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# First report on Basidiomycota fungi in sorghum and millet from Southwest Nigeria

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**Abstract.** Trametes species are mushroom fungi with several biotechnological applications. This includes decolourisation of wastewater from olive mill and elimination of endocrine-disrupting hormones. This study reports the presence of two Trametes species, namely *Trametes polyzona* and *Trametes villosa* from the phylum Basidiomycota in sorghum and millet in Southwest Nigeria. These fungal isolates were identified culturally and further identified through phylogenetic characterisation. *Trametes* spp. occurred in 10% of sorghum samples and 20% of millet samples. The two species were morphologically similar but distantly related phylogenetically. Most fungal species present in cereal crops belong to the division Ascomycota. However, two *Trametes* species belonging to division Basidiomycota are being reported for the first time in cereal crops. *Trametes* spp. can be harnessed for their health benefits such as the treatment of cancer and the reduction of viral activity in humans.

**Keywords:** *Trametes* species, Basidiomycota, sorghum, millet, Southwest Nigeria

## 1. Introduction

Sorghum (*Sorghum bicolor*), is the fourth most cultivated crop after paddy, wheat, and maize in the world [1]. Nigeria, which is ranked third among the world producer of sorghum after



United States of America and India, produces 40% of the total sorghum recorded in Africa [2]. Millet (*Pennisetum glaucum*) is the sixth most important grain in the world, produced mainly by China, India, and Nigeria [1, 3]. Fungal contamination occurs in these cereals in the field and also during the process of drying, processing, transporting and storage, which thus enhance the production of mycotoxins under favourable conditions [4]. Fungi which colonise cereals include species of *Fusarium*, *Aspergillus*, and *Penicillium*, which significantly reduce crop yield, quality and safety owing to their capacity to produce mycotoxins [4]. Sorghum and millet are among cereals that get contaminated both on the field and during storage [5, 6]. These cereals are staple foods consumed in Southwest Nigeria in the form of fermented gruels such as 'Ogi', fermented drinks such as 'kunu' and snacks such as 'donwka' and 'kokoro'. Oranusiet *al.* [6] attributed the presence of fungi in cereal grains to the normal flora of the plants as well as contamination of the grains from the field or during storage and display for sales.

## 2. Sample collection

One hundred samples each of sorghum and millet were collected from the six states in the South-west of Nigeria, namely: Lagos, Ogun, Osun, Ondo, Ekiti and Oyo. After that, twenty composite samples each of sorghum and millet were separately prepared for the isolation of the fungal species.

## 3. Materials and Methods

### 3.1 Isolation of Fungi from the cereal grains

One gram of each ground composite grain sample was homogenised in 9 mL of sterile 0.1% peptone water solution, vortexed and serial dilutions to  $10^{-10}$  were carried out. Aliquots of the various dilutions were inoculated onto freshly prepared Potato Dextrose Agar plates and incubated for 5 days at 25°C. The inoculated plates were observed for fungal colonies after 5 days of incubation. Distinct colonies from the culture plates were subcultured onto fresh Czapek Yeast Extract agar plates and incubated at 25°C for 5 days. The identification of pure isolates was carried out based on their macroscopic and microscopic features according to the keys of Pitt and Hockings [7].

### 3.2 Genomic DNA Extraction

Fresh mycelium from purified culture was collected in sterile 2 mL Eppendorf tubes. Approximately 500  $\mu$ l of cetyltrimethylammonium bromide (CTAB) (1 M Tris-HCl, pH 8.0), 0.5M EDTA, 5M NaCl, 20 g CTAB) buffer was added to the tubes. Mycelium was then ground against the wall of the tube using sterile pipette tips for genomic DNA (gDNA) isolation using the method described by Aamir *et al.* [8]. A Nanodrop spectrophotometer was used to quantify the DNA of the fungi.

### 3.3 Polymerase Chain Reaction (PCR) Analysis

The isolated DNA was subjected to Polymerase Chain Reaction (Veriti thermal cycler, Applied Biosystems) as described by Aamir *et al.* [8]. Isolates were putatively identified by analysis of sequences of the Internal Transcribed Spacer (ITS) gene. These sequences were amplified using the primers ITS1 F (TCCGTAGGTGAACCTGCGG) and ITS4 R (TCCTCCGCTTATTGATATGC). Amplicons were confirmed with gel electrophoresis using a 1% agarose gel. The PCR products were then purified with gel filtration columns loaded with 6% Sephadex G-50 (50-150  $\mu$ m bead size) (Sigma).

