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3 **Main Manuscript for**

4 Appraisal of sedimentary alkenones for the quantitative  
5 reconstruction of phytoplankton biomass

6

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15

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21 Maria Raja: Methodology, Software, Validation, Formal Analysis, Investigation, Data Curation,  
22 Writing-Original Draft, Writing-Review&Editing, Visualization.

23 Antoni Rosell-Melé: Conceptualization, Validation, Resources, Writing-Review&Editing,  
24 Supervision, Funding Acquisition.

25 **This PDF file includes:**

26 Main Text  
27 Fig.s 1 to 5  
28 Table 1  
29

30 **Abstract**

31 Marine primary productivity is the driving factor in the global marine carbon cycle. Its  
32 reconstruction in past climates relies on biogeochemical proxies that are not considered to  
33 provide an unequivocal signal. These are often based on the water column flux of biogenic  
34 components to sediments (organic carbon, biogenic opal, biomarkers), although other factors  
35 than productivity are posited to control the sedimentary contents of the components, and their flux  
36 is related to the fraction of export production buried in sediments. Moreover, most flux proxies  
37 have not been globally appraised. Here we assess for the first time a proxy to quantify past  
38 phytoplankton biomass by correlating the concentration of C37 alkenones in a global suite of  
39 core-top sediments with sea-surface chlorophyll-*a* (SSchl<sub>a</sub>) estimates over the last 20 years.  
40 SSchl<sub>a</sub> is the central metric to calculate phytoplankton biomass and is directly related to primary  
41 productivity. We show that the global spatial distribution of sedimentary alkenones is primarily  
42 correlated to SSchl<sub>a</sub> rather than diagenetic factors such as the oxygen concentration in bottom  
43 waters, which challenges previous assumptions on the role of preservation on driving  
44 concentrations of sedimentary organic compounds. Moreover, our results suggest that the rate of  
45 global carbon export to sediments is not regionally constrained, and that alkenones producers  
46 play a dominant role in the global export of carbon buried in the sea-floor. This study shows the  
47 potential of using sedimentary alkenones to estimate past phytoplankton biomass, which in turn  
48 can be used to infer past primary productivity (PP) in the global ocean.

49

50 **Significance Statement**

51 Biomarker proxies, namely alkenones, are commonly used to reconstruct past primary  
52 productivity. However, their value has been questioned as they are posited to be controlled by  
53 diagenetic processes and often limited to draw qualitative inferences on the carbon cycle. In fact,  
54 most proxies have not been globally appraised with ocean biogeochemical data. Here we use a  
55 combination of remote sensing and geochemical data to provide the first spatial-based global  
56 calibration that show the use of alkenones to quantify past sea-surface chlorophyll-*a*  
57 concentration, which ultimately can be used to infer quantitatively past primary productivity in  
58 paleorecords. This calibration paves the way to clarify the relative role of the marine carbon cycle  
59 in climate variability using field data, and test biogeochemical models.

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61

62 **Main Text**

63

64 **Introduction**

65

66 Global carbon distribution between the ocean and the atmosphere regulates global climate on  
67 Earth. This distribution is primarily controlled by marine primary productivity (PP) and  
68 phytoplanktonic organisms, which transforms atmospheric CO<sub>2</sub> into organic matter. Only a  
69 fraction of this produced organic matter is exported to the deep ocean. Global models estimate  
70 that 48 PgC·yr<sup>-1</sup> are produced in ocean surface waters (1) while 6 PgC·yr<sup>-1</sup> (2) are exported out  
71 from the photic zone, and 0.15 PgC·yr<sup>-1</sup> are buried in sediments (3). Exported organic carbon is  
72 out of contact with the atmosphere on decadal to millennial timescales or longer once is buried in

73 the seafloor, which exerts a major control on global climate by regulating the partial pressure of  
74 atmospheric CO<sub>2</sub> (4). Hence, estimating marine PP, export and burial productivity changes during  
75 past key climatic periods (e.g. glacial-interglacial transitions) is essential to understand our  
76 present climate and predict its evolution in the future.

77 To infer past PP a range of proxies based on the fluxes of biogenic components are available (5–  
78 7). As flux proxies, they are related to changes in past export productivity, which are assumed to  
79 be proportional to surface PP in paleoreconstructions. However, depositional factors such as  
80 oxygen or ballasting effect are thought to be important in controlling organic matter export from  
81 the upper water column to sediments (8–10) and thus, organic proxies sedimentary  
82 concentration. The relative weights that control the spatial variability of organic matter  
83 concentration in sediments, are still unconstrained, which leads to some uncertainty on the  
84 applicability of organic matter proxies to infer PP (7). Consequently, available proxies are  
85 sometimes interpreted to infer either changes in PP or depositional conditions (11).

86 One of the common approaches to reconstruct PP relies on the measurement of C<sub>37</sub> di- and tri-  
87 unsaturated methyl ketones (i.e. C<sub>37</sub> alkenones) concentrations or fluxes in sediments (12–18).  
88 These organic molecules are biomarkers of the ubiquitous coccolithophore *Emiliana huxleyi*,  
89 which is the principal source of alkenones and the most abundant coccolithophore in the modern  
90 pelagic ocean (19–24). *Geophyrocapsa oceanica* and other coccolithophoral species from the  
91 same genera are also considered important alkenones producers nowadays (20).

92 In this study, we evaluate for the first time the potential use of sedimentary C<sub>37</sub> alkenones  
93 contents to infer past phytoplankton biomass at a global scale through the comparison of their  
94 spatial variability in a global compilation of core-top sediments with sea-surface chlorophyll-a  
95 (SSchla) (Fig. 1). This is the primary pigment of photosynthesis and is present in all  
96 photosynthetic phytoplankton species. Its concentration in surface waters is commonly used as  
97 an indicator of phytoplankton biomass and to infer primary productivity (25, 26). On a global  
98 scale, its concentration in surface waters is estimated by remote sensing (27). We also assess  
99 the effect of oxygen on the spatial accumulation of alkenones in sediments by comparing its  
100 concentration in bottom waters with alkenones abundance on a global scale.

## 101 102 103 **Results**

### 104 105 **Global comparison of SSchla with sedimentary alkenones**

106 The comparison between sedimentary alkenones and SSchla concentration estimated over the  
107 last 20 years is shown in Fig. 2. Log-transformation was needed, as the distribution of SSchla is  
108 close to lognormal in our dataset (SI Appendix, Fig. S1). This is concordant with the natural  
109 distribution of ocean chlorophyll (28). We applied standard major axis regressions (Type II  
110 regressions), which assume that both variables are measured and include some error. For this  
111 analysis we took into account that the remote sensing SSchla estimates using global standard  
112 algorithms can have a wide range of error values and specific regional biases (29–35). To  
113 minimize uncertainty in the correlation analysis, we focused our study on SSchla data located in  
114 well-performing remote sensing regions. We applied the threshold established in (36), where  
115 basins associated with lower  
116 root-mean square logarithmic errors (RMS log errors) than the global mean (31%), are  
117 considered well-performing regions. This value is within the range of 30-35% threshold values  
118 proposed in the remote sensing literature, including the Global Climate Observing System  
119 (GCOS) and the Ocean Colour Climate Change Initiative (OC-CCI), as the desired error  
120 benchmark for open ocean waters, or optically clear waters (30, 31, 36–38). Consequently, we  
121 did not included in our final analysis (see Fig. 2b) SSchla data from the North Atlantic (36, 39, 40)  
122 and the Gulf of Alaska (41). Nor from the western South American margin, as the SSchla error  
123 estimate of this region has not been thoroughly studied. Although data for the equatorial Atlantic  
124 presents a RMS log error of 48%, it lowers to 23% when removing samples located offshore of  
125 the north-eastern coast of South America (36). Thus, as our samples are not located in this  
126 region, we included samples from the equatorial Atlantic in the correlation analysis. We obtained

