



REVIEW

published: 16 April 2018 doi: 10.3389/fphar.2018.00267



Amyotrophic Lateral Sclerosis (ALS) and Adenosine Receptors

Ana M. Sebastião 1,2*, Nádia Rei 1,2 and Joaquim A. Ribeiro 1,2

¹ Instituto de Farmacologia e Neurociências, Faculdade de Medicina, Universidade de Lisboa, Lisbon, Portugal, ² Instituto de Medicina Molecular, Universidade de Lisboa, Lisbon, Portugal

In the present review we discuss the potential involvement of adenosinergic signaling, in particular the role of adenosine receptors, in amyotrophic lateral sclerosis (ALS). Though the literature on this topic is not abundant, the information so far available on adenosine receptors in animal models of ALS highlights the interest to continue to explore the role of these receptors in this neurodegenerative disease. Indeed, all motor neurons affected in ALS are responsive to adenosine receptor ligands but interestingly, there are alterations in pre-symptomatic or early symptomatic stages that mirror those in advanced disease stages. Information starts to emerge pointing toward a beneficial role of A_{2A} receptors (A_{2A}R), most probably at early disease states, and a detrimental role of caffeine, in clear contrast with what occurs in other neurodegenerative diseases. However, some evidence also exists on a beneficial action of $A_{2A}R$ antagonists. It may happen that there are time windows where A_{2A}R prove beneficial and others where their blockade is required. Furthermore, the same changes may not occur simultaneously at the different synapses. In line with this, it is not fully understood if ALS is a dying back disease or if it propagates in a centrifugal way. It thus seems crucial to understand how motor neuron dysfunction occurs, how adenosine receptors are involved in those dysfunctions and whether the early changes in purinergic signaling are compensatory or triggers for the disease. Getting this information is crucial before starting the design of purinergic based strategies to halt or delay disease progression.

Keywords: adenosine receptors, amyotrophic lateral sclerosis (ALS), spinal cord motor neurons, neuromuscular junction, glial cells

OPEN ACCESS

Edited by:

Kenneth A. Jacobson, National Institutes of Health (NIH), United States

Reviewed by:

Rashid Giniatullin, University of Eastern Finland, Finland Yijuang Chern, National Research Program for Biopharmaceuticals, Taiwan

*Correspondence:

Ana M. Sebastião anaseb@medicina.ulisboa.pt

Specialty section:

This article was submitted to Experimental Pharmacology and Drug Discovery, a section of the journal Frontiers in Pharmacology

> Received: 15 December 2017 Accepted: 09 March 2018 Published: 16 April 2018

Citation

Sebastião AM, Rei N and Ribeiro JA (2018) Amyotrophic Lateral Sclerosis (ALS) and Adenosine Receptors. Front. Pharmacol. 9:267. doi: 10.3389/fphar.2018.00267

INTRODUCTION

In the present review, we will address the potential role of adenosine on amyotrophic lateral sclerosis (ALS), also known as Lou Gehrig's disease. This is one of the most devastating neurodegenerative disorders and is the most common form of Motor Neuron Diseases group. During ALS progression, both the upper motor neurons (motor neurons in the cortex) and the lower motor neurons (motor neurons in the spinal cord) degenerate or die. As a consequence of deficient input from the motor neuron, there is a progressive and terminal atrophy of skeletal muscles. All muscles under voluntary control are affected, and individuals lose their strength and the ability to move their arms, legs and body. When diaphragm and the muscles in the chest wall fail, people lose the ability to breathe without ventilation support (Nichols et al., 2013).

1

The incidence of this disease is 2–3/100,000 and the prevalence 4–7/100,000 (Nalini et al., 2008). The onset of symptoms is predominantly between 55 and 65 years old and the median survival of ALS patients, after the diagnosis, is 3–5 years; however, about 10% of the patients survive for 10 or more years (Wijesekera and Leigh, 2009).

The majority of cases of ALS are sporadic (sALS) (90-95%), but some cases have a positive familial history (fALS) (5-10%) (Kiernan et al., 2011). Both forms, sALS and fALS, present similar pathological and clinical features. In the case of fALS, the inheritance is autossomal dominant, but the penetrance is variable. About 20% of fALS are caused by a missense mutation in SOD1 gene, encoding for the Cu/Zn superoxide dismutase 1 enzyme. This finding led to the first and most used rodent model for ALS, the SOD1^{G93A} mouse, which in the symptomatic phase recapitulate most features of the disease, including neuromuscular dysfunction (Naumenko et al., 2011). There is still a debate whether motor neuron degeneration initially results from failure of enzymatic machinery at the level of the cell body and proximal parts of the axon that then propagates in a centrifugal way due to impaired axonal transport (Braak et al., 2013), or results from an early dysfunction at the level of the nerve terminals, with consequent synaptic dysfunction and then progressing in a 'dying back' process (Frey et al., 2000; Dadon-Nachum et al., 2011), or both (Baker, 2014). As pointed out (Dadon-Nachum et al., 2011), if cell nerve terminal degeneration precedes with axonal and cell body degeneration, early intervention focusing on motor neurons terminals could potentially delay or prevent the progressive loss of motor neurons. A functional study focusing on the activity of single end-plates demonstrated that neuromuscular transmission impairment starts long before symptomatic onset (Rocha et al., 2013). Importantly, in pre-symptomatic SOD^{G92A} mice there are functional hallmarks of dysregulated intraterminal calcium levels (Rocha et al., 2013) in consonance with the hypothesis that nerve terminal machinery designed to buffer calcium might be impaired at very early disease stages. The pathophysiological alterations at the neuromuscular junction of SOD G92A mice are accompanied with alterations in the activity of adenosine receptors at the motor nerve terminals in these mice (Nascimento et al., 2014, 2015).

Mutations in the gene coding for a nuclear protein, TAR DNAbinding protein-43 are also frequent in fALS (Leblond et al., 2014). This protein has several functions in regulation of gene expression, which thus indicates a clear dysfunction at the level of the neuronal soma in some forms of ALS. ROS induce the mislocalization of TAR-DNA binding protein 43 (TDP-43) from the nucleus into the cytoplasm (Ayala et al., 2011), where it forms aggregates, an early hallmark of ALS (Neumann et al., 2006; Chou et al., 2018). Cytoplasmic mislocalization of TDP-43 is accompanied by further ROS production and overactivation AMP kinase (AMPK), an action suppressed by A_{2A}R activation (Liu et al., 2015a). Importantly, A2AR activation in a TDP-43Tg mouse model of ALS, improved motor function (Liu et al., 2015a). Interestingly also, activation of AMPK changed the location of a mRNA stabilizer in the motor neurons of ALS patients, in mouse motor neurons, and in a motor neuron cell line, and this mislocalization was also suppressed by activation of $A_{2A}Rs$ (Liu et al., 2015b). This again reinforces the idea of a putative neuroprotective role of $A_{2A}Rs$ in ALS since impaired RNA homeostasis is a major pathway for ALS pathogenesis (Liu et al., 2017).

Mutations in *C9ORF72* are also frequently found in fALS and accounts for 10% of the sporadic cases (DeJesus-Hernandez et al., 2011; Smith et al., 2013). Motor neurons with *C9ORF72* mutations have increased expression and activity of NMDA receptors and calcium permeable AMPA receptors (Selvaraj et al., 2018; Shi et al., 2018), known to be targets for adenosine receptor-mediated modulation (Dias et al., 2012, 2013). No information so far exists on the possibility that adenosine receptor ligands may influence motor neuronal survival in this model of ALS.

Being adenosine a ubiquitous neuromodulator, affecting synaptic transmission at pre- post and non-synaptic levels, having adenosine receptors not only neuroprotective but also excitotoxic and neuroinflammatory actions, it is somehow unexpected that only relatively few studies concentrated their attention on the influence of adenosine on ALS progression. Evidence so far available highlights early dysfunctions of neuromodulation by adenosine and the ability to influence those dysfunctions through manipulation of adenosine receptors. In this review we will critically analyze data so far available.

PATHOPHYSIOLOGY OF ALS

ALS is a complex, multifactorial and multi-systemic disease, the pathophysiological mechanisms of motor neurons degeneration remaining yet incompletely known. Some of these mechanisms include RNA dysfunction, protein misfolding and aggregation, marked neuromuscular junction abnormalities, immune system deficiency, mitochondrial dysfunction, neuroinflammation, cytoskeletal derangements, desregulation of growth factors, oxidative stress, axonal disruption and apoptosis, excitotoxity (induced by glutamate), activation of nucleases and proteases and abnormal calcium metabolism (Pasinelli and Brown, 2006; Costa et al., 2010; Chou et al., 2018). It is becoming clear that ALS is a disease that involves different cell types and the communication between them, the damage that occurs in each cell population contributing to ALS pathogenicity and phenotype, thus to the progression of the disease (Taylor et al., 2016). Indeed, non-motor neuron cells, like interneurons, astrocytes, microglia, Schwann cells, skeletal muscle cells and oligodendrocytes play a crucial role in motor neuron survival, and their dysfunction impact upon motor neuron degeneration. In practical terms, a major problem that is shared with many neurodegenerative diseases is that when patients are diagnosed, the neuronal degeneration, in the case of ALS motor neuron degeneration, has already started and progressed.