### 3.4 Sequencing of PCR Products

The purified PCR products were sequenced using forward and reverse primers with ITS1 and ITS4 primers, respectively, using the ABI Prism<sup>TM</sup> 3500 x 1 Genetic Analyzer (Applied Biosystems, CA, USA). The ABI Prism<sup>TM</sup> BigDye® Terminator v3.1 Cycle Sequencing Kit (Applied Biosystems) was used with protocols supplied by the manufacturer.

### 3.5 Sequence Data Analysis

Sequence reads generated from ITS1, and ITS4 primers were assembled into consensus sequences using the BioEdit software package. These sequences were then compared to those in the National Center for Biotechnology Information (NCBI) (<http://www.ncbi.nlm.nih.gov>) nucleotide database using BLAST searches. The putative identity of isolates was based on the highest similarity score to the newly produced sequences.

### 3.6 Phylogenetic Analyses

The evolutionary relationship of fungi isolated from maize, sorghum and millet were obtained from a Neighbour-Joining analysis as reported in Casaet *al.* [9]. The evolutionary distances were determined by the Maximum Composite Likelihood method, as referenced by Tamuraet *al.* [10]. The phylogenetic and molecular evolution analyses were conducted using MEGA X Software referenced by Kumar *et al.* [11].

## 4. Results and Discussion

The phylogenetic tree for the *Trametes* species which was grouped into six clades revealed that *Trametespolyzona* MKO20685 was distantly related to other species such as *Trametespolyzona* voucher\_OH272sp, *Trametespolyzona* strain DMC367, *Trametespolyzona* voucher BKW004 and *Neofomitellapolyzonata* voucher Dai\_10980 all in clade 1 as shown in Fig. 2. *Trametesvillosa* MKO20684 was distantly related to *Trametesvillosa* culture collection\_BRFMFRA\_1375, *Trametesvillosa* strain CBS\_334.49, *Trametesvillosa* strain\_Sc10 and *Trametesvillosaisolate\_P4*. Two different species of *Trametes* were isolated from millet in this study. These are *Trametesvillosa* and *Trametespolyzona* (Plate 1 and 2, respectively).

There is a lack of information on the presence of *Trametespolyzona* and *T. villosa* in cereals. However, these *Trametes* species were isolated in some sorghum and millet samples in this study. *Trametes* species have been reported in forests and dead and decaying hardwood [12]; thus, it may be considered as one of the field fungi in cereals. *Trametespolyzona*, previously named *Corioloropsispolyzona*, was first described as Basionym *Polyporuspolyzonus* [13]. *Trametes* species isolated in the sorghum and millet grains were most likely introduced as spores from the field where the cereals were grown before the grains were bagged and transported for sale in the markets. Cereal grains for sale in the market are usually kept in stores away from moisture until they are sold. However, these grains may also be affected by climatic conditions such as the temperature of the storage environment, which is temperate. Several fungi in grains are pathogenic and are mycotoxin producers.