127 a linear correlation between C<sub>37</sub> alkenones and SSchla concentration with an RMS log error of  
128 38% and a coefficient of determination ( $R^2$ ) of 0.60 (Table 1). Hence, our data show that changes  
129 in the spatial variability of C<sub>37</sub> alkenones are mainly related to changes in SSchla.  
130 Several studies report the occurrence of regional variability in the vertical attenuation rate of  
131 organic matter flux from the surface to the deep ocean (10, 42–44). This spatial variability could  
132 lead to different regional correlations between sedimentary alkenones and SSchla concentration.  
133 To investigate this issue in our data set, we divided the data by biogeochemical regions as  
134 defined in (44), which are classified by factors influencing transfer efficiency, such as  
135 phytoplankton community, nutrient concentration and temperature. The defined regions are as  
136 follows: the tropics, the subtropics, the subarctic and the Southern Ocean. Note that the  
137 correlation for the subarctic region was not calculated due to insufficient data. The data from the  
138 subtropics show the lower RMS log error (31%) and is the region that can explain more SSchla  
139 variability (59%). The tropics present the higher RMS log error (46%), with a coefficient of  
140 determination of 0.49, and the Southern Ocean is the region that can explain less SSchla  
141 variability (33%), with a log RMS error of 35% (Table 1). However, no significant differences were  
142 found between sedimentary alkenones and SSchla correlations when comparing different oceanic  
143 biogeochemical regions, as it is shown in Fig. 3.

144

#### 145 **Global comparison of bottom water oxygen with sedimentary alkenones**

146 To study the influence of oxygen in the spatial accumulation of alkenones in sediment, we  
147 correlated our global core-top sedimentary alkenones compilation with oxygen concentration in  
148 bottom waters (Fig. 4). Data on oxygen concentration was extracted from the World Ocean Atlas  
149 2018, which provide oxygen data with a spatial resolution of 1°. The relative low resolution of the  
150 data may introduce some uncertainty in our analyses in some regions, as strong oxygen  
151 gradients may occur that are smaller than the 1° grid of the dataset, such as those located in  
152 oceanic ridges. However, our data do provide information about the influence of bottom oxygen  
153 content on alkenones abundance at regional and global scales.

154 The global comparison between sedimentary alkenones and oxygen concentration in bottom  
155 waters is shown in Fig. 4. Since our study comprises several biogeochemical regions (Fig. 1), it  
156 covers a wide range of oxygen concentration conditions (from 18.72 to 268.11  $\mu\text{mol}\cdot\text{kg}^{-1}$ ).  
157 Consequently, Fig. 4 shows different patterns that might reflect the influence of oxygen  
158 concentration in different regions. Nevertheless, we do not observe a simple global overall trend  
159 as in the case of SSchla, and we obtained a  $R^2$  of 0.02. Thus, oxygen concentration in bottom  
160 waters is not the primary cause of alkenones abundance spatial variability on a global scale.

161

162

## 163 **Discussion**

164

### 165 **Alkenones as a proxy for past phytoplankton biomass**

166 Alkenones concentration in sediments is commonly used to qualitatively infer PP changes (14,  
167 17, 18). However, as it is commonly stated in the literature, sedimentary alkenones concentration  
168 may also be affected by depositional and burial processes, which could complicate the  
169 interpretation of the proxy. Thus, alkenones concentration fluxes decrease with depth in the water  
170 column (45, 46). In here, we do not appraise directly the link between PP and alkenones  
171 sedimentary concentration, but indirectly through the use of SSchla which is a metric of  
172 phytoplankton biomass (Fig. 2). The data span several oceanic biogeochemical regions (44), and  
173 show that the spatial variability in sedimentary alkenones is primarily related to changes in  
174 SSchla, following a linear relationship with a  $R^2$  of 0.60 and RMS log error of 38% (Table 2). Our  
175 data permit to quantitatively reconstruct past SSchla concentration (equation 2). Therefore, our  
176 results confirm previous assumptions on the use of sedimentary alkenones concentrations as a  
177 proxy to qualitatively infer PP changes rather than redox conditions.

178 The correlations in Fig. 2 also allow the quantitative comparison of phytoplankton biomass data  
179 from different sites and regions through time, which hitherto was considered hindered by  
180 differences in depositional conditions between locations. Thus, one of the implications of

181 obtaining a global correlation between C<sub>37</sub> alkenones and SSchla concentrations is that the  
182 vertical rate of degradation of C<sub>37</sub> alkenones from the sea-surface to sediments follows a similar  
183 spatial pattern across different biogeochemical regions in the global ocean. We have obtained no  
184 significantly different correlations between C<sub>37</sub> alkenones and SSchla concentration for the  
185 different oceanic biogeochemical regions defined in (44) (Fig. 3). This is consistent with studies  
186 that have indicated that there are no regional differences in the vertical attenuation rate of organic  
187 matter flux (47–49), even though other studies have found spatial variability in the export  
188 efficiency of surface biomass to the deep ocean (10, 42–44).  
189 Hence, our data support the results obtained in (50) and other earlier papers, which showed that  
190 spatial variability in the vertical flux of particulate organic carbon decreases with increasing water  
191 column depth. Our results are also in agreement with (51), who showed that the proportion of  
192 primary productivity that reaches the deep sea does not vary greatly with latitude in the North  
193 Atlantic. Besides, these results show that the global correlation is independent of the  
194 biogeographic region, which implies that it is applicable globally despite any changes in  
195 biogeochemical regions through time. In this sense, we assume the similar pattern of the vertical  
196 degradation rate over different biogeochemical regions is also maintained after burial processes.  
197 Future studies will evaluate to which extent subsurface degradation process might significantly  
198 influence or bias the relationships described in Fig.s 2 and 3, and constrain their application in  
199 paleo studies.

200

#### 201 **The relevant role of alkenones producers in the global carbon export**

202 In contrast to SSchla, alkenones are mainly produced by coccolithophores (*E. huxleyi* and  
203 *Geophyrocapsa species*) (20). Therefore, sedimentary alkenones concentration are often  
204 considered an exclusive coccolithophore productivity proxy, which may not provide information  
205 about total past phytoplanktonic biomass changes (15, 18, 52, 53). In contrast, C<sub>37</sub> alkenones  
206 concentration were also suggested to track PP not only from coccolithophores, but also from the  
207 wider phytoplanktonic community in upwelling tropical areas (12). Subsequently, such  
208 interpretation was corroborated at two more sites: ODP Site 982 in the North Atlantic and ODP  
209 Site 846 in the eastern tropical Pacific (54). In spite of their findings, the authors suggested a  
210 linear relationship would be not expected in oligotrophic areas, where alkenone-synthesising  
211 coccolithophores constitute a small proportion of the coccolithophore population.

212 Contrary to previous assumptions, our global correlation shows that C<sub>37</sub> alkenones sedimentary  
213 concentrations can be interpreted as a proxy for total phytoplankton biomass in different  
214 biogeochemical regions (Fig. 2), and suggests that there are no differences between samples  
215 located in oligotrophic (SSchla annual average < 0.1 mg·m<sup>-3</sup>) and non-oligotrophic regimes (SI  
216 Appendix, Fig. S2).

217 Obtaining a global correlation between SSchla and alkenones indicates a proportionality between  
218 total phytoplankton biomass and alkenones accumulation in sediments. In terms of total  
219 phytoplankton biomass and PP, photosynthetic picoeukaryotes, which include *E. huxleyi* and *G.*  
220 *oceanica*, have been reported to be the dominant contributors. For instance, photosynthetic  
221 picoeukaryotes are responsible for more than the half the total PP in the North Atlantic (68%)  
222 (55), southern California Bight (76%) (56), the eastern northatlantic subtropical gyre (54%) (57),  
223 the southern Bay of Biscay (51%) (58) and the South East Pacific Ocean (>60%) (59).