Functional and structural changes at the level of the endplate are major events in ALS. It has now been realized that the functional changes may precede not only the structural ones but also the onset of symptoms. Studies conducted in our lab (Rocha et al., 2013) to investigate transmission at the diaphragm neuromuscular junction in SOD1^{G93A} mice revealed that the

during the pre-symptomatic phase there is an enhanced release of ACh from nerve terminals, as determined by the mean quantal content of end-plate potentials. Also in these neuromuscular junctions the frequency of giant miniature endplate potentials (GMEPPs) was markedly increased, suggesting that the ability of the nerve terminal machinery to control cytoplasmatic [Ca²⁺] is dysregulated. In early symptomatic (defined as moderate changes performance in the RotaRod test) SOD1^{G93A} mice two groups of neuromuscular junctions were identified, both of them co-existing in the same innervated diaphragm (Rocha et al., 2013): one group, designated as group A (SOD1a group), presented significant reduction of the mean amplitude of EPP, a significant reduction of the mean amplitude of MEPPS, and a slight but not significant reduction of the quantal content of EPPs. All these changes indicate a predominant post-synaptic impairment. The frequency of GMEPPs was significantly reduced in SOD1a neuromuscular junctions, even as compared with wild type siblings, indicating that presynaptic calcium dysregulations persisted, though with opposite signal in pre-symptomatic and in symptomatic mice. The other group of neuromuscular junctions, designated as group B (SOD1b) (Rocha et al., 2013) displayed electrophysiological characteristics somehow in between those of age-matched wild type and presymptomatic mice, being not significantly different from any of these groups. Summarizing, data suggest (Rocha et al., 2013) an exacerbated, but already dysregulated, presynaptic activity at the mice neuromuscular junctions that do not yet display motor symptoms; once symptoms start to appear some neuromuscular junctions are already hypofunctional, some others being probably in a transition between hyper and hypofunction.

Presynaptic facilitation of neuromuscular transmission preceding motor symptoms was further confirmed in using a late onset slow progressing disease model, the SOD1G37R mice (Arbour et al., 2015). A recent study (Arbour et al., 2017) taking advantage of this late onset slow progressing disease model, systematically analyzed the changes in synaptic properties over the course of the disease progression and as a function of the motor unit type. Indeed, different motor units have different susceptibility to the disease, and it became clear the time course and the sequence of events associated with neuromuscular transmission dysfunction depend on the motor unit type (Arbour et al., 2017). The slow motor neurons are those that degenerate later and interestingly, they display an early and transient increase in the quantal content of endplate potentials that disappear at disease onset. In contrast, fast fatigue motor neurons, those that degenerate first, have reduced quantal content even before disease onset. Somehow in the between, fatigue resistant motor units only evidence neuromuscular transmission dysfunction after disease onset (Arbour et al., 2017). Interestingly only slow-type motor neurons display intrinsic hyperexcitability in pre-symptomatic mice (Leroy et al., 2014), pointing toward the possibility that early intrinsic hyperexcitability does not contribute to motor neuron degeneration, but rather represents an early compensatory process. It is worthwhile to note that as previously seen while using the SOD1^{G93A} mice (Rocha et al., 2013), in SOD1^{G37R} mice the neuromuscular transmission alterations precede the onset of

motor symptoms, as well as precede motor neuron loss, axonal degeneration, and NMJ structural changes (Arbour et al., 2017).

It is nowadays clear that motor neuron degeneration in ALS also involves glial cells, namely astrocytes, oligodendrocytes, microglia, Schwann Cells and, in the case of the neuromuscular junction, perisynaptic Schwann cells (PSCs). PSCs have neuromodulatory properties close to those of astrocytes in the central nervous system, closely interacting with nerve terminals. Thus, PSCs respond to ATP and ACh released during nerve activity, which act through P2Y and muscarinic ACh receptors (mAChRs) coupled to transient increases in intracellular calcium concentration, a process that has to be kept under proper control to keep PSCs competent for their functions to modulate and to repair neuromuscular transmission (Son et al., 1996; Georgiou et al., 1999; Feng and Ko, 2008a,b). Interestingly, PSC of SOD1 mice displayed exacerbated mAChR responses and exacerbated calcium signaling, which precede functional and morphological changes at the neuromuscular junction, suggesting that the impairments of PSC functions may contribute to NMJ dysfunction and ALS pathogenesis (Arbour et al., 2015).

At the level of spinal cord and focusing in neuroinflammatory and cellular intercommunication hallmarks, it has recently been also shown a clear difference between pre-symptomatic and symptomatic stages (Cunha et al., 2017). Thus, before onset of motor symptoms, alterations in both astrocytes and microglia have been detected in the spinal cord of SOD1^{G93A} mice, which comprise decreased expression of astrocytic, microglia, inflammatory and cell communication markers together with upregulation of a glutamate transporter-1 marker. In contrast, in the symptomatic stage, increased markers of inflammation became evident (Cunha et al., 2017). Microglia activation, suggestive of a switch from M1 to M2-like microglia subpopulations, have been detected in control cells exposed to exosomes derived from motor neuron-like cells transfected with mutant SOD1^{G93A} (Cunha et al., 2017), thus suggesting that disease progression is promoted by cell to cell communication events. There is also a growing body of evidence showing that astrocytes expressing ALS-associated proteins impair motor neuron survival and potentiate ALS progression (Di Giorgio et al., 2007; Nagai et al., 2007; Marchetto et al., 2008; Qian et al., 2017).

Recent attention has been paid to two other ALS-related dysfunctions, TDP-43 aggregation and *C9ORF72* repeated expansion. Mutations in the gene that codes for TDP-43, *TARDBP*, may favor TDP-43 aggregation but phosphorylated TDP-43 containing aggregates also occur in sALS (Neumann et al., 2006; Chou et al., 2018). Indeed, TDP-43 aggregates are found in 97% of ALS cases of diverse etiology and constitute a major component of protein inclusions in this disease (Arai et al., 2006; Maekawa et al., 2009). TDP-43 is a nuclear protein that interacts with RNA molecules and is involved in a wide variety of relevant cellular pathways related to RNA and protein homeostasis. TDP-43 C-terminal missense mutations have been identified in ALS patients, which promote mislocalization of TDP-43 from the nucleus to the cytoplasm and lead to neurotoxicity (Gitcho et al., 2008; Nonaka et al., 2009;

Guo et al., 2011). Oxidative stress and ROS formation also favor mislocalization of TDP-43, from the nucleus to the cytoplasm (Ayala et al., 2011), being thus not surprising that SOD mutations also lead to TDP-43 pathology in mice and humans (Shan et al., 2009; Pokrishevsky et al., 2012). ROS upregulates AMP kinase (AMPK) and abnormal AMPK activity has been shown to induce TDP-43 mislocalization in a motor neuron cell line and in the spinal cord of ALS patients (Liu et al., 2015a). Interestingly, mislocalized TDP-43 in the cytoplasm triggers a positive feedback loop, leading to further ROS production and AMPK activation (Liu et al., 2015a).

The present understanding of the mechanisms that underlie motor neuron degeneration associated to expansion of a G4C2 intronic hexanucleotide of the C9ORF72 gene, is scarce. This mutation is a common cause of fALS but also accounts for near 10% of sALS cases (DeJesus-Hernandez et al., 2011; Smith et al., 2013). A recent study has shown that the C9ORF72 mutation is associated with an increase in GluA1 AMPA receptor subunit expression, functional expression of Ca²⁺permeable AMPA receptors and motor neuron vulnerability to excitotoxicity (Selvaraj et al., 2018). Interestingly, early intrinsic hyperexcitability has been detected in mixed cultures of neurons and glia derived from C9ORF72 repeat expansion patient iPSCs (Wainger et al., 2014), but not in neuronal cultures with negligible number of glial cells (Selvaraj et al., 2018). Also, conditional medium from astrocytes expressing mutant SOD1 can induce motor neuron hyperexcitability (Fritz et al., 2013). Altogether these studies reinforce the idea that diseased astrocytes trigger excitotoxicity.

Motor neurons, astrocytes, microglia and perisynaptic Schwann cells are known to release and respond to purines, namely ATP and adenosine. Adenosine, by activating membrane receptors (A₁, A_{2A}, A_{2B}, A₃), is an endogenous and ubiquitous modulator of synaptic signaling. Released ATP is a source for extracellular adenosine, but acts on its own purinergic P2 (P2Y and P2X) receptors. So the challenging question is: How purinergic receptors affect the motor neurons and how this impacts in ALS? In this review we will only refer to adenosine receptors since the putative involvement of ATP receptors in the pathophysiology of ALS has been matter of a recent review (Volonté et al., 2016).

ADENOSINE AND ADENOSINE RECEPTORS

Adenosine is a ubiquitous endogenous neuroprotective agent, which has a central role as a neuromodulator of synaptic transmission at the central and peripheral nervous systems, protecting organs and tissues at both physiological and pathophysiological conditions. Adenosine exerts its biological effect via a class of purinergic G protein coupled receptors, which includes A₁R, A_{2A}R, A_{2B}R and A₃R, all belonging to family of receptors named by Burnstock (1976) as P₁ purinoceptors (adenosine-sensitive) as opposed to the ATP-sensitive P2 purinoceptors. The A₁R couple to G_{i/o} proteins, inhibiting the production of cyclic AMP. The A_{2A} and A₂B

receptors couple to G_s , stimulating the production of cyclic AMP, whereas the A_3R may couple to $G_{i/o}$ or G_q proteins. Adenosine acts like an extracellular signaling molecule, modulating the action of several neurotransmitters (Ribeiro et al., 2003).

Adenosine A₁R are widely distributed in the brain having a widely recognized inhibitory action in synaptic transmission associated to a neuroprotective role, while A2AR have more restrict localizations, being predominant at the basal ganglia. However, both receptors are present in the cortex, the A₁R being the predominant in this brain area (Dunwiddie and Masino, 2001; Sebastião and Ribeiro, 2009a). A2AR may exacerbate excitotoxicity in several brain areas (Ribeiro et al., 2016b) as well as in motor neurons (Mojsilovic-Petrovic et al., 2006). However, A_{2A}R are also able to gate the action of neuroprotective molecules as the neurotrophic factors, a mechanism known to occur in the brain (Diógenes et al., 2004) as well as in motor neurons (Wiese et al., 2007). Trophic actions of A_{2A}R in cortical neurons, some of them being independent of the interplay with neurotrophic factors, have been recently identified (Ribeiro et al., 2016a). Expression of A_{2A}R in spinal motor neurons is intense, being higher than that of A_1R (Kaelin-Lang et al., 1999; Ng et al., 2015).