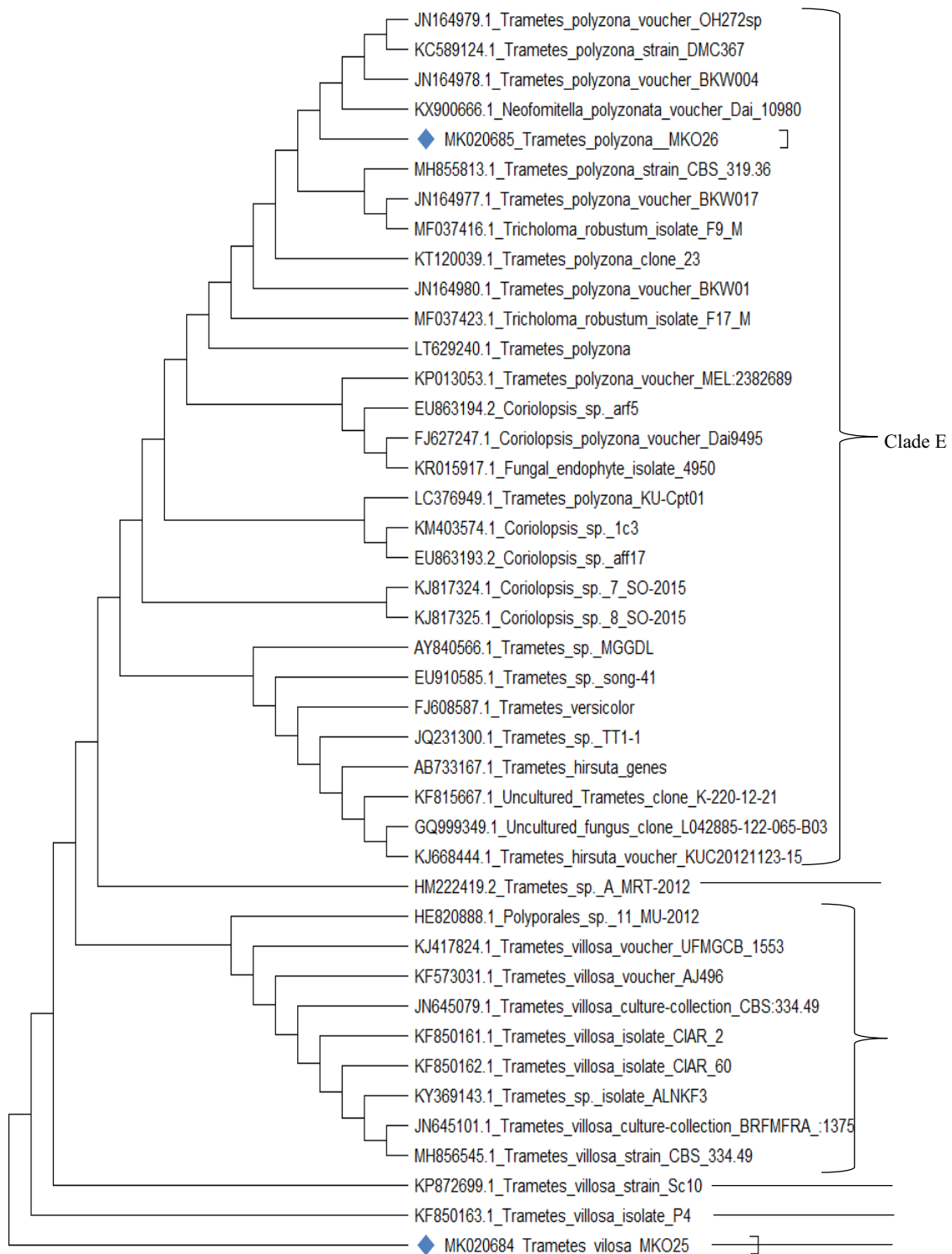


Figure 1: Phylogenetic tree of *Trametes* spp created by the Neighbour-joining Method

Therefore, various measures have been employed to ensure food sustainability such as fungal decontamination methods using *Cymbopogon citratus* in maize [14] and mycotoxin decontamination methods in cereal grains using a combination of Montmorillonite clay and *Cymbopogon citratus* [15, 16]. However, the presence of *Trametes* sp in cereals such as sorghum and millet may not pose a health risk as they have not been reported to produce mycotoxins but rather have potential uses in medicine and the industry.

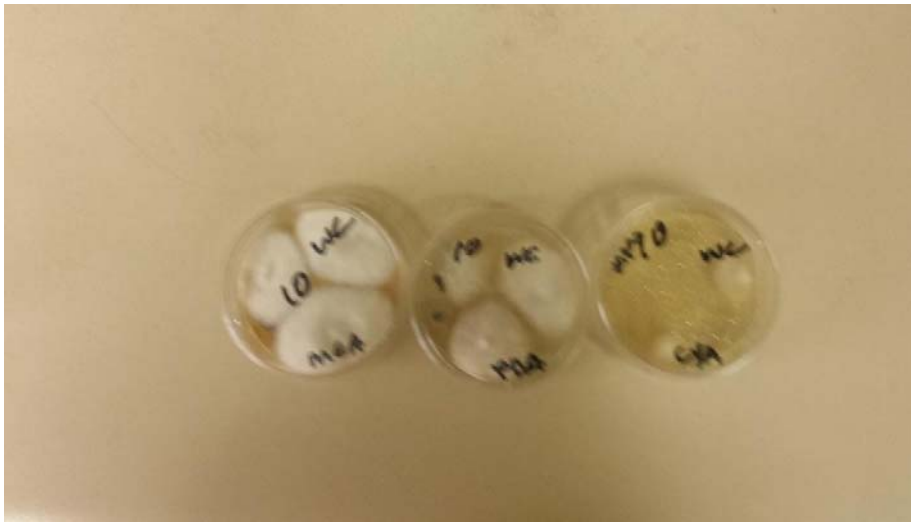


Plate 1: *Trametes villosa* on Malt Extract Agar, Potato Dextrose Agar and Czapek Yeast Agar