224 However, there is not a consensus on which class of phytoplankton is the dominant carbon  
225 exporter to the sea floor. Large phytoplankton, such as diatoms, were recognised to dominate  
226 global carbon export to sediments (60, 61) and picoplankton (photosynthetic picoeukaryotes and  
227 cyanobacteria) were claimed not to contribute significantly to carbon export because of their small  
228 size (62). In contrast, more recent studies have suggested that picoplankton plays a major role in  
229 the sedimentary carbon fluxes. For instance, massive picoplankton sedimentation in east of New  
230 Zealand (63), and picoplankton export carbon fluxes dominance were found in the eastern  
231 equatorial Pacific (64) and in the Arabian sea (65). Besides, subsequent studies showed that the  
232 downward flux of organic carbon via small particles often constitutes the bulk of the total

233 particulate organic carbon flux in the North Atlantic (66), and that the export of small particles in  
234 the Norwegian Sea contributed to long-term carbon sequestration (67).  
235 We are not aware of any evidence that endorse the notion that alkenones producers are one of  
236 the main contributors to global carbon sequestration. However, our results in Fig. 2 indicate that  
237 alkenones producers export and burial contribution is proportional to total phytoplankton biomass  
238 at global scale. Our results are in agreement to previous findings that show primary productivity to  
239 be proportional to picoplankton biomass export from the euphotic zone at two different locations:  
240 the equatorial Pacific and the Arabian Sea (68). We suggest that such proportionality is indicative  
241 of the putative dominant role of photosynthetic picoeukaryotes to global carbon sequestration and  
242 burial in the sea-floor.

243

#### 244 **The role of oxygen in the spatial accumulation of alkenones in sediment**

245 Oxygen enhances organic matter degradation and its concentration in bottom waters is  
246 considered a key factor in the preservation and accumulation of organic matter in sediments (69–  
247 73). However, despite its strong influence in the vertical degradation of organic matter, its role in  
248 the global spatial distribution of organic matter concentration remains unconstrained.  
249 Some studies located in oxygen minimum zones, such as the Arabian Sea, reported empirical  
250 relationships between oxygen concentration in bottom waters and sedimentary organic matter or  
251 alkenones concentration (69, 74–76). Although these relationships only show a correlation when  
252 oxygen is present at very low concentrations ( $\leq 50 \mu\text{mol}\cdot\text{kg}^{-1}$ ), alkenones concentration is  
253 sometimes used to reconstruct preservation conditions in other environments, such as the Pacific  
254 and the Atlantic oceans (11, 77, 78).

255 Our results in Fig. 4 show that there is not a straightforward relationship between the spatial  
256 concentration patterns of sedimentary alkenones and bottom water oxygen. In contrast to  
257 previous studies, our data are located in different oceanic biogeochemical regions and include  
258 low and high oxygen content environments (from 19 to  $268 \mu\text{mol}\cdot\text{kg}^{-1}$ ). Therefore, the wide range  
259 of oxygen concentration presented in this study would be much more representative of the global  
260 ocean conditions than those previously evaluated. These results can be interpreted as evidence  
261 of the minor role of oxygen concentration in the spatial distribution of sedimentary alkenones for  
262 the greater part of the ocean. Thus, our data do not support the global use of alkenones  
263 concentration to reconstruct preservation conditions in the deep ocean.

264

#### 265 **Conclusions**

266 The primary driver explaining the global spatial distribution of sedimentary alkenones is  
267 phytoplankton biomass as reflected by SSchl<sub>a</sub> concentration. Different biogeographic regions do  
268 not show significant differences in their correlations between sedimentary alkenones and SSchl<sub>a</sub>  
269 concentration. Moreover, oxygen concentration in bottom waters does not strongly influence the  
270 spatial concentration of alkenones in sediments on a global scale. Hence, our results do support  
271 the use of alkenones sedimentary abundance to study primary productivity, and do not support  
272 the use of alkenones as an indicator of preservation conditions in the deep ocean. Besides, our  
273 data indicate that the global vertical attenuation rate of organic matter flux from sea-surface to  
274 sediments is not regionally constrained, and suggest that the global carbon export and burial is  
275 dominated by photosynthetic picoeukaryotes.

276 Thus, this study provides the first spatial-based global calibration that show the use of  
277 sedimentary alkenones to quantify sea-surface chlorophyll-*a*, which ultimately can be used to  
278 infer quantitatively PP in paleorecords. This calibration is independent of biogeographic regions,  
279 which implies that besides relative PP changes through downcore, it can also be used for the  
280 spatial comparison of PP between different locations at any site in the global ocean. Furthermore,  
281 it is applicable despite any putative changes in biogeochemical regions through time.

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#### 285 **Materials and Methods**

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### **Sedimentary alkenones**

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We compiled alkenone data from 226 locations widely distributed around the global ocean, and across diverse biogeochemical regions as defined in (44) (Fig. 1). Most of the data are obtained from previous published studies (95% of the compilation), while the rest were analysed in our laboratory following the methodology explained hereafter. We assume that the interlaboratory reproducibility of the different available methods to measure the absolute alkenones abundance in sediments is 32% (79). The samples in the compilation generally correspond to the upper 1 cm of a sediment core, and approximately 90% of them were retrieved using devices specially designed for a minimum disturbance of the sediment surface, i.e. box corer or multicorer. The compilation appears to be dominated by samples along continental margins, since they represent the most intensively studied regions. However, sample sites span a wide range of alkenones concentration distributed in the diverse biogeochemical regions defined in Weber et al. (2016) around the global ocean (Fig. 1). In addition, over 80% of the compiled surface sediments are in sites with a water column deeper than 1,000 meters, while over 90% of the samples were retrieved from waters deeper than 500 meters (SI Appendix, Dataset S1). Consequently, the compilation is mainly representative of the water column of the open ocean. We estimate that in average, a sedimentary sample represents a time span of sedimentation of ca. 71 years. This has been obtained from the sedimentation rate average of all sediments ( $14 \text{ cm}\cdot\text{ka}^{-1}$ ) using the map published in (80). However, variability of sedimentation rates could play a role in driving the concentration of alkenones in sediments. Consequently, to analyse changes in the flux of a chemical or substance to the sea, the mass accumulation rates (MAR) must be used instead of concentrations. Alkenones mass accumulation rates (MAR<sub>alkenones</sub>) were estimated by multiplying alkenones concentration by sedimentation rate and dry bulk density. In the absence of these data for each sample site, we extracted sedimentation rates from two global maps previously published (51, 80) and discussed in (81). However, sedimentation rates could only be extracted for 54% of the sites. Dry bulk densities were assumed to be  $0.9 \text{ g}\cdot\text{cm}^{-3}$ , as it corresponds to the mean dry bulk density for marine sediments in the global sediment core database published in (81), which has been created by retrieving available data from online data repositories. It is important to point out that these estimated accumulation rates are highly uncertain, given that sedimentation rates can vary significantly over relatively short distances on the seafloor due to winnowing and focusing, and the locations of cores are often biased towards the highest accumulation rates in the search of retrieving records with high temporal resolution. Nonetheless, in the absence of more accurate data, we include the estimates to provide a first-order information on the effect of sedimentation rates in our compilation. We compared MAR<sub>alkenones</sub> against alkenones concentration in SI Appendix, Fig. S3. The correlation between the two variables is high as attested by their coefficient of determination of  $R^2=0.92$  and  $R^2=0.87$ , depending on the sources of sedimentation rates (51, 80). Consequently, overall for our compilation, the differences in using MAR<sub>alkenones</sub> and alkenones concentration when compared to values of SSchl<sub>a</sub> concentration are not likely to lead to significantly different results. To maximize the size of data points in our compilation we used the alkenones concentration rather than its MAR.