 A_1R in the spinal cord inhibit excitatory inputs (Deuchars et al., 2001) whereas $A_{2A}R$ facilitate inhibitory inputs (Brooke et al., 2004) to sympathetic preganglonic neurons, suggesting that at least in what concerns autonomic nervous system control, the opposing actions of A_1R and $A_{2A}R$ at the synaptic level may both contribute to a common outcome – a decrease in preganglionic neuron activity. The antinociceptic actions of adenosine A_1R are widely known, which involve A_1R at the dorsal horn of the spinal cord. A_3R receptors also mediate antinociception through spinal and supraspinal mechanisms (Little et al., 2015) that also involve anti-inflammation (Janes et al., 2015).

Concerning motor pattern generation, it has been for a long time described that ATP and adenosine have opposite actions in motor pattern generation, the adenosine actions being inhibitory, lowering the excitability of motor circuits (Dale and Gilday, 1996). Recent studies have shown that adenosine, via A₁R, has a general inhibitory action on ventral horn interneurons while potentially maintaining motor neuron excitability (Witts et al., 2015). Interestingly, glial cells are a predominant source of adenosine at to control motor neuron networks at the spinal cord (Witts et al., 2012; Carlsen and Perrier, 2014; Acton and Miles, 2015).

The motor nerve terminals at the neuromuscular junction have served as a model to the pioneer studies on the inhibitory action of adenosine and ATP on neurotransmitter release (Ginsborg and Hirst, 1972; Ribeiro and Walker, 1973, 1975), being now well known that endogenous adenosine inhibits neuromuscular transmission (Sebastião and Ribeiro, 1985), that ATP is a source of extracellular adenosine at the endplate (Ribeiro and Sebastião, 1987), and that adenosine can also be independently released from stimulated motor nerve endings (Cunha and Sebastião, 1993). Motor nerve terminals at the mammalian skeletal neuromuscular junction possess both inhibitory A_1R (Sebastião et al., 1990) and excitatory A_2AR (Correia-de-Sá et al., 1991), the A_2AR gaining particular relevance at high frequencies of nerve stimulation

(Correia-de-Sá et al., 1996). A_{2B} and A_{3} receptors were also more recently detected at the mammalian neuromuscular junction (Garcia et al., 2014).

Adenosine Receptors in Health and Disease

At physiological conditions, the adenosine exerts its neuromodulation action by the activation of A₁ and A_{2A}R high affinity receptors (Sebastião and Ribeiro, 2009a), which mediate dual actions of adenosine both in health and disease (Fredholm, 2014). At pre-synaptic level, adenosine can inhibit or facilitate transmitter release through A₁R and A_{2A}R respectively; at post-synaptic level, it can modulate the actions of several neurotransmitters and it can hyperpolarize or depolarize neurons. All these receptors can be found in neurons and glia cells. While the A₁R is frequently associated to neuroprotective actions, which extend to conditions as hyperexcitability, seizures and ischemia/hypoxia, protecting neurons in response to excitotoxic injury (Boison, 2016; Cunha, 2016; Ribeiro et al., 2016b), the A_{2A}R is often associated to enhancement of excitotoxicity, since these receptors facilitate glutamate release and inhibit glutamate uptake (Popoli et al., 2003; Matos et al., 2012). However, through their ability to gate the actions of neurotrophic factors, A_{2A}R may confer a protective role under specific pathologic conditions (Sebastião and Ribeiro, 2009b; Rodrigues et al., 2014). A1R, A2AR and A2BR can also influence oxygen deliver into the brain due to their influence in neurovascular coupling (Pelligrino et al., 2010).

Concerning inflammation, A2AR have a duality of actions, depending on the cell type and inflammatory conditions (Haskó et al., 2008). In immune cells the global trend for A_{2A}Rmediated actions is to interrupt the proinflammatory cascade and hence limiting tissue inflammation (Sitkovsky et al., 2004; Haskó et al., 2008), contrasting with A₂B receptors that may promote overproduction of proinflammatory cytokines (Haskó et al., 2008). In the central nervous system, however, A_{2A}R mediate anti-inflammatory effects on T cells, and thus protection at early stages of inflammatory diseases; during later stages of disease, however, may contribute to sustained tissue damage within the inflamed central nervous system (Ingwersen et al., 2016). In microglia, A_{2A}R activation leads to their activation, and in such way contributing to foster neuroinflamatory cascades, thus to neurodegeneration (Dai and Zhou, 2011; Rebola et al., 2011; Santiago et al., 2014). Similarly, A_{2B}R and A₃R may mediate either anti-inflammatory or pro-inflammatory actions (Borea et al., 2017).

Adenosine Receptors in ALS

Adenosine Receptors at Spinal Cord: Implications for ALS

In accordance with the expected facilitatory influence of $A_{2A}R$ upon excitotoxicity, $A_{2A}R$ antagonists protect against excitotoxicity-induced motor neuron death (Mojsilovic-Petrovic et al., 2006). However, motor neuron survival after mechanical lesioning has also been shown to be increased by an $A_{2A}R$ activation, through a mechanism that involves interplay with

neurotrophines (Wiese et al., 2007). Survival of motor neurons in culture is also facilitated by A2AR activation, this mechanism requiring the activity of the canonical A_{2A}R signaling pathway, cyclic AMP/protein kinase activity, as well as the activity of neurotrophin receptors, again highlighting an interplay between A_{2A}R and neurotrophins to promote motor neuron survival (Komaki et al., 2012). The apparent discrepancies between the neuroprotective actions of A_{2A}R antagonists and A2AR agonists probably result from the diversity of actions that the A2AR have, namely modulation of neuronal activity, neuronal survival, excitotoxicity, neuroinflammation. While facilitating the actions of neurotrophic factors A_{2A}R may promote survival and regeneration, but while enhancing excitotoxicity phenomena A2AR will favor neuronal death. In addition, if neuroinflammation interferes with in the regeneration/degeneration balance A2AR may again have a dual role, depending on the degree and characteristics of the inflammatory process, that it to say, if activation of the inflammatory cascade is beneficial or detrimental. Unraveling all these aspects by taking into consideration the nature of the insult and the time window for treatment is of uttermost importance to understand the role of adenosine receptors in neurodegeneration and in particular in ALS. Our poor understanding of the pathophysiology of the disease itself makes it more difficult to understand the role of adenosine receptors in this disease.

Adenosine levels are significantly elevated in the cerebrospinal fluid of ALS patients (Yoshida et al., 1999), a finding that raised the interest on the understanding the role of adenosine and adenosine receptors in ALS. The expression of A2A receptors is also enhanced in lymphocytes of ALS patients (Vincenzi et al., 2013) as well as in post-mortem spinal cord of ALS patients (Ng et al., 2015). A2AR expression in the spinal cord of early symptomatic SOD1^{G93A} mice was also markedly increased (Ng et al., 2015), but in end-stage SOD1^{G93A} mice a marked decrease was reported (Potenza et al., 2013). Mice data thus may suggest that there is an early enhancement of A2AR expression in the spinal cord, followed by a decrease in latter disease states. A comparison of data obtained in the same laboratory conditions and same animal housing conditions is, however, needed to firmly conclude on biphasic changes in A_{2A}R expression during disease progression. Post-mortem human data (Ng et al., 2015), by definition related to end-stages of the disease, seems to contradict the conclusion of a late-phase decrease in A2AR expression in the spinal cord. Whether the discrepancy arrives from methodological issues (post-mortem time of analysis, tissue collection procedures), species differences (mice vs. human) or from inappropriate disease model, is unknown.

Caffeine is a non-selective adenosine receptors antagonist (Pelligrino et al., 2010; Ribeiro and Sebastião, 2010; Fredholm et al., 2017) and is one of the most consumable psychoactive substances of the western diet. There is thus high interest in understanding the actions of caffeine in neurodegenerative diseases. Indeed, caffeine consumption in humans has been negatively associated with the incidence of some neurodegenerative diseases, as Alzheimer's Disease and Parkinson's Disease (de Mendonça and Cunha, 2010; Flaten et al., 2014; Laurent et al., 2014; Ascherio and Schwarzschild, 2016).

Accordingly, chronic caffeine consumption has been shown to protect several hallmarks of neurodegeneration in animal models of disease, these actions being frequently mimicked by selective A_{2A}R antagonists or A_{2A}R deletion (Canas et al., 2009; Kachroo and Schwarzschild, 2012; Laurent et al., 2016; Ferreira et al., 2017). Concerning ALS, however, conflicting data had recently emerged in the literature, and there are several points that are far from being understood. In clear contrast to what could be expected from previous evidence in other neurodegenerative diseases, chronic administration of caffeine significantly shortens the survival of SOD1^{G93A} mice (Potenza et al., 2013). Also somehow unexpected, chronic administration of caffeine caused a marked decrease in A2AR levels in the spinal cord (Potenza et al., 2013) and in this aspect seems to mimic the disease itself since SOD1^{G93A} mice also displayed a strong reduction in A2AR in the spinal cord. Both conditions (caffeine treatment and SOD^{G93A} mutation) were not additive since there were no further reduction in spinal cord A2AR levels in SOD1^{G93A} mice treated with caffeine (Potenza et al., 2013). Caffeine can inhibit all adenosine receptors, and thus one cannot preclude that those actions of caffeine were due to antagonism of the usually neuroprotective A₁R rather than antagonism of the A2AR. Indeed, A1 and A2AR are known to strongly interact with each other (Ferre et al., 2008), including in motor neurons (Pousinha et al., 2010, 2012), and the loss of inhibitory control of A₁R upon A_{2A}R may cause marked unbalance of the fine-tuning of neuronal function exerted by adenosine receptors. A selective A_{2A}R antagonist, KW6002, administered daily, was recently reported to increase of motor neuron survival, delay the onset of disease and increase in the lifespan that SOD1G93A mice (Ng et al., 2015), which is in accordance with the usual finding that A2AR may exacerbate excitotoxicity.