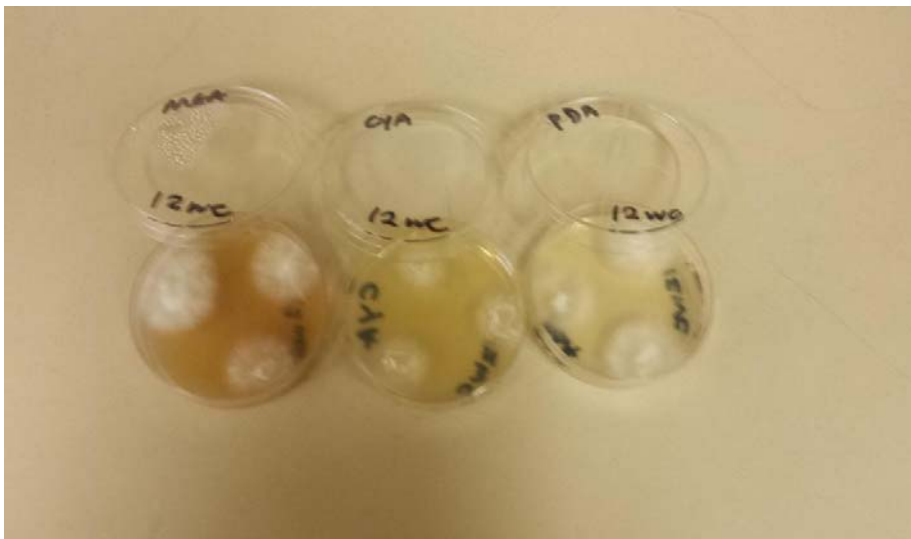


Plate 2: *Trametes polyzona* on Malt Extract Agar, Czapek Yeast Agar and Potato Dextrose Agar

## 5. Conclusion and Recommendation

This study documents for the first time, the occurrence of *Trametes* in cereals from Southwest Nigeria. *Trametes* species have biotechnological potentials in bioremediation and medicine. Hence, it can be beneficial in the economic development of the nation. Cereal grains across other geopolitical zones in Nigeria should be screened for the presence of *Trametes* species.

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## References

- [1] Food and Agriculture Organization. FAO (2020). Sorghum. Post-Harvest Operations. Accessed [www.fao.org](http://www.fao.org) > file admin > user\_upload > inpho > docs, February 27, 2020.
- [2] Food and Agriculture Organization. FAO. (2016). Food Security Assessment in the Northeastern States of Nigeria; FAO Rapid Seed System Assessment Report; FAO: Rome, Italy, pp. 16–64.
- [3] Makun H. A., Gbodi T. A., Akanya O. H., Salako A. E. and Ogbadu G. H. (2007). Fungi and some mycotoxins contaminating rice (*Oryza sativa*) in Niger State, Nigeria. *African Journal of Biotechnology*, 6(2): 99 – 108.
- [4] Alkenz S., Sassi A. A., Abugnah Y. S. and Alryani M. B. (2015). Isolation and identification of fungi associated with some Libyan foods. *African Journal of Food Science*, 9(7): 406-410.
- [5] Makun H. A., Gbodi T. A., Akanya O. H., Salako A. E. and Ogbadu G. H. (2009). Health implication of toxigenic fungi found in two Nigerian staples: guinea corn and rice. *African Journal of Food Science*, 3: 250 – 256.
- [6] Oranusi S., Nwankwo U. E., Onu-Okpara I. and Olopade B. K. (2016). Assessment of Microflora, Deoxynivalenol (DON) and Fumonisin Contamination of Grains sold in Local Markets, Nigeria. *Covenant Journal of Physical and Life Sciences*, 4(2): 42-49.
- [7] Pitt J. I. and Hocking A. D. (2009). Fungi and Food Spoilage. (2009). Springer-Verlag, USA.
- [8] Aamir S., Sutar S., Singh S. K. and Baghela, A. (2015). A rapid and efficient method of fungal genomic extraction suitable for PCR-based molecular methods. *Plant Pathology and Quarantine*, 5(2): 74–81.
- [9] Casa A. M., Mitchell S. E., Hamblin M. T., Sun H., Bowers J. E., Paterson A. H., Aquadro C. F. and Kresovich S. (2005). Diversity and selection in sorghum: Simultaneous analyses using simple sequence repeats. *Theoretical and Applied Genetics*, 111(1): 23-30.
- [10] Tamura K., Nei M. and Kumar S. (2004). Prospects for inferring very large phylogenies by using the neighbour-joining method. *Proceedings of the National Academy of Sciences (USA)* 101: 11030-11035.
- [11] Kumar S., Stecher G., Li M., Knyaz C. and Tamura K. (2018). MEGA X: Molecular Evolutionary Genetics Analysis across Computing Platforms. *Molecular Biology and Evolution*, 35(6): 1547–1549.