In the samples analysed for the study, sediments (1-4 g) were extracted using 10 mL of a mixture of dichloromethane and methanol (3:1) (GC grade, Suprasolv) in a MARS5 microwave accelerated extraction system (CEM Corporation). Before extraction,  $25 \text{ ng}\cdot\mu\text{L}^{-1}$  of the internal standard 2-nonadecanone (Fluka, purity 0.97%) was added to the sediment placed in a Teflon vessel. During extraction the mixture was stirred continuously with a magnetic bar, while temperature was increased from ambient to  $70^\circ\text{C}$  for 2.5 minutes and left at this temperature for a further 5 minutes. After extraction, samples' vessels were left to cool down at room temperature, and the supernatants were decanted into glass tubes and centrifuged. Extracts were then dried under vacuum and cleaned up using  $\text{SiO}_2$  columns eluted sequentially with hexane (first fraction) and dichloromethane (second fraction). Extracts were dried again and  $50 \mu\text{L}$  of isooctane was added to the second fraction before their analysis with a gas chromatograph (GC) fitted with a



340 flame ionization detector at 320°C (Agilent Technologies 7820A GC System). Samples were  
341 injected in splitless mode in a GC column (HP-1 GC column, 60m length, 250µm internal  
342 diameter, 0.25µm film thickness) with a flow rate of 1.5 mL·min<sup>-1</sup>. The GC method consists of 2  
343 ramps, the first one from 80°C to 120°C with a temperature rate of 30 °C·min<sup>-1</sup>, followed by a  
344 second ramp that increases temperature at 6°C min<sup>-1</sup> until 320°C, and held at this temperature  
345 during 21 minutes.

346  
347

### 348 **Ocean surface chlorophyll**

349 Satellite observation of ocean-colour, in the open ocean, is primarily dependent on phytoplankton  
350 occurrence. These unicellular algae contain photosynthetic pigments, primarily chlorophyll-a,  
351 which coexist together with other pigments, associated detrital, and coloured dissolved organic  
352 matter (82). To infer SSchla from ocean-colour data, many algorithms have been constructed  
353 relating characteristics of the water signal to the property of interest (83). There are large  
354 uncertainties in the estimation of SSchla in some regions, particularly where the optical and  
355 biological properties are complex (84, 85). For instance, some of the factors that affect the  
356 properties of the surface waters are coloured dissolved organic matter, radiance-absorbing  
357 aerosols, phytoplankton species diversity, suspended sediments and minerals, clouds, ice, sun  
358 glint, and navigation/time space mismatches (86–89). Furthermore, desert dust (90) and bubbles  
359 (91) make the water appear greener. Other sources of scatter are the remote sensing limitation of  
360 detecting SSchla at different depths in the water column (92, 93).

361 Global SSchla algorithms, calibrated with global *in situ* datasets, show average root mean square  
362 (RMS) log errors of 34% (29). The Global Climate Observing System uncertainty requirement for  
363 SSchla is 30% ([www.ncdc.noaa.gov/gosic/gcos-essential-climate-variable-ecv-data-access-](http://www.ncdc.noaa.gov/gosic/gcos-essential-climate-variable-ecv-data-access-matrix/gcos-ocean-biogeochemistry-ecv-ocean-color)  
364 [matrix/gcos-ocean-biogeochemistry-ecv-ocean-color](http://www.ncdc.noaa.gov/gosic/gcos-essential-climate-variable-ecv-data-access-matrix/gcos-ocean-biogeochemistry-ecv-ocean-color)). End-users of SSchla data commonly *quote*  
365 uncertainty requirement of 35% (83). There are a number of challenges in the calibration of  
366 remote sensing data, like the diversity of optical properties in various water types, comparability  
367 of ocean *in situ* measurements using different analytical approaches, and the different spatial and  
368 temporal scales of the ocean-in situ and satellite measurements (83).

369 The available ocean colour satellite data since 1997 have been merged in the GlobColour project  
370 (<http://globcolour.info>), which combines measurements from the Sea-viewing Wide Field-of-view  
371 Sensor (SeaWiFS; 1997-2010), the MEdium Resolution Imaging Spectrometer (MERIS; 2002-  
372 2012), the MODerate-resolution Imaging Spectroradiometer (MODIS; 2002-2017), and the Visible  
373 Infrared Imaging Radiometer Suite (VIIRS; 2012-2017) missions. The spatial resolution of the  
374 available data for end-users is 1/24° (4.63 km at the equator). The algorithms used for obtaining  
375 SSchla are OC4v5 for SeaWiFS, OC4Me for MERIS and OC3v5 for MODIS and VIIRS (94).

376 SSchla values for every sediment location were extracted from merged global-scale matrices of  
377 the Globcolour Project, which contain monthly data. For this paper, we extracted SSchla  
378 concentration data from September 1997 to December 2017 for every core location. GlobColour  
379 data used in this study have been developed, validated, and distributed by ACRI-ST, France. We  
380 used the Matlab script provided by the Monterey Bary Aquarium Research Institute  
381 (<https://www.mbari.org/index-of-downloadable-files/>) to calculate the standard major axis  
382 regressions (type II regressions) between SSchla and sedimentary alkenones.

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398 **The authors declare no competing interest.**

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## References

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1. J. L. Sarmiento, N. Gruber, *Ocean Biogeochemical Dynamics* (Princeton Univ. Press, 2006) <https://doi.org/10.1063/1.2754608>.

404

2. D. A. Siegel, *et al.*, Global assessment of ocean carbon export by combining satellite observations and food-web models. *Global Biogeochem. Cycles* **28**, 181–196 (2014).

405

3. F. E. Muller-Karger, *et al.*, The importance of continental margins in the global carbon cycle. *Geophys. Res. Lett.* **32**, 1–4 (2005).

406

4. P. W. Boyd, H. Claustre, M. Levy, D. A. Siegel, T. Weber, Multi-faceted particle pumps drive carbon sequestration in the ocean. *Nature* **568**, 327–335 (2019).

407

5. K. E. Kohfeld, C. Le Quéré, S. P. Harrison, R. F. Anderson, Role of marine biology in glacial-interglacial CO<sub>2</sub> cycles. *Science (80-. )*. **308**, 74–78 (2005).

408

6. M. Zhao, J. L. Mercer, G. Eglinton, M. J. Higginson, C. Y. Huang, Comparative molecular biomarker assessment of phytoplankton paleoproductivity for the last 160 kyr off Cap Blanc, NW Africa. *Org. Geochem.* **37**, 72–97 (2006).

409

7. S. D. Schoepfer, *et al.*, Total organic carbon, organic phosphorus, and biogenic barium fluxes as proxies for paleomarine productivity. *Earth-Science Rev.* **149**, 23–52 (2015).

410

8. T. DeVries, T. Weber, The export and fate of organic matter in the ocean: New constraints from combining satellite and oceanographic tracer observations. *Global Biogeochem. Cycles* **31**, 535–555 (2017).

411

9. R. Francois, S. Honjo, R. Krishfield, S. Manganini, Factors controlling the flux of organic carbon to the bathypelagic zone of the ocean. *Glob. Biogeochem. Cycles* **16**, 1087 (2002).

412

10. S. A. Henson, R. Sanders, E. Madsen, Global patterns in efficiency of particulate organic carbon export and transfer to the deep ocean. *Global Biogeochem. Cycles* **26**, 1–14 (2012).

413

11. R. F. Anderson, *et al.*, Deep-Sea Oxygen Depletion and Ocean Carbon Sequestration During the Last Ice Age. *Global Biogeochem. Cycles* **33**, 301–317 (2019).

