5 library(emmeans)
            6 library(pastecs)
            7
            8
            
```
- Console:** Displays the R startup message:


```

            R 4.1.0 · ~/Desktop/LEAD-ME/

            R is free software and comes with ABSOLUTELY NO WARRANTY.
            You are welcome to redistribute it under certain conditions.
            Type 'license()' or 'licence()' for distribution details.

            Natural language support but running in an English locale

            R is a collaborative project with many contributors.
            Type 'contributors()' for more information and
            'citation()' on how to cite R or R packages in publications.

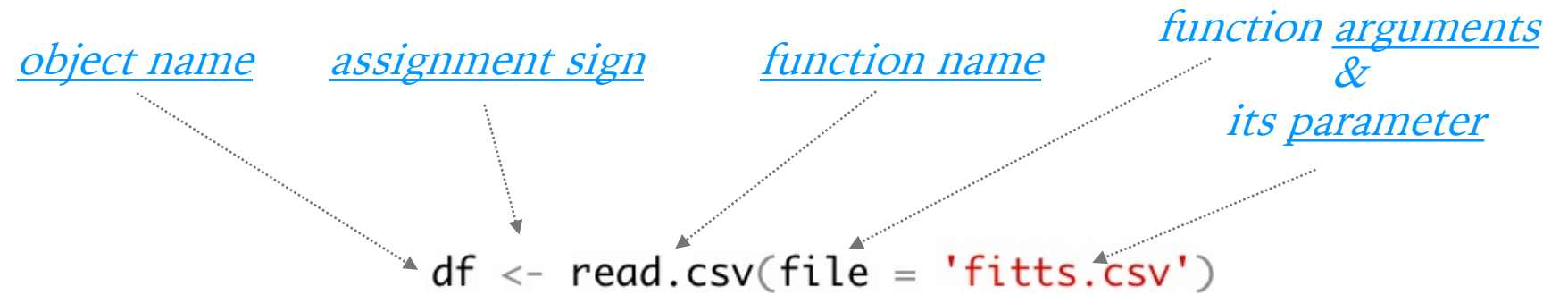
            Type 'demo()' for some demos, 'help()' for on-line help, or
            'help.start()' for an HTML browser interface to help.
            Type 'q()' to quit R.

            > |
            
```
- Environment:** Shows the Global Environment with the message "Environment is empty".
- Files, Plots, Packages & Help:** Shows a file browser view of the LEAD-ME directory:

Name	Size	Modified
..		
LEAD-ME.Rproj	205 B	Jul 3, 2021, 1:05 PM
data		
analysis.R	107 B	Jul 3, 2021, 1:10 PM

Talking to R

```
lunch <- make.pizza(dough = "thin",  
                    sauce = "tomato",  
                    main.ingredient = "mozzarella",  
                    doubleCheese = FALSE)
```


Ordering pizza (*oops...* analyses) with R

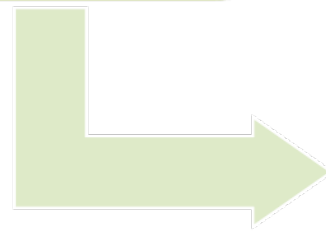
I want ... an object `df` ... made by ... reading the csv file ... called `fitts.csv`

Talking to R

libraries
functions
parameters

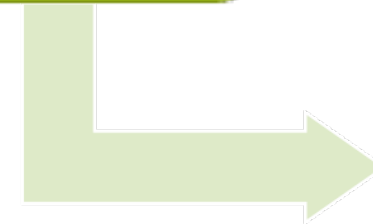
Libraries
(packages)

- All functions are stored in packages.
- Load packages to R memory to make its functions available
- There are packages for everything – you can download them from Internet



Functions

- Functions names are described in packages' help files



Parameters

- All you will find in the 'help' files

libraries

functions

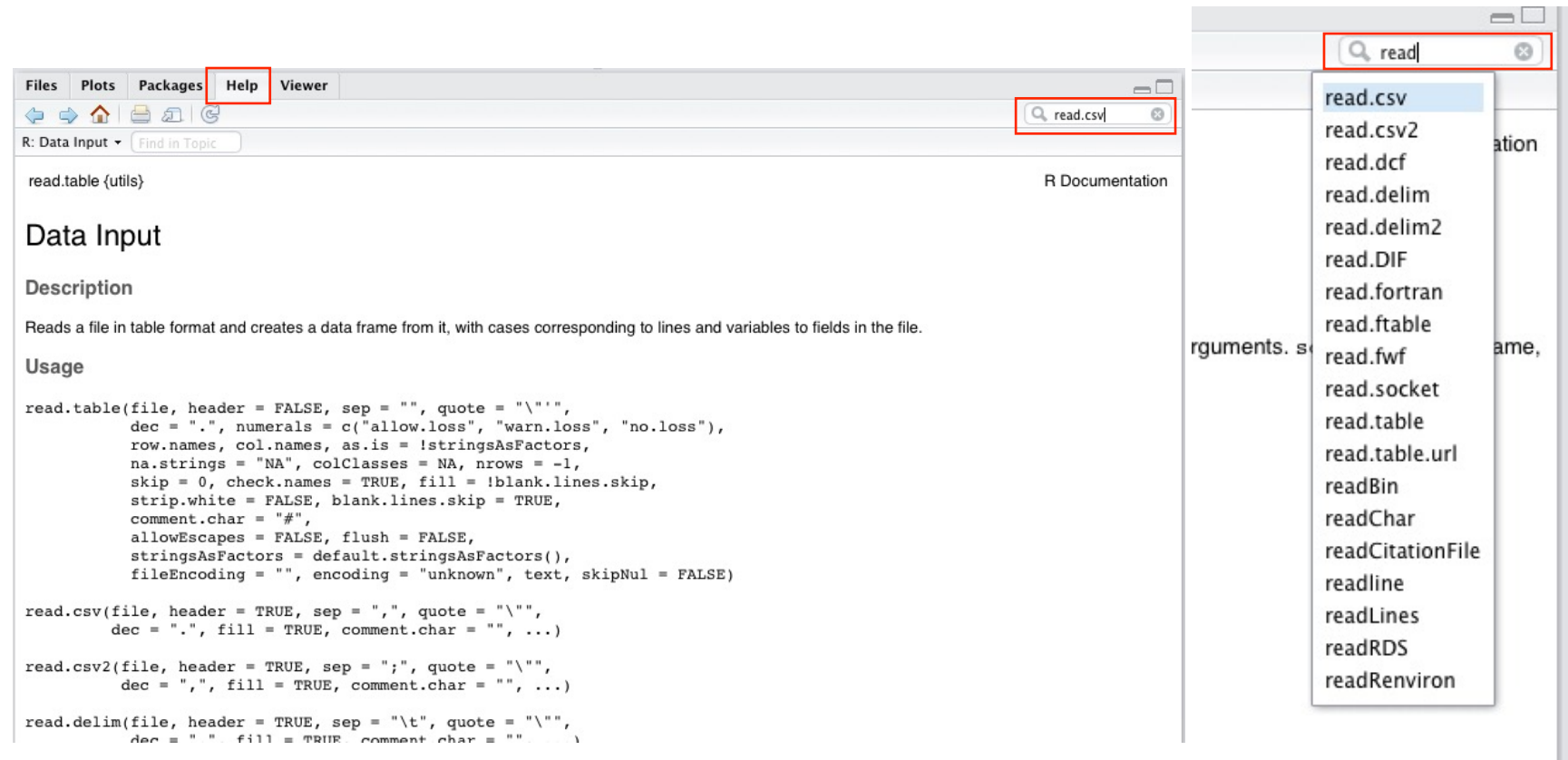
parameters

`library(psych)`

`describe(x, na.rm = TRUE, interp=FALSE, skew = TRUE, ranges = TRUE, trim=.1,
type=3, check=TRUE, fast=NULL, quant=NULL, IQR=FALSE)`

Searching for functions' help: definition, arguments, examples of usage

Using help



The screenshot shows the RStudio Help viewer interface. The 'Help' menu item is highlighted with a red box. A search bar in the top right corner contains the text 'read.csv', also highlighted with a red box. A dropdown menu is open, listing various R functions related to reading data, with 'read.csv' selected. The main content area displays the documentation for the 'read.csv' function, including its description and usage examples.

Data Input

Description

Reads a file in table format and creates a data frame from it, with cases corresponding to lines and variables to fields in the file.

Usage

```
read.table(file, header = FALSE, sep = "", quote = "\"",
  dec = ".", numerals = c("allow.loss", "warn.loss", "no.loss"),
  row.names, col.names, as.is = !stringsAsFactors,
  na.strings = "NA", colClasses = NA, nrows = -1,
  skip = 0, check.names = TRUE, fill = !blank.lines.skip,
  strip.white = FALSE, blank.lines.skip = TRUE,
  comment.char = "#",
  allowEscapes = FALSE, flush = FALSE,
  stringsAsFactors = default.stringsAsFactors(),
  fileEncoding = "", encoding = "unknown", text, skipNul = FALSE)

read.csv(file, header = TRUE, sep = ",", quote = "\"",
  dec = ".", fill = TRUE, comment.char = "", ...)

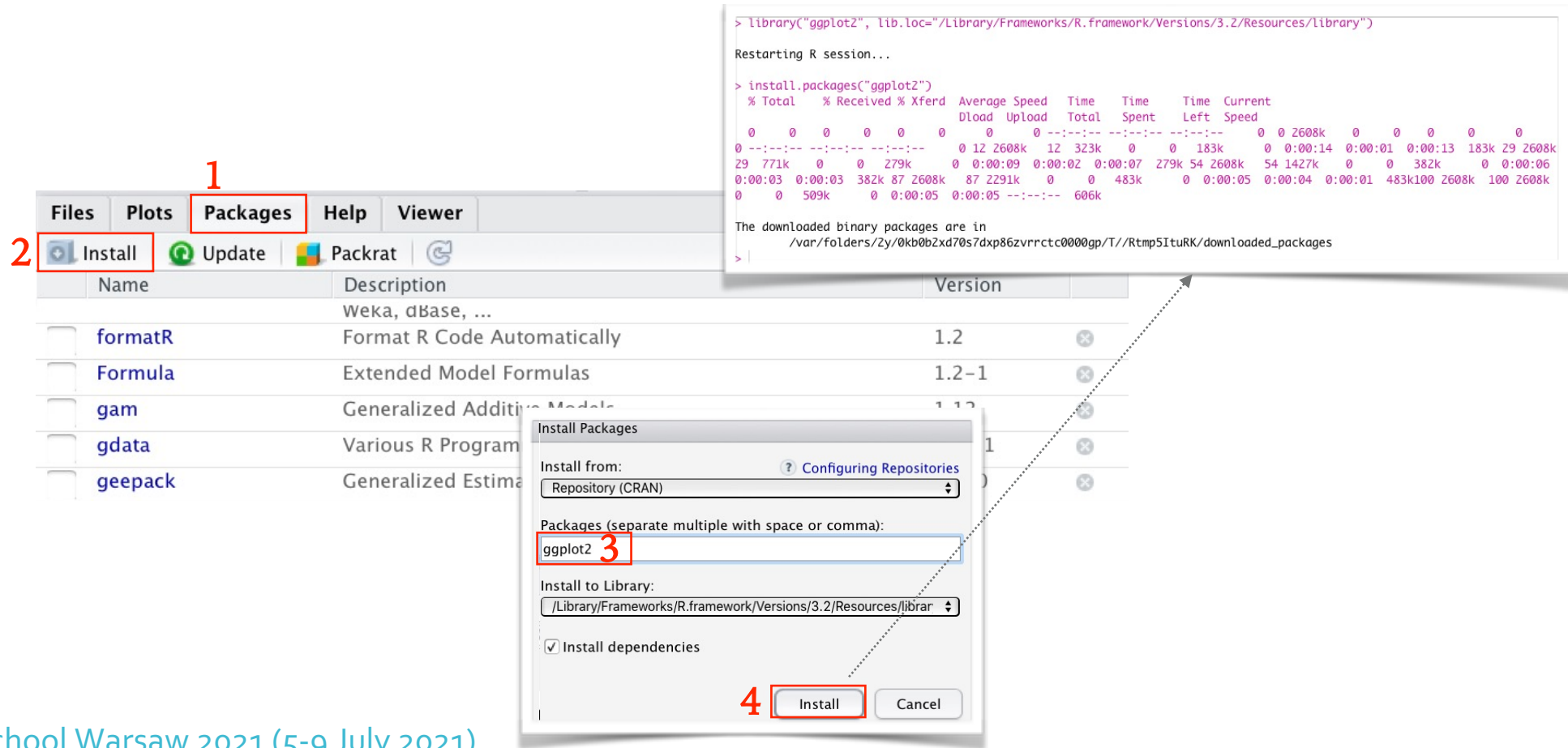
read.csv2(file, header = TRUE, sep = ";", quote = "\"",
  dec = ",", fill = TRUE, comment.char = "", ...)

read.delim(file, header = TRUE, sep = "\t", quote = "\"",
  dec = ".", fill = TRUE, comment.char = "", ...)
```

Installing R libraries (packages)

```
install.packages(pkgs = "ggplot2", dependencies = TRUE)
```

if TRUE will install also all dependent packages



1 Packages tab is selected.

2 Install button is clicked.

3 "ggplot2" is entered in the Packages field.

4 Install button is clicked in the dialog.

```
> library("ggplot2", Lib.Loc="/Library/Frameworks/R.framework/Versions/3.2/Resources/Library")
Restarting R session...

> install.packages("ggplot2")
% Total % Received % Xferd Average Speed Time Time Time Current
% Dload Upload Total Spent Left Speed
0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0
0 12 2608k 12 323k 0 0 183k 0 0:00:14 0:00:01 0:00:13 183k 29 2608k
29 771k 0 0 279k 0 0:00:09 0:00:02 0:00:07 279k 54 2608k 54 1427k 0 0 382k 0 0:00:06
0:00:03 0:00:03 382k 87 2608k 87 2291k 0 0 483k 0 0:00:05 0:00:04 0:00:01 483k100 2608k 100 2608k
0 0 509k 0 0:00:05 0:00:05 ----- 606k

The downloaded binary packages are in
/var/folders/Zy/0kb0b2xd70s7dpx86zvrctc0000gp/T//Rtmp5IuRK/downloaded_packages
>
```

Name	Description	Version
formatR	Format R Code Automatically	1.2
Formula	Extended Model Formulas	1.2-1
gam	Generalized Additive Models	1.12
gdata	Various R Programs	1
geepack	Generalized Estimating Equations	1

Loading installed libraries (packages)

Before using functions from the library we have to load them to the working memory.

`library(ggplot2)`

```
> library(ggplot2)
> |
```

```
> library("ggplot2", lib.loc="/Library/Frameworks/R.framework/Versions/3.2/Resources/Library")
```

1

Name	Description	Version
<input type="checkbox"/> formatR	Format R Code Automatically	1.2
<input type="checkbox"/> Formula	Extended Model Formulas	1.2-1
<input type="checkbox"/> gam	Generalized Additive Models	1.12
<input type="checkbox"/> gdata	Various R Programming Tools for Data Manipulation	2.16.1
<input type="checkbox"/> geeppack	Generalized Estimating Equation Package	1.2-0
2 <input checked="" type="checkbox"/> ggplot2	An Implementation of the Grammar of Graphics	1.0.1
<input type="checkbox"/> gmodels	Various R programming tools for model fitting	2.15.4.1

- Some libraries contain other useful packages e.g., tidyverse