- [12] Park J. H., Pavlov I. N., Kim M. J., Park M. S., Oh S. Y., Park K. H., Fong, J. J. and Lim, Y. W. (2020). Investigating Wood Decaying Fungi Diversity in Central Siberia, Russia Using ITS Sequence Analysis and Interaction with Host Trees. *Sustainability*, 12, 2535. <http://dx.doi.org/10.3390/su12062535>.
- [13] Justo A. and Hibbett D. S. (2011). Phylogenetic classification of *Trametes* (Basidiomycota, Polyporales) based on a five-gene dataset. *Taxon*, 60(6): 1567-1583.
- [14] Atanda, O. O. and Olopade, T. A. (2013). Effect of lemongrass (*Cymbopogon citratus* (DC) Stapf.) treatments on *Aspergillus flavus* (SGS-421) infestation and Aflatoxin B<sub>1</sub> content of maize grains. *International Food Research Journal*, 20(4): 1933-1939.
- [15] Olopade B.K., Oranusi S.U., Nwinyi O.C., Njobeh P.B. (2019a). Modification of montmorillonite clay with *Cymbopogon citratus* for the decontamination of zearalenone in millet. *AIMS Agriculture and Food*, 4(3): 643–657. DOI: 10.3934/agrfood.2019.3.643.
- [16] Olopade, B. K., Oranusi, S. U., Nwinyi, O.C., Lawal, I. A, Gbashi, S., Njobeh, P. B. (2019). Decontamination of T-2 toxin in maize using modified Montmorillonite clay. *Toxins*, 11(11): 616. <https://doi.org/10.3390/toxins11110616>.
- [17] Bukh C., Lund M. and Bjerrum M. J. (2006). Kinetic studies on the reaction between *Trametes villosa* laccase and dioxygen, *Journal of Inorganic Biochemistry*, 100: 1547–1557.
- [18] Silva M. L. C., Souza V. B., Santos V. S, Kamida H. M., Vasconcellos-Neto J. R. T., Góes-Neto A. and Koblitz M. G. B. (2014). Production of manganese peroxidase by *Trametes villosa* on inexpensive substrate and its application in the removal of lignin from agricultural wastes. *Advances in Bioscience Biotechnology*, 5: 1067–1077.
- [19] Carneiro R. T., Lopes M. A., Silva M. L., Santos V. D., Souza V. B., Sousa A. O., Pirovani C. P., Koblitz M. G., Benevides R. G. and Góes-Neto A. (2017). *Trametes villosa* lignin peroxidase (TvLiP): genetic and molecular characterisation. *Journal of Microbiology and Biotechnology*, 27(1): 179–188.
- [20] Heemken O.P., Reincke H., Stachel B. and Theobald N. (2001). “The occurrence of xenoestrogens in the Elbe river and the North Sea,” *Chemosphere*, vol. 45, no. 3, pp. 245–259.
- [21] Zeng S., Zhao J. and Xia L. (2017). Simultaneous production of laccase and degradation of bisphenol A with *Trametes versicolor* cultivated on agricultural wastes. *Bioprocess and Biosystems Engineering*, 40: 1237–1245. <https://doi.org/10.1007/s00449-017-1783-1>.