414

12. C. T. Bolton, *et al.*, Glacial-interglacial productivity changes recorded by alkenones and microfossils in late Pliocene eastern equatorial Pacific and Atlantic upwelling zones. *Earth Planet. Sci. Lett.* **295**, 401–411 (2010).

415

13. K. T. Lawrence, Evolution of the Eastern Tropical Pacific Through Plio-Pleistocene Glaciation. *Science (80-. )*. **312**, 79–83 (2006).

416

14. A. Moreno, I. Cacho, M. Canals, J. O. Grimalt, A. Sanchez-Vidal, Millennial-scale variability in the productivity signal from the Alboran Sea record, Western Mediterranean Sea. *Palaeogeogr. Palaeoclimatol. Palaeoecol.* **211**, 205–219 (2004).

417

15. B. Petrick, *et al.*, Oceanographic and climatic evolution of the southeastern subtropical Atlantic over the last 3.5 Ma. *Earth Planet. Sci. Lett.* **492**, 12–21 (2018).

418

16. F. G. Prahl, R. B. Collier, J. Dymond, M. Lyle, M. A. Sparrow, A biomarker perspective on prymnesiophyte productivity in the northeast pacific ocean. *Deep. Res. Part I* **40**, 2061–2076 (1993).

419

17. F. Rostek, E. Bard, L. Beaufort, C. Sonzogni, G. Ganssen, Sea surface temperature and productivity records for the past 240kyr in the Arabian Sea. *Deep. Res.* **44**, 1461–1480 (1997).

420

18. O. Seki, *et al.*, Reconstruction of paleoproductivity in the Sea of Okhotsk over the last 30 kyr. *Paleoceanography* **19** (2004).

421

- 446 19. S. Schmidt, *et al.*, Particle export during a bloom of *Emiliana huxleyi* in the North-West  
447 European continental margin. *J. Mar. Syst.* **109–110**, S182–S190 (2013).
- 448 20. J. K. Volkman, S. M. Barrett, S. I. Blackburn, E. L. Sikes, Alkenones in *Gephyrocapsa*-  
449 *Oceanica* - Implications for Studies of Paleoclimate. *Geochim. Cosmochim. Acta* **59**, 513–  
450 520 (1995).
- 451 21. J. K. Volkman, G. Eglinton, E. D. S. Corner, T. E. V. Forsberg, Long-chain alkenes and  
452 alkenones in the marine coccolithophorid *Emiliana huxleyi*. *Phytochemistry* **19**, 2619–  
453 2622 (1980).
- 454 22. I. T. Marlowe, *et al.*, Long chain (n-c37-c39) alkenones in the prymnesiophyceae.  
455 distribution of alkenones and other lipids and their taxonomic significance. *Br. Phycol. J.*  
456 **19**, 203–216 (1984).
- 457 23. L. L. Rhodes, B. M. Peake, A. L. MacKenzie, M. Simon, A. R. Edwards, Coccolithophores  
458 *Gephyrocapsa oceanica* and *Emiliana huxleyi* (Prymnesiophyceae = Haptophyceae) in  
459 New Zealand's coastal waters: Characteristics of blooms and growth in laboratory culture.  
460 *New Zeal. J. Mar. Freshw. Res.* **29**, 345–357 (1995).
- 461 24. M. H. Conte, J. K. Volkman, G. Eglinton, Lipid biomarkers of the Prymnesiophyceae. *The*  
462 *Haptophyte Algae* **51**, 351–377 (1994).
- 463 25. M. J. Behrenfeld, E. Boss, D. A. Siegel, D. M. Shea, Carbon-based ocean productivity and  
464 phytoplankton physiology from space. *Global Biogeochem. Cycles* **19**, 1–14 (2005).
- 465 26. Y. Huot, *et al.*, Relationship between photosynthetic parameters and different proxies of  
466 phytoplankton biomass in the subtropical ocean. *Biogeosciences* **4**, 853–868 (2007).
- 467 27. Z. Lee, J. Marra, M. J. Perry, M. Kahru, Estimating oceanic primary productivity from  
468 ocean color remote sensing: A strategic assessment. *J. Mar. Syst.* **149**, 50–59 (2015).
- 469 28. J. W. Campbell, The lognormal distribution as a model for bio-optical variability in the sea.  
470 *J. Geophys. Res.* **100**, 13237–13254 (1995).
- 471 29. R. J. W. Brewin, *et al.*, Uncertainty in Ocean-Color Estimates of Chlorophyll for  
472 Phytoplankton Groups. *Front. Mar. Sci.* **4** (2017).
- 473 30. S. Dutkiewicz, A. E. Hickman, O. Jahn, Modelling ocean-colour-derived chlorophyll a.  
474 *Biogeosciences* **15**, 613–630 (2018).
- 475 31. Global Climate Observing System, “Systematic Observation Requirements For Satellite-  
476 Based Data Products for Climate - 2011 Update” (2011).
- 477 32. IOCCG, *Partition of the Ocean into Ecological Provinces : Role of Ocean-Colour*  
478 *Radiometry*, M. Dowell, T. Platt, Eds. (Reports of the International Ocean-Colour  
479 Coordinating Group, No. 9, IOCCG, 2009).
- 480 33. T. Jackson, S. Sathyendranath, F. Mélin, “An improved optical classification scheme for  
481 the Ocean Colour Essential Climate Variable and its applications” (Elsevier Inc., 2017).
- 482 34. T. Moore, J. Campbell, M. Dowell, A class-based approach to characterizing and mapping  
483 the uncertainty of the MODIS ocean chlorophyll product. *Remote Sens. Environ.* **113**,  
484 2424–2430 (2009).
- 485 35. I. Shulman, *et al.*, Bio-Optical Data Assimilation With Observational Error Covariance  
486 Derived From an Ensemble of Satellite Images. *J. Geophys. Res. Ocean.* **123**, 1801–1813  
487 (2018).
- 488 36. W. W. Gregg, N. W. Casey, Global and regional evaluation of the SeaWiFS chlorophyll  
489 data set. *Remote Sens. Environ.* **93**, 463–479 (2004).
- 490 37. C. Hu, L. Feng, Z. Lee, Uncertainties of SeaWiFS and MODIS remote sensing reflectance:  
491 Implications from clear water measurements. *Remote Sens. Environ.* **133**, 168–182  
492 (2013).
- 493 38. C. R. McClain, A Decade of Satellite Ocean Color Observations. *Ann. Rev. Mar. Sci.* **1**,  
494 19–42 (2009).
- 495 39. M. Stramska, D. Stramski, H. Ryszard, S. Kaczmarek, S. Joanna, Bio-optical relationships  
496 and ocean color algorithms for the north polar region of the Atlantic. *J. Geophys. Res.*  
497 **108**, 3143 (2003).
- 498 40. A. Mendonça, *et al.*, Evaluation of ocean color and sea surface temperature sensors  
499 algorithms using in situ data: a case study of temporal and spatial variability on two