Concerning the influence of A2AR agonists in ALS progression, it has been shown that treatment of SOD1^{G93A} mice with CGS21680 at a very early symptomatic stage of the disease, delayed disease onset of motor symptoms and enhanced motor neuron survival (Yanpallewar et al., 2012). Interestingly, the effect of the A2AR agonist was similar to that resulting from genetic deletion of the truncated form of the TrkB receptor, TrkB.T1, which is overexpressed in the spinal cord of the SOD1^{G93A} mice. The TrkB.T1 act as negative regulator of fulllength TrkB receptor signaling, impairing the ability of its ligand, the neurotrophin Brain Derived Neurotrophic factor (BDNF), to promote neuronal survival, neuronal maturation and neuronal plasticity. Most of the actions of BDNF at synapses are triggered by A_{2A}R activation (Diógenes et al., 2004; Sebastião et al., 2011; Ribeiro et al., 2016b). It is thus likely that the protective influence of A2AR receptor agonist in very early stages of ALS progression in SOD^{G93A} mice results from the ability of A_{2A}R to facilitate full length TrkB receptor activation, thus compensating the negative influence the TrkB.T1 receptor overexpression. That A_{2A}R and A_{2A}R antagonists may have strict time windows to exert its beneficial influences in neurodegeneration has been already proposed (Sebastião and Ribeiro, 2009b) but data from experiments specifically addressing this point have not yet appeared.

Also particularly relevant in the context of understanding the role of $A_{2A}R$ in ALS progression was the finding that $A_{2A}R$ activation, through PKA signaling, suppresses aberrant AMPK activity as well as suppresses mislocalization of TDP-43, in the TDP-43 mouse model of ALS (Liu et al., 2015a). Taking into consideration that mislocalization of TDP-43 from the nucleus to the cytoplasm can be induced not only by mutations in the gene that codes for TDP-43 but also by ROS production, there is some overlap in the disease triggering mechanisms in the SOD mouse models and in the TDP-43 model.

Summarizing, in two models of ALS (SOD1^{G93A} and TDP-43), two different A_{2A}R agonists proved beneficial to improve motor neuron survival (Yanpallewar et al., 2012; Liu et al., 2015a). One study clearly showed detrimental effects of the adenosine receptor antagonist, caffeine in the survival of SOD1^{G93A} mice (Potenza et al., 2013). Thus, as whole and in the specific context of ALS, A_{2A}R activation seems beneficial. Importantly, A_{2A}R, through cAMP/PKA signaling and subsequent inhibition of AMPK may contribute to cut the positive feedback mechanism where AMPK activation by ROS leads to further mislocalization of TDP-43, which leads to further ROS production, thus to further AMPK activation. In this way, A2AR activation may hit a key pathophysiologic mechanism of ALS, thus reducing or even halting disease progression. Unfortunately, in the study by Liu et al. (2015a), the influence of A_{2A}R activation in the time of appearance of first symptoms or in the life-span of ALS mice was not assessed. Results from such studies would indeed complement the evidence already obtained on enhancement of motor neuron survival by A2AR activation in two models of ALS (Yanpallewar et al., 2012; Liu et al., 2015a). Also, and taking into consideration that the agonist used by Liu et al. (2015a) already entered clinical trials for other neurodegenerative diseases, a clinical trial specifically designed for a fast devastating disease as ALS, would be of high relevance.

 $A_{2A}R$ and D_2 dopamine receptors (D_2R) co-exist in motor neurons of the spinal cord of normal subjects and ALS patients (Chern et al., 2017). Activation of D_2R reverse the $A_{2A}R$ -mediated prevention of ROS-induced AMPK activation and TDP-43 mislocalization in cells co-transfected with D_2R and $A_{2A}R$ (Chern et al., 2017). A_{2A}/D_2 negative interactions are well known in the basal ganglia (Fuxe et al., 2007). The evidence that they also exist at the level of the spinal cord and in a context of ALS pathophysiology (Chern et al., 2017), point toward the need of care in the use of D_2R agonists in ALS patients, as well as may suggest a beneficial role of D_2R antagonists in this disease.

From the studies mentioned above, it appears that $A_{2A}R$ agonists may prove of therapeutic value in ALS. Yet, there are several discrepancies in the literature, as the report that a selective $A_{2A}R$ antagonist may delay disease progression. To solve some discrepancies, it will be important to compare in the same colony and under the same experimental conditions, and always against age-matched control littermates, time-dependent and disease state-dependent changes in the activity and expression of adenosine receptors. The comparisons among different mouse models are also of uttermost importance to allow starting to understand the different roles of $A_{2A}R$ as a function of the etiology of the disease itself. It will be also necessary to follow

those changes in mice treated with either selective receptor ligands as well as with caffeine. Since ALS progression may not be similar at different synaptic levels, a better understanding of the time-dependent changes in $A_{2A}R$ signaling at upper and lower motor neuron levels would also allow a better understanding in the role of $A_{2A}R$ in ALS. Information of the role of $A_{1}R$ in this neurodegenerative disease is scarce, which is somehow surprising on the light of the known neuroprotection exerted by these receptors. Their ability to inhibit synaptic transmission, may, however, exert a negative influence in a degenerative disease that markedly compromises information flow in the motor circuit.

Adenosine Receptors at the Neuromuscular Junction: Implications for ALS

The neuromuscular junction early served as a model to understand synaptic transmission mechanisms (Katz and Miledi, 1965), as model to understand neuromodulation by adenosine (Ginsborg and Hirst, 1972), as a model to understand the relevance of endogenous adenosine to control synaptic transmission (Ribeiro and Sebastião, 1987), and as a model to start identifying A_{2A}R at synapses outside the basal ganglia (Correia-de-Sá et al., 1991; Correia-de-Sá and Ribeiro, 1994). Reasons for this are the simplicity of the model, allowing to record from a single synapse receiving input from only one nerve terminal, with a clearly identified neurotransmitter - acetylcholine. In spite of this simplicity, there are several neuromodulators released by either the pre-synaptic, the postsynaptic, the muscle itself and the perisynaptic Schwann cells, which finely tune synaptic transmission and synaptic maintenance. As widely known, ATP and adenosine are among such neuromodulators. Neurotrophic factors are also present at the neuromuscular junction and A2AR are also indispensable promoters of the facilitatory action BDNF, upon neuromuscular transmission (Pousinha et al., 2006), as it was firstly observed hippocampal synapses (Diógenes et al., 2004).

Having studied the action of adenosine for more than 40 years, using the neuromuscular junction as a model, and considering that this simple synaptic model could be as a sort of key (Arbour et al., 2017) to investigate novel approaches for disease therapy, and also that this particular synapse is compromised in ALS, we hypothesized (Nascimento et al., 2014, 2015) that ALS progression could be accompanied by changes in the functioning of adenosine A₁R and A_{2A}R, the two high affinity adenosine receptors known to be expressed at motor nerve endings (Correia-de-Sá and Ribeiro, 1994). We used a functional approach, recording endplate potentials (EPPs) and miniature endplate potentials (MEPPs), and quantifying quantal content of EPPs, from Mg²⁺-paralyzed hemidiaphragm preparations of SOD1^{G93A} mice at pre-symptomatic (absence of motor symptoms assessed by the Rota-Rod) and early symptomatic (mild but significant dysfunction at the Rota-Rod) phases of the disease. We found that at the pre-symptomatic stage the A_{2A}R-mediated presynaptic facilitatory action on neuromuscular transmission is exacerbated, whereas in the early symptomatic phase, this excitatory action disappears, indicating that indeed that A2AR function changes upon ALS progression. We then further investigated the role of A₁R in ALS using the same

approach and the same disease model (Nascimento et al., 2015) and found that the A₁R/A_{2A}R functional cross-talk is lost in the pre-symptomatic phase, so that the ability of A₁R to brake the action of A2AR is lost, which might explain the exacerbation of the A_{2A}R signaling at this disease state. In addition, there was an increase of A₁R tonic activation in the symptomatic phase, suggesting that the changes adenosine modulation mediated by A₁R may be contributing to disease progression and aggravating symptoms in late disease stage. Whether the lack of A₁Rmediated inhibition of A2AR function, with the corresponding exacerbation of A_{2A}R-mediated function, corresponds to an early compensatory process that may facilitate neuromuscular transmission and confer some neuroprotection through an enhancement of the action of neurotrophic factors, remains unknown. Also unknown is if these early excitatory changes, accompanied by dysruptions of the calcium buffering at the nerve terminal (Rocha et al., 2013) and at perisynaptic Schwann Cells (Arbour et al., 2015), are a fast-track to neurodegeneration.

Summarizing, data (Nascimento et al., 2014, 2015) suggest that in ALS there is an unbalanced adenosine receptors modulation at the neuromuscular junction (**Figure 1**) that may start before first signs of motor impairment. We anticipate early compensatory alterations followed by a disruption of neuromodulatory control that may then act as an aggravating factor to exacerbate excitotoxicity, fasting neuronal death. A clear identification of the nature of these adenosine receptors changes, at different diseases states, would allow to identify new therapeutic targets based on the cause of neurodegeneration and thus to halt it.

Interestingly, adenosine A_1R at motor nerve endings, besides inhibiting the release probability, also synchronize release by removing quantal events with long latencies, an action that is sensitive to the redox potential, being abolished by oxidants (Tsentsevitsky et al., 2013). How this impacts in oxidative-stress related motor neuron diseases, as ALS, is yet unknown.