```
> library(tidyverse)
```

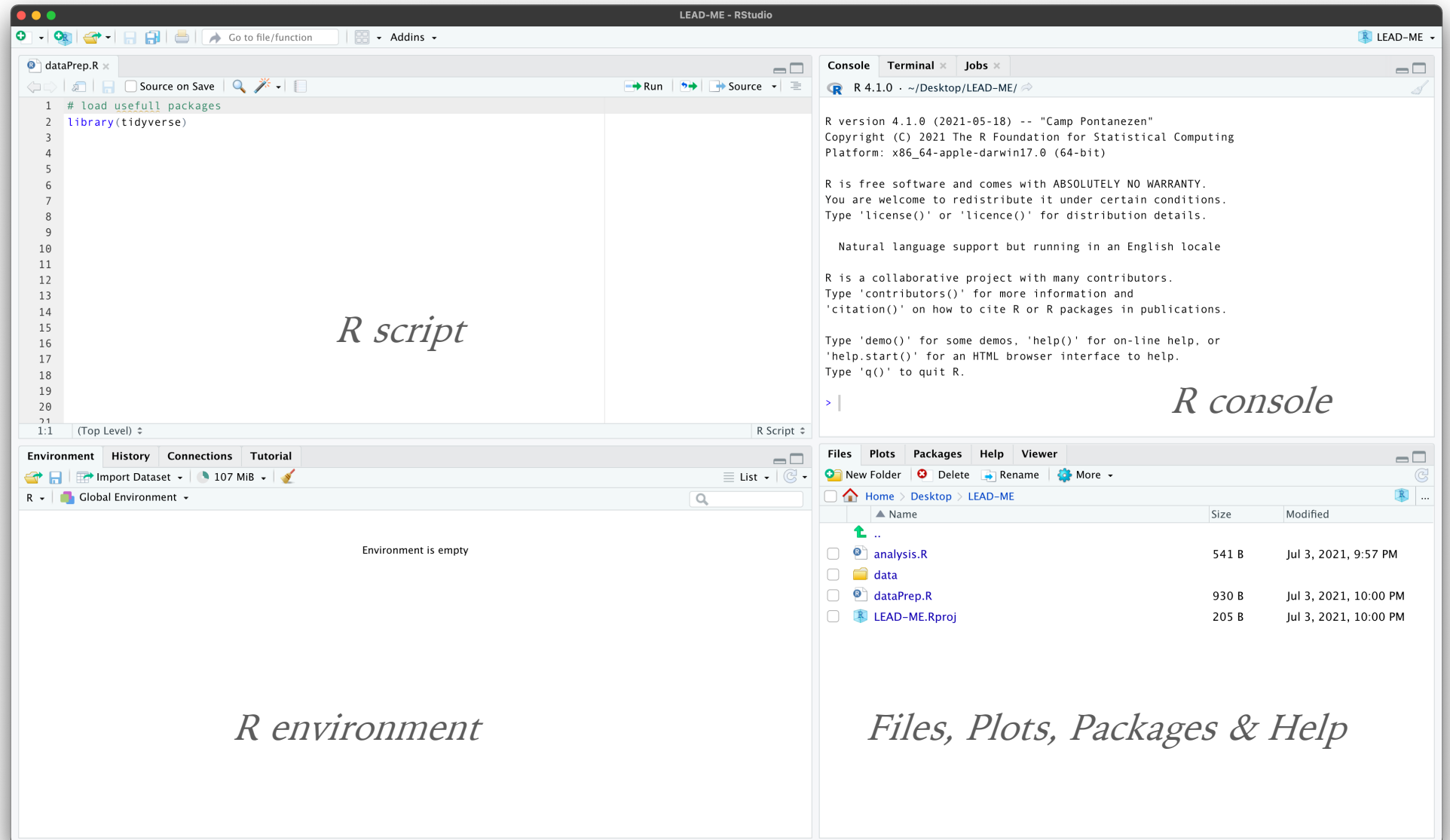
```
— Attaching packages —
```

```
✓ ggplot2 3.3.4    ✓ purrr  0.3.4  
✓ tibble  3.1.2    ✓ dplyr  1.0.6  
✓ tidyr   1.1.3    ✓ stringr 1.4.0  
✓ readr   1.4.0    ✓ forcats 0.5.1
```

```
tidyverse 1.3.1 —
```

Loading
large
libraries

Complete
RStudio IDE
ready for
statistical
analyses



The screenshot shows the RStudio IDE interface with the following components:

- Source Editor:** Contains an R script with the following code:

```
1 # load usefull packages
2 library(tidyverse)
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
1:1 (Top Level) ↕
```
- Console:** Displays the R version information and license text:

```
R 4.1.0 ~~/Desktop/LEAD-ME/ ↵
R version 4.1.0 (2021-05-18) -- "Camp Pontanezen"
Copyright (C) 2021 The R Foundation for Statistical Computing
Platform: x86_64-apple-darwin17.0 (64-bit)

R is free software and comes with ABSOLUTELY NO WARRANTY.
You are welcome to redistribute it under certain conditions.
Type 'license()' or 'licence()' for distribution details.

Natural language support but running in an English locale

R is a collaborative project with many contributors.
Type 'contributors()' for more information and
'citation()' on how to cite R or R packages in publications.

Type 'demo()' for some demos, 'help()' for on-line help, or
'help.start()' for an HTML browser interface to help.
Type 'q()' to quit R.

> |
```
- Environment:** Shows the Global Environment with the message "Environment is empty".
- Files, Plots, Packages & Help:** A file browser showing the contents of the LEAD-ME project folder:

Name	Size	Modified
..		
analysis.R	541 B	Jul 3, 2021, 9:57 PM
data		
dataPrep.R	930 B	Jul 3, 2021, 10:00 PM
LEAD-ME.Rproj	205 B	Jul 3, 2021, 10:00 PM

R script

R console

R environment

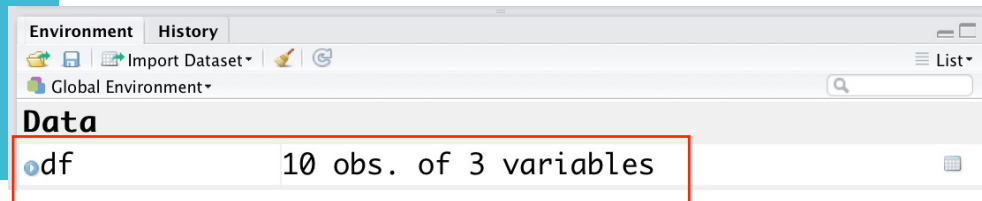
Files, Plots, Packages & Help

- **Data frame** is a quadratic table of data.
 - Different columns can be of different classes (numeric, character, factor, etc.).
 - All variables have to have the same length.
 - This is similar to SPSS datasets or other spreadsheets

Data frames

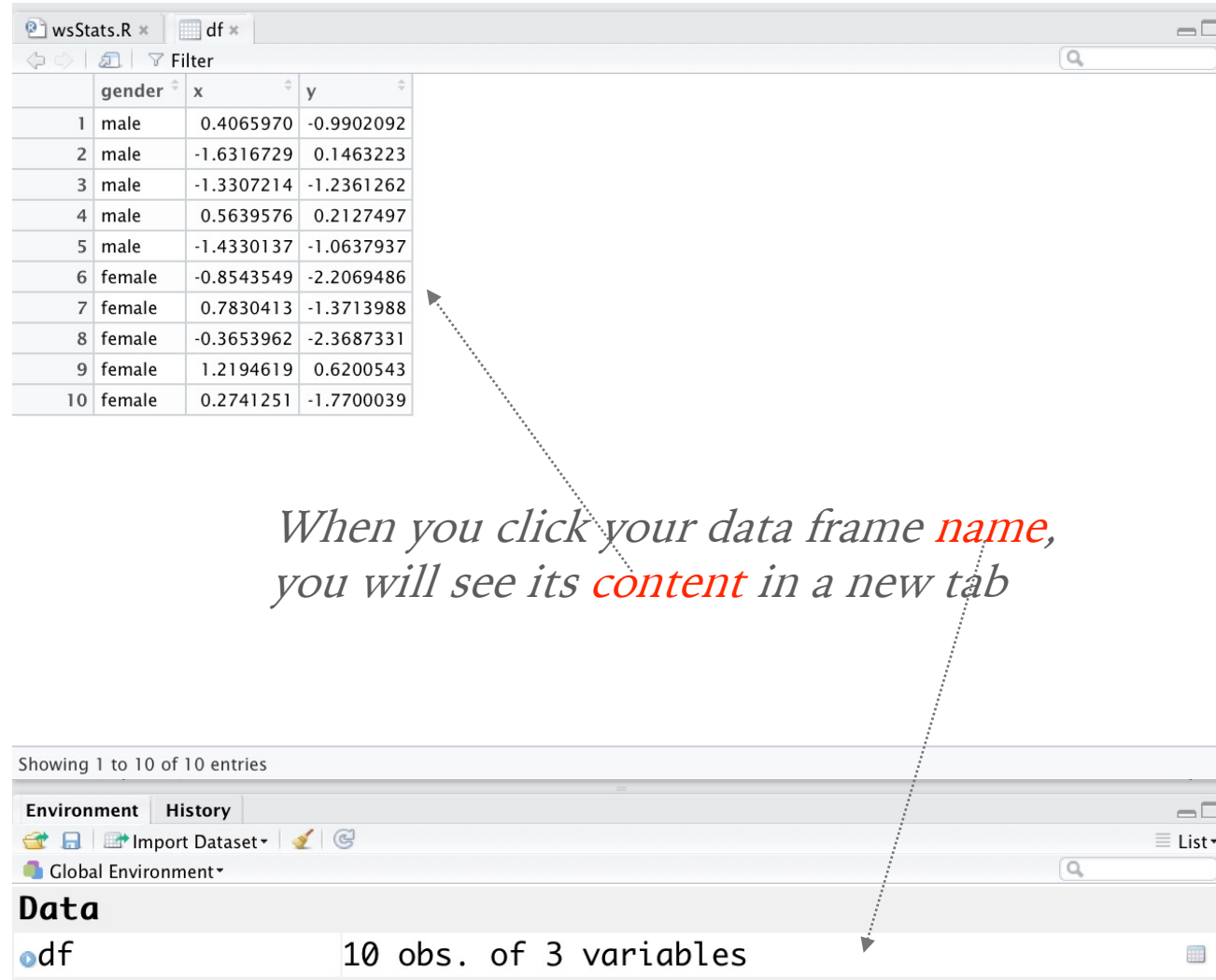
```

20 # Data frames
21 x <- rnorm(10) .....> numeric vector of random 10 values
22 y <- rnorm(10) .....> numeric vector of random 10 values
23 gender <- c(rep("male", 5), rep("female", 5)) .....> character vector of 5 „male” and 5 „female” values
24 gender <- factor(gender) .....> factor of gender
25 df <- data.frame(gender, x, y) .....> data frame with 3 variables: gender, x, and y
    
```



When done you'll see your data frame in Environment tab + information about its dimensions

Data frames



	gender	x	y
1	male	0.4065970	-0.9902092
2	male	-1.6316729	0.1463223
3	male	-1.3307214	-1.2361262
4	male	0.5639576	0.2127497
5	male	-1.4330137	-1.0637937
6	female	-0.8543549	-2.2069486
7	female	0.7830413	-1.3713988
8	female	-0.3653962	-2.3687331
9	female	1.2194619	0.6200543
10	female	0.2741251	-1.7700039

Showing 1 to 10 of 10 entries

Environment History

Global Environment

Data

df 10 obs. of 3 variables

*When you click your data frame **name**, you will see its **content** in a new tab*

Data frames & variables

A set of useful functions

(good to remember
all of them)

27	<code>class(df)</code>>	<i>information about the object class</i>
28	<code>dim(df)</code>>	<i>dimensions of data frame or matrix</i>
29	<code>names(df)</code>>	<i>names of variables in the data frame</i>
30	<code>str(df)</code>>	<i>structure of data frame and all its variables</i>
31	<code>head(df)</code>>	<i>displays first 6 rows of data frame</i>

```
> names(df)
[1] "gender" "x"      "y"
```

```
> head(df)
  gender      x      y
1  male  0.4065970 -0.9902092
2  male -1.6316729  0.1463223
3  male -1.3307214 -1.2361262
4  male  0.5639576  0.2127497
5  male -1.4330137 -1.0637937
6 female -0.8543549 -2.2069486
```

```
> str(df)
'data.frame':  10 obs. of  3 variables:
 $ gender: Factor w/ 2 levels "female","male": 2 2 2 2 2 1 1 1 1 1
 $ x      : num  0.407 -1.632 -1.331 0.564 -1.433 ...
 $ y      : num  -0.99 0.146 -1.236 0.213 -1.064 ...
```

Data frames

Calling the variable

- Calling/using a specific variable from the data frame can be done in two ways:
 - referring to the name of the variable

`dataFrame$variableName`

```
36 df$gender
```

```
> df$gender  
[1] male  male  male  male  male  female female female female female  
Levels: female male
```

- referring to the index (number) of the column which stores the variable in the data frame

`dataFrame[row, column]`

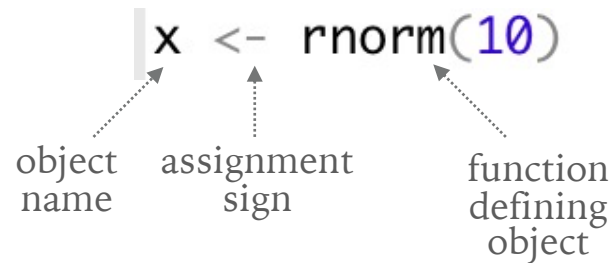
```
38 df[,1]
```