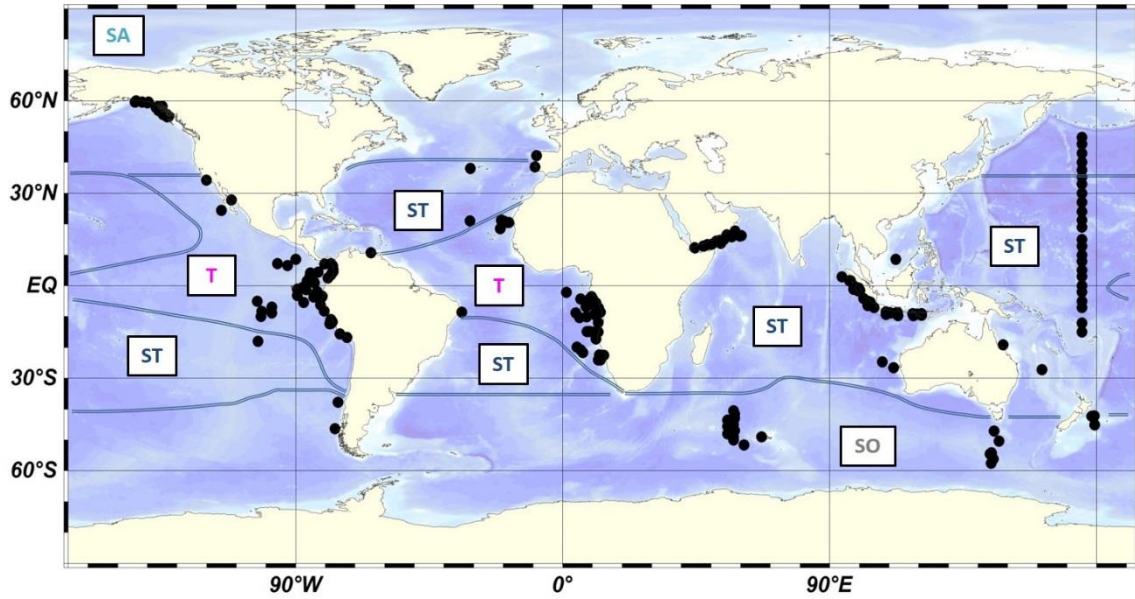
- 500 northeast Atlantic seamounts. *J. Appl. Remote Sens.* **4**, 043506 (2010).
- 501 41. J. N. Waite, F. J. Mueter, Spatial and temporal variability of chlorophyll-a concentrations in  
502 the coastal Gulf of Alaska, 1998-2011, using cloud-free reconstructions of SeaWiFS and  
503 MODIS-Aqua data. *Prog. Oceanogr.* **116**, 179–192 (2013).
- 504 42. P. J. Lam, S. C. Doney, J. K. B. Bishop, The dynamic ocean biological pump: Insights  
505 from a global compilation of particulate organic carbon, CaCO<sub>3</sub>, and opal concentration  
506 profiles from the mesopelagic. *Global Biogeochem. Cycles* **25**, 1–14 (2011).
- 507 43. C. M. Marsay, *et al.*, Attenuation of sinking particulate organic carbon flux through the  
508 mesopelagic ocean. *Proc. Natl. Acad. Sci.* **112**, 1089–1094 (2015).
- 509 44. T. Weber, J. A. Cram, S. W. Leung, T. DeVries, C. Deutsch, Deep ocean nutrients imply  
510 large latitudinal variation in particle transfer efficiency. *Proc. Natl. Acad. Sci.* **113**, 8606–  
511 8611 (2016).
- 512 45. M. Yamamoto, *et al.*, Seasonal and depth variations in molecular and isotopic alkenone  
513 composition of sinking particles from the western North Pacific. *Deep. Res. Part I*  
514 *Oceanogr. Res. Pap.* **54**, 1571–1592 (2007).
- 515 46. P. J. Müller, G. Fischer, A 4-year sediment trap record of alkenones from the filamentous  
516 upwelling region off Cape Blanc, NW Africa and a comparison with distributions in  
517 underlying sediments. *Deep. Res. Part I Oceanogr. Res. Pap.* **48**, 1877–1903 (2001).
- 518 47. E. Suess, Particulate organic carbon flux in the oceans—surface productivity and oxygen  
519 utilization. *Nature* **288**, 260–263 (1980).
- 520 48. J. H. Martin, G. A. Knauer, D. M. Karl, W. W. Broenkow, VERTEX: carbon cycling in the  
521 northast Pacific. **34**, 267–285 (1987).
- 522 49. R. S. Lampitt, A. N. Antia, Particle flux in deep seas: Regional characteristics and  
523 temporal variability. *Deep. Res. Part I Oceanogr. Res. Pap.* **44**, 1377–1403 (1997).
- 524 50. M. Lutz, R. Dunbar, K. Caldeira, Regional variability in the vertical flux of particulate  
525 organic carbon in the ocean interior. *Global Biogeochem. Cycles* **16**, 11-1-11–18 (2002).
- 526 51. R. A. Jahnke, The global ocean flux of particulate organic carbon : Areal distribution and  
527 magnitude. **10**, 71–88 (1996).
- 528 52. C. Pelejero, J. O. Grimalt, M. Sarnthein, L. Wang, J. A. Flores, Molecular biomarker record  
529 of sea surface temperature and climatic change in the South China Sea during the last  
530 140,000 years. *Mar. Geol.* **156**, 109–121 (1999).
- 531 53. M. J. Higginson, M. A. Altabet, Initial test of the silicic acid leakage hypothesis using  
532 sedimentary biomarkers. *Geophys. Res. Lett.* **31**, 4–7 (2004).
- 533 54. C. T. Bolton, K. T. Lawrence, S. J. Gibbs, P. A. Wilson, T. D. Herbert, Biotic and  
534 geochemical evidence for a global latitudinal shift in ocean biogeochemistry and export  
535 productivity during the late Pliocene. *Earth Planet. Sci. Lett.* **308**, 200–210 (2011).
- 536 55. W. K. W. Li, Composition of ultraphytoplankton in the central North Atlantic. **122**, 1–8  
537 (1995).
- 538 56. A. Z. Worden, J. K. Nolan, B. Palenik, Assessing the dynamic and ecology of marine  
539 picoplankton : the importance of eukaryotic component. *Limnol. Ocean.* **49**, 168–79.  
540 (2004).
- 541 57. E. Teira, *et al.*, Variability of chlorophyll and primary production in the Eastern North  
542 Atlantic Subtropical Gyre: Potential factors affecting phytoplankton activity. *Deep. Res.*  
543 *Part I Oceanogr. Res. Pap.* **52**, 569–588 (2005).
- 544 58. X. A. G. Morán, Annual cycle of picophytoplankton photosynthesis and growth rates in a  
545 temperate coastal ecosystem: A major contribution to carbon fluxes. *Aquat. Microb. Ecol.*  
546 **49**, 267–279 (2007).
- 547 59. Y. M. Rii, *et al.*, Diversity and productivity of photosynthetic picoeukaryotes in  
548 biogeochemically distinct regions of the South East Pacific Ocean. *Limnol. Oceanogr.* **61**,  
549 806–824 (2016).
- 550 60. V. S. Smetacek, Role of sinking in diatom life-history cycles: ecological, evolutionary and  
551 geological significance. *Mar. Biol.* **84**, 239–251 (1985).
- 552 61. C. Sancetta, T. Villareal, P. Falkowski, Massive fluxes of rhizosolenid diatoms : A common  
553 occurrence ? *Notes*, 1452–1457 (1989).