CONCLUSION AND PERSPECTIVES

A major finding in what concerns adenosinergic signaling in ALS is the existence of very early synaptic alterations, which precede motor symptoms and which are evident at the level of the neuromuscular junction as well as, most probably, at central synapses. Whether those changes are triggers for the disease or represent early compensatory modifications is yet unknown. The evidence of a positive influence of A_{2A}R agonists and negative influence of caffeine, may suggest that an early enhancement of A_{2A}R functioning may represent an endogenous attempt to fight the disease. The positive influence of a selective A_{2A}R antagonist may, however, contradict this possibility. A better clarification of the nature of the purinergic signaling dysfunctions, their implications and their time-windows, is for sure relevant not only for a better understanding of the pathophysiology of the disease and the implications of caffeine intake, but also to guide the design of putative therapeutic strategies to halt disease progression. Specifically, it is important to know if the early exacerbation of A2AR signaling detected at the motor nerve endings also occurs at central synapses, in particular, at the spinal

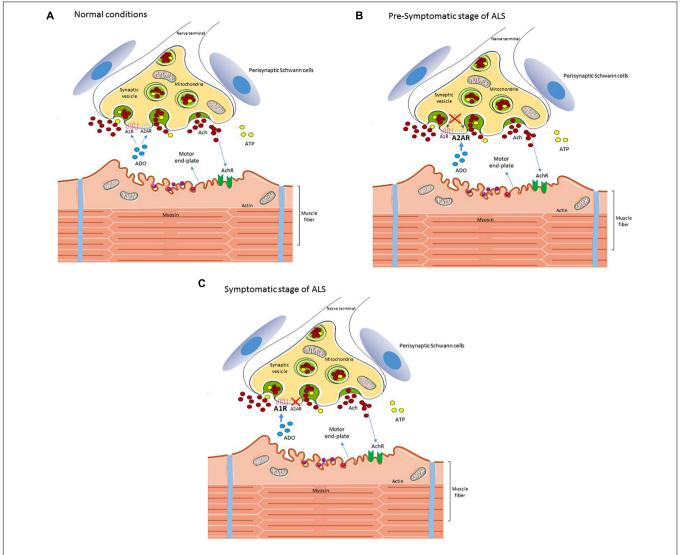


FIGURE 1 Schematic diagram of the reported changes in adenosine signaling at the neuromuscular junction in an ALS mice model. In wild type (A) neuromuscular junctions, adenosine activates both inhibitory A_1R and excitatory A_2AR , modulating acetylcholine release from motor nerve terminals. In ALS mice at the pre-symptomatic stage (B) there is a loss of A_1R/A_2AR functional cross-talk at the neuromuscular junction and the excitatory action of A_2AR is exacerbated. At the symptomatic stage (C) there is an increase of A_1R tonic activation, which may be contributing to neurotransmission failure on late disease stages, whereas the excitatory action of A_2AR disappears. For details see Nascimento et al. (2014, 2015). Perisynaptic Schwann cells contribute to the pool of ATP at the synaptic cleft but ATP signaling in these cells seems unchanged in ALS, in contrast with mACh signaling (Arbour et al., 2015). Interestingly, the exacerbated A_2AR signaling at the pre-symptomatic stage correlates with an enhanced release of ACh from nerve terminals and with signs of dysregulated calcium buffering at motor nerve endings (Rocha et al., 2013; Arbour et al., 2017).

cord and motor cortex, and how selective blockade of these receptors at different disease states affects disease progression. Keeping in mind the intriguing data reported while testing the influence of caffeine in animal models, it is also of uttermost importance to understand the molecular basis of this action of caffeine, whether it has an impact in the signaling of other neuroprotective molecules, or in the signaling mediated by other adenosine receptors. This knowledge is not only important for the design of future drugs but also to guide research in patients since caffeine is present in many widely consumed beverages and is usually regarded as protective against other neurodegenerative diseases. Lastly, the putative influence of $A_{2A}R$ and $A_{3}R$ in

the inflammatory cascade associated to ALS has to be better understood. Indeed, recent studies have demonstrated the presence of inflammation propagating substrates in the central nervous system of patients afflicted with ALS (Khalid et al., 2017). $A_{2A}R$ receptors mediate anti-inflammatory actions and are up-regulated in lymphocytes from ALS patients(Vincenzi et al., 2013). The $A_{3}R$ is a promising therapeutic target for inflammatory diseases (Jacobson et al., 2017) but these receptors have been mostly disregarded in what concerns ALS.

In conclusion, though one starts to envisage changes in the adenosinergic system at different stages of ALS progression, much more needs to be known before understanding the causal relationship of those changes and which of them can be targeted to develop novel therapeutics toward this devastating disease.

AUTHOR CONTRIBUTIONS

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

REFERENCES

- Acton, D., and Miles, G. B. (2015). Stimulation of glia reveals modulation of mammalian spinal motor networks by adenosine. PLoS One 10:e0134488. doi: 10.1371/journal.pone.0134488
- Arai, T., Hasegawa, M., Akiyama, H., Ikeda, K., Nonaka, T., Mori, H., et al. (2006). TDP-43 is a component of ubiquitin-positive tau-negative inclusions in frontotemporal lobar degeneration and amyotrophic lateral sclerosis. *Biochem. Biophys. Res. Commun.* 351, 602–611. doi: 10.1016/j.bbrc.2006.10.093
- Arbour, D., Tremblay, E., Martineau, E., Julien, J. P., and Robitaille, R. (2015). Early and persistent abnormal decoding by glial cells at the neuromuscular junction in an ALS model. *J. Neurosci.* 35, 688–706. doi: 10.1523/JNEUROSCI.1379-14. 2015
- Arbour, D., Vande Velde, C., and Robitaille, R. (2017). New perspectives on amyotrophic lateral sclerosis: the role of glial cells at the neuromuscular junction. *J. Physiol.* 595, 647–661. doi: 10.1113/JP270213
- Ascherio, A., and Schwarzschild, M. A. (2016). The epidemiology of Parkinson's disease: risk factors and prevention. *Lancet Neurol.* 15, 1257–1272. doi: 10.1016/ S1474-4422(16)30230-7
- Ayala, V., Granado-Serrano, A. B., Cacabelos, D., Naudí, A., Ilieva, E. V., Boada, J., et al. (2011). Cell stress induces TDP-43 pathological changes associated with ERK1/2 dysfunction: implications in ALS. Acta Neuropathol. 122, 259–270. doi: 10.1007/s00401-011-0850-y
- Baker, M. R. (2014). ALS—dying forward, backward or outward? *Nat. Rev. Neurol.* 10:660. doi: 10.1038/nrneurol.2013.221-c1
- Boison, D. (2016). The biochemistry and epigenetics of epilepsy: focus on adenosine and glycine. Front. Mol. Neurosci. 9:26. doi: 10.3389/fnmol.2016. 00026
- Borea, P. A., Gessi, S., Merighi, S., Vincenzi, F., and Varani, K. (2017). Pathological overproduction: the bad side of adenosine. *Br. J. Pharmacol.* 174, 1945–1960. doi: 10.1111/bph.13763
- Braak, H., Brettschneider, J., Ludolph, A. C., Lee, V. M., Trojanowski, J. Q., and Del Tredici, K. (2013). Amyotrophic lateral sclerosis-a model of corticofugal axonal spread. Nat. Rev. Neurol. 9, 708-714. doi: 10.1038/nrneurol.2013.221
- Brooke, R. E., Deuchars, J., and Deuchars, S. A. (2004). Input-specific modulation of neurotransmitter release in the lateral horn of the spinal cord via adenosine receptors. J. Neurosci. 24, 127–137. doi: 10.1523/JNEUROSCI.4591-03. 2004
- Burnstock, G. (1976). Do some nerve cells release more than one transmitter? Neuroscience 1, 239–248.
- Canas, P. M., Porciúncula, L. O., Cunha, G. M., Silva, C. G., Machado, N. J., Oliveira, J. M., et al. (2009). Adenosine A_{2A} receptor blockade prevents synaptotoxicity and memory dysfunction caused by beta-amyloid peptides via p38 mitogen-activated protein kinase pathway. *J. Neurosci.* 29, 14741–14751. doi: 10.1523/JNEUROSCI.3728-09.2009
- Carlsen, E. M., and Perrier, J. F. (2014). Purines released from astrocytes inhibit excitatory synaptic transmission in the ventral horn of the spinal cord. Front. Neural Circuits 8:60. doi: 10.3389/fncir.2014.00060
- Chern, Y., Lai, C., Liu, Y., and Lai, H. (2017). Activation of the D2 dopamine receptor hampers the protective effect of the A2A adenosine receptor on TDP-43 mislocalization. FASEB J. 31:LB140.
- Chou, C. C., Zhang, Y., Umoh, M. E., Vaughan, S. W., Lorenzini, I., Liu, F., et al. (2018). TDP-43 pathology disrupts nuclear pore complexes and nucleocytoplasmic transport in ALS/FTD. *Nat. Neurosci.* 21, 228–239. doi: 10. 1038/s41593-017-0047-3

FUNDING

This work was supported by LISBOA-01-0145-FEDER-007391, project co-funded by FEDER through POR Lisboa 2020 (Programa Operacional Regional de Lisboa) from PORTUGAL 2020 and Fundação para a Ciência e Tecnologia (FCT) and by a Twinning action (SynaNet) from the EU H2020 program (Project Number: 692340). NR is in receipt of an FCT fellowship (PD/BD/113463/2015) and is a fellow of the M2B-Ph.D. Program.