```
> df[,1]  
[1] male  male  male  male  male  female female female female female  
Levels: female male
```

- Objects are *bricks*, which you can use for the analyses (data frames, variables, statistics results, functions, plots, etc.)
- Objects are stored temporarily in the R environment
- Objects can be loaded, eg. data frames or created by R

Objects in R

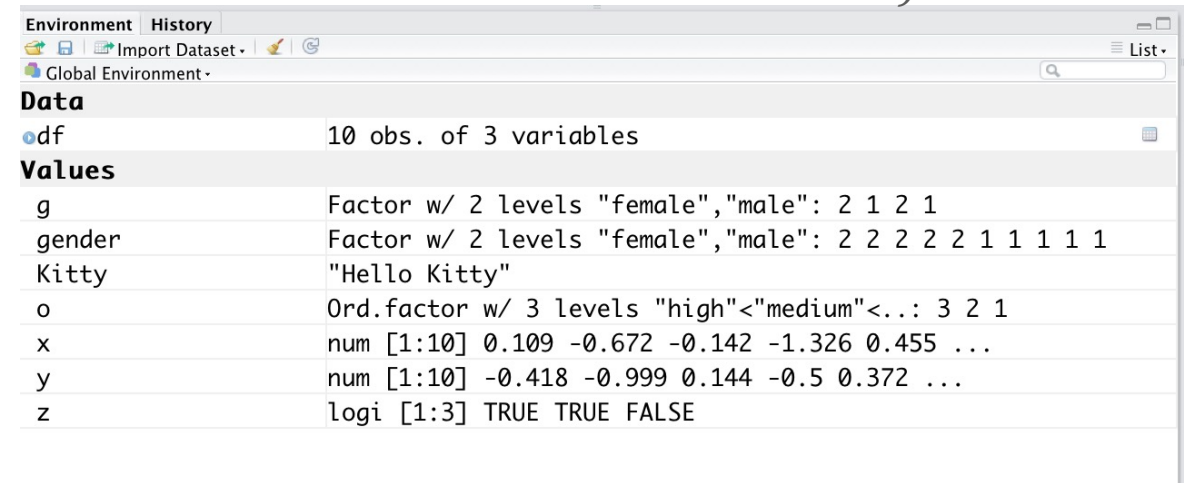
create the object



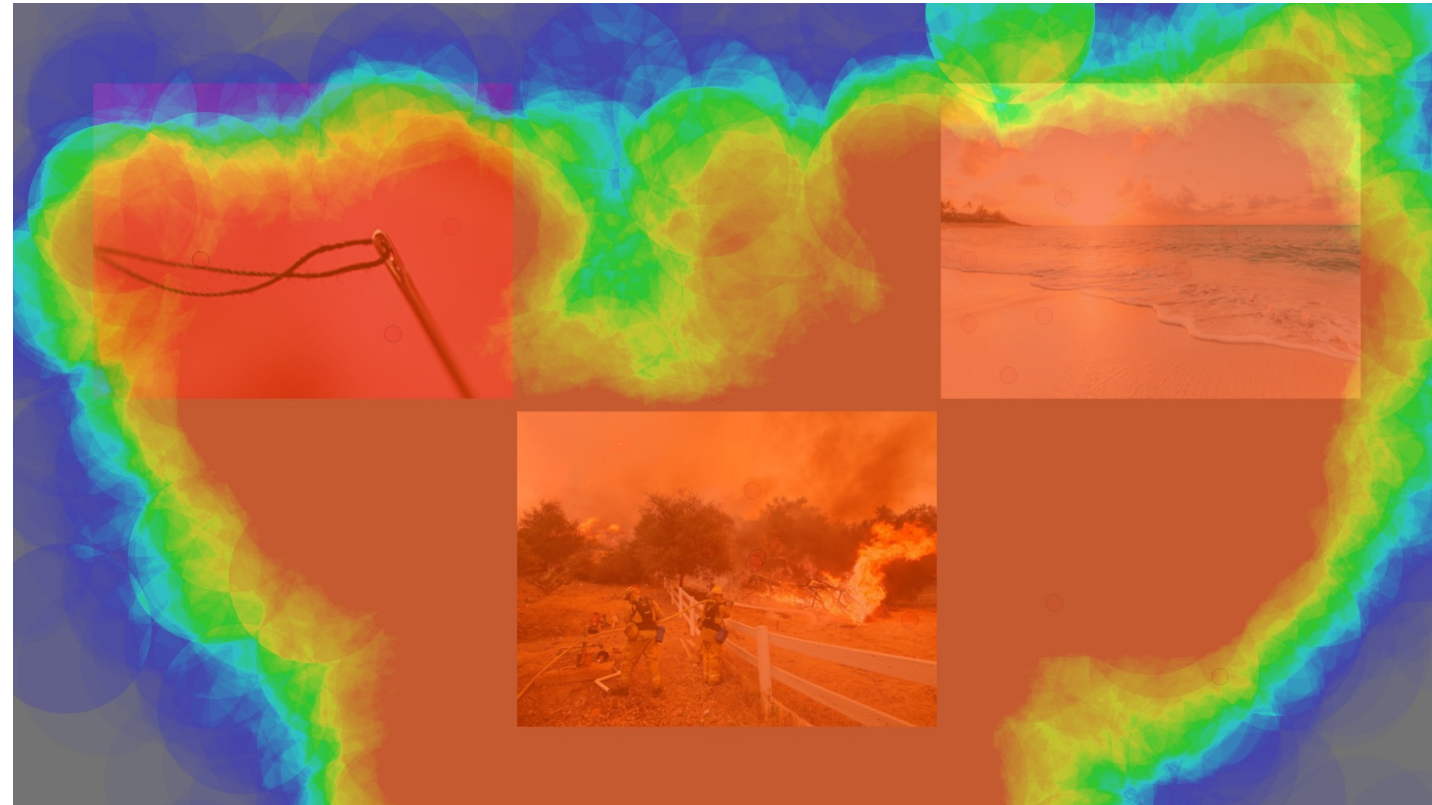
remove the object

`rm(x)`

the list of available objects

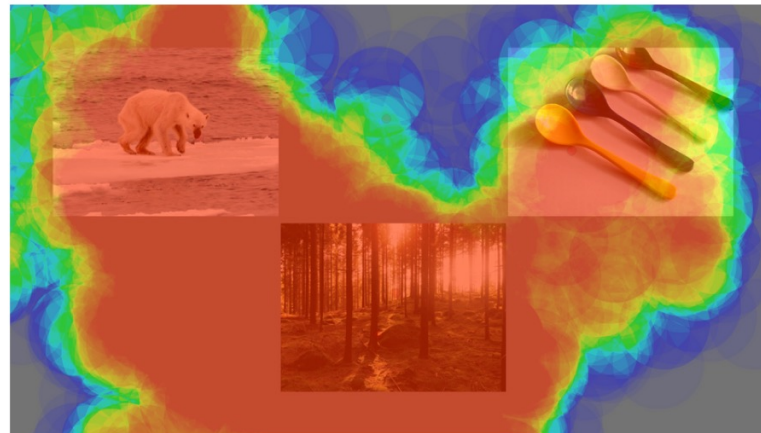
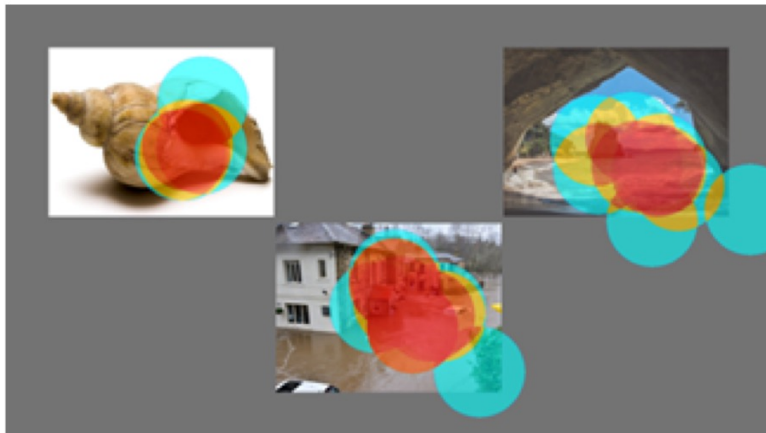


Environment		History
Global Environment		
Data		
df	10 obs. of 3 variables	
Values		
g	Factor w/ 2 levels "female","male": 2 1 2 1	
gender	Factor w/ 2 levels "female","male": 2 2 2 2 2 1 1 1 1 1	
Kitty	"Hello Kitty"	
o	Ord.factor w/ 3 levels "high"<"medium"<...: 3 2 1	
x	num [1:10] 0.109 -0.672 -0.142 -1.326 0.455 ...	
y	num [1:10] -0.418 -0.999 0.144 -0.5 0.372 ...	
z	logi [1:3] TRUE TRUE FALSE	



Exemplary eye tracking project

Climate change overlooked. The role of attitudes and mood regulation in visual attention to global warming



Anna Mazurowska

SWPS University of Social Sciences and Humanities, Warsaw, Poland

Research Advisor:
dr. Krzysztof Krejtz



Hypothesis

Individuals with **environmental concerns** will be more likely to place visual attention on images depicting **negative consequences of climate change** than individuals without such concerns, where pro-environmental attitude is the mediator.

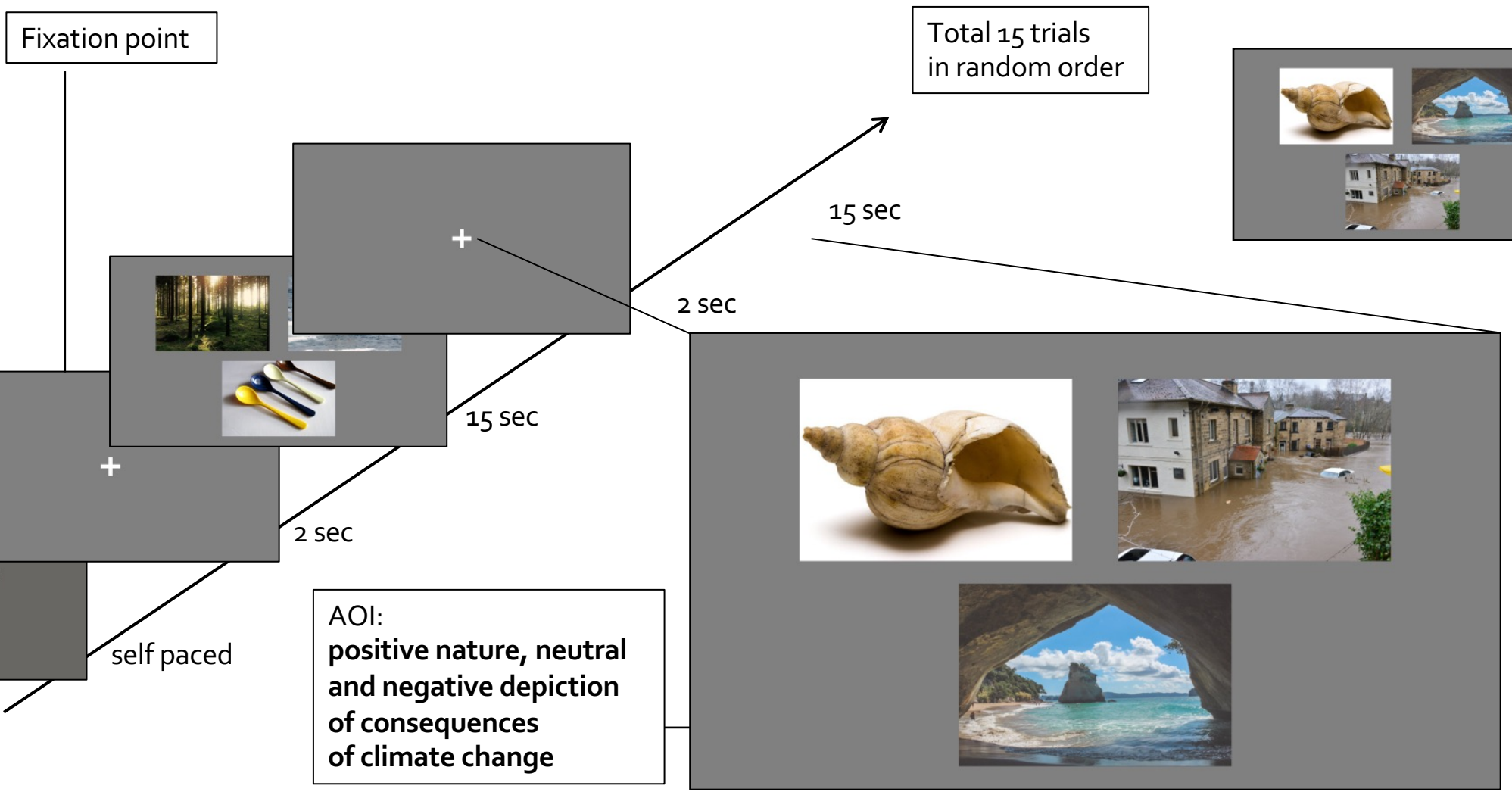
Method

- Participants:
 - students of psychology at SWPS University (N = 48)
- Two stage online study:
 - Stage 1: Eye tracking experiment with RealEye
 - Stage 2: New Ecological Paradigm Scale (NEP) – a measure of pro-ecological attitudes (sensitivity to climate change)



Experimental procedure

Instruction: "Please take a look at the presented photos, especially those that seem to be the most interesting".



Pro-environmental attitude measurement:

New Ecological Paradigm Scale (Dunlap et al. 2000), e.g.:

- "We are approaching the limit of the number of people the Earth can support".
- "Plants and animals have as much right as humans to exist".

Questionnaire

	Strongly disagree	Disagree	Neither agree nor disagree	Agree	Strongly agree
We are approaching the limit of the number of people the Earth can support	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input checked="" type="radio"/>
Plants and animals have as much right as humans to exist	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input checked="" type="radio"/>	<input type="radio"/>

Exemplary
recording
from
RealEye



Areas of interest definition



Neutral:
neutral_igla



Positive:
pozytyw_ocean



Negative:
negatyw_pozar



orange.biz

[This Photo](#) by Unknown Author is licensed under [CC BY](#)

Preparing data for the analyses

read / filter / select / mutate / join / write

Read data frame from file

```
# read data frame
df <- read
```

◆ read_builtin	{readr}
◆ read_csv	{readr}
◆ read_csv_chunked	{readr}
◆ read_csv2	{readr}
◆ read_csv2_chunked	{readr}
◆ read_delim	{readr}
◆ read_delim_chunked	{readr}
◆ read_file	{readr}

```
read_csv(file, col_names = TRUE, col_types = NULL,
         locale = default_locale(), na = c("", "NA"),
         quoted_na = TRUE, quote = "\"", comment = "",
         trim_ws = TRUE, skip = 0, n_max = Inf, guess_max =
         min(1000, n_max), progress = show_progress(),
         skip_empty_rows = TRUE)

read_csv() and read_tsv() are special cases of the general
read_delim(). They're useful for reading the most common types
of flat file data, comma separated values and tab separated values,
Press F1 for additional help
```

```
# read data frame
df <- read_csv(file = "data/|")
```

all_data.csv	data
all_data.Rda	data
qualtrics.csv	data
realeye.csv	data
.DS_Store	data

Read data frame from file

```
> df <- read_csv(file = "data/realeye.csv")
```

— Column specification

```
cols(  
  ID = col_double(),  
  aoi_id = col_character(),  
  aoi_name = col_character(),  
  aoi_size_percents = col_double(),  
  aoi_fixation_total_count = col_double(),  
  aoi_fixation_average_duration_ms = col_double(),  
  aoi_fixation_ttff_ms = col_double(),  
  aoi_fixation_average_total_time_spent_ms = col_double(),  
  aoi_fixation_first_fixation_average_duration_ms = col_double(),  
  notes = col_character()  
)
```


Environment History Connections Tutorial

Import Dataset 344 MiB

R Global Environment

Data

df 6026 obs. of 10 variables

analysis.R x df x

Filter

ID	aoi_id	aoi_name	aoi_size_percents	aoi_fixation_total_count	aoi_fixation_averag
1	56827	7499dfa7-a9bd-4e17-a26a-9a62a9200d1f	negatyw_pozar	13.51776	7
2	56827	c4d7557d-835a-496b-9757-be63a5a20754	neutral_igla	13.51776	7
3	56827	7a6fb176-eed4-4ebc-871b-e1bdc483d039	pozytyw_ocean	13.48463	1
4	63148	7499dfa7-a9bd-4e17-a26a-9a62a9200d1f	negatyw_pozar	13.51776	11
5	63148	c4d7557d-835a-496b-9757-be63a5a20754	neutral_igla	13.51776	0
6	63148	7a6fb176-eed4-4ebc-871b-e1bdc483d039	pozytyw_ocean	13.48463	2
7	46018	7499dfa7-a9bd-4e17-a26a-9a62a9200d1f	negatyw_pozar	13.51776	4
8	46018	c4d7557d-835a-496b-9757-be63a5a20754	neutral_igla	13.51776	0
9	46018	7a6fb176-eed4-4ebc-871b-e1bdc483d039	pozytyw_ocean	13.48463	2
10	45940	7499dfa7-a9bd-4e17-a26a-9a62a9200d1f	negatyw_pozar	13.51776	13
11	45940	c4d7557d-835a-496b-9757-be63a5a20754	neutral_igla	13.51776	3
12	45940	7a6fb176-eed4-4ebc-871b-e1bdc483d039	pozytyw_ocean	13.48463	12
13	54370	7499dfa7-a9bd-4e17-a26a-9a62a9200d1f	negatyw_pozar	13.51776	10
14	54370	c4d7557d-835a-496b-9757-be63a5a20754	neutral_igla	13.51776	2

Showing 1 to 13 of 6,026 entries, 10 total columns

First look at
the data



Create AOI type independent variable

- There were three different types of AOI (positive, negative and neutral picture)
- Currently the data frame contains the following AOI names:

```
> unique(df$aoi_name)
[1] "negatyw_pozar"      "neutral_igla"      "pozytyw_ocean"    "negatyw_lodowiec"
[5] "neutral_kosci"     "pozytyw_gory"     "negatyw_niedzwiedz" "neutral_lyzeczeki"
[9] "pozytyw_las"       "negatyw_podtopienia" "neutral_muszla"   "pozytyw_morze"
[13] "negatyw_susza"     "neutral_zegar"    "pozytyw_rzeka"
```

- We want to make AOI type variable which will be used as a factor in further analyses
- We will use the first element of unique AOI names as values of this new variable
- Technically we will split aoi_name variable into two columns by “_”, creating new “aoi_type” and “picture” variables