- 554 62. A. F. Michaels, M. W. Silver, Primary production, sinking fluxes and the microbial food  
555 web. *Deep. Res.* **35**, 473–490 (1988).
- 556 63. A. M. Waite, K. A. Safi, J. A. Hall, S. D. Nodder, Mass sedimentation of picoplankton  
557 embedded in organic aggregates. *Limnol. Oceanogr.* **45**, 87–97 (2000).
- 558 64. T. L. Richardson, G. A. Jackson, H. W. Ducklow, M. R. Roman, Carbon fluxes through  
559 food webs of the eastern equatorial Pacific: An inverse approach. *Deep. Res. Part I*  
560 *Oceanogr. Res. Pap.* **51**, 1245–1274 (2004).
- 561 65. T. L. Richardson, G. A. Jackson, H. W. Ducklow, M. R. Roman, Spatial and seasonal  
562 patterns of carbon cycling through planktonic food webs of the Arabian Sea determined by  
563 inverse analysis. *Deep. Res. Part II Top. Stud. Oceanogr.* **53**, 555–575 (2006).
- 564 66. S. L. C. Giering, *et al.*, High export via small particles before the onset of the North Atlantic  
565 spring bloom. *J. Geophys. Res. Ocean.* **121**, 2268–2285 (2016).
- 566 67. G. Dall’Olmo, K. A. Mork, Carbon export by small particles in the Norwegian Sea.  
567 *Geophys. Res. Lett.*, 1–6 (2014).
- 568 68. T. L. Richardson, G. A. Jackson, Small Phytoplankton and Carbon Export from the  
569 Surface Ocean. *Science (80-. )*, 838–840 (2007).
- 570 69. K. G. J. Nierop, G. J. Reichart, H. Veld, J. S. Sinninghe Damsté, The influence of oxygen  
571 exposure time on the composition of macromolecular organic matter as revealed by  
572 surface sediments on the Murray Ridge (Arabian Sea). *Geochim. Cosmochim. Acta* **206**,  
573 40–56 (2017).
- 574 70. E. Kristensen, M. Holmer, Decomposition of plant materials in marine sediment exposed  
575 to different electron acceptors (O<sub>2</sub>, NO<sub>3</sub><sup>-</sup>, and SO<sub>4</sub><sup>2-</sup>), with emphasis on substrate origin,  
576 degradation kinetics, and the role of bioturbation. *Geochim. Cosmochim. Acta* **65**, 419–  
577 433 (2001).
- 578 71. B. Dauwe, J. J. Middelburg, P. M. J. Herman, Effect of oxygen on the degradability of  
579 organic matter in subtidal and intertidal sediments of the North Sea area. *Mar. Ecol. Prog.*  
580 *Ser.* **215**, 13–22 (2001).
- 581 72. D. E. Canfield, Factors influencing organic carbon preservation in marine sediments.  
582 *Chem. Geol.* **114**, 315–329 (1994).
- 583 73. H. E. Hartnett, R. G. Keil, J. I. Hedges, A. H. Devol, Influence of oxygen exposure time on  
584 organic carbon preservation in continental margin sediments. *Nature* **391**, 572–574  
585 (1998).
- 586 74. K. A. Koho, *et al.*, Microbial bioavailability regulates organic matter preservation in marine  
587 sediments. *Biogeosciences* **10**, 1131–1141 (2013).
- 588 75. M. M. M. Kuypers, R. D. Pancost, I. A. Nijenhuis, J. S. Sinninghe Damsté, Enhanced  
589 productivity led to increased organic carbon burial in the euxinic North Atlantic basin  
590 during the late Cenomanian oceanic anoxic event. *Paleoceanography* **17**, 3-1-3–13  
591 (2002).
- 592 76. M. Rodrigo-Gámiz, S. W. Rampen, S. Schouten, J. S. Sinninghe Damsté, The impact of  
593 oxic degradation on long chain alkyl diol distributions in Arabian Sea surface sediments.  
594 *Org. Geochem.* **100**, 1–9 (2016).
- 595 77. A. W. Jacobel, *et al.*, Deep Pacific storage of respired carbon during the last ice age:  
596 Perspectives from bottom water oxygen reconstructions. *Quat. Sci. Rev.* **230** (2020).
- 597 78. W. Lu, *et al.*, I/Ca in epifaunal benthic foraminifera: A semi-quantitative proxy for bottom  
598 water oxygen in a multi-proxy compilation for glacial ocean deoxygenation. *Earth Planet.*  
599 *Sci. Lett.* **533**, 116055 (2020).
- 600 79. A. Rosell-Melé, *et al.*, Precision of the current methods to measure the alkenone proxy  
601 U37K’ and absolute alkenone abundance in sediments: Results of an interlaboratory  
602 comparison study. *Geochemistry, Geophys. Geosystems* **2** (2001).
- 603 80. J. P. Dunne, B. Hales, J. R. Toggweiler, Global calcite cycling constrained by sediment  
604 preservation controls. *Global Biogeochem. Cycles* **26**, 1–14 (2012).
- 605 81. O. Cartapanis, D. Bianchi, S. L. Jaccard, E. D. Galbraith, Global pulses of organic carbon  
606 burial in deep-sea sediments during glacial maxima. *Nat. Commun.* **7**, 10796 (2016).
- 607 82. IOCCG, “Remote Sensing of Ocean Colour in Coastal , and Other Optically-Complex ,

- 608 Waters" (2000).
- 609 83. S. B. Groom, *et al.*, Satellite ocean colour: Current status and future perspective. *Front.*  
610 *Mar. Sci.* **6** (2019).
- 611 84. C. W. Brown, J. A. Yoder, Cocolithophorid blooms in the global ocean. *J. Geophys. Res.*  
612 **99**, 7467–7482 (1994).
- 613 85. D. Blondeau-Patissier, J. F. R. Gower, A. G. Dekker, S. R. Phinn, V. E. Brando, A review  
614 of ocean color remote sensing methods and statistical techniques for the detection,  
615 mapping and analysis of phytoplankton blooms in coastal and open oceans. *Prog.*  
616 *Oceanogr.* **123**, 123–144 (2014).
- 617 86. H. M. Dierssen, R. C. Smith, Bio-optical properties and remote sensing ocean color  
618 algorithms for Antarctic Peninsula waters. **105**, 26301–26312 (2000).
- 619 87. S. Sathyendranath, G. Cota, V. Stuart, H. Maass, T. Platt, Remote sensing of  
620 phytoplankton pigments: A comparison of empirical and theoretical approaches. *Int. J.*  
621 *Remote Sens.* **22**, 249–273 (2001).
- 622 88. H. Claustre, S. Maritoner, The Many Shades of Ocean Blue. *Ocean Sci.* **302**, 1514–1515  
623 (2003).
- 624 89. G. Volpe, *et al.*, The colour of the Mediterranean Sea: Global versus regional bio-optical  
625 algorithms evaluation and implication for satellite chlorophyll estimates. *Remote Sens.*  
626 *Environ.* **107**, 625–638 (2007).
- 627 90. H. Claustre, *et al.*, Is desert dust making oligotrophic waters greener? *Geophys. Res. Lett.*  
628 **29**, 107-1-107-4 (2002).
- 629 91. D. Stramski, J. Tegowski, Effects of intermittent entrainment of air bubbles by breaking  
630 wind waves on ocean reflectance and underwater light field. *J. Geophys. Res.* **106**,  
631 31345–31360 (2001).
- 632 92. J. J. Cullen, The deep chlorophyll maximum: Comparing vertical profiles of chlorophyll a.  
633 *Can. J. Fish. Aquat. Sci.* **39**, 791–803 (1982).
- 634 93. J. Huisman, N. N. Pham Thi, D. M. Karl, B. Sommeijer, Reduced mixing generates  
635 oscillations and chaos in the oceanic deep chlorophyll maximum. *Nature* **439**, 322–325  
636 (2006).
- 637 94. J. E. O'Reilly, *et al.*, SeaWiFS Postlaunch Calibration and Validation Analyses, part 3.  
638 *NASA Tech. Memo.* **11**, 1–49 (2000).
- 639