- Correia-de-Sá, P., and Ribeiro, J. A. (1994). Evidence that the presynaptic A_{2a} -adenosine receptor of the rat motor nerve endings is positively coupled to adenylate cyclase. *Naunyn Schmiedebergs Arch. Pharmacol.* 350, 514–522. doi: 10.1007/BF00173021
- Correia-de-Sá, P., Sebastião, A. M., and Ribeiro, J. A. (1991). Inhibitory and excitatory effects of adenosine receptor agonists on evoked transmitter release from phrenic nerve endings of the rat. *Br. J. Pharmacol.* 103, 1614–1620. doi: 10.1111/j.1476-5381.1991.tb09836.x
- Correia-de-Sá, P., Timóteo, M. A., and Ribeiro, J. A. (1996). Presynaptic A_1 inhibitory/ A_{2A} facilitatory adenosine receptor activation balance depends on motor nerve stimulation paradigm at the rat hemidiaphragm. *J. Neurophysiol.* 76, 3910–3919. doi: 10.1152/jn.1996.76.6.3910
- Costa, J., Gomes, C., and de Carvalho, M. (2010). Diagnosis, pathogenesis and therapeutic targets in amyotrophic lateral sclerosis, CNS and neurological disorders. *Drug Targets* 9, 764–778. doi: 10.2174/187152710793237502
- Cunha, C., Santos, C., Gomes, C., Fernandes, A., Correia, A. M., Sebastião, A. M., et al. (2017). Downregulated glia interplay and increased miRNA-155 as promising markers to track ALS at an early stage. *Mol. Neurobiol.* doi: 10.1007/s12035-017-0631-2 [Epub ahead of print].
- Cunha, R. A. (2016). How does adenosine control neuronal dysfunction and neurodegeneration? J. Neurochem. 139, 1019–1055. doi: 10.1111/jnc. 13724
- Cunha, R. A., and Sebastião, A. M. (1993). Adenosine and adenine nucleotides are independently released from both the nerve terminals and the muscle fibres upon electrical stimulation of the innervated skeletal muscle of the frog. *Pflugers Arch.* 424, 503–510. doi: 10.1007/BF00374914
- Dadon-Nachum, M., Melamed, E., and Offen, D. (2011). The "dying-back" phenomenon of motor neurons in ALS. J. Mol. Neurosci. 43, 470–477. doi: 10.1007/s12031-010-9467-1
- Dai, S. S., and Zhou, Y. G. (2011). Adenosine 2A receptor: a crucial neuromodulator with bidirectional effect in neuroinflammation and brain injury. Rev. Neurosci. 22, 231–239. doi: 10.1515/RNS.2011.020
- Dale, N., and Gilday, D. (1996). Regulation of rhythmic movements by purinergic neurotransmitters in frog embryos. *Nature* 383, 259–263. doi: 10.1038/ 383259a0
- de Mendonça, A., and Cunha, R. A. (2010). Putative neuroprotective effects of caffeine in clinical trials. Concluding remarks. J. Alzheimers Dis. 20(Suppl. 1), S249–S252.
- DeJesus-Hernandez, M., Mackenzie, I. R., Boeve, B. F., Boxer, A. L., Baker, M., Rutherford, N. J., et al. (2011). Expanded GGGGCC hexanucleotide repeat in noncoding region of C9ORF72 causes chromosome 9p-linked FTD and ALS. *Neuron* 72, 245–256. doi: 10.1016/j.neuron.2011.09.011
- Deuchars, S. A., Brooke, R. E., and Deuchars, J. (2001). Adenosine A₁ receptors reduce release from excitatory but not inhibitory synaptic inputs onto lateral horn neurons. J. Neurosci. 21, 6308–6320.
- Di Giorgio, F. P., Carrasco, M. A., Siao, M. C., Maniatis, T., and Eggan, K. (2007). Non-cell autonomous effect of glia on motor neurons in an embryonic stem cell-based ALS model. *Nat. Neurosci.* 10, 608–614. doi: 10.1038/nn1885
- Dias, R. B., Ribeiro, J. A., and Sebastião, A. M. (2012). Enhancement of AMPA currents and GluR1 membrane expression through PKA- coupled adenosine A_{2A} receptors. *Hippocampus* 22, 276–291. doi: 10.1002/hipo.20894
- Dias, R. B., Rombo, D. M., Ribeiro, J. A., and Sebastião, A. M. (2013). Ischemia-induced synaptic plasticity drives sustained expression of calcium-permeable AMPA receptors in the hippocampus. *Neuropharmacology* 65, 114–122. doi: 10.1016/j.neuropharm.2012.09.016

- Diógenes, M. J., Fernandes, C. C., Sebastião, A. M., and Ribeiro, J. A. (2004). Activation of adenosine A_{2A} receptor facilitates brain-derived neurotrophic factor modulation of synaptic transmission in hippocampal slices. *J. Neurosci.* 24, 2905–2913. doi: 10.1523/JNEUROSCI.4454-03.2004
- Dunwiddie, T. V., and Masino, S. A. (2001). The role and regulation of adenosine in the central nervous system. *Annu. Rev. Neurosci.* 24, 31–55. doi: 10.1146/ annurev.neuro.24.1.31
- Feng, Z., and Ko, C. P. (2008a). Schwann cells promote synaptogenesis at the neuromuscular junction via transforming growth factor-β1. *J. Neurosci.* 28, 9599–9609. doi: 10.1523/JNEUROSCI.2589-08.2008
- Feng, Z., and Ko, C. P. (2008b). The role of glial cells in the formation and maintenance of the neuromuscular junction. Ann. N. Y. Acad. Sci. 1132, 19–28. doi: 10.1196/annals.1405.016
- Ferre, S., Ciruela, F., Borycz, J., Solinas, M., Quarta, D., Antoniou, K., et al. (2008). Adenosine A₁-A_{2A} receptor heteromers: new targets for caffeine in the brain. *Front. Biosci.* 13, 2391–2399. doi: 10.2741/2852
- Ferreira, D. G., Batalha, V. L., Vicente Miranda, H., Coelho, J. E., Gomes, R., Gonçalves, F. Q., et al. (2017). Adenosine A_{2A} receptors modulate α -synuclein aggregation and toxicity. *Cereb. Cortex* 27, 718–730.
- Flaten, V., Laurent, C., Coelho, J. E., Sandau, U., Batalha, V. L., Burnouf, S., et al. (2014). From epidemiology to pathophysiology: what about caffeine in Alzheimer's disease? *Biochem. Soc. Trans.* 42, 587–592. doi: 10.1042/BST20130229
- Fredholm, B. B. (2014). Adenosine–a physiological or pathophysiological agent? *J. Mol. Med.* 92, 201–206. doi: 10.1007/s00109-013-1101-6
- Fredholm, B. B., Yang, J., and Wang, Y. (2017). Low, but not high, dose caffeine is a readily available probe for adenosine actions. *Mol. Aspects Med.* 55, 20–25. doi: 10.1016/j.mam.2016.11.011
- Frey, D., Schneider, C., Xu, L., Borg, J., Spooren, W. I., and Caroni, P. (2000).Early and selective loss of neuromuscular synapse subtypes with low sprouting competence in motoneuron diseases. *J. Neurosci.* 20, 2534–2542.
- Fritz, E., Izaurieta, P., Weiss, A., Mir, F. R., Rojas, P., Gonzalez, D., et al. (2013). Mutant SOD1-expressing astrocytes release toxic factors that trigger motoneuron death by inducing hyperexcitability. *J. Neurophysiol.* 109, 2803– 2814. doi: 10.1152/jn.00500.2012
- Fuxe, K., Ferré, S., Genedani, S., Franco, R., and Agnati, L. F. (2007). Adenosine receptor-dopamine receptor interactions in the basal ganglia and their relevance for brain function. *Physiol. Behav.* 92, 210–217. doi: 10.1016/j. physbeh.2007.05.034
- Garcia, N., Priego, M., Hurtado, E., Obis, T., Santafe, M. M., Tomàs, M., et al. (2014). Adenosine A2B and A3 receptor location at the mouse neuromuscular junction. J. Anat. 225, 109–117. doi: 10.1111/joa.12188
- Georgiou, J., Robitaille, R., and Charlton, M. P. (1999). Muscarinic control of cytoskeleton in perisynaptic glia. J. Neurosci. 19, 3836–3846.
- Ginsborg, B. L., and Hirst, G. D. S. (1972). The effect of adenosine on the release of the transmitter from the phrenic nerve of the rat. *J. Physiol.* 224, 629–645. doi: 10.1113/jphysiol.1972.sp009916
- Gitcho, M. A., Baloh, R. H., Chakraverty, S., Mayo, K., Norton, J. B., Levitch, D., et al. (2008). TDP-43 A315T mutation in familial motor neuron disease. Ann. Neurol. 63, 535–538. doi: 10.1002/ana.21344
- Guo, W., Chen, Y., Zhou, X., Kar, A., Ray, P., Chen, X., et al. (2011). An ALS associated mutation affecting TDP-43 enhances protein aggregation, fibril formation and neurotoxicity. *Nat. Struct. Mol. Biol.* 18, 822–830. doi: 10.1038/ nsmb.2053
- Haskó, G., Linden, J., Cronstein, B., and Pacher, P. (2008). Adenosine receptors: therapeutic aspects for inflammatory and immune diseases. *Nat. Rev. Drug Discov.* 7, 759–770. doi: 10.1038/nrd2638
- Ingwersen, J., Wingerath, B., Graf, J., Lepka, K., Hofrichter, M., Schröter, F., et al. (2016). Dual roles of the adenosine A2a receptor in autoimmune neuroinflammation. J. Neuroinflammation 13:48. doi: 10.1186/s12974-016-0512-z
- Jacobson, K. A., Merighi, S., Varani, K., Borea, P. A., Baraldi, S., Aghazadeh Tabrizi, M., et al. (2017). A₃ adenosine receptors as modulators of inflammation: from medicinal chemistry to therapy. *Med. Res. Rev.* doi: 10. 1002/med.21456 [Epub ahead of print].
- Janes, K., Wahlman, C., Little, J. W., Doyle, T., Tosh, D. K., Jacobson, K. A., et al. (2015). Spinal neuroimmune activation is independent of T-cell infiltration and attenuated by ${\rm A_3}$ adenosine receptor agonists in a model of oxaliplatin-induced