```
df <- df %>%
  separate(aoi_name, sep = "_", into = c("aoi_type", "picture"))
```

- Sometimes you want to delete from your data frame some variables (columns)
- This time we want to get rid off three variables "notes", "aoi_id", and "picture"
- Also we want to remove four subjects who were not following the instruction

```
df <- df %>%  
  select(-notes, -aoi_id, -picture) %>%  
  filter(ID != "61567" & ID!="64828" & ID!="66001" & ID != "54127")
```

- The list of new df variables names shows that the operation was successful

```
> names(df)  
[1] "ID" "aoi_size_percents" "aoi_fixation_average_duration_ms" "aoi_fixation_average_total_time_spent_ms"  
[2] "aoi_type" "aoi_fixation_total_count" "aoi_fixation_ttff_ms" "aoi_fixation_first_fixation_average_duration_ms"
```

Select & filter

- Missing values are annotated with NA
- NAs are not welcomed
- Always try to find out why you have NAs in your data set
- This time NAs mean that a participant did not fixate on AOI
 - It make sense to replace NAs with zeros

```
df <- df %>%  
  replace_na(list(aoi_fixation_ttff_ms = 0, aoi_fixation_average_duration_ms = 0,  
                 aoi_fixation_average_total_time_spent_ms = 0,  
                 aoi_fixation_first_fixation_average_duration_ms = 0))
```

Missing
values
(NAs)
a special
case

```
# read data file with additional variables
dq <- read_csv(file = "data/qualtrics.csv")
```

— Column specification —

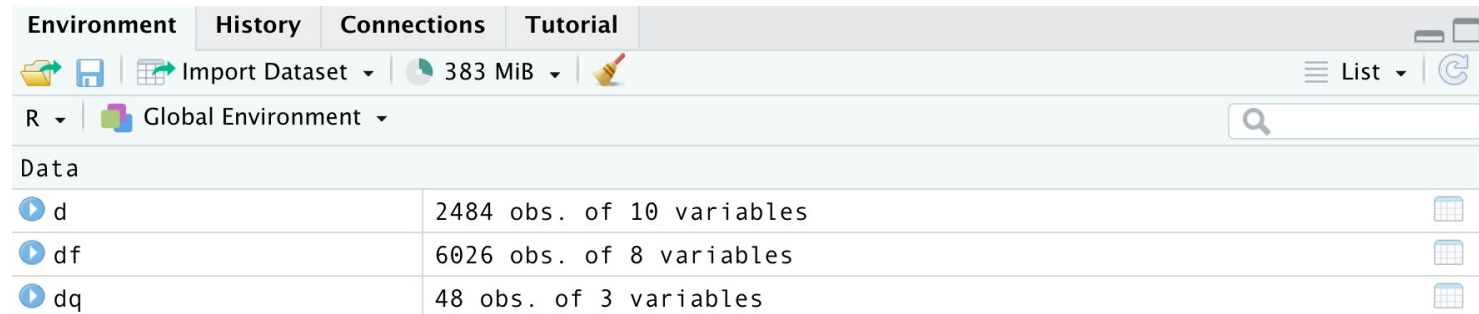
```
cols(
  Q1 = col_double(),
  Q3 = col_double(),
  Q5 = col_double(),
  Q7 = col_double(),
  Q9 = col_double(),
  Q11 = col_double(),
  Q13 = col_double(),
  Q15 = col_double(),
  ID = col_double()
)
```

	Q1	Q3	Q5	Q7	Q9	Q11	Q13	Q15	ID
1	4	5	4	5	5	5	4	4	41428
2	4	5	2	5	4	5	3	2	64666
3	4	4	4	5	4	4	2	4	32185
4	2	2	4	2	5	4	3	3	65176
5	4	4	4	5	3	5	5	5	45652
6	1	5	5	4	5	4	2	5	60466
7	3	5	4	5	4	4	4	4	63148
8	3	4	4	4	4	4	5	4	45940
9	2	4	5	5	2	4	3	5	14539
10	3	4	4	2	4	3	4	3	29236
11	4	2	3	3	5	3	4	4	54619
12	4	4	3	4	2	5	5	4	46417
13	4	4	4	4	5	4	2	4	65905
14	2	4	4	4	4	2	2	4	52011

Read new data
file with
questionnaire
answers

- We need to join two data frames (with eye tracking data and questionnaire data)
- We will join two data frames by subject ID variable

```
# join two data frames by ID (subject ID variable)
d <- inner_join(df, dq, by = "ID")
```



The screenshot shows the RStudio environment pane with the following data frames:

Environment	History	Connections	Tutorial
R	Global Environment	383 MiB	
Data			
d		2484 obs. of 10 variables	
df		6026 obs. of 8 variables	
dq		48 obs. of 3 variables	

- Note that new data frame “d” has lower number of rows.
- It is due to the fact that not all subjects who participated in eye tracking study completed the questionnaire.

Joining two data frames by subject ID

Calculate independent variables / factors

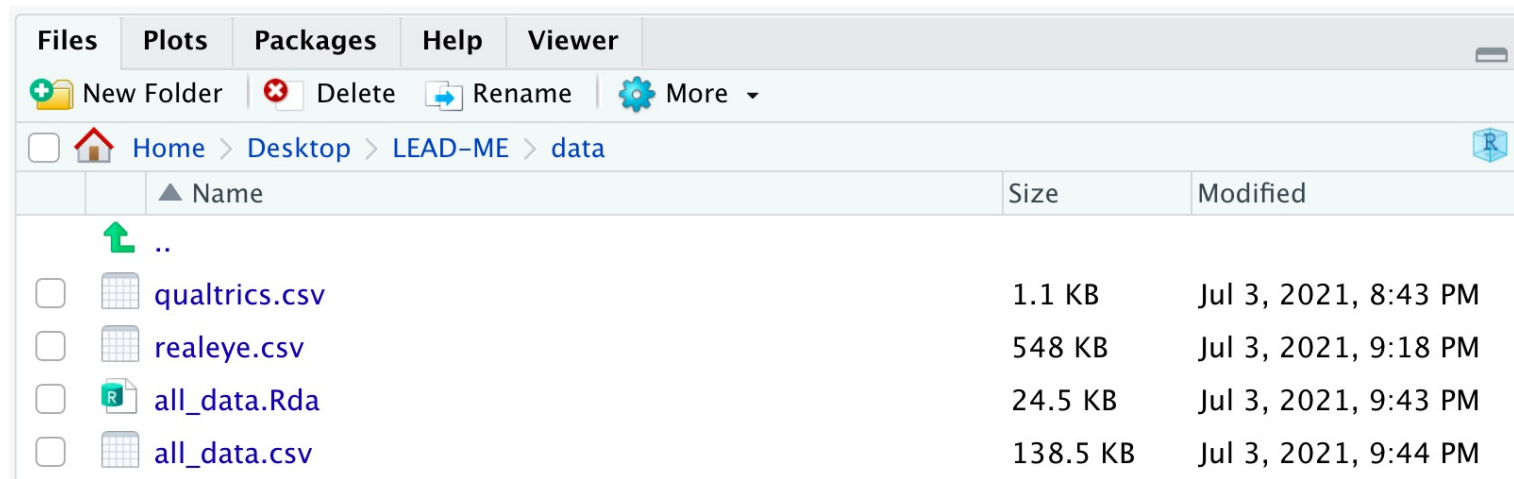
- Questionnaire data contains answers from the New Ecological Paradigm (NEP) questionnaire and **subject ID**
 - The answers were on 1-5 Likert-type scale (the higher value the higher sensitivity to climate change)
- We want to calculate one score of the sensitivity to climate change (NEP) which will be a mean of all given answers
- Next, we want to perform median split on NEP score (low vs high sensitivity to climate change) to use it as a factor in further analyses.
- We need also to set `aoi_type` as factor and make “neutral” value of it as a reference point (important for statistical analyses)
- Last, we do not need all raw answers to each NEP question (they all starts with “Q”).

```
d <- d %>%  
  mutate(NEP = rowSums(select(., starts_with("Q")))) %>%  
  mutate(NEPsplit = cut(NEP, 2, labels = c("low", "high"))) %>%  
  mutate(aoi_type = factor(aoi_type)) %>%  
  mutate(aoi_type = relevel(aoi_type, ref = "neutral")) %>%  
  select(-starts_with("Q"))
```

- Save the entire data frame into a new file
- We can do it in several formats. The most useful are:
 - .csv files (great for sharing even with those who do not use R)
 - Data format (.Rda) - great for further use within R and hard drive space saver

Write data files

```
# write the entire data base into RData file
save(d, file = "data/all_data.Rda")
# or csv file
write_csv(d, file = "data/all_data.csv")
```



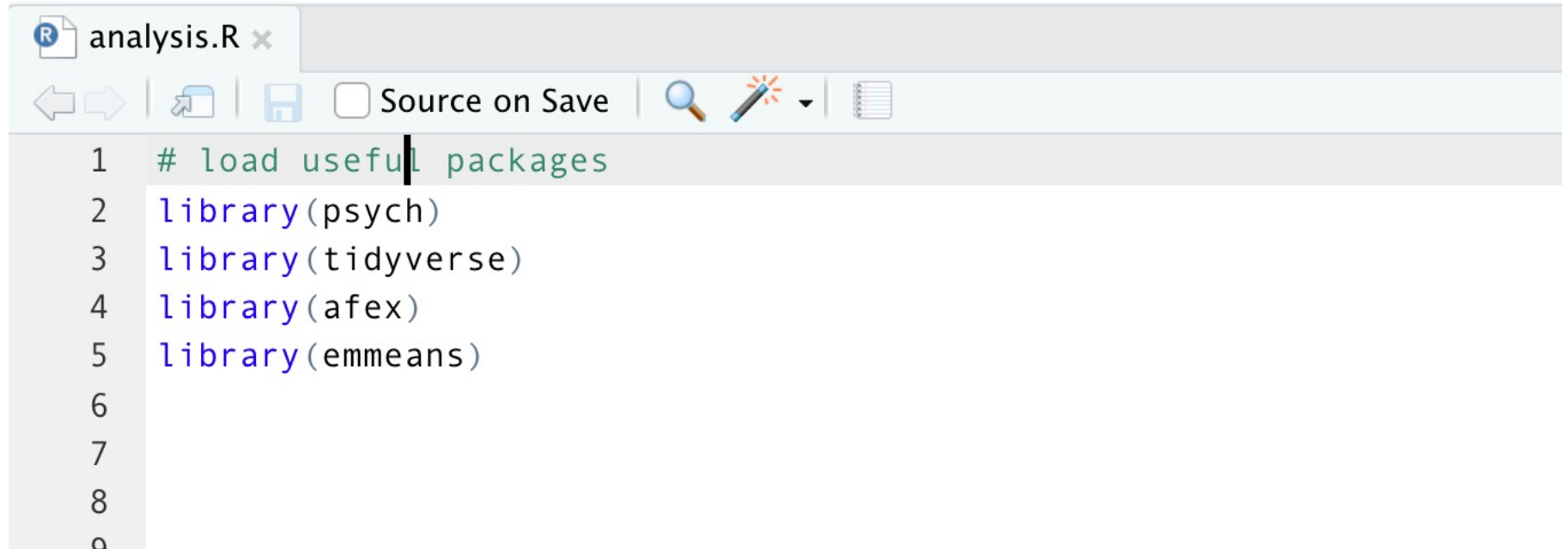
Name	Size	Modified
..		
qualtrics.csv	1.1 KB	Jul 3, 2021, 8:43 PM
realeye.csv	548 KB	Jul 3, 2021, 9:18 PM
all_data.Rda	24.5 KB	Jul 3, 2021, 9:43 PM
all_data.csv	138.5 KB	Jul 3, 2021, 9:44 PM

Descriptive statistics

With visualisations

- It is good to have several R scripts for different purposes.
- We will create a new R script named “analysis.R”
- Start the script with useful libraries

Create new R script “analysis.R”

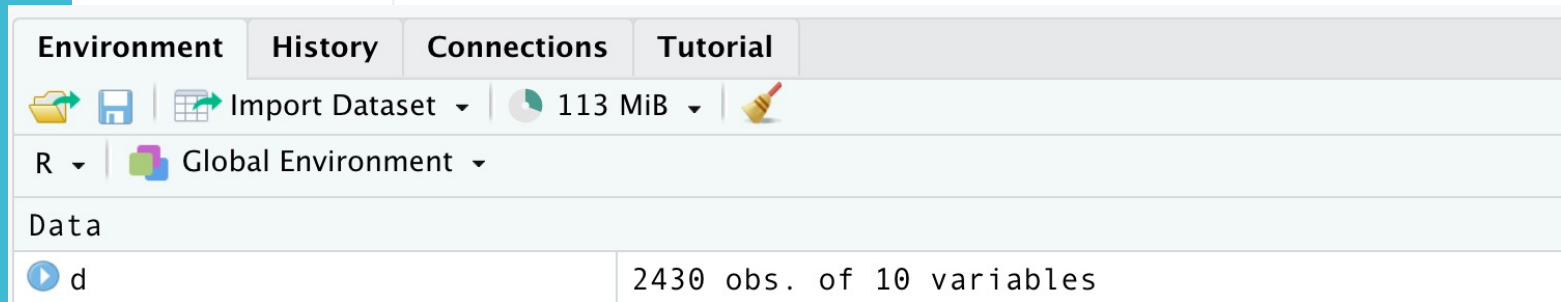


```
analysis.R x
← → | 📄 | 💾  Source on Save | 🔍 ✏️ | 📄
1 # load useful packages
2 library(psych)
3 library(tidyverse)
4 library(afex)
5 library(emmeans)
6
7
8
9
```

- Reading RData format will automatically load data frame name

Read data
frame of
RData
format

```
# read data frame from RData format
load("data/all_data.Rda")
```



Environment | History | Connections | Tutorial

Import Dataset | 113 MiB

R | Global Environment

Data

d | 2430 obs. of 10 variables

ID	aoi_type	aoi_size_percents	aoi_fixation_total_count	aoi_fixation_average_duration_ms	aoi_fixation_tfff_ms	
1	63148	negatyw	13.51776	11	155	520
2	63148	neutral	13.51776	0	NA	NA
3	63148	pozytyw	13.48463	2	149	7384
4	45940	negatyw	13.51776	13	192	1106
5	45940	neutral	13.51776	3	150	747
6	45940	pozytyw	13.48463	12	173	4859
7	54370	negatyw	13.51776	10	145	767
8	54370	neutral	13.51776	2	137	540
9	54370	pozytyw	13.48463	9	151	1343
10	61567	negatyw	13.51776	2	157	2392
11	61567	neutral	13.51776	4	148	973

Describe data frame

- Summary is a generic function used to produce summaries of the results of various model fitting functions.
- The function invokes particular methods which depend on the class of the first argument.
- When applied to data frame it returns basic descriptive statistics for all numerical variables

```
> summary(d)
      ID      aoi_type      aoi_size_percents aoi_fixation_total_count aoi_fixation_average_duration_ms aoi_fixation_ttff_ms
Min.   :10564  Length:2430  Min.     :11.97  Min.     : 0.000  Min.     :100.0  Min.     : 500
1st Qu.:41710  Class :character  1st Qu.:12.18  1st Qu.: 1.000  1st Qu.:144.0  1st Qu.: 1044
Median :55152  Mode  :character  Median :12.30  Median : 4.000  Median :162.0  Median : 1918
Mean   :51814  Mean   :12.46  Mean   : 6.779  Mean   :169.1  Mean   : 3119
3rd Qu.:63148  3rd Qu.:12.80  3rd Qu.:10.000  3rd Qu.:183.0  3rd Qu.:3998
Max.   :67201  Max.   :13.52  Max.   :44.000  Max.   :583.0  Max.   :14882
                                     NA's   :486                                     NA's   :486

aoi_fixation_average_total_time_spent_ms aoi_fixation_first_fixation_average_duration_ms      NEP      NEPsplit
Min.   : 100  Min.   : 100.0  Min.   :3.125  low :1125
1st Qu.: 450  1st Qu.: 123.0  1st Qu.:3.750  high:1305
Median : 1048  Median : 143.0  Median :4.125
Mean   : 1492  Mean   : 163.9  Mean   :4.113
3rd Qu.: 2020  3rd Qu.: 179.0  3rd Qu.:4.500
Max.   :13419  Max.   :2051.0  Max.   :5.000
NA's   :486  NA's   :486
```

Can be hard to read however

The **describe** function in the **psych** package is meant to produce the most frequently requested stats in psychometric and psychology studies, and to produce them in an easy to read data.frame.

```
> describe(d[,4:8])
```

	vars	n	mean	sd	median	trimmed	mad	min	max	range	skew	kurtosis	se
aoi_fixation_total_count	1	2430	6.78	7.42	4.0	5.50	5.93	0	44	44	1.52	2.34	0.15
aoi_fixation_average_duration_ms	2	1944	169.09	42.81	162.0	163.86	28.17	100	583	483	3.16	18.95	0.97
aoi_fixation_ttff_ms	3	1944	3119.15	3063.71	1918.5	2495.53	1650.88	500	14882	14382	1.80	2.84	69.49
aoi_fixation_average_total_time_spent_ms	4	1944	1492.11	1536.47	1048.5	1230.19	1027.44	100	13419	13319	2.86	14.39	34.85
aoi_fixation_first_fixation_average_duration_ms	5	1944	163.92	79.41	143.0	151.01	35.58	100	2051	1951	8.71	169.99	1.80

The **fast=TRUE** option will lead to a speed up of about 50% for larger data sets

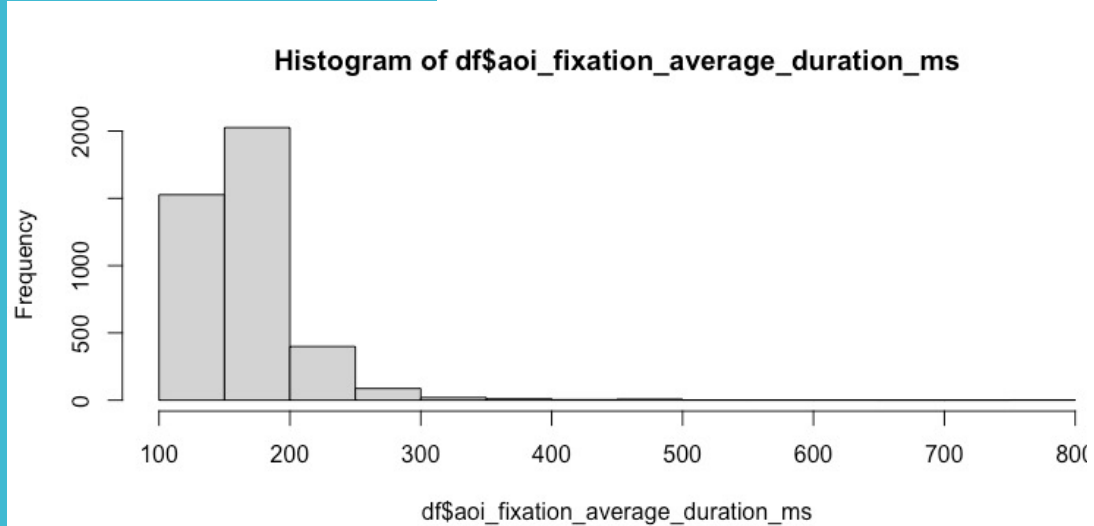
It is also easy to read!

```
> describe(d[,4:8], fast = TRUE)
```

	vars	n	mean	sd	min	max	range	se
aoi_fixation_total_count	1	2430	6.78	7.42	0	44	44	0.15
aoi_fixation_average_duration_ms	2	1944	169.09	42.81	100	583	483	0.97
aoi_fixation_ttff_ms	3	1944	3119.15	3063.71	500	14882	14382	69.49
aoi_fixation_average_total_time_spent_ms	4	1944	1492.11	1536.47	100	13419	13319	34.85
aoi_fixation_first_fixation_average_duration_ms	5	1944	163.92	79.41	100	2051	1951	1.80

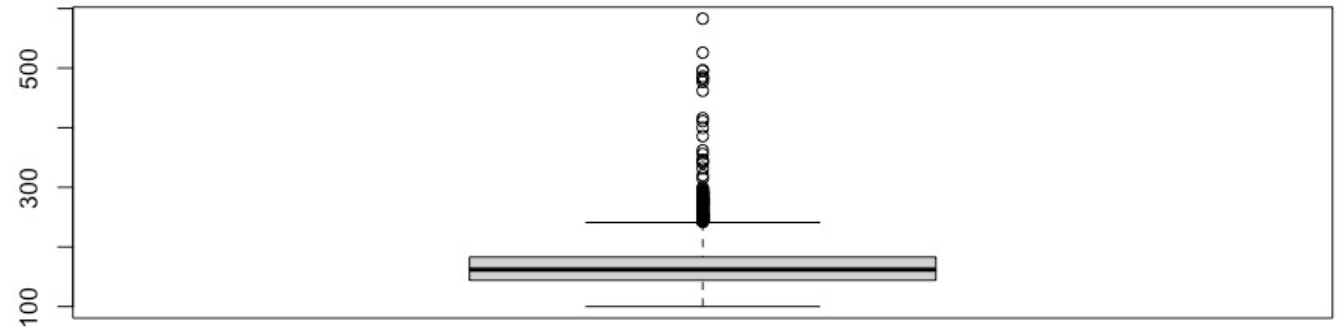
Descriptive
 statistics
 of selected
 variables

```
# plot distribution of variable values with histogram  
hist(d$aoi_fixation_average_duration_ms)
```



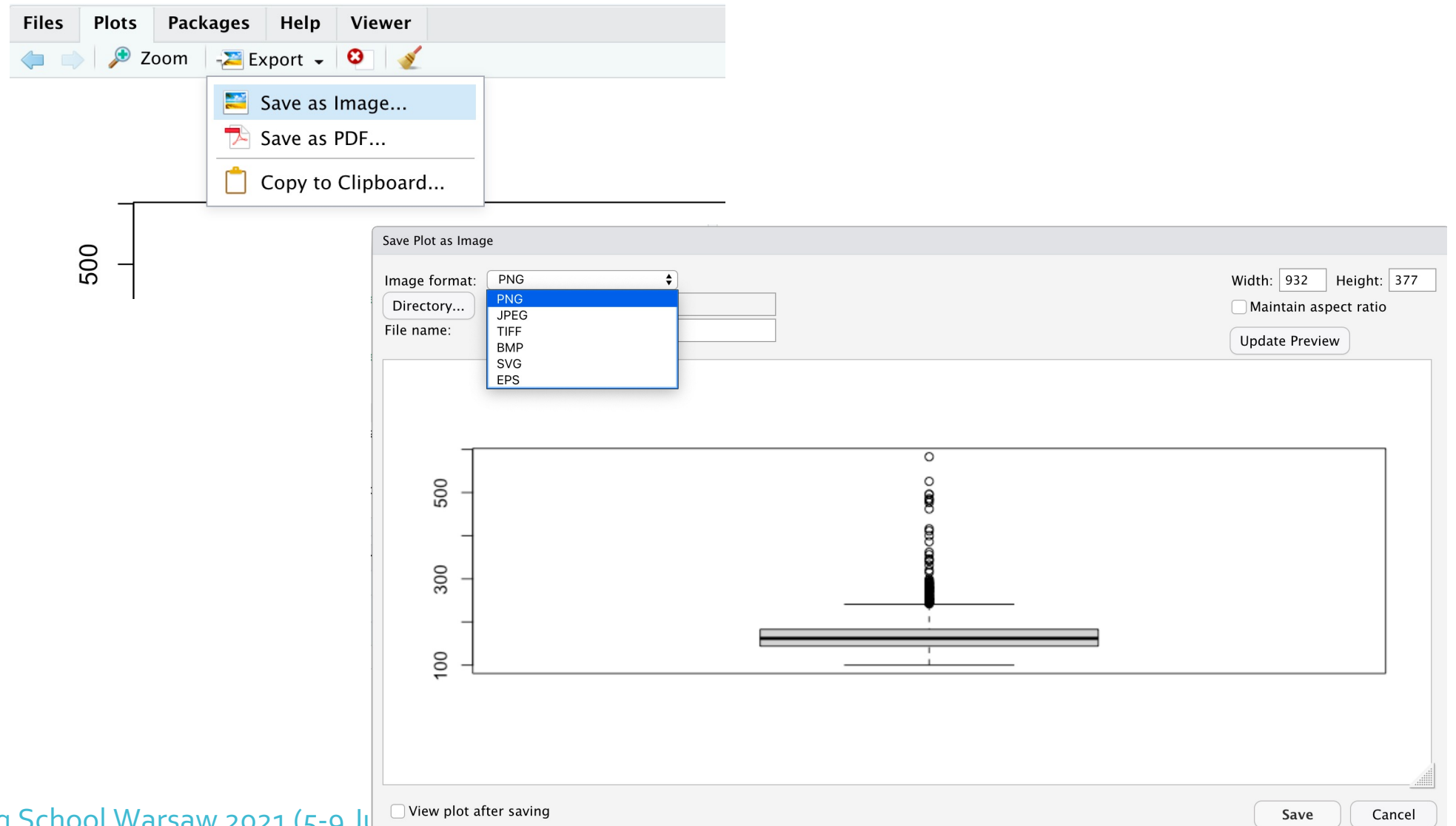
Useful to visually inspect normality of distribution

```
# plot distribution of variable values with boxplot  
boxplot(d$aoi_fixation_average_duration_ms)
```



Useful to identify outlying values

Saving graphs in RStudio graphical interface



The screenshot shows the RStudio graphical user interface. The top menu bar includes 'Files', 'Plots', 'Packages', 'Help', and 'Viewer'. The 'Export' menu is open, showing options: 'Save as Image...', 'Save as PDF...', and 'Copy to Clipboard...'. A vertical line with a bracket indicates the height of the plot area is 500. The 'Save Plot as Image' dialog box is open, showing the following details:

- Image format: PNG (selected in a dropdown menu)
- Directory: [empty field]
- File name: [empty field]
- Width: 932
- Height: 377
- Maintain aspect ratio
- Update Preview button
- View plot after saving
- Save and Cancel buttons

The dialog box contains a preview of a box plot. The y-axis of the plot is labeled with values 100, 300, and 500. The plot shows a distribution with a median around 150, a box from approximately 120 to 180, whiskers extending from 100 to 250, and several outliers above the whiskers, reaching up to 500.

Moderation analysis with ANOVA

Mixed-design Analysis of Variance (ANOVA)

Basic concept of ANOVA

- ANalysis Of VAriance generalizes t-tests to more than two groups.
- Tests variation among the means of several groups
 - Provides a statistical test of whether or not the means of several groups are all equal, or is there a difference between them?
- Null hypothesis
 - Several populations have the same means

Variability of results and its sources

- **Between group variability**
 - Differences between group means
- **Within group variability / error variance**
 - Differences between each score in the sample and the sample mean
- **Between group variance** can come from:
 - The effect of IV 😊
 - Differences between group means – our expected effect
 - Individual differences 😞
 - Everyone is different, different motivations, reactions to tasks, etc.
 - Measurement error 😞
 - Nobody's perfect – our studies neither, non standardised instructions

F - ratio

- Ratio of the between-groups variance (explained variability) to the within-groups variance (unexplained variability)

$$\text{statistic} = \frac{\text{between-groups variability}}{\text{error variability}}$$

- When the F is below or close to 1 ...
- Reducing the error variability in the denominator of the equation will result in a larger computed statistical value, thereby making it easier to reject the Null hypothesis.

Usefull libraries for ANOVA analysis

```
library(tidyverse)  
library(afex)  
library(emmeans)
```

```
> library(tidyverse)  
— Attaching packages —  
—— tidyverse 1.3.0 ——  
✓ ggplot2 3.3.3      ✓ purrr 0.3.4  
✓ tibble 3.0.5       ✓ dplyr 1.0.3  
✓ tidyr 1.1.2        ✓ stringr 1.4.0  
✓ readr 1.4.0        ✓ forcats 0.5.0
```

Load data

```
# read data frame from RData format  
load("data/all_data.Rda")
```

```
> names(d)  
[1] "ID"  
[2] "aoi_type"  
[3] "aoi_size_percents"  
[4] "aoi_fixation_total_count"  
[5] "aoi_fixation_average_duration_ms"  
[6] "aoi_fixation_ttff_ms"  
[7] "aoi_fixation_average_total_time_spent_ms"  
[8] "aoi_fixation_first_fixation_average_duration_ms"  
[9] "NEP"  
[10] "NEPsplit"
```

ID	aoi_type	aoi_size_percents	aoi_fixation_total_count	aoi_fixation_average_duration_ms
63148	negatyw	13.51776	11	155
63148	neutral	13.51776	0	0
63148	pozytyw	13.48463	2	149
45940	negatyw	13.51776	13	192
45940	neutral	13.51776	3	150
45940	pozytyw	13.48463	12	173
54370	negatyw	13.51776	10	145
54370	neutral	13.51776	2	137

- To run ANOVA in R you have plenty of options.
- We are going to use the `aov_ez()` function within the **afex package**.

Simple function for ANOVA in R

```
library(afex)
```

afex package provides convenience functions for analyzing factorial experiments using ANOVA or mixed models. `aov_ez()`, `aov_car()`, and `aov_4()` allow specification of between, within, or mixed ANOVAs for data in long format.

- Generic form of `aov_ez()` function

```
aov_ez(id = "PARTICIPANTS ID VARIABLE",  
      dv = "DEPENDENT VARIABLE",  
      data = "DATA FRAME",  
      between = "BETWEEN-SUBJECTS FACTOR",  
      within = "WITHIN-SUBJECTS FACTOR",  
      covariate = "BETWEEN-SUBJECTS COVARIATES",  
      type = "ERROR TYPE - DEFAULT SET TO 3")
```

Test the hypotheses about fixation duration

- We want to test two hypotheses:
 - In general, negative and positive pictures of environment will evoke longer fixation durations than neutral ones.
 - Negative and positive pictures of environment will evoke longer fixation durations than neutral ones BUT ONLY among people with high sensitivity to climate change.
 - *sensitivity to climate change will be a moderator of the effect of picture valence*

Run two-way mixed-design ANOVA

RUN ANOVA TEST

```
# fixation count dependent on aoi valence and NEP score
fit <- aov_ez(data = d,
             id = "ID",
             dv = "aoi_fixation_average_duration_ms",
             within = "aoi_type",
             between = "NEPsplit")
```

Data

Subject ID variable

Dependent variable

Within-subject factor

Between-subject factor

PRINT ANOVA TEST RESULTS

```
> print(fit)
Anova Table (Type 3 tests)

Response: aoi_fixation_average_duration_ms
      Effect      df    MSE      F ges p.value
1      NEPsplit    1, 42 1300.67    0.39 .005    .535
2      aoi_type 1.83, 76.89  546.00 10.54 *** .098    <.001
3 NEPsplit:aoi_type 1.83, 76.89  546.00    2.59 + .026    .086
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '+' 0.1 ' ' 1

Sphericity correction method: GG
```


Estimated means and pairwise comparisons for a main effect of picture valence

```
em <- emmeans(fit, ~aoi_type)
```

```
# post-hoc for main effect  
pairs(em)
```

```
> print(em)  
aoi_type emmean SE df lower.CL upper.CL  
neutral 127 4.19 102 118 135  
negatyw 141 4.19 102 133 150  
pozytyw 148 4.19 102 140 157
```

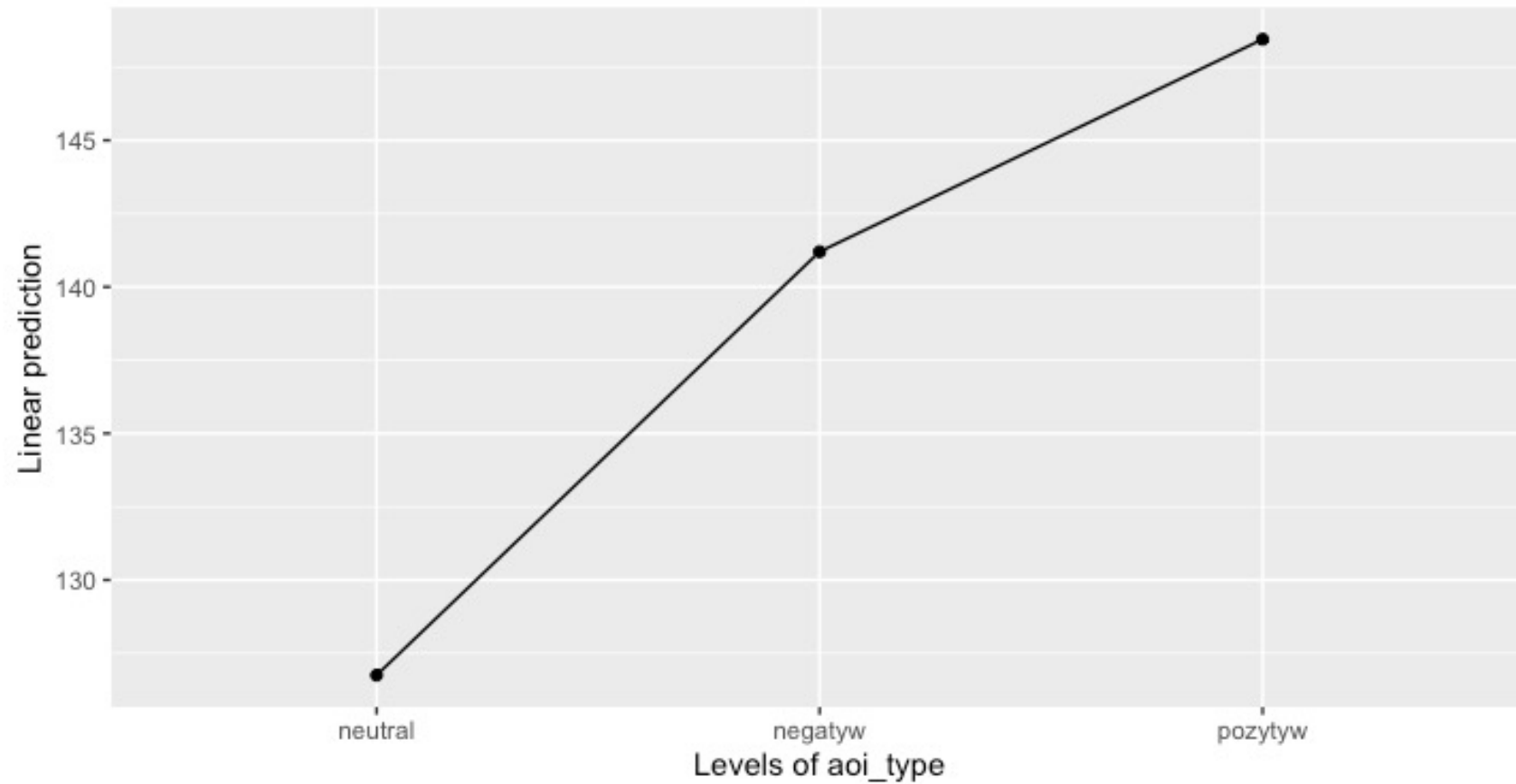
Results are averaged over the levels of: NEPsplit
Warning: EMMs are biased unless design is perfectly balanced
Confidence level used: 0.95

```
> pairs(em)  
contrast estimate SE df t.ratio p.value  
neutral - negatyw -14.45 4.81 84 -3.003 0.0098  
neutral - pozytyw -21.70 4.81 84 -4.510 0.0001  
negatyw - pozytyw -7.25 4.81 84 -1.506 0.2931
```

Results are averaged over the levels of: NEPsplit
P value adjustment: tukey method for comparing a family of 3 estimates

Analytical
graph of the
main effect

```
emmip(fit, ~ aoi_type)
```



```
em <- emmeans(fit, ~aoi_type:NEPsplit)
```

Estimated
means for
interaction
effect

```
> print(em)
```

aoi_type	NEPsplit	emmean	SE	df	lower.CL	upper.CL
neutral	low	134	6.12	107.1	122	147
negatyw	low	138	6.12	107.1	126	150
pozytyw	low	150	6.12	107.1	138	162
neutral	high	119	5.76	95.7	108	131
negatyw	high	144	5.76	95.7	133	156
pozytyw	high	147	5.76	95.7	136	158

Pairwise comparisons for the interaction effect

```
pairs(em, by = "NEPsplit")
```

```
> pairs(em, by = "NEPsplit")
```

```
NEPsplit = low:
```

contrast	estimate	SE	df	t.ratio	p.value
neutral - negatyw	-3.54	7.25	84	-0.488	0.8774
neutral - pozytyw	-15.48	7.25	84	-2.135	0.0890
negatyw - pozytyw	-11.95	7.25	84	-1.647	0.2318

```
NEPsplit = high:
```

contrast	estimate	SE	df	t.ratio	p.value
neutral - negatyw	-25.36	6.32	84	-4.011	0.0004
neutral - pozytyw	-27.91	6.32	84	-4.414	0.0001
negatyw - pozytyw	-2.55	6.32	84	-0.403	0.9144

```
P value adjustment: tukey method for comparing a family of 3 estimates
```

```
pairs(em, by = "aoi_type")
```

```
> pairs(em, by = "aoi_type")
```

```
aoi_type = neutral:
```

contrast	estimate	SE	df	t.ratio	p.value
low - high	15.38	8.43	101	1.825	0.0709

```
aoi_type = negatyw:
```

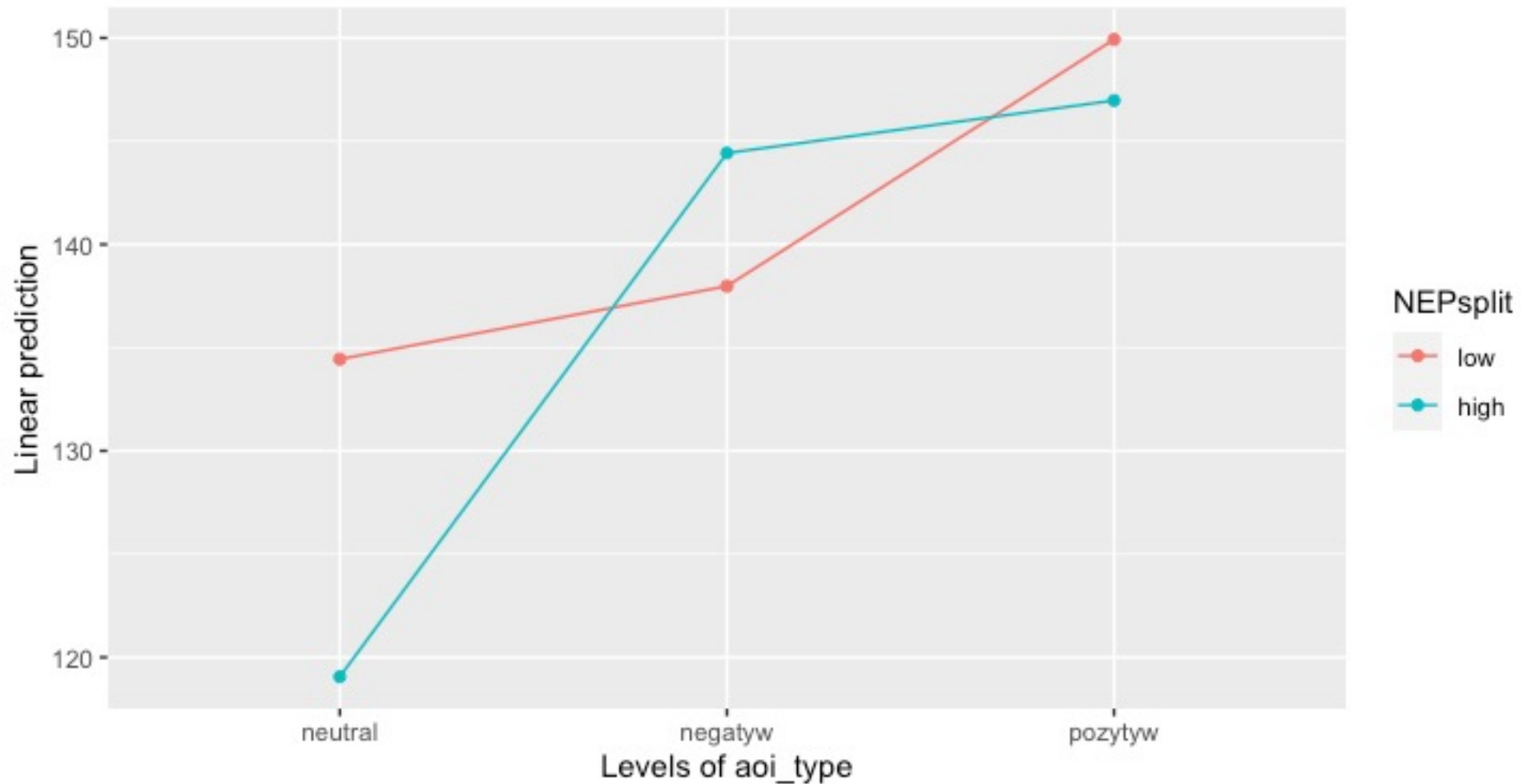
contrast	estimate	SE	df	t.ratio	p.value
low - high	-6.44	8.43	101	-0.765	0.4463

```
aoi_type = pozytyw:
```

contrast	estimate	SE	df	t.ratio	p.value
low - high	2.95	8.43	101	0.350	0.7267

Analytical
graph of the
interaction
effect

```
emmip(fit, NEPsplit ~ aoi_type)
```



Publication ready plot of interaction effect

```
tab <- summary(em)
```

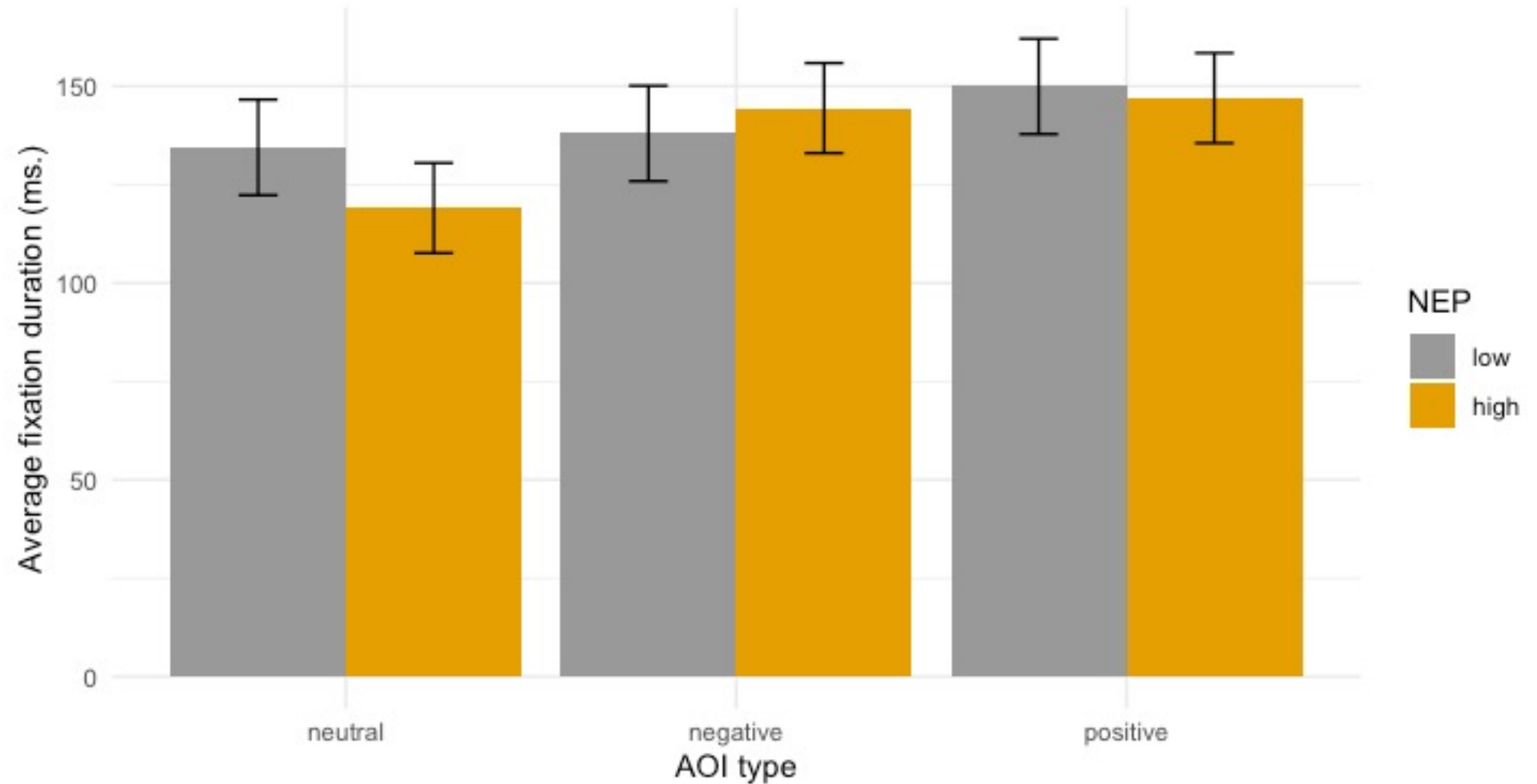
```
> print(tab)
```

aoi_type	NEPsplit	emmean	SE	df	lower.CL	upper.CL
neutral	low	134	6.12	107.1	122	147
negatyw	low	138	6.12	107.1	126	150
pozytyw	low	150	6.12	107.1	138	162
neutral	high	119	5.76	95.7	108	131
negatyw	high	144	5.76	95.7	133	156
pozytyw	high	147	5.76	95.7	136	158

```
plo <- ggplot(tab, aes(x=aoi_type, y=emmean, fill=NEPsplit)) +  
  geom_bar(position=position_dodge(), stat="identity") +  
  geom_errorbar(aes(ymin=lower.CL, ymax=upper.CL),  
                width=.2, position=position_dodge(.9)) +  
  scale_y_continuous(name = "Average fixation duration (ms.)") +  
  scale_x_discrete(name = "AOI type",  
                  labels = c("neutral", "negative", "positive")) +  
  scale_fill_manual(name = "NEP",  
                   values=c("#999999", "#E69F00")) +  
  theme_minimal()
```

Publication ready plot of interaction effect

(bargraph with error bars representing confidence intervals)



Saving the graph to the file with a code.

You can choose a wide variety of formats (jpg, png, tiff, pdf, eps, etc.)

Save the
graph to
a file