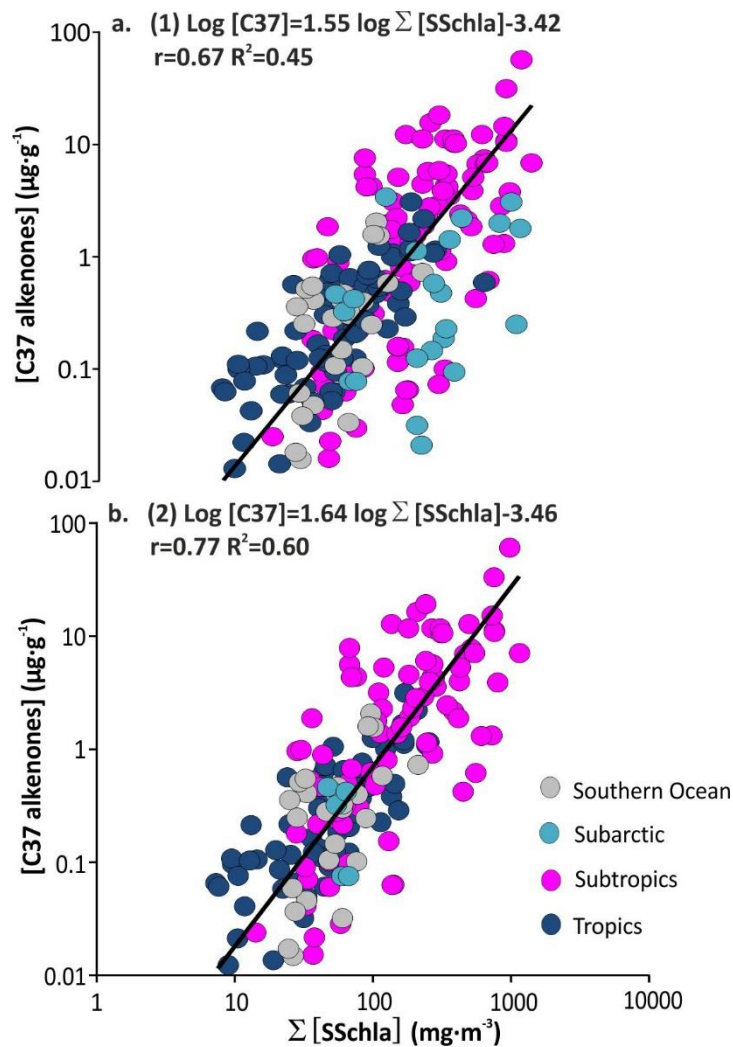
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Figures and Tables



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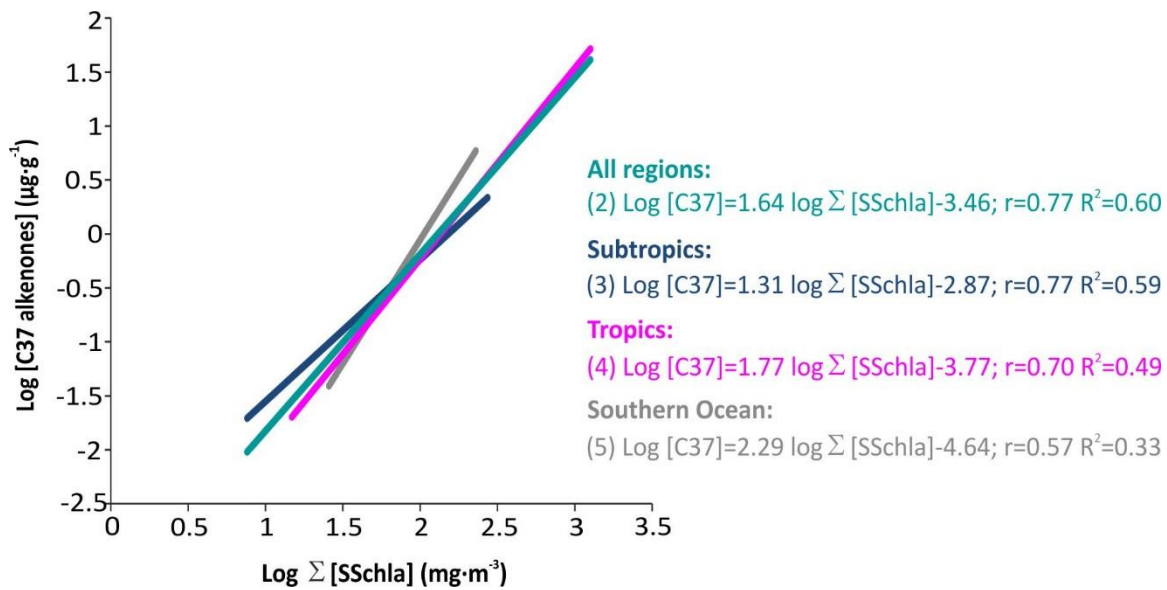
**Fig. 1.** Global core-top sediments distribution. Lines delineate distinct biogeochemical regions defined on the basis of temperature and nutrient concentration (44) Abbreviations: subarctic (SA), subtropics (ST), tropics (T) and Southern Ocean (SO).



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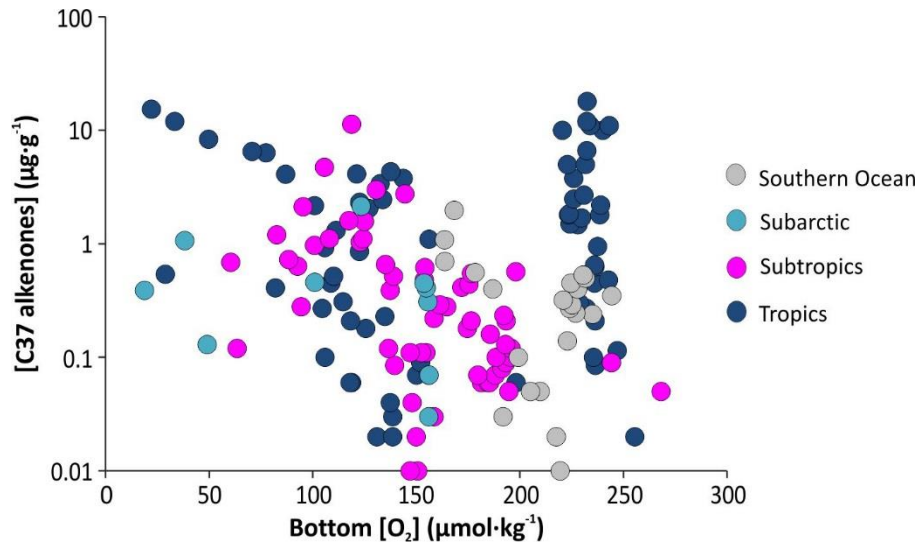
**Fig. 2.** Global correlations between sedimentary C<sub>37</sub> alkenones concentration and the sum of sea-surface chlorophyll-a (SSchla) concentration from 1997 to 2017. **a.** includes the whole compilation, and **b.** includes regions presenting RMS log errors lower than 31% between remote sensing and *in situ* SSchla. Samples are classified by biogeochemical regions as defined in Fig. 1.





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**Fig. 3.** Regional correlations between sedimentary C<sub>37</sub> alkenones concentration and the sum of sea-surface chlorophyll-a (SSchla) concentration from 1997 to 2017. Different colours for lines and text indicate the evaluated biogeochemical regions, which are defined in Fig. 1.



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**Fig. 4.** Global comparison of sedimentary C<sub>37</sub> alkenones concentration and oxygen concentration in bottom waters. Different colours for lines and text indicate the evaluated biogeochemical regions, which are defined in Fig. 1.

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675 **Table 1.** Equation coefficients and errors from correlations in Fig. 2 and 3.  $\text{Log}[C_{37}$   
676  $\text{alkenones}] = a \cdot \text{log}[\text{SSchla}] + b$ . Abbreviations: sea-surface chlorophyll-a (SSchla), coefficient of  
677 correlation ( $r$ ), coefficient of determination ( $R^2$ ), root-mean square logarithmic error (RMS log error),  
678 number of samples ( $n$ ).  
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Equation	Region	a	b	r	$R^2$	RMS log error (%)	n
1	Global	1.55	-3.42	0.67	0.45	53	226
2	Global	1.64	-3.46	0.77	0.60	38	194
3	Subtropics	1.31	-2.87	0.77	0.59	31	75
4	Tropics	1.77	-3.77	0.70	0.49	46	88
5	Southern Ocean	2.29	-4.64	0.57	0.33	35	26

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