- peripheral neuropathy. Brain Behav. Immun. 44, 91–99. doi: 10.1016/j.bbi.2014.
- Kachroo, A., and Schwarzschild, M. A. (2012). Adenosine A_{2A} receptor gene disruption protects in an α-synuclein model of Parkinson's disease. Ann. Neurol. 71, 278–282. doi: 10.1002/ana.22630
- Kaelin-Lang, A., Lauterburg, T., and Burgunder, J. M. (1999). Expression of adenosine A2a receptors gene in the olfactory bulb and spinal cord of rat and mouse. *Neurosci. Lett.* 261, 189–191. doi: 10.1016/S0304-3940(99)00022-1
- Katz, B., and Miledi, R. (1965). Release of acetylcholine from a nerve terminal by electric pulses of variable strength and duration. *Nature* 207, 1097–1098. doi: 10.1038/2071097a0
- Khalid, S. I., Ampie, L., Kelly, R., Ladha, S. S., and Dardis, C. (2017). Immune modulation in the treatment of amyotrophic lateral sclerosis: a review of clinical trials. Front. Neurol. 8:486. doi: 10.3389/fneur.2017.00486
- Kiernan, M. C., Vucic, S., Cheah, B. C., Turner, M. R., Eisen, A., Hardiman, O., et al. (2011). Amyotrophic lateral sclerosis. *Lancet* 377, 942–955. doi: 10.1016/S0140-6736(10)61156-7
- Komaki, S., Ishikawa, K., and Arakawa, Y. (2012). Trk and cAMP-dependent survival activity of adenosine A_{2A} agonist CGS21680 on rat motoneurons in culture. *Neurosci. Lett.* 522, 21–24. doi: 10.1016/j.neulet.2012.06.003
- Laurent, C., Burnouf, S., Ferry, B., Batalha, V. L., Coelho, J. E., Baqi, Y., et al. (2016).
 A_{2A} adenosine receptor deletion is protective in a mouse model of Tauopathy.
 Mol. Psychiatry 21, 97–107. doi: 10.1038/mp.2014.151
- Laurent, C., Eddarkaoui, S., Derisbourg, M., Leboucher, A., Demeyer, D., Carrier, S., et al. (2014). Beneficial effects of caffeine in a transgenic model of Alzheimer's disease-like tau pathology. *Neurobiol. Aging* 35, 2079–2090. doi: 10.1016/j.neurobiolaging.2014.03.027
- Leblond, C. S., Kaneb, H. M., Dion, P. A., and Rouleau, G. A. (2014). Dissection of genetic factors associated with amyotrophic lateral sclerosis. *Exp. Neurol.* 262, 91–101. doi: 10.1016/j.expneurol.2014.04.013
- Leroy, F., Lamotte d'Incamps, B., Imhoff-Manuel, R. D., and Zytnicki, D. (2014). Early intrinsic hyperexcitability does not contribute to motoneuron degeneration in amyotrophic lateral sclerosis. eLife 3:e04046. doi: 10.7554/eLife. 04046
- Little, J. W., Ford, A., Symons-Liguori, A. M., Chen, Z., Janes, K., Doyle, T., et al. (2015). Endogenous adenosine A3 receptor activation selectively alleviates persistent pain states. *Brain* 138, 28–35. doi: 10.1093/brain/awu330
- Liu, Y. J., Ju, T. C., Chen, H. M., Jang, Y. S., Lee, L. M., Lai, H. L., et al. (2015a). Activation of AMP-activated protein kinase α1 mediates mislocalization of TDP-43 in amyotrophic lateral sclerosis. *Hum. Mol. Genet.* 24, 787–801. doi: 10.1093/hmg/ddu497
- Liu, Y. J., Lee, L. M., Lai, H. L., and Chern, Y. (2015b). Aberrant activation of AMP-activated protein kinase contributes to the abnormal distribution of HuR in amyotrophic lateral sclerosis. FEBS Lett. 589, 432–439. doi: 10.1016/j.febslet. 2014.12.029
- Liu, Y. J., Tsai, P. Y., and Chern, Y. (2017). Energy homeostasis and abnormal RNA metabolism in amyotrophic lateral sclerosis. Front. Cell. Neurosci. 11:126. doi: 10.3389/fncel.2017.00126
- Maekawa, S., Leigh, P. N., King, A., Jones, E., Steele, J. C., Bodi, I., et al. (2009).
 TDP-43 is consistently co-localized with ubiquitinated inclusions in sporadic and Guam amyotrophic lateral sclerosis but not in familial amyotrophic lateral sclerosis with and without SOD1 mutations. *Neuropathology* 29, 672–683. doi: 10.1111/j.1440-1789.2009.01029.x
- Marchetto, M. C., Muotri, A. R., Mu, Y., Smith, A. M., Cezar, G. G., and Gage, F. H. (2008). Non-cell-autonomous effect of human SOD1 G37R astrocytes on motor neurons derived from human embryonic stem cells. *Cell Stem Cell* 3, 649–657. doi: 10.1016/j.stem.2008.10.001
- Matos, M., Augusto, E., Santos-Rodrigues, A. D., Schwarzschild, M. A., Chen, J. F., Cunha, R. A., et al. (2012). Adenosine $A_{\rm 2A}$ receptors modulate glutamate uptake in cultured astrocytes and gliosomes. *Glia* 60, 702–716. doi: 10.1002/glia.22290
- Mojsilovic-Petrovic, J., Jeong, G. B., Crocker, A., Arneja, A., David, S., Russell, D., et al. (2006). Protecting motor neurons from toxic insult by antagonism of adenosine A2a and Trk receptors. *J. Neurosci.* 26, 9250–9263. doi: 10.1523/JNEUROSCI.1856-06.2006
- Nagai, M., Re, D. B., Nagata, T., Chalazonitis, A., Jessell, T. M., Wichterle, H., et al. (2007). Astrocytes expressing ALS-linked mutated SOD1 release factors selectively toxic to motor neurons. *Nat. Neurosci.* 10, 615–622. doi: 10.1038/pp.1876

- Nalini, A., Thennarasu, K., Gourie-Devi, M., Shenoy, S., and Kulshreshtha, D. (2008). Clinical characteristics and survival pattern of 1,153 patients with amyotrophic lateral sclerosis: experience over 30 years from India. *J. Neurol. Sci.* 272, 60–70. doi: 10.1016/j.jins.2008.04.034
- Nascimento, F., Pousinha, P. A., Correia, A. M., Gomes, R., Sebastião, A. M., and Ribeiro, J. A. (2014). Adenosine A_{2A} receptors activation facilitates neuromuscular transmission in the pre-symptomatic phase of the SOD1(G93A) ALS mice, but not in the symptomatic phase. *PLoS One* 9:e104081. doi: 10.1371/journal.pone.0104081
- Nascimento, F., Sebastiao, A. M., and Ribeiro, J. A. (2015). Presymptomatic and symptomatic ALS SOD1 (G93A) mice differ in adenosine A_1 and A_{2A} receptor-mediated tonic modulation of neuromuscular transmission. *Purinergic Signal*. 11, 471–480. doi: 10.1007/s11302-015-9465-4
- Naumenko, N., Pollari, E., Kurronen, A., Giniatullina, R., Shakirzyanova, A., Magga, J., et al. (2011). Gender-specific mechanism of synaptic impairment and its prevention by GCSF in a mouse model of ALS. Front. Cell. Neurosci. 5:26. doi: 10.3389/fncel.2011.00026
- Neumann, M., Sampathu, D. M., Kwong, L. K., Truax, A. C., Micsenyi, M. C., Chou, T. T., et al. (2006). Ubiquitinated TDP-43 in frontotemporal lobar degeneration and amyotrophic lateral sclerosis. *Science* 314, 130–133. doi:10.1126/science.1134108
- Nichols, N. L., Van Dyke, J., Nashold, L., Satriotomo, I., Suzuki, M., and Mitchell, G. S. (2013). Ventilatory control in ALS. Respir. Physiol. Neurobiol. 189, 429–437. doi: 10.1016/j.resp.2013.05.016
- Ng, S. K., Higashimori, H., Tolman, M., and Yang, Y. (2015). Suppression of adenosine 2a receptor (A_{2A}R)-mediated adenosine signaling improves disease phenotypes in a mouse model of amyotrophic lateral sclerosis. *Exp. Neurol.* 267, 115–122. doi: 10.1016/j.expneurol.2015.03.004
- Nonaka, T., Kametani, F., Arai, T., Akiyama, H., and Hasegawa, M. (2009). Truncation and pathogenic mutations facilitate the formation of intracellular aggregates of TDP-43. Hum. Mol. Genet. 18, 3353–3364. doi: 10.1093/hmg/ ddp275
- Pasinelli, P., and Brown, R. H. (2006). Molecular biology of amyotrophic lateral sclerosis: insights from genetics. *Nat. Rev. Neurosci.* 7, 710–723. doi: 10.1038/ nrn1971
- Pelligrino, D. A., Xu, H. L., and Vetri, F. (2010). Caffeine and the control of cerebral hemodynamics. J. Alzheimers Dis. 20(Suppl. 1), S51–S62. doi: 10.3233/JAD-2010-091261
- Pokrishevsky, E., Grad, L. I., Yousefi, M., Wang, J., Mackenzie, I. R., and Cashman, N. R. (2012). Aberrant localization of FUS and TDP43 is associated with misfolding of SOD1 in amyotrophic lateral sclerosis. *PLoS One* 7:e35050. doi: 10.1371/journal.pone.0035050
- Popoli, P., Frank, C., Tebano, M. T., Potenza, R. L., Pintor, A., Domenici, M. R., et al. (2003). Modulation of glutamate release and excitotoxicity by adenosine A_{2A} receptors. *Neurology* 61(11 Suppl. 6), S69–S71. doi: 10.1212/01.WNL. 0000095216.89483.A2
- Potenza, R. L., Armida, M., Ferrante, A., Pèzzola, A., Matteucci, A., Puopolo, M., et al. (2013). Effects of chronic caffeine intake in a mouse model of amyotrophic lateral sclerosis. *J. Neurosci. Res.* 91, 585–592. doi: 10.1002/jnr.23185
- Pousinha, P. A., Correia, A. M., Sebastião, A. M., and Ribeiro, J. A. (2010). Predominance of adenosine excitatory over inhibitory effects on transmission at the neuromuscular junction of infant rats. *J. Pharmacol. Exp. Ther.* 332, 153–163. doi: 10.1124/jpet.109.157255
- Pousinha, P. A., Correia, A. M., Sebastião, A. M., and Ribeiro, J. A. (2012). Neuromuscular transmission modulation by adenosine upon aging. Neurobiol. Aging 33, 2869–2880. doi: 10.1016/j.neurobiolaging.2012. 01.008
- Pousinha, P. A., Diogenes, M. J., Ribeiro, J. A., and Sebastião, A. M. (2006). Triggering of BDNF facilitatory action on neuromuscular transmission by adenosine A_{2A} receptors. *Neurosci. Lett.* 404, 143–147. doi: 10.1016/j.neulet. 2006.05.036
- Qian, K., Huang, H., Peterson, A., Hu, B., Maragakis, N. J., Ming, G. L., et al. (2017). Sporadic ALS astrocytes induce neuronal degeneration in vivo. *Stem Cell Rep.* 8, 843–855. doi: 10.1016/j.stemcr.2017.03.003
- Rebola, N., Simões, A. P., Canas, P. M., Tomé, A. R., Andrade, G. M., Barry, C. E., et al. (2011). Adenosine A_{2A} receptors control neuroinflammation and consequent hippocampal neuronal dysfunction. *J. Neurochem.* 117, 100–111. doi: 10.1111/j.1471-4159.2011.07178.x