```
print(plo)
ggsave(plot = plo, filename = "graphs/AverageFixationDuration_NEP_AOI.jpg")
dev.off()
```


Test hypotheses about fixation count

- We want to test two hypotheses:
 - In general, negative and positive pictures of environment will evoke more fixations than neutral pictures
 - Negative and positive pictures of environment will evoke more fixations than neutral pictures BUT ONLY for people with high sensitivity to climate change
 - *sensitivity to climate change will be a moderator for the effect of picture valence*

Run two- way mixed- design ANOVA

RUN ANOVA TEST

```
fit <- aov_ez(data = d,  
             id = "ID",  
             dv = "aoi_fixation_total_count",  
             within = "aoi_type",  
             between = "NEPsplit")
```

Data

Subject ID variable

Dependent variable

Within-subject factor

Between-subject factor

PRINT ANOVA TEST RESULTS

```
> print(fit)  
Anova Table (Type 3 tests)  
  
Response: aoi_fixation_total_count  
          Effect          df    MSE          F ges p.value  
1          NEPsplit           1, 42 12.08          0.57 .005   .455  
2          aoi_type 1.76, 74.09 12.19 23.08 *** .260  <.001  
3 NEPsplit:aoi_type 1.76, 74.09 12.19          0.67 .010   .497  
---  
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '+' 0.1 ' ' 1  
  
Sphericity correction method: GG
```

Estimated means and pairwise comparisons for main effect

```
em <- emmeans(fit, ~aoi_type)
```

```
> print(em)
```

aoi_type	emmean	SE	df	lower.CL	upper.CL
neutral	4.19	0.507	126	3.18	5.19
negatyw	8.16	0.507	126	7.15	9.16
pozytyw	8.50	0.507	126	7.50	9.50

```
# post-hoc for main effect  
pairs(em)
```

```
> pairs(em)
```

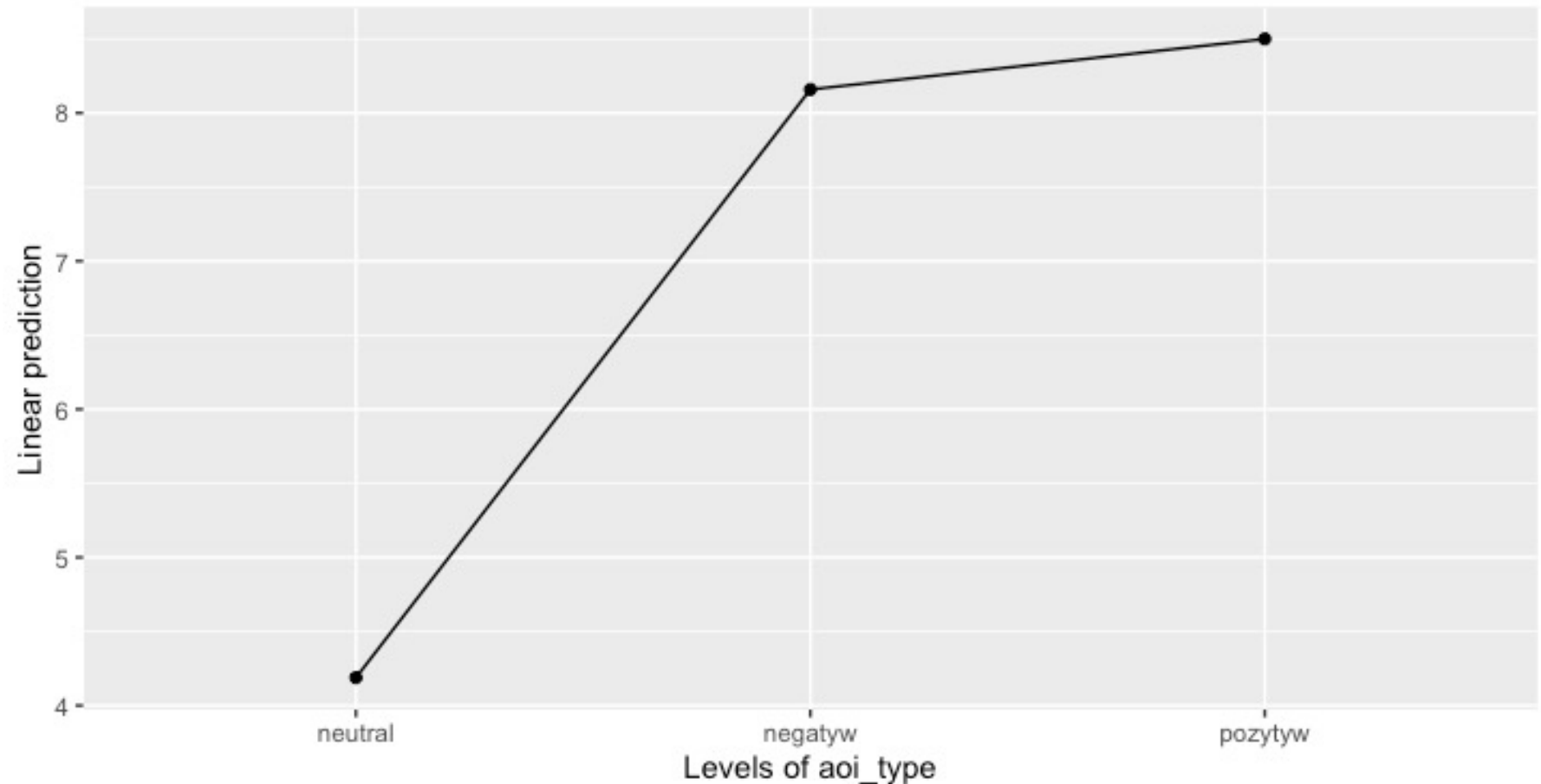
contrast	estimate	SE	df	t.ratio	p.value
neutral - negatyw	-3.971	0.706	84	-5.626	<.0001
neutral - pozytyw	-4.312	0.706	84	-6.110	<.0001
negatyw - pozytyw	-0.341	0.706	84	-0.484	0.8792

Results are averaged over the levels of: NEPsplit

P value adjustment: tukey method for comparing a family of 3 estimates

Analytical
graph of the
main effect

```
emmip(fit, ~ aoi_type)
```



Publication ready plot of the main effect

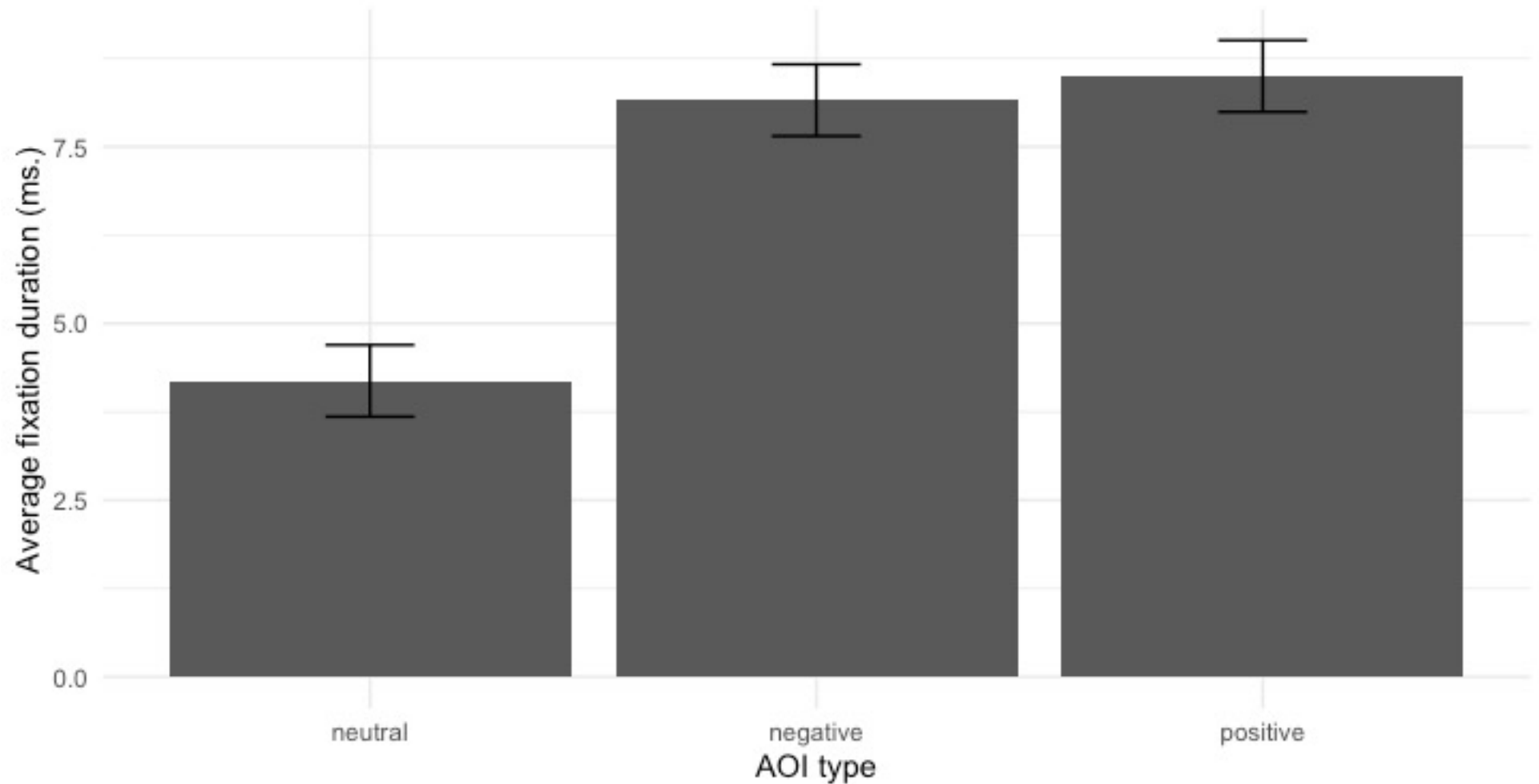
```
tab <- summary(em)
```

```
> print(tab)
```

aoi_type	emmean	SE	df	lower.CL	upper.CL
neutral	4.19	0.507	126	3.18	5.19
negatyw	8.16	0.507	126	7.15	9.16
pozytyw	8.50	0.507	126	7.50	9.50

```
plo <- ggplot(tab, aes(x=aoi_type, y=emmean)) +  
  geom_bar(position=position_dodge(), stat="identity") +  
  geom_errorbar(aes(ymin=emmean-SE, ymax=emmean+SE),  
               width=.2, position=position_dodge(.9)) +  
  scale_y_continuous(name = "Average fixation duration (ms.)") +  
  scale_x_discrete(name = "AOI type",  
                  labels = c("neutral", "negative", "positive")) +  
  theme_minimal()
```

Publication
ready plot of
the main
effect



Saving the graph to a file

You can choose a wide variety of formats (jpg, png, tiff, pdf, eps, etc.)

Save the
graph to a
file

```
print(plo)  
ggsave(plot = plo, filename = "graphs/fixationCount_A0I.jpg")  
dev.off()
```

Moderation analysis with linear regression

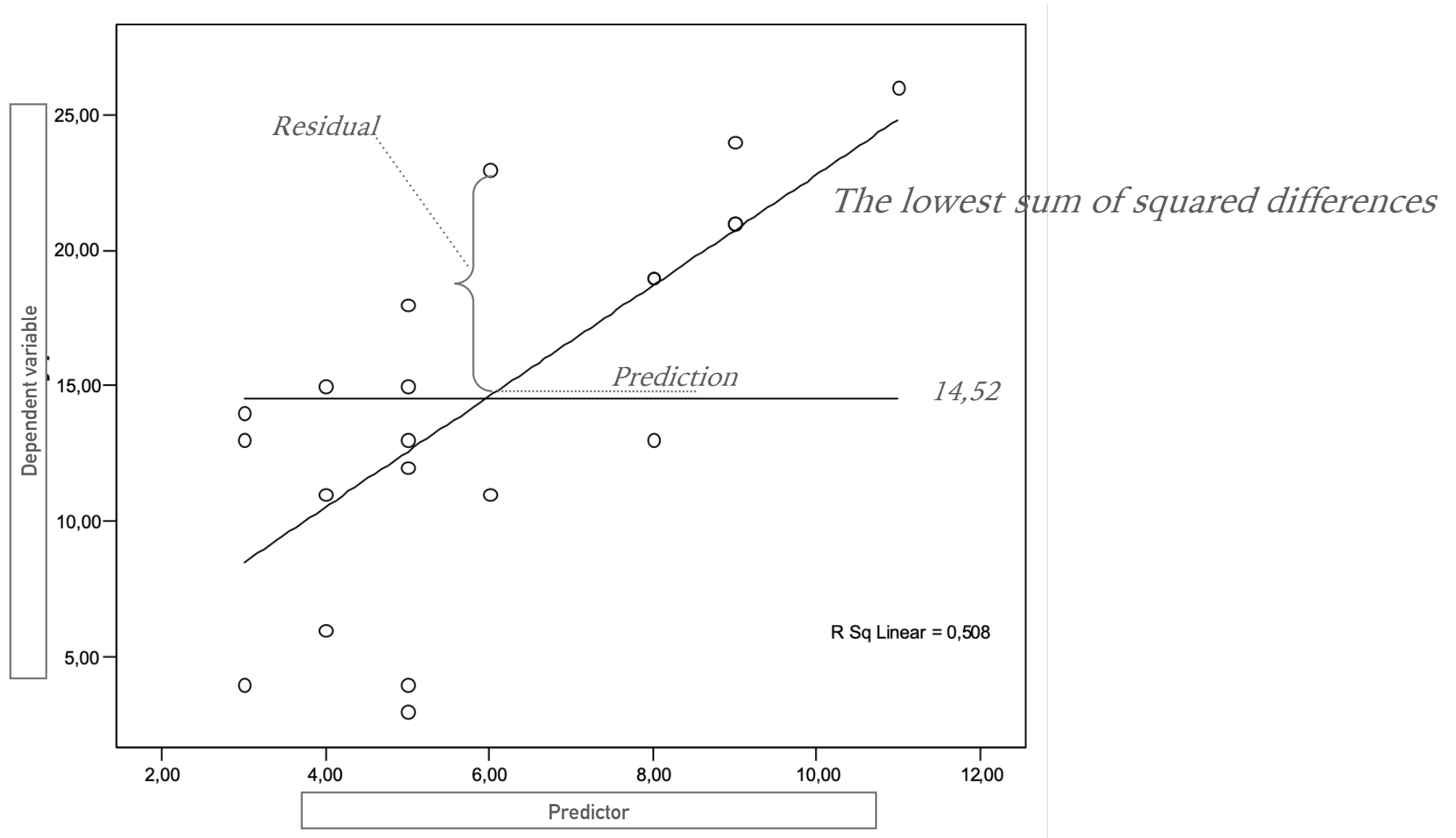
Mixed-design linear regression with continuous and nominal predictors and interaction term

Linear regression

- In regression we are fitting a model to our data
 - not loosing variance if predictor is continuous and does not need to be splitted into a categorical variable
 - predicting values of the dependent variable from one or more independent variable

$$\hat{Y} = a + bX + e$$

Least squares Method



- Each predictor has its own regression coefficient
 - For every extra predictor you include, another coefficient need to be estimated
- We are looking for linear combination of predictors that correlate maximally with the outcome variable
 - multiple regression formula with two predictors (no interaction term)

$$\hat{Y} = a + \beta_1 X_1 + \beta_2 X_2 + \epsilon$$

- multiple regression formula with two predictors and interaction term

$$\hat{Y} = a + \beta_1 X_1 + \beta_2 X_2 + \beta_3 X_1 X_2 + \epsilon$$

Multiple regression without and with interaction

Useful libraries

```
library(emmeans)
```

```
library(lme4)
```

lme4 provides functions for fitting and analyzing mixed models:
linear (lmer),
generalized linear (glmer), and
nonlinear (nlmer.)

Hypotheses

- We want to test the following hypotheses:
 - The more sensitive to climate change people are the longer fixation duration on the environment pictures
 - Positive and negative pictures of environment will evoke longer fixation duration than neutral ones
 - Sensitivity to climate change will predict fixation duration differently while looking at pictures of different valence (interaction hypothesis).

Longer form of full model definition

```
fit <- lmer(aoi_fixation_average_duration_ms ~ NEP + aoi_type + NEP:aoi_type + (1 | ID),
           data = d)
```

Data frame

Dependent variable

Predictor 1

Predictor 2

Interaction of predictors

Random intercept for subjects

Shorter form of full model definition

```
fit <- lmer(aoi_fixation_average_duration_ms ~ NEP*aoi_type + (1 | ID),
           data = d)
```

Multiple
regression.

Model
definition

Model
results.

Basic table

```
> anova(fit)
Type III Analysis of Variance Table with Satterthwaite's method
              Sum Sq Mean Sq NumDF DenDF F value  Pr(>F)
NEP              2269   2269.4     1    42  0.3989 0.53107
aoi_type         35625  17812.5     2   1932  3.1312 0.04389 *
NEP:aoi_type     51223  25611.7     2   1932  4.5022 0.01120 *
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

Model
results.

Detailed
table

```

> summary(fit)
Linear mixed model fit by REML. t-tests use Satterthwaite's method ['lmerModLmerTest']
Formula: aoi_fixation_average_duration_ms ~ NEP + aoi_type + NEP:aoi_type + (1 | ID)
Data: d

REML criterion at convergence: 22769

Scaled residuals:
    Min       1Q   Median       3Q      Max
-2.2882 -0.1971  0.2103  0.5200  5.7394

Random effects:
 Groups   Name      Variance Std.Dev.
 ID       (Intercept) 307.1   17.52
 Residual                5688.8  75.42
Number of obs: 1980, groups: ID, 44

Fixed effects:
              Estimate Std. Error      df t value Pr(>|t|)
(Intercept)  161.0240    31.1653  104.5102  5.167 1.15e-06 ***
NEP          -1.0629     0.9374  104.5102 -1.134 0.25946
aoi_typenegatyw -81.3558    32.7634 1932.0000 -2.483 0.01311 *
aoi_typepozytyw -31.8670    32.7634 1932.0000 -0.973 0.33085
NEP:aoi_typenegatyw  2.9503     0.9855 1932.0000  2.994 0.00279 **
NEP:aoi_typepozytyw  1.6500     0.9855 1932.0000  1.674 0.09424 .
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Correlation of Fixed Effects:
              (Intr) NEP    a_typn a_typp NEP:_typn
NEP          -0.992
ao_typngtyw -0.526  0.521
a_typpzytyw -0.526  0.521  0.500
NEP:_typngt  0.521 -0.526 -0.992 -0.496
NEP:_typpzy  0.521 -0.526 -0.496 -0.992  0.500
    
```

NOTE: In the report remember to provide all the values from the coefficients table: coefficient value, standard error, t-test value, degrees of freedom and p-value.

Code

```
em <- emtrends(fit, ~ NEP:aoi_type, var = "NEP")
```

Results of trends analysis

```
> print(em)
  NEP aoi_type NEP.trend   SE  df lower.CL upper.CL
  33 neutral    -1.063 0.937 105  -2.9218   0.796
  33 negatyw     1.887 0.937 105   0.0286   3.746
  33 pozytyw     0.587 0.937 105  -1.2718   2.446
```

Degrees-of-freedom method: kenward-roger
Confidence level used: 0.95

Statistical comparison of slopes

```
> pairs(em)
contrast
32.9772727272727 neutral - 32.9772727272727 negatyw    estimate   SE  df t.ratio p.value
32.9772727272727 neutral - 32.9772727272727 pozytyw    -1.65 0.986 1932 -1.674  0.2154
32.9772727272727 negatyw - 32.9772727272727 pozytyw     1.30 0.986 1932  1.319  0.3845
```

Degrees-of-freedom method: kenward-roger
P value adjustment: tukey method for comparing a family of 3 estimates

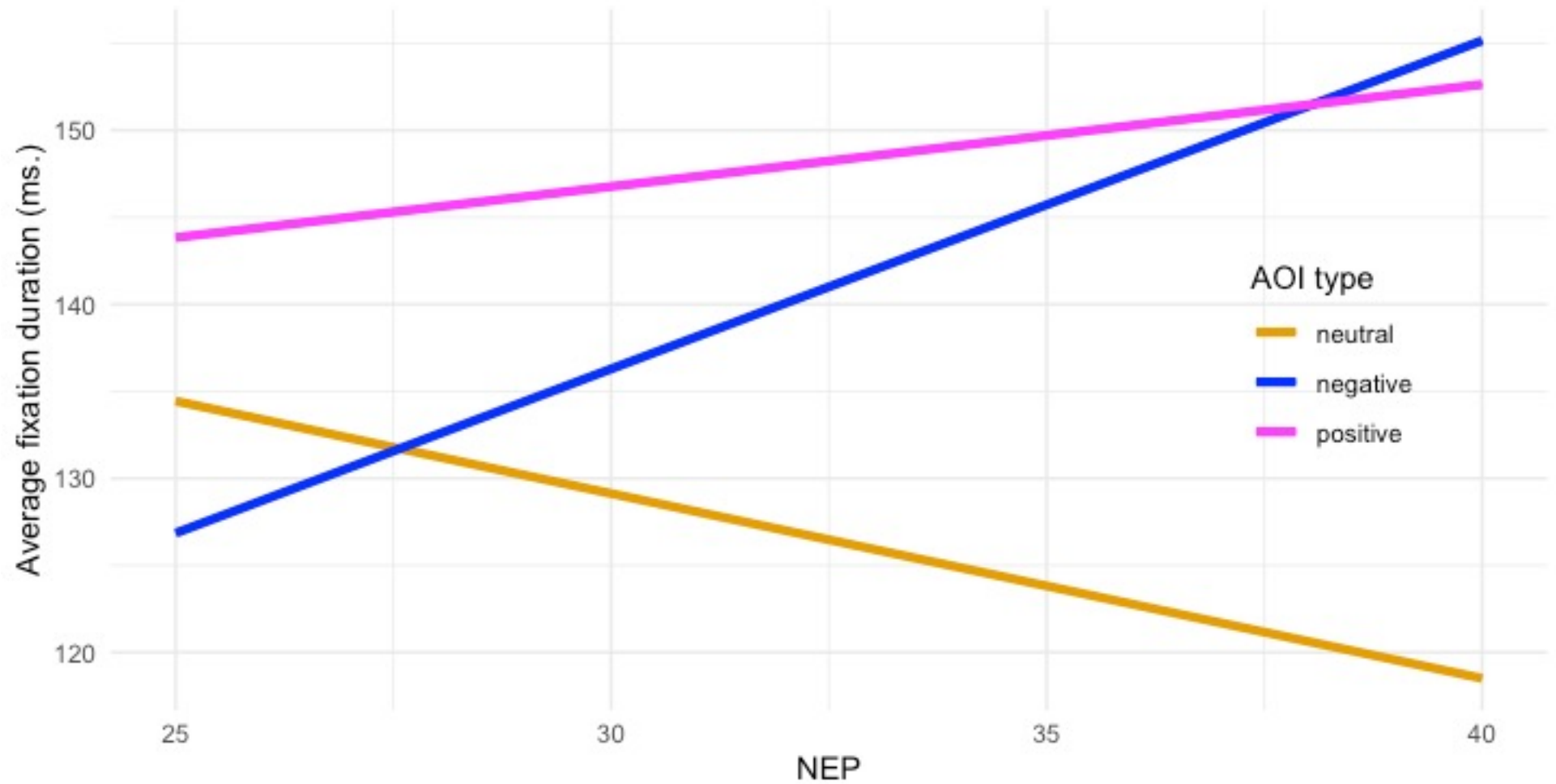
Trends
analysis for
interaction

Interaction
visualization.

Publication
ready

```
plo <- emmip(fit, aoi_type ~ NEP, cov.reduce = range) +  
  geom_line(size=1.5) +  
  scale_y_continuous(name = "Average fixation duration (ms.)") +  
  scale_x_continuous(name = "NEP") +  
  scale_color_manual(name = "AOI type",  
                     labels = c("neutral", "negative", "positive"),  
                     values = c("#E69F00", "#0000FF", "#FF00FF")) +  
  theme_minimal() +  
  theme(legend.position = c(.85, .5))
```

Save the
graph



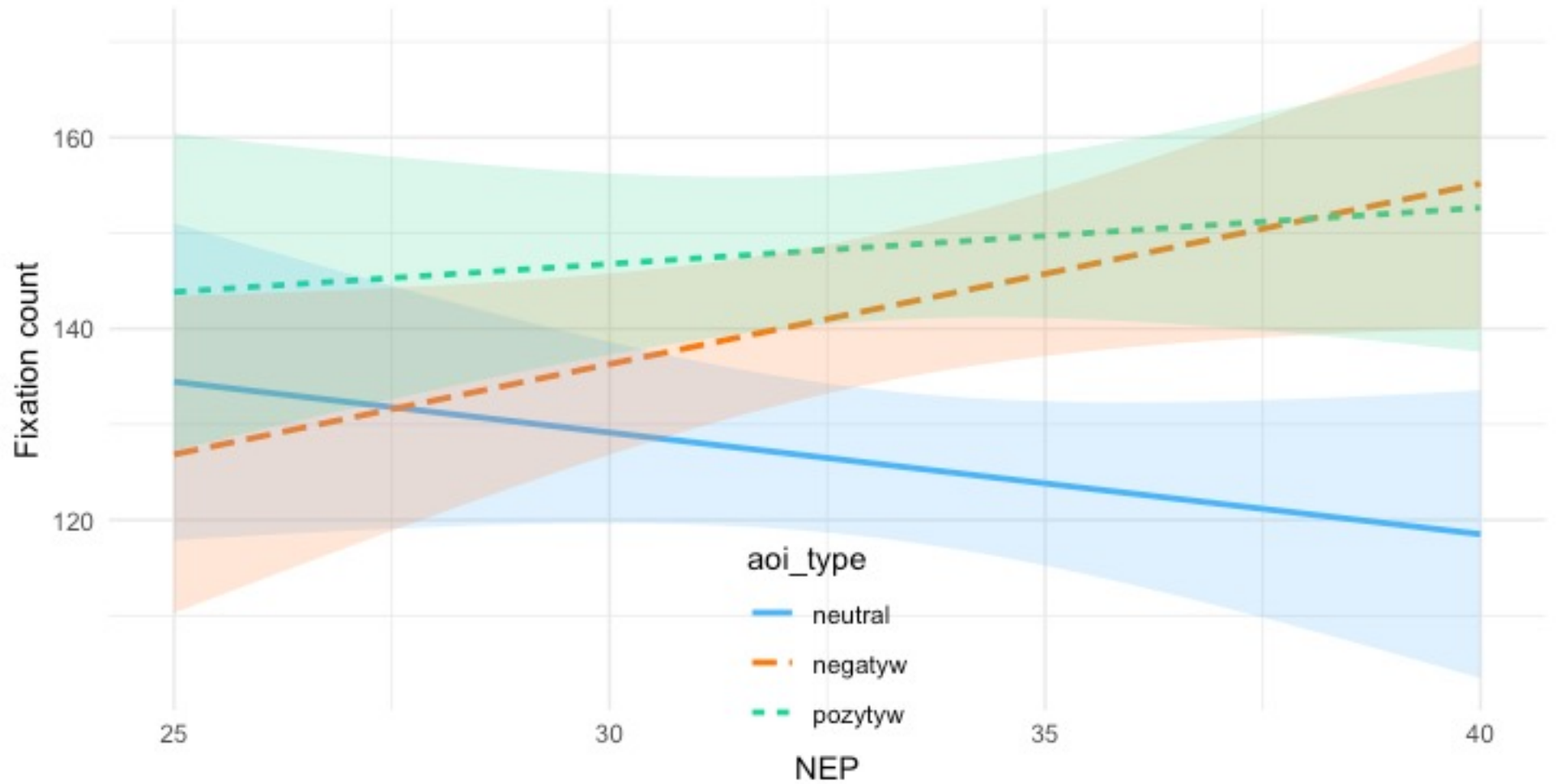
```
print(plo)
ggsave(plot = plo, filename = "graphs/fixationDuration_regression.jpg")
dev.off()
```

Alternative plot of the interaction

```
# alternatively
require(interactions)
interact_plot(fit, pred = NEP, modx = aoi_type, interval = TRUE) +
  scale_y_continuous(name = "Fixation count") +
  scale_x_continuous(name = "NEP") +
  theme_minimal() +
  theme(legend.position = c(.5, .1))
```

Alternative plot of interaction

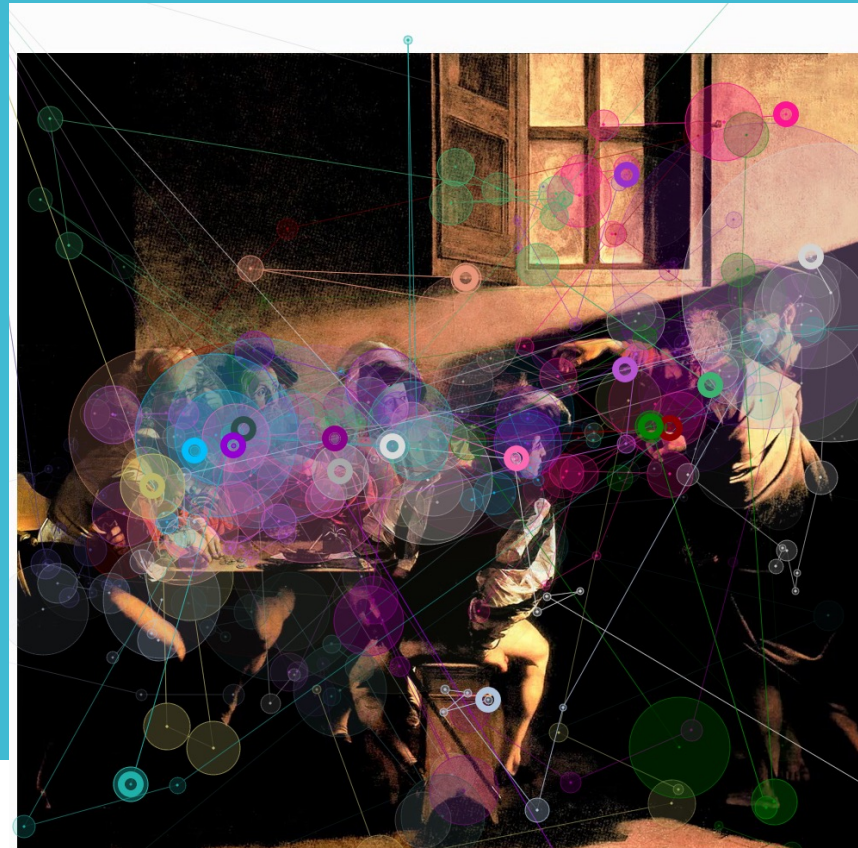
(with confidence intervals for slopes and different predefined line types)



After this
workshop ...
you are able to
perform
mixed-design
ANOVA and
multiple
regression with
interaction
effects

- **Prepare the data frames for the analysis**
 - Read/load data into R
 - Calculate new variables
 - Prepare dichotomous factor with median-split
 - Select variables and filter observations
 - Merge two data frames
 - Write data frame to a file
- **Perform mixed-design ANOVA**
 - Read and interpret the results
 - Estimate means
 - Run pairwise comparisons (post hoc tests) for significant effects
 - Prepare publication ready bar graphs
- **Perform mixed-design linear multiple regression analysis**
 - Read and interpret the results
 - Perform trends (simple slopes) analysis
 - Compare statistically simple slopes
 - Prepare publication ready line graphs with confidence intervals

It's your turn to
practice statistical
analysis during
hand-on sessions



Thank you!

Any questions?