- Ribeiro, F. F., Neves-Tomé, R., Assaife-Lopes, N., Santos, T. E., Silva, R. F. M., Brites, D., et al. (2016a). Axonal elongation and dendritic branching is enhanced by adenosine $A_{2\Lambda}$ receptors activation in cerebral cortical neurons. *Brain Struct. Funct.* 221, 2777–2799. doi: 10.1007/s00429-015-1072-1
- Ribeiro, F. F., Xapelli, S., Miranda-Lourenço, C., Tanqueiro, S. R., Fonseca-Gomes, J., Diógenes, M. J., et al. (2016b). Purine nucleosides in neuroregeneration and neuroprotection. *Neuropharmacology* 104, 226–242. doi: 10.1016/j.neuropharm.2015.11.006
- Ribeiro, J. A., and Sebastião, A. M. (1987). On the role, inactivation and origin of endogenous adenosine at the frog neuromuscular junction. *J. Physiol.* 384, 571–585. doi: 10.1113/jphysiol.1987.sp016470
- Ribeiro, J. A., and Sebastião, A. M. (2010). Caffeine and adenosine. *J. Alzheimers Dis.* 20(Suppl. 1), S3–S15. doi: 10.3233/JAD-2010-1379
- Ribeiro, J. A., Sebastião, A. M., and De Mendonça, A. (2003). Adenosine receptors in the nervous system: pathophysiological implications. *Prog. Neurobiol.* 68, 377–392. doi: 10.1016/S0301-0082(02)00155-7
- Ribeiro, J. A., and Walker, J. (1973). Action of adenosine triphosphate on endplate potentials recorded from muscle fibres of the rat-diaphragm and frog sartorius. Br. J. Pharmacol. 49, 724–725. doi: 10.1111/j.1476-5381.1973.tb08 555.x
- Ribeiro, J. A., and Walker, J. (1975). The effects of adenosine triphosphate and adenosine diphosphate on transmission at the rat and frog neuromuscular junctions. *Br. J. Pharmacol.* 54, 213–218. doi: 10.1111/j.1476-5381.1975. tb06931.x
- Rocha, M. C., Pousinha, P. A., Correia, A. M., Sebastião, A. M., and Ribeiro, J. A. (2013). Early changes of neuromuscular transmission in the SOD1(G93A) mice model of ALS start long before motor symptoms onset. *PLoS One* 8:e73846. doi: 10.1371/journal.pone.0073846
- Rodrigues, T. M., Jerónimo-Santos, A., Outeiro, T. F., Sebastião, A. M., and Diógenes, M. J. (2014). Challenges and promises in the development of neurotrophic factor-based therapies for Parkinson's disease. *Drugs Aging* 31, 239–261. doi: 10.1007/s40266-014-0160-x
- Santiago, A. R., Baptista, F. I., Santos, P. F., Cristóvão, G., Ambrósio, A. F., Cunha, R. A., et al. (2014). Role of microglia adenosine A_{2A} receptors in retinal and brain neurodegenerative diseases. *Mediators Inflamm*. 2014:465694. doi: 10.1155/2014/465694
- Sebastião, A. M., Assaife-Lopes, N., Diógenes, M. J., Vaz, S. H., and Ribeiro, J. A. (2011). Modulation of brain-derived neurotrophic factor (BDNF) actions in the nervous system by adenosine A_{2A} receptors and the role of lipid rafts. *Biochim. Biophys. Acta* 1808, 1340–1349. doi: 10.1016/j.bbamem.2010.06.028
- Sebastião, A. M., and Ribeiro, J. A. (1985). Enhancement of transmission at the frog neuromuscular junction by adenosine deaminase: evidence for an inhibitory role of endogenous adenosine on neuromuscular transmission. *Neurosci. Lett.* 62, 267–270. doi: 10.1016/0304-3940(85)90366-0
- Sebastião, A. M., and Ribeiro, J. A. (2009a). Adenosine receptors and the central nervous system. Handb. Exp. Pharmacol. 193, 471–534. doi: 10.1007/978-3-540-89615-9_16
- Sebastião, A. M., and Ribeiro, J. A. (2009b). Triggering neurotrophic factor actions through adenosine A_{2A} receptor activation: implications for neuroprotection. Br. J. Pharmacol. 158, 15–22. doi: 10.1111/j.1476-5381.2009.00157.x
- Sebastião, A. M., Stone, T. W., and Ribeiro, J. A. (1990). On the inhibitory adenosine receptor at the neuromuscular junction and hippocampus of the rat: antagonism by 1,3,8-substituted xanthines. *Br. J. Pharmacol.* 101, 453–459. doi: 10.1111/j.1476-5381.1990.tb12729.x
- Selvaraj, B. T., Livesey, M. R., Zhao, C., Gregory, J. M., James, O. T., Cleary, E. M., et al. (2018). C9ORF72 repeat expansion causes vulnerability of motor neurons to Ca²⁺-permeable AMPA receptor-mediated excitotoxicity. Nat. Commun. 9:347. doi: 10.1038/s41467-017-02729-0
- Shan, X., Vocadlo, D., and Krieger, C. (2009). Mislocalization of TDP-43 in the G93A mutant SOD1 transgenic mouse model of ALS. *Neurosci. Lett.* 458, 70–74. doi: 10.1016/j.neulet.2009.04.031
- Shi, Y., Lin, S., Staats, K. A., Li, Y., Chang, W. H., Hung, S. T., et al. (2018). Haploinsufficiency leads to neurodegeneration in C9ORF72 ALS/FTD human induced motorneurons. *Nat. Med.* 24, 313–325. doi: 10.1038/nm.4490
- Sitkovsky, M. V., Lukashev, D., Apasov, S., Kojima, H., Koshiba, M., Caldwell, C., et al. (2004). Physiological control of immune response and inflammatory tissue damage by hypoxia-inducible factors and adenosine A_{2A} receptors. *Annu. Rev. Immunol.* 22, 657–682. doi: 10.1146/annurev.immunol.22.012703.104731

- Smith, B. N., Newhouse, S., Shatunov, A., Vance, C., Topp, S., Johnson, L., et al. (2013). The C9ORF72 expansion mutation is a common cause of ALS+/-FTD in Europe and has a single founder. Eur. J. Hum. Genet. 21, 102–108. doi: 10.1038/ejhg.2012.98
- Son, Y. J., Trachtenberg, J. T., and Thompson, W. J. (1996). Schwann cells induce and guide sprouting and reinnervation of neuromuscular junctions. *Trends Neurosci.* 19, 280–285. doi: 10.1016/S0166-2236(96)10032-1
- Taylor, J. P., Brown, R. H. Jr., and Cleveland, D. W. (2016). Decoding ALS: from genes to mechanism. *Nature* 539, 197–206. doi: 10.1038/nature20413
- Tsentsevitsky, A., Kovyazina, I., Nikolsky, E., Bukharaeva, E., and Giniatullin, R. (2013). Redox-sensitive synchronizing action of adenosine on transmitter release at the neuromuscular junction. *Neuroscience* 248, 699–707. doi: 10.1016/j.neuroscience.2013.05.065
- Vincenzi, F., Corciulo, C., Targa, M., Casetta, I., Gentile, M., Granieri, E., et al. (2013). A_{2A} adenosine receptors are up-regulated in lymphocytes from amyotrophic lateral sclerosis patients. *Amyotroph. Lateral Scler. Frontotemporal Degener.* 14, 406–413. doi: 10.3109/21678421.2013.793358
- Volonté, C., Apolloni, S., Parisi, C., and Amadio, S. (2016). Purinergic contribution to amyotrophic lateral sclerosis. *Neuropharmacology* 104, 180–193. doi: 10.1016/j.neuropharm.2015.10.026
- Wainger, B. J., Kiskinis, E., Mellin, C., Wiskow, O., Han, S. S., Sandoe, J., et al. (2014). Intrinsic membrane hyperexcitability of amyotrophic lateral sclerosis patient-derived motorneurons. *Cell Rep.* 7, 1–11. doi: 10.1016/j.celrep.2014. 03.019
- Wiese, S., Jablonka, S., Holtmann, B., Orel, N., Rajagopal, R., Chao, M. V., et al. (2007). Adenosine receptor A_{2A}-R contributes to motoneuron survival by transactivating the tyrosine kinase receptor TrkB. *Proc. Natl. Acad. Sci. U.S.A.* 104, 17210–17215. doi: 10.1073/pnas.0705267104

- Wijesekera, L. C., and Leigh, P. N. (2009). Amyotrophic lateral sclerosis. *Orphanet J. Rare Dis.* 4:3. doi: 10.1186/1750-1172-4-3
- Witts, E. C., Nascimento, F., and Miles, G. B. (2015). Adenosine-mediated modulation of ventral horn interneurons and spinal motoneurons in neonatal mice. J. Neurophysiol. 114, 2305–2315. doi: 10.1152/jn.00574.2014
- Witts, E. C., Panetta, K. M., and Miles, G. B. (2012). Glial-derived adenosine modulates spinal motor networks in mice. J. Neurophysiol. 107, 1925–1934. doi: 10.1152/jn.00513.2011
- Yanpallewar, S. U., Barrick, C. A., Buckley, H., Becker, J., and Tessarollo, L. (2012). Deletion of the BDNF truncated receptor TrkB.T1 delays disease onset in a mouse model of amyotrophic lateral sclerosis. *PLoS One* 7:e39946. doi: 10.1371/journal.pone.0039946
- Yoshida, Y., Une, F., Utatsu, Y., Nomoto, M., Furukawa, Y., Maruyama, Y., et al. (1999). Adenosine and neopterin levels in cerebrospinal fluid of patients with neurological disorders. *Intern. Med.* 38, 133–139. doi: 10.2169/internalmedicine.38.133

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Sebastião, Rei and Ribeiro. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms